Optimum temperature and salinity conditions for growth of green algae *Chlorella ellipsoidea* and *Nannochloris oculata*

Sung Hwoan CHO, 1* Sung-Choon JI, 1 Sung Bum HUR, 2 Jeanhee BAE, 2 In-Seok PARK 1 AND Young-Chae SONG 3

¹Division of Marine Environment and Bioscience, College of Ocean Science and Technology, Korea Maritime University, Busan 606-791, ²Department of Aquaculture, Pukyong National University, Busan 608-737, and ³Division of Civil and Environmental System Engineering, College of Engineering, Korea Maritime University, Busan 606-791, Korea

ABSTRACT: The effects of temperature and salinity on growth of green algae Chlorella ellipsoidea and Nannochloris oculata were determined to compare the optimum culture conditions. A fourtemperature (15, 20, 25, and 30°C) × three-salinity (10, 20, and 30) factorial design with triplicates was applied. Specific growth rate (SGR), maximum density, and duration to reach maximum density of C. ellipsoidea were significantly affected by both temperature and salinity. The highest SGR was observed in C. ellipsoidea at 25°C and salinity 10, but the maximum density was very low. The highest maximum density was achieved in C. ellipsoidea at 15°C and 10. The slope constant of the linear relationship between semilogarithmic growth of C. ellipsoidea and day of culture was highest at 15°C and 10. The SGR and duration to reach maximum density of N. oculata were significantly affected by both temperature and salinity. However, maximum density of N. oculata was significantly affected by temperature, but not salinity. The highest maximum density was achieved in N. oculata at 25°C and 30, but SGR was significantly lower than that of N. oculata at 25°C and 10. The slope constant of the linear relationship between semilogarithmic growth of N. oculata and day of culture was highest at 25°C and 30. Based on these results, the condition of 15°C and salinity 10 seemed to be optimal for maximum density of C. ellipsoidea, and the condition of 25°C and 10 and 30 for SGR and maximum density for N. oculata, respectively.

KEY WORDS: Chlorella ellipsoidea, Nannochloris oculata, salinity, temperature.

INTRODUCTION

Application of green algae has been tried for marine fish hatcheries because the algae are commonly used as food for the first live foods (such as rotifers and *Artemia* nauplii) for early larval fish, or directly added to the rearing tanks for larval fish production to achieve the 'green water effect'. Supplementation of algae into the larval rearing tanks improved survival and/or growth of larval fish. ^{2–5}

In addition, the use of microalgae lead to a decrease in numbers of bacteria associated with the rotifer culture when compared with the rotifer culture fed yeast-based diets in the rotifer culture tanks.⁶ The high nutrient content in microalgae

*Corresponding author: Tel: 82-51-404-4755. Fax: 82-51-404-3988. Email: chosunh@hhu.ac.kr Received 20 November 2006. Accepted 27 April 2007. improved the lipid and fatty acid compositions of rotifers when either fed to rotifers, or supplied to the larval fish tanks, and eventually resulted in an improvement of fish production.^{7–11} Therefore, the constant production of high-quality microalgae is needed in marine fish hatcheries, and application of microalgae is highly recommended for larval fish production.

Growth of microalgae is likely to be affected by culture conditions such as temperature, salinity, illumination, and/or nutrients. 12-22 Green algae commonly used for marine fish hatcheries are *Chlorella ellipsoidea* and *Nannochloris oculata*, class Chlorophyceae, 23 because of the ease of mass culture and management, and high nutrient content leading to improvement in larval fish production. 5,9,24-28 Hur¹⁹ demonstrated that *N. oculata* was superior to *C. ellipsoidea* as food for rotifers in terms of rotifer growth rate and fatty acid

composition during summer. However, no study on the combined effect of temperature and salinity on growth of *C. ellipsoidea* and *N. oculata* have been reported.

In this study, therefore, the effects of temperature and salinity on growth of *C. ellipsoidea* and *N. oculata* were determined to compare the optimum culture conditions.

MATERIALS AND METHODS

Preparation of green algae and culture conditions

Chlorella ellipsoidea (KMCC C-20 clone) and Nannochloris oculata (KMCC C-31 clone) were purchased from the Korea Marine Microalgae Culture Center (KMCC, Busan, Korea). Algal cultures were performed in f/2 media²⁹ and intensity of illumination was maintained at 31 μmol photons/m² per second by a digital lux meter (INS DX-100, Japan) with a 24:0 h (light–dark) cycle at various temperature zones in a low temperature incubator (Dongwon Scientific System, Busan, Korea). Salinity was adjusted by mixing the filtered sea water and distilled water while monitoring with a salinometer (SM-2000, TS-EL, Japan).

Before inoculation with green algae, all flasks with 100-mL working volumes were sterilized by autoclave at 121°C for 20 min. *Chlorella ellipsoidea* and *N. oculata* were initially inoculated into thirty-six 250-mL flasks at a concentration of 100×10^4 cells/mL each. All flasks were handagitated twice a day (09:00 and 17:00 hours) and alternated within the same temperature zone every day to minimize the effects of differences in intensity of illumination on growth of algae.

Design of experiment

A four-temperature (15, 20, 25, 30° C) × three-salinity (10, 20, 30) factorial design with triplicates was prepared for this study.

Criteria measured for growth of green algae

Growth of green algae was measured by using a hematocytometer (0.0025 mm², Superior, Marienfeld, Germany). Samples of green algae from each flask were taken then dropped on the hematocytometer by means of a straw. Ten minutes were allowed for focusing before counting the green algae. Density of green algae at each condition was the mean of three counts observed under a micro-

scope (CH20, Olympus, Tokyo, Japan) and specific growth rate (SGR/day) was calculated by the formula of Guillard:³⁰ SGR/day = $3.322 \times \log_{10}(N_1/$ N_2)/ $t_1 - t_2$, where N_1 and N_2 are the cell concentrations at t_1 and t_2 , and t_1 and t_2 are final and initial days of the experiment, respectively. The optimum conditions for SGR, maximum density (×10⁴ cells/ mL) and duration to reach maximum density (d) were determined for each alga. Data on daily growth of algae were semilogarithmically transferred, and relationships between semilogarithmic growth of algae and days of culture were calculated. The cell sizes of C. ellipsoidea and N. oculata were measured as approximately 3 and 1 µm in diameter, respectively, using a microscope (Zeiss, Jena, Germany).

Statistical analysis

One-way and two-way analysis of variance (ANOVA) and Duncan's multiple range test³¹ were used to analyze the significance of the difference among the means of treatments. In addition, regression analysis for SGR and maximum density of each alga were conducted by using regression analysis with SAS v9.12 (SAS Institute, Cary, NC, USA).

RESULTS

Growth of Chlorella ellipsoidea

SGR, maximum density, and duration to reach maximum density of C. ellipsoidea at the various temperature and salinity conditions are given in Table 1. SGR, maximum density, and duration to reach maximum density of C. ellipsoidea were significantly (P < 0.05) affected by both temperature and salinity. The highest SGR (0.60) was observed in C. ellipsoidea at 25°C and salinity 10, but the maximum density was very low (Fig. 1). A similar trend was observed in C. ellipsoidea at 30°C at all salinity conditions tested. Maximum density and duration to reach maximum density of C. ellipsoidea at 15°C and 10 were 131.7 × 106 cells/mL and 18 days after inoculation, respectively. Relatively high SGR and maximum density was achieved in C. ellipsoidea at 25°C and 30. Significant (P < 0.0001) effects of temperature and salinity on SGR, maximum density, and duration to reach maximum density of C. ellipsoidea were observed.

Results of regression analysis for specific growth rate (SGR) and maximum density of *C. ellipsoidea* on salinity levels at constant temperature are given in Table 2. A linear regression was obtained

Table 1 Specific growth rate, maximum density and duration to reach maximum density of *Chlorella ellipsoidea* at various temperature and salinity conditions

