

# main

September 1, 2021

## 1 Plotting eQTLs, increase font sizes

### 1.0.1 Kynon Jade Benjamin and Apuã Paquola

```
[1]: import re
import functools
import subprocess
import numpy as np
import pandas as pd
from plotnine import *
from pandas_plink import read_plink
from warnings import filterwarnings
from matplotlib.cbook import mplDeprecation

filterwarnings("ignore",category=mplDeprecation)
filterwarnings('ignore', category=UserWarning, module='plotnine.*')
filterwarnings('ignore', category=DeprecationWarning, module='plotnine.*')
```

### 1.1 Configuration

```
[2]: tissue = "dlpfc"; feature = "genes"
config = {
    'biomart_file': '../_h/biomart.csv',
    'residual_expression_file': "../../../../../prep_eqtl_analysis/%s/%s/
↳covariates/" % (tissue, feature)+\
    "residualized_expression/_m/%s_residualized_expression.csv" % feature,
    'phenotype_file': '/ceph/projects/v4_phase3_paper/inputs/phenotypes/_m/
↳merged_phenotypes.csv',
    'plink_file_prefix': '/ceph/projects/v4_phase3_paper/inputs/genotypes/_m/
↳LIBD_Brain_TopMed',
    'eqtl_output_file': '../../../../../summary_table/_m/
↳Brainseq_sex_interacting_4features_3regions.eFeatures.txt.gz',
    'gwas_snp_file': '/ceph/projects/v4_phase3_paper/inputs/sz_gwas/pgc2_clozuk/
↳map_phase3/_m/libd_hg38_pgc2sz_snps_p5e_minus8.tsv'
}
```

## 1.2 Functions

### 1.2.1 Expression functions

```
[3]: @functools.lru_cache()
def tissue_map(tissue):
    return {"caudate": "Caudate", "dlpfc": "DLPFC",
            "hippocampus": "Hippocampus"}[tissue]

@functools.lru_cache()
def feature_map(feature):
    return {"genes": "Gene", "transcripts": "Transcript",
            "exons": "Exon", "junctions": "Junction"}[feature]

@functools.lru_cache()
def get_biomart_df():
    biomart = pd.read_csv(config['biomart_file'], index_col=0)
    biomart['description'] = biomart['description'].str.replace('\[Source.
→*$', '', regex=True)
    return biomart

@functools.lru_cache()
def get_residual_expression_df():
    return pd.read_csv(config['residual_expression_file'], index_col=0).
→transpose()

@functools.lru_cache()
def get_pheno_df():
    return pd.read_csv(config['phenotype_file']).set_index("BrNum").loc[:,
→["RNum", "Sex", "Dx"]]

@functools.lru_cache()
def get_expression_and_pheno_df():
    return pd.merge(get_pheno_df(), get_residual_expression_df(),
                    left_index=True, right_index=True)

@functools.lru_cache()
def get_gene_id_df():
    return pd.DataFrame({'gene_id': get_residual_expression_df().columns,
                        'ensembl_gene_id': get_residual_expression_df().
→columns.str.replace('\.+$', '', regex=True)})
```

```

@functools.lru_cache()
def gene_info_from_symbol(gene_symbol):
    return
    ↳get_biomart_df()[get_biomart_df()['external_gene_name']==gene_symbol]\
        .merge(get_gene_id_df(), on='ensembl_gene_id', how='left')

@functools.lru_cache()
def gene_id_from_symbol(gene_symbol):
    df = gene_info_from_symbol(gene_symbol)
    assert df.shape[0] == 1
    return df[['gene_id']].iloc[0].values[0]

```

## 1.2.2 Genotype and eQTL functions

```

[4]: def letter_snp(number, a0, a1):
    '''
    Example:
    letter_snp(0, 'A', 'G') is 'AA'
    letter_snp(1, 'A', 'G') is 'AG'
    letter_snp(2, 'A', 'G') is 'GG'
    '''
    if np.isnan(number):
        return np.nan
    if len(a0)==1 and len(a1)==1:
        sep = ''
    else:
        sep = ' '
    return sep.join(sorted([a0]*int(number) + [a1]*(2-int(number)))))

@functools.lru_cache()
def get_plink_tuple():
    '''
    Usage: (bim, fam, bed) = get_plink_tuple()
    '''
    return read_plink(config['plink_file_prefix'])

@functools.lru_cache()
def get_eFeature_df():
    eqtl_df = pd.read_csv(config["eqtl_output_file"], sep='\t')
    return eqtl_df[(eqtl_df["Type"] == feature_map(feature)) &
                    (eqtl_df["Tissue"] == tissue_map(tissue))]

```

```

@functools.lru_cache()
def get_gwas_snps():
    return pd.read_csv(config['gwas_snp_file'], sep='\t', index_col=0)

@functools.lru_cache()
def get_risk_allele(snp_id):
    gwas_snp = get_gwas_snp(snp_id)
    if gwas_snp['OR'].iloc[0] > 1:
        ra = gwas_snp['A1'].iloc[0]
    else:
        ra = gwas_snp['A2'].iloc[0]
    return ra

@functools.lru_cache()
def get_snp_df(snp_id):
    """
    Returns a dataframe containing the genotype on snp snp_id.
    The allele count is the same as in the plink files.

    Example:
    get_snp_df('rs653953').head(5)

    rs653953_num rs653953_letter rs653953
    Br5168      0                GG    0\nGG
    Br2582      1                AG    1\nAG
    Br2378      1                AG    1\nAG
    Br5155      2                AA    2\nAA
    Br5182      2                AA    2\nAA
    """
    (bim, fam, bed) = get_plink_tuple()
    brain_ids = list(set(get_expression_and_pheno_df().index).
    ↪ intersection(set(fam['fid'])))
    snp_info = bim[bim['snp']==snp_id]
    snp_pos = snp_info.iloc[0]['i']
    fam_pos = list(fam.drop_duplicates(subset="fid").set_index('fid').
    ↪ loc[brain_ids]['i'])
    dfsnp = (pd.DataFrame(bed[[snp_pos]].compute()[:,fam_pos],
                          columns=brain_ids, index=[snp_id + '_num'])
              .transpose().dropna())
    my_letter_snp = functools.partial(letter_snp, a0=snp_info.iloc[0]['a0'],
    ↪ a1=snp_info.iloc[0]['a1'])
    dfsnp[[snp_id + '_num']] = 2 - dfsnp[[snp_id + '_num']].astype('int')
    dfsnp[snp_id + '_letter'] = dfsnp[snp_id + '_num'].apply(my_letter_snp)
    dfsnp[snp_id] = (dfsnp[snp_id + '_num'].astype('str') + '\n' +
                     dfsnp[snp_id + '_letter'].astype('str')).astype('category')

```

```

return dfsnp

@functools.lru_cache()
def get_gwas_ordered_snp_df(snp_id):
    '''
    Returns a dataframe containing the genotype on snp snp_id.
    The allele count is the number of risk alleles according to GWAS.

    Example:
    get_gwas_ordered_snp_df('rs653953').head(5)

           rs653953_num rs653953_letter rs653953
    Br5168             2             GG      2\nGG
    Br2582             1             AG      1\nAG
    Br2378             1             AG      1\nAG
    Br5155             0             AA      0\nAA
    Br5182             0             AA      0\nAA
    '''
    pgc = get_gwas_snps()
    dfsnp = get_snp_df(snp_id).copy()
    gwas_snp = get_gwas_snp(snp_id)
    if gwas_snp['pgc2_a1_same_as_our_counted'].iloc[0]:
        if gwas_snp['OR'].iloc[0] > 1:
            pass
        else:
            dfsnp[[snp_id + '_num']] = 2 - dfsnp[[snp_id + '_num']]
    else:
        if gwas_snp['OR'].iloc[0] > 1:
            dfsnp[[snp_id + '_num']] = 2 - dfsnp[[snp_id + '_num']]
        else:
            pass
    dfsnp[snp_id] = (dfsnp[snp_id + '_num'].astype('str') + '\n' +
                    dfsnp[snp_id + '_letter'].astype('str')).astype('category')
    return dfsnp

