main

August 16, 2021

1 Differential Expression with limma-voom pipeline - Genes

1.1 Prepare Data

1.2 Functions

1.2.1 General functions

```
[2]: # Function from jaffelab github
     merge_rse_metrics <- function(rse) {</pre>
         stopifnot(is(rse, 'RangedSummarizedExperiment'))
             c('concordMapRate', 'overallMapRate', 'mitoRate', 'rRNA_rate',
               'totalAssignedGene', 'numMapped', 'numReads', 'numUnmapped',
               'mitoMapped', 'totalMapped') %in%
                 colnames(SummarizedExperiment::colData(rse))
         )
         stopifnot(all(sapply(c(
             'concordMapRate', 'overallMapRate', 'mitoRate', 'rRNA_rate',
             'totalAssignedGene', 'numMapped', 'numReads', 'numUnmapped',
             'mitoMapped', 'totalMapped'), function(var) {
                 is(colData(rse)[, var], 'List')
             })
         ))
         rse$concordMapRate = mapply(function(r, n) {
             sum(r*n)/sum(n)
         }, rse$concordMapRate, rse$numReads)
         rse$overallMapRate = mapply(function(r, n) {
```

```
sum(r*n)/sum(n)
    }, rse$overallMapRate, rse$numReads)
    rse$mitoRate = mapply(function(r, n) {
        sum(r*n)/sum(n)
    }, rse$mitoRate, rse$numMapped)
    rse$rRNA_rate = mapply(function(r, n) {
        sum(r*n)/sum(n)
    }, rse$rRNA_rate, rse$numMapped)
    rse$totalAssignedGene = mapply(function(r, n) {
        sum(r*n)/sum(n)
    }, rse$totalAssignedGene, rse$numMapped)
    rse$numMapped = sapply(rse$numMapped, sum)
    rse$numReads = sapply(rse$numReads, sum)
    rse$numUnmapped = sapply(rse$numUnmapped, sum)
    rse$mitoMapped = sapply(rse$mitoMapped, sum)
    rse$totalMapped = sapply(rse$totalMapped, sum)
    return(rse)
}
```

```
[3]: save_volcanoPlot <- function(top, label, dirname){
         pdf(file=paste0(dirname, "/volcanoPlot_", label, ".pdf"), 8, 6)
         with(top, plot(logFC, -log10(P.Value), pch=20, cex=0.6))
         with(subset(top, adj.P.Val<=0.05), points(logFC, -log10(P.Value),
                                                    pch=20, col='red', cex=0.6))
         with(subset(top, abs(logFC)>0.50), points(logFC, -log10(P.Value),
                                                    pch=20, col='orange', cex=0.6))
         with(subset(top, adj.P.Val<=0.05 & abs(logFC)>0.50),
              points(logFC, -log10(P.Value), pch=20, col='green', cex=0.6))
         dev.off()
     }
     save_MAplot <- function(top, label, dirname){</pre>
         pdf(file=paste0(dirname, "/MAplot_", label, ".pdf"), 8, 6)
         with(top, plot(AveExpr, logFC, pch=20, cex=0.5))
         with(subset(top, adj.P.Val<0.05),</pre>
              points(AveExpr, logFC, col="red", pch=20, cex=0.5))
         dev.off()
     }
     extract_de <- function(contrast, label, efit, seed_int){</pre>
         dirname = paste0("permutation_", stringr::str_pad(seed_int, 2, pad = "0"))
         se <- as.data.frame(sqrt(efit$s2.post) * efit$stdev.unscaled) %>%
             rename("SE"="CtrlvsSZ") %>% rownames_to_column()
         top <- topTable(efit, coef=1, number=Inf, sort.by="P") %>%
             rownames_to_column() %>% inner_join(se, by="rowname") %>%
             column_to_rownames("rowname")
```

