# Introduction

1. Eutrophication can have serious consequences on aquatic ecosystem health including decreased levels of dissolved oxygen, formation of toxic compounds, changes in abundance and composition of various aquatic organisms, and overall loss of biodiversity (Camargo & Alonso, 2006).
   1. Anthropogenically caused nutrient enrichment is one of the biggest threats to freshwaters today (Smith & Schindler, 2009) and can lead to more frequent and intense harmful algal blooms, one of the greatest risks to freshwaters biodiversity across the world (Reid et al., 2019).
   2. In the United States, an estimated $2.2 billion in losses because of eutrophication is likely an underestimate of the actual amount. Costs are associated with recreation, fisheries, property values, loss of biodiversity, and drinking water treatment (Dodds et al., 2008).
      1. And determining economic value of freshwater is difficult with many important factors often excluded from these analyses (Keiser et al., 2019).
   3. Many freshwaters across the United States are generally at serious risk of or are already plagued by eutrophication.
      1. Median TN and TP concentrations in lakes exceeded reference values across all ecoregions in 2008 study (Dodds et al., 2008).
      2. In streams, chlorophyll-a concentrations were found to be substantially higher above the thresholds of 30 µg/L of P and 40 µg/L of N (Dodds et al., 2011).
2. Enrichment of N and P are mainly responsible for eutrophication (Wetzel, 2001). The elements are not mutually exclusive, however, as their cycles are coupled in the environment (Oviedo-Vargas et al., 2013); and studying the relative abundances may unfold large-scale patterns that would otherwise be unseen.
   1. Reductions in P pollution in large lakes may lead to the accumulation of N (Finlay et al., 2013), negating the attempt to decrease nutrient pollution.
   2. Large-scale nutrient stoichiometry integrates biogeochemical processes and serves as the backdrop for many smaller-scale processes to occur (Sterner & Elser, 2002).
      1. Ecosystem stoichiometry varies with temporal scale and can be impacted by things such as biotic dynamics, precipitation, geological weathering, anthropogenic influences (Sterner & Elser, 2002).
   3. Although single nutrient concentration patterns are relatively well-known across regions in the US, nutrient stoichiometry is much more difficult to predict (Collins et al., 2017).
3. Behavior of nutrients in freshwaters can be influenced depending on a variety of factors including the surrounding landscape, legacy storage (Lin et al., 2021), precipitation, biogeochemistry, source water (Basu et al., 2011), and residence time (Maranger et al., 2018).
   1. Omernik’s development of ecoregions provides a qualitative understanding of spatial patterns and regional homogeneities that can be used to inform freshwater researchers (Omernik, 1987).
   2. Although difficult to predict, nutrient stoichiometry does show some patterns that may be useful for assessing trophic status of lakes.
      1. Additions of N and P to lakes has demonstrated much greater impact on productivity in lakes than addition of a single element (Elser et al., 2011).
      2. TN:TP ratios tend to be high in oligotrophic lakes and low in eutrophic lakes, indicating potential shifts in limitation from P to N as trophic status increases (Downing & McCauley, 1992).
         1. TN:TP can be used to indicate nutrient deficiency based on the Redfield ratio, which illustrates balanced growth of marine algal cells have a 106C:16N:1P molar ratio (Redfield, 1958).
      3. Generally, increased residence time correlates with increased C:N, C:P, and N:P. Residence time may also promote burial of P and lead to higher rates of primary productivity (Maranger et al., 2018).
4. In this study, we use US Environmental Protection Agency (EPA) National Lakes Assessment (NLA) data to evaluate patterns of nutrient stoichiometry in relation to trophic status in lakes across the US, as balancing nutrient stoichiometry may assist in eutrophication remediation (Carpenter et al., 2011).
   1. Intended to “support efforts to assess nutrient water quality and more effectively protect and restore waters from nutrient pollution.” (wording from challenge description)
   2. We aim to answer the following questions:
      1. How does nutrient limitation/enrichment vary across ecoregions and what are the underlying mechanisms?
      2. Is trophic status (based on chlorophyll) more influenced by nitrogen or phosphorus and how/why does this relationship vary spatially?
      3. What are the trends of stoichiometry and trophic levels across ecoregional and the national scale?

# Methods

1. Data
   1. US EPA NLA data 2007, 2012, 2017
   2. # lakes sampled
      1. 2007: # lakes surveyed = 1156, 95 resampled in same year, 124 reference lakes
      2. 2012: # lakes surveyed = 1038, 100 resampled in same year, 0 reference lakes
      3. 2017: # lakes surveyed = 1112, 97 resampled in same year, 108 (hand) reference lakes?
      4. Lakes in 2007 and 2012 = 364
      5. Lakes in 2012 and 2017 = 473
      6. Lakes in 2007 and 2017 = 282
      7. Lakes in all 3 years = 234
   3. Lake sizes sampled
      1. In 2007, lakes greater than 4 ha were sampled. This changed in the 2012 and 2017 surveys and lakes with surface area > 1 ha and 1-m deep were sampled
         1. Sampling programs often exhibit similar biases including when and which lakes are sampled. Most lake data are collected throughout in the summer and from large lakes (>20 ha) (Stanley et al., 2019). – NLA data follows the collection during summer, but breaks away from the large lakes sampling only by including smaller lakes (<20 ha).
   4. Lakes sampled during the summer (May-September, with a handful of sampling events in October – 4 in 2017 and 9 in 2007) in each year
2. Site selection
   1. Generalized Random Tessellation Stratified survey design (p. 3 technical doc) to randomly choose sampling sites. (USEPA, 2022)
      1. Stratification based on omernik level-3 aggregated ecoregions, state, and lake size
      2. Each lake is assigned a weight to indicate the # lakes it represents
         1. NLA adjusted site weights will be used to broaden the results to regional and national extents
3. Sampling and laboratory methods
   1. Standardized sampling protocols p.37 manuals (USEPA, 2007b, 2011, 2017a)
      1. Water was collected using an integrated sampler within the euphotic zone (up to 2 m).
      2. Chlorophyll sample is placed in a dark 2L bottle and stored on ice until filtration occurred -- Chlorophyll samples filtered in the field with 0.4 µm pore size polycarbonate filters??? Double check this.
      3. Nutrients sample is placed ino a 250 mL bottle and sulfuric acid is added to stabilize the sample at pH <2 and stored on ice
   2. Standardized lab protocols p.51 manuals (USEPA, 2007a, 2012, 2017b)
      1. Shipped overnight to approved laboratories and processed within 24 hours
         1. Laboratory processing procedures must maintain quality assurance/control outlined by the EPA.
      2. Chlorophyll a is analyzed via extraction in 90% acetone followed by fluorometry
      3. TN and TP (no3 and nh4) analyzed via persulfate digestion then automaticed colorimetric analysis
4. Trophic state calculation
   1. p.80 technical doc
5. Statistical/Data analyses
   1. R programming (R Core Team, 2022)
   2. tidyverse (Wickham et al., 2019)
   3. Spsurvey (Dumelle et al., 2022)
      1. Calculated change in trophic levels at the aggregated ecoregional scale and national scale using change.analysis function in the R package spsurvey
         1. This analysis and package uses the stratified randomized weighting of lakes p 133 tech doc
   4. More TBD as results come in
   5. Use N:P to assess nutrient limitation and stoichiometry at national and ecoregional scales
   6. Analyze nutrient limitation in relation to trophic state
   7. Analyze stoichiometric shifts across time to evaluate the condition of waters

# Results – very preliminary – I need help determining what tests to run and what else to look at

1. How does nutrient limitation/enrichment vary across ecoregions and what are the underlying mechanisms?
   1. Figure showing N limitation and P limitation + citations used for justification (e.g. Bergstrom, McCauley)
   2. Sp survey of limitation shifts over time – national, ecoregional
   3. Trophic status in N-limited, P-limited
      1. Ecoregional, national
      2. Changes in these numbers through the years
2. Is trophic status (based on chlorophyll) more influenced by nitrogen or phosphorus and how/why does this relationship vary spatially?
   1. TN:TP ratio vs TS
      1. Ecoregion, nationally (all data?)
   2. TN vs TS
      1. Ecoregion, nationally (all data?)
   3. TP vs TS
      1. Ecoregion, nationally (all data?)
3. What are the trends of stoichiometry and trophic levels across ecoregional and the national scale?
   1. Trophic status across ecoregions, national
   2. Look at urban vs. non urban
   3. % development and % ag
   4. Elevation

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