main

July 12, 2021

1 Differential Expression with limma-voom pipeline

1.1 Functions

1.1.1 Simple functions

```
[2]: # Function from jaffelab github
     merge_rse_metrics <- function(rse) {</pre>
         stopifnot(is(rse, 'RangedSummarizedExperiment'))
         rse$overallMapRate = mapply(function(r, n) {
             sum(r*n)/sum(n)
         }, rse$overallMapRate, rse$numReads)
         rse$mitoRate = mapply(function(r, n) {
             sum(r*n)/sum(n)
         }, rse$mitoRate, rse$numMapped)
         rse$rRNA_rate = mapply(function(r, n) {
             sum(r*n)/sum(n)
         }, rse$rRNA_rate, rse$numMapped)
         rse$totalAssignedGene = mapply(function(r, n) {
             sum(r*n)/sum(n)
         }, rse$totalAssignedGene, rse$numMapped)
         rse$numMapped = sapply(rse$numMapped, sum)
         rse$numReads = sapply(rse$numReads, sum)
         rse$numUnmapped = sapply(rse$numUnmapped, sum)
         rse$mitoMapped = sapply(rse$mitoMapped, sum)
         rse$totalMapped = sapply(rse$totalMapped, sum)
         return(rse)
     }
```

```
save_volcanoPlot <- function(top, label, feature){</pre>
    pdf(file=paste0(feature, "/volcanoPlot_", label, ".pdf"), 8, 6)
    with(top, plot(logFC, -log10(P.Value), pch=20, cex=0.6))
    with(subset(top, adj.P.Val<=0.05), points(logFC, -log10(P.Value),
                                                pch=20, col='red', cex=0.6))
    with(subset(top, abs(logFC)>0.50), points(logFC, -log10(P.Value),
                                                pch=20, col='orange', cex=0.6))
    with(subset(top, adj.P.Val<=0.05 & abs(logFC)>0.50),
         points(logFC, -log10(P.Value), pch=20, col='green', cex=0.6))
    dev.off()
}
save_MAplot <- function(top, label, feature){</pre>
    pdf(file=paste0(feature, "/MAplot_", label, ".pdf"), 8, 6)
    with(top, plot(AveExpr, logFC, pch=20, cex=0.5))
    with(subset(top, adj.P.Val<0.05),</pre>
         points(AveExpr, logFC, col="red", pch=20, cex=0.5))
    dev.off()
}
extract_de <- function(contrast, label, efit, feature){</pre>
    top <- topTable(efit, coef=contrast, number=Inf, sort.by="P")</pre>
    top <- top[order(top$P.Value), ]</pre>
    top.fdr <- top %>% filter(adj.P.Val<=0.05)</pre>
    print(paste("Comparison for:", label))
    print(paste('There are:', dim(top.fdr)[1], 'DE features!'))
    data.table::fwrite(top, file=paste0(feature, "/diffExpr_", label, "_full.
sep='\t', row.names=TRUE)
    data.table::fwrite(top.fdr, file=paste0(feature, "/diffExpr_", label, __
\hookrightarrow" FDR05.txt"),
                        sep='\t', row.names=TRUE)
    save_volcanoPlot(top, label, feature)
    save_MAplot(top, label, feature)
}
```

