

Clockor2: Inferring global and local strict molecular clocks using root-to-tip regression

Leo A. Featherstone^{*,1}, Andrew Rambaut², Sebastian Duchene^{†,1}, Wytamma Wirth^{†,1}

¹ Peter Doherty Institute for Infection and Immunity, University of Melbourne, Australia.

² Institute of Ecology and Evolution, University of Edinburgh, Edinburgh, UK

*email: leo.featherstone@unimelb.edu.au

[†] These authors supervised this work equally.

Abstract

Molecular sequence data from rapidly evolving organisms are often sampled at different points in time. Sampling times can then be used for molecular clock calibration. The root-to-tip (RTT) regression is an essential tool to assess the degree to which the data are amenable to such analyses and behave in a clock-like fashion. Here, we introduce Clockor2, a client-side web application for conducting RTT regression. Clockor2 uniquely allows users to quickly fit local and global molecular clocks, thus handling the increasing complexity of genomic datasets that sample beyond the assumption homogeneous host populations. Clockor2 is efficient, handling trees of up to the order of 10^4 tips, with significant speed increases compared to other RTT regression applications. Although clockor2 is written as a web application, all data processing happens on the client-side, meaning that data never leaves the user's computer. Clockor2 is freely available at <https://clockor2.github.io/>.

Keywords: Molecular clock, evolutionary rate heterogeneity, root-to-tip regression.

Introduction

Phylogenetic analyses make use of genetic sequence data to understand the evolution, epidemiological, and ecological dynamics of a pathogen. Importantly, phylogenetics achieves its greatest value when generating insight about infectious disease dynamics outside the purview of epidemiology. This frequently occurs at population interfaces, such as during transmission across host sub-populations, geographical boundaries or host species. Despite the increasing complexity of such datasets, the essential component to all phylogenetic modelling is the assumption of a molecular clock relating epidemiological and evolutionary timescales (?).

The simplest molecular clock model is the strict clock, which assumes a constant rate of substitution per unit time, known as the 'evolutionary rate'. When the evolutionary rate is constant throughout a phylogenetic tree, the term global molecular clock is used. In contrast, a local strict clock refers to the situation where different substitution rates apply to different monophyletic groups within a tree (Ho and Duchêne, 2014). The branches of local clocks are sometimes referred to as 'foreground' while the remaining branches are known as the 'background' (Yoder and Yang, 2000). The assumption of a local clock may for example reflect samples from different host populations, host species, or pathogen lineages (Worobey et al., 2014). For example, Clockor2 includes a default example from Porter et al. (2023), where local clocks are fit to human and mink SARS-CoV-2 host populations. Clockor2 enables rapid inference of global and local strict molecular clocks from phylogenetic trees where tips are annotated with sampling times, using root-to-tip regression (RTT). Several other tools allow for the inference of strict molecular clocks via RTT, but none readily offer the ability to fit local clocks models (Rambaut et al., 2016; Hadfield et al., 2018; Sagulenko et al., 2018; Volz and Frost, 2017).

Phylogenetic datasets are and will continue to grow in size and scope Featherstone et al. (2022). For

example, datasets of thousands to tens-of-thousands have been used to understand the spread of SARS-CoV-2 at national and international scales, as well as study the emergence of variants of concern (VOC), and transmission to other species (du Plessis et al., 2021; Hill et al., 2022; Nadeau et al., 2023; Porter et al., 2023). Larger datasets, as a function of their size, are more likely to be sampled from increasingly distinct populations, meaning that local clocks will become increasingly important. Currently, testing the fit of a local clock over alternative models, such as global or relaxed clocks, frequently requires intensive computational efforts using common Bayesian phylodynamic applications such as BEAST or RevBayes (Bouckaert et al., 2019; Suchard et al., 2018; Höhna et al., 2016; ?; ?), each requiring hours to days of computational time. Clockor2 uniquely offers a scalable and accessible client-side web application to perform the same function, with results available in seconds to minutes to inform subsequent phylodynamic analysis.