Temperature			Maximum density	Duration to reach maximum
(°C)	Salinity	SGR	(10 ⁶ cells/mL)	density (d)
15	10	$0.37 \pm 0.013^{e,f}$	131.7 ± 12.36^{a}	$18 \pm 0.0^{\rm b}$
	20	$0.36 \pm 0.010^{ m e,f,g}$	$115.2 \pm 7.60^{\mathrm{a,b}}$	$18\pm0.0^{ m b}$
	30	$0.31 \pm 0.004^{\mathrm{g}}$	$097.3 \pm 1.78^{\mathrm{b,c}}$	$20\pm0.0^{\mathrm{a}}$
20	10	$0.40\pm0.002^{ m d,e,f}$	$112.5 \pm 6.91^{\mathrm{a,b}}$	$16\pm0.0^{ m b}$
	20	$0.37\pm0.007^{ m e,f}$	$113.3 \pm 11.94^{a,b}$	$17 \pm 0.0^{ m b,c}$
	30	$0.35\pm0.009^{ m f,g}$	$087.3 \pm 5.49^{\circ}$	$17 \pm 0.0^{ m b,c}$
25	10	$0.60\pm0.042^{\mathrm{a}}$	$010.2 \pm 0.99^{\mathrm{e}}$	$05\pm0.0^{ m g}$
	20	$0.41\pm0.017^{ m d,e}$	$041.2 \pm 15.80^{ m d}$	12 ± 2.1^{e}
	30	$0.45\pm0.006^{ m c,d}$	$103.1 \pm 27.11^{\mathrm{b,c}}$	$14 \pm 1.5^{ m d}$
30	10	$0.48\pm0.078^{ m b,c}$	$2.6 \pm 0.44^{\rm e}$	$3\pm0.6^{ m h}$
	20	$0.52 \pm 0.044^{\rm b}$	$012.9 \pm 1.80^{\rm e}$	$07\pm0.0^{\mathrm{f}}$
	30	$0.44\pm0.030^{ m c,d}$	050.8 ± 10.69^{d}	$12\pm0.0^{ m e}$
Two-way anova				
Temperature		P < 0.0001	P < 0.0001	P < 0.0001
Salinity		P < 0.0001	P < 0.0006	P < 0.0001
Interaction		P < 0.0001	P < 0.0001	P < 0.0001

Values (mean \pm standard deviation) with different superscript letters in the same column are significantly different (P < 0.05). SGR, specific growth rate.

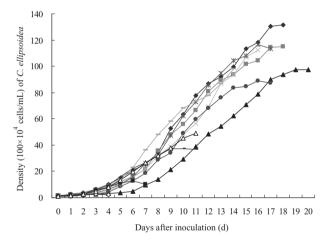


Fig. 1 Density of *Chlorella ellipsoidea* after inoculation at temperature and salinity conditions: 15°C and salinity $10 \, (\spadesuit)$, $20 \, (\blacksquare)$, and $30 \, (\blacktriangle)$; 20°C and salinity $10 \, (\times)$, $20 \, (★)$, and $30 \, (Φ)$; 25°C and salinity $10 \, (+)$, $20 \, (-)$, and $30 \, (-)$; 30°C and salinity $10 \, (\diamondsuit)$, $20 \, (\blacksquare)$, and $30 \, (△)$.

between SGR of *C. ellipsoidea* and salinity at 15°C and 20°C (P < 0.0003 and P < 0.0001, respectively), but quadratic regression was obtained at 25°C (P < 0.0009). Linear regression was also obtained between maximum density of *C. ellipsoidea* and salinity at 15, 20, 25, and 30°C (P < 0.003, P < 0.02, P < 0.0009, and P < 0.0001, respectively).

Relationships between semilogarithmic growth of *C. ellipsoidea* and day of culture at various temperature and salinity conditions are presented in

Table 3. The slope of the linear relationship was highest in *C. ellipsoidea* at 15°C and salinity 10, and lowest for this alga at 30°C and 10. However, a relatively lower constant was observed in *C. ellipsoidea* at 25°C and 10, at which the highest SGR was achieved.

Growth of Nannochloris oculata

SGR, maximum density, and duration to reach maximum density of N. oculata at various temperatures and salinities are presented in Table 4. SGR and duration to reach maximum density of N. oculata were significantly (P < 0.05) affected by both temperature and salinity. However, maximum density of *N. oculata* was significantly (P < 0.05) affected by temperature, but not salinity. The highest SGR (0.46) was observed in N. oculata at 25°C and salinity 10, and its maximum density was 227.0×10^6 cells/mL (Fig. 2). A relatively poor SGR was achieved in N. oculata at 15°C at all salinities tested. However, the highest maximum density, 308.0×10^6 cells/mL was achieved in N. oculata at 25°C and 30, but SGR was significantly (P < 0.05)lower than that of N. oculata at 25°C and 10. Further, duration to reach maximum density of N. oculata at 25°C and 30 was longest, 21 days after inoculation. Significant effect of temperature and salinity on maximum density and duration to reach maximum density of N. oculata were observed.

Table 2 Regression analysis for specific growth rate and maximum density of *Chlorella ellipsoidea* and *Nannochloris oculata* on salinity levels at constant temperature

Item	Temperature (°C)	Chlorella ellipsoidea	Nannochloris oculata
SGR	15	L (P < 0.0003)	Q (P < 0.005)
	20	L(P < 0.0001)	ns
	25	Q (P < 0.0009)	L(P < 0.03)
	30	ns	Q $(P < 0.0004)$
Maximum density	15	L (P < 0.003)	L(P < 0.02)
•	20	L (P < 0.02)	ns
	25	L(P < 0.0009)	L(P < 0.03)
	30	L (P < 0.0001)	L $(P < 0.02)$

L, linear regression; ns, no significant difference; Q, quadratic regression; SGR, specific growth rate.

Table 3 Relationships between semilogarithmic growth of *Chlorella ellipsoidea* and *Nannochloris oculata* and day of culture at various temperature and salinity conditions

Temperature (°C)	Salinity	Chlorella ellipsoidea	Nannochloris oculata
15	10	$Y = 8.30X - 17.63, R^2 = 0.97$	$Y = 8.34X - 21.60, R^2 = 0.95$
	20	$Y = 7.47X - 15.91, R^2 = 0.96$	$Y = 8.08X - 18.57$, $R^2 = 0.97$
	30	$Y = 5.73X - 18.01, R^2 = 0.94$	$Y = 7.04X - 24.94$, $R^2 = 0.92$
20	10	$Y = 7.54X - 18.91$, $R^2 = 0.93$	$Y = 10.10X - 18.54$, $R^2 = 0.97$
	20	$Y = 8.09X - 18.49$, $R^2 = 0.95$	$Y = 10.25X - 20.35, R^2 = 0.95$
	30	$Y = 6.32X - 14.12, R^2 = 0.94$	$Y = 9.50X - 18.97, R^2 = 0.97$
25	10	$Y = 1.88X + 0.67, R^2 = 0.90$	$Y = 15.31X - 41.64, R^2 = 0.95$
	20	$Y = 3.96X - 2.60$, $R^2 = 0.96$	$Y = 14.59X - 33.29$, $R^2 = 0.97$
	30	$Y = 7.59X - 12.94$, $R^2 = 0.96$	$Y = 17.19X - 49.98, R^2 = 0.96$
30	10	$Y = 0.34X + 1.08$, $R^2 = 0.66$	$Y = 12.10X - 29.04$, $R^2 = 0.96$
	20	$Y = 1.73X + 0.57, R^2 = 0.88$	$Y = 14.05X - 35.73, R^2 = 0.95$
	30	$Y = 4.74X - 5.89$, $R^2 = 0.96$	$Y = 15.15X - 32.93, R^2 = 0.97$

 R^2 , residual.

Results of regression analysis for SGR and maximum density of N. oculata on salinity levels at constant temperature are given in Table 2. Linear regression was obtained between SGR of N. oculata and salinity at 25°C (P < 0.03), but quadratic regression was obtained at 15°C and 30°C (P < 0.005 and P < 0.0004, respectively). Linear regression was obtained between the maximum density of N. oculata and salinity at 15, 25, and 30°C (P < 0.02, P < 0.03, and P < 0.02, respectively).

Relationships between semilogarithmic growth of *N. oculata* and day of culture at the various temperature and salinity conditions are presented in Table 3. The slope of the linear relationship was highest in *N. oculata* at 25°C and salinity 30, and lowest for this alga at 15°C and 30, respectively.

DISCUSSION

Green algae such as *C. ellipsoidea* and *N. oculata* have been widely used not only as a food for live

foods such as rotifers and *Artemia* nauplii for early larval fish, but also as a source for the green water effect in larval production of marine fish. Therefore, preparation of these algae in high quantity and quality is needed for successful seedling production of marine larval fish.