1.1.2 Cached functions

```
return (mds)
}
memMDS <- memoise::memoise(get_mds)</pre>
prep_data <- function(feature){</pre>
    counts_lt = list("genes"="/ceph/projects/v4_phase3_paper/inputs/counts/_m/
 →hippo_brainseq_phase2_hg38_rseGene_merged_n447.rda",
                     "transcripts"="/ceph/projects/v4_phase3_paper/inputs/
"exons"="/ceph/projects/v4_phase3_paper/inputs/counts/_m/
→hippo_brainseq_phase2_hg38_rseExon_merged_n447.rda",
                     "junctions"="/ceph/projects/v4_phase3_paper/inputs/counts/
 → m/hippo_brainseq_phase2_hg38_rseJxn_merged_n447.rda")
    tx_file = "/ceph/projects/v4_phase3_paper/inputs/counts/_m/
⇔transcripts_counts/hippo_counts.txt"
   load(counts lt[[feature]])
    if(exists("rse gene")){
       rse_df = rse_gene
   } else if (exists("rse_tx")){
       counts <- data.table::fread(tx_file) %>%_
 →column_to_rownames("transcript_id") %>%
            select(colnames(rse tx)) %>% as.matrix
       annot <- data.table::fread(tx file) %>%|
→column_to_rownames("transcript_id") %>%
            select(-starts_with("R"))
       rse_df <- SummarizedExperiment(assays=SimpleList(counts=counts),</pre>
                                      rowData=annot,
                                       colData=colData(rse_tx)) %>%
            as("RangedSummarizedExperiment")
   } else if (exists("rse_exon")){
       rse_df = rse_exon
   } else {
       rse_df = rse_jxn
   keepIndex = which((rse df$Dx %in% c("Control", "Schizo")) &
                      rse_df$Age > 17 & rse_df$Race %in% c("AA", "CAUC"))
   rse_df = rse_df[, keepIndex]
   rse_df$Dx = factor(rse_df$Dx, levels = c("Control", "Schizo"))
   rse_df$Sex <- factor(rse_df$Sex)</pre>
   rse_df <- merge_rse_metrics(rse_df)</pre>
    colData(rse_df)$RIN = sapply(colData(rse_df)$RIN,"[",1)
   rownames(colData(rse_df)) <- sapply(strsplit(rownames(colData(rse_df)),__
 \rightarrow"_"), "[", 1)
   pheno = colData(rse_df) %>% as.data.frame %>%
        inner_join(memMDS(), by=c("BrNum"="FID")) %>%
```

```
distinct(RNum, .keep_all = TRUE)
    # Generate DGE list
    x <- DGEList(counts=assays(rse_df)$counts[, pheno$RNum],
                  genes=rowData(rse_df), samples=pheno)
    # Filter by expression
    design0 <- model.matrix(~Dx*Sex, data=x$samples)</pre>
    keep.x <- filterByExpr(x, design=design0)</pre>
    x <- x[keep.x, , keep.lib.sizes=FALSE]</pre>
    print(paste('There are:', sum(keep.x), 'features left!', sep=' '))
    # Normalize library size
    x <- calcNormFactors(x, method="TMM")</pre>
    return(x)
}
memo_prepData <- memoise::memoise(prep_data)</pre>
SVA_model <- function(feature){</pre>
    x <- memo_prepData(feature)</pre>
    # Design matrix
    mod = model.matrix(~Dx*Sex + Age + mitoRate + rRNA_rate +
                         totalAssignedGene + RIN + overallMapRate +
                         snpPC1 + snpPC2 + snpPC3, data = x$samples)
    colnames(mod) <- gsub("Dx", "", colnames(mod))</pre>
    colnames(mod) <- gsub("SexM", "Male", colnames(mod))</pre>
    colnames(mod) <- gsub("Schizo:Male", "Schizo_Male", colnames(mod))</pre>
    colnames(mod) <- gsub("\\(Intercept\\)", "Intercept", colnames(mod))</pre>
    # Null model
    null.model = mod %>% as.data.frame %>%
        select(-c("Male", "Schizo", "Schizo_Male")) %>% as.matrix
    n.sv <- num.sv(x$counts, mod, method="be")</pre>
    ## Fit SVA
    svobj <- svaseq(x$counts, mod, null.model, n.sv=n.sv)</pre>
    ## Add to model
    print(paste('Adding SV to design matrix ...', Sys.time(), sep=' '))
    modQsva <- cbind(mod, svobj$sv)</pre>
    len.d <- length(colnames(modQsva))</pre>
    colnames(modQsva)[((len.d - n.sv)+1):len.d] <- make.names(paste0("sv",1:n.</pre>
⇒sv))
    return(modQsva)
}
memSVA <- memoise::memoise(SVA_model)</pre>
get_voom <- function(feature){</pre>
    ### Preform voom
    x <- memo_prepData(feature)</pre>
    modQsva <- memSVA(feature)</pre>
```

