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Deep Learning Project
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Malaria Cells Image Detection

Mit CNN Algorithm



Inhalte

- Motivation
- Datensatz erklären
 - Code erklären
- Visualisierungen erklären
 - Interpretation
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Malaria Cells Image Detection

- Die Problematik Malaria, ist immernoch ein Thema in Tropische und Sud Sahara Länder.
 - Malaria ist ein wichtiger Faktor für die höhe Sterberate in Afrika beispielweise.
 - Jede effektive Methode Malaria infizierte Zellen zu identifizieren, hilft bei der Bekämpfung
- ...

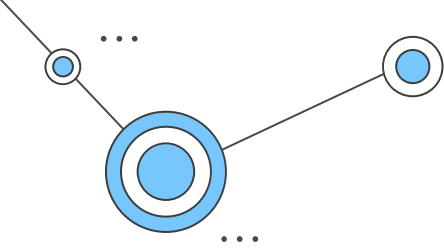
WORLD MALARIA REPORT

2019

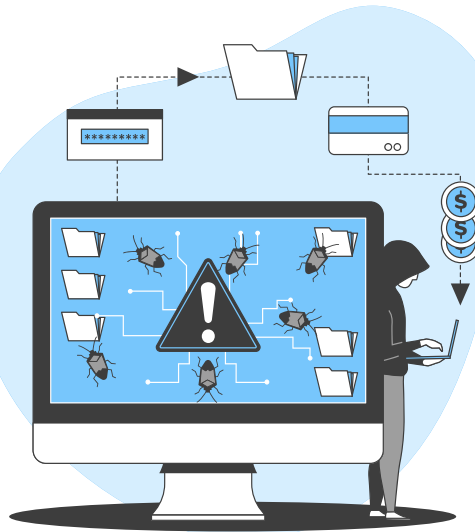


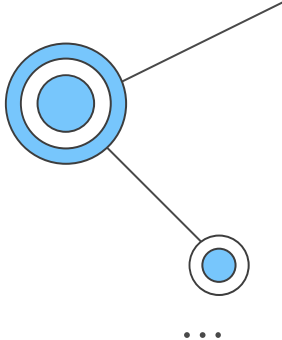
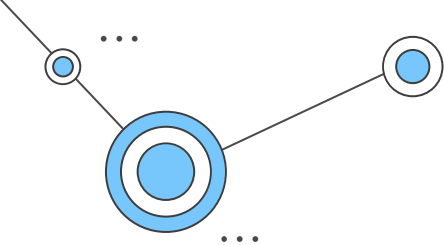
World Health
Organization



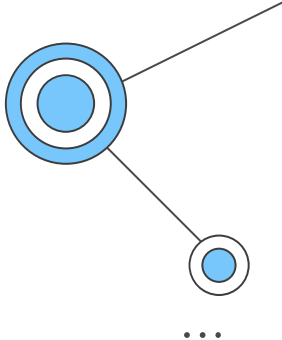
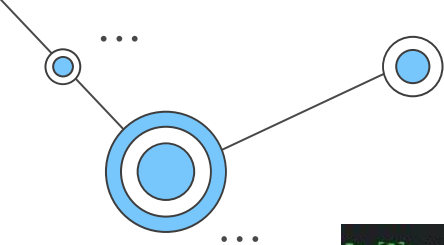


```
untitled0.py X  Projekt Thesis_DeepLearning_V1.py* X  Projekt Thesis_DeepLearning_V1_1.py* X
31
32 # Import Libraries
33
34 import matplotlib.pyplot as plt
35 import pandas as pd
36 import numpy as np
37 import seaborn as sns
38 import cv2
39 import os
40 import PIL
41 import pathlib
42 import tensorflow as tf
43 from tensorflow import keras
44 from sklearn.metrics import confusion_matrix, ConfusionMatrixDisplay
45 from sklearn.model_selection import train_test_split
46
47 import warnings
48 warnings.filterwarnings("ignore")
49 from matplotlib.pyplot import figure
50
51
52
53
54 data = pathlib.Path('C:/Users/alfa/Desktop/Project/cell_images') # Naming the path of the dataset
55
56
57
58 image_count = len(list(data.glob('*/*.png'))) # Counting the dataset
59 print(image_count)
60
61
62 parasitized = list(data.glob('Parasitized/*')) # Visualizing the morphology of a parasitized cell
63 PIL.Image.open(str(parasitized[0]))
64
65 uninfected = list(data.glob('Uninfected/*')) #Visualizing the morphology of an uninfected cell
66 PIL.Image.open(str(uninfected[0]))
67
```





```
2022-08-04 22:27:41.945688: W tensorflow/stream_executor/platform/default/dso_loader.cc:64] Could not load
dynamic library 'cudart64_110.dll'; dlerror: cudart64_110.dll not found
2022-08-04 22:27:41.945907: I tensorflow/stream_executor/cuda/cudart_stub.cc:29] Ignore above cudart dlerror
if you do not have a GPU set up on your machine.
27558
Found 27558 files belonging to 2 classes.
Using 22047 files for training.
```



```
In [2]: parasitized = list(data.glob('Parasitized/*'))    # Visualizing the morphology of a parasitized cell
...: PIL.Image.open(str(parasitized[0]))
Out[2]:
```

