

main_female

November 24, 2021

1 Tissue comparison for differential expression analysis

```
[1]: import functools
import numpy as np
import pandas as pd
from gtftparse import read_gtf

[2]: config = {
    'caudate': '../.../caudate/female_analysis/metrics_summary/_m/
    ↪female_specific_DE_4features.txt',
    'dlpfc': '../.../dlpfc/female_analysis/metrics_summary/_m/
    ↪female_specific_DE_4features.txt',
    'hippo': '../.../hippocampus/female_analysis/metrics_summary/_m/
    ↪female_specific_DE_4features.txt',
    'cmc_dlpfc': '../.../cmc_dlpfc/female_analysis/metrics_summary/_m/
    ↪female_specific_DE_genes.txt'
}

[3]: @functools.lru_cache()
def get_deg(filename):
    dft = pd.read_csv(filename, sep='\t', index_col=0)
    dft = dft[(dft['Type'] == 'gene')].copy()
    dft['Feature'] = dft.index
    dft['Dir'] = np.sign(dft['t'])
    if 'gene_id' in dft.columns:
        dft['ensemblID'] = dft.gene_id.str.replace('\\.*', '', regex=True)
    return dft[['Feature', 'ensemblID', 'adj.P.Val', 'logFC', 't', 'Dir']]

@functools.lru_cache()
def get_deg_sig(filename):
    dft = get_deg(filename)
    return dft[(dft['adj.P.Val'] < 0.05)]

@functools.lru_cache()
def merge_dataframes(tissue1, tissue2):
    return get_deg(config[tissue1]).merge(get_deg(config[tissue2]),
```

```

                                on='Feature',
                                suffixes=['_%s' % tissue1, '%s' %
→tissue2])

@functools.lru_cache()
def merge_dataframes_sig(tissue1, tissue2):
    return get_deg_sig(config[tissue1]).merge(get_deg_sig(config[tissue2]),
                                                on='Feature',
                                                suffixes=['_%s' % tissue1, '%s'
→% tissue2])

```

```

[4]: def tissue_annotation(tissue):
    return {'dlpfc': 'DLPFC', 'hippo': 'Hippocampus',
            'caudate': 'Caudate', 'cmc_dlpfc': 'CMC DLPFC'}[tissue]

def save_plot(p, fn, width=7, height=7):
    '''Save plot as svg, png, and pdf with specific label and dimension.'''
    for ext in ['.svg', '.png', '.pdf']:
        p.save(fn+ext, width=width, height=height)

```

1.1 BrainSeq Comparison

```

[5]: caudate = get_deg(config['caudate'])
    caudate.groupby('Dir').size()

```

```

[5]: Dir
-1.0    16
 1.0    14
dtype: int64

```

```

[6]: caudate[(caudate['adj.P.Val'] < 0.05)].shape

```

INFO:numexpr.utils:Note: NumExpr detected 60 cores but "NUMEXPR_MAX_THREADS" not set, so enforcing safe limit of 8.

INFO:numexpr.utils:NumExpr defaulting to 8 threads.

```

[6]: (30, 6)

```

```

[7]: dlpfc = get_deg(config['dlpfc'])
    dlpfc.groupby('Dir').size()

```

```

[7]: Series([], dtype: int64)

```

```

[8]: dlpfc[(dlpfc['adj.P.Val'] < 0.05)].shape

```

```

[8]: (0, 6)

```

```
[9]: hippo = get_deg(config['hippo'])
hippo.groupby('Dir').size()
```

```
[9]: Series([], dtype: int64)
```

```
[10]: hippo[(hippo['adj.P.Val'] < 0.05)].shape
```

```
[10]: (0, 6)
```

1.1.1 Upset Plot

```
[11]: phase2_dlpfc = dlpfc[(dlpfc['adj.P.Val'] < 0.05)].copy()
phase2_dlpfc['DLPFC'] = 1
phase2_dlpfc = phase2_dlpfc[['ensemblID', 'DLPFC']]

phase2_hippo = hippo[(hippo['adj.P.Val'] < 0.05)].copy()
phase2_hippo['Hippocampus'] = 1
phase2_hippo = phase2_hippo[['ensemblID', 'Hippocampus']]

phase3_caodate = caodate[(caodate['adj.P.Val'] < 0.05)].copy()
phase3_caodate['Caudate'] = 1
phase3_caodate = phase3_caodate[['ensemblID', 'Caudate']]
```

```
[12]: geneList = pd.merge(phase3_caodate[['ensemblID']], phase2_dlpfc[['ensemblID']],
                        on=['ensemblID'], how='outer')\
                        .merge(phase2_hippo[['ensemblID']], on=['ensemblID'], how='outer')\
                        .groupby(['ensemblID']).first().reset_index()

newC = pd.merge(geneList, phase3_caodate, on=['ensemblID'], how='outer').
    ↪fillna(0)
newC['Caudate'] = newC['Caudate'].astype('int')

newD1 = pd.merge(geneList, phase2_dlpfc, on=['ensemblID'], how='outer').
    ↪fillna(0)
newD1['DLPFC'] = newD1['DLPFC'].astype('int')

newH = pd.merge(geneList, phase2_hippo, on=['ensemblID'], how='outer').fillna(0)
newH['Hippocampus'] = newH['Hippocampus'].astype('int')

print(newC.shape, newH.shape, newD1.shape)
```

```
(30, 2) (30, 2) (30, 2)
```

```
[13]: df = pd.concat([newC.set_index(['ensemblID']), newD1.set_index(['ensemblID']),
                    newH.set_index(['ensemblID'])], axis=1, join='outer')
df.head(2)
```

```
[13]:
```

	Caudate	DLPFC	Hippocampus
ensemblID			
ENSG00000003137	1	0	0
ENSG000000070915	1	0	0

```
[14]: %load_ext rpy2.ipython
```

```
[15]: %%R
library(ComplexHeatmap)
library(tidyverse)
subset_pvalue <- function(filename, fdr_cutoff){
  df <- data.table::fread(filename) %>%
    filter(Type == 'gene', adj.P.Val < fdr_cutoff)
  return(df$ensemblID)
}

caudate = subset_pvalue('../.../caudate/female_analysis/metrics_summary/_m/
  ↳female_specific_DE_4features.txt',
                        0.05)
dlpfc = subset_pvalue('../.../dlpfc/female_analysis/metrics_summary/_m/
  ↳female_specific_DE_4features.txt',
                      0.05)
hippo = subset_pvalue('../.../hippocampus/female_analysis/metrics_summary/_m/
  ↳female_specific_DE_4features.txt',
                      0.05)

lt = list(Caudate = caudate,
          DLPFC = dlpfc,
          Hippocampus = hippo)

m = make_comb_mat(lt)
cbb_palette <- c("#000000", "#E69F00", "#56B4E9", "#009E73", "#F0E442",
                 "#0072B2", "#D55E00", "#CC79A7")
```

WARNING:rpy2.rinterface_lib.callbacks:R[write to console]: Loading required package: grid

WARNING:rpy2.rinterface_lib.callbacks:R[write to console]:

=====

ComplexHeatmap version 2.10.0

Bioconductor page: <http://bioconductor.org/packages/ComplexHeatmap/>

Github page: <https://github.com/jokergoo/ComplexHeatmap>

Documentation: <http://jokergoo.github.io/ComplexHeatmap-reference>

If you use it in published research, please cite:

Gu, Z. Complex heatmaps reveal patterns and correlations in multidimensional genomic data. *Bioinformatics* 2016.

The new InteractiveComplexHeatmap package can directly export static complex heatmaps into an interactive Shiny app with zero effort. Have a try!

This message can be suppressed by:

```
suppressPackageStartupMessages(library(ComplexHeatmap))
=====
```

```
WARNING:rpy2.rinterface.lib.callbacks:R[write to console]:  Attaching packages
                    tidyverse 1.3.1
```

```
WARNING:rpy2.rinterface.lib.callbacks:R[write to console]:  ggplot2 3.3.5
purrr 0.3.4
tibble 3.1.6      dplyr 1.0.7
tidyr 1.1.4      stringr 1.4.0
readr 2.1.0      forcats 0.5.1
```

```
WARNING:rpy2.rinterface.lib.callbacks:R[write to console]:  Conflicts
                    tidyverse_conflicts()
dplyr::filter() masks stats::filter()
dplyr::lag()    masks stats::lag()
```

```
[16]: %%R
right_annot = upset_right_annotation(
  m, ylim = c(0, 150),
  gp = gpar(fill = "black"),
  annotation_name_side = "top",
  axis_param = list(side = "top"))

top_annot = upset_top_annotation(
  m, height=unit(7, "cm"),
  ylim = c(0, 150),
  gp=gpar(fill=cbb_palette[comb_degree(m)]),
  annotation_name_rot = 90)

pdf('BrainSeq_sex_tissue_upsetR_DEgenes_femaleSpecific.pdf', width=6, height=4)
ht = draw(UpSet(m, pt_size=unit(4, "mm"), lwd=3,
               comb_col=cbb_palette[comb_degree(m)],
               set_order = c("Caudate", "DLPFC", "Hippocampus"),
               comb_order = order(-comb_size(m)),
               row_names_gp = gpar(fontsize = 14, fontface='bold'),
               right_annotation = right_annot,
               top_annotation = top_annot))
od = column_order(ht)
cs = comb_size(m)
decorate_annotation("intersection_size", {
```

```

        grid.text(cs[od], x = seq_along(cs), y = unit(cs[od], "native") +
                  unit(6, "pt"),
                  default.units = "native", just = "bottom", gp = gpar(fontsize = 11))
    })
    dev.off()

```

png
2

```

[17]: %%%R
right_ha = rowAnnotation(
  "Intersection\nsize" = anno_barplot(comb_size(m), border=F,
                                     ylim = c(0, 150),
                                     ↪gp=gpar(fill=cbb_palette[comb_degree(m)]),
                                     width = unit(7, "cm")))
top_ha = HeatmapAnnotation(
  "Set size" = anno_barplot(set_size(m), border=F,
                           ylim = c(0, 150),
                           gp = gpar(fill = "black"),
                           height = unit(2, "cm")),
  gap = unit(2, "mm"), annotation_name_side = "left",
  annotation_name_rot = 90)

pdf("BrainSeq_sex_tissue_upsetR_DEgenes_transpose_femaleSpecific.pdf", width=5, ↪
    ↪height=10)
ht = draw(UpSet(t(m), pt_size=unit(5, "mm"), lwd=3,
               comb_order = order(-comb_size(m)),
               comb_col=cbb_palette[comb_degree(m)],
               set_order = c("Caudate", "DLPFC", "Hippocampus"),
               column_names_gp = gpar(fontsize = 16, fontface='bold'),
               right_annotation = right_ha, top_annotation=top_ha))

od = rev(row_order(ht))
cs = comb_size(m)
decorate_annotation("Intersection\nsize", {
  grid.text(cs[od], y = seq_along(cs), x = unit(cs[od], "native") +
            unit(6, "pt"),
            default.units = "native", just = "left", gp = gpar(fontsize = 11))
})
dev.off()

```

png
2

1.1.2 Shared features

```
[18]: @functools.lru_cache()
def get_gtf(gtf_file):
    return read_gtf(gtf_file)
```

```
[19]: def gene_annotation(gtf_file, feature):
    gtf0 = get_gtf(gtf_file)
    gtf = gtf0[gtf0["feature"] == feature]
    return gtf[["gene_id", "gene_name", "transcript_id", "exon_id",
    ↪ "gene_type", "seqname", "start", "end", "strand"]]

gtf_file = '/ceph/genome/human/gencode25/gtf.CHR/_m/gencode.v25.annotation.gtf'
gtf_annot = gene_annotation(gtf_file, 'gene')
gtf_annot.head(2)
```

```
INFO:root:Extracted GTF attributes: ['gene_id', 'gene_type', 'gene_status',
'gene_name', 'level', 'havana_gene', 'transcript_id', 'transcript_type',
'transcript_status', 'transcript_name', 'transcript_support_level', 'tag',
'havana_transcript', 'exon_number', 'exon_id', 'ont', 'protein_id', 'ccdsid']
```

```
[19]:      gene_id gene_name transcript_id exon_id \
0   ENSG00000223972.5   DDX11L1
12  ENSG00000227232.5   WASH7P

      gene_type seqname  start  end strand
0   transcribed_unprocessed_pseudogene   chr1  11869  14409      +
12      unprocessed_pseudogene   chr1  14404  29570      -
```

```
[20]: dft = caudate.merge(gtf_annot[['gene_id', 'gene_name', 'seqname']],
    left_index=True, right_on='gene_id')
dft.head(2)
```

```
[20]:      Feature      ensemblID  adj.P.Val  logFC      t \
1928121  ENSG00000070915.9  ENSG00000070915  0.006380  0.909953  4.668428
1468999  ENSG00000111181.12  ENSG00000111181  0.009994 -0.432628 -4.405278

      Dir      gene_id gene_name seqname
1928121  1.0  ENSG00000070915.9  SLC12A3  chr16
1468999 -1.0  ENSG00000111181.12  SLC6A12  chr12
```

1.2 Comparison with CommonMind

```
[21]: cmc_dlpfc = get_deg(config['cmc_dlpfc'])
cmc_dlpfc.groupby('Dir').size()
```

```
[21]: Dir
-1.0    227
```

```
1.0    356
dtype: int64
```

```
[22]: cmc_dlpfc[(cmc_dlpfc['adj.P.Val'] < 0.05)].shape
```

```
[22]: (583, 6)
```

1.2.1 Upset Plot

```
[23]: cmc = cmc_dlpfc[(cmc_dlpfc['adj.P.Val'] < 0.05)].copy()
cmc['CMC DLPFC'] = 1
cmc = cmc[['ensemblID', 'CMC DLPFC']].groupby('ensemblID').first().reset_index()
```

```
[24]: geneList = pd.merge(phase3_caudate[['ensemblID']], phase2_dlpfc[['ensemblID']],
    ↳ on=['ensemblID'], how='outer')\
        .merge(phase2_hippo[['ensemblID']], on=['ensemblID'], how='outer')\
        .merge(cmc[['ensemblID']], on=['ensemblID'], how='outer')\
        .groupby(['ensemblID']).first().reset_index()
```

```
newC = pd.merge(geneList, phase3_caudate, on=['ensemblID'], how='outer').
    ↳ fillna(0)
newC['Caudate'] = newC['Caudate'].astype('int')
```

```
newD1 = pd.merge(geneList, phase2_dlpfc, on=['ensemblID'], how='outer').
    ↳ fillna(0)
newD1['DLPFC'] = newD1['DLPFC'].astype('int')
```

```
newH = pd.merge(geneList, phase2_hippo, on=['ensemblID'], how='outer').fillna(0)
newH['Hippocampus'] = newH['Hippocampus'].astype('int')
```

```
newCMC = pd.merge(geneList, cmc, on=['ensemblID'], how='outer').fillna(0)
newCMC['CMC DLPFC'] = newCMC['CMC DLPFC'].astype('int')
```

```
print(newC.shape, newH.shape, newD1.shape, newCMC.shape)
```

```
(610, 2) (610, 2) (610, 2) (610, 2)
```

```
[25]: df = pd.concat([newC.set_index(['ensemblID']), newD1.set_index(['ensemblID']),
    ↳ newH.set_index(['ensemblID']), newCMC.set_index(['ensemblID'])],
    axis=1, join='outer')
df.head(2)
```

```
[25]:
```

	Caudate	DLPFC	Hippocampus	CMC DLPFC
ensemblID				
ENSG00000003137	1	0	0	0
ENSG00000003147	0	0	0	1


```
[26]: %R
cmc = subset_pvalue('../.../cmc_dlpfc/female_analysis/metrics_summary/_m/
  ↳female_specific_DE_genes.txt',
                    0.05)

lt = list(Caudate = caudate,
          DLPFC = dlpfc,
          Hippocampus = hippo,
          `CMC DLPFC` = cmc)

m = make_comb_mat(lt)
```

```
[27]: %R
right_annot = upset_right_annotation(
  m, ylim = c(0, 800),
  gp = gpar(fill = "black"),
  annotation_name_side = "bottom",
  axis_param = list(side = "bottom"))

top_annot = upset_top_annotation(
  m, height=unit(7, "cm"),
  ylim = c(0, 800),
  gp=gpar(fill=cbb_palette[comb_degree(m)]),
  annotation_name_rot = 90)

pdf('cmc_sex_tissue_upsetR_DEgenes_femaleSpecific.pdf', width=8, height=5)
ht = draw(UpSet(m, pt_size=unit(6, "mm"), lwd=3,
               comb_col=cbb_palette[comb_degree(m)],
               set_order = c("Caudate", "DLPFC", "Hippocampus", "CMC DLPFC"),
               comb_order = order(-comb_size(m)),
               row_names_gp = gpar(fontsize = 16, fontface='bold'),
               right_annotation = right_annot,
               top_annotation = top_annot))
od = column_order(ht)
cs = comb_size(m)
decorate_annotation("intersection_size", {
  grid.text(cs[od], x = seq_along(cs), y = unit(cs[od], "native") +
    unit(6, "pt"),
    default.units = "native", just = "bottom", gp = gpar(fontsize = 11))
})
dev.off()
```

png
2

```
[28]: %R
right_ha = rowAnnotation(
```

```

    "Intersection\nsize" = anno_barplot(comb_size(m), border=F,
                                       ylim = c(0, 800),
                                       gp=gpar(fill=cbb_palette[comb_degree(m)]),
                                       width = unit(7, "cm")))
top_ha = HeatmapAnnotation(
  "Set size" = anno_barplot(set_size(m), border=F,
                           ylim = c(0, 800),
                           gp = gpar(fill = "black"),
                           height = unit(2, "cm")),
  gap = unit(2, "mm"), annotation_name_side = "left",
  annotation_name_rot = 90)

pdf("cmc_sex_tissue_upsetR_DEgenes_transpose_femaleSpecific.pdf", width=5,
    height=10)
ht = draw(UpSet(t(m), pt_size=unit(5, "mm"), lwd=3,
               comb_order = order(-comb_size(m)),
               comb_col=cbb_palette[comb_degree(m)],
               set_order = c("Caudate", "DLPFC", "Hippocampus", "CMC DLPFC"),
               column_names_gp = gpar(fontsize = 16, fontface='bold'),
               right_annotation = right_ha, top_annotation=top_ha))

od = rev(row_order(ht))
cs = comb_size(m)
decorate_annotation("Intersection\nsize", {
  grid.text(cs[od], y = seq_along(cs), x = unit(cs[od], "native") +
            unit(6, "pt"),
            default.units = "native", just = "left", gp = gpar(fontsize = 11))
})
dev.off()

```

png
2

```

[29]: dft = pd.read_csv('.././././cmc_dlpfc/female_analysis/metrics_summary/_m/
    female_specific_DE_genes.txt',
    sep='\t')
dft['Dir'] = np.sign(dft['t'])
dft.head()

```

```

[29]:

```

	Feature	gencodeID	Symbol	ensemblID	Chrom \
0	ENSG000000153132.12	ENSG000000153132.12	CLGN	ENSG000000153132	chr4
1	ENSG000000179083.6	ENSG000000179083.6	FAM133A	ENSG000000179083	chrX
2	ENSG000000165733.7	ENSG000000165733.7	BMS1	ENSG000000165733	chr10
3	ENSG000000183023.18	ENSG000000183023.18	SLC8A1	ENSG000000183023	chr2
4	ENSG000000236268.5	ENSG000000236268.5	LINC01361	ENSG000000236268	chr1

	logFC	t	adj.P.Val	Male_Pval	Male_FDR	Type	Dir
0	0.389937	5.559139	0.000123	0.283020	0.313711	gene	1.0
1	0.261268	5.004488	0.000535	0.272019	0.302973	gene	1.0
2	0.150918	4.986552	0.000535	0.169787	0.205428	gene	1.0
3	0.245819	4.925477	0.000632	0.082759	0.108963	gene	1.0
4	0.404532	4.865744	0.000700	0.110632	0.140404	gene	1.0

```
[30]: dft.loc[:, ['Feature', 'ensemblID', 'Symbol', 'Chrom', 'Dir']] \
      .merge(pd.DataFrame({'ensemblID': list(set(phase3_caodate['ensemblID']) &
                                             set(cmc['ensemblID']))}),
            on='ensemblID')
```

```
[30]:
```

	Feature	ensemblID	Symbol	Chrom	Dir
0	ENSG000000263006.6	ENSG000000263006	ROCK1P1	chr18	1.0
1	ENSG000000249669.9	ENSG000000249669	CARMN	chr5	-1.0
2	ENSG000000167703.14	ENSG000000167703	SLC43A2	chr17	-1.0

```
[31]: shared_df = dft.loc[:, ['Feature', 'ensemblID', 'Chrom', 'Symbol', 'Dir']] \
      .merge(pd.DataFrame({'ensemblID':
      ↪ list(set(phase3_caodate['ensemblID']) &
                                             set(cmc['ensemblID']))}),
            on='ensemblID')
shared_df.to_csv('cmc_shared_caodate_degs_annotation_femaleSpecific.txt',
      ↪ sep='\t',
            index=False, header=True)
shared_df
```

```
[31]:
```

	Feature	ensemblID	Chrom	Symbol	Dir
0	ENSG000000263006.6	ENSG000000263006	chr18	ROCK1P1	1.0
1	ENSG000000249669.9	ENSG000000249669	chr5	CARMN	-1.0
2	ENSG000000167703.14	ENSG000000167703	chr17	SLC43A2	-1.0

```
[32]: ##### 6 out of 41 are autosomal
dd = np.sum(shared_df.Chrom.isin(['chrX', 'chrY'])) / shared_df.shape[0] * 100
print("%0.2f%% of shared DEG are allosomal!" % dd)
```

0.00% of shared DEG are allosomal!

```
[33]: gtf_annot['ensemblID'] = gtf_annot.gene_id.str.replace("\\..*", "", regex=True)
gtf_annot[["gene_id", 'ensemblID', 'gene_name', 'seqname', 'gene_type']] \
      .merge(df, left_on='ensemblID', right_index=True) \
      .to_csv('cmc_all_deg_across_tissues_femaleSpecific.csv')
```

```
[ ]:
```