

# main

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## 1 Plotting eQTLs, increase font sizes

### 1.0.1 Kynon Jade Benjamin and Apuã Paquola

```
[1]: import re
import functools
import subprocess
import numpy as np
import pandas as pd
from plotnine import *
from pandas_plink import read_plink
from warnings import filterwarnings
from matplotlib.cbook import mplDeprecation

filterwarnings("ignore",category=mplDeprecation)
filterwarnings('ignore', category=UserWarning, module='plotnine.*')
filterwarnings('ignore', category=DeprecationWarning, module='plotnine.*')
```

### 1.1 Configuration

```
[2]: tissue = "caudate"; feature = "genes"
config = {
    'biomart_file': '../_h/biomart.csv',
    'residual_expression_file': "../../../../../prep_eqtl_analysis/%s/%s/
↳covariates/" % (tissue, feature)+\
    "residualized_expression/_m/%s_residualized_expression.csv" % feature,
    'phenotype_file': '/ceph/projects/v4_phase3_paper/inputs/phenotypes/_m/
↳merged_phenotypes.csv',
    'plink_file_prefix': '/ceph/projects/v4_phase3_paper/inputs/genotypes/_m/
↳LIBD_Brain_TopMed',
    'eqtl_output_file': '../../../../../summary_table/_m/
↳Brainseq_sex_interacting_4features_3regions.eFeatures.txt.gz',
    'gwas_snp_file': '/ceph/projects/v4_phase3_paper/inputs/sz_gwas/pgc2_clozuk/
↳map_phase3/_m/libd_hg38_pgc2sz_snps_p5e_minus8.tsv'
}
```

## 1.2 Functions

### 1.2.1 Expression functions

```
[3]: @functools.lru_cache()
def tissue_map(tissue):
    return {"caudate": "Caudate", "dlpfc": "DLPFC",
            "hippocampus": "Hippocampus"}[tissue]

@functools.lru_cache()
def feature_map(feature):
    return {"genes": "Gene", "transcripts": "Transcript",
            "exons": "Exon", "junctions": "Junction"}[feature]

@functools.lru_cache()
def get_biomart_df():
    biomart = pd.read_csv(config['biomart_file'], index_col=0)
    biomart['description'] = biomart['description'].str.replace('\[Source.
→*$', '', regex=True)
    return biomart

@functools.lru_cache()
def get_residual_expression_df():
    return pd.read_csv(config['residual_expression_file'], index_col=0).
→transpose()

@functools.lru_cache()
def get_pheno_df():
    return pd.read_csv(config['phenotype_file']).set_index("BrNum").loc[:,
→["RNum", "Sex", "Dx"]]

@functools.lru_cache()
def get_expression_and_pheno_df():
    return pd.merge(get_pheno_df(), get_residual_expression_df(),
                    left_index=True, right_index=True)

@functools.lru_cache()
def get_gene_id_df():
    return pd.DataFrame({'gene_id': get_residual_expression_df().columns,
                        'ensembl_gene_id': get_residual_expression_df().
→columns.str.replace('\.+$', '', regex=True)})
```

```

@functools.lru_cache()
def gene_info_from_symbol(gene_symbol):
    return
    ↳get_biomart_df()[get_biomart_df()['external_gene_name']==gene_symbol]\
        .merge(get_gene_id_df(), on='ensembl_gene_id', how='left')

@functools.lru_cache()
def gene_id_from_symbol(gene_symbol):
    df = gene_info_from_symbol(gene_symbol)
    assert df.shape[0] == 1
    return df[['gene_id']].iloc[0].values[0]

```

## 1.2.2 Genotype and eQTL functions

```

[4]: def letter_snp(number, a0, a1):
    '''
    Example:
    letter_snp(0, 'A', 'G') is 'AA'
    letter_snp(1, 'A', 'G') is 'AG'
    letter_snp(2, 'A', 'G') is 'GG'
    '''
    if np.isnan(number):
        return np.nan
    if len(a0)==1 and len(a1)==1:
        sep = ''
    else:
        sep = ' '
    return sep.join(sorted([a0]*int(number) + [a1]*(2-int(number)))))

@functools.lru_cache()
def get_plink_tuple():
    '''
    Usage: (bim, fam, bed) = get_plink_tuple()
    '''
    return read_plink(config['plink_file_prefix'])

@functools.lru_cache()
def get_eFeature_df():
    eqtl_df = pd.read_csv(config["eqtl_output_file"], sep='\t')
    return eqtl_df[(eqtl_df["Type"] == feature_map(feature)) &
                    (eqtl_df["Tissue"] == tissue_map(tissue))]

