

# main

June 8, 2023

## 1 Examine overlaps with published data

```
[1]: import numpy as np
import pandas as pd
import session_info
from pyhere import here
```

### 1.1 Public si-eQTL analysis

```
[2]: shen = ["GDAP2", "AIM2", "SLAMF6", "RLF", "ATG4C", "FUT7",
            "TMEM218", "C11orf74", "RAB35", "TMEM5", "HNRNPK",
            "CDCA3", "ERCC5", "GJB6", "SNTB2", "SPNS3",
            "XAF1", "RBBP8", "RUFY4", "CA2", "RAPGEF1"]
print("Shen et al.:")
print(len(shen))

kukurba = ["NOD2", "WDR36", "BSCL2", "MAP7D3", "RHOXF1", "DNAH1"]
print("Kukurba et al.:")
print(len(kukurba))

yao = ["NOD2", "HLA-DRB5", "HLA-DRB5", "KIAA0586", "PPP2R5A",
       "TSNAXIP1", "MUT", "GRIK2", "C15orf37", "LIMA1", "IL6ST",
       "HCG8", "BLOC1S3", "NKX3-1", "CXorf23"]
print("Yao et al.:")
print(len(np.unique(yao)))
print("Total of Yao + Kukurba:")
len(set(yao) | set(kukurba))
```

Shen et al.:

21

Kukurba et al.:

6

Yao et al.:

14

Total of Yao + Kukurba:

[2]: 19

## 1.2 Load BrainSeq si-eQTL results

### 1.2.1 Interacting variant-gene pairs

```
[3]: bs0 = pd.read_csv("../_m/BrainSeq_sexGenotypes_4features_3regions.txt.gz",  
    ↪sep='\t')  
bs0["ensembl_gene_id"] = bs0.gene_id.str.replace("\\\\.*", "", regex=True)  
biomart = pd.read_csv("../_h/biomart.csv", index_col=0)  
bs = bs0.merge(biomart, on="ensembl_gene_id").drop_duplicates(subset="gene_id")  
print(bs.shape)  
bs.tail(2)
```

(692, 15)

```
[3]:
```

	region	gene_id	variant_id	gencode_id	\
8285	Caudate	ENSG00000270605.1	chr1:28102893:G:C	ENSG00000270605.1	
8286	DLPFC	ENSG00000187498.16	chr13:109650494:C:T	ENSG00000187498.16	

  

	gene_name	seqnames	start	end	lfsr	\
8285	ENSG00000270605	chr1	28239509	28241453	0.049873	
8286	COL4A1	chr13	110148963	110307157	0.048863	

  

	posterior_mean	feature_type	ensembl_gene_id	external_gene_name	\
8285	-0.261352	Gene	ENSG00000270605	AL353622.1	
8286	0.193807	Gene	ENSG00000187498	COL4A1	

  

	entrezgene	description
8285	NaN	NaN
8286	1282.0	collagen type IV alpha 1 chain [Source:HGNC Sy...

```
[4]: bs[(bs['external_gene_name'].isin(shen))].to_csv("siEQTL_Shen_comparison.csv",  
    ↪index=False)
```

```
[5]: bs[(bs['external_gene_name'].isin(kukurba))]
```

```
[5]: Empty DataFrame  
Columns: [region, gene_id, variant_id, gencode_id, gene_name, seqnames, start,  
end, lfsr, posterior_mean, feature_type, ensembl_gene_id, external_gene_name,  
entrezgene, description]  
Index: []
```

```
[6]: bs[(bs['external_gene_name'].isin(yao))]
```

```
[6]: Empty DataFrame
Columns: [region, gene_id, variant_id, gencode_id, gene_name, seqnames, start,
end, lfsr, posterior_mean, feature_type, ensembl_gene_id, external_gene_name,
entrezgene, description]
Index: []
```

```
[7]: bs[(bs['external_gene_name'].isin(shen+kukurba+yao))]
```

```
[7]:      region      gene_id      variant_id      gencode_id \
5787  Caudate  ENSG00000125703.15  chr1:63060301:G:A  ENSG00000125703.15
8130  Caudate  ENSG00000104267.10  chr8:84966439:A:T  ENSG00000104267.10

      gene_name seqnames      start      end      lfsr  posterior_mean \
5787      ATG4C      chr1  62784132  62865516  0.019839      0.094136
8130       CA2      chr8  85463968  85481493  0.045578      0.289557

      feature_type  ensembl_gene_id  external_gene_name  entrezgene \
5787          Gene  ENSG00000125703              ATG4C      84938.0
8130          Gene  ENSG00000104267              CA2       760.0

                                description
5787  autophagy related 4C cysteine peptidase [Sourc...
8130  carbonic anhydrase 2 [Source:HGNC Symbol;Acc:H...
```