Specifically, Clockor2 allows users to perform RTT regression for fitting global and local clocks (Fig 1). The user begins by dropping or importing a newick tree. Sampling dates and group identifiers can be parsed from tip labels or separate files on input. Like other RTT regression applications, Clockor2 also allows users to infer the best fitting root based on the R^2 value of the RTT regression, a key indicator of clock-like evolution. It also offers users a local clock-search function to test assumptions about local clocks in a dataset.

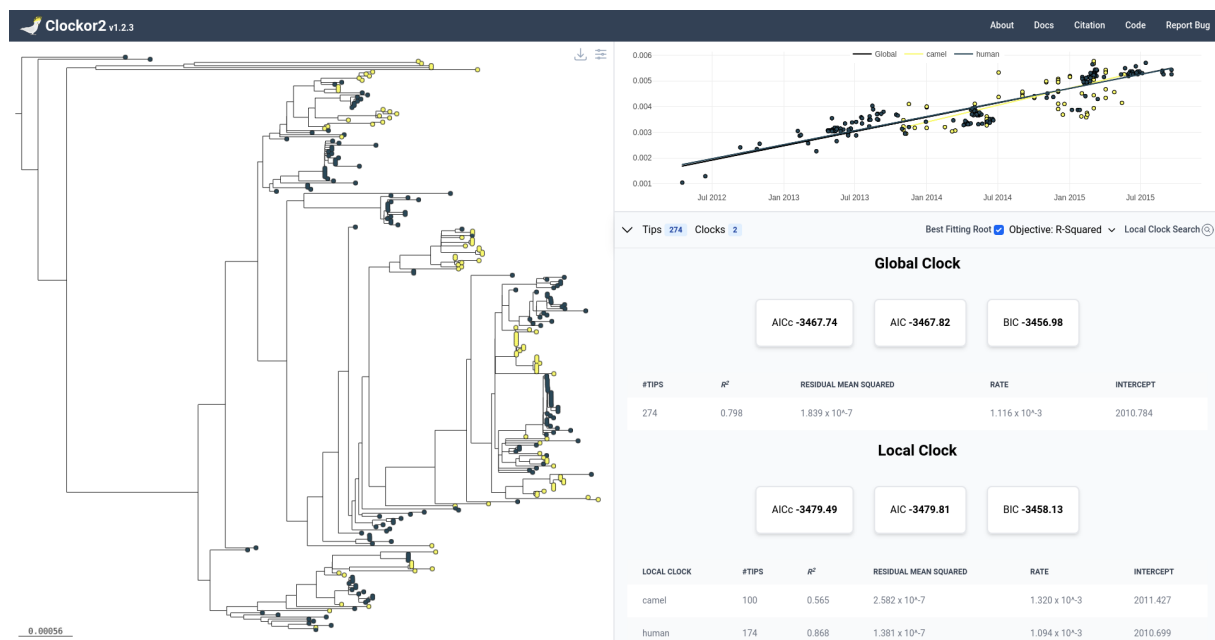


Figure 1: Clockor2 presents the tree along size RTT regression data. Users can toggle between local and global clocks and alter the appearance of the tree.