```

```

@functools.lru_cache()
def get_gwas_snps():
    return pd.read_csv(config['gwas_snp_file'], sep='\t', index_col=0)

@functools.lru_cache()
def get_risk_allele(snp_id):
    gwas_snp = get_gwas_snp(snp_id)
    if gwas_snp['OR'].iloc[0] > 1:
        ra = gwas_snp['A1'].iloc[0]
    else:
        ra = gwas_snp['A2'].iloc[0]
    return ra

@functools.lru_cache()
def get_snp_df(snp_id):
    """
    Returns a dataframe containing the genotype on snp snp_id.
    The allele count is the same as in the plink files.

    Example:
    get_snp_df('rs653953').head(5)

    rs653953_num rs653953_letter rs653953
    Br5168      0                GG    0\nGG
    Br2582      1                AG    1\nAG
    Br2378      1                AG    1\nAG
    Br5155      2                AA    2\nAA
    Br5182      2                AA    2\nAA
    """
    (bim, fam, bed) = get_plink_tuple()
    brain_ids = list(set(get_expression_and_pheno_df().index).
    ↪ intersection(set(fam['fid'])))
    snp_info = bim[bim['snp']==snp_id]
    snp_pos = snp_info.iloc[0]['i']
    fam_pos = list(fam.drop_duplicates(subset="fid").set_index('fid').
    ↪ loc[brain_ids]['i'])
    dfsnp = (pd.DataFrame(bed[[snp_pos]].compute()[:,fam_pos],
                          columns=brain_ids, index=[snp_id + '_num'])
              .transpose().dropna())
    my_letter_snp = functools.partial(letter_snp, a0=snp_info.iloc[0]['a0'],
    ↪ a1=snp_info.iloc[0]['a1'])
    dfsnp[[snp_id + '_num']] = 2 - dfsnp[[snp_id + '_num']].astype('int')
    dfsnp[snp_id + '_letter'] = dfsnp[snp_id + '_num'].apply(my_letter_snp)
    dfsnp[snp_id] = (dfsnp[snp_id + '_num'].astype('str') + '\n' +
                     dfsnp[snp_id + '_letter'].astype('str')).astype('category')

```

```

return dfsnp

@functools.lru_cache()
def get_gwas_ordered_snp_df(snp_id):
    '''
    Returns a dataframe containing the genotype on snp snp_id.
    The allele count is the number of risk alleles according to GWAS.

    Example:
    get_gwas_ordered_snp_df('rs653953').head(5)

           rs653953_num rs653953_letter rs653953
    Br5168             2             GG      2\nGG
    Br2582             1             AG      1\nAG
    Br2378             1             AG      1\nAG
    Br5155             0             AA      0\nAA
    Br5182             0             AA      0\nAA
    '''
    pgc = get_gwas_snps()
    dfsnp = get_snp_df(snp_id).copy()
    gwas_snp = get_gwas_snp(snp_id)
    if gwas_snp['pgc2_a1_same_as_our_counted'].iloc[0]:
        if gwas_snp['OR'].iloc[0] > 1:
            pass
        else:
            dfsnp[[snp_id + '_num']] = 2 - dfsnp[[snp_id + '_num']]
    else:
        if gwas_snp['OR'].iloc[0] > 1:
            dfsnp[[snp_id + '_num']] = 2 - dfsnp[[snp_id + '_num']]
        else:
            pass
    dfsnp[snp_id] = (dfsnp[snp_id + '_num'].astype('str') + '\n' +
                    dfsnp[snp_id + '_letter'].astype('str')).astype('category')
    return dfsnp

```

### 1.2.3 Plotting functions

```

[5]: def get_snp_gene_pheno_df(snp_id, gene_id, snp_df_func):
    pheno_columns = list(get_pheno_df().columns)
    expr_df = get_expression_and_pheno_df()[pheno_columns + [gene_id]]
    snp_df = snp_df_func(snp_id)
    return expr_df.merge(snp_df, left_index=True, right_index=True)

def simple_snp_expression_plot_impl(snp_id, gene_id, snp_df_func):
    df = get_snp_gene_pheno_df(snp_id, gene_id, snp_df_func)

```

```

y0 = df[gene_id].quantile(.01) - 0.26
y1 = df[gene_id].quantile(.99) + 0.26
p = ggplot(df, aes(x=snp_id, y=gene_id, fill='Sex')) \
+ geom_boxplot(alpha=0.4, outlier_alpha=0) \
+ geom_jitter(position=position_jitterdodge(jitter_width=0.25),
              stroke=0, alpha=0.6) \
+ ylim(y0, y1) \
+ theme_bw(base_size=15) \
+ theme(panel_grid=element_blank(),
        axis_title=element_text(face="bold"))
return p

def simple_snp_expression_plot(snp_id, gene_id):
    return simple_snp_expression_plot_impl(snp_id, gene_id, get_snp_df)

def simple_gwas_ordered_snp_expression_plot(snp_id, gene_id):
    return simple_snp_expression_plot_impl(snp_id, gene_id,
    ↪get_gwas_ordered_snp_df)

def get_gene_symbol(gene_id, biomart=get_biomart_df()):
    ensge = re.sub('\.+$', '', gene_id)
    ggg = biomart[biomart['ensembl_gene_id']==ensge]
    if ggg.shape[0]==0:
        return '', ''
    gs = ggg['external_gene_name'].values[0]
    de = ggg['description'].values[0]
    if type(de)!=str:
        de = ''
    de = re.sub('\[Source:.*$', '', de)
    return gs, de

def get_gwas_snp(snp_id):
    gwas = get_gwas_snps()
    r = gwas[gwas['our_snp_id']==snp_id]
    assert len(r) == 1
    return r

def gwas_annotation(snp_id):
    return 'SZ GWAS pvalue: %.1e' % get_gwas_snp(snp_id).iloc[0]['P']

def eqtl_annotation(snp_id, gene_id):