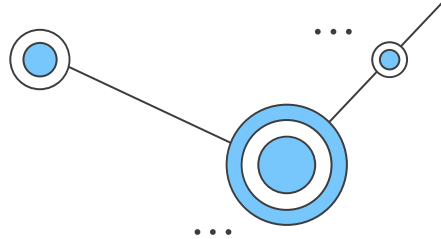
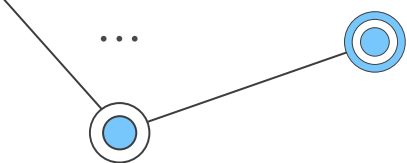


```
In [3]: uninfected = list(data.glob('Uninfected/*'))    #Visualizing the morphology of an uninfected cell
...: PIL.Image.open(str(uninfected[0]))
Out[3]:
```



```
67
68
69
70 # Data Loading
71 # Creating Dataset
72
73 batch_size = 200
74 img_height = 75
75 img_width = 75
76
77 train_ds = tf.keras.utils.image_dataset_from_directory(
78     data,
79     validation_split=0.2,
80     subset="training",
81     seed=123,
82     image_size=(img_height, img_width),
83     batch_size=batch_size)
84
85
86
87
88 val_ds = tf.keras.utils.image_dataset_from_directory(
89     data,
90     validation_split=0.2,
91     subset="validation",
92     seed=123,
93     image_size=(img_height, img_width),
94     batch_size=batch_size)
95
96
97 class_names = train_ds.class_names
98 print(class_names)
99
100
```



