main female

August 6, 2021

1 Tissue comparison for differential expression analysis

```
[1]: import functools
     import numpy as np
     import pandas as pd
     from gtfparse import read_gtf
[2]: config = {
         'caudate': '../../caudate/female_analysis/metrics_summary/_m/
      →female_specific_DE_4features.txt',
         'dlpfc': '../../dlpfc/female_analysis/metrics_summary/_m/
     →female_specific_DE_4features.txt',
         'hippo': '../../hippocampus/female analysis/metrics summary/ m/
      →female_specific_DE_4features.txt',
         'cmc_dlpfc': '../../cmc_dlpfc/female_analysis/metrics_summary/_m/

¬female_specific_DE_genes.txt'

[3]: Ofunctools.lru_cache()
     def get_deg(filename):
         dft = pd.read_csv(filename, sep='\t', index_col=0)
         dft = dft[(dft['Type'] == 'gene')].copy()
         dft['Feature'] = dft.index
         dft['Dir'] = np.sign(dft['t'])
         if 'gene id' in dft.columns:
             dft['ensemblID'] = dft.gene_id.str.replace('\\..*', '', regex=True)
         return dft[['Feature', 'ensemblID', 'adj.P.Val', 'logFC', 't', 'Dir']]
     @functools.lru_cache()
     def get_deg_sig(filename):
         dft = get_deg(filename)
         return dft[(dft['adj.P.Val'] < 0.05)]</pre>
     @functools.lru_cache()
     def merge dataframes(tissue1, tissue2):
         return get_deg(config[tissue1]).merge(get_deg(config[tissue2]),
```

```
on='Feature',
                                               suffixes=['_%s' % tissue1, '_%s' %_
      →tissue2])
     @functools.lru cache()
     def merge_dataframes_sig(tissue1, tissue2):
         return get_deg_sig(config[tissue1]).merge(get_deg_sig(config[tissue2]),
                                                   on='Feature',
                                                   suffixes=['_%s' % tissue1, '_%s'_
      →% tissue2])
[4]: def tissue_annotation(tissue):
         return {'dlpfc': 'DLPFC', 'hippo': 'Hippocampus',
                 'caudate': 'Caudate', 'cmc_dlpfc': 'CMC DLPFC'}[tissue]
     def save_plot(p, fn, width=7, height=7):
         '''Save plot as sug, png, and pdf with specific label and dimension.'''
         for ext in ['.svg', '.png', '.pdf']:
             p.save(fn+ext, width=width, height=height)
    1.1 BrainSeq Comparison
[5]: caudate = get deg(config['caudate'])
     caudate.groupby('Dir').size()
[5]: Dir
     -1.0
             68
     1.0
             56
     dtype: int64
[6]: caudate[(caudate['adj.P.Val'] < 0.05)].shape
    INFO:numexpr.utils:Note: NumExpr detected 64 cores but "NUMEXPR_MAX_THREADS" not
    set, so enforcing safe limit of 8.
    INFO:numexpr.utils:NumExpr defaulting to 8 threads.
[6]: (124, 6)
[7]: dlpfc = get_deg(config['dlpfc'])
     dlpfc.groupby('Dir').size()
[7]: Series([], dtype: int64)
[8]: dlpfc[(dlpfc['adj.P.Val'] < 0.05)].shape
[8]: (0, 6)
```

```
[9]: hippo = get_deg(config['hippo'])
      hippo.groupby('Dir').size()
 [9]: Series([], dtype: int64)
[10]: hippo[(hippo['adj.P.Val'] < 0.05)].shape
[10]: (0, 6)
     1.1.1 Upset Plot
[11]: phase2_dlpfc = dlpfc[(dlpfc['adj.P.Val'] < 0.05)].copy()</pre>
      phase2_dlpfc['DLPFC'] = 1
      phase2_dlpfc = phase2_dlpfc[['ensemblID', 'DLPFC']]
      phase2_hippo = hippo[(hippo['adj.P.Val'] < 0.05)].copy()</pre>
      phase2_hippo['Hippocampus'] = 1
      phase2_hippo = phase2_hippo[['ensemblID', 'Hippocampus']]
      phase3_caudate = caudate[(caudate['adj.P.Val'] < 0.05)].copy()</pre>
      phase3 caudate['Caudate'] = 1
      phase3_caudate = phase3_caudate[['ensemblID', 'Caudate']]
[12]: geneList = pd.merge(phase3_caudate[['ensemblID']], phase2_dlpfc[['ensemblID']],
                          on=['ensemblID'], how='outer')\
                   .merge(phase2 hippo[['ensemblID']], on=['ensemblID'], how='outer')\
                   .groupby(['ensemblID']).first().reset_index()
      newC = pd.merge(geneList, phase3_caudate, on=['ensemblID'], how='outer').
      →fillna(0)
      newC['Caudate'] = newC['Caudate'].astype('int')
      newD1 = pd.merge(geneList, phase2_dlpfc, on=['ensemblID'], how='outer').
      →fillna(0)
      newD1['DLPFC'] = newD1['DLPFC'].astype('int')
      newH = pd.merge(geneList, phase2_hippo, on=['ensemblID'], how='outer').fillna(0)
      newH['Hippocampus'] = newH['Hippocampus'].astype('int')
      print(newC.shape, newH.shape, newD1.shape)
     (124, 2) (124, 2) (124, 2)
[13]: df = pd.concat([newC.set_index(['ensemblID']), newD1.set_index(['ensemblID']),
                      newH.set_index(['ensemblID'])], axis=1, join='outer')
      df.head(2)
```

```
Γ13]:
                      Caudate DLPFC Hippocampus
     ensemblID
     ENSG00000000457
                                   0
                                                0
                            1
     ENSG00000003137
                            1
                                   0
                                                \cap
[14]: %load_ext rpy2.ipython
[15]: \%\R
     library(ComplexHeatmap)
     library(tidyverse)
     subset pvalue <- function(filename, fdr cutoff){</pre>
          df <- data.table::fread(filename) %>%
             filter(Type == 'gene', adj.P.Val < fdr_cutoff)
         return(df$ensemblID)
     }
     caudate = subset_pvalue('.../.../caudate/female_analysis/metrics_summary/_m/
      →female_specific_DE_4features.txt',
                             0.05)
     dlpfc = subset_pvalue('../../dlpfc/female analysis/metrics summary/_m/
      →female_specific_DE_4features.txt',
                           0.05)
     hippo = subset_pvalue('../../hippocampus/female_analysis/metrics_summary/_m/
      →female_specific_DE_4features.txt',
                           0.05)
     lt = list(Caudate = caudate,
               DLPFC = dlpfc,
               Hippocampus = hippo)
     m = make_comb_mat(lt)
     cbb_palette <- c("#000000", "#E69F00", "#56B4E9", "#009E73", "#F0E442",
                       "#0072B2", "#D55E00", "#CC79A7")
     WARNING:rpy2.rinterface_lib.callbacks:R[write to console]: Loading required
     package: grid
     WARNING:rpy2.rinterface lib.callbacks:R[write to console]:
     _____
     ComplexHeatmap version 2.6.2
     Bioconductor page: http://bioconductor.org/packages/ComplexHeatmap/
     Github page: https://github.com/jokergoo/ComplexHeatmap
     Documentation: http://jokergoo.github.io/ComplexHeatmap-reference
     If you use it in published research, please cite:
     Gu, Z. Complex heatmaps reveal patterns and correlations in multidimensional
       genomic data. Bioinformatics 2016.
```

```
This message can be suppressed by:
       suppressPackageStartupMessages(library(ComplexHeatmap))
     WARNING:rpy2.rinterface_lib.callbacks:R[write to console]: Attaching packages
                          tidyverse 1.3.1
     WARNING:rpy2.rinterface_lib.callbacks:R[write to console]: ggplot2 3.3.5
     purrr
           0.3.4
      tibble 3.1.2
                         dplyr 1.0.7
      tidyr 1.1.3
                         stringr 1.4.0
                         forcats 0.5.1
              1.4.0
      readr
     WARNING:rpy2.rinterface_lib.callbacks:R[write to console]:
                                                                Conflicts
                            tidyverse_conflicts()
      dplyr::filter() masks stats::filter()
      dplyr::lag()
                     masks stats::lag()
[16]: \%\R
     right_annot = upset_right_annotation(
         m, ylim = c(0, 150),
         gp = gpar(fill = "black"),
         annotation_name_side = "top",
         axis param = list(side = "top"))
     top_annot = upset_top_annotation(
         m, height=unit(7, "cm"),
         ylim = c(0, 150),
         gp=gpar(fill=cbb_palette[comb_degree(m)]),
         annotation_name_rot = 90)
     pdf('BrainSeq_sex_tissue_upsetR_DEgenes_femaleSpecific.pdf', width=6, height=4)
     ht = draw(UpSet(m, pt_size=unit(4, "mm"), lwd=3,
                     comb_col=cbb_palette[comb_degree(m)],
                     set_order = c("Caudate", "DLPFC", "Hippocampus"),
                     comb_order = order(-comb_size(m)),
                     row_names_gp = gpar(fontsize = 14, fontface='bold'),
                     right_annotation = right_annot,
                     top_annotation = top_annot))
     od = column order(ht)
     cs = comb_size(m)
     decorate_annotation("intersection_size", {
         grid.text(cs[od], x = seq_along(cs), y = unit(cs[od], "native") +
                   unit(6, "pt"),
             default.units = "native", just = "bottom", gp = gpar(fontsize = 11))
```

```
})
      dev.off()
     png
[17]: %%R
      right_ha = rowAnnotation(
          "Intersection\nsize" = anno_barplot(comb_size(m), border=F,
                                              ylim = c(0, 150),
      →gp=gpar(fill=cbb_palette[comb_degree(m)]),
                                              width = unit(7, "cm")))
      top_ha = HeatmapAnnotation(
          "Set size" = anno_barplot(set_size(m), border=F,
                                    ylim = c(0, 150),
                                    gp = gpar(fill = "black"),
                                    height = unit(2, "cm")),
          gap = unit(2, "mm"), annotation_name_side = "left",
          annotation name rot = 90)
      pdf("BrainSeq_sex_tissue_upsetR_DEgenes_transpose_femaleSpecific.pdf", width=5,_
      →height=10)
      ht = draw(UpSet(t(m), pt_size=unit(5, "mm"), lwd=3,
                      comb_order = order(-comb_size(m)),
                      comb_col=cbb_palette[comb_degree(m)],
                      set_order = c("Caudate", "DLPFC", "Hippocampus"),
                      column_names_gp = gpar(fontsize = 16, fontface='bold'),
                      right_annotation = right_ha, top_annotation=top_ha))
      od = rev(row_order(ht))
      cs = comb size(m)
      decorate_annotation("Intersection\nsize", {
          grid.text(cs[od], y = seq_along(cs), x = unit(cs[od], "native") +
                    unit(6, "pt"),
              default.units = "native", just = "left", gp = gpar(fontsize = 11))
      })
      dev.off()
```

png

1.1.2 Shared features

```
[18]: Ofunctools.lru cache()
     def get_gtf(gtf_file):
         return read_gtf(gtf_file)
[19]: def gene_annotation(gtf_file, feature):
         gtf0 = get_gtf(gtf_file)
         gtf = gtf0[gtf0["feature"] == feature]
         return gtf[["gene_id", "gene_name", "transcript_id", "exon_id", u
      gtf_file = '/ceph/genome/human/gencode25/gtf.CHR/_m/gencode.v25.annotation.gtf'
     gtf_annot = gene_annotation(gtf_file, 'gene')
     gtf_annot.head(2)
     INFO:root:Extracted GTF attributes: ['gene_id', 'gene_type', 'gene_status',
     'gene_name', 'level', 'havana_gene', 'transcript_id', 'transcript_type',
     'transcript_status', 'transcript_name', 'transcript_support_level', 'tag',
     'havana_transcript', 'exon_number', 'exon_id', 'ont', 'protein_id', 'ccdsid']
[19]:
                   gene_id gene_name transcript_id exon_id \
         ENSG00000223972.5
                             DDX11L1
     12 ENSG00000227232.5
                             WASH7P
                                 gene_type seqname start
                                                             end strand
                                              chr1 11869
     0
         transcribed_unprocessed_pseudogene
                                                           14409
     12
                     unprocessed_pseudogene
                                              chr1 14404
                                                           29570
[20]: dft = caudate.merge(gtf_annot[['gene_id', 'gene_name', 'seqname']],
                         left_index=True, right_on='gene_id')
     dft.head(2)
[20]:
                         Feature
                                       ensemblID adj.P.Val
                                                               logFC
                                                                                \
     1468999 ENSG00000111181.12 ENSG00000111181
                                                   0.001106 -0.456838 -4.917349
     1928121
               ENSG00000070915.9 ENSG00000070915
                                                   0.001680 0.848495 4.756160
              Dir
                             gene_id gene_name seqname
     1468999 -1.0 ENSG00000111181.12
                                       SLC6A12
                                                 chr12
     1928121 1.0
                    ENSG00000070915.9
                                       SLC12A3
                                                 chr16
     1.2 Comparison with CommonMind
[21]: cmc_dlpfc = get_deg(config['cmc_dlpfc'])
     cmc_dlpfc.groupby('Dir').size()
[21]: Dir
     -1.0
             227
```

```
1.0
             356
      dtype: int64
[22]: cmc_dlpfc[(cmc_dlpfc['adj.P.Val'] < 0.05)].shape
[22]: (583, 6)
     1.2.1 Upset Plot
[23]: cmc = cmc_dlpfc[(cmc_dlpfc['adj.P.Val'] < 0.05)].copy()
      cmc['CMC DLPFC'] = 1
      cmc = cmc[['ensemblID', 'CMC DLPFC']].groupby('ensemblID').first().reset_index()
[24]: geneList = pd.merge(phase3_caudate[['ensemblID']], phase2_dlpfc[['ensemblID']],
      .merge(phase2 hippo[['ensemblID']], on=['ensemblID'], how='outer')\
                   .merge(cmc[['ensemblID']], on=['ensemblID'], how='outer')\
                   .groupby(['ensemblID']).first().reset_index()
      newC = pd.merge(geneList, phase3 caudate, on=['ensemblID'], how='outer').
      \rightarrowfillna(0)
      newC['Caudate'] = newC['Caudate'].astype('int')
      newD1 = pd.merge(geneList, phase2_dlpfc, on=['ensemblID'], how='outer').