```

### 1.2.3 Plotting functions

```

[5]: def get_snp_gene_pheno_df(snp_id, gene_id, snp_df_func):
    pheno_columns = list(get_pheno_df().columns)
    expr_df = get_expression_and_pheno_df()[pheno_columns + [gene_id]]
    snp_df = snp_df_func(snp_id)
    return expr_df.merge(snp_df, left_index=True, right_index=True)

def simple_snp_expression_plot_impl(snp_id, gene_id, snp_df_func):
    df = get_snp_gene_pheno_df(snp_id, gene_id, snp_df_func)

```

```

y0 = df[gene_id].quantile(.01) - 0.26
y1 = df[gene_id].quantile(.99) + 0.26
p = ggplot(df, aes(x=snp_id, y=gene_id, fill='Sex')) \
+ geom_boxplot(alpha=0.4, outlier_alpha=0) \
+ geom_jitter(position=position_jitterdodge(jitter_width=0.25),
              stroke=0, alpha=0.6) \
+ ylim(y0, y1) \
+ theme_bw(base_size=15) \
+ theme(panel_grid=element_blank(),
        axis_title=element_text(face="bold"))
return p

def simple_snp_expression_plot(snp_id, gene_id):
    return simple_snp_expression_plot_impl(snp_id, gene_id, get_snp_df)

def simple_gwas_ordered_snp_expression_plot(snp_id, gene_id):
    return simple_snp_expression_plot_impl(snp_id, gene_id,
    ↪get_gwas_ordered_snp_df)

def get_gene_symbol(gene_id, biomart=get_biomart_df()):
    ensge = re.sub('\.+$', '', gene_id)
    ggg = biomart[biomart['ensembl_gene_id']==ensge]
    if ggg.shape[0]==0:
        return '', ''
    gs = ggg['external_gene_name'].values[0]
    de = ggg['description'].values[0]
    if type(de)!=str:
        de = ''
    de = re.sub('\[Source:.*$', '', de)
    return gs, de

def get_gwas_snp(snp_id):
    gwas = get_gwas_snps()
    r = gwas[gwas['our_snp_id']==snp_id]
    assert len(r) == 1
    return r

def gwas_annotation(snp_id):
    return 'SZ GWAS pvalue: %.1e' % get_gwas_snp(snp_id).iloc[0]['P']

def eqtl_annotation(snp_id, gene_id):

```

```

eqtl_df = get_eFeature_df()
r = eqtl_df[(eqtl_df['variant_id']==snp_id) & (eqtl_df['gene_id']==gene_id)]
assert len(r)==1
return 'eQTL adjusted p-value: %.1e' % r.iloc[0]['BF']

def risk_allele_annotation(snp_id):
    return 'SZ risk allele: %s' % get_risk_allele(snp_id)

def annotated_eqtl_plot(snp_id, gene_id):
    p = simple_snp_expression_plot(snp_id, gene_id)
    gene_symbol, gene_description = get_gene_symbol(gene_id)
    title = "\n".join([gene_symbol,
                       eqtl_annotation(snp_id, gene_id)
                      ])
    p += ggtitle(title) + ylab('Residualized Expression')
    return p

def gwas_annotated_eqtl_plot(snp_id, gene_id):
    p = simple_gwas_ordered_snp_expression_plot(snp_id, gene_id)
    gene_symbol, gene_description = get_gene_symbol(gene_id)
    title = "\n".join([gene_symbol,
                       eqtl_annotation(snp_id, gene_id),
                       gwas_annotation(snp_id),
                       risk_allele_annotation(snp_id)
                      ])
    p += ggtitle(title) + ylab('Residualized Expression')
    return p

def save_plot(p, fn):
    for ext in ['png', 'pdf', 'svg']:
        p.save(fn + '.' + ext)

```

## 1.3 Plot eQTLs

### 1.3.1 DRD2

```
[6]: get_eFeature_df()[get_eFeature_df()["gene_id"] == gene_id_from_symbol('DRD2')]
```

```
[6]: Empty DataFrame
Columns: [variant_id, gene_id, gencodeID, slope, statistic, pval_nominal, BF,
eigenMT_BH, TESTS, Type, Tissue]
Index: []
```

### 1.3.2 Top 5 eQTLs

```
[7]: eqtl_df = get_eFeature_df()
eqtl_df.head()
```

```
[7]:
```

	variant_id	gene_id	gencodeID	\
29020	chr1:196175713:C:T	ENSG00000000971.15	ENSG00000000971.15	
29021	chrX:65756506:T:A	ENSG00000001497.16	ENSG00000001497.16	
29022	chr4:11426453:A:AG	ENSG00000002587.9	ENSG00000002587.9	
29023	chr2:200747038:C:T	ENSG00000003400.14	ENSG00000003400.14	
29024	chr1:22575250:CTTATTA:C	ENSG00000004487.15	ENSG00000004487.15	

	slope	statistic	pval_nominal	BF	eigenMT_BH	TESTS	Type	\
29020	0.380554	14.718097	0.000132	0.020039	0.586787	152	Gene	
29021	-0.454968	-5.143391	0.000160	0.011876	0.560779	74	Gene	
29022	-0.732540	-8.886183	0.000042	0.027774	0.612625	667	Gene	
29023	0.455473	10.846644	0.000148	0.044193	0.656748	298	Gene	
29024	0.432105	11.118354	0.000077	0.036023	0.646335	466	Gene	

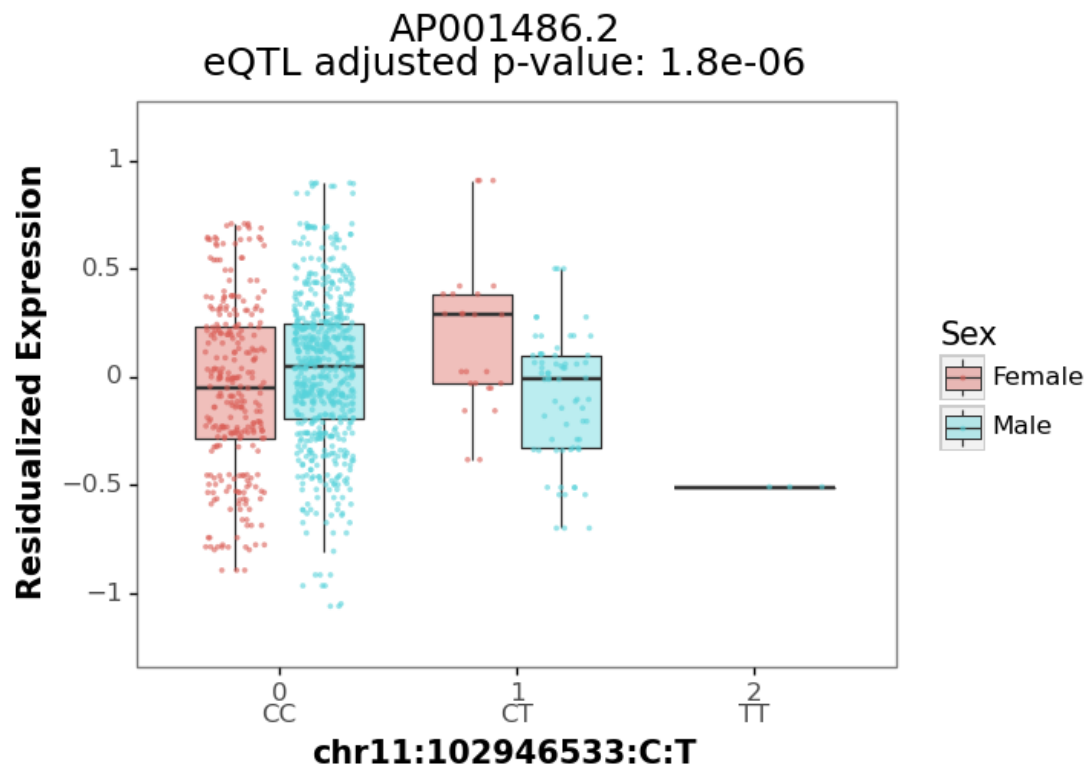
	Tissue
29020	DLPFC
29021	DLPFC
29022	DLPFC
29023	DLPFC
29024	DLPFC

```
[8]: top_5 = eqtl_df.sort_values('pval_nominal').reset_index(drop=True).head(5)
for x in top_5.itertuples():
    filename = "top_%d_eqtl_%s" % (x.Index, tissue)
    p = annotated_eqtl_plot(x.variant_id, x.gene_id)
    print(filename, x.Index, x.variant_id, x.gene_id)
    print(p)
    save_plot(p, filename)
```