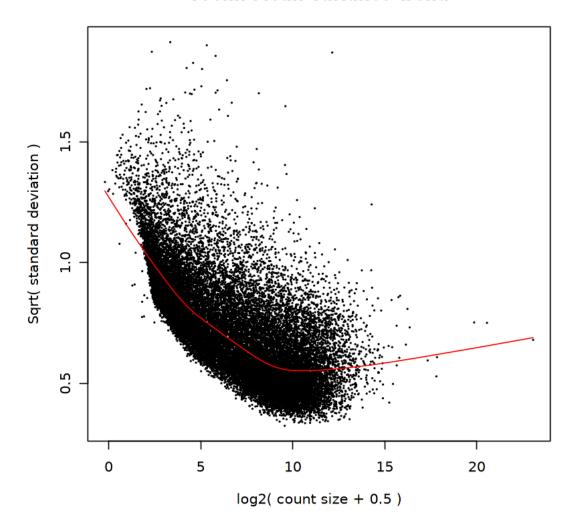
```
v <- voom(x[, rownames(modQsva)], modQsva, plot=TRUE)</pre>
    return(v)
}
memo_voom <- memoise::memoise(get_voom)</pre>
cal res <- function(feature){</pre>
    ### Calculate residuals
    v <- memo voom(feature)</pre>
    null model <- v$design %>% as.data.frame %>%
         select(-c("Male", "Schizo", "Schizo_Male")) %>% as.matrix
    fit_res <- lmFit(v, design=null_model)</pre>
    res = v$E - ( fit_res$coefficients %*% t(null_model) )
    res_sd = apply(res, 1, sd)
    res_mean = apply(res, 1, mean)
    res_norm = (res - res_mean) / res_sd
    write.table(res_norm, file=paste0(feature, '/residualized_expression.tsv'),
                 sep="\t", quote=FALSE)
}
memo_res <- memoise::memoise(cal_res)</pre>
fit_voom <- function(feature){</pre>
    v <- memo voom(feature)</pre>
    modQsva <- memSVA(feature)</pre>
    fit0 <- lmFit(v, modQsva)</pre>
    contr.matrix <- makeContrasts(CtrlvsSZ = Schizo,</pre>
                                     MvsF = Male,
                                     Interaction = Schizo_Male,
                                     levels=colnames(modQsva))
    fit <- contrasts.fit(fit0, contrasts=contr.matrix)</pre>
    esv <- eBayes(fit)</pre>
    return(esv)
}
memo_efit <- memoise::memoise(fit_voom)</pre>
```

1.2 Differential Expression Analysis

```
[4]: groups = c('szVctl', 'maleVfemale', 'interaction')
for(feature in c('genes', 'transcripts', 'junctions', 'exons')){
    dir.create(feature)
    # Preform voom
    v <- memo_voom(feature)
    save(v, file=pasteO(feature,'/voomSVA.RData'))
    # Fit model and apply eBayes
    efit = memo_efit(feature)</pre>
```

```
# Save differential expression
    for(group in seq_along(groups)){
        print(groups[group])
        extract_de(group, groups[group], efit, feature)
    # Calculate residuals
    memo_res(feature)
}
[1] "There are: 24937 features left!"
Number of significant surrogate variables is: 1
Iteration (out of 5 ):1 2 3 4 5 [1] "Adding SV to design matrix ...
2021-07-12 09:22:08"
[1] "szVctl"
[1] "Comparison for: szVctl"
[1] "There are: 0 DE features!"
[1] "maleVfemale"
[1] "Comparison for: maleVfemale"
[1] "There are: 92 DE features!"
[1] "interaction"
[1] "Comparison for: interaction"
[1] "There are: 0 DE features!"
[1] "There are: 96295 features left!"
Number of significant surrogate variables is: 1
Iteration (out of 5):1 2 3 4 5 [1] "Adding SV to design matrix ...
2021-07-12 09:33:50"
[1] "szVctl"
[1] "Comparison for: szVctl"
[1] "There are: 1 DE features!"
[1] "maleVfemale"
[1] "Comparison for: maleVfemale"
[1] "There are: 231 DE features!"
[1] "interaction"
```