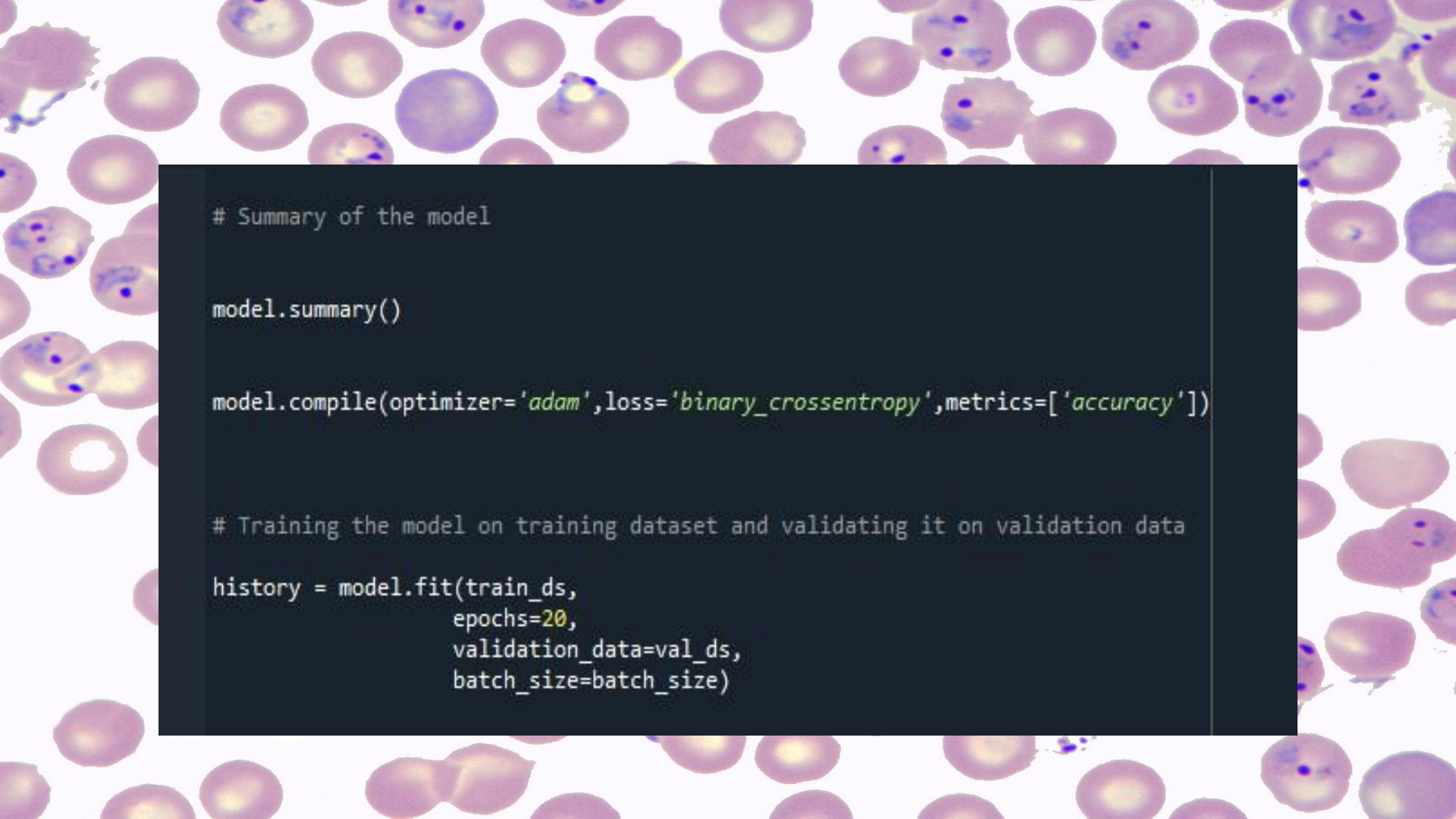


```
To enable them in other operations, rebuild TensorFlow with the appropriate compiler flags.  
Found 27558 files belonging to 2 classes.  
Using 5511 files for validation.  
['Parasitized', 'Uninfected']  
Model: "sequential"
```



```
100
101
102 # Defining the model architecture
103
104 model = keras.Sequential()
105
106 model.add(keras.layers.Conv2D(16, kernel_size=3, strides=1, activation=None,
107                               input_shape=(75,75,3)))
108 model.add(keras.layers.BatchNormalization())
109 model.add(keras.layers.Activation('relu'))
110 model.add(keras.layers.MaxPool2D(2,2))
111 model.add(keras.layers.Dropout(0.2))
112
113 model.add(keras.layers.Conv2D(32, kernel_size=3, strides=1, activation=None))
114 model.add(keras.layers.BatchNormalization())
115 model.add(keras.layers.Activation('relu'))
116
117 model.add(keras.layers.Conv2D(64, kernel_size=3, strides=1, activation=None))
118 model.add(keras.layers.BatchNormalization())
119 model.add(keras.layers.Activation('relu'))
120 model.add(keras.layers.MaxPool2D(2,2))
121 model.add(keras.layers.Dropout(0.3))
122
123 model.add(keras.layers.Conv2D(32, kernel_size=3, strides=1, activation=None))
124 model.add(keras.layers.BatchNormalization())
125 model.add(keras.layers.Activation('relu'))
126
127 model.add(keras.layers.Conv2D(16, kernel_size=3, strides=1, activation=None))
128 model.add(keras.layers.BatchNormalization())
129 model.add(keras.layers.Activation('relu'))
130 model.add(keras.layers.MaxPool2D(2,2))
131 model.add(keras.layers.Dropout(0.3))
132
133 model.add(keras.layers.Conv2D(8, kernel_size=3, strides=1, activation=None))
134 model.add(keras.layers.BatchNormalization())
135 model.add(keras.layers.Activation('relu'))
136
137 model.add(keras.layers.Flatten())
138 model.add(keras.layers.Dense(64, activation='relu'))
139 model.add(keras.layers.Dropout(0.5))
140
141 model.add(keras.layers.Dense(units=1))
142 model.add(keras.layers.Activation('sigmoid'))
143
```



A microscopic background image showing numerous cells, likely blood cells, with purple nuclei and yellowish cytoplasm or extracellular matrix.

```
# Summary of the model
```

```
model.summary()
```

```
model.compile(optimizer='adam', loss='binary_crossentropy', metrics=['accuracy'])
```

```
# Training the model on training dataset and validating it on validation data
```

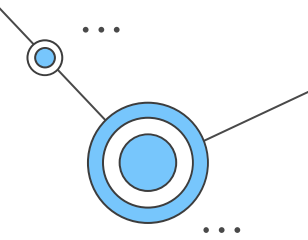
```
history = model.fit(train_ds,  
                    epochs=20,  
                    validation_data=val_ds,  
                    batch_size=batch_size)
```

Model: "sequential"

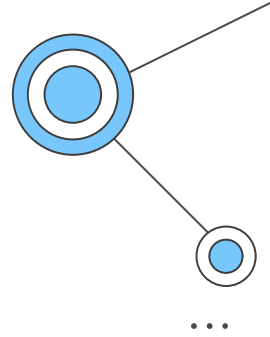
Layer (type)	Output Shape	Param #
conv2d (Conv2D)	(None, 73, 73, 16)	448
batch_normalization (Batch Normalization)	(None, 73, 73, 16)	64
activation (Activation)	(None, 73, 73, 16)	0
max_pooling2d (MaxPooling2D)	(None, 36, 36, 16)	0
dropout (Dropout)	(None, 36, 36, 16)	0
conv2d_1 (Conv2D)	(None, 34, 34, 32)	4640
batch_normalization_1 (Batch Normalization)	(None, 34, 34, 32)	128
activation_1 (Activation)	(None, 34, 34, 32)	0
conv2d_2 (Conv2D)	(None, 32, 32, 64)	18496
batch_normalization_2 (Batch Normalization)	(None, 32, 32, 64)	256
activation_2 (Activation)	(None, 32, 32, 64)	0
max_pooling2d_1 (MaxPooling2D)	(None, 16, 16, 64)	0
dropout_1 (Dropout)	(None, 16, 16, 64)	0
conv2d_3 (Conv2D)	(None, 14, 14, 32)	18464
batch_normalization_3 (Batch Normalization)	(None, 14, 14, 32)	128
activation_3 (Activation)	(None, 14, 14, 32)	0
conv2d_4 (Conv2D)	(None, 12, 12, 16)	4624
batch_normalization_4 (Batch Normalization)	(None, 12, 12, 16)	64
activation_4 (Activation)	(None, 12, 12, 16)	0

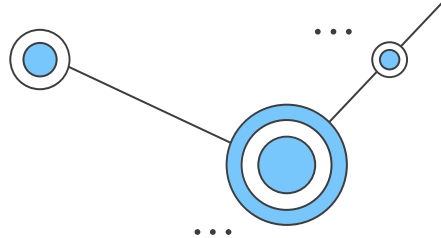
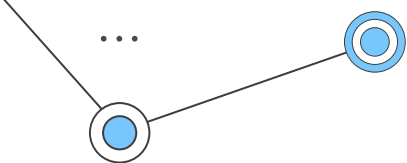
activation_4 (Activation)	(None, 12, 12, 16)	0
max_pooling2d_2 (MaxPooling2D)	(None, 6, 6, 16)	0
dropout_2 (Dropout)	(None, 6, 6, 16)	0
conv2d_5 (Conv2D)	(None, 4, 4, 8)	1160
batch_normalization_5 (Batch Normalization)	(None, 4, 4, 8)	32
activation_5 (Activation)	(None, 4, 4, 8)	0
flatten (Flatten)	(None, 128)	0
dense (Dense)	(None, 64)	8256
dropout_3 (Dropout)	(None, 64)	0
dense_1 (Dense)	(None, 1)	65
activation_6 (Activation)	(None, 1)	0

=====
Total params: 56,825
Trainable params: 56,489
Non-trainable params: 336



```
Epoch 1/20
111/111 [=====] - 122s 1s/step - loss: 0.5730 - accuracy: 0.7006 - val_loss: 0.7713 -
val_accuracy: 0.6788
Epoch 2/20
111/111 [=====] - 116s 1s/step - loss: 0.2544 - accuracy: 0.9060 - val_loss: 0.1823 -
val_accuracy: 0.9363
Epoch 3/20
111/111 [=====] - 113s 1s/step - loss: 0.1770 - accuracy: 0.9405 - val_loss: 0.1843 -
val_accuracy: 0.9316
Epoch 4/20
111/111 [=====] - 112s 1s/step - loss: 0.1632 - accuracy: 0.9460 - val_loss: 0.1520 -
val_accuracy: 0.9505
Epoch 5/20
111/111 [=====] - 112s 1s/step - loss: 0.1547 - accuracy: 0.9493 - val_loss: 0.1851 -
val_accuracy: 0.9365
Epoch 6/20
111/111 [=====] - 113s 1s/step - loss: 0.1472 - accuracy: 0.9528 - val_loss: 0.1735 -
val_accuracy: 0.9443
Epoch 7/20
111/111 [=====] - 124s 1s/step - loss: 0.1432 - accuracy: 0.9531 - val_loss: 0.1324 -
val_accuracy: 0.9563
Epoch 8/20
111/111 [=====] - 123s 1s/step - loss: 0.1389 - accuracy: 0.9542 - val_loss: 0.1329 -
val_accuracy: 0.9550
Epoch 9/20
111/111 [=====] - 113s 1s/step - loss: 0.1362 - accuracy: 0.9563 - val_loss: 0.1361 -
val_accuracy: 0.9550
Epoch 10/20
111/111 [=====] - 110s 990ms/step - loss: 0.1331 - accuracy: 0.9566 - val_loss: 0.2308 -
val_accuracy: 0.9385
Epoch 11/20
111/111 [=====] - 113s 1s/step - loss: 0.1309 - accuracy: 0.9566 - val_loss: 0.1422 -
val_accuracy: 0.9510
Epoch 12/20
111/111 [=====] - 129s 1s/step - loss: 0.1273 - accuracy: 0.9590 - val_loss: 0.1992 -
val_accuracy: 0.9452
Epoch 13/20
111/111 [=====] - 152s 1s/step - loss: 0.1261 - accuracy: 0.9587 - val_loss: 0.1266 -
val_accuracy: 0.9552
Epoch 14/20
111/111 [=====] - 147s 1s/step - loss: 0.1247 - accuracy: 0.9586 - val_loss: 0.1384 -
val_accuracy: 0.9552
Epoch 15/20
111/111 [=====] - 152s 1s/step - loss: 0.1223 - accuracy: 0.9595 - val_loss: 0.2261 -
val_accuracy: 0.9392
Epoch 16/20
111/111 [=====] - 149s 1s/step - loss: 0.1204 - accuracy: 0.9604 - val_loss: 0.1372 -
val_accuracy: 0.9559
```





```
Epoch 16/20
111/111 [=====] - 149s 1s/step - loss: 0.1204 - accuracy: 0.9604 - val_loss: 0.1372 -
val_accuracy: 0.9559
Epoch 17/20
111/111 [=====] - 150s 1s/step - loss: 0.1202 - accuracy: 0.9594 - val_loss: 0.1620 -
val_accuracy: 0.9510
Epoch 18/20
111/111 [=====] - 149s 1s/step - loss: 0.1137 - accuracy: 0.9611 - val_loss: 0.1572 -
val_accuracy: 0.9525
Epoch 19/20
111/111 [=====] - 154s 1s/step - loss: 0.1191 - accuracy: 0.9601 - val_loss: 0.1618 -
val_accuracy: 0.9459
Epoch 20/20
111/111 [=====] - 151s 1s/step - loss: 0.1151 - accuracy: 0.9609 - val_loss: 0.1333 -
val_accuracy: 0.9574
```