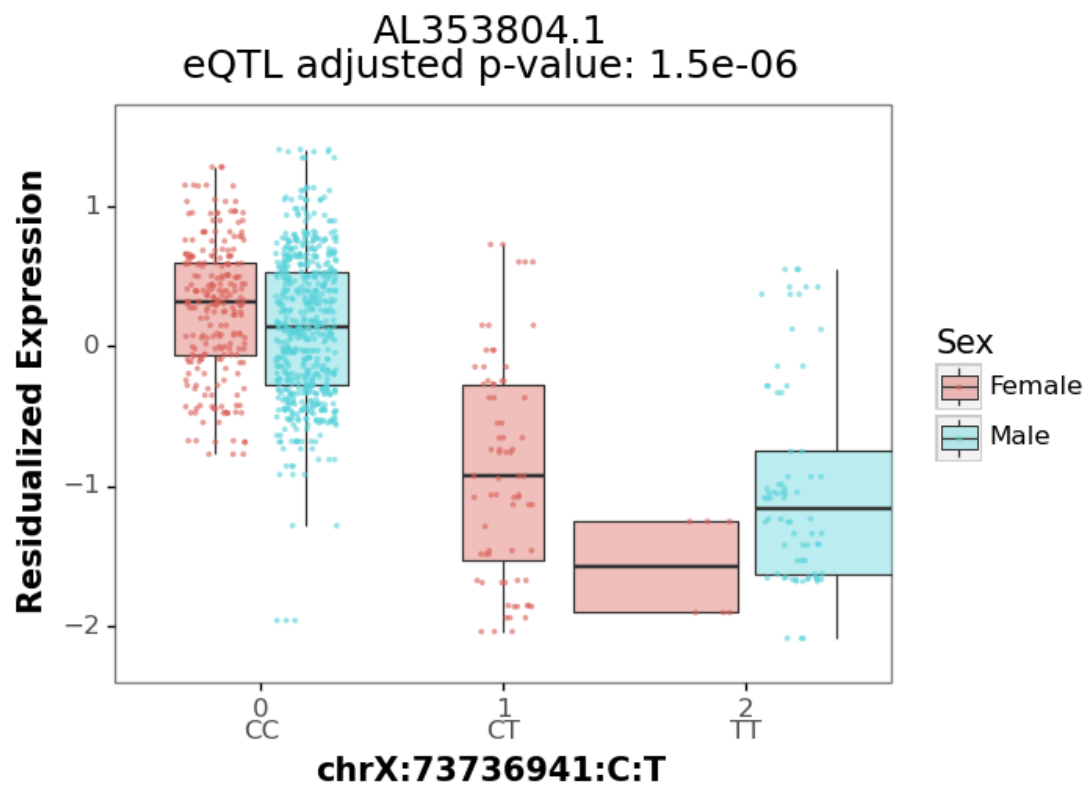
Mapping files: 100%| | 3/3 [00:26<00:00, 8.76s/it]

top\_0\_eqtl\_dlpfc 0 chr11:102946533:C:T ENSG00000260966.1

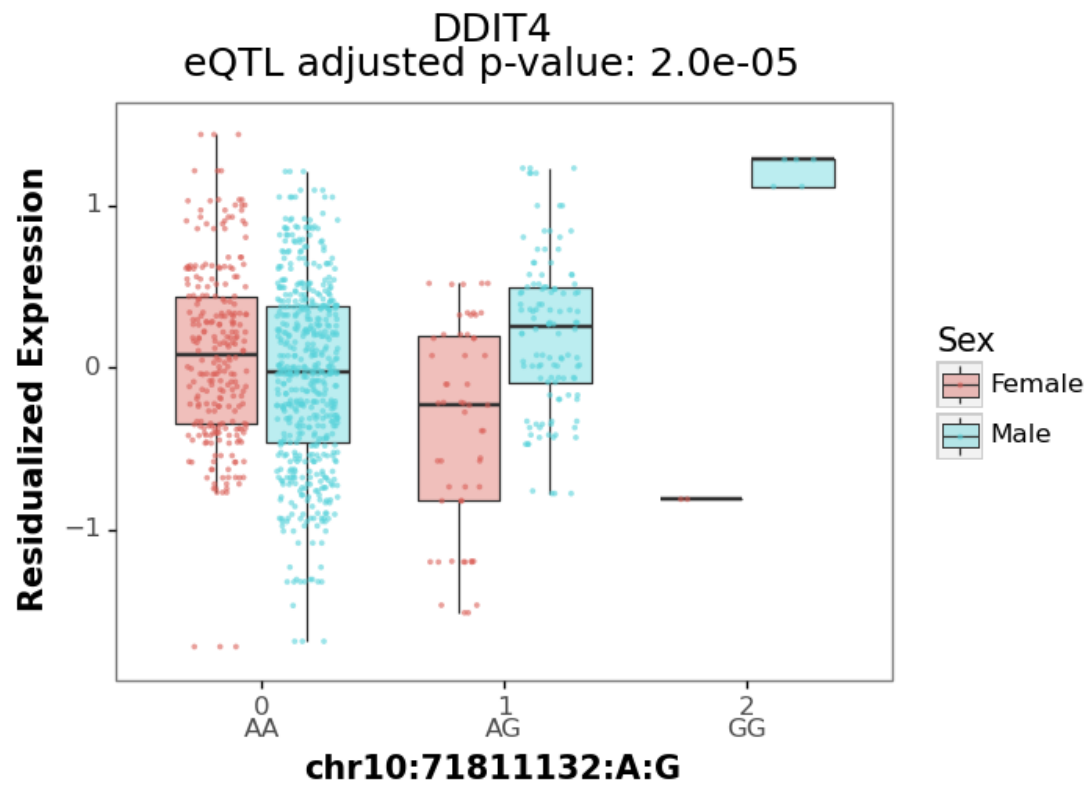




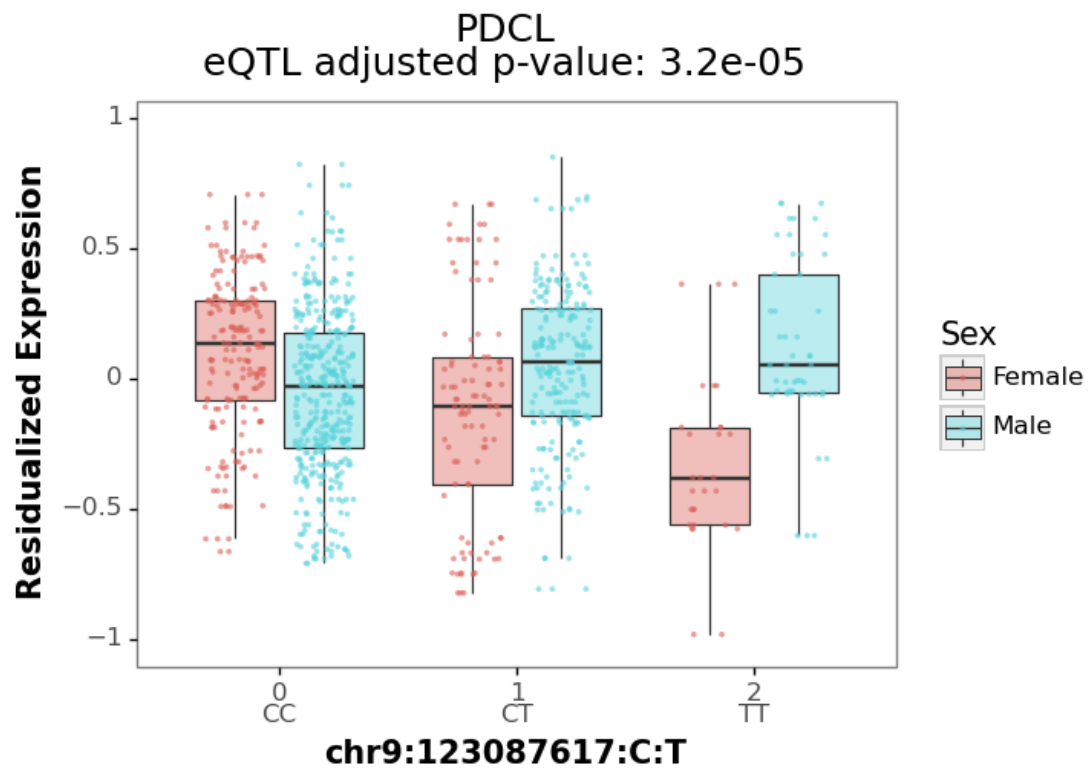
```
<ggplot: (8739662760394)>
top_1_eqtl_dlpfc 1 chrX:73736941:C:T ENSG00000228906.1
```



