

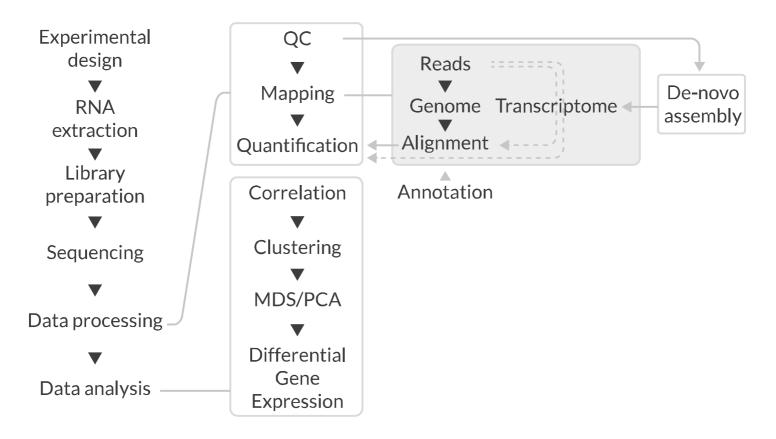
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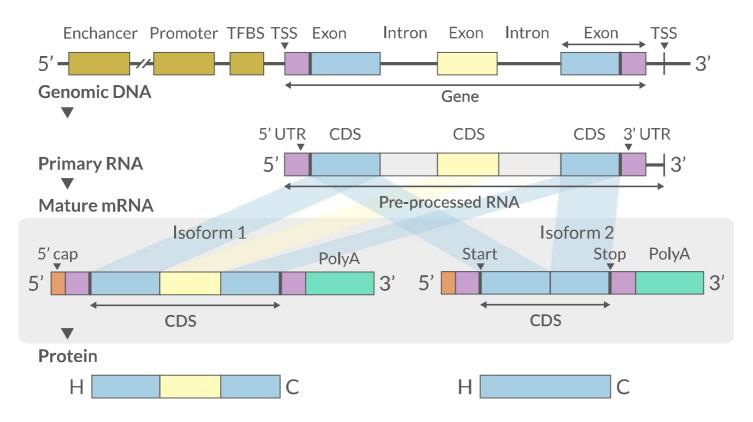
Workflow





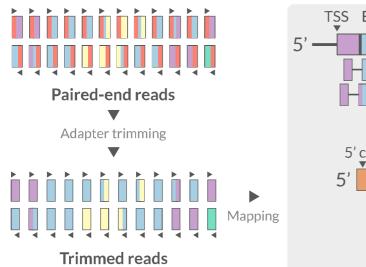
Mapping

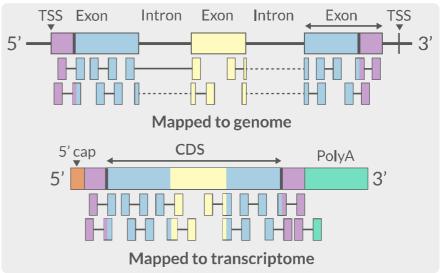




Mapping







- Aligning reads back to a reference sequence
- Mapping to genome vs transcriptome
- Splice-aware alignment (genome)

Aligners



Considerations

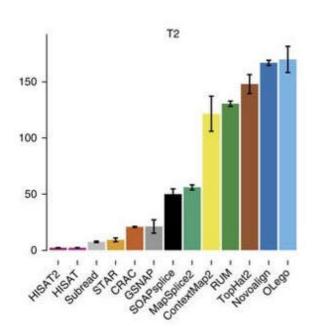
- Speed
- Accuracy
- Resources
- Settings
- Purpose (General/Specific)
- Support & Community

Features

- Reference index
- Read pair alignment
- Consider base quality scores
- Sophisticated indexing to decrease CPU and memory usage
- Resolving multi-mappers
 - Report first X alignments and flag read as multi-mapping
- Use known annotations (junctions)
- 2-pass approach