Many algae grew well over the broad ranges in both temperature and salinity, showing high tolerance to changes in these factors. 12 SGR, maximum density, and duration to reach maximum density of C. ellipsoidea and N. oculata were significantly affected by both temperature and salinity, except for maximum density of N. oculata in this study. Regardless of salinity, the maximum density was very low in C. ellipsoidea at 30°C in this study. This result probably indicates that high temperature is not optimum for growth of C. ellipsoidea, and agrees with Hur's 19 study. SGR and maximum density of C. ellipsoidea linearly decreased with an increase in salinity at 15°C and 20°C, but maximum density linearly increased with increase in salinity at 25°C and 30°C (Table 2). However, since

Table 4 Specific growth rate, maximum density, and duration to reach maximum density of *Nannochloris oculata* at various temperature and salinity conditions

Temperature			Maximum density	Duration to reach maximum density
(°C)	Salinity	SGR	(10^6 cells/mL)	(d)
15	10	$0.36 \pm 0.004^{ m e,f}$	154.0 ± 19.32^{d}	$19 \pm 0.6^{\rm b,c}$
	20	$0.36 \pm 0.014^{ m e,f}$	139.4 ± 4.30^{d}	$20\pm0.6^{\rm a,b,c}$
	30	$0.35 \pm 0.005^{\mathrm{f}}$	121.0 ± 4.67^{d}	$20\pm0.0^{\mathrm{a,b}}$
20	10	$0.44\pm0.014^{ m a,b}$	154.2 ± 5.72^{d}	$16\pm0.0^{ m d,e}$
	20	$0.44\pm0.012^{\mathrm{a,b,c}}$	150.7 ± 7.15^{d}	$16\pm0.0^{ m f}$
	30	$0.41 \pm 0.009^{\mathrm{b,c,d}}$	139.8 ± 2.67^{d}	$17\pm0.0^{ m e,f}$
25	10	0.46 ± 0.017^{a}	$227.0 \pm 45.03^{\mathrm{b,c}}$	$18\pm1.7^{ m f}$
	20	$0.43 \pm 0.003^{ m b,c}$	$238.1 \pm 38.11^{\text{b}}$	$19\pm0.6^{ m b,c}$
	30	$0.42 \pm 0.023^{\mathrm{b,c}}$	308.0 ± 11.51^{a}	21 ± 0.6^{a}
30	10	$0.41 \pm 0.009^{c,d}$	$192.7 \pm 11.31^{\circ}$	$19\pm0.6^{\mathrm{c,d}}$
	20	$0.37 \pm 0.012^{e,f}$	$208.0 \pm 16.17^{\mathrm{b,c}}$	$19\pm0.6^{\mathrm{b,c}}$
	30	$0.38 \pm 0.036^{ m d,e}$	$230.7 \pm 1947^{\text{b}}$	$18\pm0.0^{ m d,e}$
Two-way anova				
Temperature		P < 0.0001	P < 0.0001	P < 0.0001
Salinity		P < 0.0020	P < 0.0800	P < 0.0070
Interaction		P < 0.2000	P < 0.0010	P < 0.0040

Values (mean \pm standard deviation) with different superscript letters in the same column are significantly different (P < 0.05). SGR, specific growth rate.

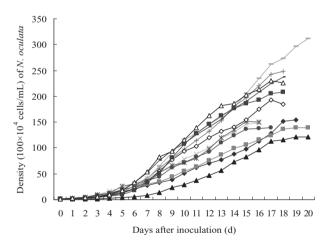


Fig. 2 Density of *Nannochloris oculata* after inoculation at temperature and salinity conditions: 15°C and salinity $10 \ (\clubsuit)$, $20 \ (\blacksquare)$, and $30 \ (\blacktriangle)$; 20°C and salinity $10 \ (×)$, and $30 \ (Φ)$; 25°C and salinity $10 \ (+)$, $20 \ (-)$, and $30 \ (Φ)$; 25°C and salinity $10 \ (+)$, $20 \ (-)$, and $30 \ (Φ)$.

maximum density was achieved in *C. ellipsoidea* at 15°C and salinity 10, which was the lowest temperature in this study, it is possible that the optimum temperature for this alga is lower than 15°C.