1.2.2 Cached functions

```
[4]: load counts <- function(){
         counts_file = paste0("/ceph/projects/v3_phase3_paper/inputs/phase3/_m/
      "caudate_brainseq_phase3_hg38_rseGene_merged_n464.rda")
         load(counts file)
         rse_df = rse_gene
         return(rse_df)
     memCounts <- memoise(load_counts)</pre>
     get random samples <- function(seed int, new dir=TRUE){</pre>
         set.seed(seed_int + 113) # seed for reproducibility
         dirname = paste0("permutation_", stringr::str_pad(seed_int, 2, pad = "0"))
         if(new_dir){
              dir.create(dirname)
         }
         rse_df <- memCounts()</pre>
         keepIndex = which((rse_df$Dx %in% c("Control", "Schizo")) &
                             rse_df$Age > 17 & rse_df$Sex == "M" &
                             rse df$Race %in% c("AA", "CAUC"))
         snames = sample(keepIndex, 121, replace=FALSE) # subsampling to Female N_{\sqcup}
      \hookrightarrow (sample size)
         return(snames)
     memSamples <- memoise(get_random_samples)</pre>
     get_mds <- function(){</pre>
         mds_file = "/ceph/projects/v4_phase3_paper/inputs/genotypes/mds/_m/
      {\scriptstyle \hookrightarrow LIBD\_Brain\_TopMed.mds"}
         mds = data.table::fread(mds file) %>%
              rename_at(.vars = vars(starts_with("C")),
                        function(x){sub("C", "snpPC", x)}) %>%
```

```
mutate_if(is.character, as.factor)
    return(mds)
}
memMDS <- memoise::memoise(get_mds)</pre>
prep_data <- function(seed_int){</pre>
    rse_df <- memCounts()</pre>
    keepIndex <- memSamples(seed_int)</pre>
    rse_df = rse_df[, keepIndex]
    rse_df$Dx = factor(rse_df$Dx, levels = c("Control", "Schizo"))
    rse_df$Sex <- factor(rse_df$Sex)</pre>
    rse_df <- merge_rse_metrics(rse_df)</pre>
    rse_df$ERCCsumLogErr <- mapply(function(r, n) {</pre>
        sum(r * n)/sum(n)
    }, rse_df$ERCCsumLogErr, rse_df$numReads)
    colData(rse_df)$RIN = sapply(colData(rse_df)$RIN,"[",1)
    rownames(colData(rse_df)) <- sapply(strsplit(rownames(colData(rse_df)),__
 →"_"), "[", 1)
    pheno = colData(rse_df) %>% as.data.frame %>%
        inner_join(memMDS(), by=c("BrNum"="FID")) %>%
        distinct(RNum, .keep all = TRUE)
    # Generate DGE list
    x <- DGEList(counts=assays(rse_df)$counts[, pheno$RNum],</pre>
                  genes=rowData(rse_df), samples=pheno)
    # Filter by expression
    design0 <- model.matrix(~Dx, data=x$samples)</pre>
    keep.x <- filterByExpr(x, design=design0)</pre>
    x <- x[keep.x, , keep.lib.sizes=FALSE]</pre>
    print(paste('There are:', sum(keep.x), 'features left!', sep=' '))
    # Normalize library size
    x <- calcNormFactors(x, method="TMM")</pre>
    return(x)
}
memo_prepData <- memoise(prep_data)</pre>
SVA_model <- function(seed_int){</pre>
    x <- memo_prepData(seed_int)</pre>
    # Design matrix
    mod = model.matrix(~Dx + Age + mitoRate + rRNA_rate + RIN +
                        totalAssignedGene + overallMapRate + ERCCsumLogErr +
                        snpPC1 + snpPC2 + snpPC3, data=x$samples)
    colnames(mod) <- gsub("Dx", "", colnames(mod))</pre>
    colnames(mod) <- gsub("\\(Intercept\\)", "Intercept", colnames(mod))</pre>
    # Calculated SVs
    null.model = mod %>% as.data.frame %>% select(-c("Schizo")) %>% as.matrix
    n.sv <- num.sv(x$counts, mod, method="be")</pre>
    svobj <- svaseq(x$counts, mod, null.model, n.sv=n.sv)</pre>
```

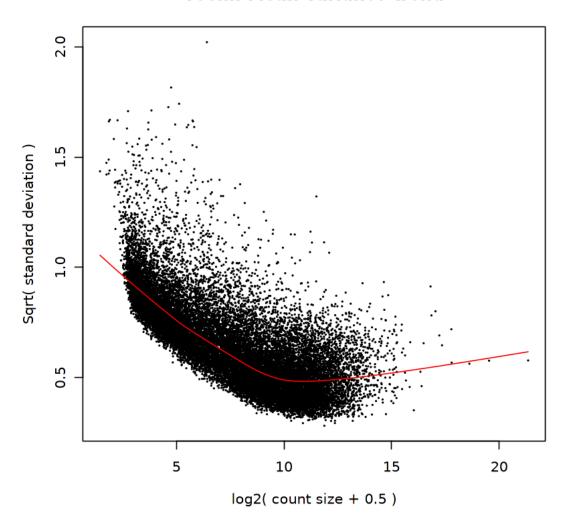
```
if(svobj$sv == 0){
        modQsva <- mod
    } else {
        modQsva <- cbind(mod, svobj$sv)</pre>
        len.d <- length(colnames(modQsva))</pre>
        colnames(modQsva)[((len.d - n.sv)+1):len.d] <- make.names(paste0("sv",1:</pre>
 \rightarrown.sv))
    }
    return (modQsva)
memo_svaModel <- memoise(SVA_model)</pre>
get_voom <- function(seed_int){</pre>
    ### Preform voom
    x <- memo_prepData(seed_int)</pre>
    modQsva <- memo_svaModel(seed_int)</pre>
    v <- voom(x[, rownames(modQsva)], modQsva, plot=TRUE)</pre>
    return(v)
}
memo_voom <- memoise(get_voom)</pre>
cal res <- function(seed int){</pre>
    ### Calculate residuals
    v <- memo_voom(seed_int)</pre>
    null_model <- v$design %>% as.data.frame %>% select(-c("Schizo")) %>% as.
 →matrix
    fit res <- lmFit(v, design=null model)</pre>
    res = v$E - ( fit_res$coefficients %*% t(null_model) )
    res_sd = apply(res, 1, sd)
    res_mean = apply(res, 1, mean)
    res_norm = (res - res_mean) / res_sd
    dirname = paste0("permutation_", stringr::str_pad(seed_int, 2, pad = "0"))
    write.table(res_norm, file=paste0(dirname, '/residualized_expression.tsv'),
                 sep="\t", quote=FALSE)
memo_res <- memoise(cal_res)</pre>
fit_voom <- function(seed_int){</pre>
    v <- memo_voom(seed_int)</pre>
    modQsva <- memo_svaModel(seed_int)</pre>
    fit0 <- lmFit(v, modQsva)</pre>
    contr.matrix <- makeContrasts(CtrlvsSZ = Schizo,</pre>
                                      levels=colnames(modQsva))
    fit <- contrasts.fit(fit0, contrasts=contr.matrix)</pre>
    esv <- eBayes(fit)</pre>
    return(esv)
```

```
memo_efit <- memoise(fit_voom)</pre>
```

1.3 Differential Expression Analysis

[1] "Comparison for: szVctl"
[1] "There are: 38 DE features!"

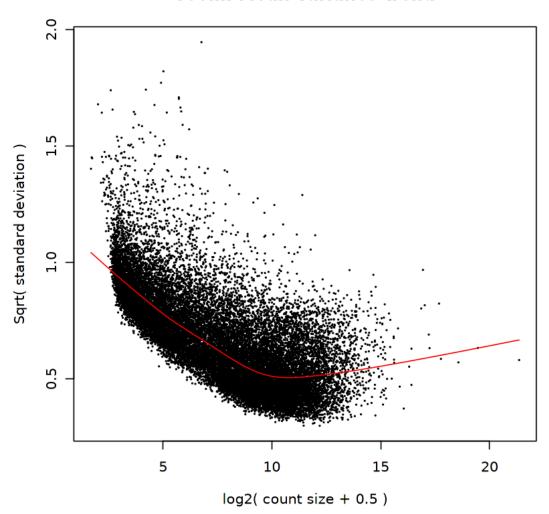
```
[5]: for(seed_int in seq(1, 10)){
         # Preform voom
         v <- memo_voom(seed_int)</pre>
         dirname = paste0("permutation_", stringr::str_pad(seed_int, 2, pad = "0"))
         save(v, file=paste0(dirname, '/voomSVA.RData'))
         # Fit model and apply eBayes
         efit = memo_efit(seed_int)
         # Save differential expression
         extract_de(1, "szVctl", efit, seed_int)
         # Calculate residuals
         memo_res(seed_int)
     }
    [1] "There are: 22898 features left!"
    No significant surrogate variables
    [1] "Comparison for: szVctl"
    [1] "There are: 324 DE features!"
    [1] "There are: 22849 features left!"
    No significant surrogate variables
```



[1] "There are: 22940 features left!" No significant surrogate variables

[1] "Comparison for: szVctl"

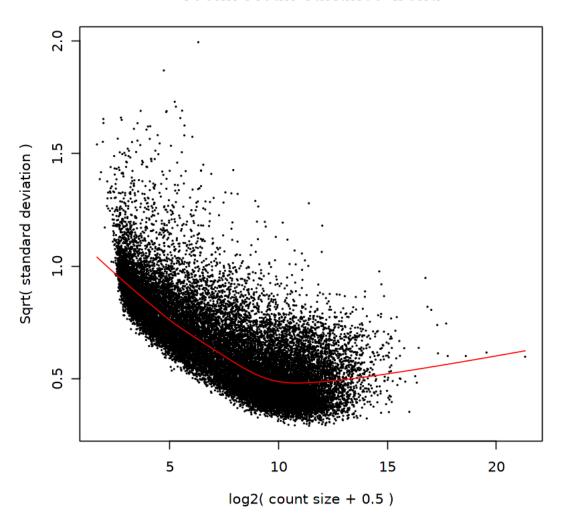
[1] "There are: 173 DE features!"



[1] "There are: 22662 features left!" No significant surrogate variables

[1] "Comparison for: szVctl"

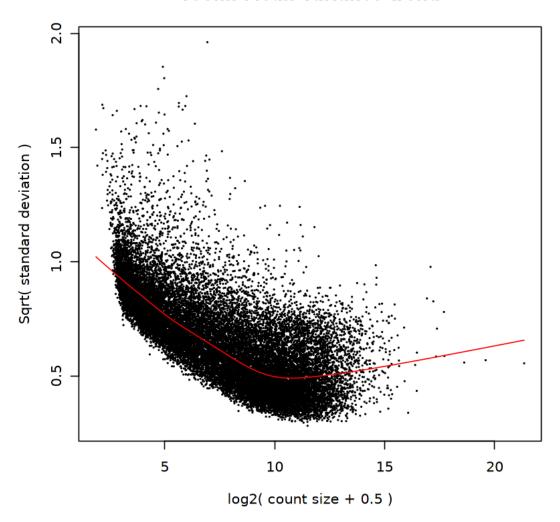
[1] "There are: 44 DE features!"



[1] "There are: 22792 features left!" No significant surrogate variables

[1] "Comparison for: szVctl"

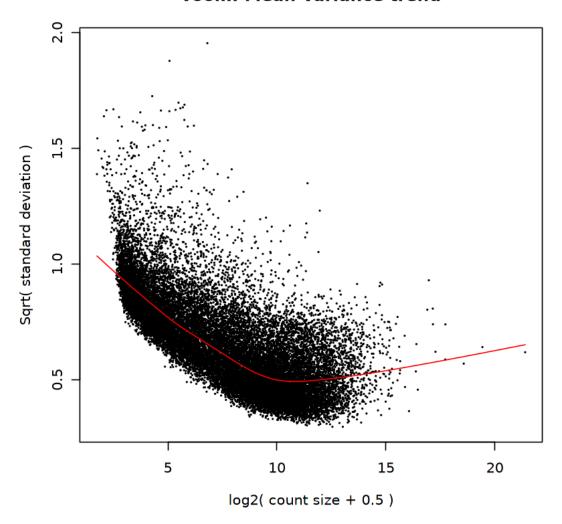
[1] "There are: 65 DE features!"



[1] "There are: 22602 features left!" No significant surrogate variables

[1] "Comparison for: szVctl"

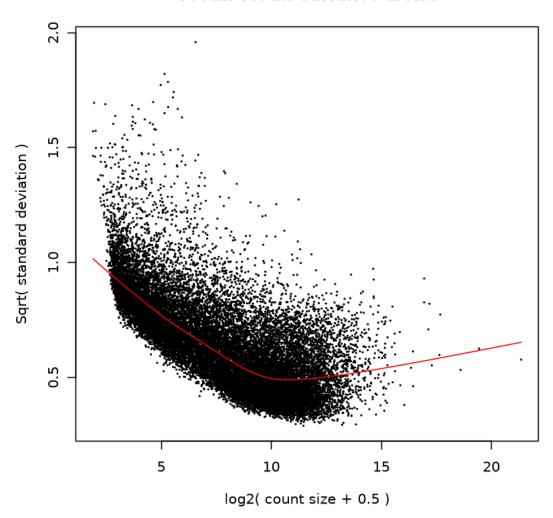
[1] "There are: 343 DE features!"



[1] "There are: 22840 features left!" No significant surrogate variables

[1] "Comparison for: szVctl"

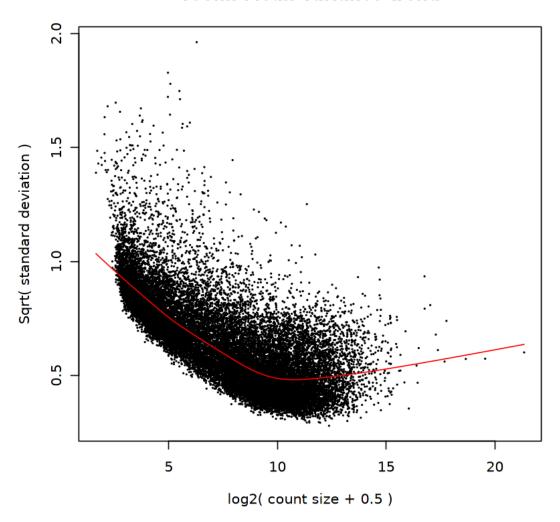
[1] "There are: 1688 DE features!"



[1] "There are: 23199 features left!" No significant surrogate variables

[1] "Comparison for: szVctl"

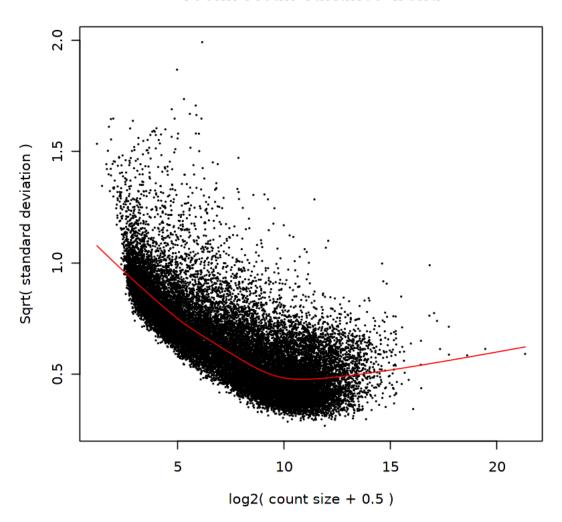
[1] "There are: 105 DE features!"



[1] "There are: 22409 features left!" No significant surrogate variables

[1] "Comparison for: szVctl"

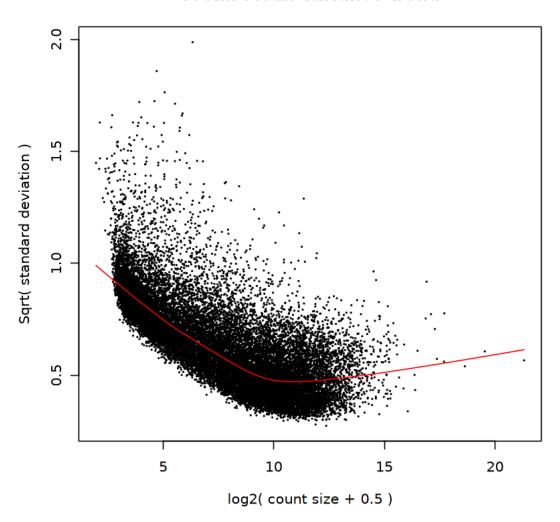
[1] "There are: 121 DE features!"

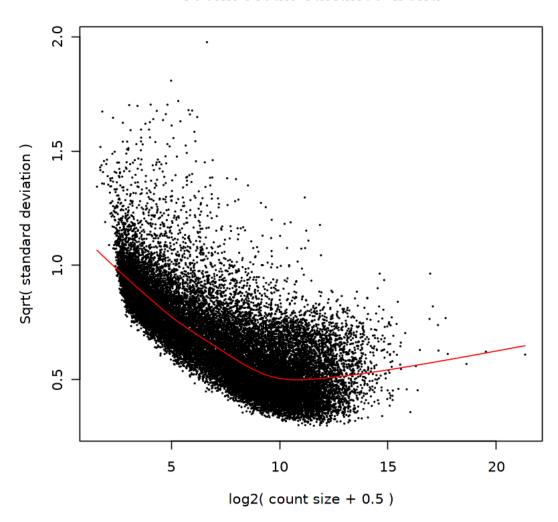


[1] "There are: 23161 features left!" No significant surrogate variables

[1] "Comparison for: szVctl"

[1] "There are: 187 DE features!"





1.4 Reproducibility Information

```
[6]: Sys.time()
    proc.time()
    options(width = 120)
    sessioninfo::session_info()

[1] "2021-08-16 16:55:45 EDT"

    user system elapsed
9056.674 648.935 2068.548

    Session info
    setting value
```

version R version 4.0.3 (2020-10-10)

os Arch Linux

system x86_64, linux-gnu

ui X11 language (EN)

collate en_US.UTF-8
ctype en_US.UTF-8
tz America/New_York

date 2021-08-16

Packages package

package	*	version	date	lib	source
annotate		1.68.0	2020-10-27	[1]	Bioconductor
AnnotationDbi		1.52.0	2020-10-27	[1]	Bioconductor
assertthat		0.2.1	2019-03-21	[1]	CRAN (R 4.0.2)
backports		1.2.1	2020-12-09	[1]	CRAN (R 4.0.2)
base64enc		0.1-3	2015-07-28	[1]	CRAN (R 4.0.2)
Biobase	*	2.50.0	2020-10-27	[1]	Bioconductor
BiocGenerics	*	0.36.1	2021-04-16	[1]	Bioconductor
BiocParallel	*	1.24.1	2020-11-06	[1]	Bioconductor
bit		4.0.4	2020-08-04	[1]	CRAN (R 4.0.2)
bit64		4.0.5	2020-08-30	[1]	CRAN (R 4.0.2)
bitops		1.0-7	2021-04-24	[1]	CRAN (R 4.0.3)
blob		1.2.1	2020-01-20	[1]	CRAN (R 4.0.2)
broom		0.7.8	2021-06-24	[1]	CRAN (R 4.0.3)
cachem		1.0.5	2021-05-15	[1]	CRAN (R 4.0.3)
Cairo		1.5-12.2	2020-07-07	[1]	CRAN (R 4.0.2)
cellranger		1.1.0	2016-07-27	[1]	CRAN (R 4.0.2)
cli		3.0.0	2021-06-30	[1]	CRAN (R 4.0.3)
colorspace		2.0-2	2021-06-24	[1]	CRAN (R 4.0.3)
crayon		1.4.1	2021-02-08	[1]	CRAN (R 4.0.3)
data.table	*	1.14.0	2021-02-21	[1]	CRAN (R 4.0.3)
DBI		1.1.1	2021-01-15	[1]	CRAN (R 4.0.2)
dbplyr		2.1.1	2021-04-06	[1]	CRAN (R 4.0.3)
DelayedArray		0.16.3	2021-03-24	[1]	Bioconductor
digest		0.6.27	2020-10-24	[1]	CRAN (R 4.0.2)
dplyr	*	1.0.7	2021-06-18	[1]	CRAN (R 4.0.3)
edgeR	*	3.32.1	2021-01-14	[1]	Bioconductor
ellipsis		0.3.2	2021-04-29	[1]	CRAN (R 4.0.3)
evaluate		0.14	2019-05-28	[1]	CRAN (R 4.0.2)
fansi		0.5.0	2021-05-25	[1]	CRAN (R 4.0.3)
fastmap		1.1.0	2021-01-25	[1]	CRAN (R 4.0.2)
forcats	*	0.5.1	2021-01-27	[1]	CRAN (R 4.0.2)
fs		1.5.0	2020-07-31	[1]	CRAN (R 4.0.2)
genefilter	*	1.72.1	2021-01-21	[1]	Bioconductor
generics		0.1.0	2020-10-31	[1]	CRAN (R 4.0.2)
GenomeInfoDb	*	1.26.7	2021-04-08	[1]	Bioconductor
${\tt GenomeInfoDbData}$		1.2.4	2021-02-02	[1]	Bioconductor

GenomicRanges	*	1.42.0	2020-10-27	[1]	Bioconductor
ggplot2		3.3.5	2021-06-25	[1]	CRAN (R 4.0.3)
glue	.,.	1.4.2	2020-08-27	[1]	CRAN (R 4.0.2)
gtable		0.3.0	2019-03-25	[1]	CRAN (R 4.0.2)
haven		2.4.1	2019 03 23	[1]	CRAN (R 4.0.3)
hms		1.1.0	2021-04-23		CRAN (R 4.0.3)
htmltools		0.5.1.1	2021-03-17		CRAN (R 4.0.2)
httr		1.4.2	2021-01-22		CRAN (R 4.0.2)
	J.	2.24.1			Bioconductor
IRanges	•			[1]	CRAN (R 4.0.2)
IRdisplay IRkernel		1.0	2021-01-20		
			2021-05-11	[1]	CRAN (R 4.0.3)
jsonlite		1.7.2	2020-12-09		CRAN (R 4.0.2)
lattice		0.20-41	2020-04-02		CRAN (R 4.0.3)
lifecycle		1.0.0	2021-02-15	[1]	CRAN (R 4.0.3)
limma	*	3.46.0	2020-10-27		Bioconductor
locfit		1.5-9.4	2020-03-25		CRAN (R 4.0.2)
lubridate		1.7.10	2021-02-26		CRAN (R 4.0.3)
magrittr		2.0.1	2020-11-17		CRAN (R 4.0.2)
Matrix		1.3-4	2021-06-01	[1]	CRAN (R 4.0.3)
MatrixGenerics	*	1.2.1	2021-01-30	[1]	
matrixStats	*	0.59.0	2021-06-01	[1]	CRAN (R 4.0.3)
memoise	*	2.0.0	2021-01-26	[1]	CRAN (R 4.0.2)
mgcv	*	1.8-33	2020-08-27		CRAN (R 4.0.3)
modelr		0.1.8	2020-05-19		
munsell		0.5.0	2018-06-12		CRAN (R 4.0.2)
nlme	*	3.1-152	2021-02-04	[1]	CRAN (R 4.0.3)
pbdZMQ		0.3-5	2021-02-10	[1]	CRAN (R 4.0.3)
pillar		1.6.1	2021-05-16	[1]	CRAN (R 4.0.3)
pkgconfig		2.0.3	2019-09-22	[1]	CRAN (R 4.0.2)
purrr	*	0.3.4	2020-04-17	[1]	CRAN (R 4.0.2)
R6		2.5.0	2020-10-28	[1]	CRAN (R 4.0.2)
Rcpp		1.0.7	2021-07-07		CRAN (R 4.0.3)
RCurl		1.98-1.3	2021-03-16	[1]	CRAN (R 4.0.3)
readr	*	1.4.0	2020-10-05	[1]	CRAN (R 4.0.2)
readxl		1.3.1	2019-03-13	[1]	CRAN (R 4.0.2)
repr		1.1.3	2021-01-21	[1]	CRAN (R 4.0.2)
reprex		2.0.0	2021-04-02	[1]	CRAN (R 4.0.3)
rlang		0.4.11	2021-04-30	[1]	CRAN (R 4.0.3)
RSQLite		2.2.7	2021-04-22	[1]	CRAN (R 4.0.3)
rstudioapi		0.13	2020-11-12	[1]	CRAN (R 4.0.2)
rvest		1.0.0	2021-03-09	[1]	CRAN (R 4.0.3)
S4Vectors	*	0.28.1	2020-12-09	[1]	Bioconductor
scales		1.1.1	2020-05-11		CRAN (R 4.0.2)
sessioninfo		1.1.1			CRAN (R 4.0.2)
stringi		1.7.3	2021-07-16		
stringr	*	1.4.0	2019-02-10		
SummarizedExperiment					Bioconductor
survival		3.2-7			CRAN (R 4.0.3)
					, , , , , , , , , , , , , , , , , , , ,

```
* 3.38.0
                                2020-10-27 [1] Bioconductor
sva
tibble
                     * 3.1.2
                                2021-05-16 [1] CRAN (R 4.0.3)
                     * 1.1.3
                                2021-03-03 [1] CRAN (R 4.0.3)
tidyr
tidyselect
                       1.1.1
                                2021-04-30 [1] CRAN (R 4.0.3)
tidyverse
                     * 1.3.1
                                2021-04-15 [1] CRAN (R 4.0.3)
                                2021-03-12 [1] CRAN (R 4.0.3)
utf8
                       1.2.1
uuid
                       0.1-4
                                2020-02-26 [1] CRAN (R 4.0.2)
                                2021-04-29 [1] CRAN (R 4.0.3)
                       0.3.8
vctrs
withr
                       2.4.2
                                2021-04-18 [1] CRAN (R 4.0.3)
XML
                       3.99-0.6 2021-03-16 [1] CRAN (R 4.0.3)
                                2020-04-23 [1] CRAN (R 4.0.2)
xm12
                       1.3.2
xtable
                       1.8-4
                                2019-04-21 [1] CRAN (R 4.0.2)
                                2020-10-27 [1] Bioconductor
XVector
                       0.30.0
                       1.36.0
                                2020-10-27 [1] Bioconductor
zlibbioc
```

- [1] /home/jbenja13/R/x86_64-pc-linux-gnu-library/4.0
- [2] /usr/lib/R/library