```

```

eqtl_df = get_eFeature_df()
r = eqtl_df[(eqtl_df['variant_id']==snp_id) & (eqtl_df['gene_id']==gene_id)]
assert len(r)==1
return 'eQTL adjusted p-value: %.1e' % r.iloc[0]['BF']

def risk_allele_annotation(snp_id):
    return 'SZ risk allele: %s' % get_risk_allele(snp_id)

def annotated_eqtl_plot(snp_id, gene_id):
    p = simple_snp_expression_plot(snp_id, gene_id)
    gene_symbol, gene_description = get_gene_symbol(gene_id)
    title = "\n".join([gene_symbol,
                       eqtl_annotation(snp_id, gene_id)
                      ])
    p += ggtitle(title) + ylab('Residualized Expression')
    return p

def gwas_annotated_eqtl_plot(snp_id, gene_id):
    p = simple_gwas_ordered_snp_expression_plot(snp_id, gene_id)
    gene_symbol, gene_description = get_gene_symbol(gene_id)
    title = "\n".join([gene_symbol,
                       eqtl_annotation(snp_id, gene_id),
                       gwas_annotation(snp_id),
                       risk_allele_annotation(snp_id)
                      ])
    p += ggtitle(title) + ylab('Residualized Expression')
    return p

def save_plot(p, fn):
    for ext in ['png', 'pdf', 'svg']:
        p.save(fn + '.' + ext)

```

## 1.3 Plot eQTLs

### 1.3.1 DRD2

```
[6]: get_eFeature_df()[get_eFeature_df()["gene_id"] == gene_id_from_symbol('DRD2')]
```

```
[6]: Empty DataFrame
Columns: [variant_id, gene_id, gencodeID, slope, statistic, pval_nominal, BF,
eigenMT_BH, TESTS, Type, Tissue]
Index: []
```

### 1.3.2 Top 5 eQTLs

```
[7]: eqtl_df = get_eFeature_df()
eqtl_df.head()
```

```
[7]:
```

	variant_id	gene_id	gencodeID	\
7125	chr7:42936690:A:C	ENSG000000002746.14	ENSG000000002746.14	
7126	chr17:48075934:C:T	ENSG000000002919.14	ENSG000000002919.14	
7127	chr7:7934340:A:C	ENSG000000003147.17	ENSG000000003147.17	
7128	chr16:89536451:CGGTGAGGCG:C	ENSG000000003249.13	ENSG000000003249.13	
7129	chr8:17754947:C:T	ENSG000000003989.16	ENSG000000003989.16	

	slope	statistic	pval_nominal	BF	eigenMT_BH	TESTS	Type	\
7125	0.436502	7.820971	0.000023	0.010399	0.445692	445	Gene	
7126	-0.536903	-7.959488	0.000127	0.037424	0.549367	294	Gene	
7127	0.213896	11.357107	0.000022	0.022195	0.503766	1002	Gene	
7128	-0.524657	-9.921271	0.000028	0.014577	0.470811	529	Gene	
7129	-0.497351	-11.806077	0.000007	0.007416	0.417015	1040	Gene	

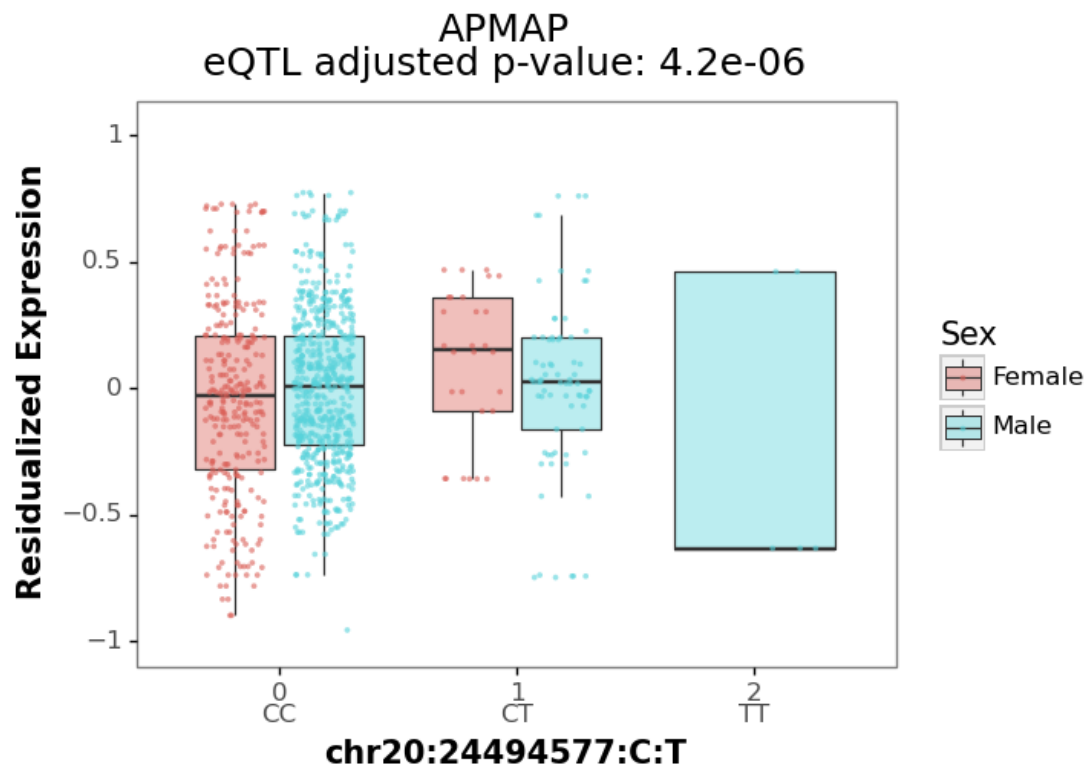
	Tissue
7125	Caudate
7126	Caudate
7127	Caudate
7128	Caudate
7129	Caudate

```
[8]: top_5 = eqtl_df.sort_values('pval_nominal').reset_index(drop=True).head(5)
for x in top_5.itertuples():
    filename = "top_%d_eqtl_%s" % (x.Index, tissue)
    p = annotated_eqtl_plot(x.variant_id, x.gene_id)
    print(filename, x.Index, x.variant_id, x.gene_id)
    print(p)
    save_plot(p, filename)
```

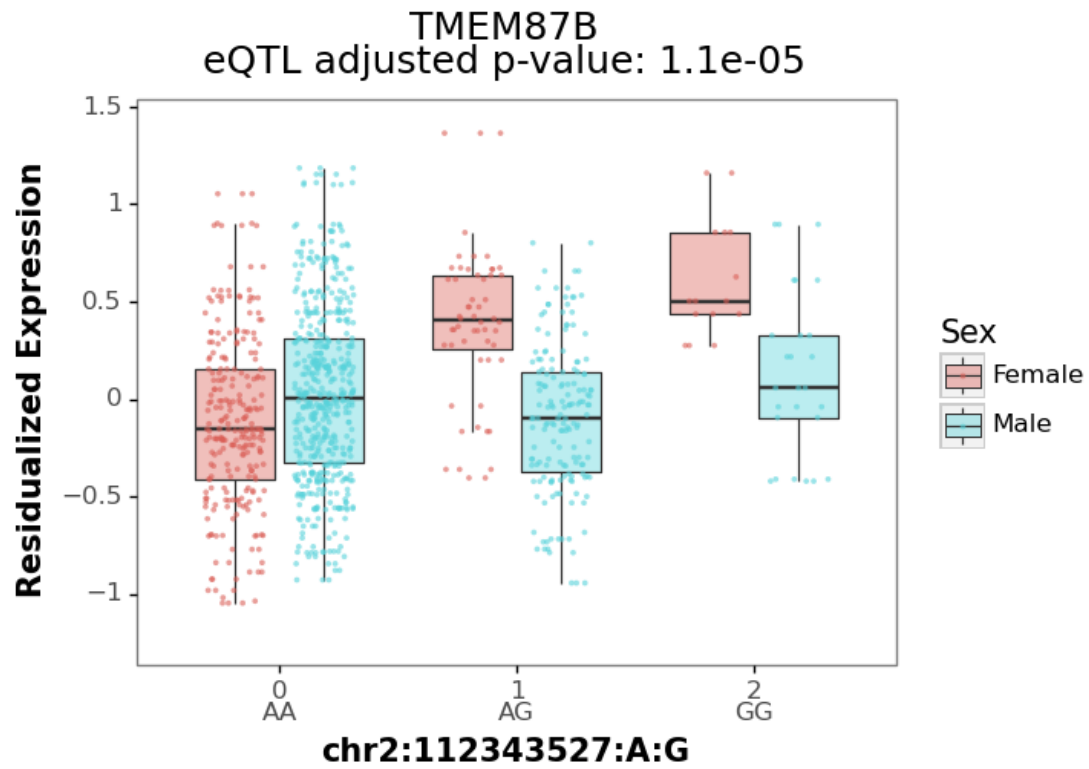
Mapping files: 100%| | 3/3 [00:26<00:00, 8.79s/it]