Warning

Figures now render in the Plots pane by default. To make them also appear inline in the Console, uncheck "Mute Inline Plotting" under the Plots pane options menu.

Infected
liver cells

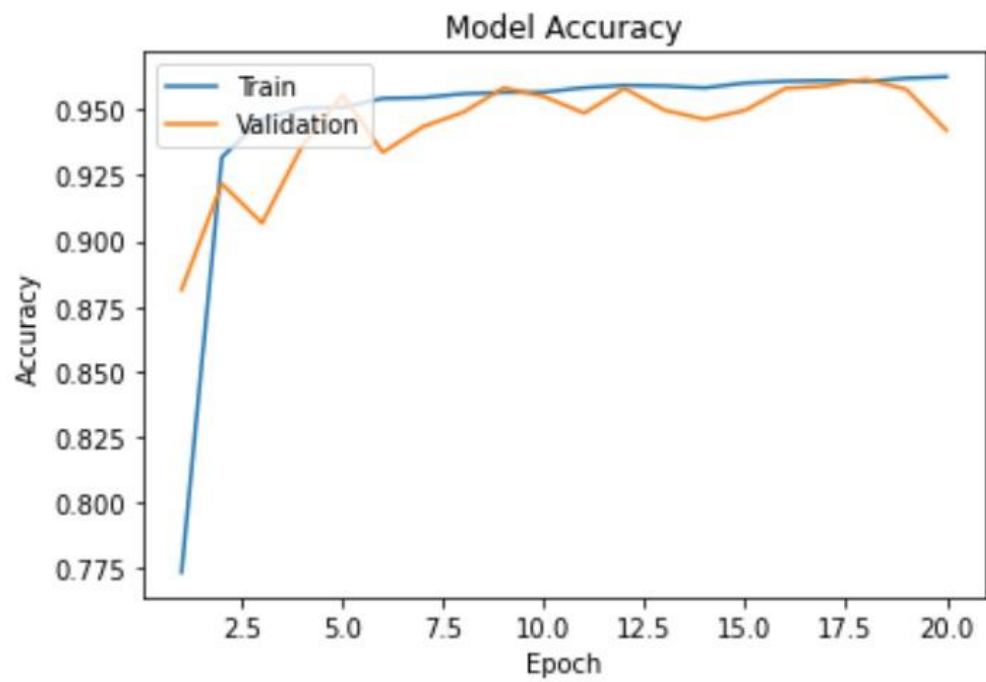
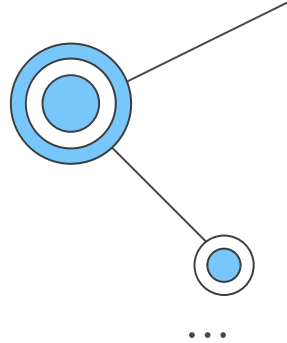
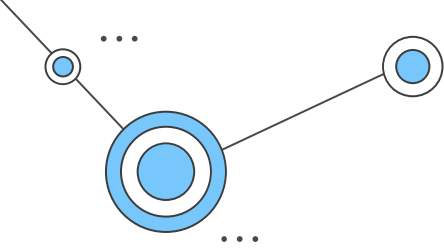
Infected
red blood cells

