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SMAIL COMMUNITY PAGE

The *Tara* Pacific expedition—A pan-ecosystemic approach of the "-omics" complexity of coral reef holobionts across the Pacific Ocean

authors

Serge Planes 1,2‡*, Denis Allemand³‡, Sylvain Agostini⁴, Bernard Banaigs¹, Emilie Boissin 1, Emmanuel Boss 5, Guillaume Bourdin⁵,6, Chris Bowler 2,7, Eric Douville 8, J. Michel Flores 9, Didier Forcioli¹0, Paola Furla 10, Pierre E. Galand 2,1¹, Jean-François Ghiglione²¹², Eric Gilson¹0, Fabien Lombard6, ©lémentine Moulin¹³, Stephane Pesant 14,1⁵, Julie Poulain¹⁶, Stéphanie Reynaud³, Sarah Romac²,¹७, Matthew B. Sullivan 18, Shinichi Sunagawa 19, Olivier P. Thomas Romain Troublé²,¹³, Colomban de Vargas²,¹¹७, Rebecca Vega Thurber 1², Christian R. Voolstra 2², Patrick Wincker²,¹⁶, Didier Zoccola 3, the Tara Pacific Consortium¹¹

institutions and Publishers

1 Laboratoire d'Excellence "CORAIL," PSL Research University: EPHE-UPVD-CNRS, USR 3278 CRIOBE, Université de Perpignan, Perpignan Cedex, France, 2 Research Federation for the study of Global Ocean Systems Ecology and Evolution, FR2022/Tara Oceans-GOSEE, Paris, France, 3 Centre Scientifique de Monaco, Monte Carlo, Principality of Monaco, 4 Shimoda Marine Research Center, Shimoda, Japan, 5 School of Marine Sciences, University of Maine, Orono, Maine, United States of America, 6 Sorbonne Université, Institut de la Mer de Villefranche sur mer, Laboratoire d'Océanographie de Villefranche, Villefranche-sur-Mer, France, 7 Institut de Biologie de l'Ecole Normale Supérieure (IBENS), Ecole normale supérieure, CNRS, INSERM, Université PSL, Paris, France, 8 Laboratoire des Sciences du Climat et de L'Environnement, LSCE/IPSL, CEA-CNRS-UVSQ, Université Paris-Saclay, Gif-sur-Yvette, France, 9 Weizmann Institute of Science, Dept. Earth and Planetary Science, Rehovot, Israel, 10 Université Côte d'Azur-CNRS-INSERM, IRCAN, Medical School, Nice, France and Department of Medical Genetics, CHU of Nice, Nice, France, 11 Sorbonne Université, CNRS, Laboratoire d'Ecogéochimie des Environnements Benthiques (LECOB), Observatoire Océanologique de Banyuls, Banyuls sur mer, France, 12 Sorbonne Université Laboratoire d'Océanographie Microbienne LOMIC, UMR 7621, Observatoire Océanologique de Banyuls, Banyuls sur mer, France, 13 La Fondation Tara Expéditions, "Base Tara" 11, Paris, France, 14 PANGEA, Data Publisher for Earth and Environment Science, Bremen, Germany, 15 MARUM—Center for Marine Environmental Sciences, Universität Bremen, Bremen, Germany, 16 Génomique Métabolique, Genoscope, Institut François Jacob, CEA, CNRS, Université Evry, Université Paris-Saclay, Evry, France, 17 Sorbonne Université, CNRS, Station Biologique de Roscoff, AD2M, UMR 7144, ECOMAP, Roscoff, France, 18 Departments of Microbiology and Civil, Environmental and Geodetic Engineering, The Ohio State University, Columbus, Ohio, United States of America, 19 Department of Biology and Swiss Institute of Bioinformatics, ETH Zürich, Zürich, Switzerland, 20 Marine Biodiscovery Laboratory, School of Chemistry and Ryan Institute, National University of Ireland, Galway (NUI Galway), Galway, Ireland, 21 Department of Microbiology, Oregon State University, Corvallis, Oregon, United States of America, 22 Department of Biology, University of Konstanz, Konstanz, Germany

institutions

These authors share first authorship on this work.

Membership of the Tara Pacific Consortium is listed in the Acknowledgments.

* planes@univ-perp.fr

Abstract

Coral reefs are the most diverse habitats in the marine realm. Their productivity, structural complexity, and biodiversity critically depend on ecosystem services provided by corals that are threatened because of climate change effects—in particular, ocean warming and acidification. The coral holobiont is composed of the coral animal host, endosymbiotic dinoflagellates,



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footnote Abbreviations: CBD, Convention on Biological Diversity; FAIR, findable, accessible, interoperable, reusable; GLODAP, Global Ocean Data Analysis Project; MODIS, Moderate Resolution Imaging Spectroradiometer; MS, mass spectrometry; NASA, National Aeronautics and Space Administration; NMR, nuclear magnetic resonance; OTU, operational taxonomic unit; SST, sea surface temperature.

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associated viruses, bacteria, and other microeukaryotes. In particular, the mandatory photosymbiosis with microalgae of the family Symbiodiniaceae and its consequences on the evolution, physiology, and stress resilience of the coral holobiont have yet to be fully elucidated. The functioning of the holobiont as a whole is largely unknown, although bacteria and viruses are presumed to play roles in metabolic interactions, immunity, and stress tolerance. In the context of climate change and anthropogenic threats on coral reef ecosystems, the Tara Pacific project aims to provide a baseline of the "-omics" complexity of the coral holobiont and its ecosystem across the Pacific Ocean and for various oceanographically distinct defined areas. Inspired by the previous Tara Oceans expeditions, the Tara Pacific expedition (2016-2018) has applied a pan-ecosystemic approach on coral reefs throughout the Pacific Ocean, drawing an east-west transect from Panama to Papua New Guinea and a south-north transect from Australia to Japan, sampling corals throughout 32 island systems with local replicates. Tara Pacific has developed and applied state-of-the-art technologies in very-highthroughput genetic sequencing and molecular analysis to reveal the entire microbial and chemical diversity as well as functional traits associated with coral holobionts, together with various measures on environmental forcing. This ambitious project aims at revealing a massive amount of novel biodiversity, shedding light on the complex links between genomes, transcriptomes, metabolomes, organisms, and ecosystem functions in coral reefs and providing a reference of the biological state of modern coral reefs in the Anthropocene.

heading Introduction

The 20th century has seen the earth enter into the now widely called Anthropocene [1]. Anthropogenically induced changes are happening on both global and local scales and are altering the physiology of organisms and ecosystems by modifying the entire earth's physical, chemical, and biological processes [2]. Among marine ecosystems, coral reefs have the unfortunate privilege of being highly sensitive to these environmental modifications [3]. In particular, the thermally mediated process of coral bleaching—i.e., the loss of the obligate photosynthetic microalgal endosymbionts—is increasingly decimating corals. Projections estimate that approximately 25% of reefs have already been lost, and up to 99% will be threatened and dramatically transformed by 2050 [4]. Even though they cover only approximately 0.2% of the ocean's surface [5], coral reefs harbor approximately 30% of ocean biodiversity [6], providing ecological services (fisheries, tourism, coastal protection) to nearly 1 billion people [7], and are estimated to be worth USD 30 billion per year [8].

The late 20th century has also seen our world entering into the "-omics" revolution sparked by the development of high-throughput analyses of DNA, RNA, proteins, and metabolites. This revolution has changed our approach to investigate organisms, and it is about to change the descriptions of ecosystems as genes-to-ecosystem modeling improves [9]. It will advance our capabilities to investigate the biodiversity and functioning of our oceans in a holistic way. Holistic approaches, interrogating components across the various levels of organization of an ecosystem, appear accessible today, as exemplified in the *Tara* Oceans project [10], which continues to decipher biodiversity and structural networks across all plankton organisms in the ocean [11].

Tara Pacific is a unique scientific expedition inspired by earlier maritime explorations that uncovered the unchartered territories of marine biodiversity. Coral reef research



started with the contribution of Charles Darwin during "the voyage of the *Beagle*" (1831–1836), when Darwin explored many reefs in the Indian and Pacific Oceans and established his biophysical theory of the formation of coral reefs and atolls [12]. He also highlighted the paradoxical high diversity of reef organisms living in a nutrient-poor system. Later, the Great Barrier Reef Expedition (1928–1929) led by Charles Yonge was a landmark in coral reef research, as it established the scientific basis of coral physiology (nutrition, symbiosis, growth, etc.). The "polypifers," later called coral polyps, were then demonstrated to host a large population of organisms (protists, viruses, bacteria, archaea, and all sorts of unicellular and multicellular eukaryotes), which are intimately involved in the physiology of the coral animal host in a complex and still mysterious system named the holobiont [13].

The *Tara* Pacific expedition has built on these early tracks, applying the most recent technologies to map the "-omics" complexity of the coral holobiont within its ecosystem and across the Pacific Ocean. Through the exploration of marine biodiversity at scales spanning from organisms to genes to biomolecules, *Tara* Pacific is undertaking the first pan-ecosystemic study of coral reef diversity across an entire ocean basin (Fig 1). Given that a holistic approach integrating all components of the reef biota is unrealistic, the *Tara* Pacific expedition focuses its approach on widely distributed coral and fish holobionts and their contextual biological (plankton) and physicochemical environment, including modifications in the context of global changes (Fig 2).

The geographic and sequencing extent of *Tara* Pacific will bring an exceptionally comprehensive description of coral holobionts, together with the surrounding fish and plankton biota. Such unprecedented sequencing coverage and depth will place coral reefs among the first ecosystems with a comprehensive description of their hologenomic diversity (i.e., the totality of genomes making up the coral holobiont) across their natural environments. We will go beyond the mere description of species composition to investigate in detail the gene content and gene expression of these communities, as well as the interconnectedness with their hosts and habitats. Broad emerging patterns will then be accessible through large-scale data mining and network analyses.

heading

The coral holobiont

The coral host

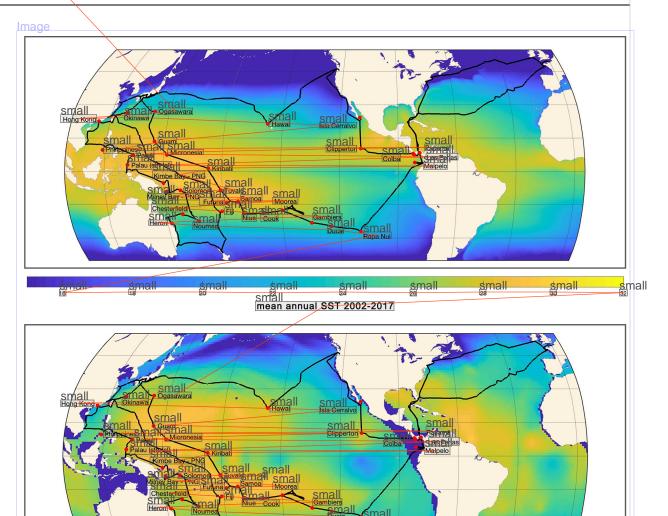
Scleractinian corals evolved over 450 million years ago [14] and belong to the phylum Cnidaria, animals located close to the root of the metazoans. Because of their place in the early Eumetazoa and as a sister group of bilaterians, the cnidarians allow interesting evolutionary analyses regarding the origins of animal complexity and the coevolution of early animals with microbes. Shinzato and colleagues [15] were the first to show that the coral genome is as complex as the one of vertebrates and that it has retained many ancestral genes lost in other lineages. Subsequent studies confirmed the deep genomic differences between coral taxa [16]. Moreover, the exceptional longevity of corals, reaching several hundred to thousands of years, makes them attractive emerging models for aging studies.

Headline

The coral endosymbiont microalgae

Reef corals host symbiotic unicellular algae of the family Symbiodiniaceae inside their endodermal cells [17], raising intriguing questions on animal–plant interactions, photosymbiosis, coevolution, and speciation [18]. The divergence and genetic variation within and between populations of coral species is also investigated, allowing us to firmly ground the exact taxonomic status of each coral host [19].





imageDescription
Fig 1. Map showing the route of the *Tara* Pacific expedition and the sampling sites (red spots) throughout the Pacific Ocean (July 2016 to October 2018) as well as the mean annual SST (top) and pH (bottom). Global annual SSTs were extracted from the MODIS-Aqua satellite and correspond to global mapped climatologies of the period from 2002 to 2018 (NASA Goddard Space Flight Center), whereas pH values originate from the GLODAPv2 database [59, 60], with mean data displayed corresponding to the 0-30 m depths between 2000 and 2013. GLODAP, Global Ocean Data Analysis Project; MODIS, Moderate Resolution Imaging Spectroradiometer; NASA, National Aeronautics and Space Administration; PNG, Papua New Guinea; SST, sea surface temperature.

small (2000-2013)

https://doi.org/10.1371/journal.pbio.3000483.g001

heading

The coral microbiome

In addition to the Symbiodiniaceae, corals host diverse groups of microorganisms, bacteria, archaea, fungi, and protists, together forming the so-called coral holobiont [13]. The importance of the microbiome in the normal steady-state functioning of corals or during environmental stress is not well understood, but recent research suggests that microbiome structure aligns with and contributes to stress resilience [20, 21, 22] (in particular, bleaching [23, 24]), as seen in humans [25]. Recent work provides evidence that corals have coevolved with certain groups of microbes [26–28], but the exact nature of these host–microbe symbioses and their involvement in coral resilience remains unclear. Therefore, Tara Pacific will draw, using



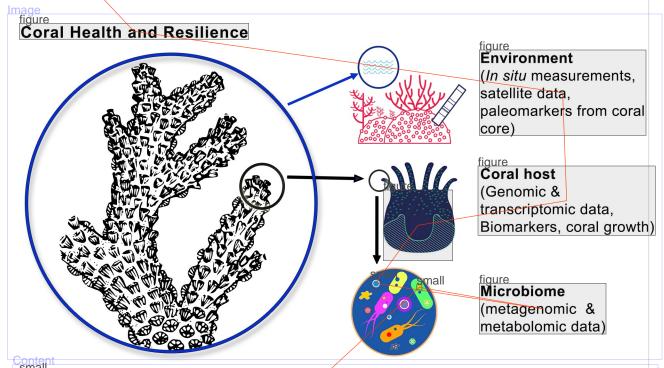


Fig 2. Schematic representation of the frame and major goals of *Tara* Pacific, which is investigating jointly the coral, microbiome, and environment.

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marker gene and metagenomic/metatranscriptomic approaches, a nearly exhaustive census of the microbial diversity associated with targeted coral taxa, allowing the understanding of the major drivers of the microbiome diversity and its role in coral health, resilience, and evolution.

heading The coral virome

Viruses infect all known cellular organisms and are the most abundant biological entities on our planet [29]. Yet even the basic diversity of viruses associated with marine species and habitats is shockingly unknown. This is particularly true in tropical reef systems, where research on viral abundance, diversity, and dynamics is still in its infancy (for review, see [30]). *Tara* Pacific is using metagenomics and microscopy to interrogate the diversity of viruses across Pacific waters and within the foundation species of coral reefs.

heading

Research objectives of Tara Pacific

Our goal is to unveil the entire organismal diversity of eukaryotes, prokaryotes, and viruses associated with the targeted coral and fish holobionts and their surrounding waters and to assess variation across the explored ecological and geographical gradients. The main objectives of *Tara* Pacific are as follows:

Draw a near-exhaustive census of the biodiversity composing and surrounding coral holobionts. This includes the coral hosts, their endosymbiotic microalgae (zooxanthellae of the family Symbiodiniaceae), and the associated microeukaryotes, bacteria, archaea, and viruses found either as endo- or exosymbionts, or in the plankton in surrounding water, using a metabarcoding approach.



- Determine the spatial patterns of the holobionts and the environmental diversity within
 coral reefs throughout the Pacific Ocean and derive large-scale biogeographic patterns to be
 compared with macrofauna patterns of diversity (i.e., the Pacific biodiversity gradient).
- Compare metatranscriptomes and metagenomes of coral reefs throughout the Pacific Ocean
 and elucidate the contribution of microbial diversity to local versus basin-scale adaptation in
 response to climate change. Importantly, the
 entire plankton community surrounding the
 corals and the islands has been sampled and integrated with physicochemical contextual
 data, providing critical context to advance our
 understanding of the environmental contribution to coral reef resilience.
- Investigate divergence and standing genetic variation within and between populations of the targeted coral species and their Symbiodiniac eae symbionts across the Pacific Ocean basin via marker gene barcoding as well as genome and transcriptome sequencing (population genomics using SNPs). Extensive metadata resources collected during *Tara* Pacific will allow the coral holobiont composition to be modeled, taking environmental and coral trait data into account (e.g., temperature, pH, bleaching, symbiont acquisition, reproduction cycle, etc.) to identify drivers of adaptation/selection or acclimatization, providing insights into coral stress resilience.
- Investigate the health status of corals by measuring growth parameters of the recent coral skeleton and stress biomarkers (antioxidant capacity, apoptosis, stress response pathways, transcriptomic signatures) as well as by determining the telomere status of each sample (telomeric DNA length and damage as potential proxies for stress). These data will be studied in relation to holobiont biodiversity, transcriptomic and metabolomics data, modern and/or historical environmental parameters, and stresses determined from a geochemical analysis of the coral skeleton.
- Generate a holistic coral metabolome for coral species as a foundation to the identification
 of the links between holobiont metabolism and prevailing environmental conditions.
- Identify how the environment (physical, chemical, biogeochemical, and biological) is influencing coral holobiont diversity, physiology, and evolution using an extensive compilation of environmental conditions.

-leadline

heading

A unique, pan-ecosystemic sampling strategy

The Pacific Ocean covers approximately one-third of the earth's surface, with nearly 25,000 islands, most of which harbor coral reefs. *Tara* Pacific equipped the schooner *Tara* to explore 32 islands across the entire Pacific over a period of 2.5 years (Fig 1). The route of the *Tara* schooner was chosen to maximize the number of visited remote islands and atolls and to perform the widest possible comparative survey, from the equator to the temperate and more acidic regions, encompassing most of the environmental range where scleractinian coral species can live. It also covers a biodiversity gradient, from the low diversity present in the eastern Pacific reefs to the highly diverse western Pacific "warm pool" [14]. This sampling strategy also encompasses a wide variety of environments, from high temperature/low seasonality to low temperature/high seasonality, and also a full range of trophic status and physicochemical environmental parameters that affect coral reef ecosystems (sea surface temperature [SST], pH, nutrients, lights, pollutants, etc.).

Across its entire route, the *Tara* Pacific expedition targeted two species of scleractinian corals (Cnidaria, Anthozoa: *Pocillopora meandrina* and *Porites lobata*), one species of hydrocoral



(Cnidaria, Hydrozoa: *Millepora platyphylla*), and two species of reef fish (*Acanthurus triostegus* and *Zanclus cornutus*) (Fig 3). The chosen taxa are among the few species that occur across most of the Pacific Ocean and are usually abundant on reefs. Around each of the 32 islands, three sites were sampled, collecting 10 colonies of each coral species and 5–10 individuals of each fish species, as well as water samples (see below) (Fig 3). Coral fragments and fish samples were preserved immediately on board, using specific buffers (e.g., DNA/RNA shield for genomics, glutaraldehyde for microscopy, etc.), and or flash frozen for subsequent laboratory analyses (e.g., metabolomics, biomarkers, etc.).

In addition, seawater biogeochemistry and plankton microbiomes (viruses to zooplankton) were assessed in oceanic waters upstream and downstream of each island, as well as from water above each site (i.e., "surface water") and surrounding two colonies of sampled *P. meandrina* corals at each site (i.e., "coral-surrounding water"). Underwater propeller-driven plankton nets (for surface water) and water pumping systems (for coral-surrounding water) were developed to collect plankton on reef sites, whereas the preparation of plankton samples into different size classes (<0.2, 0.2–3, 3–20, 20–2000 µm) and their preservation followed the *Tara* Oceans protocols [31] in order to maximize comparability and integration of data. Samples will be analyzed using high-throughput imaging and genomic techniques.

Finally, key physical, chemical, and biological parameters were measured continuously from surface water and from aerosols using specific instruments installed on board (e.g., thermosalinograph, mass- and spectrophotometers), as well as samples for imaging and genetic analysis of aerosol particles. Environmental context will also be obtained from satellite images and operational oceanographic products from the European Copernicus Marine Service and Mercator Ocean.

Headline heading

Tara Pacific samples and data resources for the immediate and long-term future

Tara Pacific endorses the findable, accessible, interoperable, reusable (FAIR) principles for scientific data management [32] as well as ethical and responsible use of data. With a few exceptions, all physical samples of coral, plankton, fish, and sediments were preserved on board Tara and sent back to partner laboratories for subsequent analyses. Controlled vocabularies describing sampling devices and sample preparation protocols were used throughout the expedition to capture provenance metadata on customized log sheets, and samples were assigned unique identifiers to facilitate their traceability. Legal documents regulating the collection, export, and import of samples, as well as links to the Convention on Biological Diversity's (CBD) access and benefit-sharing clearing house (https://absch.cbd.int/) are in the process of being provided for each sample as a first step toward ethical and responsible use of the *Tara* Pacific data. The detailed registry of all samples, including their provenance and environmental and legal context, is curated manually using simple semantics that enable machine- and human-readable data discovery services. Sequencing data will be deposited at the free, openaccess European Nucleotides Archive (https://www.ebi.ac.uk/ena); environmental data are deposited at the free, open-access PANGAEA database (https://www.pangaea.de/); and both archives will be interlinked via the sample registry available online at BioSamples (https:// www.ebi.ac.uk/biosamples/). Metabolomic data (mass spectrometry [MS] and nuclear magnetic resonance [NMR]) and their annotations will also be accessible through the Metabolights portal (https://www.ebi.ac.uk/metabolights/). The submission of data in a relational, openaccess, updated, and cured database is critical and necessary for our aim to establish a reference of the biological state of coral reefs in the Anthropocene for the broader research community.



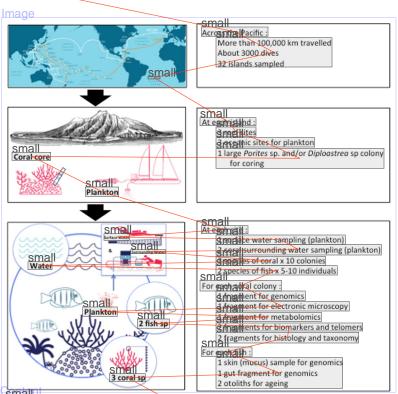


Fig 3. Schematic representation of the sampling design allowing the comparison of various components of coral reef ecosystems. Using the developed protocol, we collected 2,500 oceanic samples (32 islands × 3 oceanic sites × 5 size fractions × 2–5 protocols), 7,500 coral-surrounding and surface water samples (32 islands × 3 coral sites × 3 environments × 5 size fractions × 2–5 protocols), 40 coral core samples (32 islands × 1 *Porites* sp. and/or *Diploastrea sp* × 1–2 core samples), 20,160 coral fragments (32 islands × 3 coral sites × 3 species of corals × 10 colonies × 7 protocols), and 9,600 fish tissue samples (32 islands × 3 coral sites × 2 species of fish × 5–10 individuals × 5 protocols). Taken together, the dataset comprises a total of approximately 40,000 samples. *Bottom figure copyright to Agence DATCHA/taraexpeditions.org*.

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heading

Metagenomics and metatranscriptomics to explore the diversity and physiology of coral reef holobionts

Building upon expertise gained in the previous *Tara* Oceans project [33–36], we are applying Illumina HiSeq technology to sequence a series of barcodes designed recently to explore the diversity of bacteria and archaea (V4–V5 region of the nuclear 16S rRNA gene [37]), eukaryotes (V9 region of nuclear 18S rRNA gene [33]), Symbiodiniaceae (ITS2 region of the nuclear ribosomal DNA [38-40]), and metazoan species (mitochondrial COX1 gene [41]), as well as metagenome assembly-defined viral populations [42, 43]. Barcode-specific PCRs are performed on DNA from coral tissues, coral-surrounding water, fish tissues, and surface water above the reef in order to assess interkingdom diversity associated with the holobiont at different degrees of proximity from the coral animal host, from endosymbiotic to drifting in coralsurrounding and surface waters. Biogeographic gradients will also be investigated between the studied island systems. The sequence reads are assembled into operational taxonomic units (OTUs) for bacteria, archaea, and eukaryotes and into ITS2-type profiles (using the SymPortal framework at symportal.org [40]) for Symbiodiniaceae and taxonomically annotated by comparison to reference databases. Classical numerical ecology methods are applied to employ OTU richness and abundance data to (1) assess and compare the total diversity of bacteria, archaea, and eukaryotes associated with various compartments of the different coral



holobionts; (2) compare biodiversity patterns across coral colonies, hosts, reefs, islands (geographic and Lagrangian oceanic circulation distances), and all contextual environmental parameters measured; (3) integrate the *Tara* Pacific data into the *Tara* Oceans global plankton dataset. In addition, co-occurrence graph techniques inspired from systems biology [44, 9] are used to reconstruct OTU interaction (sub)networks, and we will aim to disentangle the intrinsic (symbiotic auto-organization) from the extrinsic (environmental, historical contingencies) forces on holobiont biodiversity. Overall, this study is the first attempt to characterize coral reef and holobiont diversity across comprehensive taxonomic, ecological, and geographical scales over an entire oceanic basin. Comparison with *Tara* Oceans plankton data will also reveal which part of the coral holobiont biodiversity is found in the open ocean and thus bring fundamental information about reef connectivity, as recently shown for Symbiodiniaceae, which are found all over the plankton [34].

The sequencing of the holotranscriptomes of coral holobionts sampled along the transect will provide important genetic information to uncover ecological and evolutionary questions at both intraspecific and community levels. Two protocols are successively applied on total RNA from three specimens per species and per site. In a first step, a polyA+ enrichment protocol is applied to provide eukaryotic mRNA in order to study the coral and Symbiodiniaceae dual transcriptome, and in a second step, rRNA is removed from the remaining polyA – fraction to sequence microbial mRNA. We are applying different metatranscriptomics protocols to capture the gene expression of the Cnidaria, Symbiodiniaceae, bacteria and archaea, other microbes, and even viruses from the same initial tissue samples [45, 46]. In addition, we are using similar analysis procedures after filtration of coral-surrounding water samples. This may potentially reveal how microbes and viruses interact with the coral host and how the coral host adapts or acclimatizes to their presence. On top of providing deep insight on gene expression networks at the holobiont level, mRNA samples will also be used to generate transcriptomewide SNPs for the coral host, allowing in-depth characterization of the patterns of diversity, connectivity, demographic history, and local adaptation of sampled coral species. Hence, the resulting patterns will provide a solid foundation on which to base the study of partner relationships within the coral holobiont, and possibly even allow for the identification of the selective forces acting on key genes for these symbiotic associations, plus an account on standing genetic diversity within and between reefs to highlight regions of high genetic endemicity and diversity important for conservation efforts.

A metabolomic approach

We will also assess the metabolome (i.e., the metabolic diversity) of the three targeted coral species (two stony and one fire coral, see above). Small molecules (i.e., specialized metabolites) are the end products of unique metabolic pathways. Most of these presumably possess an ecological role, and as such, they represent key phenotypic traits for the specimens. The broad geographical cover of the collected coral species will enable us to provide some insight into the relative contribution of environmental factors and the genetic information expressed in the metabolome by comparison with genomic data. Recent advances in analytical techniques and bioinformatics led to the development of global metabolomics approaches capable of providing an overview of the thousands of metabolites present in a minute amount of sample [47, 48]. A nontargeted metabolomics approach is applied to the three species of corals collected across the Pacific Ocean using both MS and NMR. The subsequent identification of the chemomarkers unveiled by these comparative approaches are performed using not only databases of experimental spectra but also comparison with in silico databases of metabolites. The results will finally contribute to the assessment of the reef health status when combined with other "-omics" data.



heading

Identifying the interaction among environmental stressors, the holobiont, and coral resilience

A key question to be addressed by Tara Pacific is how reef corals change geographically and how corals locally adapt or acclimate to environmental changes and increasing stress. This question has been rarely studied at the scale of a complete ocean and across multiple populations under consideration of phenotypic traits [49]. The physiological status of corals will be assessed by measuring the parameters of coral growth such as density, linear extension, and calcification rates of P. lobata (with added Diploastrea heliopora in few places) colonies by analyzing coral cores recovering the last 50–150 years by sclerochronology. The annual density bands and growth parameters of the massive coral sampled along the transect will be determined using X-ray radiography and 3D-computed tomography of growth rings on sliced coral cores [50-52], allowing the calculation of recent reef growth over the last decades and century. In parallel, ambient sea temperature and pH changes over the same period of time will be documented by analyzing, at an annual or monthly timescale, the most advanced geochemical tracers along the cores, such as boron isotopes or trace elements, trapped along the cores during the coral life [53–56]. Recent stress events will be identified by a series of conventional markers of physiological stress or damage (see above) [57]. In addition, telomere length measurements of coral and microalgal symbiont genomes will be conducted. Notably, although measurements of telomere length as a method of assessing stress accumulation is currently being employed in numerous human cohort studies, the importance of telomere length for stress resilience and biodiversity in ecological studies is in its infancy and remains to be determined [58]. Thus, it will be highly informative to compare telomere length variations with coral growth measurements assessed by sclerochronology.

More broadly, the integration of environmental and physicochemical with biological data will allow us to pinpoint the specific adaptations that enable corals to live and survive across environmental gradients. Comparison of current to historical data throughout the Pacific will then show the biological cost of this adaptation and the associated time span. This in turn will make for a much better understanding of the capacity of coral holobionts to adapt to adverse environmental conditions and the required time scales.

In summary, *Tara* Pacific is poised to build the most comprehensive morphomolecular inventory of the phenotypic and genotypic biodiversity of coral reef ecosystems, including a wide spectrum of life from viruses to bacteria and from unicellular eukaryotes to metazoans and covering nested spatial scales from coral holobiont colonies, their surrounding water (i.e., coral-surrounding water), reef surface water, and neighboring oceanic surface waters (upstream and downstream of the sampled islands). The project focuses on selected key species that are ubiquitous in coral reef ecosystems throughout the Pacific and can be traced from an environmental, physicochemical, biological perspective down to the gene. This ambitious project has the potential to reveal substantial uncharted biodiversity; to shed light on the complex links between genomes, transcriptomes, metabolomes, organisms, and ecosystem functions in coral reef systems; and to provide a basis for the biodiversity and biological state of modern coral reefs for the research community at large. In addition, this project will contribute significantly to other fields of research such as stress and aging biology. We expect that the unprecedented scale of this project will help to decipher the complex interactions that together ensure a healthy state of coral reefs, a quest for which advances are urgently needed.

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institutions

Department of Earth Sciences, University of Southern California, Los Angeles, California, United States of America; (30) Marine Chemistry and Geochemistry Department, Woods Hole Oceanographic Institution; (31) Institut Pierre Simon Laplace, CNRS/IRD/MNHN (LOCEAN)Sorbonne-Université Paris Cedex 05, France; (32) Laboratoire Evolution, Génomes, Comportement et Ecologie, CNRS/Université Paris-Saclay, Avenue de la Terrasse, Gif sur Yvette, France. S. Planes and D. Allemand are joint scientific directors of *Tara* Pacific.

references

References

- 1. Crutzen PJ, Stoermer EF (2000) The "Anthropocene". Global Change 41: 17–18.
- Steffen W, Grinevald J, Crutzen P, McNeill J (2011) The Anthropocene: conceptual and historical perspectives. Phil Trans R Soc A 369: 842–867. https://doi.org/10.1098/rsta.2010.0327 PMID: 21282150
- Gattuso JP, Magnan A, Billé R, Cheung WWL, Howes EL, Joos F, et al. (2015) Contrasting futures for ocean and society from different anthropogenic CO₂ emissions scenarios. Science 349: aac4722. https://doi.org/10.1126/science.aac4722 PMID: 26138982
- 4. IPCC (2018) Global Warming of 1.5°C: An IPCC Special Report on the impacts of global warming of 1.5°C above pre-industrial levels and related global greenhouse gas emission pathways, in the context of strengthening the global response to the threat of climate change, sustainable development, and efforts to eradicate poverty. Masson-Delmotte V, Pirani A, Moufouma-Okia W, Péan C, Pidcock R, Connors S, Matthews JBR, Chen Y, Zhou X, Gomis MI, Lonnoy E, Maycock T, Tignor M, Waterfield T, editors. Geneva, Switzerland: IPCC.
- Smith L (1978) Coral reef area and the contributions of reefs to processes and resources of the world's oceans. Nature 273: 225–226.
- 6. Porter JW, Tougas JI (2001) Reef ecosystems: threats to their biodiversity. In: Encyclopedia of Biodiversity. Volume 5, Levin S.A., editor. San Diego: Academic Press. p. 73–95.
 7. Wilkinson C, editor (2004) Status of Coral Reefs of the World 2004.
- Wilkinson C, editor (2004) Status of Coral Reefs of the World: 2004. Volume 1. Townsville, Queensland, Australia: Australian Institute of Marine Science. p. 301.
- Chen P-Y, Chen C-C, Chu LF, McCarl B (2015) Evaluating the economic damage of climate change on global coral reefs. Global Environ Change 30: 12

 –20.

 references
- 9. Guidi L, Chaffron S, Bittner L, Eveillard D, Larhlimi A, Roux S, et al. (2016) Plankton networks driving carbon export in the oligotrophic ocean. Nature 532: 465–470. https://doi.org/10.1038/nature16942 PMID: 26863193
- Karsenti E, Acinas SG, Bork P, Bowler C, De Vargas C, Raes J, et al. (2011) A holistic approach to marine eco-systems biology. PLoS Biol 9(10): e1001177. https://doi.org/10.1371/journal.pbio.1001177
 PMID: 22028628
- 11. Bork P, Bowler C, de Vargas C, Gorsky G, Karsenti E, Wincker P (2015) Tara Oceans studies plankton at planetary scale. Science 348: 873–873. https://doi.org/10.1126/science.aac5605 PMID: 25999501
- 12. Darwin C (1842) The Structure and Distribution of Coral Reefs. Being the First Part of the Geology of the Voyage of the 'Beagle', under the command of capt. Fitzroy R. N. London: Smith, Elder & Co.
- 13. Rohwer F, Seguritan V, Azam F, Knowlton N (2002) Diversity and Distribution of Coral-Associated Bacteries Mar Ecol Progress Ser 243: 1–10.
- 14. Huang D, Roy K (2015) The future of evolution ary diversity in reef corals. Phil Trans R Soc B 370: 20140010
- Shinzato C, Shoguchi E, Kawashima T, Hamada M, Hisata K, Tanaka M, et al. (2011) Using the Acropora digitifera genome to understand coral responses to environmental change. Nature 476: 320–323. https://doi.org/10.1038/nature10249 PMID: 21785439
- 16. Voolstra CR, Li Y, Liew YJ, Baumgarten S, Zoccola D, Flot J-F, et al. (2017) Comparative analysis of the genomes of *Stylophora pistillata* and *Acropora digitifera* provides evidence for extensive differences between species of corals. Scientific Reports 7: 17583. https://doi.org/10.1038/s41598-017-17484-x
 PMID: 29242500
- 17. LaJeunesse TC, Parkinson JE, Gabrielson PW, Jeong HJ, Reimer JD, Voolstra CR, Santos SR (2018) Systematic Revision of Symbiodiniaceae Highlights the Antiquity and Diversity of Coral Endosymbionts. Current Biol 28: 873–875.
- Thornhill DJ, Howells EJ, Wham DC, Steury TD, Santos SR (2017) Population genetics of reef coral endosymbionts (Symbiodinium, Dinophyceae). Mol Ecol 26: 2640–2659. https://doi.org/10.1111/mec 14055 PMID: 28188662



- references
- 19. Pante E, Puillandre E, Viricel A, Arnaud-Haond S, Aurelle D, Castelin M, et al. (2015) Species are hypotheses: avoid connectivity assessments based on pillars of sand. Mol Ecol 24: 525–544. https://referencesrg/10.1111/mec.13048 PMID: 25529046
- 20. Jessen C, Villa Lizcano JF, Bayer T, Roder C, Aranda M, Wild C, Voolstra CR (2013) In situ Effects of Eutrophication and Overfishing on Physiology and Bacterial Diversity of the Red Sea Coral Acropora refere hemorichii. PLoS ONE 8(4): e62091. https://doi.org/10.1371/journal.pone.0062091 PMID: 23630625
- 21. Zaneveld JR, Burkepile DE, Shantz AA, Pritchard CE, McMinds R, Payet JP, et al. (2016) Overfishing and nutrient pollution interact with temperature to disrupt coral reefs down to microbial scales. Nature Comunications 7: 11833.
- 22. Pogoreutz C, Rädecker N, Cárdenas A, Gärdes A, Wild C, Voolstra CR (2018) Dominance of Endozoi-comonas bacteria throughout coral bleaching and mortality suggests structural inflexibility of the Pocillopora verrucosa microbiome. Ecology and Evolution 8: 2240–2252. https://doi.org/10.1002/ece3.3830 references.
- 23. Ziegler M, Seneca FO, Yum LK, Palumbi SR, Voolstra CR (2017) Bacterial community dynamics are linked to patterns of coral heat tolerance. Nature Communications 8: 14213. https://doi.org/10.1038/referencesms14213 PMID: 28186132
- 24. Pogoreutz C, Rädecker N, Cárdenas A, Gärdes A, Voolstra CR, Wild C (2017) Sugar enrichment provides evidence for a role of nitrogen fixation in coral bleaching. Global Change Biology 23: 3838–3848. https://doi.org/10.1111/gcb.13695 PMID: 28429531
- 25. McDervitt-Irwin J, Garren M, Baums J, Vega Thurber R (2017) Responses of coral-associated bacterial references munity responses to local and global stressors. Frontiers of Marine Science 4(262): e00143.
- **26.** Pollock JF, McMinds R, Smith S, Bourne DG, Willis BL, Medina M, et al. (2018) Coral-associated bacterefre demonstrate phylosymbiosis and cophylogery. Nature Com 9: 4921.
- 27. Neave MJ, Michell CT, Apprill A, Voolstra CR (2017) Endozoicomonas genomes reveal functional adaptation and plasticity in bacterial strains symbiotically associated with diverse marine hosts. Scientific Reports 7: 40579. https://doi.org/10.1038/srep40579 PMID: 28094347
- 28. Neave MJ, Rachmawati R, Xun L, Michell CT, Bourne DG, Apprill A, Voolstra CR (2017) Differential specificity between closely related corals and abundant *Endozoicomonas* endosymbionts across global scales. ISME J 11: 186–200. https://doi.org/10.1038/ismej.2016.95 PMID: 27392086
- 29. Güemes AGB, Youle M, Cantu VA, Felts B, Nulton J, Rohwer F (2016) Viruses as the Winners in the Game of Life. Annual Reviews of Virology 3: 197–214. references
- 30. Vega Thurber R, Payet JP, Thurber AR, Correa AMS (2017) Virus—host interactions and their roles in coral reef health and disease. Nature Reviews Microbiology 15: 205–216. https://doi.org/10.1038/references
- 31. Pesant S, Not F, Picheral M, Kandels-Lewis S, Le Bescot N, Gorsky G, et al. (2015) Open science resources for the discovery and analysis of Tara Oceans data. Sci Data 2: 150023. https://doi.org/10.1038/sdata.2015.23 PMID: 26029378 references
- 32. Wilkinson MD, Dumontier M, Aalbersherg IJ, Appleton G, Axton A, Blomberg N, et al. (2016) The fair guiding principles for scientific data management and stewardship. Science Data 3: 160018.
- 33. de Vargas C, Audic S, Henry N, Decelle J, Mahe F, Logares R, et al. (2015) Ocean plankton. Eukaryotic plankton diversity in the sunlit ocean. Science 348: 1261605. https://doi.org/10.1126/science.1261605
- 34. Decelle J, Carradec Q, Pochon X, Henry N, Romac S, Mahé S, et al. (2018) Worldwide Occurrence and Activity of the Reef-Building Coral Symbiont Symbiodinium in the Open Ocean. Current Biol 28: 3625—referen 3633.
- 35. Sunagawa S, Karsenti E, Bowler C, Bork P (2015) Computational eco-systems biology in Tara Oceans: translating data into knowledge. Mol Syst Biol 11: 809. https://doi.org/10.15252/msb.20156272 PMID: references
- 36. Brum JR, Ignacio-Espinoza JC, Roux S, Doulcier G, Acinas SG, Alberti A, et al. (2015) Ocean plankton. Patterns and ecological drivers of ocean viral communities. Science 348: 1261498. https://doi.org/10.1126/science.1261498 PMID: 25999515
- 37. Walters W, Hyde ER, Berg-Lyons D, Ackermann G, Humphrey G, Parada A, et al. (2015) Improved bacterial 16S rRNA gene (V4 and V4-5) and fungal internal transcribed spacer marker gene primers for microbial community surveys. mSystems 1(1):e00009–15. https://doi.org/10.1128/mSystems.00009-15 PMID: 27822518
- Arif C, Daniels C, Bayer T, Banguera-Hinestroza E, Barbrook A, Howe CJ, et al. (2014) Assessing Symbiodinium diversity in scleractinian corals via next-generation sequencing-based genotyping of the ITS2 rDNA region. Molecular Ecology 23(17): 4418–33. https://doi.org/10.1111/mec.12869 PMID: 25052021



<u>references</u>

- Hume CC, Ziegler M, Poulain J, Pochon X, Romac S, Boissin E., et al. (2018) An improved primer set and amplification protocol with increased specificity and sensitivity targeting the Symbiodinium ITS2 region. PeerJ 6: e4816. https://doi.org/10.7717/peerj.4816 PMID: 29844969
- Hume BCC, Smith EG, Ziegler M, Warrington HJM, Burt JA, LaJeunesse TC, et al. (2019) SymPortal: A 40. novel analytical framework and platform for coral algal symbiont next-generation sequencing ITS2 profiling. refere Molecular Ecology Resources 19(4): 1063–80. https://doi.org/10.1111/1755-0998.13004 PMID: 30740899
- Leray M, Knowlton N (2015) DNA barcoding and metabarcoding of standardized samples reveal patterns of marine benthic diversity. Proc Natl Acad Sci U S A 112(7): 2076–81. https://doi.org/10.1073/ pnas.1424997112 PMID: 25646458
- Gregory A, Zayed A, Jang HB, Conceicao-Neto N, Temperton B, Bolduc B, et al. (2019) Marine DNA viral macro- and micro-diversity from pole to pole. Cell 177: 1109-1123. https://doi.org/10.1016/j.cell. 2019.03.040 PMID: 31031001 references
- Roux S, Brum JR, Dutilh BE, Sunagawa S, Duhaime MB, Loy A, et al. (2016) Ecogenomics and potential biogeochemical impacts of globally abundant ocean viruses. Nature 537: 689-693. https://doi.org/ references 10.1038/nature19366 PMID: 27654921
- Lima-Mendez G, Faust K, Henry N, Decelle J, Colin S, Carcillo F, et al. (2015) Determinants of community structure in the global plankton interactome. Science 348: 1262073. https://doi.org/10.1126/ referencesce.1262073 PMID: 25999517
- Alberti A, Poulain J, Engelen S, Labadie K, Romac S, Ferrera I, et al. (2017) Viral to metazoan marine plankton nucleotide sequences from the Tara Oceans expedition. Scientific Data 4: 170093. https://doi org/10.1038/sdata.2017.93 PMID: 28763055
- Carradec Q, Pelletier E, Da Silva C, Alberti A, Seeleuthner Y, Blanc-Mathieu R, et al. (2018) A global ocean atlas of eukaryotic genes. Nat Com 9: 373.
- Wegley KL, Haas AF, Nelson CE (2018) Ecosystem microbiology of coral reefs: linking genomic, metabolomic, and biogeochemical dynamics from animal symbioses to reefscape processes. mSystems 3: e00162–17. https://doi.org/10.1128/mSystems.00162-17 PMID: 29556542
- Quinn RA, Vermeij MJA, Hartmann AC, Galtier d'Auriac I, Benler S, Haas A, et al. (2016) Metabolomic of reef benthic interactions reveals a bioactive lipid involved in coral defence. Proc R Soc B 283: 20160469. https://doi.org/10.1098/rspb.2016.0469 PMID: 27122568
- Houle D, Govindaraju DR, Omholt S (2010) Phenomics: the next challenge. Nature Reviews Genetics 49. 11: 855–866. https://doi.org/10.1038/nrg2897/PMID: 21085204
- **50.** Buddemeier RW, Kinzie RA (1976) Coral growth. Oceanogr Mar Biol Ann Rev 14: 183–225, references
- 51. Bessat F, Buigues D (2001) Two centuries of variation in coral growth in a massive Porites colony from Palaeogeography, Palaeoclimatology, Palaeoecology 175: 381–392.

 The standard of the standard Moorea (French Polynesia): a response of ocean-atmosphere variability from south central Pacific.
- Lough JM, Cantin NE (2014) Perspectives on Massive Coral Growth Rates in a Changing Ocean. Biol Bull 226: 187–202. https://doi.org/10.1086/BBLv226n3p187 PMID: 25070864
- 53. Montagna P. McCulloch M. Douville E. López Correa M. Trotter J. Rodolfo-Metalpa R. et al. (2014) Li/ Mg systematics in scleractinian corals: Calibration of the thermometer. Geochim Cosmochim Acta 132: references 310
- D'Olivo JP, Sinclair DJ, Rankenburg K, McCulloch MT (2018) A universal multi-trace element calibration for reconstructing sea surface temperatures from long-lived *Porites* corals: Removing 'vital-effects'. references Cosmochim Acta 239: 109-135.
- Wu HC, Dissard D, Douville E, Blamart D, Bordier L, Tribollet A, et al. (2018) Surface ocean pH variations since 1689 CE and recent ocean acidification in the tropical south Pacific. Nature Communications 9: 2543. https://doi.org/10.1038/s41467-018-04922-1 PMID: 29959313
- D'Olivo JP, Georgiou L, Falter J, DeCarlo TM, Irigoien X, Voolstra CR, et al. (2019) Long-Term Impacts of 56. the 1997–1998 Bleaching Event on the Growth and Resilience of Massive Porites Corals From the Central Red Sea. Geochemistry, Geophysics, Geosystems 2019. https://doi.org/10.1029/2019GC008312
- Hook SE, Gallagher EP, Batley GE (2014) The role of biomarkers in the assessment of aquatic ecosystem health. Integr Environ Assess Manag 10: 327–341. https://doi.org/10.1002/ieam.1530 PMID: 24574147
- Ye J, Renault V, Jamet K, Gilson É (2014) Transcriptional outcome of telomere signalling. Nature Rev 58. Genetics 15: 491–503.
- Key RM, Olsen A, van Heuven S, Lauvset SK, Velo A, Lin X, et al. (2015) Global Ocean Data Analysis Project, Version 2 (GLODAPv2), ORNL/CDIAC+162, NDP-P093. Oak Ridge, Tennessee: Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, US Department of Energy. references://doi.org/10.3334/CDIAC/OTG.NDP093_GLODAPv2
- Lauvset SK, Key RM, Olsen A, van Heuven S, Velo A, Lin X, Schirnick C, et al. (2016) A new global interior ocean mapped climatology: the 1°x1° GLODAP version 2. Earth System Science Data 8, 325-340.