top\_0\_eqtl\_caudate 0 chr20:24494577:C:T ENSG00000101474.11

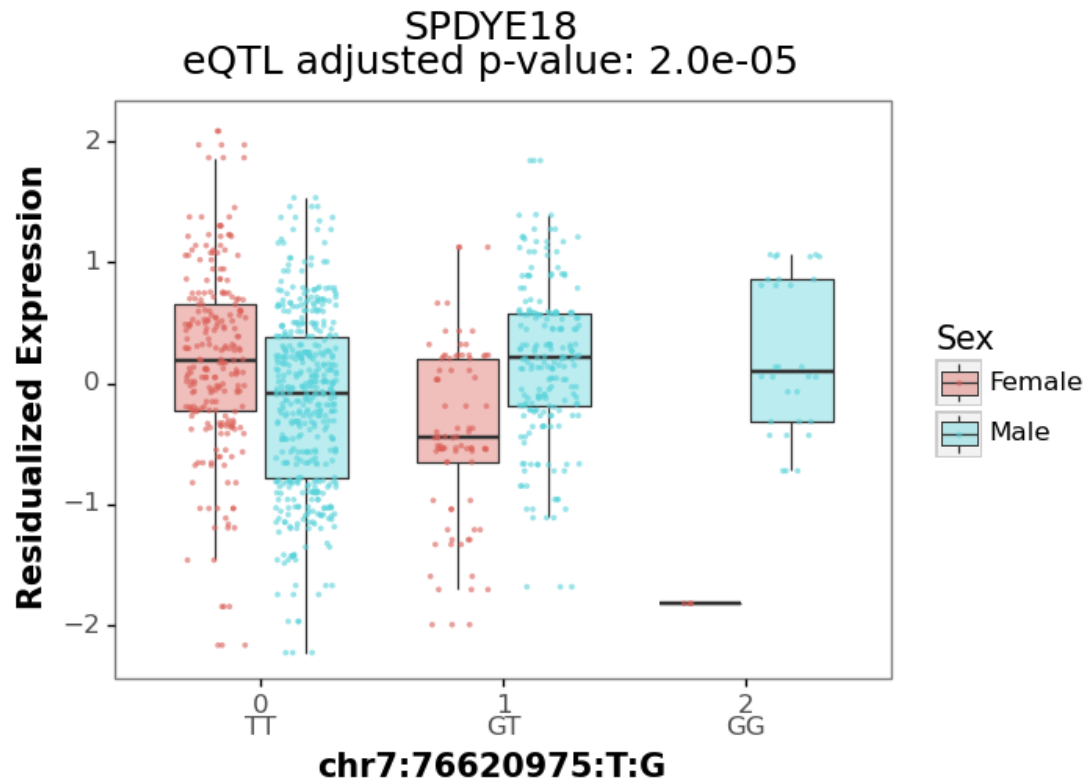




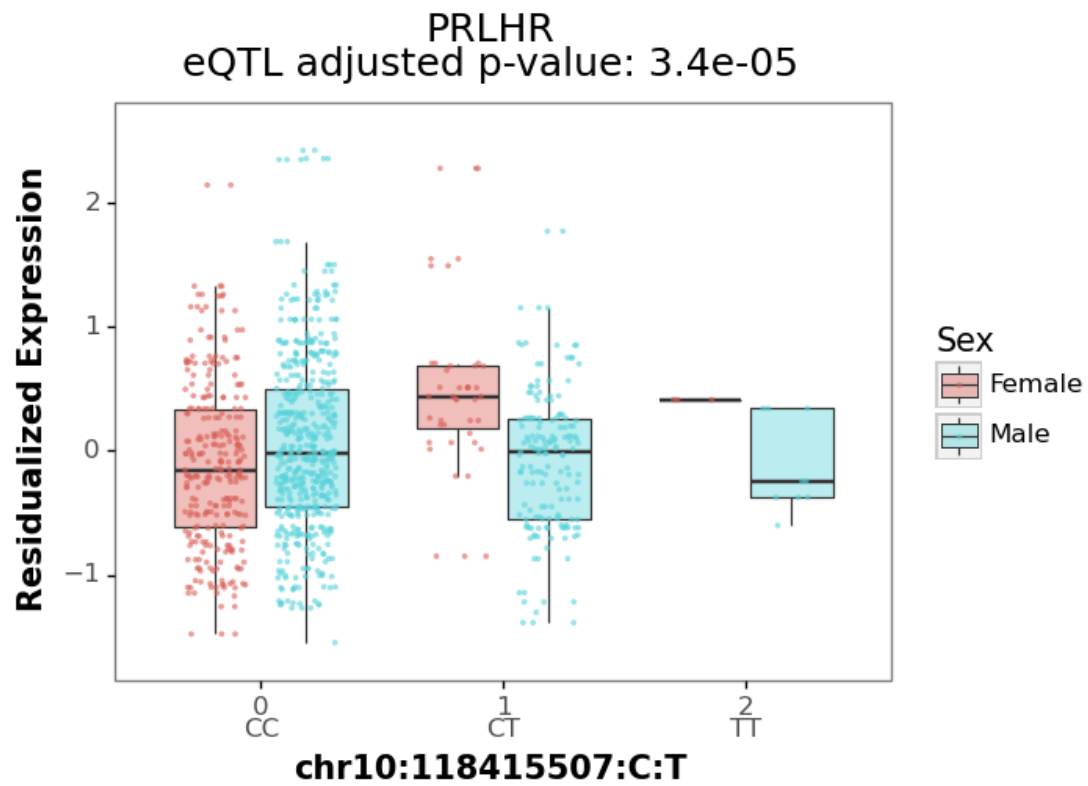
```
<ggplot: (8728063515606)>
top_1_eqtl_caudate 1 chr2:112343527:A:G ENSG00000153214.9
```