### 1.3 GTEx comparison

```
[8]: gtex = pd.read_csv(here("input/public_results/gtex_results/_m",
                             "GTEx_Analysis_v8_sbeQTLs/GTEx_Analysis_v8_sbeQTLs.
                             ↪txt"),
                             sep='\t')
gtex.iloc[0:2, 0:10]
```

```
[8]:      ensembl_gene_id  hugo_gene_id      gene_type \
0  ENSG00000241860.6  RP11-34P13.13  processed_transcript
1  ENSG00000227232.5      WASH7P  unprocessed_pseudogene

      variant_id      rs_id      Tissue      maf \
0  chr1_14677_G_A_b38  rs201327123  Adipose_Subcutaneous  0.051635
1  chr1_64764_C_T_b38  rs769952832  Adipose_Subcutaneous  0.061102

      pval_nominal_sb  slope_sb  slope_se_sb
0      0.847114  0.055080      0.285537
1      0.316881  0.222928      0.222511
```

```
[9]: gtex.iloc[0:2, 10:14]
```

```
[9]:      numtested  pvals.corrected      qval  pval_nominal_f
0           1           0.847114  1.000000      0.022302
1           1           0.316881  0.981254      0.003978
```

```
[10]: ## qual threshold equal to number of published sb-eQTL
gtex[(gtex['qval'] < 0.25) & (gtex["Tissue"].str.contains("Brain"))]\
      .loc[:, ["ensembl_gene_id", "hugo_gene_id", "Tissue", "pvals.corrected",
↪ 'qval']].head(10)
```

```
[10]:      ensembl_gene_id  hugo_gene_id  \
62155  ENSG00000026025.15      VIM
116842  ENSG00000160818.16  GPATCH4
121904  ENSG00000141562.17  NARF
122123  ENSG00000267174.5  CTC-510F12.4

      Tissue  pvals.corrected      qval
62155      Brain_Amygdala      0.000004  0.012836
116842  Brain_Nucleus_accumbens_basal_ganglia      0.000088  0.198445
121904  Brain_Nucleus_accumbens_basal_ganglia      0.000056  0.198445
122123  Brain_Nucleus_accumbens_basal_ganglia      0.000083  0.198445
```

```
[11]: ## qual threshold equal to number of published sb-eQTL
gtex[(gtex['qval'] < 0.25) & (gtex["Tissue"].str.contains("Whole"))]\
      .loc[:, ["ensembl_gene_id", "hugo_gene_id", "Tissue", "pvals.corrected",
↪ 'qval']].head(10)
```

```
[11]:      ensembl_gene_id  hugo_gene_id      Tissue  pvals.corrected  \
362961  ENSG00000221571.3  RNU6ATAC35P  Whole_Blood      0.000039
365043  ENSG00000196743.8      GM2A  Whole_Blood      0.000011
367164  ENSG00000148459.15  PDSS1  Whole_Blood      0.000027

      qval
362961  0.139762
365043  0.116825
367164  0.139762
```

```
[12]: gtex_sig = gtex[(gtex['qval'] < 0.25)]
gtex_sig.shape
```

```
[12]: (369, 22)
```

```
[13]: gtex_sig.head(10)
```

```
[13]:      ensembl_gene_id  hugo_gene_id      gene_type  \
1096  ENSG00000076356.6  PLXNA2  protein_coding
5262  ENSG00000170632.13  ARMC10  protein_coding
5644  ENSG00000120907.17  ADRA1A  protein_coding
```

6414	ENSG00000136830.11	FAM129B	protein_coding
7220	ENSG00000166787.3	SAA3P	transcribed_unprocessed_pseudogene
8540	ENSG00000183463.5	URAD	protein_coding
9191	ENSG00000282651.2	IGHV5-10-1	IG_V_gene
14611	ENSG00000143933.16	CALM2	protein_coding
15082	ENSG00000144410.4	CPD	protein_coding
17452	ENSG00000211698.2	TRGV4	TR_V_gene

	variant_id	rs_id	Tissue \
1096	chr1_208030492_G_A_b38	rs3811383	Adipose_Subcutaneous
5262	chr7_103076937_C_T_b38	rs6958836	Adipose_Subcutaneous
5644	chr8_26839198_G_A_b38	rs117380715	Adipose_Subcutaneous
6414	chr9_127584339_G_A_b38	rs10739693	Adipose_Subcutaneous
7220	chr11_18269355_T_C_b38	rs34068567	Adipose_Subcutaneous
8540	chr13_27990205_T_A_b38	rs7335293	Adipose_Subcutaneous
9191	chr14_106114510_A_G_b38	rs4573838	Adipose_Subcutaneous
14611	chr2_46225349_C_T_b38	rs12477148	Adipose_Visceral_Omentum
15082	chr2_206822186_C_T_b38	rs12470278	Adipose_Visceral_Omentum
17452	chr7_38361995_A_C_b38	rs10233345	Adipose_Visceral_Omentum