[1] "Comparison for: interaction"
[1] "There are: 0 DE features!"



```
[1] "There are: 159519 features left!"

Number of significant surrogate variables is: 4

Iteration (out of 5 ):1 2 3 4 5 [1] "Adding SV to design matrix ...

2021-07-12 09:55:26"

[1] "szVctl"

[1] "Comparison for: szVctl"

[1] "There are: 26 DE features!"

[1] "maleVfemale"

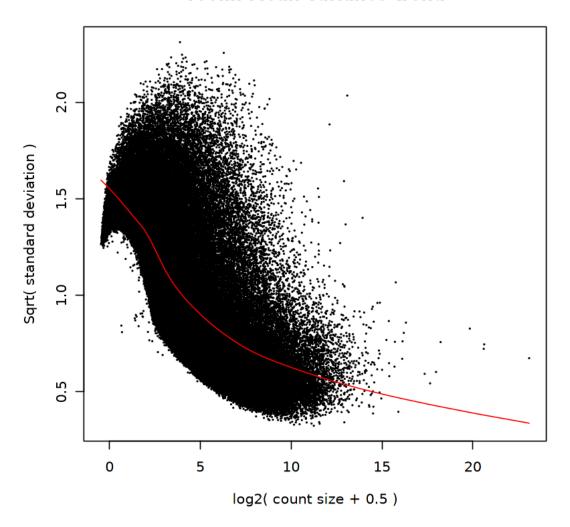
[1] "Comparison for: maleVfemale"

[1] "There are: 633 DE features!"

[1] "interaction"

[1] "Comparison for: interaction"
```

[1] "There are: 6 DE features!"

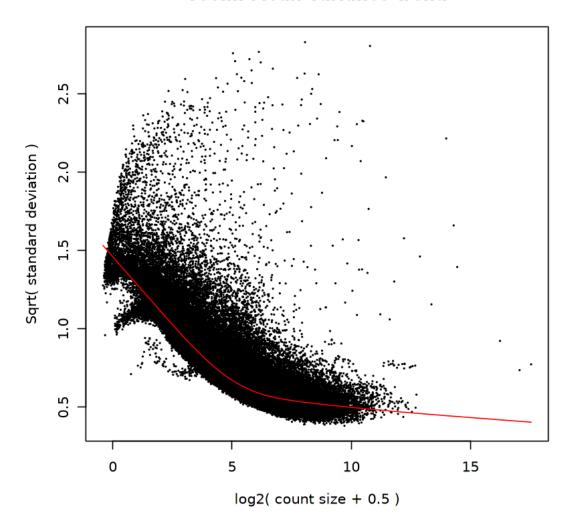


- [1] "There are: 363747 features left!"
- [1] "There are: 363747 features left!"

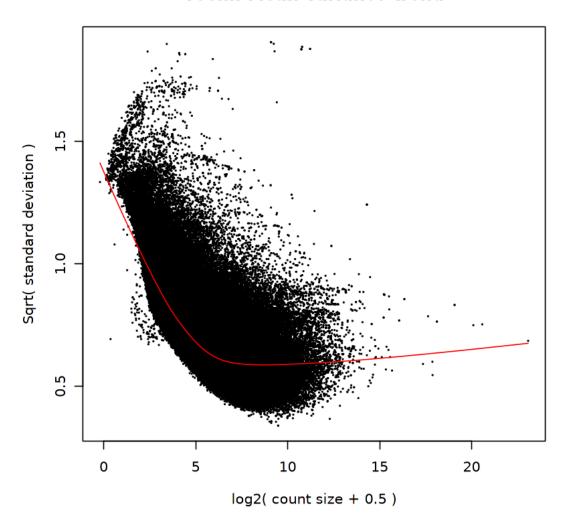
Number of significant surrogate variables is: 1

Iteration (out of 5):1 2 3 4 5 [1] "Adding SV to design matrix ...

2021-07-12 10:45:23"



- [1] "There are: 363747 features left!"
- [1] "szVctl"
- [1] "Comparison for: szVctl"
- [1] "There are: 0 DE features!"
- [1] "maleVfemale"
- [1] "Comparison for: maleVfemale"
- [1] "There are: 906 DE features!"
- [1] "interaction"
- [1] "Comparison for: interaction"
- [1] "There are: 0 DE features!"
- [1] "There are: 363747 features left!"



1.3 Repreducibility Information

```
[5]: Sys.time()
    proc.time()
    options(width = 120)
    sessioninfo::session_info()

[1] "2021-07-12 11:06:45 EDT"

    user system elapsed
    23720.308 100541.382 6512.117

    Session info
    setting value
```

version R version 4.0.3 (2020-10-10)

os Arch Linux

system x86_64, linux-gnu

ui X11 language (EN)

collate en_US.UTF-8
ctype en_US.UTF-8
tz America/New_York

date 2021-07-12

Packages

package	*	version	date	lib	source
annotate		1.68.0	2020-10-27	[1]	Bioconductor
AnnotationDbi		1.52.0	2020-10-27	[1]	Bioconductor
assertthat		0.2.1	2019-03-21	[1]	CRAN (R 4.0.2)
backports		1.2.1	2020-12-09	[1]	CRAN (R 4.0.2)
base64enc		0.1-3	2015-07-28	[1]	CRAN (R 4.0.2)
Biobase	*	2.50.0	2020-10-27	[1]	Bioconductor
BiocGenerics	*	0.36.1	2021-04-16	[1]	Bioconductor
BiocParallel	*	1.24.1	2020-11-06	[1]	Bioconductor
bit		4.0.4	2020-08-04	[1]	CRAN (R 4.0.2)
bit64		4.0.5	2020-08-30	[1]	CRAN (R 4.0.2)
bitops		1.0-7	2021-04-24	[1]	CRAN (R 4.0.3)
blob		1.2.1	2020-01-20	[1]	CRAN (R 4.0.2)
broom		0.7.8	2021-06-24	[1]	CRAN (R 4.0.3)
cachem		1.0.5	2021-05-15	[1]	CRAN (R 4.0.3)
Cairo		1.5-12.2	2020-07-07	[1]	CRAN (R 4.0.2)
cellranger		1.1.0	2016-07-27	[1]	CRAN (R 4.0.2)
cli		3.0.0	2021-06-30	[1]	CRAN (R 4.0.3)
colorspace		2.0-2	2021-06-24	[1]	CRAN (R 4.0.3)
crayon		1.4.1	2021-02-08	[1]	CRAN (R 4.0.3)
data.table		1.14.0	2021-02-21	[1]	CRAN (R 4.0.3)
DBI		1.1.1	2021-01-15	[1]	CRAN (R 4.0.2)
dbplyr		2.1.1	2021-04-06	[1]	CRAN (R 4.0.3)
DelayedArray		0.16.3	2021-03-24	[1]	Bioconductor
digest		0.6.27	2020-10-24	[1]	CRAN (R 4.0.2)
dplyr	*	1.0.7	2021-06-18	[1]	CRAN (R 4.0.3)
edgeR	*	3.32.1	2021-01-14	[1]	Bioconductor
ellipsis		0.3.2	2021-04-29	[1]	CRAN (R 4.0.3)
evaluate		0.14	2019-05-28	[1]	CRAN (R 4.0.2)
fansi		0.5.0	2021-05-25	[1]	CRAN (R 4.0.3)
fastmap		1.1.0	2021-01-25	[1]	CRAN (R 4.0.2)
forcats	*	0.5.1	2021-01-27	[1]	CRAN (R 4.0.2)
fs		1.5.0	2020-07-31	[1]	CRAN (R 4.0.2)
genefilter	*	1.72.1	2021-01-21	[1]	Bioconductor
generics		0.1.0	2020-10-31	[1]	CRAN (R 4.0.2)
GenomeInfoDb	*	1.26.7	2021-04-08	[1]	Bioconductor
${\tt GenomeInfoDbData}$		1.2.4	2021-02-02	[1]	Bioconductor