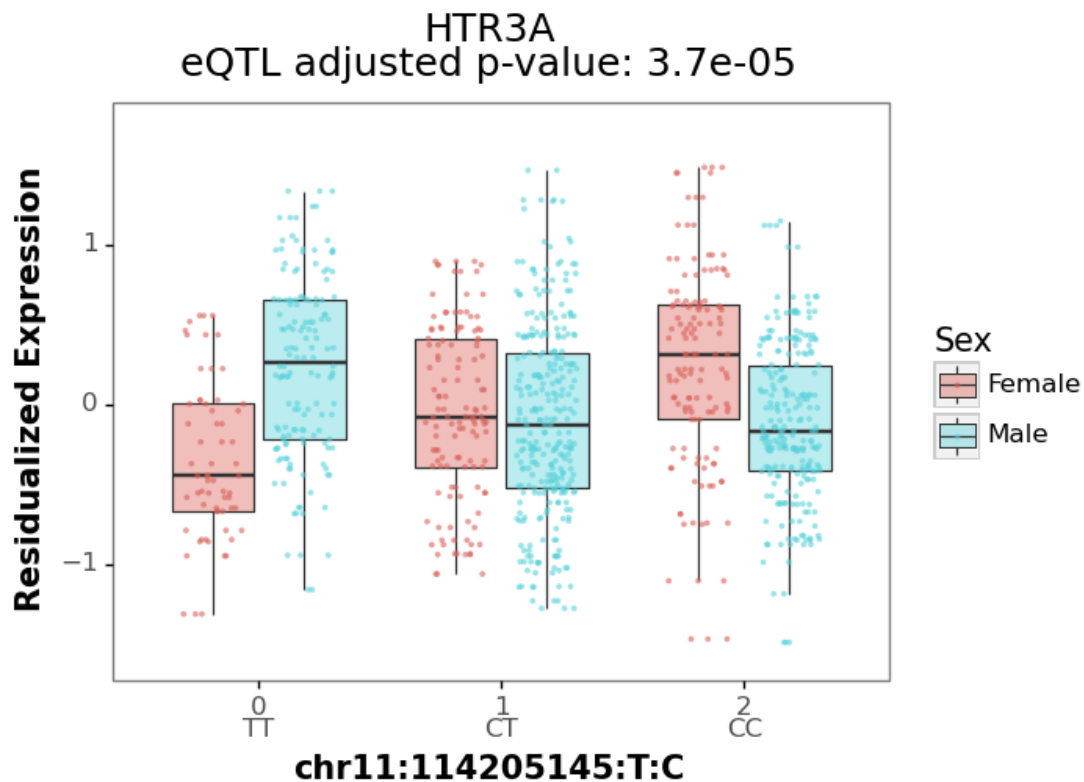
```
<ggplot: (8728063342174)>
top_2_eqtl_caudate 2 chr7:76620975:T:G ENSG00000205482.9
```



```
<ggplot: (8728063254492)>
top_3_eqtl_caudate 3 chr10:118415507:C:T ENSG00000119973.5
```



```
<ggplot: (8728062620182)>
top_4_eqtl_caudate 4 chr11:114205145:T:C ENSG00000166736.11
```



<ggplot: (8728062803366)>

### 1.3.3 Top 5 X-linked genes

```
[9]: top_5_x = eqtl_df[eqtl_df['variant_id'].str.contains("chrX")].
      ↪sort_values("pval_nominal").reset_index(drop=True).head(5)
for x in top_5_x.itertuples():
    filename = "top_%d_eqtl_xlinked_%s" % (x.Index, tissue)
    p = annotated_eqtl_plot(x.variant_id, x.gene_id)
    print(filename, x.Index, x.variant_id, x.gene_id)
    print(p)
    save_plot(p, filename)
```

### 1.3.4 Top 5 eQTL with GWAS significant index SNP

```
[10]: gwas_eqtl_df = eqtl_df.merge(get_gwas_snps(), left_on = 'variant_id',
                                   right_on = 'our_snps_id', suffixes=['', '_gwas'])
print(gwas_eqtl_df.shape)
gwas_eqtl_df.head()
```

(3, 33)

```
[10]:
```

	variant_id	gene_id	gencodeID	slope	\
0	chr22:42119633:G:C	ENSG00000183172.8	ENSG00000183172.8	0.215764	
1	chr6:32140074:G:A	ENSG00000204371.11	ENSG00000204371.11	-0.509024	
2	chr6:31867387:G:C	ENSG00000244731.7	ENSG00000244731.7	-0.632753	

	statistic	pval_nominal	BF	eigenMT_BH	TESTS	Type	...	A2	OR	\
0	10.770408	5.788120e-05	0.024426	0.516399	422	Gene	...	C	1.0629	
1	-8.596720	5.852160e-05	0.029436	0.536823	503	Gene	...	A	1.1331	
2	-13.250431	1.564220e-07	0.000079	0.123254	502	Gene	...	C	1.1744	

	SE	P	hg19chrc	hg38chrc	hg38pos	\
0	0.010193	2.130000e-09	chr22	chr22	42119633	
1	0.014200	1.370000e-18	chr6	chr6	32140074	
2	0.014203	1.050000e-29	chr6	chr6	31867387	

	pgc2_a1_same_as_our_counted	rsid	is_index_snp
0	False	rs8143153	False
1	False	rs3134962	False
2	False	rs693906	False

[3 rows x 33 columns]

```
[11]: top_gwas_eqtl_df = gwas_eqtl_df[(gwas_eqtl_df['is_index_snp'])].
      ↪sort_values(['BF', 'P'])
      print(top_gwas_eqtl_df.shape)
      top_gwas_eqtl_df.head()
```

(0, 33)

```
[11]: Empty DataFrame
Columns: [variant_id, gene_id, gencodeID, slope, statistic, pval_nominal, BF,
eigenMT_BH, TESTS, Type, Tissue, chrN, our_snp_id, cm, pos, our_counted,
our_alt, chrom, SNP, Freq.A1, CHR, BP, A1, A2, OR, SE, P, hg19chrc, hg38chrc,
hg38pos, pgc2_a1_same_as_our_counted, rsid, is_index_snp]
Index: []
```

[0 rows x 33 columns]

```
[12]: top_gwas_eqtl_df = gwas_eqtl_df.sort_values(['BF', 'P']).reset_index(drop=True)
      print(top_gwas_eqtl_df.shape)
      top_gwas_eqtl_df.head(10)
```