In addition, a relatively high maximum density of *C. ellipsoidea* at low temperature (15 and 20°C) rather than high temperature (25 and 30°C) at all salinities tested in this study, except for 25°C and salinity 30, indicated that lower temperature is

better to achieve the maximum density of *C. ellipsoidea*. Although the highest SGR was achieved in *C. ellipsoidea* at 25°C and 10, the relatively low slope between growth of this alga and days of culture, and maximum density probably indicate that the condition of 25°C and 10 is not optimum for growth of *C. ellipsoidea*.

SGR and duration to reach maximum density of *C. ellipsoidea* at 25°C and 30 were relatively high and short, respectively, compared to those of *C. ellipsoidea* at 15°C and 20°C at all salinities tested in this study. Hur³² also showed that *C. ellipsoidea* achieved satisfactory growth at 28°C, but there was exponential growth retardation above 30°C. However, the condition of 15°C and salinity 10, which achieved the highest slope between growth of *C. ellipsoidea* and days of culture, seemed to be the best for growth of *C. ellipsoidea* at all conditions tested in this study.

Unlike growth of *C. ellipsoidea*, *N. oculata* seemed to grow relatively well at all conditions tested in this study. SGR of *N. oculata* linearly decreased with the increase in salinity at 25°C (Table 2). The maximum density linearly decreased with increase in salinity at 15°C, but linearly increased at 25°C and 30°C. The highest SGR and maximum density were achieved for *N. oculata* at 25°C and salinities 10 and 30, respectively, indicating the optimal temperature was 25°C for growth of *N. oculata*. This result is supported by the relatively high slope of the linear relationship between growth of *N. oculata* and day of culture

at 25°C for all salinities tested in this study. Similarly, Terlizzi and Karlander³³ reported a positive correlation between temperature and growth of *N. oculata* from 20–25°C. Thus, the optimal salinity condition for growth of *N. oculata* in this study could vary depending on its purpose. For the purpose of achieving the maximum density of *N. oculata*, the optimal salinity condition was 30, and it was 10 for the purpose of fast growth (high SGR), at 25°C.

The relatively high maximum density of *N. oculata* at high temperature (25 and 30°C) rather than low temperature (15 and 20°C) at all salinities tested in this study indicate that high temperature is better for achieving the maximum density of *N. oculata*. Similarly, Hur¹⁹ mentions that *N. oculata* is more stable than marine *Chlorella* as food for rotifers above 30°C.

The results of two-way anova of growth of C. ellipsoidea and N. oculata on temperature and salinity showed significant effects of temperature, salinity, and a temperature–salinity interaction on the SGR, maximum density, and duration to reach maximum density (Tables 1 and 4). For both C. ellipsoidea and N. oculata, the main effect of temperature was significant at P < 0.01%, which was equal to or greater than that of salinity and two–factor interaction, and it probably indicates that temperature is more important than salinity in determining growth of both algae. A similar result was observed in red tide flagellates Heterocapsa circularisquama and Chattonella verruculosa. 34

The maximum density of *C. ellipsoidea* was lower than that of *N. oculata* at the same culture conditions in this study, probably resulting from the differences in cell size of the two algae. The cell size of *C. ellipsoidea* (3 μ m diameter) was three times larger than that of *N. oculata* (1 μ m diameter). Optimal culture conditions (temperature and salinity) for growth of two green algae were determined in 250-mL flasks (i.e. lab scale) in this study. Since large quantities of green algae are needed for marine fish hatcheries, further study is needed to determine optimal culture conditions for outdoor tanks or at a commercial scale.

Direct application of these algae as food for the first live foods, or to the rearing tanks for marine larval fish should be avoided due to high gap in salinity, although relatively high maximum density was achieved in *C. ellipsoidea* and *N. oculata* at low salinities (10 and 20) in this study. The chemical composition (fatty acid composition) of microalgae is also affected by environmental culture conditions such as temperature, salinity, light, and nutrients. ^{16,18,35} Therefore, further studies on nutrient composition of green algae at the various culture conditions are needed.

CONCLUSION

Low temperature (15 and 20°C) with low salinity (10 and 20) conditions were better for growth of *C. ellipsoidea*. On the contrary, high temperature (25 and 30°C) with high salinity (30) were better conditions for growth of *N. oculata*. Optimal temperature and salinity to achieve maximum density of *C. ellipsoidea* were 15°C and 10, respectively. However, for *N. oculata*, optimal temperature and salinity conditions were, respectively, 25°C and 10 for SGR, and 25°C and 30 for maximum density.

ACKNOWLEDGMENT

This work was supported by the Korea Research Foundation Grant funded by the Korean Government (MOEHRD, Basic Research Promotion Fund) (KRF-2006-005-J00501).

REFERENCES

- Reitan KI, Rainuzzo JR, Oie G, Olsen Y. A review on the nutritional effects of algae in marine fish larvae. *Aquaculture* 1997; 155: 207–221.
- Reitan KI, Rainuzzo JR, Oie G, Olsen Y. Nutritional effects of algal addition in first feeding of turbot (*Scophthalmus maximus* L.) larvae. *Aquaculture* 1993; 118: 257–275.
- 3. Tamaru CS, Murashinge R, Lee CS. The paradox of using background phytoplankton during the larval culture of striped mullet, *Mugil cephalus* L. *Aquaculture* 1994; **119**: 167–174.
- Oie G, Makridis P, Reitan KI, Olsen Y. Protein and carbon utilization of rotifers (*Brachionus plicatilis*) in first feeding of turbot larvae (*Scophthalmus maximus* L.). *Aquaculture* 1997; 153: 103–122.
- Cho SH, Hur SB, Jo J. Effect of enriched live feeds on survival and growth rates in larval Korean rockfish Sebastes schlegeli Hilgendorf. Aquacult. Res. 2001; 32: 199–208.
- Oie G, Reitan KI, Olsen Y. Comparison of rotifer culture quality with yeast-plus oil and algal based diet cultivation diets. *Aquacult. Internat.* 1994; 2: 225–238.
- Watanabe T, Kitajima C, Fujita S. Nutritional values of live food organisms used in Japan for mass propagation of fish: a review. *Aquaculture* 1983; 34: 115–143.
- Ben-Amotz A, Fishler R, Schneller A. Chemical composition of dietary species of marine unicellular algae and rotifers with emphasis on fatty acids. *Mar. Biol.* 1987; 95: 31–36.
- Hur SB, Kim H. Chlorella cultivation for mass culture of rotifer, Brachionus plicatilis. I. Selection of suitable Chlorella species. J. Aquacult. 1988; 1: 27–35.
- Whyte JNC, Nagata WD. Carbohydrate and fatty acid composition of the rotifer, *Brachionus plicatilis* fed monospecific diets of yeast or phytoplankton. *Aquaculture* 1990; 89: 263–272.
- Reitan KI, Rainuzzo JR, Olsen Y. Influence of lipid composition of live feed on growth, survival and pigmentation of turbot larvae. *Aquacult. Internat.* 1994; 2: 33–48.

- McLachlan J. Effect of salinity on growth and chlorophyll content in representative classes of unicellular marine algae. Can. J. Microbiol. 1961; 7: 399–406.
- Jitts HR, McAllister CD, Stephens K, Strickland JDH. The cell division of some marine phytoplankton as a function of light and temperature. Fish. Res. Can. 1963; 21: 139–157.
- Laing I, Utting SD. The influence of salinity on the production of two commercially important unicellular marine algae. Aquaculture 1980; 21: 79–86.
- Brank MR, Guillard RRL. The effects of continuous light and light intensity on the reproduction rates of twenty-two species of marine phytoplankton. *J. Exp. Mar. Biol. Ecol.* 1981; 50: 119–132.
- Richmond A. Microalgae of economic potential. In: Richmond A (ed.). *Handbook of Microalgal Mass Cultures*. CRC Press Inc, Boca Raton, FL. 1986; 199–243.
- Latala A, Hamoud N, Plinski M. Growth dynamics and morphology of plankton green algae from brackishwaters under the influence of salinity, temperature and light. *Acta Ichythyol. Piscator.* 1991; XXI (Suppl.): 101–116.
- Renaud SM, Zhou HC, Parry DL, Thinh LV, Woo KC. Effect of temperature on the growth, total lipid content and fatty acid composition of recently isolated tropical microalgae *Isochrysis* sp., *Nitzschia closterium*, *Nitzschia paleacea* and commercial species *Isochrysis* sp. (clone T.ISO). *J. Appl. Phycol.* 1995; 7: 595–602.
- Hur SB. Present status of larval-rearing technology in Korea. Hydrobiology 1997; 358: 21–26.
- 20. Lee Y. Commercial production of microalgae in the Asia-Pacific rim. *J. Appl. Phycol.* 1997; **9**: 403–411.
- Abu-Rezq TS, Al-Musallan L, Al-Shimmari J, Dias P. Optimum production conditions for different high-quality marine algae. *Hydrobiology* 1999; 403: 97–107.
- Sobrino C, Neale PJ, Lubian LM. Interaction of UV radiation and inorganic carbon supply in the inhibition of photosynthesis: spectral and temporal responses of two marine picoplankters. *Photochem. Photobiol.* 2005; 81: 384–393.
- 23. Chihara M, Murano M. *An Illustrated Guide to Marine Plankton in Japan*. Tokai University Press, Tokyo. 1997.
- 24. Lee Y, Chin P, Sim W. Effect of micronutritional-element deficiencies on the metabolism of *Chlorella* cells. (I) On the growth rate, respiration and photosynthesis. *Korean J. Microbiol.* 1967; **5**: 15–19.

- 25. Hirayama K, Funamoto H. Supplementary effect of several nutrients on nutritive deficiency of baker's yeast for population growth of the rotifer *Brachionus plicatilis*. *Nippon Suisan Gakkaishi* 1983; **49**: 505–510.
- James CM, Abu-Rezq TS. Effect of different cell density of Chlorella capsulata and a marine Chlorella sp. for feeding the rotifer Brachionus plicatilis. Aquaculture 1988; 69: 43–56.
- Ferreiro MJ, Fernandez-Reiriz MJ, Planas M, Labarta U, Garrido JL. Biochemical composition of enriched and starved rotifers. Larvi "91– Fish and Crustacean Larviculture Symposium. Eur. Aquacult. Soc. Spec. Publ. 1991; 15: 36–38.
- Korstad J, Neyts A, Danielsen T, Overrein I, Olsen Y. Use of swimming speed and egg ratio as predictors of the status of rotifer cultures in aquaculture. *Hydrobiology* 1995; 313/314: 395–398.
- Guillard R, Ryther J. Studies of marine planktonic diatoms.
 I. Cyclotella nana Hustedt and Detonula confervacea Cleve. Can. J. Microbiol. 1962; 8: 229–239.
- Guillard R. Division rates. In: Stein J (ed.). Handbook of Phycological Methods. Cambridge University Press, New York, NY. 1973; 289–311.
- 31. Duncan DB. Multiple range and multiple F tests. *Biometrics* 1955; **11**: 1–42.
- 32. Hur SB. The selection of optimum phytoplankton species for rotifer culture during cold and warm seasons and their nutritional value for marine finfish larval. In: Fulks W, Main K (eds). *Rotifer and Microalgae Culture Systems, Proceedings of a U.S.–Asia Workshop.* Hawaiian Oceanic Institute, Honolulu. HI. 1991: 163–173.
- Terlizzi DE, Karlander EP. Growth of a coccoid nanoplankter (Eustigmatophyceae) from the Chesapeake Bay as influenced by light, temperature, salinity, and nitrogen source in factorial combination. *J. Phycol.* 1980; 16: 364–368.
- 34. Yamaguchi M, Itakura S, Nagasaki K, Matsuyama Y, Uchida T, Imai I. Effects of temperature and salinity on the growth of the red tide flagellates *Heterocapsa circulcrisquama* (Dinophyceae) and *Chattonella verruculosa* (Raphidophyceae). *J. Plankton Res.* 1997; 19: 1167–1174.
- 35. Thompson PA, Guo M, Harrison PJ, Whyte JNC. Effects of variation in temperature. II. On the fatty acid composition of eight species of marine phytoplankton. *J. Phycol.* 1992; **28**: 488–497.