      \rightarrowfillna(0)
      newD1['DLPFC'] = newD1['DLPFC'].astype('int')
      newH = pd.merge(geneList, phase2_hippo, on=['ensemblID'], how='outer').fillna(0)
      newH['Hippocampus'] = newH['Hippocampus'].astype('int')
      newCMC = pd.merge(geneList, cmc, on=['ensemblID'], how='outer').fillna(0)
      newCMC['CMC DLPFC'] = newCMC['CMC DLPFC'].astype('int')
      print(newC.shape, newH.shape, newD1.shape, newCMC.shape)
     (702, 2) (702, 2) (702, 2) (702, 2)
[25]: | df = pd.concat([newC.set_index(['ensemblID']), newD1.set_index(['ensemblID']),
                     newH.set_index(['ensemblID']), newCMC.set_index(['ensemblID'])],
                     axis=1, join='outer')
      df.head(2)
[25]:
                      Caudate DLPFC Hippocampus CMC DLPFC
      ensemblID
      ENSG00000000457
                                                            0
                             1
                                    0
      ENSG00000003137
                             1
                                    0
                                                            0
```

```
[26]: \%\R
      cmc = subset_pvalue('../../cmc_dlpfc/female_analysis/metrics_summary/_m/
      →female_specific_DE_genes.txt',
                          0.05)
      lt = list(Caudate = caudate,
                DLPFC = dlpfc,
                Hippocampus = hippo,
                `CMC DLPFC` = cmc)
      m = make_comb_mat(lt)
[27]: %%R
      right_annot = upset_right_annotation(
          m, ylim = c(0, 800),
          gp = gpar(fill = "black"),
          annotation_name_side = "bottom",
          axis_param = list(side = "bottom"))
      top_annot = upset_top_annotation(
          m, height=unit(7, "cm"),
          ylim = c(0, 800),
          gp=gpar(fill=cbb_palette[comb_degree(m)]),
          annotation_name_rot = 90)
      pdf('cmc_sex_tissue_upsetR_DEgenes_femaleSpecific.pdf', width=8, height=5)
      ht = draw(UpSet(m, pt_size=unit(6, "mm"), lwd=3,
                      comb_col=cbb_palette[comb_degree(m)],
                      set_order = c("Caudate", "DLPFC", "Hippocampus", "CMC DLPFC"),
                      comb_order = order(-comb_size(m)),
                      row_names_gp = gpar(fontsize = 16, fontface='bold'),
                      right_annotation = right_annot,
                      top_annotation = top_annot))
      od = column_order(ht)
      cs = comb size(m)
      decorate_annotation("intersection_size", {
          grid.text(cs[od], x = seq_along(cs), y = unit(cs[od], "native") +
                    unit(6, "pt"),
              default.units = "native", just = "bottom", gp = gpar(fontsize = 11))
      })
      dev.off()
     png
[28]: %%R
      right_ha = rowAnnotation(
```

```
"Intersection\nsize" = anno_barplot(comb_size(m), border=F,
                                              ylim = c(0, 800),
      →gp=gpar(fill=cbb_palette[comb_degree(m)]),
                                              width = unit(7, "cm")))
      top ha = HeatmapAnnotation(
          "Set size" = anno_barplot(set_size(m), border=F,
                                    ylim = c(0, 800),
                                    gp = gpar(fill = "black"),
                                    height = unit(2, "cm")),
         gap = unit(2, "mm"), annotation_name_side = "left",
         annotation_name_rot = 90)
      pdf("cmc_sex_tissue_upsetR_DEgenes_transpose_femaleSpecific.pdf", width=5,__
       →height=10)
      ht = draw(UpSet(t(m), pt_size=unit(5, "mm"), lwd=3,
                      comb_order = order(-comb_size(m)),
                      comb_col=cbb_palette[comb_degree(m)],
                      set_order = c("Caudate", "DLPFC", "Hippocampus", "CMC DLPFC"),
                      column_names_gp = gpar(fontsize = 16, fontface='bold'),
                      right_annotation = right_ha, top_annotation=top_ha))
      od = rev(row_order(ht))
      cs = comb_size(m)
      decorate_annotation("Intersection\nsize", {
         grid.text(cs[od], y = seq_along(cs), x = unit(cs[od], "native") +
                    unit(6, "pt"),
              default.units = "native", just = "left", gp = gpar(fontsize = 11))
      })
      dev.off()
     png
       2
[29]: dft = pd.read_csv('../../cmc_dlpfc/female_analysis/metrics_summary/_m/
      →female_specific_DE_genes.txt',
                        sep='\t')
      dft['Dir'] = np.sign(dft['t'])
      dft.head()
[29]:
                                                    Symbol
                    Feature
                                      gencodeID
                                                                  ensemblID Chrom
      0 ENSG00000153132.12 ENSG00000153132.12
                                                      CLGN ENSG00000153132
                                                                              chr4
      1
         ENSG00000179083.6
                             ENSG00000179083.6
                                                   FAM133A ENSG00000179083