(3, 33)

```
[12]:
```

	variant_id	gene_id	gencodeID	slope	\
0	chr6:31867387:G:C	ENSG00000244731.7	ENSG00000244731.7	-0.632753	
1	chr22:42119633:G:C	ENSG00000183172.8	ENSG00000183172.8	0.215764	
2	chr6:32140074:G:A	ENSG00000204371.11	ENSG00000204371.11	-0.509024	

	statistic	pval_nominal	BF	eigenMT_BH	TESTS	Type	...	A2	OR	\
0	-13.250431	1.564220e-07	0.000079	0.123254	502	Gene	...	C	1.1744	
1	10.770408	5.788120e-05	0.024426	0.516399	422	Gene	...	C	1.0629	
2	-8.596720	5.852160e-05	0.029436	0.536823	503	Gene	...	A	1.1331	

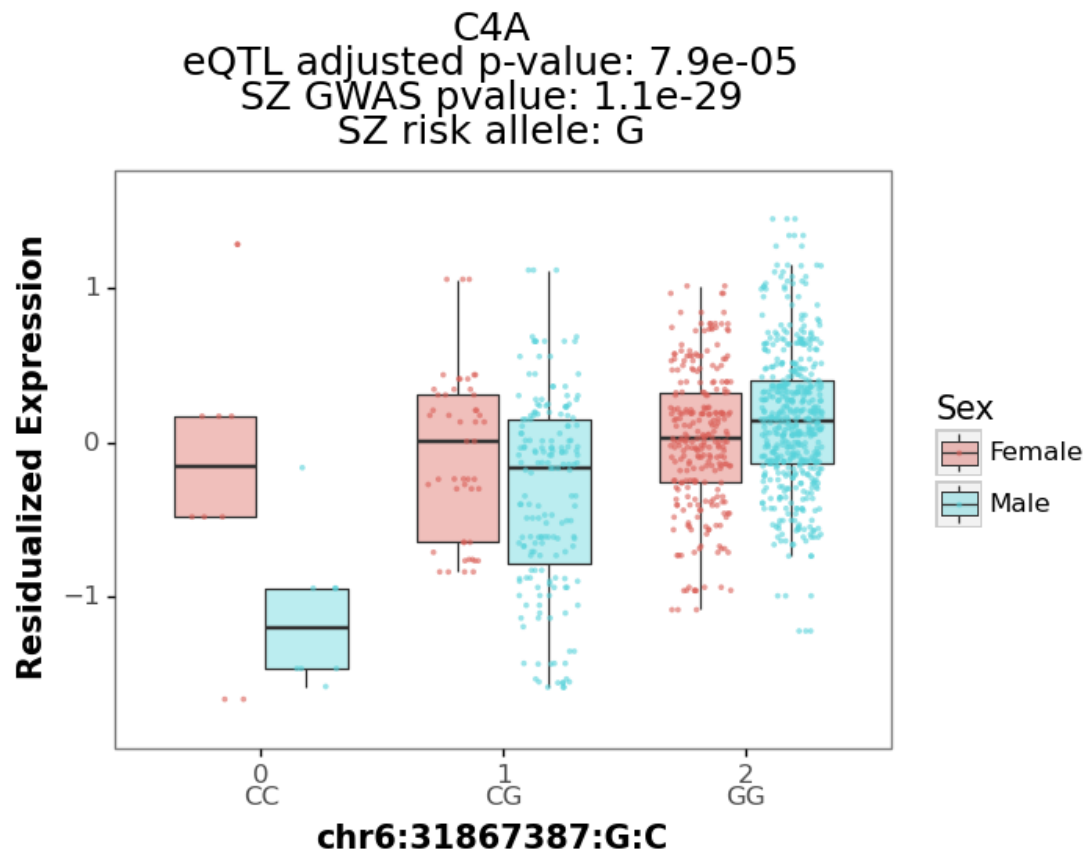
	SE	P	hg19chr	hg38chr	hg38pos	\
0	0.014203	1.050000e-29	chr6	chr6	31867387	
1	0.010193	2.130000e-09	chr22	chr22	42119633	
2	0.014200	1.370000e-18	chr6	chr6	32140074	

	pgc2_a1_same_as_our_counted	rsid	is_index_snp
0	False	rs693906	False
1	False	rs8143153	False
2	False	rs3134962	False

[3 rows x 33 columns]

```
[13]: top_5_gwas = top_gwas_eqtl_df.head(5)
for x in top_5_gwas.itertuples():
    filename = "top_%d_eqtl_in_gwas_significant_snps_%s" % (x.Index, tissue)
    p = gwas_annotated_eqtl_plot(x.variant_id, x.gene_id)
    print(filename, x.Index, x.variant_id, x.gene_id)
    print(p)
    save_plot(p, filename)
```