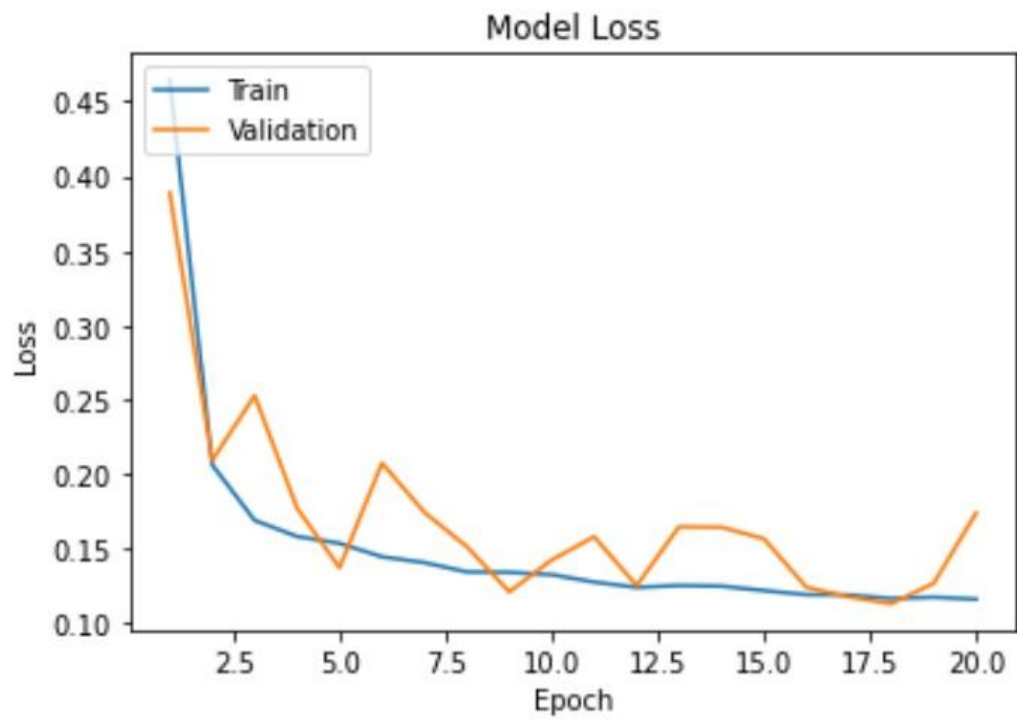
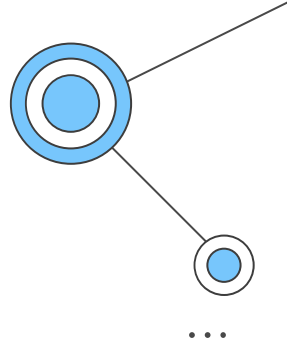
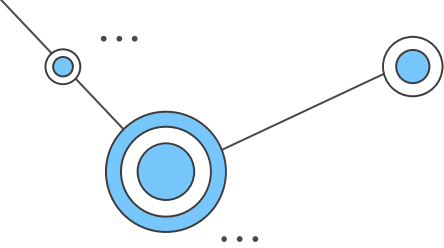
The life cycle of **MALARIA** parasite

Second infected
mosquito

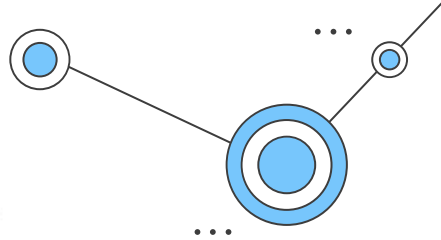
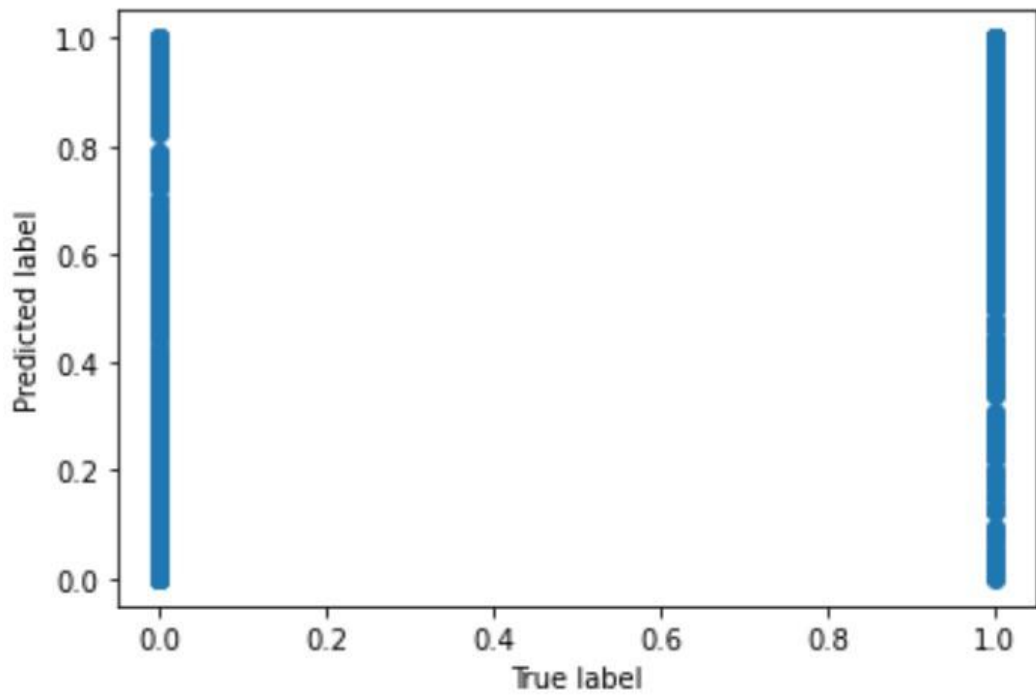
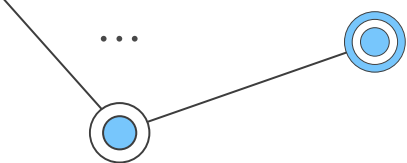
Second
infected
person

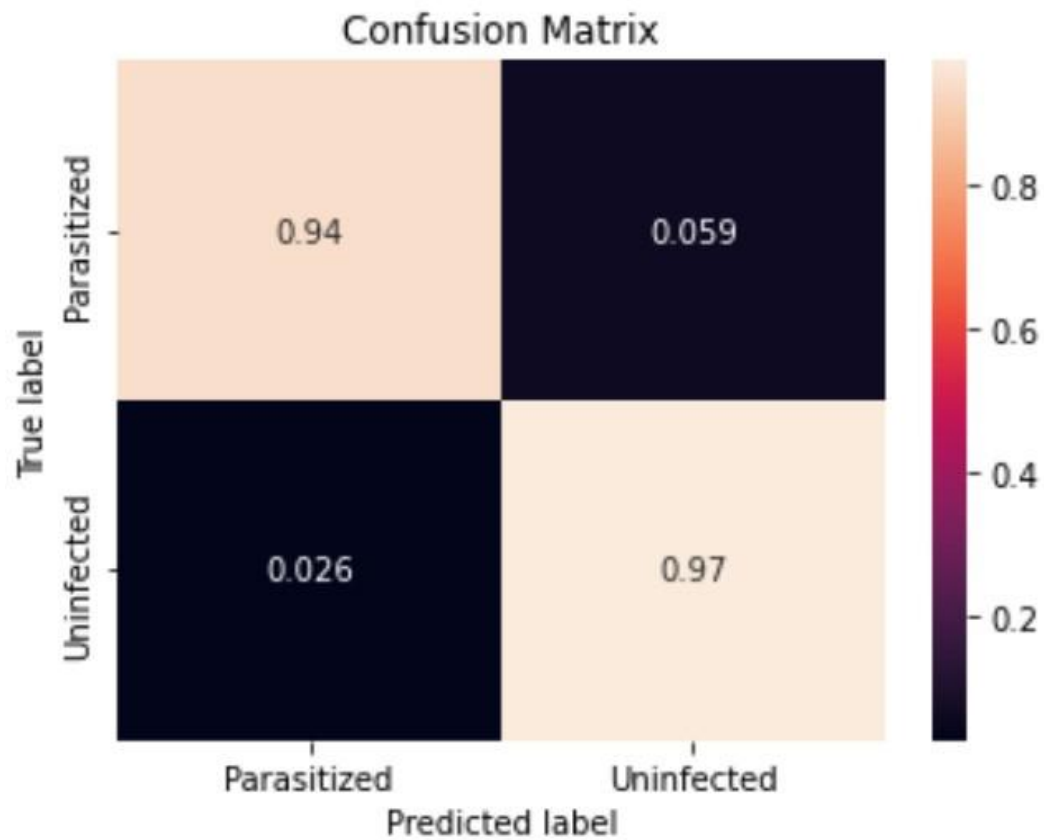
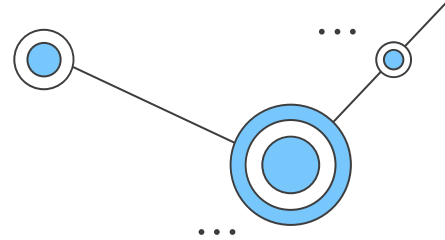
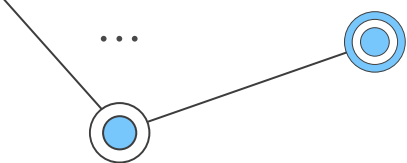
```
163 # Visualizing the Learning Curve
164
165
166 def plotLearningCurve(history,epochs):
167     epochRange = range(1,epochs+1)
168     plt.plot(epochRange,history.history['accuracy'])
169     plt.plot(epochRange,history.history['val_accuracy'])
170     plt.title('Model Accuracy')
171     plt.xlabel('Epoch')
172     plt.ylabel('Accuracy')
173     plt.legend(['Train','Validation'],loc='upper left')
174     plt.show()
175
176     plt.plot(epochRange,history.history['loss'])
177     plt.plot(epochRange,history.history['val_loss'])
178     plt.title('Model Loss')
179     plt.xlabel('Epoch')
180     plt.ylabel('Loss')
181     plt.legend(['Train','Validation'],loc='upper left')
182     plt.show()
183
184 plotLearningCurve(history,20)
185
186
187 # Confusion Matrix
188
```






```
186 # Confusion Matrix
187
188
189
190 all_labels = []
191 all_y_preds = []
192
193 for images, labels in val_ds:
194     print('images.shape:', images.shape)
195     print('labels.shape:', labels.shape)
196
197     y_test_pred = model.predict(images)
198     all_labels.extend(labels)
199     all_y_preds.extend(y_test_pred)
200
201 plt.scatter(all_labels, all_y_preds)
202 plt.xlabel('True Label')
203 plt.ylabel('Predicted Label')
204 plt.show()
205
206
207 y_test_pred = model.predict(val_ds)
208 y_test = np.concatenate([y for x, y in val_ds], axis=0)
209
210 y_test_pred_class = np.round(all_y_preds).reshape(-1,1)
211 y_test_class      = np.round(all_labels).reshape(-1,1)
212
213
214
215 # Creates a confusion matrix
216 cm = confusion_matrix(y_test_class, y_test_pred_class, normalize='true')
217
218 # Transform to df for easier plotting
219 cm_df = pd.DataFrame(cm,
220                      index = ['Parasitized', 'Uninfected'],
221                      columns = ['Parasitized', 'Uninfected'])
222
223 plt.figure(figsize=(5.5,4))
224 sns.heatmap(cm_df, annot=True)
225 plt.title('Confusion Matrix')
226 plt.ylabel('True Label')
227 plt.xlabel('Predicted Label')
228 plt.show()
229
230
```







```
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 1s 50ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 43ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 45ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 43ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 45ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 43ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 44ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 44ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 44ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 46ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 44ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 46ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 43ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 42ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 47ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
```

```
labels.shape: (200,)
7/7 [=====] - 0s 47ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 48ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 50ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 46ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 48ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 44ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 48ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 54ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 50ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 48ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 47ms/step
images.shape: (200, 75, 75, 3)
labels.shape: (200,)
7/7 [=====] - 0s 46ms/step
images.shape: (111, 75, 75, 3)
labels.shape: (111,)
4/4 [=====] - 0s 49ms/step
28/28 [=====] - 11s 364ms/step
```

In [16]:

Interpretation:

- Aus der Confusion Matrix, kann man erkennen wo TP, FP, TN, FN liegen:

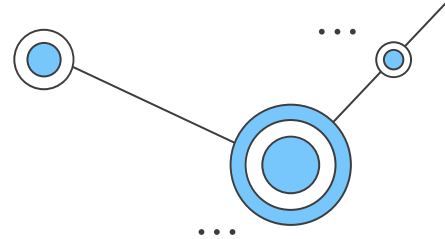
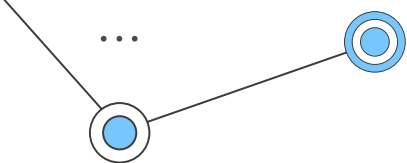
TP	FN
FP	TN

- Die Precision kann ausgerechnet werden: $\text{Precision} = \text{TP} / (\text{TP} + \text{FP})$
 $\Rightarrow 0.94 / (0.94 + 0.026) = 0.9731$
Entspricht ein Precision von 97.3%
- Der Recall kann auch ausgerechnet werden: $\text{Recall} = \text{TP} / (\text{TP} + \text{FN})$
 $\Rightarrow 0.94 / (0.94 + 0.059) = 0.9409$
Entspricht ein Recall von 94.1%

Fazit

- Ziel der vorliegenden Projekt war Malaria infizierte Zellen von nicht infizierte
- Zellen zu unterscheiden/erkennen (Binary Image Classification of Malaria cells).
- Die Ergebnisse des Projekts zeigen dass CNN (Convolutional Neural Networks), ein sehr guter Modell ist für die Klassifikation von Cell Images (Mikroskopische Bilder).
- Die Convolutional und Pooling Layers helfen bei der Data Preprocessing.
- Die Ergebnisse der Binary Image Classification: accuracy von 0.9609, val_loss: 0.1333, val_accuracy : 0.9574
- Entspricht ein Prozentsatz von 95.7%.
- Ein Vergleich mit anderen Modellen ist mit der erzielten 95.7%, nicht nötig.
- Außerdem habe ich leider ein Mangel an Hardware Memory Leistung gehabt.
- Versuche mit weiteren Modellen hätte größte Einfluss auf das Zeitmanagement gehabt.





Vielen Dank für
Ihre Aufmerksamkeit



Quelle Datensatz:
<https://www.kaggle.com/>