	maf	pval_nominal_sb	slope_sb	slope_se_sb	...	qval \
1096	0.123924	5.391600e-05	0.338278	0.083064	...	0.121068
5262	0.169535	5.011130e-05	0.357403	0.087384	...	0.192900
5644	0.216867	1.045890e-05	-0.323552	0.072676	...	0.084548
6414	0.304647	7.387010e-07	-0.283660	0.056579	...	0.004976
7220	0.278830	2.207290e-05	0.323030	0.075427	...	0.074347
8540	0.500000	9.078700e-09	-0.444892	0.076123	...	0.000122
9191	0.419105	2.025150e-05	-0.406760	0.094541	...	0.074347
14611	0.072495	4.497930e-05	-0.480557	0.116471	...	0.161955
15082	0.097015	3.204120e-05	0.682291	0.162191	...	0.115370
17452	0.335821	6.438100e-05	0.427491	0.105837	...	0.139089

	pval_nominal_f	slope_f	slope_se_f	pval_nominal_m	slope_m \
1096	1.718880e-08	0.456729	0.075705	9.155700e-01	0.009739
5262	4.933240e-01	-0.054539	0.079379	3.219220e-07	-0.429800
5644	4.637410e-18	-0.779707	0.076596	3.976660e-10	-0.469672
6414	1.978000e-06	-0.333315	0.066772	1.653380e-01	-0.082625
7220	6.409400e-08	0.453034	0.078725	3.138000e-01	0.063002
8540	9.982650e-21	-0.887723	0.078738	1.892290e-09	-0.457733
9191	5.248710e-12	-0.682629	0.089412	2.805060e-03	-0.289091
14611	4.747150e-04	-0.491287	0.134732	9.165740e-01	0.013043
15082	1.165430e-01	0.280837	0.176978	4.431060e-06	-0.558002
17452	7.011230e-06	-0.481758	0.100091	8.857670e-15	-1.068840

	slope_se_m	pval_nominal	slope	slope_se
1096	0.091682	2.747400e-05	0.171830	0.040604
5262	0.079545	8.797530e-08	-0.216374	0.039857

5644	0.069091	5.637370e-52	-0.568916	0.033334
6414	0.059205	1.393160e-08	-0.168762	0.029260
7220	0.062292	2.433600e-08	0.211910	0.037395
8540	0.070571	3.077310e-53	-0.640604	0.036976
9191	0.094806	3.458420e-21	-0.445408	0.045073
14611	0.124116	2.197750e-05	-0.246023	0.057281
15082	0.113158	7.896400e-06	-0.320288	0.070745
17452	0.112111	1.630390e-49	-0.838766	0.049090

[10 rows x 22 columns]

### 1.3.1 mashr

```
[14]: gtex_overlap = bs[(bs['gene_id'].isin(gtex_sig.ensembl_gene_id))].
      ↪drop_duplicates()
      print(gtex_overlap.shape)
      gtex_overlap
```

(2, 15)

```
[14]:
```

	region	gene_id	variant_id	gencode_id	\
4943	Caudate	ENSG000000272977.1	chr22:25059120:A:C	ENSG000000272977.1	
8285	Caudate	ENSG000000270605.1	chr1:28102893:G:C	ENSG000000270605.1	

  

	gene_name	seqnames	start	end	lfsr	posterior_mean	\
4943	ENSG000000272977	chr22	25476218	25479971	0.011928	0.323847	
8285	ENSG000000270605	chr1	28239509	28241453	0.049873	-0.261352	

  

	feature_type	ensembl_gene_id	external_gene_name	entrezgene	description
4943	Gene	ENSG000000272977	AL008721.2	NaN	NaN
8285	Gene	ENSG000000270605	AL353622.1	NaN	NaN

```
[15]: gtex_overlap.shape[0]/bs.shape[0] * 100
```

```
[15]: 0.2890173410404624
```

```
[16]: gtex_sig[(gtex_sig['ensembl_gene_id'].isin(bs.gene_id))]
```

```
[16]:
```

	ensembl_gene_id	hugo_gene_id	gene_type	\
297207	ENSG000000270605.1	RP5-1092A3.4	antisense	
338770	ENSG000000272977.1	CTA-390C10.10	sense_intronic	

  

	variant_id	rs_id	Tissue	\
297207	chr1_28223937_C_T_b38	rs481640	Skin_Not_Sun_Exposed_Suprapubic	
338770	chr22_25459662_G_A_b38	rs6004655	Spleen	

	maf	pval_nominal_sb	slope_sb	slope_se_sb	...	qval	\
297207	0.323985	0.000237	-0.241524	0.065161	...	0.208195	
338770	0.167401	0.000038	0.413128	0.097856	...	0.212883	

  

	pval_nominal_f	slope_f	slope_se_f	pval_nominal_m	slope_m	\
297207	4.046590e-21	-0.760755	0.063142	6.395880e-13	-0.599143	
338770	7.931210e-14	-1.129030	0.107327	5.155260e-16	-1.276590	

  

	slope_se_m	pval_nominal	slope	slope_se
297207	0.07247	9.917080e-60	-0.591873	0.031047
338770	0.10469	4.896640e-51	-1.193670	0.056659

[2 rows x 22 columns]

```
[17]: gtex_sig[(gtex_sig['ensembl_gene_id'].isin(bs.gene_id))]\
      .to_csv("siEQTL_gtex_comparison.csv", index=False)
```

## 2 Session information

```
[18]: session_info.show()
```

```
[18]: <IPython.core.display.HTML object>
```