main

August 16, 2021

1 Differential Expression with limma-voom pipeline - Genes

1.1 Prepare Data

1.2 Functions

1.2.1 General functions

```
[2]: # Function from jaffelab github
     merge_rse_metrics <- function(rse) {</pre>
         stopifnot(is(rse, 'RangedSummarizedExperiment'))
             c('concordMapRate', 'overallMapRate', 'mitoRate', 'rRNA_rate',
               'totalAssignedGene', 'numMapped', 'numReads', 'numUnmapped',
               'mitoMapped', 'totalMapped') %in%
                 colnames(SummarizedExperiment::colData(rse))
         )
         stopifnot(all(sapply(c(
             'concordMapRate', 'overallMapRate', 'mitoRate', 'rRNA_rate',
             'totalAssignedGene', 'numMapped', 'numReads', 'numUnmapped',
             'mitoMapped', 'totalMapped'), function(var) {
                 is(colData(rse)[, var], 'List')
             })
         ))
         rse$concordMapRate = mapply(function(r, n) {
             sum(r*n)/sum(n)
         }, rse$concordMapRate, rse$numReads)
         rse$overallMapRate = mapply(function(r, n) {
```

```
sum(r*n)/sum(n)
    }, rse$overallMapRate, rse$numReads)
    rse$mitoRate = mapply(function(r, n) {
        sum(r*n)/sum(n)
    }, rse$mitoRate, rse$numMapped)
    rse$rRNA_rate = mapply(function(r, n) {
        sum(r*n)/sum(n)
    }, rse$rRNA_rate, rse$numMapped)
    rse$totalAssignedGene = mapply(function(r, n) {
        sum(r*n)/sum(n)
    }, rse$totalAssignedGene, rse$numMapped)
    rse$numMapped = sapply(rse$numMapped, sum)
    rse$numReads = sapply(rse$numReads, sum)
    rse$numUnmapped = sapply(rse$numUnmapped, sum)
    rse$mitoMapped = sapply(rse$mitoMapped, sum)
    rse$totalMapped = sapply(rse$totalMapped, sum)
    return(rse)
}
```

```
[3]: save_volcanoPlot <- function(top, label, dirname){
         pdf(file=paste0(dirname, "/volcanoPlot_", label, ".pdf"), 8, 6)
         with(top, plot(logFC, -log10(P.Value), pch=20, cex=0.6))
         with(subset(top, adj.P.Val<=0.05), points(logFC, -log10(P.Value),
                                                    pch=20, col='red', cex=0.6))
         with(subset(top, abs(logFC)>0.50), points(logFC, -log10(P.Value),
                                                    pch=20, col='orange', cex=0.6))
         with(subset(top, adj.P.Val<=0.05 & abs(logFC)>0.50),
              points(logFC, -log10(P.Value), pch=20, col='green', cex=0.6))
         dev.off()
     }
     save_MAplot <- function(top, label, dirname){</pre>
         pdf(file=paste0(dirname, "/MAplot_", label, ".pdf"), 8, 6)
         with(top, plot(AveExpr, logFC, pch=20, cex=0.5))
         with(subset(top, adj.P.Val<0.05),</pre>
              points(AveExpr, logFC, col="red", pch=20, cex=0.5))
         dev.off()
     }
     extract_de <- function(contrast, label, efit, seed_int){</pre>
         dirname = paste0("permutation_", stringr::str_pad(seed_int, 2, pad = "0"))
         se <- as.data.frame(sqrt(efit$s2.post) * efit$stdev.unscaled) %>%
             rename("SE"="CtrlvsSZ") %>% rownames_to_column()
         top <- topTable(efit, coef=1, number=Inf, sort.by="P") %>%
             rownames_to_column() %>% inner_join(se, by="rowname") %>%
             column_to_rownames("rowname")
```