Aligners | Speed

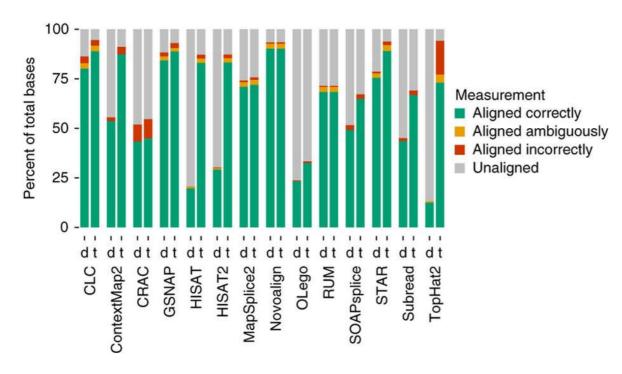




Program	Time_Min	Memory_GB
HISATx1	22.7	4.3
HISATx2	47.7	4.3
HISAT	26.7	4.3
STAR	25	28
STARx2	50.5	28
GSNAP	291.9	20.2
TopHat2	1170	4.3

Aligners | Accuracy





Increasing Accuracy

- Novel variants / RNA editing
- Allele-specific expression
- Genome annotation
- Gene and transcript discovery
- Differential expression

STAR, HiSat2, GSNAP, Novoalign (Commercial)

Mapping



Reads (FASTQ)

```
@ST-E00274:179:HHYMLALXX:8:1101:1641:1309 1:N:0:NGATGT
NCATCGTGGTATTTGCACATCTTTTCTTATCAAATAAAAAGTTTAACCTACTCAGTTATGCGCATACGTTTTTGATGG
+
#AAAFAFA<-AFFJJJAFA-FFJJJJFFFAJJJJ-<FFJJJ-A-F-7--FA7F7-----FFFJFA<FFFFJ<AJ--FF-
```

```
@instrument:runid:flowcellid:lane:tile:xpos:ypos
read:isfiltered:controlnumber:sampleid
```

Reference Genome/Transcriptome (FASTA)

Annotation (GTF/GFF)

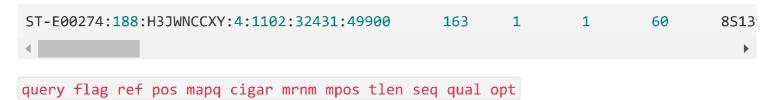
```
#!genome-build GRCz10
#!genebuild-last-updated 2016-11
4 ensembl_havana gene 6732 52059 . - . gene_id "ENS"
```

chr source feature start end score strand frame attribute

Alignment



• SAM/BAM (Sequence Alignment Map format)



Alignment formats



Format	Size_GB				
SAM	7.4				
BAM	1.9				
CRAM lossless Q	1.4				
CRAM 8 bins Q	0.8				
CRAM no Q	0.26				