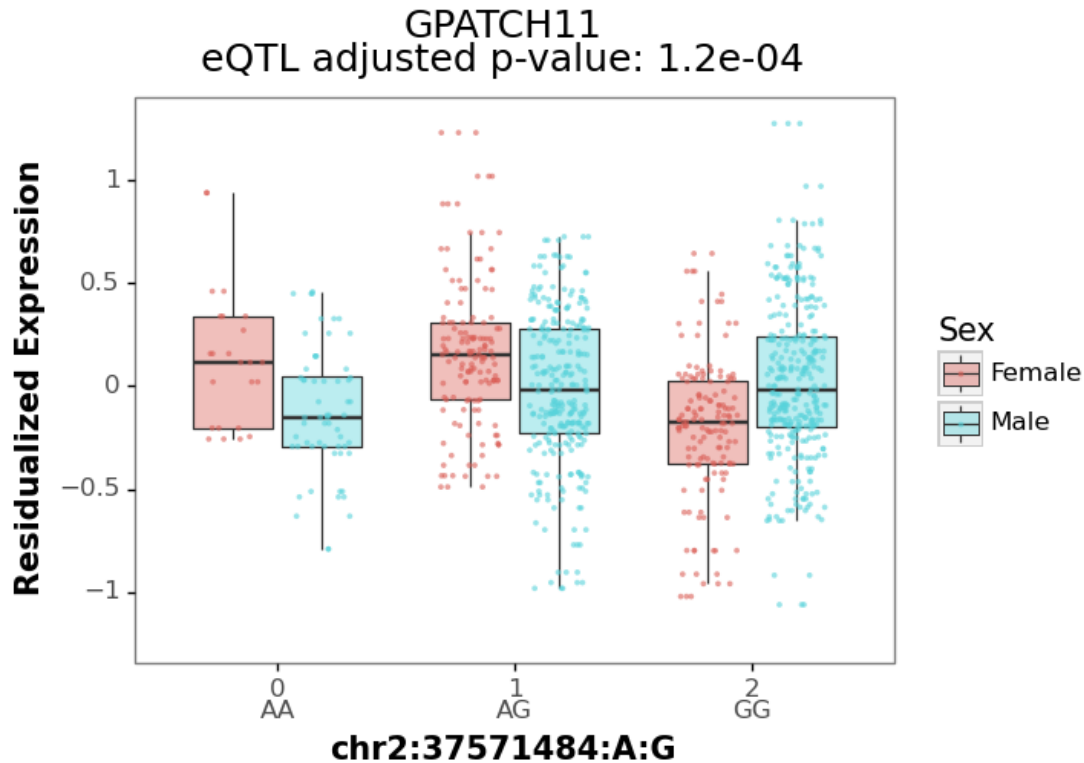
```
<ggplot: (8739662559507)>
top_2_eqtl_dlpfc 2 chr10:71811132:A:G ENSG00000168209.4
```



```
<ggplot: (8739636181008)>
top_3_eqtl_dlpfc 3 chr9:123087617:C:T ENSG00000136940.13
```



```
<ggplot: (8739636066545)>
top_4_eqtl_dlpfc 4 chr2:37571484:A:G ENSG00000152133.14
```