1.2.2 Cached functions

```
[4]: load counts <- function(){
         counts_file = paste0("/ceph/projects/v3_phase3_paper/inputs/phase2/_m/
      \hookrightarrow "dlpfc_ribozero_brainseq_phase2_hg38_rseGene_merged_n453.rda")
         load(counts_file)
         rse_df = rse_gene
         return(rse_df)
     memCounts <- memoise(load_counts)</pre>
     get_random_samples <- function(seed_int, new_dir=TRUE){</pre>
         set.seed(seed_int + 113) # seed for reproducibility
         dirname = paste0("permutation_", stringr::str_pad(seed_int, 2, pad = "0"))
         if(new dir){
             dir.create(dirname)
         rse df <- memCounts()</pre>
         keepIndex = which((rse_df$Dx %in% c("Control", "Schizo")) &
                            (rse_df$Age > 17) & (rse_df$Sex == "M") &
                            (rse_df$Race %in% c("AA", "CAUC")))
         snames = sample(keepIndex, 114, replace=FALSE) # subsampling to Female N_{\sqcup}
      \hookrightarrow (sample size)
         return(snames)
     memSamples <- memoise(get_random_samples)</pre>
     get mds <- function(){</pre>
         mds_file = "/ceph/projects/v4_phase3_paper/inputs/genotypes/mds/_m/
      mds = data.table::fread(mds_file) %>%
             rename_at(.vars = vars(starts_with("C")),
```

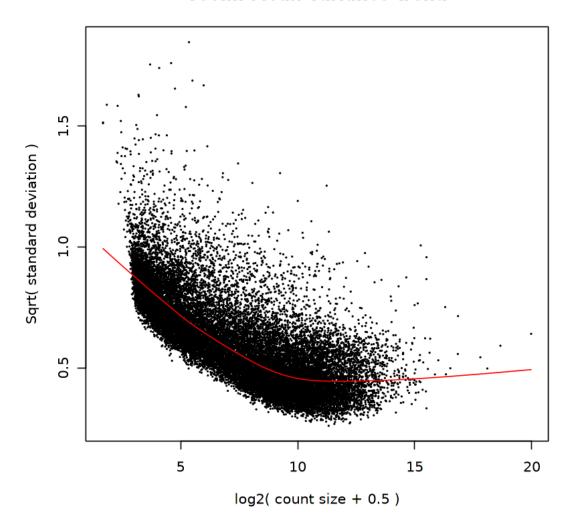
```
function(x){sub("C", "snpPC", x)}) %>%
        mutate_if(is.character, as.factor)
    return(mds)
memMDS <- memoise::memoise(get_mds)</pre>
prep_data <- function(seed_int){</pre>
    rse_df <- memCounts()</pre>
    keepIndex <- memSamples(seed int)</pre>
    rse_df = rse_df[, keepIndex]
    rse df$Dx = factor(rse df$Dx, levels = c("Control", "Schizo"))
    rse_df$Sex <- factor(rse_df$Sex)</pre>
    rse_df <- merge_rse_metrics(rse_df)</pre>
    rse_df$ERCCsumLogErr <- mapply(function(r, n) {</pre>
        sum(r * n)/sum(n)
    }, rse_df$ERCCsumLogErr, rse_df$numReads)
    colData(rse_df)$RIN = sapply(colData(rse_df)$RIN,"[",1)
    rownames(colData(rse_df)) <- sapply(strsplit(rownames(colData(rse_df)),__
 \rightarrow"_"), "[", 1)
    pheno = colData(rse_df) %>% as.data.frame %>%
        inner join(memMDS(), by=c("BrNum"="FID")) %>%
        distinct(RNum, .keep all = TRUE)
    # Generate DGE list
    x <- DGEList(counts=assays(rse_df)$counts[, pheno$RNum],</pre>
                  genes=rowData(rse_df), samples=pheno)
    # Filter by expression
    design0 <- model.matrix(~Dx, data=x$samples)</pre>
    keep.x <- filterByExpr(x, design=design0)</pre>