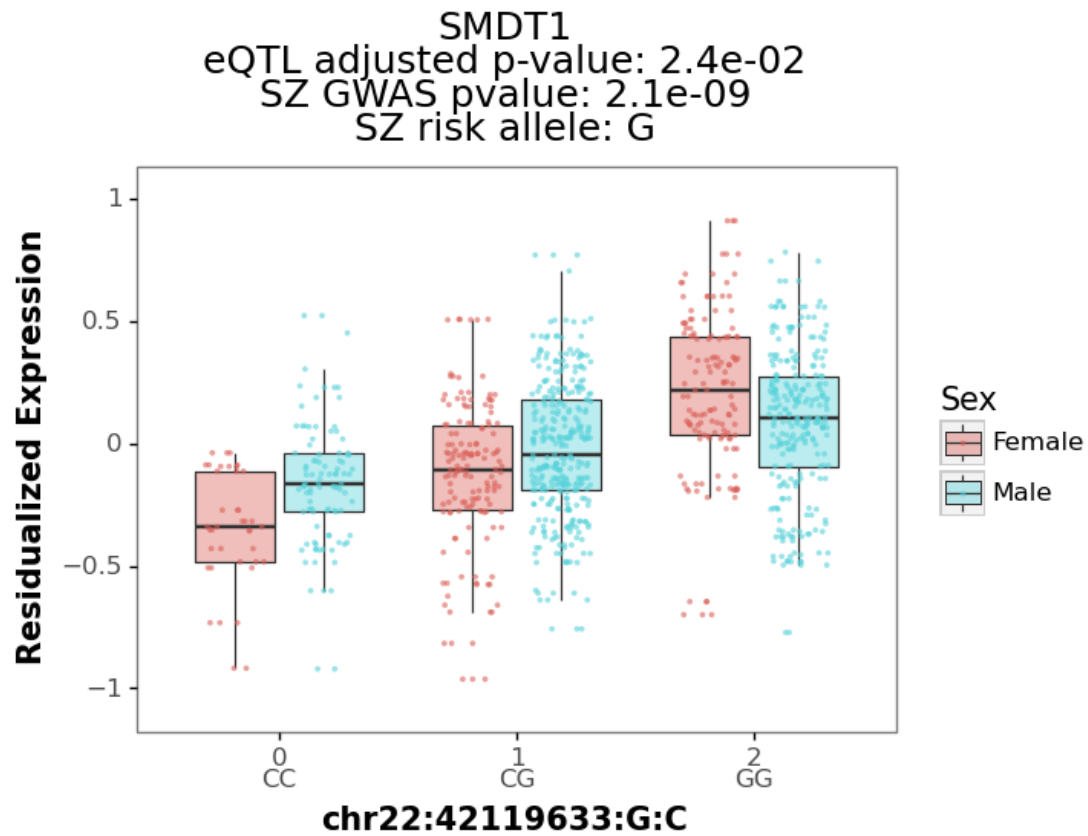
```
top_0_eqtl_in_gwas_significant_snps_caudate 0 chr6:31867387:G:C
ENSG00000244731.7
```



```
<ggplot: (8728062453709)>
```

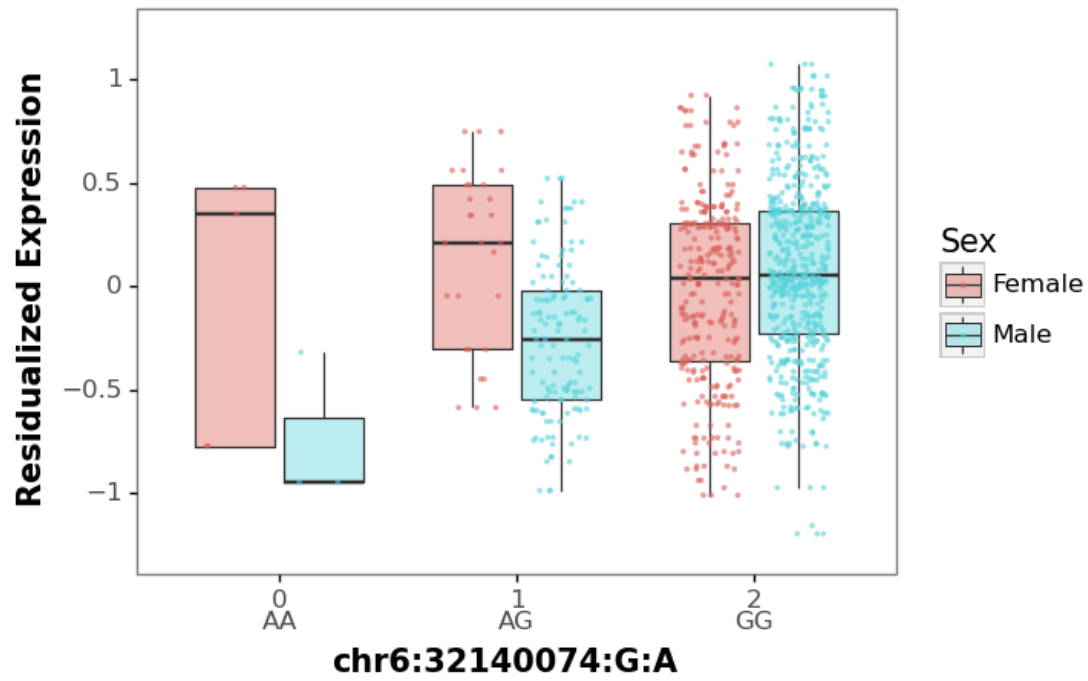
```
top_1_eqtl_in_gwas_significant_snps_caudate 1 chr22:42119633:G:C  
ENSG00000183172.8
```





```
<ggplot: (8728184152691)>
top_2_eqtl_in_gwas_significant_snps_caudate 2 chr6:32140074:G:A
ENSG00000204371.11
```

EHMT2  
eQTL adjusted p-value: 2.9e-02  
SZ GWAS pvalue: 1.4e-18  
SZ risk allele: G



<ggplot: (8728062942880)>

[ ]: