

# The Kok effect revisited

Xinyou Yin , Yuxi Niu, Peter E. L. van der Putten and Paul C. Struik 

Centre for Crop Systems Analysis, Department of Plant Sciences, Wageningen University & Research, PO Box 430, Wageningen 6700 AK, the Netherlands

Author for correspondence:

Xinyou Yin

Tel: +31 317 482348

Email: Xinyou.Yin@wur.nl

Received: 19 February 2020

Accepted: 24 April 2020

New Phytologist (2020) 227: 1764–1775

doi: 10.1111/nph.16638

**Key words:** day respiration, Kok method, photorespiration, photosystem II efficiency, reassimilation, Yin method.

## Summary

- The Kok effect refers to the abrupt decrease around the light compensation point in the slope of net photosynthetic rate vs irradiance. Arguably, this switch arises from light inhibition of respiration, allowing the Kok method to estimate day respiration ( $R_d$ ). Recent analysis suggests that increasing proportions of photorespiration (quantified as  $\Gamma^*/C_c$ , the ratio of  $\text{CO}_2$  compensation point  $\Gamma^*$  to chloroplast  $\text{CO}_2$  concentration,  $C_c$ ) with irradiance explain much of the Kok effect. Also, the Kok method has been modified to account for the decrease in PSII photochemical efficiency ( $\Phi_2$ ) with irradiance.
- Using a model that illustrates how varying  $R_d$ ,  $\Gamma^*/C_c$ ,  $\Phi_2$  and proportions of alternative electron transport could engender the Kok effect, we quantified the contribution of these parameters to the Kok effect measured in sunflower across various  $\text{O}_2$  and  $\text{CO}_2$  concentrations and various temperatures.
- Overall, the decreasing  $\Phi_2$  with irradiance explained c. 12%, and the varying  $\Gamma^*/C_c$  explained c. 25%, of the Kok effect. Maximum real light inhibition of  $R_d$  was much lower than the inhibition derived from the Kok method, but still increased with photorespiration.
- Photorespiration had a dual contribution to the Kok effect, one via the varying  $\Gamma^*/C_c$  and the other via its participation in light inhibition of  $R_d$ .

## Introduction

The Kok effect refers to the abrupt change in the slope of the linear relationship between net photosynthetic rate and irradiance that occurs at very low irradiances, as observed initially in unicellular algae (Kok, 1948, 1949; Healey & Myers, 1971). The switch is reported later in leaves of many higher plant species (e.g. Ishii & Schmid, 1981; Sharp *et al.*, 1984; Villar *et al.*, 1994; Buckley *et al.*, 2017). The slope decreases from the initial higher value to a lower value, mostly at an irradiance value around the light compensation point. This switch has been interpreted as a consequence of light inhibition of respiration, allowing the so-called Kok method to estimate respiration in the light, or day respiration ( $R_d$ ), and quantum yield of  $\text{CO}_2$  assimilation ( $\Phi_{\text{CO}_2}$ ) (see Supporting Information Table S1 for all symbol definitions), using the part of the relationship with the lower slope. The absolute value of the estimated  $R_d$  is lower than the respiration in the dark ( $R_{\text{dk}}$ ) (Fig. 1). The cost of total respiratory activities accounts for c. 40% of gross photosynthetic productivity of whole plants (Gifford, 1995; Amthor, 2010). Light inhibition of respiratory activities also occurs at a stand scale (Gong *et al.*, 2017), suggesting that it is a general phenomenon, and thus would have a significant impact on projecting the net ecosystem carbon fluxes in biomes across the globe (Heskel *et al.*, 2013). For this reason, understanding the Kok effect and its related light inhibition of respiration has continuously received attention (Tcherkez *et al.*, 2017a,b).

The lower estimates of  $R_d$  by the Kok method, relative to  $R_{\text{dk}}$ , have been confirmed by other gas exchange-based methods such as the popular Laisk method (Laisk, 1977). By applying the Laisk

method to different light intensities, it has been shown that  $R_d$  was progressively inhibited by increasing irradiance (Brooks & Farquhar, 1985; Villar *et al.*, 1995). However, this light inhibition has been challenged by the direct measurement of  $R_d$ , which exploits the differences in the time course of labelling by carbon isotopes of photosynthetic, photorespiratory and respiratory pathways. For example, using such techniques, Pinelli & Loreto (2003) suggested a significant refixation of respired and photorespired  $\text{CO}_2$  and Loreto *et al.* (2001) calculated that there would be no significant difference between  $R_d$  and  $R_{\text{dk}}$  if the refixation of  $\text{CO}_2$  released from respiration during illumination were taken into account. Similarly, a recent report using a direct method based on isotopic disequilibrium (Gong *et al.*, 2018) showed that  $R_d$  was underestimated by the Laisk method. Owing to inconsistent reports of this kind, whether the Kok effect was a result of light inhibition of leaf respiration has been under debate over years.

In fact, according to an extended form of the widely used model of Farquhar *et al.* (1980) for describing the electron transport-limited photosynthesis, several other mechanisms could also explain the Kok effect. The extended model expresses the net  $\text{CO}_2$  assimilation rate ( $A$ ) as a function of the photosynthetically absorbed irradiance ( $I_{\text{abs}}$ ) as (Yin *et al.*, 2004, 2006):

$$A = \frac{1 - \Gamma^*/C_c}{4(1 + 2\Gamma^*/C_c)} f_{\text{act}}(\Phi_2 \rho_2 I_{\text{abs}}) - R_d \quad \text{Eqn 1}$$

where  $C_c$  is the  $\text{CO}_2$  concentration at the carboxylating sites of Rubisco,  $\Gamma^*$  is the  $\text{CO}_2$  compensation point in the absence of  $R_d$ ,

$\Phi_2$  is the photochemical efficiency of photosystem II (PSII) electron transport,  $\rho_2$  is the fraction of the absorbed photons partitioned to PSII, and  $f_{\text{aet}}$  is the factor accounting for fractions of alternative electron transport. So, the term  $(\Phi_2 \rho_2 I_{\text{abs}})$  is the flux of PSII electron transport. Parameters  $f_{\text{aet}}$  and  $\rho_2$  can be quantified by the extended model as (Yin *et al.*, 2006):

$$f_{\text{aet}} = 1 - \frac{f_{\text{pseudo}}}{1 - f_{\text{cyc}}} \quad \text{Eqn 2}$$

$$\rho_2 = \frac{1 - f_{\text{cyc}}}{1 - f_{\text{cyc}} + \frac{\Phi_2}{\Phi_1}} \quad \text{Eqn 3}$$

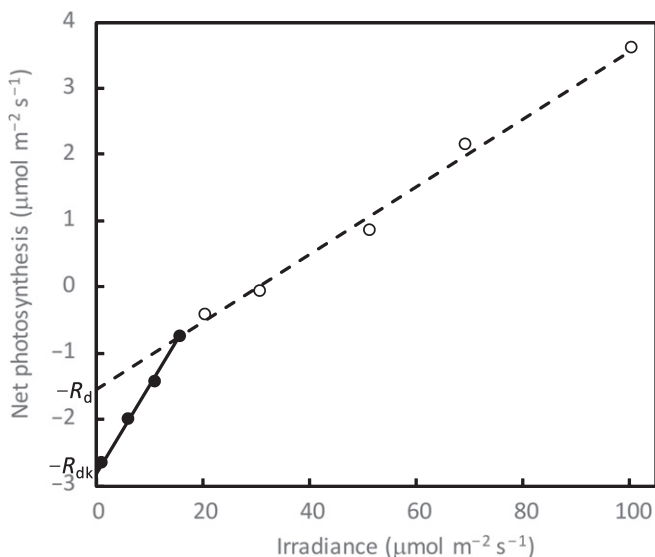
where  $\Phi_1$  is the photochemical efficiency of PSI electron transport,  $f_{\text{cyc}}$  is the fraction of the PSI electron flux that follows the cyclic electron transport around PSI, and  $f_{\text{pseudo}}$  is the fraction of the PSI electron flux that follows the pseudocyclic electron transport (defined as all noncyclic electron-consuming pathways other than the Calvin cycle or the photorespiratory cycle).

Eqn (1) suggests that changes not only in  $R_d$  (Fig. 2a), but also in  $\Gamma^*/C_c$ ,  $\Phi_2$ ,  $f_{\text{aet}}$  and  $\rho_2$ , with increasing  $I_{\text{abs}}$ , result in changes in the slope of  $A$  vs  $I_{\text{abs}}$ . Notably, Farquhar & Busch (2017) recently demonstrated that as a result of regulation of stomatal conductance ( $g_s$ ) and mesophyll conductance ( $g_m$ ),  $C_c$  decreased (thus  $\Gamma^*/C_c$  increased) sharply with increasing  $I_{\text{abs}}$  within the range of low irradiances, and that this phenomenon accounted for much of the observed Kok effect (Fig. 2b). A similar argument might be applied to  $\Phi_2$  (Fig. 2c), as  $\Phi_2$  is not constant but decreases with increasing  $I_{\text{abs}}$  (Genty & Harbinson, 1996), even within the range of low irradiances within which the Kok method is used to estimate  $R_d$  and  $\Phi_{\text{CO}_2}$  (Yin *et al.*, 2011a, 2014). Accounting for

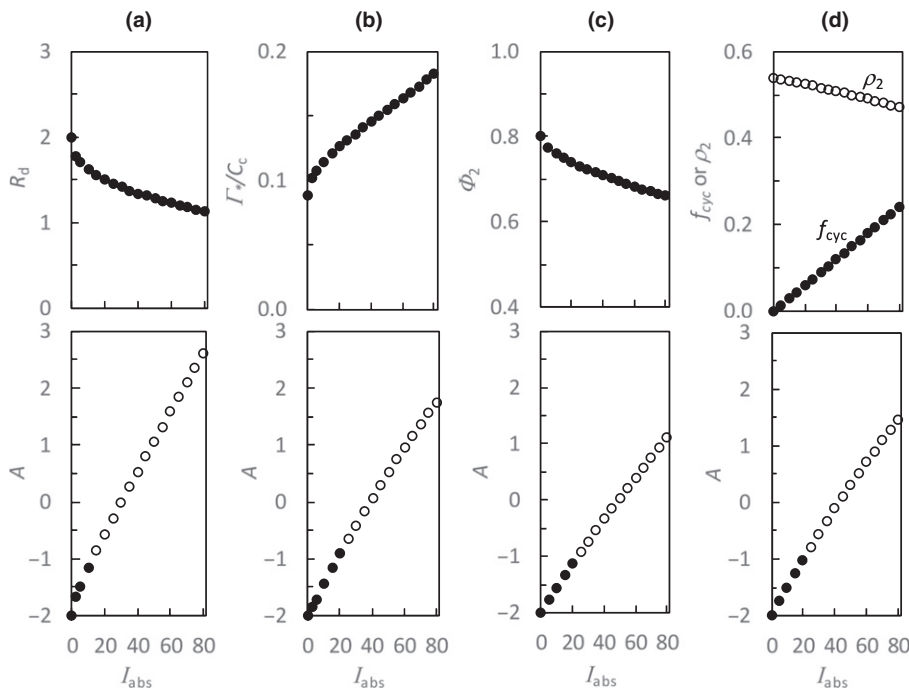
the decrease of  $\Phi_2$  with increasing irradiance has resulted in a modified method to estimate  $R_d$  (Yin *et al.*, 2009, 2011a). The analysis using the modified method, now known as the Yin method (Tcherkez *et al.*, 2017b), indicates that the inhibition of  $R_d$  by light is less than the original Kok method suggests (Yin *et al.*, 2011a).

Less information is available on the change in  $f_{\text{aet}}$  or in  $\rho_2$  with  $I_{\text{abs}}$  that could partly explain the Kok effect. Peltier & Sarrey (1988) indicated that the inhibition of chlororespiration (a process in chloroplasts that involves a respiratory electron transport chain within the thylakoid membrane) might be responsible for the Kok effect. Data of Zhang *et al.* (2018) and Ver Sagun *et al.* (2019) suggested that cyclic electron transport around PSI increased with increasing  $I_{\text{abs}}$ . If this also applies to the limiting light conditions, an increase in  $f_{\text{cyc}}$  with increasing  $I_{\text{abs}}$  would predict a part of the Kok effect (Fig. 2d). According to Eqns 2 and 3,  $f_{\text{cyc}}$  has a dual effect on the expression of the Kok effect, that is, via both terms  $f_{\text{aet}}$  and  $\rho_2$ . Eqn 3 suggests that parameter  $\rho_2$ , related to state transition, could be affected not only by  $f_{\text{cyc}}$  (Fig. 2d) but also by the  $\Phi_2/\Phi_1$  ratio. Tcherkez *et al.* (2017a) speculated the possible role of state transition in the Kok effect. The model of Eqns 1–3 predicts that an increase in  $f_{\text{cyc}}$  or in  $\Phi_2/\Phi_1$  with increasing  $I_{\text{abs}}$  leads to the state transition in favour of PSI with increasing irradiance, and this could engender part of the Kok effect (Fig. 2d).

The Kok effect is not ubiquitous. Early reports found little Kok effect at low  $\text{O}_2$  conditions and in  $\text{C}_4$  plants (Cornic & Jarvis, 1978; Ishii & Murata, 1978). These observations have led to suggestions that photorespiration might be involved in the Kok effect, as confirmed by other studies where photorespiration was manipulated by changing measurement temperatures (Ishii & Schmid, 1981; Way *et al.*, 2019) or by lowering leaf water potential (Sharp *et al.*, 1984). Again, the model analyses of Farquhar & Busch (2017) demonstrated that the change in  $\Gamma^*/C_c$ , therefore, in relative amounts of photorespiration, with increasing  $I_{\text{abs}}$  can explain much of the diminution of the Kok effect in  $\text{C}_4$  plants and at low  $\text{O}_2$  or high  $\text{CO}_2$  concentrations or low temperatures. They also showed that the change in  $\Gamma^*/C_c$  can generate the apparent inhibition of  $R_d$  as inferred by the Laisk method, which is based on  $A-C_i$  curves at two or more irradiances (where  $C_i$  is the intercellular  $\text{CO}_2$  concentration). The decrease in  $C_i$  with increasing  $I_{\text{abs}}$  is the result of stomatal regulation, and its influence on estimates of  $R_d$  by the Kok method was noted by Kirschbaum & Farquhar (1987), who proposed a method to correct for this decrease in  $C_i$ . The further drawdown in  $C_c$ , relative to  $C_i$ , is regulated by  $g_m$  (Evans & von Caemmerer, 1996). The Kok method would underestimate  $R_d$  if light-dependent changes in  $C_i$  (Villar *et al.*, 1994) or in  $C_c$  (Ayub *et al.*, 2011) are not corrected for. Simple  $g_s$  and  $g_m$  models when coupled with photosynthesis models like Eqns 1–3 can account for the refixation of  $\text{CO}_2$  released from respiration and photorespiration (von Caemmerer, 2013), and in fact, the refixation fractions of (photo) respired  $\text{CO}_2$  can be calculated analytically from stomatal, mesophyll and carboxylation resistances (Yin & Struik, 2017). As such, the light inhibition of  $R_d$  predicted for photorespiratory conditions by Farquhar & Busch (2017) and the need to correct



**Fig. 1** Illustration of a two-phase photosynthetic response to low irradiances – the Kok effect. The early interpretation of this effect, as suppressing respiration by light, gave rise to the Kok method to estimate respiration in the light (or ‘day respiration’,  $R_d$ , the intercept of phase 2; open symbols with the dashed line), which is lower than respiration in the dark ( $R_{\text{dk}}$ , the intercept of phase 1 (closed symbols with the solid line).



**Fig. 2** Illustration of the impact of varying values of four parameters (day respiration,  $R_d$ ; ratio of  $\text{CO}_2$  compensation point to chloroplast  $\text{CO}_2$  concentration,  $\Gamma^*/C_c$ ; photosystem II photochemical efficiency,  $\Phi_2$ ; and fraction for cyclic electron transport,  $f_{\text{cyc}}$ ) with absorbed irradiance  $I_{\text{abs}}$  (upper panels of a–d, respectively) on the shape of the light response curve of net photosynthesis ( $A$ , lower panels), where there seems to be a transition from a higher-slope phase (closed symbols) to a lower-slope phase (open symbols). Curves in lower panels are generated from Eqns 1–3, in which, when showing the impact of one parameter, other parameters were kept constant. Units are as follows:  $I_{\text{abs}}$ ,  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ;  $R_d$  and  $A$ ,  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ;  $\Phi_2$ ,  $\text{mol mol}^{-1}$ ;  $\Gamma^*/C_c$ , unitless; and  $f_{\text{cyc}}$  and excitation partitioning to PSII  $\rho_2$ , fractions.

for light-dependent changes in  $C_i$  and  $C_c$  are basically analogous to the statement of Loreto *et al.* (2001) that the lower  $R_d$  than  $R_{\text{dk}}$  resulted from the failure of the original Kok or Laik methods in accounting for the refixation of respired  $\text{CO}_2$  in the light.

However, there are cases where the Kok effect is not always associated with photorespiration. The change in the slope was occasionally observed to be present under high- $\text{CO}_2$  conditions (Sharp *et al.*, 1984), and in  $C_4$  leaves and under low- $\text{O}_2$  conditions albeit to a smaller extent (Yin *et al.*, 2011a). Gong *et al.* (2015) reported an even lower  $R_d : R_{\text{dk}}$  ratio in  $C_4$  than in  $C_3$  leaves. Buckley *et al.* (2017) observed a similar extent of change in the slope under both 21% and 2%  $\text{O}_2$  conditions for broad-bean (*Vicia faba*) mature leaves. Nevertheless, the Kok effects reported in the early years (Kok, 1949; Ishii & Schmid, 1981; Sharp *et al.*, 1984) are mostly associated with the abrupt transition in the slope (Fig. 1), whereas the  $g_s$ – $g_m$  photosynthesis model predicts only a smooth transition (Farquhar & Busch, 2017).

Of the possible mechanisms ( $R_d$ ,  $\Gamma^*/C_c$ ,  $\Phi_2$ ,  $f_{\text{act}}$  and/or  $\rho_2$ ) highlighted by Eqns 1–3 that potentially explain the magnitude of the Kok effect (Fig. 2),  $f_{\text{act}}$  and  $\rho_2$  are hard to measure accurately by existing equipment, especially at low irradiances along the Kok curve. Also, the pattern of changing  $R_d$  in response to  $I_{\text{abs}}$  is hard to quantify with existing methods. In this study, we will illustrate, using Eqns 1–3, that how  $R_d$  responds to  $I_{\text{abs}}$  would have relevance to the Kok effect and in estimating  $\Phi_{\text{CO}_2}$ . We surmise that if the varying  $\Gamma^*/C_c$  ratio is a major factor accounting for the Kok effect, as stated by Farquhar & Busch (2017), then the magnitude of the Kok effect should be associated with the  $\Gamma^*/C_c$  ratio, regardless of how the variation of this ratio is created. To this end, we designed an experiment in which we used various  $\text{O}_2$  and  $\text{CO}_2$  concentrations or temperatures to generate varying relative amounts of photorespiration, that is, various  $\Gamma^*/C_c$  ratios. Based on a modelling analysis of the

experimental data we quantitatively assess: whether the change of  $\Gamma^*/C_c$  and the decrease of  $\Phi_2$  with increasing  $I_{\text{abs}}$  could explain, in part, the Kok effect; if so, what the relative contribution of the two components is in determining the Kok effect; and what the maximum real inhibition of  $R_d$  by light is. We demonstrate that our results help to identify common threads explaining seemingly contradictory findings among previous studies on  $R_d$ .

## Materials and Methods

### Plant materials and measurements

Plants of sunflower (*Helianthus annuus*, cv ‘Sunspot’) were grown in pots in a growth chamber (day : night temperature, 25 : 20°C; relative humidity, 70%; photon flux density, *c.* 500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  at the soil level; photoperiod, 16 h, 06:00–22:00 h) in Wageningen. Five seeds were sown and seedlings were thinned to one plant per 7 l pot. Initial amounts of soil nitrogen (N), phosphorus (P) and potassium (K) were 0.62, 0.83, and 1.04 g per pot, respectively. Nutrient solution was added two or three times per week based on the expected plant growth. Seeds were sown weekly for 4 wk, creating four replications. Measurements were conducted on the 11<sup>th</sup> or 12<sup>th</sup> fully expanded leaf counting from the bottom, using one plant per replication.

An open gas exchange system (Li-Cor 6800; Li-Cor Inc., Lincoln, NE, USA) and an integrated fluorescence chamber head of 6  $\text{cm}^2$  were used for three sets of measurements, in which various  $\text{O}_2$  or  $\text{CO}_2$  concentrations or various temperatures were used to create different amounts of photorespiration (Table 1). The first set used five  $\text{O}_2$  concentrations. Four cylinders containing different mixtures of  $\text{O}_2$  and  $\text{N}_2$  were used. Gas from the cylinder was supplied to the Li-Cor 6800 where  $\text{CO}_2$  was blended with  $\text{O}_2$ . For the second set, five different ambient  $\text{CO}_2$  ( $C_a$ )

concentrations in the leaf chamber were used (Table 1). For the third set, four leaf temperatures were used (Table 1). A flow rate of  $200 \mu\text{mol s}^{-1}$  was used, and leaf-to-air vapour pressure difference was maintained within 0.8–1.6 kPa, for all measurements.

For a given  $\text{O}_2$ ,  $\text{CO}_2$  or temperature, a photosynthetic response curve to incident irradiance ( $A-I_{\text{inc}}$ ) was measured. Leaves were first acclimated under  $80 \mu\text{mol m}^{-2} \text{s}^{-1}$  until  $A$  reached a steady state, which took *c.* 45 min. Measurements were then undertaken using a sequence of 80, 70, 60, 50, 40, 30, 25, 20, 15, 10, and  $5 \mu\text{mol m}^{-2} \text{s}^{-1}$ , with 6 min for each step. For measurements in each of the first two sets,  $\text{O}_2$  or  $\text{CO}_2$  concentrations were chosen randomly. Measurements of the temperature set were conducted after the  $\text{O}_2$  and  $\text{CO}_2$  sets to avoid possible after-effects of high temperature on leaves. For the same reason, the four temperatures were set up from low to high rather than randomly.

After the measurements for  $A-I_{\text{inc}}$  curves,  $A-C_i$  curves were determined to provide extra data to estimate  $g_m$ . Leaves were adapted to an  $I_{\text{inc}}$  of  $100 \mu\text{mol m}^{-2} \text{s}^{-1}$  at  $25^\circ\text{C}$  and 21%  $\text{O}_2$  until  $A$  became stable, and curves were measured using a  $C_a$  sequence of 400, 200, 100, 75, 50, 400, 400, 550, 800 and  $1500 \mu\text{mol mol}^{-1}$ , with 3 min per step. Apparent  $A-C_i$  curves were also assessed with heat-killed leaves, which showed that  $\text{CO}_2$  leakage was negligible during our measurement using the Li-Cor 6800.

For each step of either the  $A-I_{\text{inc}}$  or  $A-C_i$  curve, PSII photochemical operating efficiency ( $\Phi_2$ ) was determined by Chl fluorescence as  $(1-F_s/F_m')$ , where  $F_s$  is the steady-state fluorescence and  $F_m'$  is the maximum fluorescence as revealed using the single flash of *c.*  $8500 \mu\text{mol m}^{-2} \text{s}^{-1}$  for a duration of 1.0 s. We did not use the multiphase method to determine  $F_m'$  because all measurements were undertaken at low irradiances.

Leaf spots used for measurements were punched out, and leaf discs were measured for light absorption (STS-VIS miniature spectrometer; Ocean Optics, Dunedin, FL, USA), twice per disc, to represent average absorption at this spot. After measuring their areas, leaf discs were dried in a  $70^\circ\text{C}$  oven for 24 h to determine dry matter. Each dry leaf disc was then ground into powder, and samples of 1–3 mg were analysed for N concentrations with an EA1108 CHN-O Element Analyzer (Fisons Instruments, Waltham, MA, USA) using the Dumas combustion method.

## Data and modelling analyses

All the leaf spots had a similar N content. The average leaf N was  $1.6 \text{ g m}^{-2}$  and average leaf absorptance was 85%. Variation among replications was small, and replicate average values were used for analysis.

**Table 1** Levels of  $\text{O}_2$ , ambient  $\text{CO}_2$  and leaf temperature in three sets of measurements on sunflower leaves.

Set	$\text{O}_2$ (%)	$\text{CO}_2$ ( $\mu\text{mol mol}^{-1}$ )	Temperature ( $^\circ\text{C}$ )
1	2, 10, 21, 35, 50	400	25
2	21	100, 250, 400, 550, 700	25
3	21	400	15, 25, 30, 35

Data of  $A$  vs  $I_{\text{abs}}$  were inspected to identify the irradiance at the Kok transition point ( $I_{\text{abs},t}$ ), based on the highest average  $r^2$  of linear regression on points both below and above the candidate  $I_{\text{abs},t}$  of each curve. The regression slopes below and above  $I_{\text{abs},t}$  were denoted as  $b_1$  and  $b_2$ , respectively, and the  $b_1 : b_2$  ratio was calculated. The intercept of the regression after  $I_{\text{abs},t}$  is the day respiration estimated by the Kok method. Here, the intercepts of regression lines before and after  $I_{\text{abs},t}$  are denoted as  $r_{d1}$  and  $r_{d2}$ , respectively. According to the original interpretation of the Kok effect (Fig. 1),  $r_{d1}$  is equivalent to the respiration rate in the darkness,  $R_{\text{dk}}$ .

To examine if the decrease of  $\Phi_2$  with increasing  $I_{\text{abs}}$  could partly explain the Kok effect, plots of  $A$  vs  $I_{\text{abs}}$   $\Phi_2$  were made. To be compared with the  $A-I_{\text{abs}}$  plots, data points were allocated according to  $I_{\text{abs},t}$  identified earlier, and linear regression slopes both below and above  $I_{\text{abs},t}$  were denoted as  $B_1$  and  $B_2$ , respectively. Any decrease in the  $B_1 : B_2$  ratio, relative to the  $b_1 : b_2$  ratio, would suggest that the decrease of  $\Phi_2$  with increasing  $I_{\text{abs}}$  could partly explain the Kok effect. The intercept of the linear plot of  $A$  vs  $I_{\text{abs}}$   $\Phi_2/4$  after the Kok break point is the day respiration estimated by the Yin method (Yin *et al.*, 2009, 2011a). As the intercept remains unchanged if the linear plot is made here for  $A$  vs  $I_{\text{abs}}$   $\Phi_2$ , the intercepts of  $A-I_{\text{abs}}$   $\Phi_2$  lines before and after  $I_{\text{abs},t}$  are denoted as  $R_{D1}$  and  $R_{D2}$ , respectively.

To assess the impact of  $\Gamma^*/C_c$  on the Kok effect,  $C_c$  has to be known. To that end, we estimated  $g_m$  using all data from combined gas exchange and Chl fluorescence measurements.  $g_m$  is known to vary with temperature (Bernacchi *et al.*, 2002), but whether  $g_m$  varies with  $C_i$  or with  $I_{\text{inc}}$  or  $\text{O}_2$  is uncertain. Furthermore, recent literature suggests the necessity to dissect mesophyll resistance into its components (Tholen *et al.*, 2012) and to consider the intracellular arrangements of organelles (Yin & Struik, 2017; Ubierna *et al.*, 2019; Yin *et al.*, 2020). Here we consider three  $g_m$  modes: mode i assumes that  $g_m$  varies only with temperature, but not with either  $C_i$  or  $I_{\text{inc}}$  or  $\text{O}_2$ ; mode ii assumes that  $g_m$  varies with all these factors; and mode iii is similar to mode ii but uses an additional factor  $m$  that lumps subresistance proportions and several intracellular properties of mesophyll organelles (Yin *et al.*, 2020). For mode i, we estimated  $g_m$  by fitting, the NRH-A method based on the non-rectangular hyperbolic equation for  $\text{CO}_2$ -assimilation, described by Yin & Struik (2009) to all data (including  $A-C_i$  curves). Like photosynthetic rate,  $g_m$  has generally an optimum response to temperature (e.g. Bernacchi *et al.*, 2002; Warren & Dreyer, 2006; but with caution, see von Caemmerer & Evans, 2015), and we assumed that this response followed a normal distribution function, with an optimum temperature of  $30^\circ\text{C}$ :  $g_m = g_{m30} \exp\{-[(T-30)/\Omega]^2\}$ , which has a minimum number of parameters to estimate. We incorporated these relationships into the NRH-A method to fit parameter  $\Omega$ . For modes ii and iii, we used an equation described by Yin *et al.* (2009),  $g_m = \delta(A + R_d)/(C_c - \Gamma^*)$ , which can semi-empirically accommodate the response (if observed) of  $g_m$  to  $I_{\text{inc}}$ ,  $C_i$ ,  $\text{O}_2$  and temperature. Here, it is the unitless coefficient  $\delta$  that is an explicit parameter to be estimated, and  $\delta$  represents the carboxylation resistance : mesophyll resistance ratio (Yin *et al.*, 2020). For each mode, the simultaneously estimated parameters together with  $g_m$  or  $\delta$  were: the calibration factor(s) that converts Chl fluorescence-



based PSII photochemical efficiency ( $\Phi_2$ ) into linear electron transport rate ( $J$ ), with  $J = sI_{inc} \Phi_2$  (Yin *et al.*, 2009); and Rubisco specificity at 25°C ( $S_{c/o25}$ ). The values of  $S_{c/o}$  for other temperatures were calculated from the relation  $\Gamma^* = 0.5 O/S_{c/o}$  (where  $O$  is the concentration of oxygen; Farquhar *et al.*, 1980; von Caemmerer, 2013) and the Arrhenius equation using 24 460 J mol<sup>-1</sup> of Bernacchi *et al.* (2002) as the activation energy for  $\Gamma^*$  (using other activation-energy estimates (e.g. Walker *et al.*, 2013; Yin *et al.*, 2014) had little impact on our calculated  $\Gamma^*/C_c$  ratios). In view of the reasoning of Farquhar & Busch (2017), we used  $R_{D1}$  of each curve as input for the  $R_d$  term of the model in fitting. The fitting procedures for three modes were implemented using the GAUSS method in PROC NLIN of SAS (SAS Institute Inc, Cary, NC, USA), and the SAS codes can be obtained upon request. The SAS output gave the fitted  $A$  for each measurement point, with which  $C_c$  was then solved from the model of Farquhar *et al.* (1980) as:  $C_c = \Gamma^* [J/4 + 2(A + R_d)]/[J/4 - (A + R_d)]$ .

## Results

### Forms of light inhibition of $R_d$ in relation to the Kok effect

We consider all possible scenarios in interpreting the often-said 'progressive' inhibition of respiration by light, and examine, based on Eqn 1, the consequence of these scenarios on the shape of  $A-I_{abs}$  curves within a range of the low irradiances (Fig. 3).

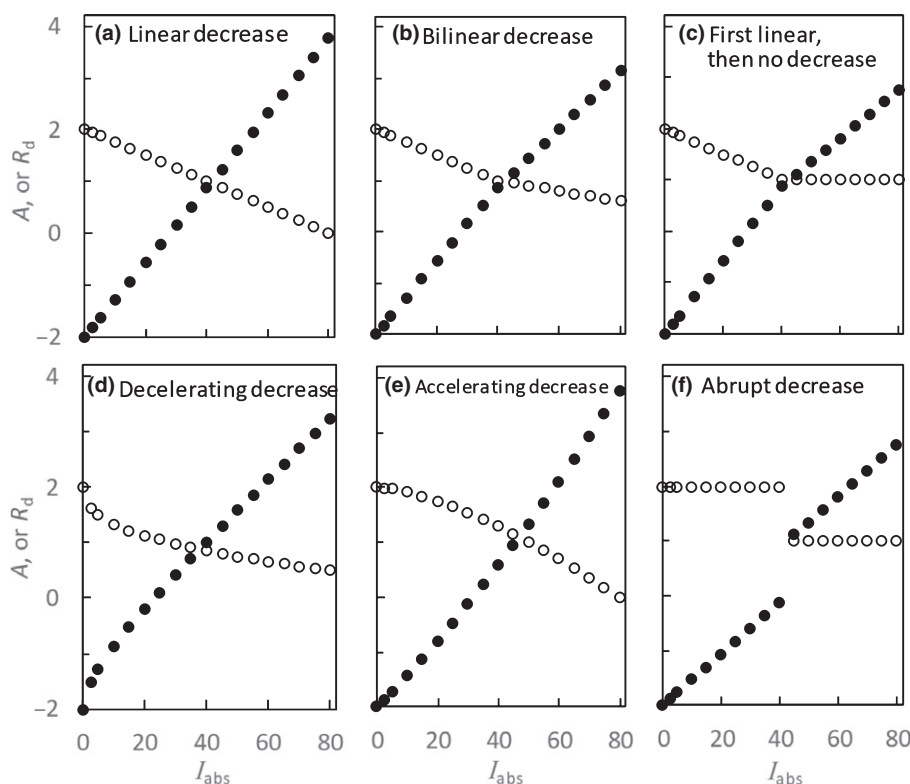
The scenario 'continuously linear decrease' of  $R_d$  with light (Fig. 3a) did not at all result in a break in the linear relationship. Only two 'bilinear' scenarios can generate the Kok effect with an abrupt transition point (Fig. 3b,c). The 'continuously

decelerating decrease' scenario also generated the Kok effect but without the abrupt break point (Fig. 3d). For an 'accelerating decrease' scenario,  $R_d$  was also progressively suppressed by light, but this scenario generated an  $A-I_{abs}$  curve where the slope did not decrease but increased (Fig. 3e), thereby being unable to reproduce the Kok curve. Finally, an 'abrupt suppression' scenario cannot be ruled out, but this scenario generated two linear discontinued segments with the same slope (Fig. 3f), thereby being unable to reproduce the Kok effect either.

As illustrated in Fig. 3, the difference in scenarios also has implications for the estimation of  $\Phi_{CO_2}$ . Only in the second 'bilinear' scenario (Fig. 3c) and the abrupt-suppression scenario (Fig. 3f) can  $\Phi_{CO_2}$  be reliably estimated by the Kok method as the slope of the  $A-I_{abs}$  curve above the break point. For other scenarios, the slope represents the combined yield of photosynthesis and of the component of light suppression of  $R_d$ . In fact, it is the scenario of Fig. 3(c) that the Kok method relies on to estimate  $R_d$  and  $\Phi_{CO_2}$ .

### The observed Kok effect across various O<sub>2</sub> and CO<sub>2</sub> concentrations and various temperatures

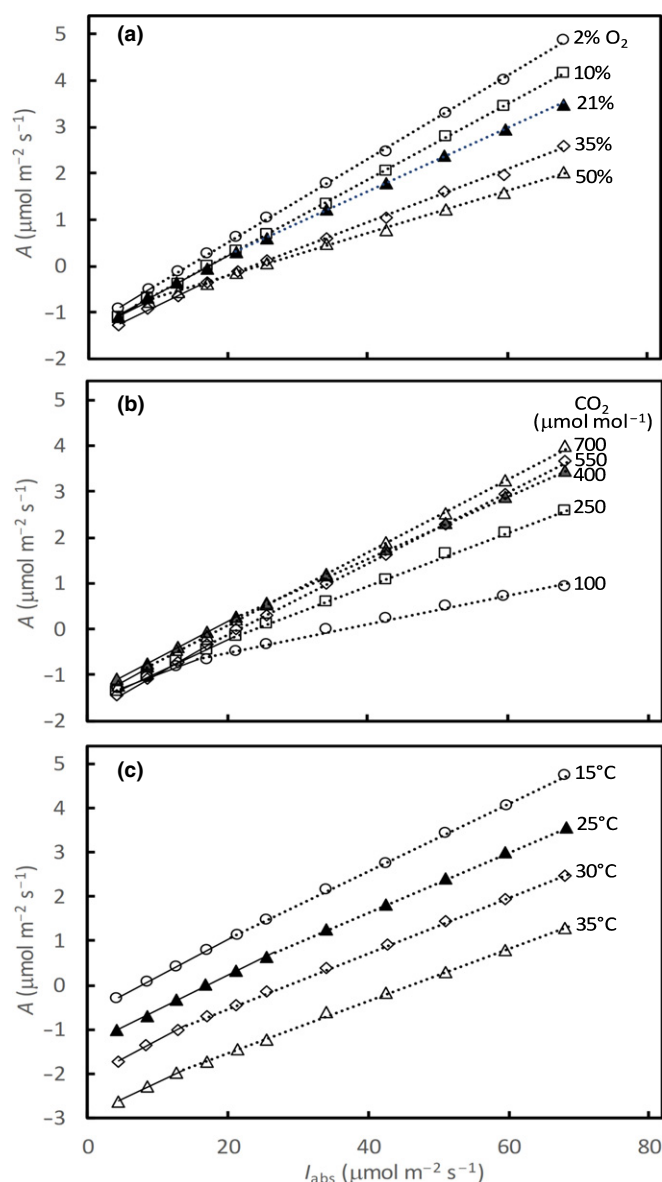
Linear plots of  $A$  vs  $I_{abs}$  using our experimental data identified the Kok break point in each curve (Fig. 4). The maximum values of the slope below (phase 1,  $b_1$ ) and above the break point (phase 2,  $b_2$ ) were achieved at 2% O<sub>2</sub>, and were 0.095 and 0.090 mol mol<sup>-1</sup>, respectively (Table 2), similar to experimentally measured (Long *et al.*, 1993) or theoretically inferred  $\Phi_{CO_2}$  (Yin *et al.*, 2006) for  $C_3$  species under nonphotorespiratory conditions. A change in the slope from phase 1 to phase 2 became more significant with



**Fig. 3** Illustration of six scenarios (a–f) for the so-called 'progressive' decrease of day respiration,  $R_d$ , with absorbed irradiance,  $I_{abs}$  (open circles), and their impact on the shape of the light response curve of net photosynthesis,  $A$  (closed circles). Units:  $I_{abs}$ ,  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ;  $R_d$  and  $A$ ,  $\mu\text{mol m}^{-2} \text{s}^{-1}$ .

increasing  $O_2$  concentrations, with decreasing  $CO_2$  concentrations, and with increasing temperature (Fig. 4). The  $b_1 : b_2$  ratio increased from 1.06 at 2%  $O_2$  to 1.69 at 50%  $O_2$ , from 1.07 at 700  $\mu\text{mol mol}^{-1}$   $CO_2$  to 1.83 at 100  $\mu\text{mol mol}^{-1}$   $CO_2$ , and from 1.10 at 15°C to > 1.30 at 30–35°C (Table 2).

Similarly, the difference in the estimated respiration for phase 1 and phase 2, denoted as  $r_{d1}$  and  $r_{d2}$ , respectively, became more significant with increasing  $O_2$  concentrations, decreasing  $CO_2$  concentrations, and increasing temperature (Table 3). With the estimated  $b_1$ ,  $b_2$ ,  $r_{d1}$  and  $r_{d2}$ , the irradiance for the Kok break point,  $I_{\text{abs},b}$  can be calculated, and it varied from 7 to 27  $\mu\text{mol m}^{-2} \text{s}^{-1}$  (Table 3).



**Fig. 4** Net photosynthesis rates ( $A$ ) vs absorbed irradiance ( $I_{\text{abs}}$ ), based on measured data from five  $O_2$  concentrations (a), five  $CO_2$  concentrations (b), and four temperatures (c) for sunflower leaves. Points represent the means of measurements on four replicated leaves. Continuous lines are for phase 1, and dotted lines are for phase 2, of the Kok plot, drawn from parameter estimates as given in Tables 2 and 3.

## The variable $\Phi_2$ as a possible cause for the Kok effect

As with previous reports (Yin *et al.*, 2011a, 2014),  $\Phi_2$  decreased with increasing irradiances in all three sets of measurements (Fig. S1). Compared with the  $A$  vs  $I_{\text{abs}}$  plots, the  $A$  vs  $I_{\text{abs}} \Phi_2$  plots had a similar shape (thus, they are not shown), but the obtained  $B_1 : B_2$  ratios were slightly lower than the  $b_1 : b_2$  ratios (Table 2). As expected, the regression of  $A$  against  $I_{\text{abs}} \Phi_2$  yielded consistently lower intercepts, and therefore higher estimates,  $R_{D1}$  and  $R_{D2}$ , compared with  $r_{d1}$  and  $r_{d2}$ , respectively, confirming the results of earlier studies (Yin *et al.*, 2011a). For the same reason, the  $R_{D1} : R_{D2}$  ratios were smaller than the  $r_{d1} : r_{d2}$  ratios (Table 3). There were no consistent trends for absolute values of  $r_{d1}$ ,  $r_{d2}$ ,  $R_{D1}$  and  $R_{D2}$  with changing  $O_2$  or  $CO_2$  concentrations; but unsurprisingly they increased consistently with increasing temperature (Table 3).

## Association of the Kok effect with the variable $\Gamma^*/C_c$

We estimated parameter values of the aforementioned three  $g_m$  modes. The estimate of the  $m$  factor for mode iii was 0, which means that modes ii and iii had identical results. To test whether a nonzero  $m$  factor influenced the calculated  $C_c$ , we fixed  $m$  to 0.3, our recent estimate of this parameter (Yin *et al.*, 2020). The three  $g_m$  modes yielded the same goodness of fit with  $R^2$  of 0.966 (Table S2). The modelled  $A$  by the three modes did not differ essentially (Fig. S2a). Using the modelled  $A$ , we calculated the  $\Gamma^*/C_c$  ratio across irradiance of all the three sets of measurements. The  $\Gamma^*/C_c$  ratios calculated by mode ii or iii at first very low irradiances were more variable than those given by mode i (results not shown), but the average  $\Gamma^*/C_c$  ratio along a given  $A-I_{\text{inc}}$  curve did not differ much between the three modes (Fig. S2b). We also used the variable  $J$  method of Harley *et al.* (1992) to inspect any variation of  $g_m$  and found no evidence that  $g_m$  varied with either  $C_i$  or with  $I_{\text{inc}}$  or with  $O_2$ . In the following analysis, we show the results based on the estimate using mode i, as they did not differ much from those using mode ii or iii.

The obtained average  $\Gamma^*/C_c$  ratio varied from 0.008 to 0.195 when  $O_2$  varied from 2% to 50%, from 0.322 to 0.049 when  $CO_2$  varied from 100 to 700  $\mu\text{mol mol}^{-1}$ , and from 0.066 to 0.110 when temperature varied from 15 to 35°C. Plotting the  $b_1 : b_2$  ratio or the  $B_1 : B_2$  ratio against the  $\Gamma^*/C_c$  ratio showed linear relationships, and because these linear relations did not differ significantly among the three sets of measurements, the common regression line was obtained, and the intercept of the line at the zero  $\Gamma^*/C_c$  ratio was close to 1 (Fig. 5a).

## The extents to which the Kok effect was explained by variable $\Phi_2$ and $\Gamma^*/C_c$

The strong correlation of the  $b_1 : b_2$  or  $B_1 : B_2$  ratio with the  $\Gamma^*/C_c$  ratio (with  $R^2 > 0.80$ ; Fig. 5a) does not mean that the varying  $\Gamma^*/C_c$  ratio can explain more than 80% of the Kok effect because other factors (such as  $R_d$  and  $f_{\text{act}}$ ) may vary with  $\Gamma^*/C_c$  as well. However, the relative difference in the slope value between the  $b_1 : b_2$  vs the  $\Gamma^*/C_c$  plot (2.559, Fig. 5a) and the  $B_1 : B_2$  vs the  $\Gamma^*/C_c$

**Table 2** Estimates of the slope values of phase 1 ( $b_1$ ) and phase 2 ( $b_2$ ) in the  $A$  vs  $I_{\text{abs}}$  plot or of the slope values of phase 1 ( $B_1$ ) and phase 2 ( $B_2$ ) in the  $A$  vs  $I_{\text{abs}}\Phi_2$  plot for sunflower leaves.

		$A$ vs $I_{\text{abs}}$ plot			$A$ vs $I_{\text{abs}}\Phi_2$ plot		
		$b_1$	$b_2$	$b_1 : b_2$	$B_1$	$B_2$	$B_1 : B_2$
$\text{O}_2$ (%)	2	0.095 (0.004)	0.090 (0.001)	1.06	0.118 (0.004)	0.115 (0.001)	1.03
	10	0.083 (0.003)	0.082 (0.001)	1.02	0.109 (0.003)	0.106 (0.001)	1.02
	21	0.081 (0.003)	0.068 (0.001)	1.19	0.103 (0.003)	0.090 (0.001)	1.15
	35	0.071 (0.005)	0.057 (0.001)	1.24	0.092 (0.006)	0.076 (0.001)	1.21
	50	0.078 (0.011)	0.046 (0.001)	1.69	0.105 (0.014)	0.063 (0.001)	1.67
$\text{CO}_2$ ( $\mu\text{mol mol}^{-1}$ )	100	0.057 (0.008)	0.031 (0.001)	1.83	0.073 (0.005)	0.042 (0.001)	1.74
	250	0.070 (0.003)	0.058 (0.001)	1.21	0.088 (0.002)	0.075 (0.001)	1.17
	400	0.080 (0.001)	0.068 (0.000)	1.18	0.099 (0.001)	0.086 (0.000)	1.15
	550	0.086 (0.003)	0.078 (0.001)	1.11	0.108 (0.003)	0.101 (0.001)	1.07
	700	0.085 (0.003)	0.080 (0.001)	1.07	0.102 (0.003)	0.103 (0.001)	0.99
Temperature ( $^{\circ}\text{C}$ )	15	0.085 (0.002)	0.077 (0.000)	1.10	0.105 (0.002)	0.098 (0.000)	1.06
	25	0.079 (0.001)	0.068 (0.001)	1.16	0.099 (0.001)	0.088 (0.001)	1.13
	30	0.086 (0.004)	0.063 (0.001)	1.37	0.112 (0.004)	0.085 (0.001)	1.32
	35	0.078 (0.006)	0.059 (0.001)	1.33	0.105 (0.006)	0.081 (0.001)	1.29

The slope values have a unit of  $\text{mol mol}^{-1}$  and standard errors of the estimates are given in brackets; data used for estimation were from the three sets of measurements as described in Table 1.

$A$ , net rate of leaf photosynthesis ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ );  $I_{\text{abs}}$ , irradiance absorbed by leaf photosynthetic pigments ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ );  $\Phi_2$ , photochemical efficiency of photosystem II electron transport ( $\text{mol mol}^{-1}$ ).

**Table 3** Estimates of the intercept values of phase 1 ( $r_{d1}$ ) and phase 2 ( $r_{d2}$ ) in the  $A$  vs  $I_{\text{abs}}$  plot or of the intercept values of phase 1 ( $R_{D1}$ ) and phase 2 ( $R_{D2}$ ) in the  $A$  vs  $I_{\text{abs}}\Phi_2$  plot for sunflower leaves.

		$A$ vs $I_{\text{abs}}$ plot			$A$ vs $I_{\text{abs}}\Phi_2$ plot			
		$r_{d1}$	$r_{d2}$	$r_{d1} : r_{d2}$	$R_{D1}$	$R_{D2}$	$R_{D1} : R_{D2}$	$I_{\text{abs},t}$
$\text{O}_2$ (%)	2	1.33 (0.04)	1.28 (0.04)	1.04	1.34 (0.03)	1.34 (0.03)	1.00	9.1
	10	1.42 (0.03)	1.41 (0.03)	1.01	1.46 (0.03)	1.46 (0.03)	1.00	6.8
	21	1.41 (0.03)	1.14 (0.03)	1.24	1.41 (0.03)	1.21 (0.02)	1.17	21.0
	35	1.55 (0.05)	1.33 (0.05)	1.17	1.56 (0.06)	1.38 (0.05)	1.13	16.0
	50	1.43 (0.07)	1.13 (0.02)	1.26	1.44 (0.07)	1.15 (0.02)	1.25	9.3
$\text{CO}_2$ ( $\mu\text{mol mol}^{-1}$ )	100	1.55 (0.07)	1.14 (0.04)	1.35	1.55 (0.05)	1.17 (0.03)	1.32	15.6
	250	1.64 (0.03)	1.39 (0.03)	1.18	1.64 (0.02)	1.43 (0.02)	1.15	20.9
	400	1.44 (0.01)	1.16 (0.02)	1.24	1.44 (0.01)	1.21 (0.01)	1.19	21.9
	550	1.83 (0.03)	1.68 (0.03)	1.09	1.83 (0.03)	1.73 (0.03)	1.06	17.4
	700	1.57 (0.04)	1.53 (0.04)	1.03	1.54 (0.04)	1.56 (0.04)	0.99	8.5
Temperature ( $^{\circ}\text{C}$ )	15	0.66 (0.02)	0.50 (0.02)	1.32	0.67 (0.01)	0.56 (0.01)	1.18	20.4
	25	1.36 (0.02)	1.06 (0.03)	1.28	1.36 (0.01)	1.12 (0.02)	1.22	26.7
	30	2.10 (0.04)	1.78 (0.02)	1.18	2.10 (0.03)	1.82 (0.02)	1.15	14.0
	35	2.98 (0.06)	2.71 (0.03)	1.10	2.98 (0.06)	2.75 (0.03)	1.08	13.6

The intercept values have a unit of  $\mu\text{mol m}^{-2} \text{s}^{-1}$  and standard errors of the estimates are given in brackets; data used for estimation were from the three sets of measurements as described in Table 1.

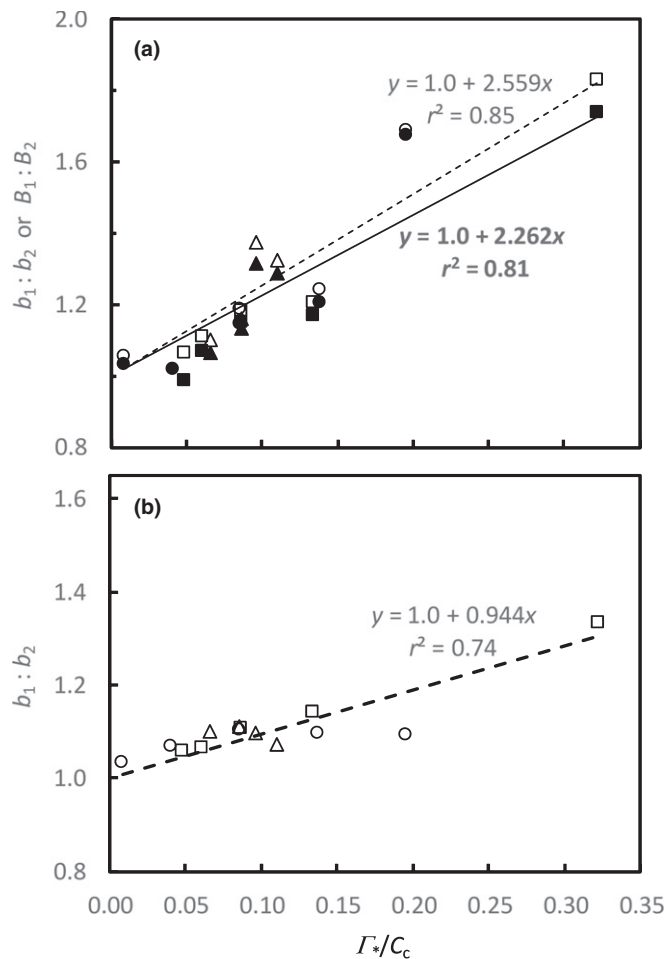
$A$ , net rate of leaf photosynthesis ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ );  $I_{\text{abs}}$ , irradiance absorbed by leaf photosynthetic pigments ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ );  $\Phi_2$ , photochemical efficiency of photosystem II electron transport ( $\text{mol mol}^{-1}$ );  $I_{\text{abs},t}$ , the calculated value of  $I_{\text{abs}}$  ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) for the transition from phase 1 to phase 2 from the  $A$  vs  $I_{\text{abs}}$  plot.

$C_c$  plot (2.262; Fig. 5a) should quantify the contribution of the decreasing  $\Phi_2$  in explaining the Kok effect. This relative difference was  $(2.559 - 2.226) / 2.559 \times 100\% = 11.6\%$ , suggesting that, overall, the varying  $\Phi_2$  explained *c.* 12% of the Kok effect across varying  $\text{O}_2$  and  $\text{CO}_2$  concentrations and varying temperatures.

We plotted the modelled  $A$  against  $I_{\text{abs}}$  to generate slope values of  $b_1$  and  $b_2$ , and thereby the modelled  $b_1 : b_2$  ratios. The modelled  $b_1 : b_2$  ratios did increase with the  $\Gamma^*/C_c$  ratio (Fig. 5b), in line with the statement of Farquhar & Busch (2017) that the changing  $\Gamma^*/C_c$  ratio explains much of the observed Kok effect.

Farquhar & Busch (2017) did not estimate quantitatively the extent of the explanation.

The modelled  $b_1 : b_2$  ratios were lower than the observed  $b_1 : b_2$  ratios shown in Table 2. Our prediction used measured  $C_i$  and  $\Phi_2$  as input and took the effect of  $g_m$  and  $\Gamma^*$  into account, and therefore the effects of varying  $\Phi_2$  and  $\Gamma^*/C_c$  were already considered in the modelling. This suggests that the plot of the modelled  $b_1 : b_2$  ratios vs the  $\Gamma^*/C_c$  ratios should reflect the combined effect of both varying  $\Phi_2$  and varying  $\Gamma^*/C_c$ . The intercept of the plot for the modelled  $b_1 : b_2$  ratios vs the  $\Gamma^*/C_c$  ratios was again close to 1; but its slope was 0.944 (Fig. 5b), much lower



**Fig. 5** The slope ratios of phase 1 to phase 2 in the net photosynthesis rate (A) vs absorbed irradiance ( $I_{\text{abs}}$ ) plot (i.e.  $b_1 : b_2$  ratios; open symbols) or in the A vs  $I_{\text{abs}}\Phi_2$  plot (i.e.  $B_1 : B_2$  ratios; closed symbols) based on measured values of A (a), or the  $b_1 : b_2$  ratios based on modelled values of A (b), plotted against ratios of  $\text{CO}_2$  compensation point to chloroplast  $\text{CO}_2$  concentration ( $\Gamma^*/C_c$ ) across various  $\text{O}_2$  concentrations (circles),  $\text{CO}_2$  concentrations (squares) and temperatures (triangles) for sunflower leaves. Equations represent the regression lines that pass the (0, 1) point.  $\Phi_2$ , photosystem II photochemical efficiency.

than 2.559 – the slope of the observed  $b_1 : b_2$  ratios vs the  $\Gamma^*/C_c$  ratios (Fig. 5a). As the intercept remained unaltered, this indicates that the combined contribution of varying  $\Phi_2$  and  $\Gamma^*/C_c$  to the observed Kok effect can be estimated from slope values, that is, c. 36.9% ( $= 0.944/2.559 \times 100\%$ ). Therefore, the effect of varying  $\Gamma^*/C_c$  alone explained c. 25.3% ( $36.9 - 11.6\%$ ) of the observed Kok effect across various  $\text{O}_2$  and  $\text{CO}_2$  concentrations and various temperatures.

### Quantifying the maximum extent of inhibition of day respiration by light

Our modelling procedure aimed to quantify the contribution of  $\Phi_2$  and varying  $\Gamma^*/C_c$ , and therefore, as is the usual case, assumed that  $R_d$  and  $f_{\text{act}}$  did not vary with irradiance or with measurement  $\text{O}_2$  and  $\text{CO}_2$  conditions. The remaining unexplained contributions (c. 63%) must be a result of light inhibition of  $R_d$  and

possibly variable  $f_{\text{act}}$  and/or  $\rho_2$ . We are not able to separate the contribution of light inhibition of  $R_d$  from the effect of variable  $f_{\text{act}}$  and/or  $\rho_2$  if the variation of  $f_{\text{act}}$  and/or  $\rho_2$  with irradiance cannot be ruled out. If we assume that the variation of either  $f_{\text{act}}$  and/or  $\rho_2$  with irradiance is negligible with the limiting light range, as is often assumed in measuring  $\Phi_{\text{CO}_2}$ , we can quantify the real inhibition of  $R_d$  by light by removing the effect of changing  $\Phi_2$  and  $\Gamma^*/C_c$ , as described in the following. As such, this estimate should be considered as the maximum real inhibition of  $R_d$  by light.

The apparent relative inhibition in case of the Yin method is:

$$\text{Inhibition}_{\text{apparent}}(\%) = \frac{R_{D1\text{measured}} - R_{D2\text{measured}}}{R_{D1\text{measured}}} \times 100 \quad \text{Eqn 4}$$

The similar apparent relative inhibition can be proposed for the Kok method. The apparent inhibition was higher according to the Kok method than according to the Yin method (Fig. 6a), owing to the fact that the Kok method ignores the decrease of  $\Phi_2$  with irradiance. Overall, the Kok method overestimated the apparent inhibition of  $R_d$  by c. 18%, as compared with the Yin method.

Plotting the modelled A against  $I_{\text{abs}}$  resulted in lower estimates of  $r_{d2}$  than  $r_{d1}$  and plotting the modelled A against  $I_{\text{abs}}\Phi_2$  also resulted in lower estimates of  $R_{D2}$  than  $R_{D1}$  than their respective estimates using the observed A (results not shown), although a single value of  $R_d$  was used for each curve in modelling. This confirmed the analysis of Farquhar & Busch (2017) that the apparent inhibition of  $R_d$  by light was partly a result of the artefact of changing  $\Gamma^*/C_c$  with irradiance. The real relative inhibition of  $R_d$  by light can be calculated as:

$$\text{Inhibition}_{\text{real}}(\%) = \frac{(R_{D1\text{measured}} - R_{D2\text{measured}}) - (R_{D1\text{modelled}} - R_{D2\text{modelled}})}{R_{D1\text{measured}}} \times 100 \quad \text{Eqn 5}$$

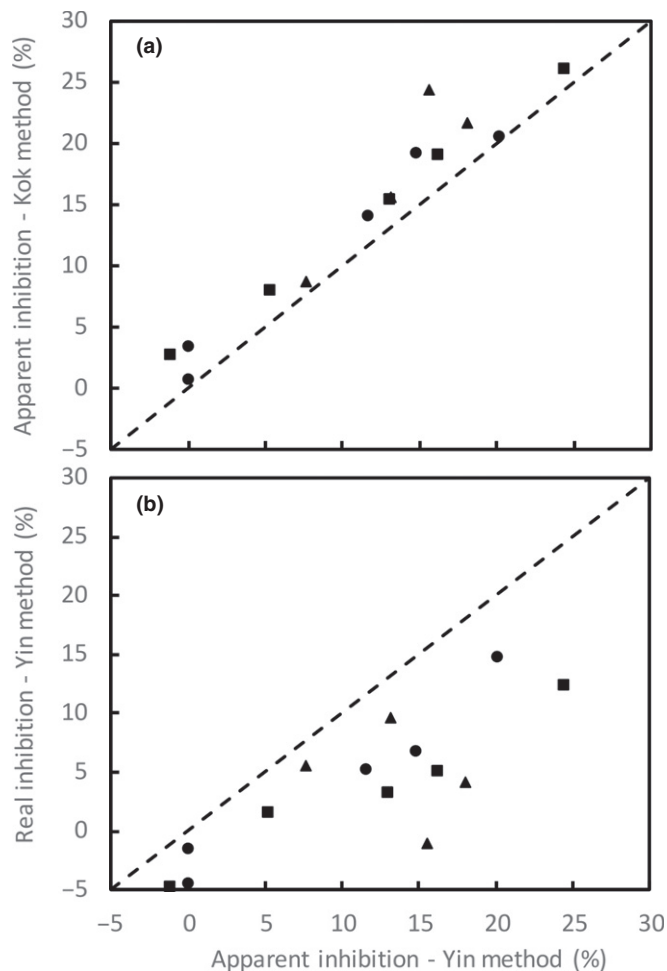
Compared with the relative apparent inhibition from the Yin method, the relative real inhibition was much lower (Fig. 6b). The results also suggested that after correcting for varying  $\Gamma^*/C_c$ , light inhibition of  $R_d$  only became lower but did not disappear: the real inhibition increased generally with relative amounts of photorespiration (Fig. 7).

## Discussion

### The 'linear decrease' of $R_d$ with light cannot generate the Kok effect

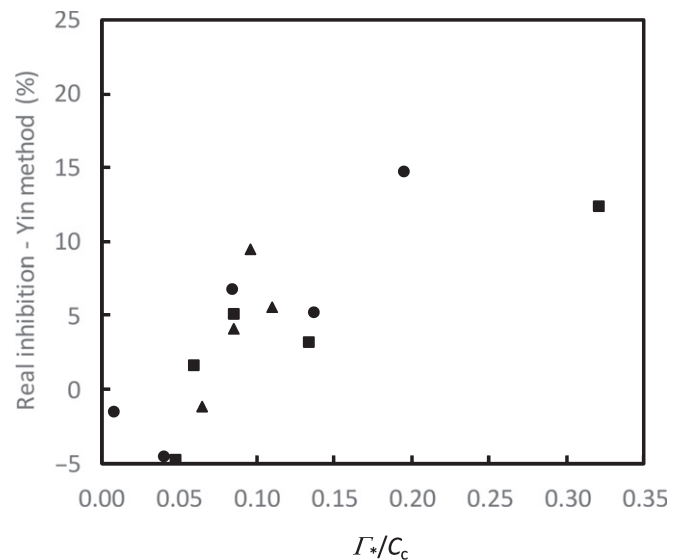
The Kok effect was initially, and is still often, hypothesized to arise from the suppression of respiration by light (Fig. 1; Sharp *et al.*, 1984; Heskell *et al.*, 2013; Tcherkez *et al.*, 2017a; Way *et al.*, 2019). This hypothesis has received support from studies that have identified several mechanisms for the metabolic down-regulation of respiratory reactions by light, as reviewed by





**Fig. 6** Relative apparent light inhibition of day respiration,  $R_d$ , identified by the Kok method vs that identified by the Yin method (a), and relative real light inhibition vs the relative apparent light inhibition of  $R_d$  both identified by the Yin method (b), across various  $O_2$  concentrations (circles),  $CO_2$  concentrations (squares) and temperatures (triangles) for sunflower leaves. The dashed diagonal represents the 1 : 1 line, at which  $y = x$ .

Tcherkez *et al.* (2012, 2017b). Gas exchange measurements have shown that  $R_d$ , relative to  $R_{dk}$ , progressively decreased with increasing  $I_{abs}$ , either in a continuously linear manner (Villar *et al.*, 1994) or in a decelerating manner (Brooks & Farquhar, 1985; Villar *et al.*, 1995; Atkin *et al.*, 2000). Using Eqns 1–3, we assessed the effect of all possible scenarios for the often-said ‘progressive’ inhibition of respiration by light on the shape of  $A-I_{abs}$  curves (Fig. 3). Of the six scenarios considered, only the three scenarios for ‘decelerating decrease’ of  $R_d$  with irradiance (Fig. 3b–d) can generate the Kok effect, thereby excluding the other three scenarios that are often considered relevant to the Kok effect. In particular, the scenario of a ‘continuously linear decrease’ of  $R_d$  with light (Fig. 3a) did not result in a break in the linear  $A-I_{abs}$  relationship. This is in contrast to the statement of Tcherkez *et al.* (2017a) in their report for the 18<sup>th</sup> New Phytologist Workshop that ‘the widely-accepted (historical) origin of the Kok effect is the inhibition of respiratory metabolism by light (linear decrease of  $R_d$  with light)’. Given the consequences of the various scenarios on the Kok effect, and thus also on the estimation of  $\Phi_{CO_2}$ , future



**Fig. 7** The relative real light inhibition of respiration identified by the Yin method plotted against ratios of  $CO_2$  compensation point to chloroplast  $CO_2$  concentration ( $\Gamma^*/C_c$ ) across various  $O_2$  concentrations (circles),  $CO_2$  concentrations (squares), and temperatures (triangles) for sunflower leaves.

studies should aim to reveal which of the three scenarios in Fig. 3 (b)–(d) is most likely for the light inhibition of  $R_d$ .

#### Several mechanisms co-contribute to the Kok effect

Our analyses suggest that not a single mechanism determines the Kok effect, but at least three mechanisms (i.e. decreasing  $\Phi_2$  with irradiance, varying  $\Gamma^*/C_c$ , and light inhibition of  $R_d$ ) co-contribute to it. Using a model, we quantitatively estimated the relative contribution of the  $CO_2$ -specific processes like refixation (reflected via  $\Gamma^*/C_c$ ) vs the light-dependent decrease in photochemical efficiency ( $\Phi_2$ ) in explaining the Kok effect. Our result suggested that varying  $\Gamma^*/C_c$  explained *c.* 25% of the Kok effect, while variable  $\Phi_2$  cannot be ignored and explained *c.* 12% of the Kok effect, across various  $CO_2$ ,  $O_2$  and temperature conditions. The appreciable contribution of variable  $\Phi_2$  is supported by decreases in the slope of phase 2, compared with Phase 1, of the  $A-I_{abs}$  plots under conditions where photorespiration is greatly suppressed, for example, for  $C_3$  species under low- $O_2$  conditions or for  $C_4$  species (Yin *et al.*, 2011a).

However, there are still small decreases in the slope of phase 2 for  $C_3$  species under low- $O_2$  conditions or for  $C_4$  species when  $A$  was plotted against  $I_{abs}\Phi_2$  (Yin *et al.*, 2011a). This effect in  $C_4$  species may reflect the low efficacy of the  $CO_2$ -concentrating mechanism (CCM) caused by a high leakiness at low irradiances (Kromdijk *et al.*, 2010; Yin *et al.*, 2011b). However, for a  $C_3$  species, Buckley *et al.* (2017) observed even more significant changes for developing leaves under 2% than under 21%  $O_2$  conditions, suggesting an involvement of other mechanisms. A fourth mechanism was shown here to potentially contribute to the Kok effect (Fig. 2d), but we were not able to verify it, as any variable  $f_{act}$  and/or  $\rho_2$  are hard to identify at the light intensities showing the Kok effect. Our results, that  $B_1 : B_2$  ratios (Table 2)

and  $R_{D1} : R_{D2}$  ratios (Table 3) were very close to 1 at 2%  $O_2$  or 700  $\mu\text{mol mol}^{-1} \text{CO}_2$  suggest that significant involvement of a fourth mechanism was highly unlikely. Thus, our remaining unexplained part (c. 63%) of the Kok effect is most likely a result of the light suppression of  $R_d$ , in agreement with the statement of Buckley *et al.* (2017) on the dominant role of this third mechanism.

### A dual effect of photorespiration in contributing to the Kok effect

Our strong linear relationships between the  $B_1 : B_2$  ratio and the  $\Gamma^*/C_c$  ratio (Fig. 5a) confirmed previous results in the literature (Cornic & Jarvis, 1978; Ishii & Murata, 1972; Ishii & Schmid, 1981; Sharp *et al.*, 1984; Farquhar & Busch, 2017; Way *et al.*, 2019) showing that the Kok effect was strongly associated with the occurrence of photorespiration. Perhaps it is because of the significant contribution of varying  $\Gamma^*/C_c$  that the Kok effect reported in the earlier days (Kok, 1949; Ishii & Schmid, 1981; Sharp *et al.*, 1984) generally had sharper transition than the recent data (Farquhar & Busch, 2017; Tcherkez *et al.*, 2017a; Way *et al.*, 2019) because  $C_a$  has been increasing over years. However, the contribution of other factors as discussed earlier means that the Kok effect will never disappear in the future high- $\text{CO}_2$  atmosphere; instead, it will continue, but to a lesser extent.

Our modelling analysis suggests that strong associations between the  $B_1 : B_2$  ratio and the  $\Gamma^*/C_c$  ratio shown in Fig. 5(a) are the combined result of a dual effect of photorespiration in contributing to the Kok effect. The first-type effect is what Farquhar & Busch (2017) discussed on the role of increasing  $\Gamma^*/C_c$  with irradiance in explaining the Kok effect, as a result of regulation of  $g_s$  and  $g_m$ . The second-type effect is what we found here – the light inhibition of  $R_d$  identified after removing the first-type effect was still positively correlated with  $\Gamma^*/C_c$  (Fig. 7). Our results suggest that the second-type effect, representing real biological inhibitions, probably contributed more to the Kok effect than the first-type effect.

Farquhar & Busch (2017) demonstrated that the first-type effect of photorespiration on the Kok effect can generate the apparent light inhibition of  $R_d$  for photorespiratory conditions. As stated in the introduction, this inhibition via regulation of  $g_s$  and  $g_m$  is the same as the importance that Loreto *et al.* (2001) emphasized for accounting for the refixation of respired  $\text{CO}_2$  when estimating  $R_d$ . Loreto *et al.* (2001) stated that there would be no significant difference between  $R_d$  and  $R_{dk}$  if the refixation of respiratory  $\text{CO}_2$  during illumination is taken into account. Our results showing that, after correcting for varying  $\Gamma^*/C_c$ , light inhibition of  $R_d$  only became lower but did not disappear (Fig. 6b), do not agree with the conclusion of Loreto *et al.* (2001). The refixation is an important means to reduce the (photo)respiratory loss under photorespiratory conditions, but its net contribution to total photosynthesis should be negligible under nonphotorespiratory conditions (Yin *et al.*, 2020). Berghuijs *et al.* (2019) showed that  $R_d$  estimated by the Kok method was closer to the estimate made by their model (that accounted for the refixation) under nonphotorespiratory than

under photorespiratory conditions. The experiment of Loreto *et al.* (2001) was conducted with maize, a  $C_4$  species where Rubisco is expected to be surrounded by a high  $\text{CO}_2$  partial pressure as a result of the  $C_4$  CCM, and thus the refixation of  $\text{CO}_2$  released from respiration and photorespiration should have little contribution to the total assimilation. Using  $^{14}\text{C}$ -labelling, Pärnik & Keerberg (1995, 2007a,b) showed that light inhibition of  $R_d$  occurs even when accounting for  $\text{CO}_2$  refixation. Gong *et al.* (2015) reported a high suppression of  $R_d$  by light in a  $C_4$  species. If refixation does occur appreciably in  $C_4$  species as Loreto *et al.* stated, it may reflect the refixation more by phosphoenolpyruvate carboxylase than by Rubisco, which might contribute to leakiness.

### Apparent vs real light inhibition of $R_d$

The suppression of  $R_d$  by light has been identified using the Kok method, in many experimental studies, including recent reports based on  $\text{CO}_2$ -exchange measurements (e.g. Buckley *et al.*, 2017) or both  $\text{CO}_2$ - and  $\text{O}_2$ -exchange measurements (e.g. Gauthier *et al.*, 2018). Light is known to suppress the activity of enzymes that involve  $\text{CO}_2$ -releasing pathways contributing to  $R_d$  (Buckley & Adams, 2011; Tcherkez *et al.*, 2012, 2017a,b). Using the model analysis, Farquhar & Busch (2017) demonstrated that at least part of the light inhibition of  $R_d$  can be generated without assuming this inhibition beforehand. Here we used the modelling approach to analyse combined  $\text{CO}_2$ -exchange and Chl fluorescence data. With such combined experimental and modelling analyses, we demonstrated quantitatively that the original Kok method that attributes the Kok effect entirely to the light inhibition of  $R_d$  overestimated the real inhibition (Fig. 6), as a result of ignoring the contribution of varying  $\Phi_2$  and  $\Gamma^*/C_c$  to the Kok effect. The effect of varying  $\Phi_2$  on the Kok method in overestimating the inhibition has been corrected simply by the Yin method, while the correction for varying  $\Gamma^*/C_c$  is more complicated. We previously stressed that both Kok and Yin methods to estimate  $R_d$  actually apply to nonphotorespiratory conditions (Yin *et al.*, 2011a). Our analysis with Eqn 5 suggests an approach to estimate the real light suppression of  $R_d$  for photorespiratory conditions, although we are unable to clarify which one of the three scenarios of suppression in Fig. 3(b–d) is most likely. Most importantly, our analysis using Eqn 5 revealed that the real suppression still increased with relative amounts of photorespiration (Fig. 7). While this new empirical trend receives the support from a theoretical analysis of Buckley & Adams (2011) that photorespiratory NADH may be involved in the suppression, there are probably other underlying biochemical mechanisms that merit further investigation.

### Author contributions

XY conceived the study, XY and PELvdP designed the experiment, YN and PELvdP implemented the experiment and conducted the measurements, XY and YN analysed the data, and XY wrote the draft and finalised it with significant input from PCS.

## ORCID

Paul C. Struik  <https://orcid.org/0000-0003-2196-547X>

Xinyou Yin  <https://orcid.org/0000-0001-8273-8022>

## References

- Amthor JS. 2010. From sunlight to phytomass: on the potential efficiency of converting solar radiation to phyto-energy. *New Phytologist* 188: 939–959.
- Atkin OK, Evans JR, Ball MC, Lambers H, Pons TL. 2000. Leaf respiration of snow gum in the light and dark. Interactions between temperature and irradiance. *Plant Physiology* 122: 915–923.
- Ayub G, Smith RA, Tissue DT, Atkin OK. 2011. Impacts of drought on leaf respiration in darkness and light in *Eucalyptus saligna* exposed to industrial-age atmospheric CO<sub>2</sub> and growth temperature. *New Phytologist* 190: 1003–1018.
- Berghuijs HNC, Yin X, Ho QT, Retta MA, Nicolai BM, Struik PC. 2019. Using a reaction-diffusion model to estimate day respiration and re-assimilation of (photo)respired CO<sub>2</sub> in leaves. *New Phytologist* 223: 619–631.
- Bernacchi CJ, Portis AR, Nakano H, von Caemmerer S, Long SP. 2002. Temperature response of mesophyll conductance. Implication for the determination of Rubisco enzyme kinetics and for limitations to photosynthesis *in vivo*. *Plant Physiology* 130: 1992–1998.
- Brooks A, Farquhar GD. 1985. Effect of temperature on the CO<sub>2</sub>/O<sub>2</sub> specificity of ribulose-1,5-bisphosphate carboxylase/oxygenase and the rate of respiration in the light. *Planta* 165: 397–406.
- Buckley TN, Adams MA. 2011. An analytical model of non-photorespiratory CO<sub>2</sub> release in the light and dark in leaves of C<sub>3</sub> species based on stoichiometric flux balance. *Plant, Cell & Environment* 34: 89–112.
- Buckley TN, Vice H, Adams MA. 2017. The Kok effect in *Vicia faba* cannot be explained solely by changes in chloroplastic CO<sub>2</sub> concentration. *New Phytologist* 216: 1064–1071.
- Cornic G, Jarvis PG. 1972. Effects of oxygen on CO<sub>2</sub> exchange and stomatal resistance in Sitka spruce and maize at low irradiances. *Photosynthetica* 6: 225–239.
- Evans JR, von Caemmerer S. 1996. Carbon dioxide diffusion inside leaves. *Plant Physiology* 110: 339–346.
- Farquhar GD, Busch FA. 2017. Changes in the chloroplastic CO<sub>2</sub> concentration explain much of the observed Kok effect: a model. *New Phytologist* 214: 570–584.
- Farquhar GD, von Caemmerer S, Berry JA. 1980. A biochemical model of photosynthetic CO<sub>2</sub> assimilation in leaves of C<sub>3</sub> species. *Planta* 149: 78–90.
- Gauthier PPG, Battle MO, Griffin KL, Bender ML. 2018. Measurement of gross photosynthesis, respiration in the light, and mesophyll conductance using H<sub>2</sub><sup>18</sup>O labelling. *Plant Physiology* 177: 62–74.
- Genty B, Harbinson J. 1996. Regulation of light utilization for photosynthetic electron transport. In: Baker NR, ed. *Photosynthesis and the Environment. Vol 5 book series 'Advances in Photosynthesis and Respiration'*. The Netherlands: Kluwer Academic Publishers, 67–99.
- Gifford RM. 1995. Whole plant respiration and photosynthesis of wheat under increased CO<sub>2</sub> concentration and temperature: long-term vs. short-term distinctions from modelling. *Global Change Biology* 1: 385–396.
- Gong XY, Schäufele R, Feneis W, Schnyder H. 2015. <sup>13</sup>CO<sub>2</sub>/<sup>12</sup>CO<sub>2</sub> exchange fluxes in a clamp-on leaf cuvette: disentangling artefacts and flux components. *Plant, Cell & Environment* 38: 2417–2432.
- Gong XY, Schäufele R, Lehmeier CA, Tcherkez G, Schnyder H. 2017. Atmospheric CO<sub>2</sub> mole fraction affects stand-scale carbon use efficiency of sunflower by stimulating respiration in light. *Plant, Cell & Environment* 40: 401–412.
- Gong XY, Tcherkez G, Wenig J, Schäufele R, Schnyder H. 2018. Determination of leaf respiration in the light: comparison between an isotopic disequilibrium method and the Laisk method. *New Phytologist* 218: 1371–1382.
- Harley PC, Loreto F, Di Marco G, Sharkey TD. 1992. Theoretical considerations when estimating the mesophyll conductance to CO<sub>2</sub> flux by analysis of the response of photosynthesis to CO<sub>2</sub>. *Plant Physiology* 98: 1429–1436.
- Healey FP, Myers J. 1971. The Kok effect in *Chlamydomonas reinhardtii*. *Plant Physiology* 47: 373–379.
- Heskel MA, Atkin OK, Turnbull MH, Griffin KL. 2013. Bringing the Kok effect to light: A review on the integration of daytime respiration and net ecosystem exchange. *Ecosphere* 4: 1–14.
- Ishii R, Murata Y. 1978. Further evidence of the Kok effect in C<sub>3</sub> plants and the effects of environmental factors on it. *Japanese Journal of Crop Science* 47: 547–550.
- Ishii R, Schmid GH. 1981. The Kok effect and its relationship to photorespiration in tomato. *Zeitschrift für Naturforschung* 36c: 450–454.
- Kirschbaum MUF, Farquhar GD. 1987. Investigation of the CO<sub>2</sub> dependence of quantum yield and respiration in *Eucalyptus pauciflora*. *Plant Physiology* 83: 1032–1036.
- Kok B. 1948. A critical consideration of the quantum yield of *Chlorella*-photosynthesis. *Enzymologia* 13: 1–56.
- Kok B. 1949. On the interrelation of respiration and photosynthesis in green plants. *Biochimica et Biophysica Acta* 3: 625–631.
- Kromdijk J, Griffiths H, Schepers HE. 2010. Can the progressive increase of C<sub>4</sub> bundle sheath leakiness at low PFD be explained by incomplete suppression of photorespiration? *Plant, Cell & Environment* 33: 1935–1948.
- Laik AK. 1977. *Kinetics of photosynthesis and photorespiration in C<sub>3</sub> plant*. Moscow, Russia: Nauka (in Russian).
- Long SP, Postl WF, Bolhár-Nordenkamp HR. 1993. Quantum yields for uptake of carbon dioxide in C<sub>3</sub> vascular plants of contrasting habitats and taxonomic groupings. *Planta* 189: 226–234.
- Loreto F, Velikova V, Di Marco G. 2001. Respiration in the light measured by <sup>12</sup>CO<sub>2</sub> emission in <sup>13</sup>CO<sub>2</sub> atmosphere in maize leaves. *Australian Journal of Plant Physiology* 28: 1103–1108.
- Pärnik T, Keerber O. 1995. Decarboxylation of primary and end products of photosynthesis at different oxygen concentrations. *Journal of Experimental Botany* 46: 1439–1477.
- Pärnik T, Keerber O. 2007a. Advanced radiogasometric method for the determination of the rates of photorespiratory and respiratory decarboxylations of primary and stored photosynthates under steady-state photosynthesis. *Physiologia Plantarum* 129: 33–44.
- Pärnik T, Keerber O. 2007b. Photorespiratory and respiratory decarboxylations in leaves of C<sub>3</sub> plants under different CO<sub>2</sub> concentrations and irradiances. *Plant, Cell & Environment* 30: 1535–1544.
- Peltier G, Sarrey F. 1988. The Kok effect and the light-inhibition of chlororespiration in *Chlamydomonas reinhardtii*. *FEBS Letters* 228: 259–262.
- Pinelli P, Loreto F. 2003. <sup>12</sup>CO<sub>2</sub> emission from different metabolic pathways measured in illuminated and darkened C<sub>3</sub> and C<sub>4</sub> leaves at low, atmospheric and elevated CO<sub>2</sub> concentration. *Journal of Experimental Botany* 54: 1761–1769.
- Sharp RE, Matthews MA, Boyer JS. 1984. Kok effect and the quantum yield of photosynthesis. Light partially inhibits dark respiration. *Plant Physiology* 75: 95–101.
- Tcherkez G, Boex-Fontvieille E, Mahe A, Hodges M. 2012. Respiratory carbon fluxes in leaves. *Current Opinion in Plant Biology* 15: 308–314.
- Tcherkez G, Gauthier P, Buckley TN, Busch FA, Barbour MM, Heskel DA, Gong XY, Crous K, Griffin KL, Way DA *et al.* 2017a. Tracking the origins of the Kok effect, 70 years after its discovery. *New Phytologist* 214: 506–510.
- Tcherkez G, Gauthier P, Buckley TN, Busch FA, Barbour MM, Heskel DA, Gong XY, Crous KY, Griffin K, Way D *et al.* 2017b. Leaf day respiration: low CO<sub>2</sub> flux but high significance for metabolism and carbon balance. *New Phytologist* 216: 986–1001.
- Tholen D, Ethier G, Genty B, Pepin S, Zhu X-G. 2012. Variable mesophyll conductance revisited: theoretical background and experimental implications. *Plant, Cell & Environment* 35: 2087–2103.
- Ubierna N, Cernusak LA, Holloway-Phillips M, Busch FA, Cousins AB, Farquhar GD. 2019. Critical review: incorporating the arrangement of mitochondria and chloroplasts into models of photosynthesis and carbon isotope discrimination. *Photosynthesis Research* 141: 5–31.
- Ver Sagun J, Badger MR, Chow WS, Ghannoum O. 2019. Cyclic electron flow and light partitioning between the two photosystems in leaves of plants with different functional types. *Photosynthesis Research* 142: 321–334.



- Villar R, Held AA, Merino J. 1994. Comparison of methods to estimate dark respiration in the light of leaves of two woody species. *Plant Physiology* 105: 167–172.
- Villar R, Held AA, Merino J. 1995. Dark leaf respiration in light and darkness of an evergreen and a deciduous plant species. *Plant Physiology* 107: 421–427.
- von Caemmerer S. 2013. Steady-state models of photosynthesis. *Plant, Cell & Environment* 36: 1617–1630.
- von Caemmerer S, Evans JR. 2015. Temperature responses of mesophyll conductance differ greatly between species. *Plant, Cell & Environment* 38: 629–637.
- Walker BJ, Ariza LS, Kaines S, Badger MR, Cousins AB. 2013. Temperature response of *in vivo* Rubisco kinetics and mesophyll conductance in *Arabidopsis thaliana*: comparisons to *Nicotiana tabacum*. *Plant, Cell & Environment* 36: 2108–2119.
- Warren CR, Dreyer E. 2006. Temperature response of photosynthesis and internal conductance to CO<sub>2</sub>: results from two independent approaches. *Journal of Experimental Botany* 57: 3057–3067.
- Way DA, Aspinwall MJ, Drake JE, Crous KY, Campy CE, Ghannoum O, Tissue DT, Tjoeker MG. 2019. Responses of respiration in the light to warming in field-grown trees: a comparison of the thermal sensitivity of the Kok and Laik methods. *New Phytologist* 222: 132–143.
- Yin X, Belay DW, van der Putten PEL, Struik PC. 2014. Accounting for the decrease of photosystem photochemical efficiency with increasing irradiance to estimate quantum yield of leaf photosynthesis. *Photosynthesis Research* 122: 323–335.
- Yin X, Harbinson J, Struik PC. 2006. Mathematical review of literature to assess alternative electron transports and interphotosystem excitation partitioning of steady-state C<sub>3</sub> photosynthesis under limiting light. *Plant, Cell & Environment* 29: 1771–1782. (Corrig. in *Plant, Cell & Environment* 29: 2252).
- Yin X, Struik PC. 2009. Theoretical reconsiderations when estimating the mesophyll conductance to CO<sub>2</sub> diffusion in leaves of C<sub>3</sub> plants by analysis of combined gas exchange and chlorophyll fluorescence measurements. *Plant, Cell & Environment* 32: 1513–1524. (Corrig. in *Plant, Cell & Environment* 33: 1595).
- Yin X, Struik PC. 2017. Simple generalisation of a mesophyll resistance model for various intracellular arrangements of chloroplasts and mitochondria in C<sub>3</sub> leaves. *Photosynthesis Research* 132: 211–220.
- Yin X, Struik PC, Romero P, Harbinson J, Evers JB, van der Putten PEL, Vos J. 2009. Using combined measurements of gas exchange and chlorophyll fluorescence to estimate parameters of a biochemical C<sub>3</sub> photosynthesis model: a critical appraisal and a new integrated approach applied to leaves in a wheat (*Triticum aestivum*) canopy. *Plant, Cell & Environment* 32: 448–464.
- Yin X, Sun Z, Struik PC, Gu J. 2011a. Evaluating a new method to estimate the rate of leaf respiration in the light by analysis of combined gas exchange and chlorophyll fluorescence measurements. *Journal of Experimental Botany* 62: 3489–3499.
- Yin X, Sun Z, Struik PC, van der Putten PEL, van Ieperen W, Harbinson J. 2011b. Using a biochemical C<sub>4</sub>-photosynthesis model and combined gas exchange and chlorophyll fluorescence measurements to estimate bundle-sheath conductance of maize leaves differing in age and nitrogen content. *Plant, Cell & Environment* 34: 2183–2199.
- Yin X, van der Putten PEL, Belay D, Struik PC. 2020. Using photorespiratory oxygen response to analyse leaf mesophyll resistance. *Photosynthesis Research* 144: 85–99.
- Yin X, van Oijen M, Schapendonk AHCM. 2004. Extension of a biochemical model for the generalized stoichiometry of electron transport limited C<sub>3</sub> photosynthesis. *Plant, Cell & Environment* 27: 1211–1222.
- Zhang M-M, Fan D-Y, Sun GY, Chow WS. 2018. Optimising the linear electron transport rate measured by chlorophyll *a* fluorescence to empirically match the gross rate of oxygen evolution in white light: towards improved estimation of the cyclic electron flux around photosystem I in leaves. *Functional Plant Biology* 45: 1138–1148.

## Supporting Information

Additional Supporting Information may be found online in the Supporting Information section at the end of the article.

**Fig. S1** Photosystem II photochemical efficiency ( $\Phi_2$ ) as a function of absorbed irradiance ( $I_{\text{abs}}$ ) across various O<sub>2</sub> and CO<sub>2</sub> concentrations and various temperatures.

**Fig. S2** Comparison of net photosynthesis rate  $A$  and the average  $\Gamma^*/C_c$  ratio modelled using three mesophyll conductance  $g_m$  modes as described in the text.

**Table S1** List of all model symbols.

**Table S2** Model parameter values estimated using three mesophyll conductance  $g_m$  modes.

Please note: Wiley Blackwell are not responsible for the content or functionality of any Supporting Information supplied by the authors. Any queries (other than missing material) should be directed to the *New Phytologist* Central Office.