    x <- x[keep.x, , keep.lib.sizes=FALSE]</pre>
    print(paste('There are:', sum(keep.x), 'features left!', sep=' '))
    # Normalize library size
    x <- calcNormFactors(x, method="TMM")</pre>
    return(x)
}
memo_prepData <- memoise(prep_data)</pre>
SVA model <- function(seed int){
    x <- memo_prepData(seed_int)</pre>
    # Design matrix
    mod = model.matrix(~Dx + Age + mitoRate + rRNA_rate + RIN +
                        totalAssignedGene + overallMapRate + ERCCsumLogErr +
                         snpPC1 + snpPC2 + snpPC3, data=x$samples)
    colnames(mod) <- gsub("Dx", "", colnames(mod))</pre>
    colnames(mod) <- gsub("\\(Intercept\\)", "Intercept", colnames(mod))</pre>
    # Calculated SVs
    null.model = mod %>% as.data.frame %>% select(-c("Schizo")) %>% as.matrix
    n.sv <- num.sv(x$counts, mod, method="be")</pre>
```

```
svobj <- svaseq(x$counts, mod, null.model, n.sv=n.sv)</pre>
    if(svobj$sv == 0){
        modQsva <- mod
    } else {
        modQsva <- cbind(mod, svobj$sv)</pre>
        len.d <- length(colnames(modQsva))</pre>
         colnames(modQsva)[((len.d - n.sv)+1):len.d] <- make.names(paste0("sv",1:</pre>
 \rightarrown.sv))
    return (modQsva)
memo_svaModel <- memoise(SVA_model)</pre>
get_voom <- function(seed_int){</pre>
    ### Preform voom
    x <- memo_prepData(seed_int)</pre>
    modQsva <- memo_svaModel(seed_int)</pre>
    v <- voom(x[, rownames(modQsva)], modQsva, plot=TRUE)</pre>
    return(v)
memo_voom <- memoise(get_voom)</pre>
cal_res <- function(seed_int){</pre>
    ### Calculate residuals
    v <- memo_voom(seed_int)</pre>
    null_model <- v$design %>% as.data.frame %>% select(-c("Schizo")) %>% as.
 →matrix
    fit_res <- lmFit(v, design=null_model)</pre>
    res = v$E - ( fit_res$coefficients %*% t(null_model) )
    res_sd = apply(res, 1, sd)
    res_mean = apply(res, 1, mean)
    res_norm = (res - res_mean) / res_sd
    dirname = paste0("permutation_", stringr::str_pad(seed_int, 2, pad = "0"))
    write.table(res_norm, file=paste0(dirname, '/residualized_expression.tsv'),
                 sep="\t", quote=FALSE)
memo_res <- memoise(cal_res)</pre>
fit_voom <- function(seed_int){</pre>
    v <- memo_voom(seed_int)</pre>
    modQsva <- memo_svaModel(seed_int)</pre>
    fit0 <- lmFit(v, modQsva)</pre>
    contr.matrix <- makeContrasts(CtrlvsSZ = Schizo,</pre>
                                     levels=colnames(modQsva))
    fit <- contrasts.fit(fit0, contrasts=contr.matrix)</pre>
    esv <- eBayes(fit)</pre>
    return(esv)
```

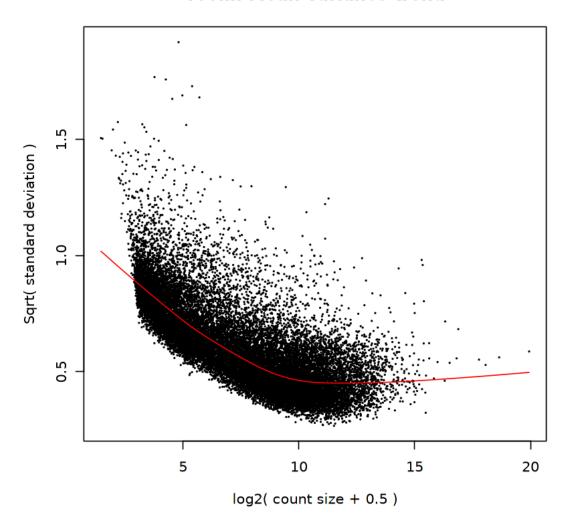
```
}
memo_efit <- memoise(fit_voom)</pre>
```

1.3 Differential Expression Analysis

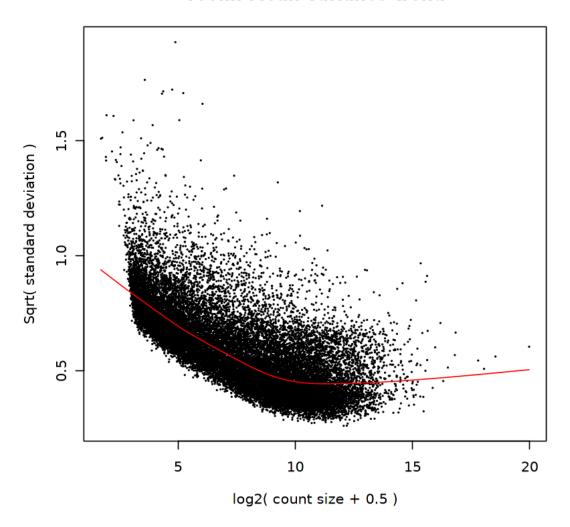
```
[5]: for(seed_int in seq(1, 10)){
         # Preform voom
        v <- memo_voom(seed_int)</pre>
        dirname = paste0("permutation_", stringr::str_pad(seed_int, 2, pad = "0"))
         save(v, file=paste0(dirname,'/voomSVA.RData'))
         # Fit model and apply eBayes
        efit = memo efit(seed int)
         # Save differential expression
        extract_de(1, "CtrlvsSZ", efit, seed_int)
        # Calculate residuals
        memo_res(seed_int)
     }
    [1] "There are: 22545 features left!"
    Number of significant surrogate variables is: 1
    Iteration (out of 5 ):1 2 3 4 5
    Warning message in if (svobj$sv == 0) {:
    "the condition has length > 1 and only the first element will be used"
    [1] "Comparison for: CtrlvsSZ"
    [1] "There are: 1 DE features!"
    [1] "There are: 22470 features left!"
    Number of significant surrogate variables is: 1
    Iteration (out of 5 ):1 2 3 4 5
    Warning message in if (svobj$sv == 0) {:
    "the condition has length > 1 and only the first element will be used"
    [1] "Comparison for: CtrlvsSZ"
    [1] "There are: 0 DE features!"
    [1] "There are: 22378 features left!"
    Number of significant surrogate variables is: 1
    Iteration (out of 5 ):1 2 3 4 5
    Warning message in if (svobj$sv == 0) {:
    "the condition has length > 1 and only the first element will be used"
```



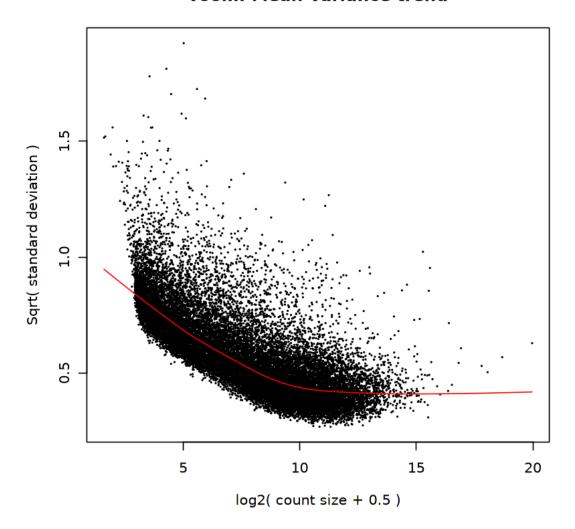
```
[1] "Comparison for: CtrlvsSZ"
[1] "There are: 0 DE features!"
[1] "There are: 22547 features left!"
Number of significant surrogate variables is: 1
Iteration (out of 5 ):1 2 3 4 5
Warning message in if (svobj$sv == 0) {:
"the condition has length > 1 and only the first element will be used"
```



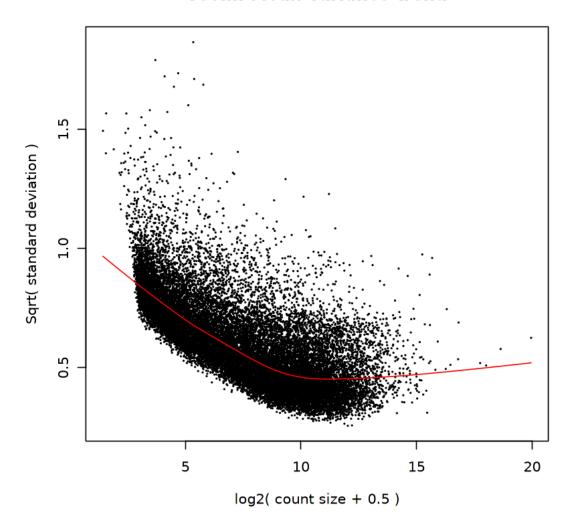
```
[1] "Comparison for: CtrlvsSZ"
[1] "There are: 328 DE features!"
[1] "There are: 22638 features left!"
Number of significant surrogate variables is: 1
Iteration (out of 5 ):1 2 3 4 5
Warning message in if (svobj$sv == 0) {:
"the condition has length > 1 and only the first element will be used"
```



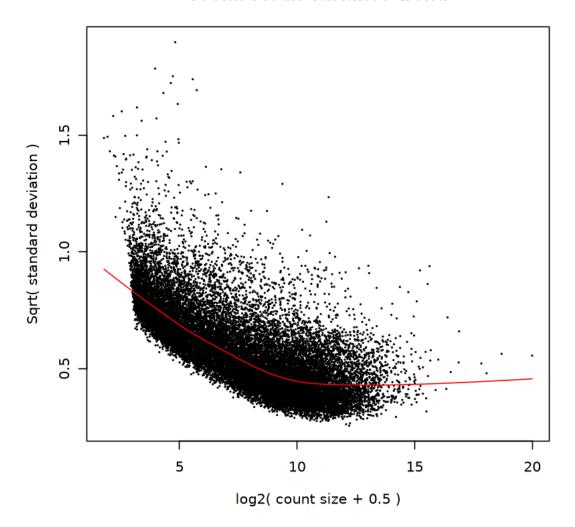
```
[1] "Comparison for: CtrlvsSZ"
[1] "There are: 6 DE features!"
[1] "There are: 22376 features left!"
Number of significant surrogate variables is: 1
Iteration (out of 5 ):1 2 3 4 5
Warning message in if (svobj$sv == 0) {:
"the condition has length > 1 and only the first element will be used"
```



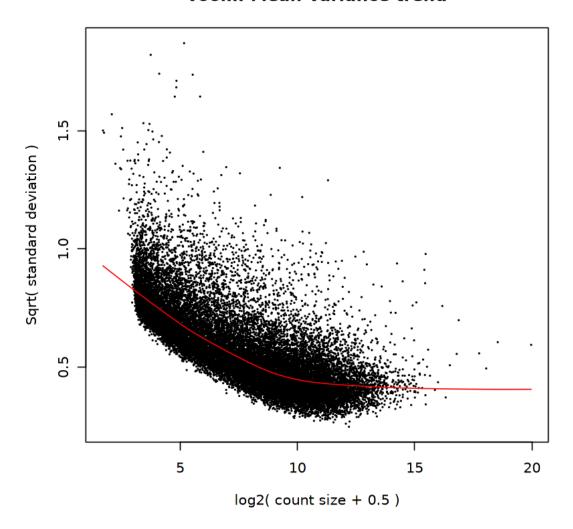
```
[1] "Comparison for: CtrlvsSZ"
[1] "There are: 0 DE features!"
[1] "There are: 22198 features left!"
Number of significant surrogate variables is: 1
Iteration (out of 5 ):1 2 3 4 5
Warning message in if (svobj$sv == 0) {:
"the condition has length > 1 and only the first element will be used"
```



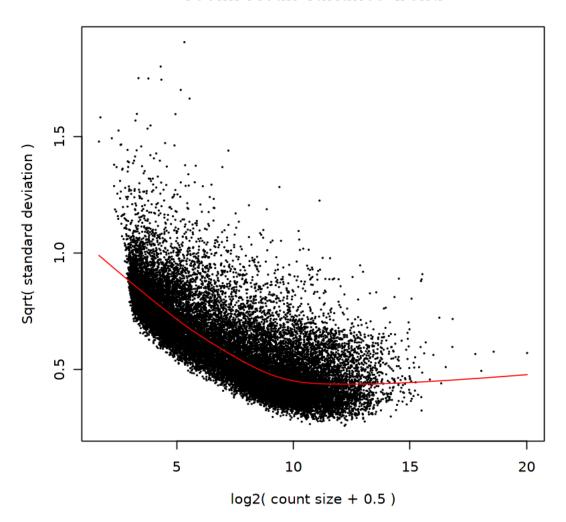
```
[1] "Comparison for: CtrlvsSZ"
[1] "There are: 2 DE features!"
[1] "There are: 22349 features left!"
Number of significant surrogate variables is: 1
Iteration (out of 5 ):1 2 3 4 5
Warning message in if (svobj$sv == 0) {:
"the condition has length > 1 and only the first element will be used"
```



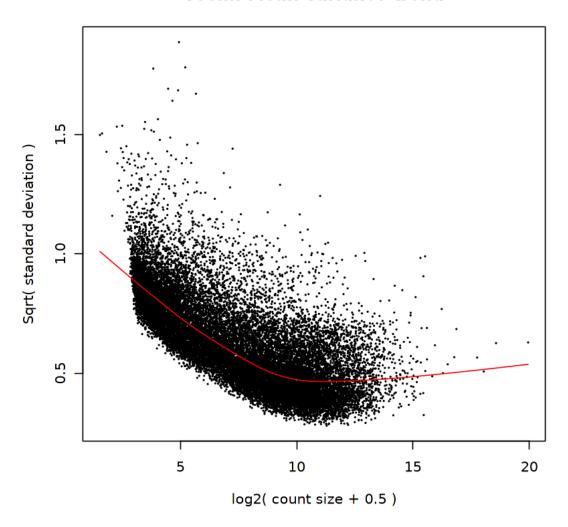
```
[1] "Comparison for: CtrlvsSZ"
[1] "There are: 146 DE features!"
[1] "There are: 22354 features left!"
Number of significant surrogate variables is: 1
Iteration (out of 5 ):1 2 3 4 5
Warning message in if (svobj$sv == 0) {:
"the condition has length > 1 and only the first element will be used"
```

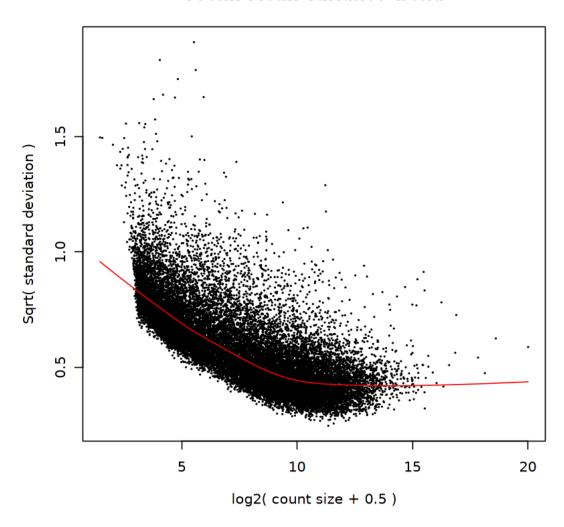


```
[1] "Comparison for: CtrlvsSZ"
[1] "There are: 2 DE features!"
[1] "There are: 22504 features left!"
Number of significant surrogate variables is: 1
Iteration (out of 5 ):1 2 3 4 5
Warning message in if (svobj$sv == 0) {:
"the condition has length > 1 and only the first element will be used"
```



- [1] "Comparison for: CtrlvsSZ"
- [1] "There are: 5 DE features!"





1.4 Repreducibility Information

```
[6]: Sys.time()
    proc.time()
    options(width = 120)
    sessioninfo::session_info()

[1] "2021-08-16 16:55:49 EDT"

    user system elapsed
    8984.190 910.264 2068.094

    Session info
    setting value
```

version R version 4.0.3 (2020-10-10)

os Arch Linux

system x86_64, linux-gnu

ui X11 language (EN)