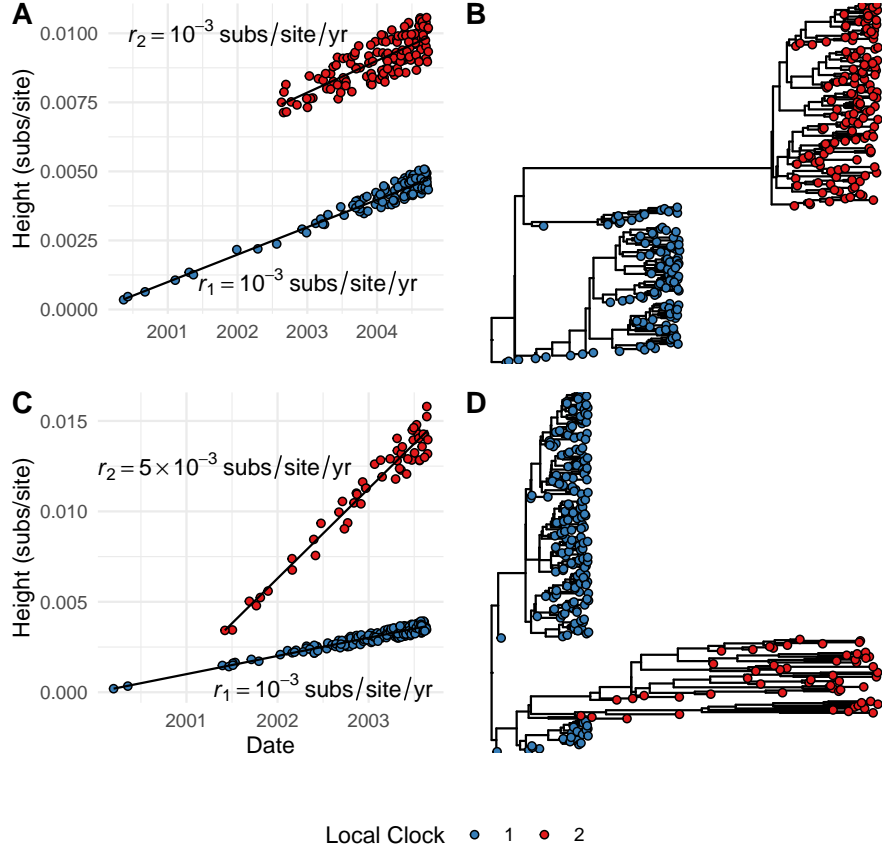


Figure 2: Simulated examples of how local clocks may manifest in trees and RTT regression data. (A) RTT regression data for two local clocks with similar rates separated by a long branch. (B) A tree characteristic of two similar local clock rates separated by a long branch. (C) RTT regression data where two local clocks have different evolutionary rates. (D) A tree characteristic of two local clocks with different rates.

General model for global and local strict clocks

RTT regression consists in modelling the evolutionary rate as the slope of a linear regression of the distance from the root to each tip (RTT distance), typically in units of substitutions per site (subs/site), against the sampling date of each tip. If we denote the evolutionary rate as r (usually in units of *subs/site/time*), RTT distance as d (usually in units of *subs/site*), o as the intercept (interpreted as origin), and sampling times as t , then the model for a global strict clock takes the form:

$$d = rt + o + \epsilon$$

where ϵ is an error term.

Clockor2 uses a generalisation of this model to accommodate local clocks. For a given tree with a set of tips T , we define local clocks as pertaining to *groups* of tips g_i and a rate parameter for each (r_i). For a strict clock model with two local clocks, we then write:

$$d = \begin{cases} r_1 t + o_1 + \epsilon, & \text{if tip} \in g_1 \\ r_2 t + o_2 + \epsilon, & \text{if tip} \in g_2 \end{cases}$$

We refer to *groups* instead of *clades* because while collections of tips belonging to one local clock necessarily share a common ancestor, they do not necessarily comprise a whole clade. This occurs when two or more local clocks are nested. The tips comprising the outer clock(s) cannot comprise a whole clade if another local clock is nested within it. For example, local clock 1 in Fig. 2 B,D does not comprise a clade (i.e. is not monophyletic) because local clock 2 is nested within it.

This general model then captures the two key scenarios where local clocks may be appropriate. The first is where rates are similar between local clocks, but separated by a long branch (Fig. 2A-B). For example, in the evolution of VOCs in SARS-CoV-2 or due to temporally-sparse sampling in the case of ancient *Yersinia pestis* samples (Tay et al., 2022; Eaton et al., 2023; Hill et al., 2022). The second scenario is where rates differ between local clocks (Fig 2 C-D). For example, this can occur when a

pathogen spreads in different host species, such as has been observed for SARS-CoV-2 in mink and human hosts (Porter et al., 2023).