                                                                              chrX
        ENSG00000165733.7
                             ENSG00000165733.7
      2
                                                      BMS1 ENSG00000165733 chr10
      3 ENSG00000183023.18 ENSG00000183023.18
                                                    SLC8A1 ENSG00000183023
                                                                              chr2
         ENSG00000236268.5
                             ENSG00000236268.5 LINC01361 ENSG00000236268
                                                                              chr1
```

```
logFC
                         t adj.P.Val Male_Pval Male_FDR
                                                           Type
                                                                 Dir
     0 0.389937 5.559139
                             0.000123
                                        0.283020
                                                  0.313711
                                                                 1.0
                                                           gene
     1 0.261268 5.004488
                             0.000535
                                        0.272019
                                                 0.302973
                                                           gene
                                                                 1.0
     2 0.150918 4.986552
                             0.000535
                                        0.169787
                                                  0.205428
                                                           gene
                                                                 1.0
     3 0.245819 4.925477
                             0.000632
                                        0.082759 0.108963
                                                           gene
                                                                 1.0
     4 0.404532 4.865744
                             0.000700
                                        0.110632 0.140404
                                                           gene
                                                                 1.0
[30]: dft.loc[:, ['Feature', 'ensemblID', 'Symbol', 'Chrom', 'Dir']]\
          .merge(pd.DataFrame({'ensemblID': list(set(phase3_caudate['ensemblID']) &
                                                set(cmc['ensemblID']))}),
                on='ensemblID')
[30]:
                   Feature
                                  ensemblID
                                               Symbol Chrom Dir
     0 ENSG00000100814.17 ENSG00000100814 CCNB1IP1
                                                      chr14 1.0
         ENSG00000263006.6 ENSG00000263006
                                              ROCK1P1 chr18 1.0
     1
     2
         ENSG00000249669.9 ENSG00000249669
                                                CARMN
                                                      chr5 -1.0
         ENSG00000269937.1 ENSG00000269937
                                                  NaN chr16 -1.0
     4 ENSG00000167703.14 ENSG00000167703
                                              SLC43A2 chr17 -1.0
[31]: shared_df = dft.loc[:, ['Feature', 'ensemblID', 'Chrom', 'Symbol', 'Dir']]\
                    .merge(pd.DataFrame({'ensemblID':___
      →list(set(phase3_caudate['ensemblID']) &
                                                          set(cmc['ensemblID']))}),
                           on='ensemblID')
      shared_df.to_csv('cmc_shared_caudate_degs_annotation_femaleSpecific.txt', __
      ⇔sep='\t',
                      index=False, header=True)
     shared_df
[31]:
                   Feature
                                  ensemblID Chrom
                                                     Symbol Dir
     0 ENSG00000100814.17 ENSG00000100814 chr14 CCNB1IP1 1.0
     1
         ENSG00000263006.6 ENSG00000263006 chr18
                                                    ROCK1P1 1.0
     2
         ENSG00000249669.9 ENSG00000249669
                                                      CARMN -1.0
                                              chr5
         ENSG00000269937.1 ENSG00000269937 chr16
                                                        NaN -1.0
     4 ENSG00000167703.14 ENSG00000167703 chr17
                                                     SLC43A2 -1.0
[32]: #### 6 out of 41 are autosomal
     dd = np.sum(shared_df.Chrom.isin(['chrX', 'chrY'])) / shared_df.shape[0] * 100
     print("%0.2f%% of shared DEG are allosomal!" % dd)
     0.00% of shared DEG are allosomal!
[33]: gtf_annot['ensemblID'] = gtf_annot.gene_id.str.replace("\\..*", "", regex=True)
     gtf_annot[["gene_id", 'ensemblID', 'gene_name', 'seqname', 'gene_type']]\
          .merge(df, left_on='ensemblID', right_index=True)\
          .to_csv('cmc_all_deg_across_tissues_femaleSpecific.csv')
 []:
```