collate en_US.UTF-8
ctype en_US.UTF-8
tz America/New_York

date 2021-08-16

Packages package

| package | * | version | date | lib | source |
|--------------------------|---|----------|------------|-----|----------------|
| annotate | | 1.68.0 | 2020-10-27 | [1] | Bioconductor |
| AnnotationDbi | | 1.52.0 | 2020-10-27 | [1] | Bioconductor |
| assertthat | | 0.2.1 | 2019-03-21 | [1] | CRAN (R 4.0.2) |
| backports | | 1.2.1 | 2020-12-09 | [1] | CRAN (R 4.0.2) |
| base64enc | | 0.1-3 | 2015-07-28 | [1] | CRAN (R 4.0.2) |
| Biobase | * | 2.50.0 | 2020-10-27 | [1] | Bioconductor |
| BiocGenerics | * | 0.36.1 | 2021-04-16 | [1] | Bioconductor |
| BiocParallel | * | 1.24.1 | 2020-11-06 | [1] | Bioconductor |
| bit | | 4.0.4 | 2020-08-04 | [1] | CRAN (R 4.0.2) |
| bit64 | | 4.0.5 | 2020-08-30 | [1] | CRAN (R 4.0.2) |
| bitops | | 1.0-7 | 2021-04-24 | [1] | CRAN (R 4.0.3) |
| blob | | 1.2.1 | 2020-01-20 | [1] | CRAN (R 4.0.2) |
| broom | | 0.7.8 | 2021-06-24 | [1] | CRAN (R 4.0.3) |
| cachem | | 1.0.5 | 2021-05-15 | [1] | CRAN (R 4.0.3) |
| Cairo | | 1.5-12.2 | 2020-07-07 | [1] | CRAN (R 4.0.2) |
| cellranger | | 1.1.0 | 2016-07-27 | [1] | CRAN (R 4.0.2) |
| cli | | 3.0.0 | 2021-06-30 | [1] | CRAN (R 4.0.3) |
| colorspace | | 2.0-2 | 2021-06-24 | [1] | CRAN (R 4.0.3) |
| crayon | | 1.4.1 | 2021-02-08 | [1] | CRAN (R 4.0.3) |
| data.table | * | 1.14.0 | 2021-02-21 | [1] | CRAN (R 4.0.3) |
| DBI | | 1.1.1 | 2021-01-15 | [1] | CRAN (R 4.0.2) |
| dbplyr | | 2.1.1 | 2021-04-06 | [1] | CRAN (R 4.0.3) |
| DelayedArray | | 0.16.3 | 2021-03-24 | [1] | Bioconductor |
| digest | | 0.6.27 | 2020-10-24 | [1] | CRAN (R 4.0.2) |
| dplyr | * | 1.0.7 | 2021-06-18 | [1] | CRAN (R 4.0.3) |
| edgeR | * | 3.32.1 | 2021-01-14 | [1] | Bioconductor |
| ellipsis | | 0.3.2 | 2021-04-29 | [1] | CRAN (R 4.0.3) |
| evaluate | | 0.14 | 2019-05-28 | [1] | CRAN (R 4.0.2) |
| fansi | | 0.5.0 | 2021-05-25 | [1] | CRAN (R 4.0.3) |
| fastmap | | 1.1.0 | 2021-01-25 | [1] | CRAN (R 4.0.2) |
| forcats | * | 0.5.1 | 2021-01-27 | [1] | CRAN (R 4.0.2) |
| fs | | 1.5.0 | 2020-07-31 | [1] | CRAN (R 4.0.2) |
| genefilter | * | 1.72.1 | 2021-01-21 | [1] | Bioconductor |
| generics | | 0.1.0 | 2020-10-31 | [1] | CRAN (R 4.0.2) |
| GenomeInfoDb | * | 1.26.7 | 2021-04-08 | [1] | Bioconductor |
| ${\tt GenomeInfoDbData}$ | | 1.2.4 | 2021-02-02 | [1] | Bioconductor |

| GenomicRanges | * | 1.42.0 | 2020-10-27 | [1] | Bioconductor |
|-----------------------|-----|----------|------------|-----|---|
| ggplot2 | | 3.3.5 | 2021-06-25 | [1] | CRAN (R 4.0.3) |
| glue | .,. | 1.4.2 | 2020-08-27 | [1] | CRAN (R 4.0.2) |
| gtable | | 0.3.0 | 2019-03-25 | [1] | CRAN (R 4.0.2) |
| haven | | 2.4.1 | 2019 03 23 | [1] | CRAN (R 4.0.3) |
| hms | | 1.1.0 | 2021-04-23 | | CRAN (R 4.0.3) |
| htmltools | | 0.5.1.1 | 2021-03-17 | | CRAN (R 4.0.2) |
| httr | | 1.4.2 | 2021-01-22 | | CRAN (R 4.0.2) |
| | J. | 2.24.1 | | | Bioconductor |
| IRanges | • | | | [1] | CRAN (R 4.0.2) |
| IRdisplay IRkernel | | 1.0 | 2021-01-20 | | |
| | | | 2021-05-11 | [1] | CRAN (R 4.0.3) |
| jsonlite | | 1.7.2 | 2020-12-09 | | CRAN (R 4.0.2) |
| lattice | | 0.20-41 | 2020-04-02 | | CRAN (R 4.0.3) |
| lifecycle | | 1.0.0 | 2021-02-15 | [1] | CRAN (R 4.0.3) |
| limma | * | 3.46.0 | 2020-10-27 | | Bioconductor |
| locfit | | 1.5-9.4 | 2020-03-25 | | CRAN (R 4.0.2) |
| lubridate | | 1.7.10 | 2021-02-26 | | CRAN (R 4.0.3) |
| magrittr | | 2.0.1 | 2020-11-17 | | CRAN (R 4.0.2) |
| Matrix | | 1.3-4 | 2021-06-01 | [1] | CRAN (R 4.0.3) |
| MatrixGenerics | * | 1.2.1 | 2021-01-30 | [1] | |
| matrixStats | * | 0.59.0 | 2021-06-01 | [1] | CRAN (R 4.0.3) |
| memoise | * | 2.0.0 | 2021-01-26 | [1] | CRAN (R 4.0.2) |
| mgcv | * | 1.8-33 | 2020-08-27 | | CRAN (R 4.0.3) |
| modelr | | 0.1.8 | 2020-05-19 | | |
| munsell | | 0.5.0 | 2018-06-12 | | CRAN (R 4.0.2) |
| nlme | * | 3.1-152 | 2021-02-04 | [1] | CRAN (R 4.0.3) |
| pbdZMQ | | 0.3-5 | 2021-02-10 | [1] | CRAN (R 4.0.3) |
| pillar | | 1.6.1 | 2021-05-16 | [1] | CRAN (R 4.0.3) |
| pkgconfig | | 2.0.3 | 2019-09-22 | [1] | CRAN (R 4.0.2) |
| purrr | * | 0.3.4 | 2020-04-17 | [1] | CRAN (R 4.0.2) |
| R6 | | 2.5.0 | 2020-10-28 | [1] | CRAN (R 4.0.2) |
| Rcpp | | 1.0.7 | 2021-07-07 | | CRAN (R 4.0.3) |
| RCurl | | 1.98-1.3 | 2021-03-16 | [1] | CRAN (R 4.0.3) |
| readr | * | 1.4.0 | 2020-10-05 | [1] | CRAN (R 4.0.2) |
| readxl | | 1.3.1 | 2019-03-13 | [1] | CRAN (R 4.0.2) |
| repr | | 1.1.3 | 2021-01-21 | [1] | CRAN (R 4.0.2) |
| reprex | | 2.0.0 | 2021-04-02 | [1] | CRAN (R 4.0.3) |
| rlang | | 0.4.11 | 2021-04-30 | [1] | CRAN (R 4.0.3) |
| RSQLite | | 2.2.7 | 2021-04-22 | [1] | CRAN (R 4.0.3) |
| rstudioapi | | 0.13 | 2020-11-12 | [1] | CRAN (R 4.0.2) |
| rvest | | 1.0.0 | 2021-03-09 | [1] | CRAN (R 4.0.3) |
| S4Vectors | * | 0.28.1 | 2020-12-09 | [1] | Bioconductor |
| scales | | 1.1.1 | 2020-05-11 | | CRAN (R 4.0.2) |
| sessioninfo | | 1.1.1 | | | CRAN (R 4.0.2) |
| stringi | | 1.7.3 | 2021-07-16 | | |
| stringr | * | 1.4.0 | 2019-02-10 | | |
| SummarizedExperiment | | | | | Bioconductor |
| survival | | 3.2-7 | | | CRAN (R 4.0.3) |
| | | | | | , |

```
* 3.38.0
                                2020-10-27 [1] Bioconductor
sva
tibble
                     * 3.1.2
                                2021-05-16 [1] CRAN (R 4.0.3)
                     * 1.1.3
                                2021-03-03 [1] CRAN (R 4.0.3)
tidyr
tidyselect
                       1.1.1
                                2021-04-30 [1] CRAN (R 4.0.3)
tidyverse
                     * 1.3.1
                                2021-04-15 [1] CRAN (R 4.0.3)
                                2021-03-12 [1] CRAN (R 4.0.3)
utf8
                       1.2.1
uuid
                       0.1-4
                                2020-02-26 [1] CRAN (R 4.0.2)
                                2021-04-29 [1] CRAN (R 4.0.3)
                       0.3.8
vctrs
withr
                       2.4.2
                                2021-04-18 [1] CRAN (R 4.0.3)
XML
                       3.99-0.6 2021-03-16 [1] CRAN (R 4.0.3)
                                2020-04-23 [1] CRAN (R 4.0.2)
xm12
                       1.3.2
xtable
                       1.8-4
                                2019-04-21 [1] CRAN (R 4.0.2)
                                2020-10-27 [1] Bioconductor
XVector
                       0.30.0
                       1.36.0
                                2020-10-27 [1] Bioconductor
zlibbioc
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- [1] /home/jbenja13/R/x86_64-pc-linux-gnu-library/4.0
- [2] /usr/lib/R/library