GenomicRanges	*	1.42.0	2020-10-27	[1]	Bioconductor
ggplot2		3.3.5		[1]	
glue		1.4.2	2020-08-27		CRAN (R 4.0.2)
gtable		0.3.0	2019-03-25	[1]	CRAN (R 4.0.2)
haven		2.4.1	2021-04-23	[1]	CRAN (R 4.0.3)
hms		1.1.0	2021-05-17	[1]	CRAN (R 4.0.3)
htmltools		0.5.1.1	2021-01-22	[1]	CRAN (R 4.0.2)
httr		1.4.2	2020-07-20	[1]	CRAN (R 4.0.2)
IRanges	*	2.24.1	2020-12-12	[1]	Bioconductor
IRdisplay		1.0	2021-01-20	[1]	CRAN (R 4.0.2)
IRkernel		1.2	2021-05-11	[1]	CRAN (R 4.0.3)
jsonlite		1.7.2	2020-12-09	[1]	CRAN (R 4.0.2)
lattice		0.20-41	2020-04-02	[2]	CRAN (R 4.0.3)
lifecycle		1.0.0	2021-02-15	[1]	CRAN (R 4.0.3)
limma	*	3.46.0	2020-10-27	[1]	Bioconductor
locfit		1.5-9.4	2020-03-25	[1]	CRAN (R 4.0.2)
lubridate		1.7.10	2021-02-26	[1]	CRAN (R 4.0.3)
magrittr		2.0.1	2020-11-17	[1]	CRAN (R 4.0.2)
Matrix		1.3-4	2021-06-01	[1]	CRAN (R 4.0.3)
MatrixGenerics	*	1.2.1	2021-01-30	[1]	Bioconductor
matrixStats	*	0.59.0	2021-06-01	[1]	CRAN (R 4.0.3)
memoise		2.0.0	2021-01-26	[1]	CRAN (R 4.0.2)
mgcv	*	1.8-33	2020-08-27	[2]	CRAN (R 4.0.3)
modelr		0.1.8	2020-05-19	[1]	CRAN (R 4.0.2)
munsell		0.5.0	2018-06-12	[1]	CRAN (R 4.0.2)
nlme	*	3.1-152	2021-02-04	[1]	CRAN (R 4.0.3)
pbdZMQ		0.3-5	2021-02-10	[1]	CRAN (R 4.0.3)
pillar		1.6.1	2021-05-16	[1]	CRAN (R 4.0.3)
pkgconfig		2.0.3	2019-09-22	[1]	CRAN (R 4.0.2)
purrr	*	0.3.4	2020-04-17	[1]	CRAN (R 4.0.2)
R6		2.5.0	2020-10-28	[1]	CRAN (R 4.0.2)
Rcpp		1.0.7	2021-07-07	[1]	CRAN (R 4.0.3)
RCurl		1.98-1.3	2021-03-16	[1]	CRAN (R 4.0.3)
readr	*	1.4.0	2020-10-05	[1]	CRAN (R 4.0.2)
readxl		1.3.1	2019-03-13	[1]	CRAN (R 4.0.2)
repr		1.1.3	2021-01-21	[1]	CRAN (R 4.0.2)
reprex		2.0.0	2021-04-02	[1]	CRAN (R 4.0.3)
rlang		0.4.11	2021-04-30	[1]	CRAN (R 4.0.3)
RSQLite		2.2.7	2021-04-22	[1]	CRAN (R 4.0.3)
rstudioapi		0.13	2020-11-12	[1]	CRAN (R 4.0.2)
rvest		1.0.0	2021-03-09	[1]	CRAN (R 4.0.3)
S4Vectors	*	0.28.1	2020-12-09	[1]	Bioconductor
scales		1.1.1	2020-05-11	[1]	CRAN (R 4.0.2)
sessioninfo		1.1.1	2018-11-05	[1]	CRAN (R 4.0.2)
stringi		1.6.2	2021-05-17	[1]	CRAN (R 4.0.3)
stringr	*	1.4.0	2019-02-10	[1]	CRAN (R 4.0.2)
${\tt SummarizedExperiment}$	*	1.20.0	2020-10-27	[1]	Bioconductor
survival		3.2-7	2020-09-28	[2]	CRAN (R 4.0.3)

```
* 3.38.0
                                2020-10-27 [1] Bioconductor
sva
tibble
                     * 3.1.2
                                2021-05-16 [1] CRAN (R 4.0.3)
                     * 1.1.3
                                2021-03-03 [1] CRAN (R 4.0.3)
tidyr
tidyselect
                       1.1.1
                                2021-04-30 [1] CRAN (R 4.0.3)
tidyverse
                     * 1.3.1
                                2021-04-15 [1] CRAN (R 4.0.3)
                                2021-03-12 [1] CRAN (R 4.0.3)
utf8
                       1.2.1
uuid
                       0.1-4
                                2020-02-26 [1] CRAN (R 4.0.2)
                                2021-04-29 [1] CRAN (R 4.0.3)
                       0.3.8
vctrs
withr
                       2.4.2
                                2021-04-18 [1] CRAN (R 4.0.3)
XML
                       3.99-0.6 2021-03-16 [1] CRAN (R 4.0.3)
                                2020-04-23 [1] CRAN (R 4.0.2)
xm12
                       1.3.2
xtable
                       1.8-4
                                2019-04-21 [1] CRAN (R 4.0.2)
                                2020-10-27 [1] Bioconductor
XVector
                       0.30.0
                       1.36.0
                                2020-10-27 [1] Bioconductor
zlibbioc
```

- [1] /home/jbenja13/R/x86_64-pc-linux-gnu-library/4.0
- [2] /usr/lib/R/library