For each group of tips defining a local clock, we independently conduct a RTT regression to estimate the evolutionary rate (slope). R^2 values for each clock are then an indication of clock-like behaviour for each local clock. Clockor2 focuses on R^2 as an indication of appropriateness of a strict clock instead of other regression summary statistics, such as root mean squared error, because it offers the most straightforward interpretation of clock-like evolution. R^2 values of one indicate perfect clock like evolution, while values of 0 indicate a lack of a molecular clock.

Local clock and or global clock configurations can then be compared using an information criterion that combines the likelihood of each local clock’s RTT regression while penalising the number of inferred parameters (three for each clock - slope, intercept, and variance). Clockor2 allows users to use either the Bayesian Information Criterion (BIC), Aikake Information Criterion (AIC), or corrected Aikake Information Criterion (AICc). We recommend using the BIC because it most heavily penalises the addition of extra parameters, and local clocks in turn.

Derivations of the above information criteria for the local clock model are given in the supplementary methods. Briefly, these exploit the assumption of independent sampling to factor the likelihood across local clocks. Note however that the assumption of independent sampling is flawed because samples necessarily share some ancestry by the assumption of a phylogenetic tree. In other words, ancestral branches are counted over many times in the calculating the distance from root to tip for each sample. However, this is a limitation of the RTT regression approach more broadly, rather than Clockor2 itself.

Algorithm for local clock search

Where it is hypothesised that a dataset contains local clocks, Clockor2 provides functionality to corroborate this hypothesis by performing a search for local clocks in the tree. Briefly, the algorithm takes a maximum number of clocks and a minimum number of tips (group size) for each local clock as

input parameters. It then iterates through all combinations of internal nodes from which local clocks can descend to induce corresponding local clock configurations. Importantly, the clock search algorithm tests for a number of clocks up to and including the maximum number so that more parsimonious configurations with fewer clocks may be found. Configurations are compared using the information criteria outlined above. Again, we recommend the BIC as it penalises additional parameters (ie. additional local clocks) most heavily. See [here for an animation](#) of the clock search algorithm.

We stress that this algorithm is intended to corroborate hypotheses about a particular local clock configuration, but is not intended to be performed as a blind search for local clocks. This is because it is highly prone to over-fitting where the maximum number of clocks is inflated, as outlined later in the results section.

The clock-search algorithm operates in polynomial time (see supplementary material). Efficiency is improved by reducing the maximum number of local clocks in the search, increasing the minimum group size, and contingent on the topology of the underlying tree. However, the former two parameters exert a far greater effect on efficiency than topology.

Clock-search algorithm simulation study

We conducted a simulation study to test the accuracy of the clock-search algorithm. We started with a core set of 100 simulated trees of 250 tips and added a local clock descending from a randomly selected node such that it would contain between 50 and 150 tips. For each, we simulated a 5-fold rate increase occurring in either the stem branch leading to the group/clade, or throughout the group. These scenarios are characterised in Fig 2 C,D respectively. For each of the resulting 200 trees, we applied the clock search algorithm with a minimum group size of 50 tips and a maximum number of of 2-5 clocks. The case of a maximum of 2 clocks tests for baseline accuracy with the search parameters match reality. Searches with a maximum of 3-5 clocks test for over-sensitivity in the algorithm where the maximum number of clocks is inflated. All clock-search tests use the BIC.

Finding the Best Fitting Root

Clockor2 selects the best fitting root based on the R^2 of a global clock model for the input tree. It follows the same algorithm as implemented in Tempest (Rambaut et al., 2016), but makes use of parallelisation to improve speed for larger trees. Briefly, the tree is rooted along the branch leading to each internal node or tip, an RTT regression is performed, and the root position along the branch leading to the highest R^2 value is selected. Clockor2 starts at the midpoint and then optimises the root position using the golden-section search algorithm.

