main

July 14, 2021

1 Visualize GO analysis

```
[1]: import numpy as np
     import pandas as pd
[2]: def get_top_GO(tissue, fn, label):
         df = pd.read_csv(fn, sep='\t').sort_values('p_value').head(10)
         df['Log10'] = -np.log10(df['p_value'])
         df['Tissue'] = tissue
         df['Direction'] = label
         return df
[3]: tissue = 'caudate'
     config = {
         'All': '../_m/DEGs_functional_enrichment.tsv',
         'Up': '../../_m/upreg_DEGs_functional_enrichment.tsv',
         'Down': '../../_m/downreg_DEGs_functional_enrichment.tsv',
     }
     df = pd.DataFrame()
     for bias in ['All', 'Up', 'Down']:
         df = pd.concat([df, get_top_GO(tissue, config[bias], bias)], axis=0)
     df.shape
[3]: (30, 17)
[4]: df.to_csv("%s_functional_analysis.tsv" % tissue, sep='\t', index=False)
    1.1 Plot
[5]: %load_ext rpy2.ipython
[6]: \%\R -i df
     library(ggplot2)
     library(tidyverse)
     save_plot <- function(p, fn, w, h){</pre>
```

```
for(ext in c('.svg', '.png', '.pdf')){
             ggsave(file=paste0(fn,ext), plot=p, width=w, height=h)
         }
     }
     plot_GO <- function(){</pre>
         cbPalette <- c("#000000", "Red", "Blue")
         gg1 = df \% > \%
             ggplot(aes(x=Log10, y=term name, color=Direction)) +
             geom_point(shape=18, alpha=0.8, size=4) + labs(y='', x='-Log10 (pu
     →adjust)') +
             theme_bw() +
             scale_colour_manual(name="Direction", values=cbPalette,
                                 labels=c("All", "Upregulated in SZ", "Downregulated ⊔
     →in SZ")) +
             geom_vline(xintercept = -log10(0.05), linetype = "dotted") +
             theme(axis.text=element_text(size=14),
                   axis.title=element_text(size=18, face='bold'),
                   strip.text=element_text(size=18, face='bold'))
         return(gg1)
     }
    R[write to console]:
                           Attaching packages
                          tidyverse 1.3.1
    R[write to console]: tibble 3.1.2
                                               dplyr 1.0.7
     tidyr 1.1.3
                         stringr 1.4.0
                         forcats 0.5.1
     readr 1.4.0
             0.3.4
     purrr
    R[write to console]:
                           Conflicts
    tidyverse_conflicts()
     dplyr::filter() masks stats::filter()
                     masks stats::lag()
     dplyr::lag()
[7]: %%R
     gg1 = plot_GO()
     print(gg1)
     save_plot(gg1, "dlpfc_GO_top10_stacked", 10, 6)
```

- taxis-
- Resistance of ERBB2 KD mutants to neratinib-
- Resistance of ERBB2 KD mutants to lapatinib-
- Resistance of ERBB2 KD mutants to afatinib-
- Resistance of ERBB2 KD mutants to AEE788-
- regulation of multicellular organismal process
 - regulation of leukocyte activation
 - regulation of cell activation-
 - positive regulation of macrophage migration
 - positive regulation of leukocyte migration
 - plasma membrane
 - membrane-

Direction

Upregulated in SZ

Downregulated in SZ

- leukocyte migration-
- leukocyte cell-cell adhesion-
- intracellular signal transduction-
 - GTPase activator activity-
- extrinsic component of plasma membrane-
- Drug-mediated inhibition of ERBB2 signaling-
- Drug resistance in ERBB2 TMD/JMD mutants-
 - Drug resistance in ERBB2 KD mutants
 - cytokine production-
 - CTNNB1-FERMT2-YBX1 complex-
 - CTNNB1-FERMT2-TCF7L2 complex
 - circulatory system process
 - chemotaxis-
 - cell periphery-
 - basal plasma membrane
 - basal part of cell-

2.6

-Log10 (p adjust)

[]: