2-25 Bioscreen for dominance

FRIDAY, 2019-08-16

- [x] Thaw all lines from aliquot 1 in the -80°C
- [x] Inoculate 10 µL of culture into 2 mL of YPAD.
- [x] Grow in shaker at 30°C for two nights.

SUNDAY, 2019-08-18

- [x] Transfer 1 mL of culture into 1.5 mL capped eppendorftube.
- [x] Arrange tubes accoring to Day 1 schedule.
- [x] Run according to Bioscreen
- [x] Arrange tubes according to Day 2 schedule
- [x] Put tubes in fridge

MONDAY, 2019-08-19

For another 11 days.

Bioscreen

Introduction

This protocol describes the prodedure of setting up a bioscreen from saturated culture.

Materials

- Bioscreen plates
- 0.2 mL PCR tubes, without lid
- ddH_20
- YPAD
- 200 µL pipette tips
- 2.5 mL Finntip stepper tip

Procedure

Dilution

1. Fill all 0.2 mL PCR tubes with 150 µL of ddH_2O using the finntip stepper (set on 3).
2. Vortex tube with culture.
3. Transfer 15 µL of saturated culture into a filled PCR tube. Pipette up and down until homogenized.

Prepare plate

4. Transfer 15 µL from diluted culture into assigned well in the Bioscreen plate (this can be done immediately after step 3, using the same pipette tip, and the PCR tube can be disposed of directly after use).
5. Repeat step 2-4 for all samples.
6. Fill all bioscreen wells with 150 µL of YPAD using Finntip stepper.

Run machine

7. Put bioscreen plate into machine (double check direction). Run for 1 day at 30°C, 200 samples, with Medium continuous shaking. Save in folder Linnea -> Dominance -> and label "dominance_dayX_machineY"