The best fitting root is inferred using a single, global clock because it presents the most parsimonious model of the evolutionary rate for a given tree. The fit of more elaborate local clock models can then be compared to this using information criteria and/or comparing the R^2 values of each model. Clockor2 does not find the best fitting root for local clock models because the search space of best fitting roots and local clock configurations quickly becomes prohibitive and is possibly unidentifiable.

Dependencies

Clockor2 has three key dependencies for handling, and plotting trees and RTT data. Trees are handled and manipulated using the phylojs library [TODO: Cite preprint/NPM repo for phylojs once up]. Phylocanvas is used to visualise trees and plotly.js is used to plot RTT data (Abudahab et al., 2021; ?).

Results

Efficiency

Clockor2 can process trees of up to the order of 10^4 tips, thus fit for the expanding size and diversity of phylodynamic datasets. Finding the best fitting root makes use of parallelisation to increase speed. Speedup is therefore proportional to the number of threads or cores available, in addition to the choice of browser and computer. For example, Clockor2 is faster than TempEst v 1.5.3 on a 2021 MacBook pro with

16Gb of memory and 8 cores running chrome v113.0.5627.126, (Table 1). However, we found Clockor2 to be comparable or slower on other combinations of processor and browser.

	R^2			
Root Mean Squared				
Tips	Clockor2	TempEst	Clockor2	TempEst
100	0.26	0.76	0.05	0.17
500	1.53	2.40	1.5	0.964
1000	5.35	10.28	5.43	2.782
5000	189.02	272.29	122	113.581
10000	422.11	1310.34	951.00	698.065

Table 1: Time taken to find the best fitting root for test trees of 100, 500, 1000, 5000, and 10000 tips using Clockor2 and TempEst v1.5.3 on a 2021 Macbook with 16Gb of memory and 8 cores running chrome v113.0.5627.126. Times vary with computer and browser. In general the relative efficiency of Clockor2 will increase with the number of cores.

The user interface also remains responsive when working with large trees. This is in large part due to the use of WebGL in the tree and plotting components, through Phylocanvas and Plotly.js respectively, which exploits GPU acceleration to render large and interactive trees and datasets (Abudahab et al., 2021; ?).

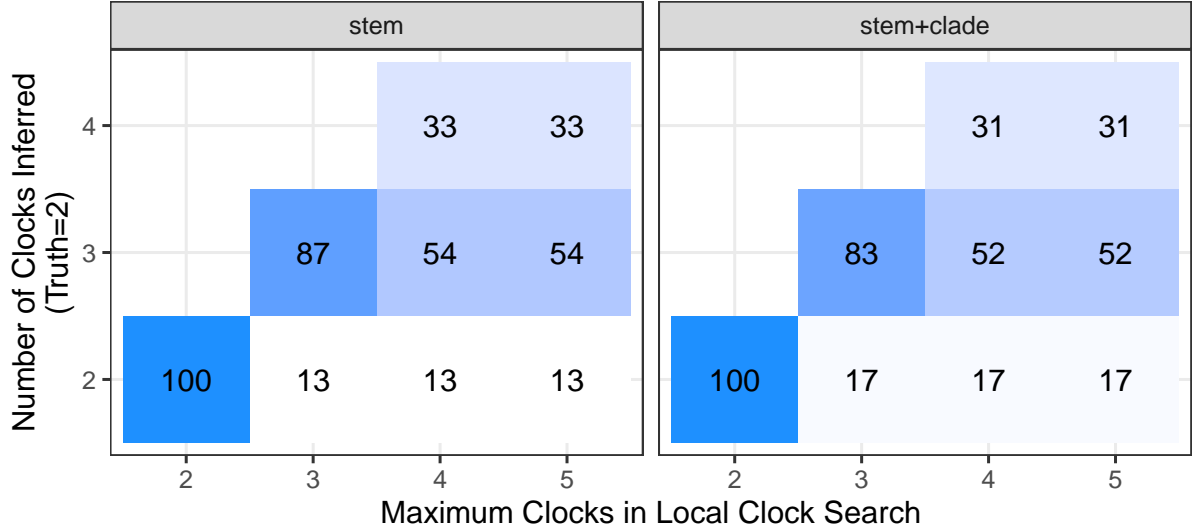


Figure 3: Confusion matrices comparing the number of inferred clocks against the maximum number of clocks allowed by each search, with a true number of 2 clocks in each search. "stem" and "stem+clade" refer to either a rate increase along only the stem of a clade, or with the rate increase continuing in the clade. The true value is 2 and accuracy decreases as the maximum number of clocks allowed by the clock-search algorithm increases. The colour of and number in each tile reflect the number of analyses out of 100 selecting the corresponding number of clocks.

142 For clock searches with a maximum number of 2 clocks, the clock-search algorithm correctly identified
 143 2 local clocks in the simulated data with 100 percent accuracy (Fig. 3). However, when the algorithm
 144 was allowed to search for configurations with 3-5 clocks, only 13 and 17 analyses correctly recovered 2
 145 clocks for the stem only and stem+clade rate increases respectively. Although we used the BIC, the most
 146 conservative information metric used in Clockor2, the clock-search algorithm is clearly still highly prone
 147 to over-fitting. This is because a higher number of clocks allows for tighter clusters of points in the RTT
 148 regression to be found, which are favoured over a lower number of clocks.

To this end, we again emphasise that the clock-search algorithm is only intended to be used as a tool testing a number of clocks up to and including the hypothesised number, but never more. It should never be used to blindly search for local clocks in the absence of a biological hypothesis. The [how-to documents at https://clockor2.github.io/](#) provide example of overfitting using the clock search function when applied to various empirical datasets. **TODO: Specify these once final list consolidated, including Hill et al.**

Discussion

Clockor2 provides a flexible and scalable front-end web based RTT regression platform. Its extension to fitting local clocks allows it to accommodate the growing complexity of phylodynamic datasets as genomic epidemiology plays a growing role in infectious disease surveillance.

As a front end application, Clockor2 is also highly accessible with no installation steps required, although users have option of saving the site to run locally. Wherever there is a browser, it is possible to conduct an RTT regression using Clockor2 with the data never leaving the user's computer.

Future Directions

In the future it will be possible to re-implement core functionality in increasingly popular and highly efficient programming languages, that can compile to Web-Assembly format. For example, as the bioinformatics ecosystem in Rust continues to develop, it will be possible to further improve the efficiency of Clockor2 using packages such as Bio-Rust (Köster, 2015).

Data availability

All code required to replicate the simulation study and figures in the paper is available at <https://github.com/LeoFeatherstone/clockor2Paper>. The code for Clockor2 is open source at <https://>

github.com/clockor2/clockor2.

References

- K. Abudahab, A. Underwood, B. Taylor, C. Yeats, and D. M. Aanensen. PhyloCanvas.gl: A WebGL-powered JavaScript library for large tree visualisation, 2021. URL <https://osf.io/nfv6m/>.
- R. Bouckaert, T. G. Vaughan, J. Barido-Sottani, S. Duchêne, M. Fourment, A. Gavryushkina, J. Heled, G. Jones, D. Kühnert, N. D. Maio, M. Matschiner, F. K. Mendes, N. F. Müller, H. A. Ogilvie, L. d. Plessis, A. Popinga, A. Rambaut, D. Rasmussen, I. Siveroni, M. A. Suchard, C.-H. Wu, D. Xie, C. Zhang, T. Stadler, and A. J. Drummond. BEAST 2.5: An advanced software platform for bayesian evolutionary analysis. *PLOS Computational Biology*, 15(4):e1006650, 2019. ISSN 1553-7358. doi: 10.1371/journal.pcbi.1006650. URL <https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1006650>. Publisher: Public Library of Science.
- L. du Plessis, J. T. McCrone, A. E. Zarebski, V. Hill, C. Ruis, B. Gutierrez, J. Raghwani, J. Ashworth, R. Colquhoun, T. R. Connor, N. R. Faria, B. Jackson, N. J. Loman, A. O’Toole, S. M. Nicholls, K. V. Parag, E. Scher, T. I. Vasylyeva, E. M. Volz, A. Watts, I. I. Bogoch, K. Khan, COVID-19 Genomics UK (COG-UK) Consortium, D. M. Aanensen, M. U. G. Kraemer, A. Rambaut, and O. G. Pybus. Establishment and lineage dynamics of the SARS-CoV-2 epidemic in the UK. *Science*, 371(6530):708–712, 2021. doi: 10.1126/science.abf2946. URL <https://www.science.org/doi/10.1126/science.abf2946>. Publisher: American Association for the Advancement of Science.
- K. Eaton, L. Featherstone, S. Duchene, A. G. Carmichael, N. Varlik, G. B. Golding, E. C. Holmes, and H. N. Poinar. Plagued by a cryptic clock: Insight and issues from the global phylogeny of yersinia pestis. *Communications Biology*, 6(1):23, 2023.
- L. A. Featherstone, J. M. Zhang, T. G. Vaughan, and S. Duchene. Epidemiological inference from

pathogen genomes: A review of phylodynamic models and applications. *Virus Evolution*, 8(1):veac045, 2022.

J. Hadfield, C. Megill, S. M. Bell, J. Huddleston, B. Potter, C. Callender, P. Sagulenko, T. Bedford, and R. A. Neher. Nextstrain: real-time tracking of pathogen evolution. *Bioinformatics*, 34(23):4121–4123, 2018. ISSN 1367-4803. doi: 10.1093/bioinformatics/bty407. URL <https://doi.org/10.1093/bioinformatics/bty407>.

V. Hill, L. Du Plessis, T. P. Peacock, D. Aggarwal, R. Colquhoun, A. M. Carabelli, N. Ellaby, E. Gallagher, N. Groves, B. Jackson, J. T. McCrone, A. O’Toole, A. Price, T. Sanderson, E. Scher, J. Southgate, E. Volz, W. S. Barclay, J. C. Barrett, M. Chand, T. Connor, I. Goodfellow, R. K. Gupta, E. M. Harrison, N. Loman, R. Myers, D. L. Robertson, O. G. Pybus, A. Rambaut, and The COVID-19 Genomics UK (COG-UK) Consortium. The origins and molecular evolution of SARS-CoV-2 lineage b.1.1.7 in the UK. *Virus Evolution*, 8(2):veac080, 2022. ISSN 2057-1577. doi: 10.1093/ve/veac080. URL <https://doi.org/10.1093/ve/veac080>.

S. Y. Ho and S. Duchêne. Molecular-clock methods for estimating evolutionary rates and timescales. *Molecular ecology*, 23(24):5947–5965, 2014.

S. Höhna, M. J. Landis, T. A. Heath, B. Boussau, N. Lartillot, B. R. Moore, J. P. Huelsenbeck, and F. Ronquist. RevBayes: Bayesian Phylogenetic Inference Using Graphical Models and an Interactive Model-Specification Language. *Systematic Biology*, 65(4):726–736, 05 2016. ISSN 1063-5157. doi: 10.1093/sysbio/syw021. URL <https://doi.org/10.1093/sysbio/syw021>.

J. Köster. Rust-Bio: a fast and safe bioinformatics library. *Bioinformatics*, 32(3):444–446, 10 2015. ISSN 1367-4803. doi: 10.1093/bioinformatics/btv573. URL <https://doi.org/10.1093/bioinformatics/btv573>.

S. A. Nadeau, T. G. Vaughan, C. Beckmann, I. Topolsky, C. Chen, E. Hodcroft, T. Schär, I. Nissen,

215 N. Santacroce, E. Burcklen, P. Ferreira, K. P. Jablonski, S. Posada-Céspedes, V. Capece, S. Seidel,
 216 N. Santamaria de Souza, J. M. Martinez-Gomez, P. Cheng, P. P. Bosshard, M. P. Levesque, V. Kufner,
 217 S. Schmutz, M. Zaheri, M. Huber, A. Trkola, S. Cordey, F. Laubscher, A. R. Gonçalves, S. Aeby,
 218 T. Pillonel, D. Jacot, C. Bertelli, G. Greub, K. Leuzinger, M. Stange, A. Mari, T. Roloff, H. Seth-
 219 Smith, H. H. Hirsch, A. Egli, M. Redondo, O. Kobel, C. Noppen, L. du Plessis, N. Beerenwinkel, R. A.
 220 Neher, C. Beisel, and T. Stadler. Swiss public health measures associated with reduced SARS-CoV-
 221 2 transmission using genome data. *Science Translational Medicine*, 15(680):eabn7979, 2023. ISSN
 222 1946-6242. doi: 10.1126/scitranslmed.abn7979.

223 A. F. Porter, D. F. Purcell, B. P. Howden, and S. Duchene. Evolutionary rate of sars-cov-2 increases
 224 during zoonotic infection of farmed mink. *Virus Evolution*, 2023.

225 A. Rambaut, T. T. Lam, L. Max Carvalho, and O. G. Pybus. Exploring the temporal structure of
 226 heterochronous sequences using TempEst (formerly path-o-gen). *Virus Evolution*, 2(1):vew007, 2016.
 227 ISSN 2057-1577. doi: 10.1093/ve/vew007.

228 P. Sagulenko, V. Puller, and R. A. Neher. TreeTime: Maximum-likelihood phylodynamic analysis. *Virus*
 229 *Evolution*, 4(1):vex042, 2018. ISSN 2057-1577. doi: 10.1093/ve/vex042. URL [https://doi.org/10.](https://doi.org/10.1093/ve/vex042)
 230 [1093/ve/vex042](https://doi.org/10.1093/ve/vex042).

231 M. A. Suchard, P. Lemey, G. Baele, D. L. Ayres, A. J. Drummond, and A. Rambaut. Bayesian phy-
 232 logenetic and phylodynamic data integration using BEAST 1.10. *Virus Evolution*, 4(1):vey016, 2018.
 233 ISSN 2057-1577. doi: 10.1093/ve/vey016. URL <https://doi.org/10.1093/ve/vey016>.

234 J. H. Tay, A. F. Porter, W. Wirth, and S. Duchene. The emergence of sars-cov-2 variants of concern is
 235 driven by acceleration of the substitution rate. *Molecular Biology and Evolution*, 39(2):msac013, 2022.

236 E. M. Volz and S. D. W. Frost. Scalable relaxed clock phylogenetic dating. *Virus Evolution*, 3(2):vex025,
 237 2017. ISSN 2057-1577. doi: 10.1093/ve/vex025. URL <https://doi.org/10.1093/ve/vex025>.

238 M. Worobey, G.-Z. Han, and A. Rambaut. A synchronized global sweep of the internal genes of modern
239 avian influenza virus. *Nature*, 508(7495):254–257, 2014. ISSN 1476-4687. doi: 10.1038/nature13016.
240 URL <https://www.nature.com/articles/nature13016>. Number: 7495 Publisher: Nature Publish-
241 ing Group.

242 A. D. Yoder and Z. Yang. Estimation of primate speciation dates using local molecular clocks. *Molecular*
243 *biology and evolution*, 17(7):1081–1090, 2000.