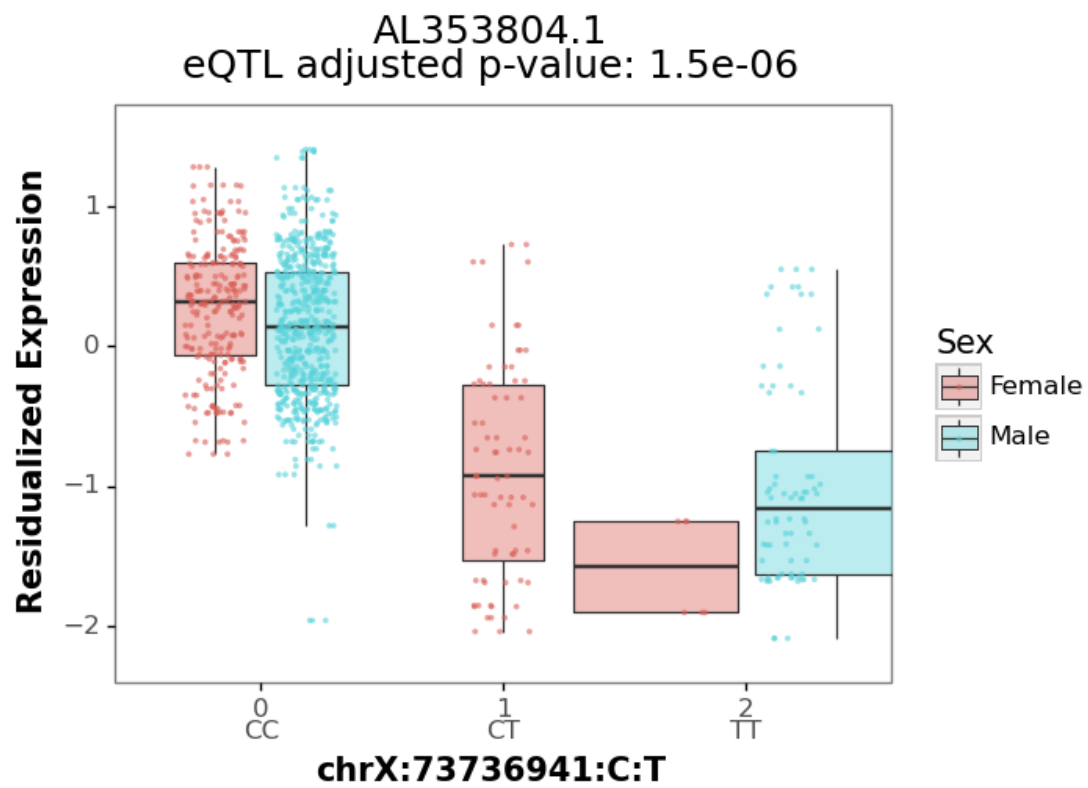


<ggplot: (8739635715780)>

### 1.3.3 Top 5 X-linked genes

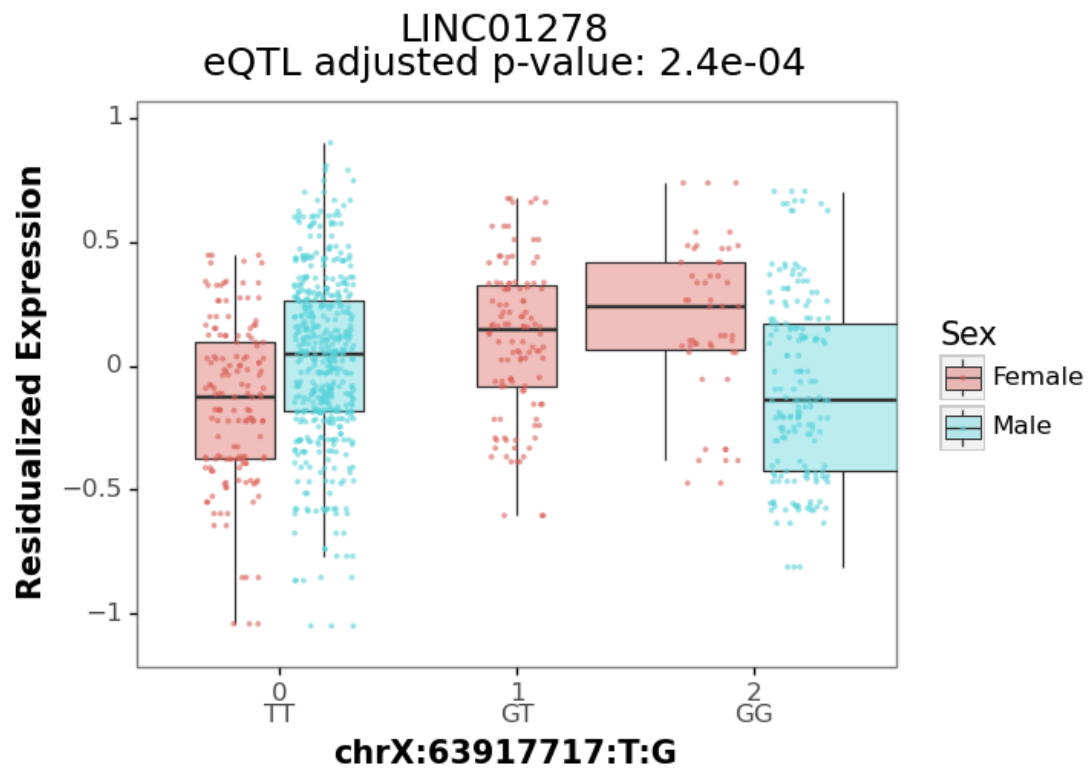
```
[9]: top_5_x = eqtl_df[eqtl_df['variant_id'].str.contains("chrX")].
      ↪sort_values("pval_nominal").reset_index(drop=True).head(5)
for x in top_5_x.itertuples():
    filename = "top_%d_eqtl_xlinked_%s" % (x.Index, tissue)
    p = annotated_eqtl_plot(x.variant_id, x.gene_id)
    print(filename, x.Index, x.variant_id, x.gene_id)
    print(p)
    save_plot(p, filename)
```

top\_0\_eqtl\_xlinked\_dlpfc 0 chrX:73736941:C:T ENSG00000228906.1

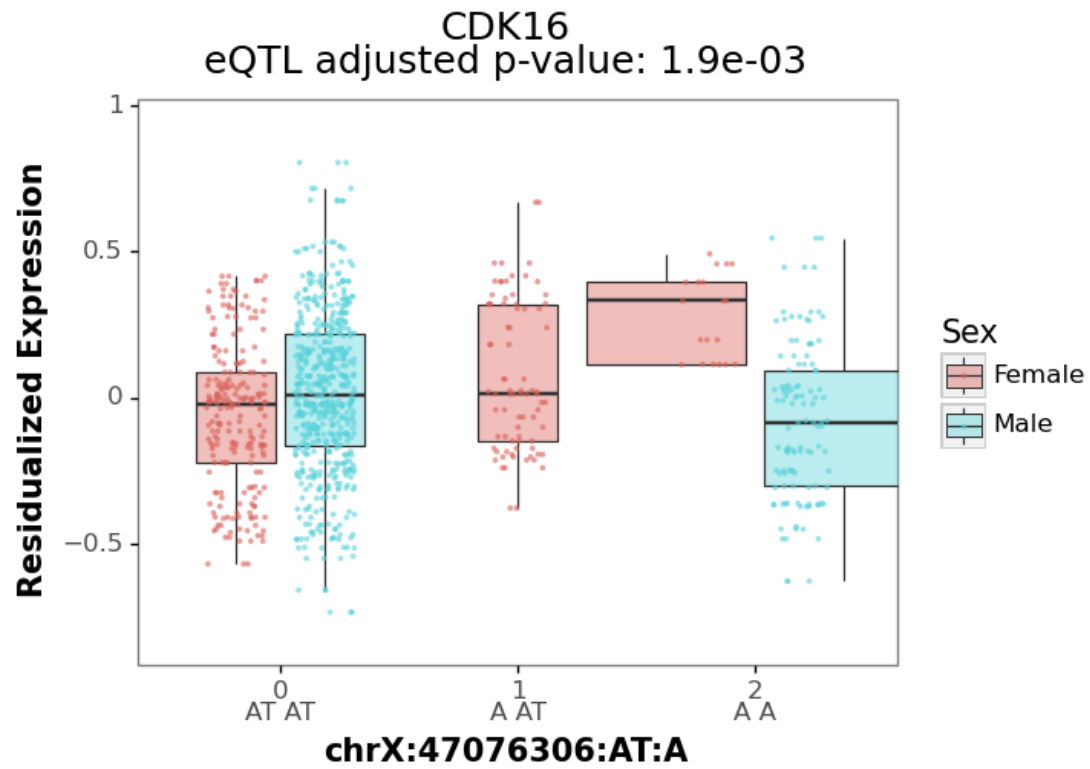


```
<ggplot: (8739764125609)>
```

```
top_1_eqtl_xlinked_dlpfc 1 chrX:63917717:T:G ENSG00000235437.7
```

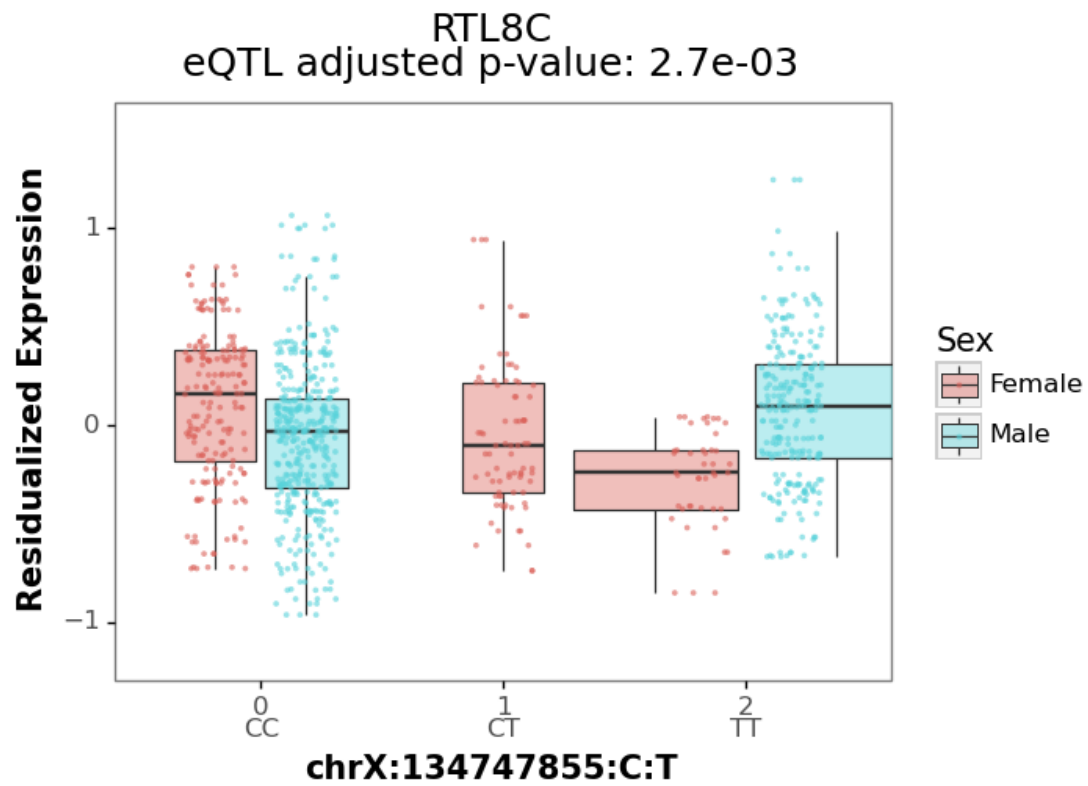


```
<ggplot: (8739662709242)>
top_2_eqtl_xlinked_dlpfc 2 chrX:47076306:AT:A ENSG00000102225.15
```

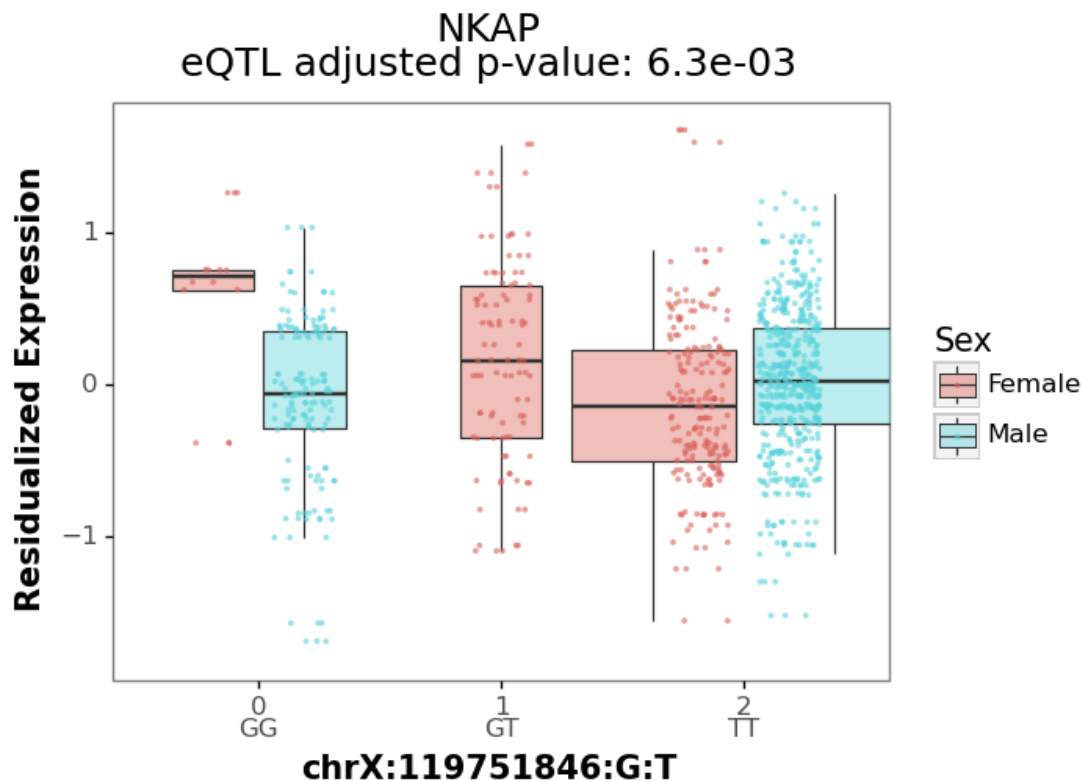


```
<ggplot: (8739662717992)>
top_3_eqtl_xlinked_dlpfc 3 chrX:134747855:C:T ENSG00000134590.13
```





```
<ggplot: (8739662229450)>
top_4_eqtl_xlinked_dlpfc 4 chrX:119751846:G:T ENSG00000101882.9
```



```
<ggplot: (8739663360139)>
```

### 1.3.4 Top 5 eQTL with GWAS significant index SNP

```
[10]: gwas_eqtl_df = eqtl_df.merge(get_gwas_snps(), left_on = 'variant_id',
                                   right_on = 'our_snp_id', suffixes=['', '_gwas'])
print(gwas_eqtl_df.shape)
gwas_eqtl_df.head()
```

```
(6, 33)
```

```
[10]:
```

	variant_id	gene_id	gencodeID	slope \
0	chr10:102875591:C:T	ENSG00000156374.14	ENSG00000156374.14	0.408080
1	chr6:31888293:G:A	ENSG00000204371.11	ENSG00000204371.11	0.360059
2	chr6:32225442:C:T	ENSG00000243649.8	ENSG00000243649.8	-0.664658
3	chr6:31494358:G:A	ENSG00000244731.7	ENSG00000244731.7	-0.762762
4	chr8:110618046:TG:T	ENSG00000254241.1	ENSG00000254241.1	-0.274868

	statistic	pval_nominal	BF	eigenMT_BH	TESTS	Type	...	A2 \
0	10.794880	0.000149	0.047714	0.666100	320	Gene	...	T
1	11.418709	0.000004	0.001779	0.434507	502	Gene	...	A
2	-14.774041	0.000015	0.008005	0.535673	525	Gene	...	T

3	-11.406318	0.000018	0.009065	0.539284	498	Gene ...	A
4	-10.339230	0.000113	0.019356	0.585158	171	Gene ...	TG

	OR	SE	P	hg19chrc	hg38chrc	hg38pos	\
0	1.08930	0.011207	2.270000e-14	chr10	chr10	102875591	
1	0.92611	0.010290	8.620000e-14	chr6	chr6	31888293	
2	1.11790	0.013264	4.480000e-17	chr6	chr6	32225442	
3	1.18310	0.015322	5.000000e-28	chr6	chr6	31494358	
4	1.08730	0.012600	3.060000e-11	chr8	chr8	110618046	

	pgc2_a1_same_as_our_counted	rsid	is_index_snp
0	False	rs12765002	False
1	False	rs486416	False
2	False	rs3130295	False
3	False	rs3130923	False
4	True	rs35771435	False

[5 rows x 33 columns]

```
[11]: top_gwas_eqtl_df = gwas_eqtl_df[(gwas_eqtl_df['is_index_snp'])].
      ↪sort_values(['BF', 'P'])
      print(top_gwas_eqtl_df.shape)
      top_gwas_eqtl_df.head()
```

(0, 33)

```
[11]: Empty DataFrame
Columns: [variant_id, gene_id, gencodeID, slope, statistic, pval_nominal, BF,
eigenMT_BH, TESTS, Type, Tissue, chrN, our_snp_id, cm, pos, our_counted,
our_alt, chrom, SNP, Freq.A1, CHR, BP, A1, A2, OR, SE, P, hg19chrc, hg38chrc,
hg38pos, pgc2_a1_same_as_our_counted, rsid, is_index_snp]
Index: []
```

[0 rows x 33 columns]

```
[12]: top_gwas_eqtl_df = gwas_eqtl_df.sort_values(['BF', 'P']).reset_index(drop=True)
      print(top_gwas_eqtl_df.shape)
      top_gwas_eqtl_df.head(10)
```

(6, 33)

	variant_id	gene_id	gencodeID	slope	\
0	chr6:31888293:G:A	ENSG00000204371.11	ENSG00000204371.11	0.360059	
1	chr6:32225442:C:T	ENSG00000243649.8	ENSG00000243649.8	-0.664658	
2	chr6:31494358:G:A	ENSG00000244731.7	ENSG00000244731.7	-0.762762	
3	chr6:26573403:C:G	ENSG00000274290.2	ENSG00000274290.2	-0.553277	
4	chr8:110618046:TG:T	ENSG00000254241.1	ENSG00000254241.1	-0.274868	
5	chr10:102875591:C:T	ENSG00000156374.14	ENSG00000156374.14	0.408080	

	statistic	pval_nominal	BF	eigenMT_BH	TESTS	Type	...	A2	\
0	11.418709	0.000004	0.001779	0.434507	502	Gene	...	A	
1	-14.774041	0.000015	0.008005	0.535673	525	Gene	...	T	
2	-11.406318	0.000018	0.009065	0.539284	498	Gene	...	A	
3	-10.810298	0.000046	0.012635	0.560779	276	Gene	...	G	
4	-10.339230	0.000113	0.019356	0.585158	171	Gene	...	TG	
5	10.794880	0.000149	0.047714	0.666100	320	Gene	...	T	

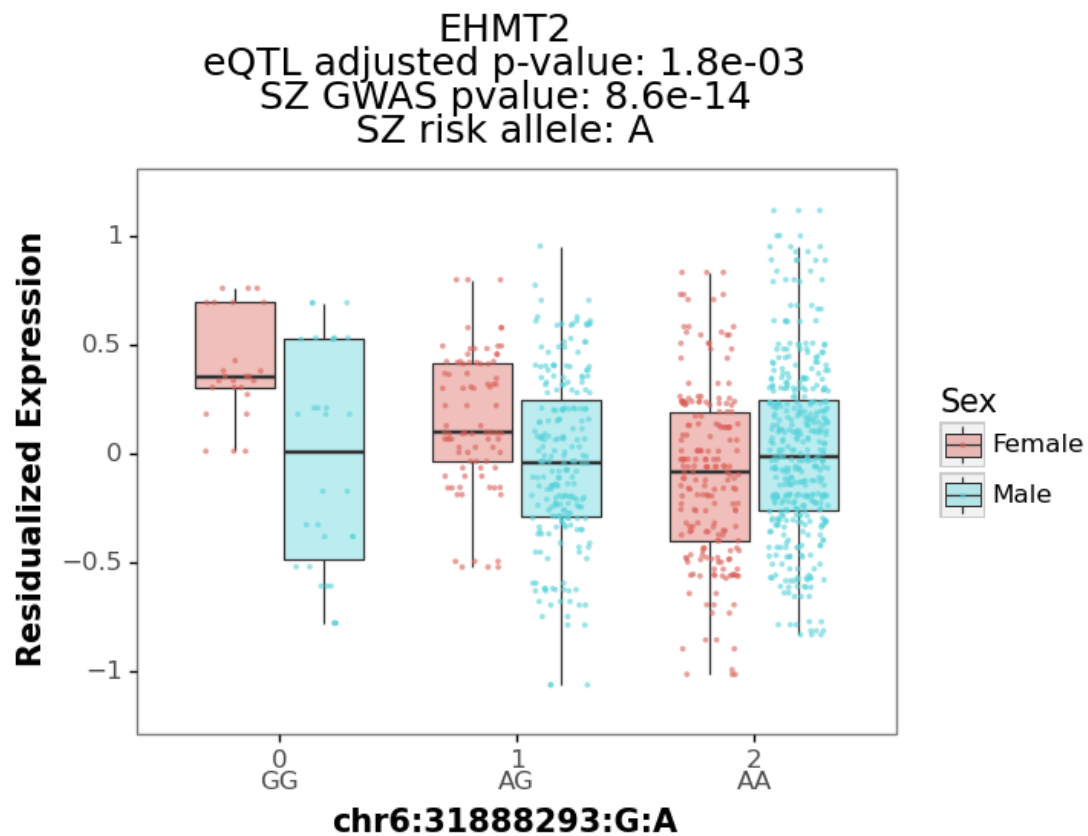
	OR	SE	P	hg19chrc	hg38chrc	hg38pos	\
0	0.92611	0.010290	8.620000e-14	chr6	chr6	31888293	
1	1.11790	0.013264	4.480000e-17	chr6	chr6	32225442	
2	1.18310	0.015322	5.000000e-28	chr6	chr6	31494358	
3	1.16530	0.014746	3.300000e-25	chr6	chr6	26573403	
4	1.08730	0.012600	3.060000e-11	chr8	chr8	110618046	
5	1.08930	0.011207	2.270000e-14	chr10	chr10	102875591	

	pgc2_a1_same_as_our_counted	rsid	is_index_snp
0	False	rs486416	False
1	False	rs3130295	False
2	False	rs3130923	False
3	False	rs13214027	False
4	True	rs35771435	False
5	False	rs12765002	False

[6 rows x 33 columns]

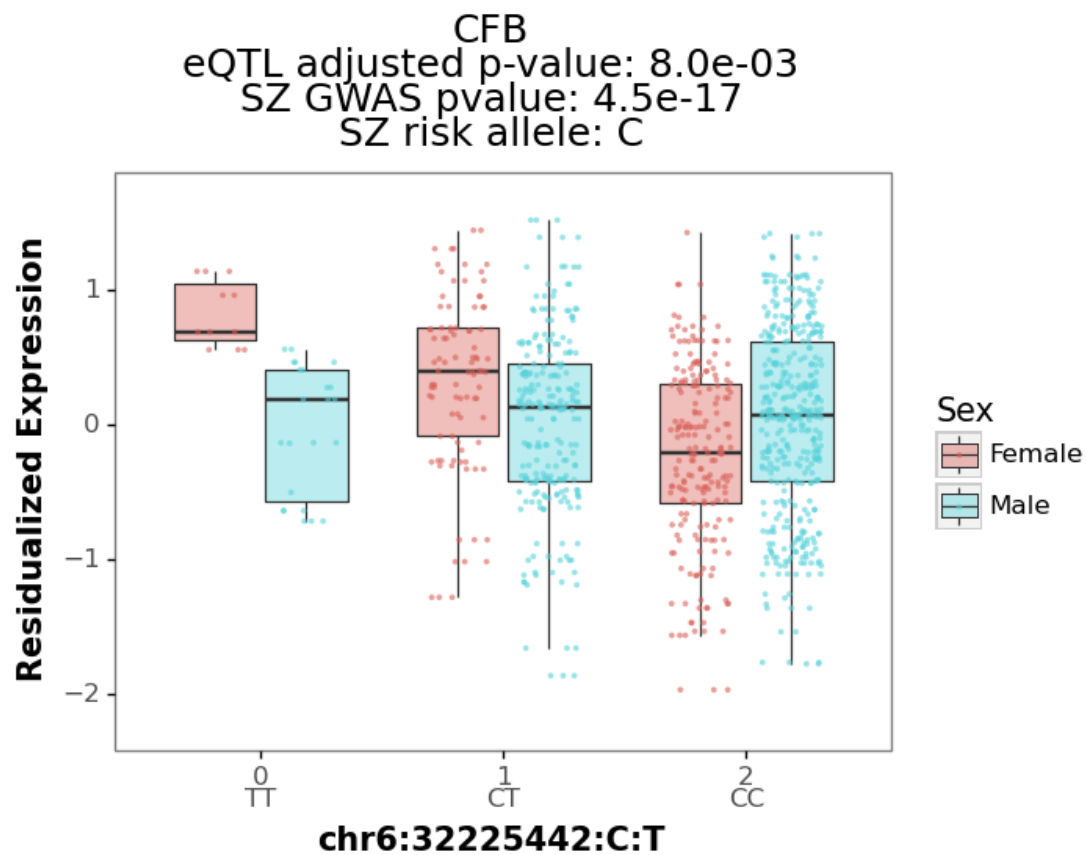
```
[13]: top_5_gwas = top_gwas_eqtl_df.head(5)
for x in top_5_gwas.itertuples():
    filename = "top_%d_eqtl_in_gwas_significant_snps_%s" % (x.Index, tissue)
    p = gwas_annotated_eqtl_plot(x.variant_id, x.gene_id)
    print(filename, x.Index, x.variant_id, x.gene_id)
    print(p)
    save_plot(p, filename)
```

top\_0\_eqtl\_in\_gwas\_significant\_snps\_dlpfc 0 chr6:31888293:G:A ENSG00000204371.11



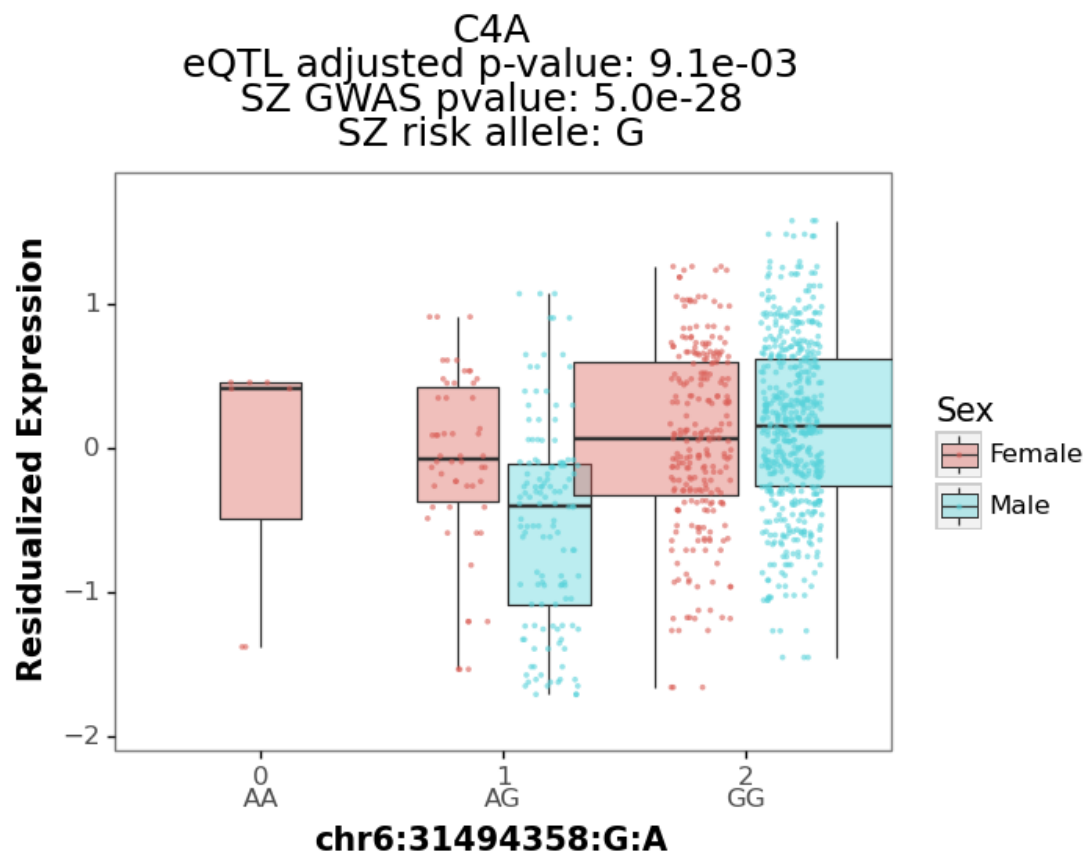
```
<ggplot: (8739663415432)>
```

```
top_1_eqtl_in_gwas_significant_snps_dlpfc 1 chr6:32225442:C:T ENSG00000243649.8
```



<ggplot: (8739634910085)>

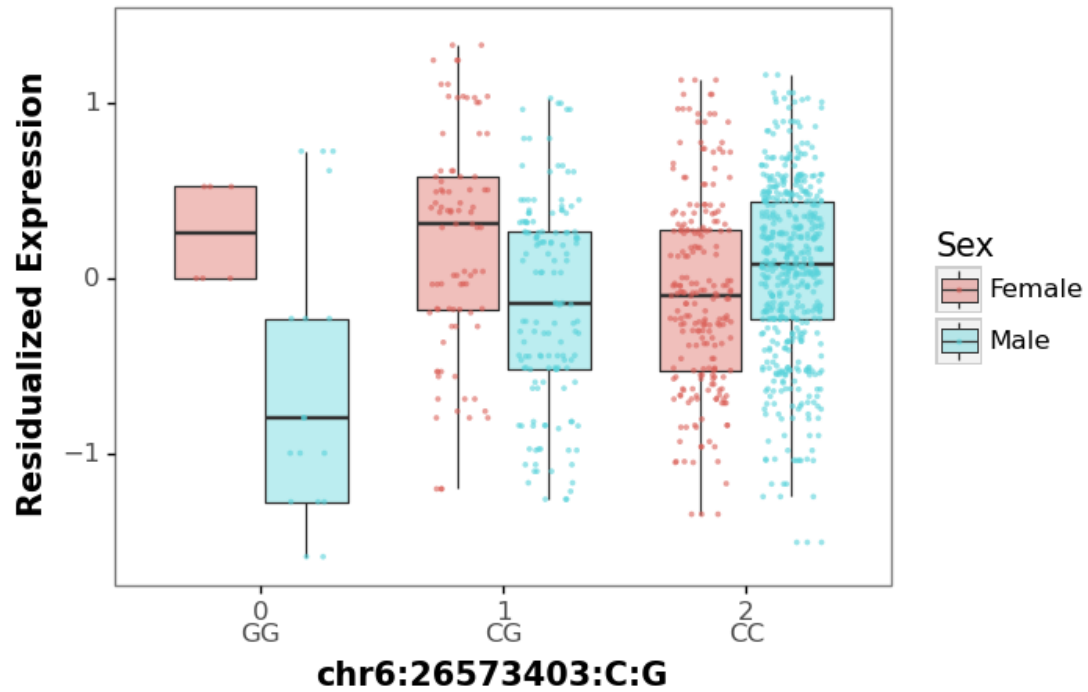
top\_2\_eqtl\_in\_gwas\_significant\_snps\_dlpfc 2 chr6:31494358:G:A ENSG00000244731.7



<ggplot: (8739635303355)>

top\_3\_eqtl\_in\_gwas\_significant\_snps\_dlpfc 3 chr6:26573403:C:G ENSG00000274290.2

HIST1H2BE  
eQTL adjusted p-value: 1.3e-02  
SZ GWAS pvalue: 3.3e-25  
SZ risk allele: C



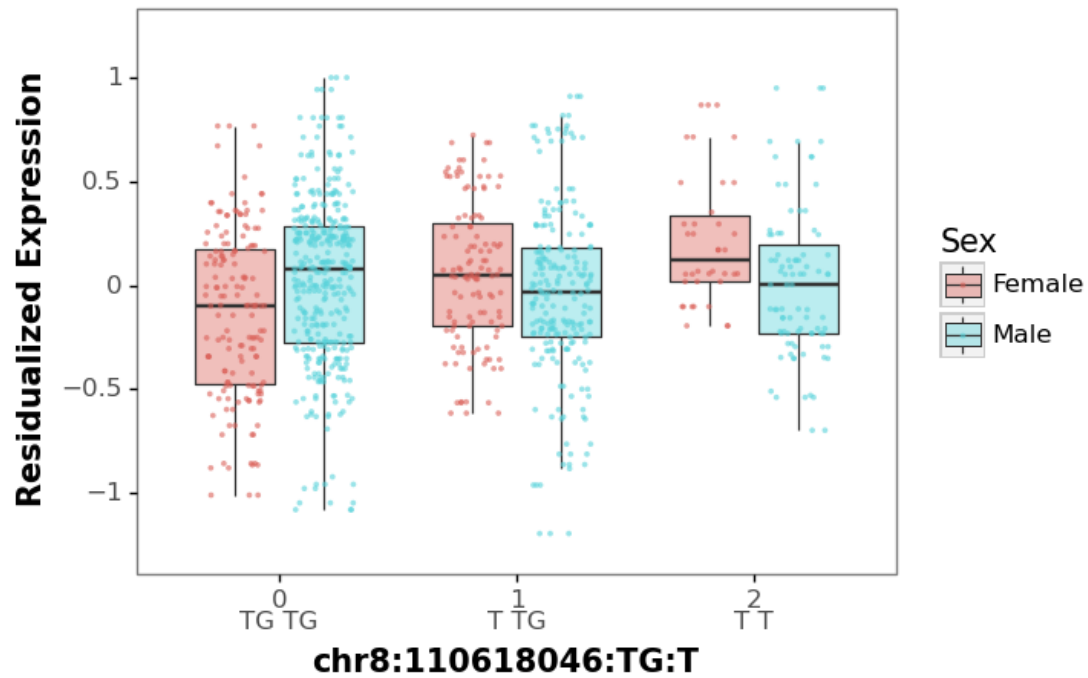
```
<ggplot: (8739636188145)>
```

```
top_4_eqtl_in_gwas_significant_snps_dlpfc 4 chr8:110618046:TG:T  

ENSG00000254241.1
```



MTCO1P47  
eQTL adjusted p-value: 1.9e-02  
SZ GWAS pvalue: 3.1e-11  
SZ risk allele: T



<ggplot: (8739635459151)>

[ ]: