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Quantifying double-stranded DNA with fluorescent dyes: Qubit on BMG LABTECH instruments

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- FLUOstar® Omega, PHERAstar® and CLARIOstar® quantify dsDNA with Qubit dye
- Microplate format increases throughput and facilitates replicate measurements



Introduction

The quantification of DNA is a standard laboratory activity and is pivotal for subsequent applications such as next generation sequencing (NGS). In addition to UV-spectrometry quantitation and qualification of nucleic acids, fluorescent probes provide the required specificity and precision to measure double-stranded DNA (dsDNA) during NGS sample preparation. The widely used Qubit™ nucleic acid staining dyes can be analyzed on a Qubit™ fluorometer. It measures fluorescence intensity directly in sample preparation tubes and automatically calculates the DNA-concentration based on two standards measured in parallel. This provides an easy, quick and intuitive way to quantitate dsDNA. However, throughput is limited using the Qubit™ fluorometer as only one sample can be analyzed at a time. This application note presents how the Qubit® dsDNA HS fluorophore can be used to quantitate dsDNA on a BMG LABTECH microplate reader.



Materials & Methods

- Qubit™ dsDNA HS Assay Kit (ThermoFisherScientific, #Q32854)
- TRIS-EDTA buffer 100x (Carl Roth GmbH & Co. KG)
- 96-well microplate (black, flat bottom, Greiner)

The Qubit working solution was prepared by mixing 60 µL of Qubit dsDNA HS reagent with 11.94 ml Qubit dsDNA buffer to obtain 12 ml working solution. Dilutions of the standard were prepared in 1x TE-buffer.

For Qubit DNA quantification in 96 well plates, 190 µL of Qubit working solution were mixed with 10 µL of DNA standards (0 and 10 ng/µL provided with the Qubit kit) or diluted DNA standard. Fluorescence intensity was measured on FLUOstar Omega, CLARIOstar, and PHERAstar with the following settings.

Instrument settings

Fluorescence intensity, endpoint protocol			
Optic settings	PERAstar	Optic Modules	FI 485 520
	CLARIOstar	Monochromator	Excitation: 483-14 Dichroic: auto 502.5 Emission: 530-30
		Filters	Excitation Ex485 Dichroic: LP504 Emission: 530-40
	FLUOstar Omega	Filters	Excitation: Ex485 Emission: Em520
	Gain and focus adjusted prior to measurement		
	Top optic		
General settings	Settling time: 0.2 s		0.5 s
	Number of flashes per well		20

Results & Discussion



1). The excitation maximum was found to be at 503 nm and the maximum of emitted light at 525 nm, which reasoned to apply 485/520 filter combination or LVF Monochromator™ settings of 483-14/530-30 for Qubit detection.

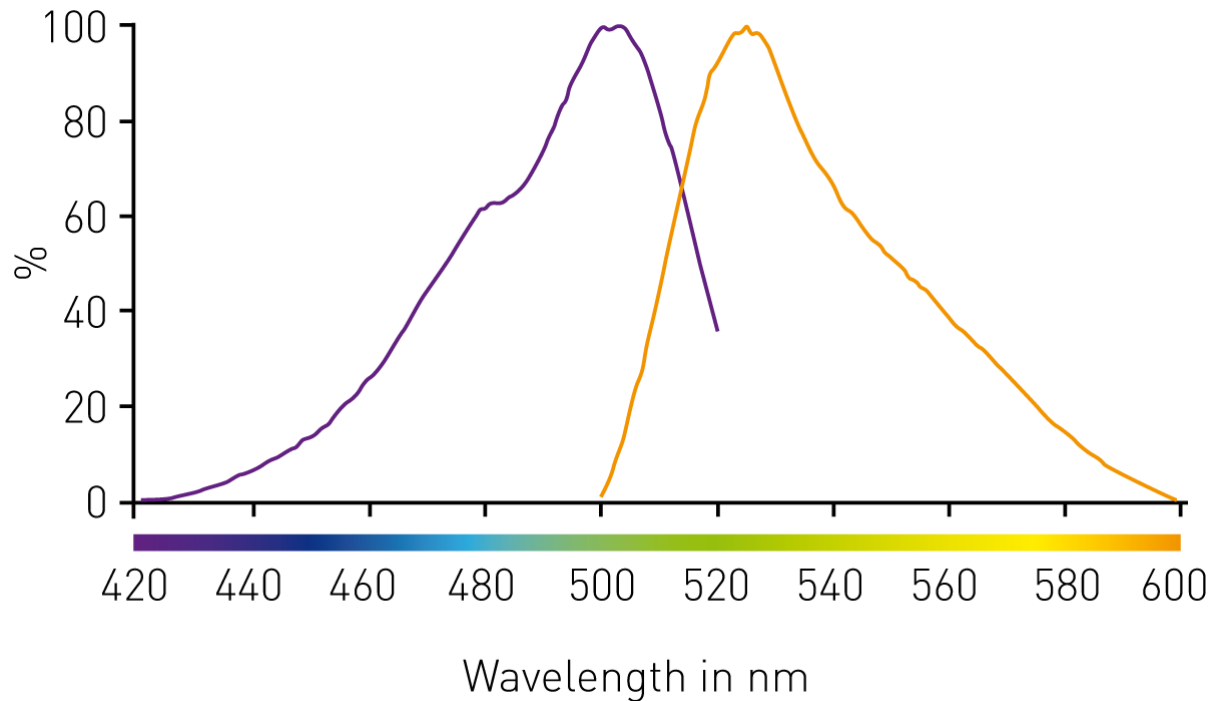


Fig. 1: Excitation (purple) and emission (orange) spectrum of Qubit DNA quantitation dye acquired on CLARIOstar microplate reader.

Next, the linearity of Qubit DNA quantification was tested. To this end, triplicates of Qubit standard DNA dilutions between 0.1 ng/μl-10 ng/μl were detected using the Qubit dye. The fluorescence signal was linear over these concentrations as indicated by a correlation coefficient $R^2 > 0.9999$ (Fig.2). This linearity was obtained on FLUOstar Omega, CLARIOstar, and PHERAstar microplate readers.

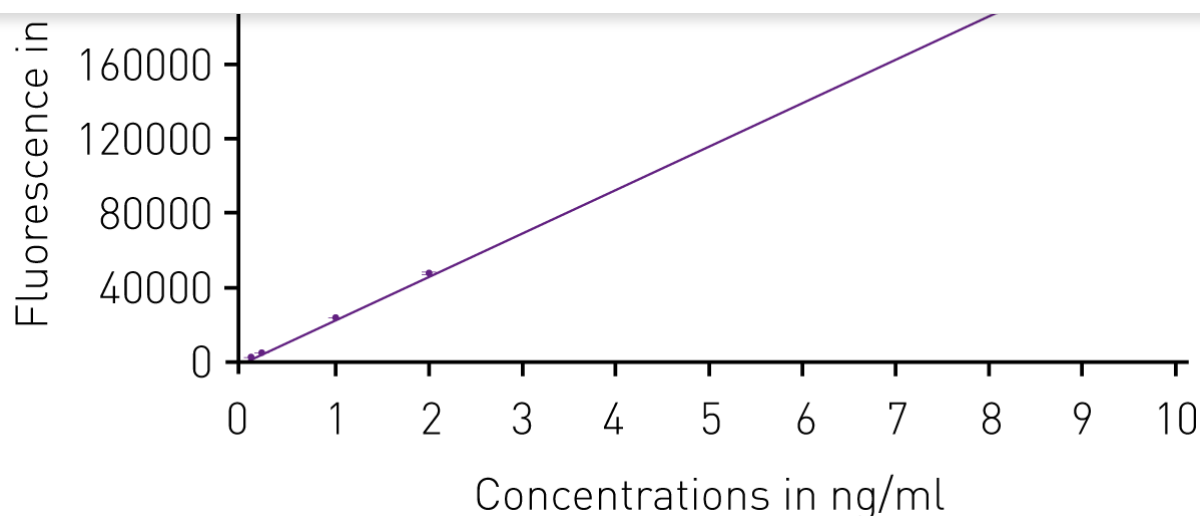


Fig. 2: Linear regression for DNA quantification with Qubit HS read on PHERAstar microplate reader.

Given the linearity of Qubit DNA quantification in the concentration range between 0.1 ng/ μ l-10 ng/ μ l we hypothesized that in accordance with the Qubit measurement, two concentrations for defining a calibration curve might be sufficient. Quantifying known amounts of DNA with Qubit dsDNA HS chemistry and with a standard curve based on triplicates of the 0 and 10 ng/ μ l DNA standard, allowed to accurately determine the amount of DNA (Fig. 3). Taking into account that the sample volume may vary in the range between 1 μ l and 20 μ l as it is suggested by the manufacturer, the measurable DNA sample concentration range that can be quantified with Qubit on BMG LABTECH readers lies between 50 pg/ μ l-100 ng/ μ l.

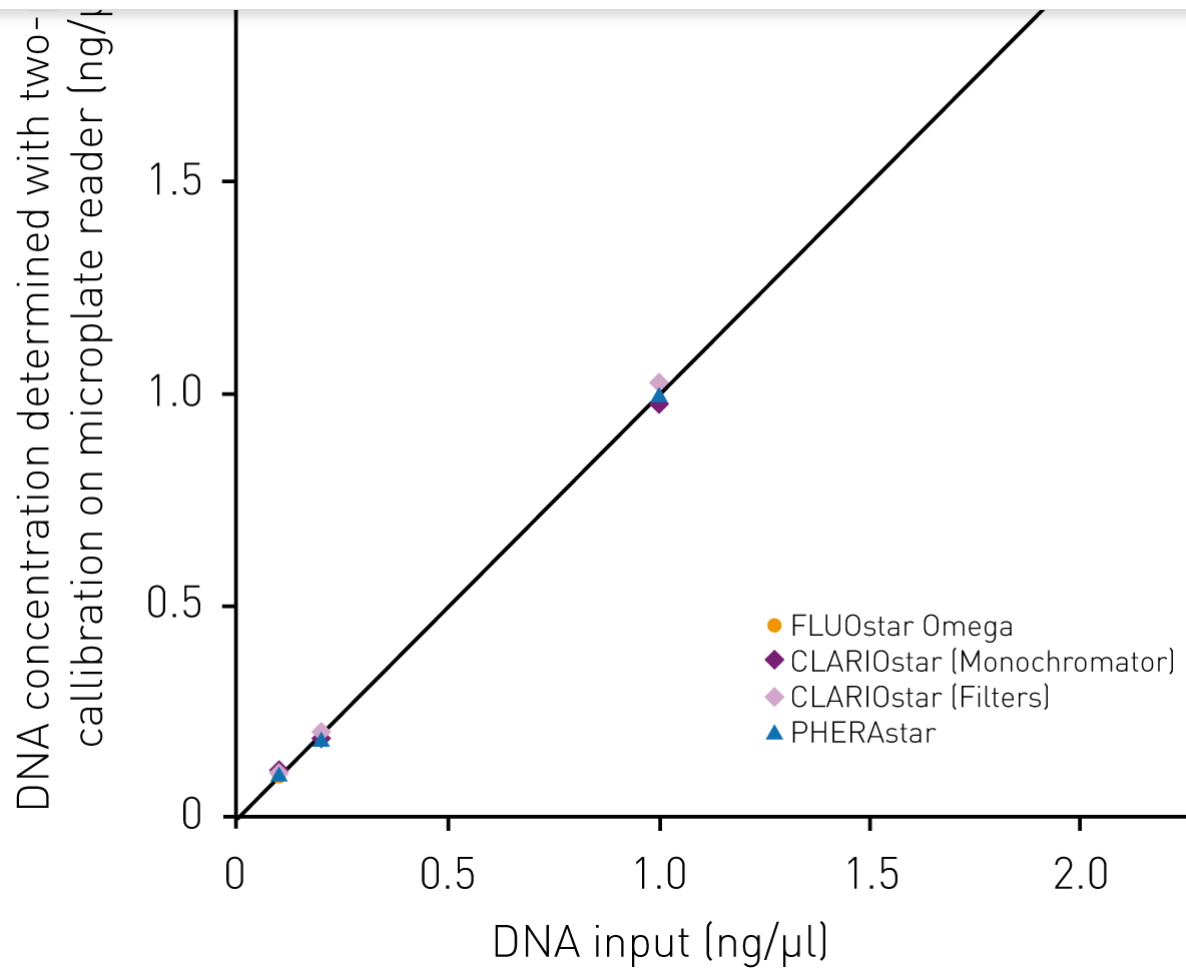


Fig. 3: DNA quantification using Qubit dye and BMG LABTECH microplate readers. Triplicates with defined DNA concentration were analyzed and DNA concentration was calculated using a two-point calibration line. Calibration curve was based on triplicates of negative control (no DNA) and 10 ng/μl DNA standard provided with the kit. Error bars indicate standard deviation.

Table 1: The table shows calculated DNA concentrations from two-point fit

DNA concentration determined on BMG LABTECH microplate readers			
Input DNA concentration	FLUOstar Omega	CLARIOstar	PERAstar
0.1 ng/μl	0.109	0.108	0.101
0.2 ng/μl	0.196	0.203	0.195
1 ng/μl	0.983	1.028	1.001
2 ng/μl	1.962	2.025	2.000

Conclusion



enables easy, repeatable measurements.



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