Visualisation | tview



samtools tview alignment.bam genome.fasta

911 921 931 GTTTAATTTCATCTTCTAATTTAGAAT		CTCTCGAAGTTGGCA											
GTTTAATTTCATCTTCTAATTTAGAAT													agacttaattga
ATTTCATCTTCTAATTTAGAAT					tgcttctgaga								agacttaattga
atttcatcttctaatttagaat													agacttaattga
atttcatcttctaatttagaat													agacttaattga
	cttgccaatcaagcc											TCCTCCATTCA	
GTTTAATTT t	cttgccaatcaagcc	ctctcgaagttggc	aatatctataac	tcaacctc	tgcttctgaga	ttctaag	CTTAGATGC	CAAGTACATTA	CTATAATTGG	TGTTATCGGG	TCTTCCAA	TCCTCCATTCA	AGACTTAA
GTTTAATTTCATCTT	cttgccaatcaagcc	ctctcgaagttggca	aatatctataac	tcaacctc	tgcttctgaga	ttctaagt	TTAGATGC	CAAGTACATTA	CTATAATTGG	TGTTATCGGG	TCTTCCAA	TCCTCCATTCA	AGACTTAAT
GTTTAATTTCATCTTC	TGCCAATCAAGCC	CTCTCGAAGTTGGCA	AATATCTATAAC	TCAACCTC	TGCTTCTGAGA	TTCTAAGTAC	ATGC	CAAGTACATTA	CTATAATTGG	TGTTATCGGG	TCTTCCAA	TCCTCCATTCA	AGACTTAATTGA
GTTTAATTTCATCTTCTAAT	TGCCAATCAAGCC	CTCTCGAAGTTGGCA	AATATCTATAA	TCAACCTC	TGCTTCTGAGA	TTCTAAGTAC	GCC	CAAGTACATTA	CTATAATTGG	TGTTATCGGG	TCTTCCAA	TCCTCCATTCA	AGACTTAATTGA
gtttaatttcatcttctaatttag	TGCCAATCAAGCC	CTCTCGAAGTTGGCA	AATATCTATAAC	TCAACCTC	TGCTTCTGAGA	TTCTAAGTAC		CATTA	CTATAATTGG	TGTTATCGGG	TCTTCCAAC	TCCTCCATTCA	AGACTTAATTGA
GTTTAATTTCATCTTCTAATTTAG		TTCTCGAAGTTGGC						catta	ctataattgg	tgttatcggg	tcttccaa	tcctccattca	agacttaattga
STTTAATTTCATCTTCTAATTTAG	CAATCAAGCC	CTCTCGAAGTTGGCA	AATATCTATAA	TCAACCTC	TGCTTCTGAGA	TTCTAAGTAC	C			tgttatcggg	tcttccaa	tcctccattca	agacttaattga
GTTTAATTTCATCTTCTAATTTAG	CAATCAAGCC	CTCTCGAAGTTGGCA	AATATCTATAA(TCAACCTC	TGCTTCTGAGA	TTCTAAGTAC	CTT						agacttaattga
GTTTAATTTCATCTTCTAATTTAG		ctctcgaagttggca											AGACTTAATTGA
STTTAATTTCATCTTCTAATTTAGAAT		CTCTCGAAGTTGGCA											agacttaattga
GTTTAATTTCATCTTCTAATTTAGAAT		ctctcgaagttggca											agacttaattga
STTTAATTTCATCTTCTAATTTAGAAT		CTCGAAGTTGGCA											AGACTTAATTGA
STTTAATTTCATCTTCTAATTTAGAAT			AATATCTATAA							g			agacttaattga
STTTAATTTCATCTTCTAATTTAGAAT			AATATCTATAA										agacttaattga
gtttaatttcatcttctaatttagaat		C	AATATCTATAA(agacttaattga
STTTAATTTCATCTTCTAATTTAGAAT			CTATAAC	TCAACCTC	TGCTTCTGAGA								AGACTTAATTGA
STITAATTTCATCTTCTAATTTAGAAT								caagtacatta					AGACTTAATTGA
gtttaatttcatcttctaatttagaat								caagtacatta					agacttaattga
STITAATTTCATCTTCTAATTTAGAAT								caagtacatta					agacttaattga
ytttaatttcatcttctaatttagaat								caagtacatta					agacttaattga
GTTTAATTTCATCTTCTAATTTAGAAT													agacttaattga
GTTTAATTTCATCTTCTAATTTAGAAT													agacttaattga
TTTAATTTCATCTTCTAATTTAGAAT								caagtacatta					agacttaattga
TTTAATTTCATCTTCTAATTTAGAAT								aagtacatta					AGACTTAATTGA
ATTTCATCTTCTAATTTAGAAT								aagtacatta					cttaattga
TTCATCTTCTAATTTAGAAT	CTTGCCAATCAAGCC	CTCTCGAAGTTGGCA	AATATCTATAA(TCAACCT				CAAGTACATTA					attga
								aagtacatta					
								aagtacatta					
					ga			aagtacatta					
						aagtac	cttagatgc	caagtacatta	ctataattgg	tgttatcggg		tcctccattca	
													agacttaattga AGACTTAATTGA
													AGACTTAATTGA
													agacttaattga
													agacttaattga agacttaattga
													agacttaattga agacttaattga
											dat		agacttaattga
													agacttaattga
												CCOLLCO	aguerraarrya
												ccattca	agacttaattga

Visualisation | IGV

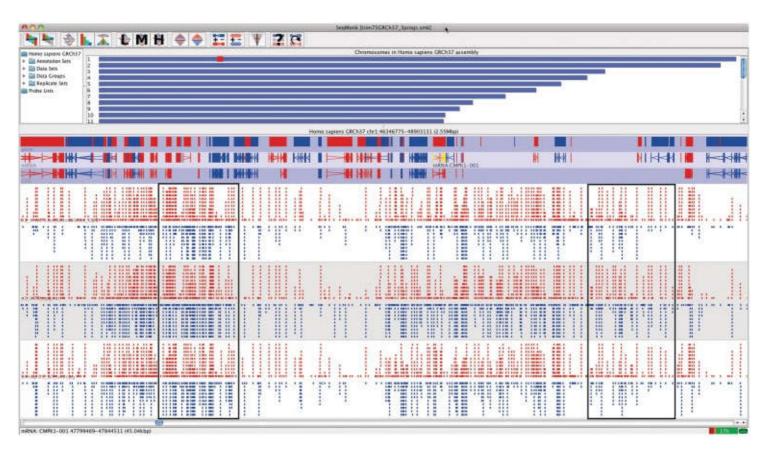




▲ IGV, UCSC Genome Browser

Visualisation | SeqMonk



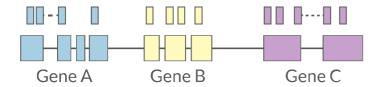




Quantification | Counts



- Read counts = gene expression
- Reads can be quantified on any feature (gene, transcript, exon etc)
- Intersection on gene models
- Gene/Transcript level

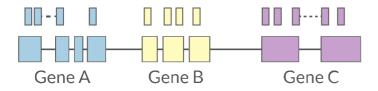


featureCounts, HTSeq

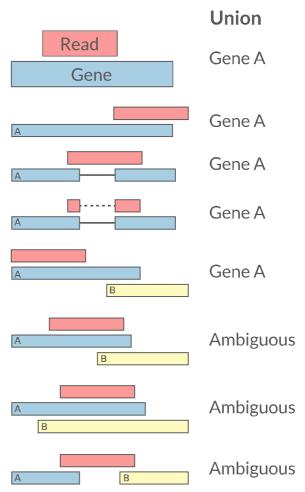
Quantification | Counts



- Read counts = gene expression
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featureCounts, HTSeq



Quantification



PCR duplicates

- Ignore for RNA-Seq data
- Computational deduplication (Don't!)
- Use PCR-free library-prep kits
- Use UMIs during library-prep

Multi-mapping

- Added (BEDTools multicov)
- Discard (featureCounts, HTSeq)
- Distribute counts (Cufflinks)
- Rescue
 - Probabilistic assignment (Rcount, Cufflinks)
 - Prioritise features (Rcount)
 - Probabilistic assignment with EM (RSEM)

Parekh, Swati, et al. "The impact of amplification on differential expression analyses by RNA-seq." Scientific reports 6 (2016): 25533

Quantification | Abundance



- Count methods
 - Provide no inference on isoforms
 - Cannot accurately measure fold change
- Probabilistic assignment
 - Deconvolute ambiguous mappings
 - Transcript-level
 - cDNA reference

Kallisto, Salmon

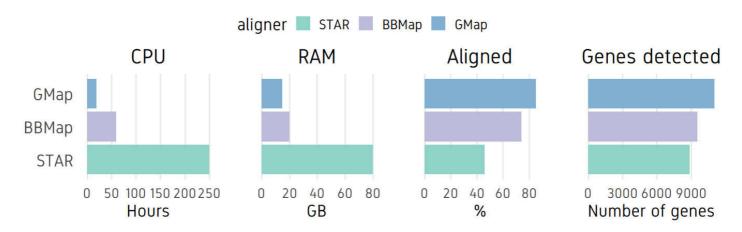
- Direct from FastQ to counts
- Ultra-fast & alignment-free
- Uses transcriptome reference
- Subsampling & quantification confidence
- Transcript-level estimates improves gene-level estimates
- Kallisto/Salmon > transcript-counts > tximport() > gene-counts

RSEM, Kallisto, Salmon, Cufflinks2

Long-Read RNA-Seq



- PacBio, Nanopore etc
- Long reads, full transcripts
- High error rate
- Expensive



Results are comparable with MinION data.

♣ GMAP, BBMap, STAR

Summary



- STAR, HISAT2 and GSNAP are good general purpose aligners
- Use HISAT2 if RAM is limited
- Consider using 2-pass mapping
- Be stringent with junction discovery criteria
- Map to genome for annotation/discovery
- For well known transcriptomes, Kallisto/Salmon offers ultra-fast quantification
- For long reads, GMAP and BBMap are good choice of aligners

