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Molecular containers assembled through the hydrophobic effect

Jacobs H. Jordan and Bruce C. Gibb*

This review focuses on molecular containers formed by assembly processes driven by the hydrophobic effect, and summarizes the progress made in the field over the last ten years. This small but growing facet of supramolecular chemistry discusses three classes of molecules used by researchers to investigate how self-assembly can be applied to form discrete, mono-dispersed, and structurally welldefined supramolecular entities. The approaches demonstrate the importance of preorganization of arrays of rigid moieties to define a specific form predisposed to bind, fold, or assemble. As the examples demonstrate, studying these systems and their properties is teaching us how to control supramolecular chemistry in water, shedding light on aspects of aqueous solutions chemistry, and illustrating novel applications that harness the unique properties of the hydrophobic effect.

1.0 Introduction

Why water?

Chemists are spoilt for choice when it comes to solvent; in contemporary chemistry there are literally dozens of different solvents to choose from, each with their own unique properties to fit a task. So why would a chemist focus on just one? And why would an organic chemist focus on one that doesn't dissolve most organic molecules? Why would an organic chemist choose water as a solvent? One reason that may be suggested is the environment. Green chemists might argue that we should aim for water as the default solvent; that organic chemists should be systematically converting - by whatever means possible - the cornucopia of organic reactions in the literature into water-based processes. Of course, whether or not this is possible, or even desirable, is debatable. And so for this and many other reasons organic chemists focus on organic solvents.

But there is no debating that supramolecular chemists should be very interested in water.2 With important exceptions, the expertise in controlling and utilizing non-covalent forces that the field has built up over the last few decades has been grounded in organic solvents. And yet aqueous chemistry is fundamental to the biosphere, hydrosphere, lithosphere and the atmosphere.3 Choose just one of these (the biosphere?), and one tiny fraction of it (Homo Sapiens?), and consider all of the key non-covalent interactions going on in aqueous solution.4 Opportunities abound.

But there are problems. Although pure water is relatively well understood, there are still some controversies arising from its dynamic structure. The idea of memory water and poly-water

may have been refuted,5 but the recent debate about the ability of water to store charge⁶ should remind us that its complicatedness - its complexity (in the sense of emergence) provides many opportunities to be led astray.

Moving away from the controversial, just the slightest complication to the pure water picture and our view becomes somewhat murky. Hence, a simple(!) air-water interface, or a change in pressure or temperature leads to debate and controversy. Dissolve something in water, be it a salt or a relatively hydrophobic, organic molecule and the water becomes murkier still. That is not to say that progress isn't being made. For example, the Jones and Ray effect (a minimum in the surface tension of electrolyte solutions at 1 mM concentrations) - a phenomenon that caused considerable debate last century - now seems to be resolved.8 More generally, it now seems clear that certain kinds of ions do accumulate at the air water interface, and that the ensuing models can now bridge the divide between physical chemists who point out that surface tension increases in solutions as a function of increasing salt concentrations can only be accounted for by a depletion of ions from the air-water interface, and atmospheric chemists who can only explain aerosol properties by having ions accumulate on droplet surfaces.9 Progress is also being made in regards to the solvation of hydrophobic surfaces, although the focus has primarily been on convex¹⁰ and flat surfaces¹¹ and is only more recently turning to concave surfaces. 12 All this noted, there is still plenty to discover about the hydrophobic effect (most simply put, why oil and water don't mix) and the Hofmeister effect (how salts effect the properties of water and aqueous solutions). Water science is a rich field barely scratched by supramolecular chemistry. But one thing is for sure, dissolve benzene and sodium chloride in water and there are six non-covalent interactions to consider

Department of Chemistry, Tulane University, New Orleans, LA 70118, USA. E-mail: bgibb@tulane.edu

(water-water, salt-salt, benzene-benzene, water-benzene, watersalt and benzene-salt). Rich pickings for supramolecular chemists.

Topic boundaries of the review

This review focuses on one small but growing area of supramolecular chemistry in aqueous solution. As the title suggests, the topic is molecular containers that are formed by assembly processes driven by the hydrophobic effect. But when is a container a container? What is an assembly? And which assemblies are, or are not, driven by the hydrophobic effect? We take the following perspectives.

The definition of a molecular container can be approached quantitatively or qualitatively, although both are subjective and arbitrary. Quantitatively, in our minds a molecular container envelops at least 83% of a smaller guest. This is based on our experiences at the macroscopic scale. Thus, in the extreme a container encloses 100% and there is no opportunity for the inner contents to escape without the opening of the container itself. Opening the container may be irreversible (an egg), but most containers can be opened and closed repeatedly because they have a screw-top or lid, and in our minds a molecular container is more akin to the plastic boxes devised by Earl Tupper (Tupperware[®]) than it is to an open cage-like structure. Our envisioned containers therefore possess the equivalent of at least five of six sides and envelop at least 83% (100% \times 5/6) of their contents. Qualitatively, we view a molecular container as fully preventing guest exchange unless the container is opened. In the case of a true molecular container bond breakage may be required to do this, but in the context of assembled containers then disassembly (removing the lid) suffices to allow guest entry or egression.

The extent by which a container envelops is not the only quality colored by our macro-scale view, so too is our definition of the type of construction material used to form the molecular walls. Thus, our definition includes structurally well-defined, mono-dispersed chemical entities with kinetically stable walls, but not those that are fluxional and/or poorly defined. We therefore do not include the topic of fluxional foldamers (but see below), even though we acknowledge that defining these as fluxional and/or poorly defined is colored by how they are analyzed.

In chemistry, self-assembly is the process in which a relatively disordered system of pre-existing molecules or ions forms a more organized structure as a result of specific, local interactions among the subunits. And so the systems under review here are principally molecules that form dimers, trimers, tetramers etc. and in doing so modify (put the lid on) or create the container. We have however intentionally chosen the more vague term "assembly" for the title rather than "self-assembly" so that structurally well-defined self-folding molecules can be included. In our minds there are too many similarities and overlaps between self-assembling and self-folding molecules to completely exclude the latter. Again, we acknowledge the vagaries in deciding what is and what is not a kinetically or thermodynamically stable entity.

Which assemblies are, or are not, driven by the hydrophobic effect? That depends on the definition of the hydrophobic

effect. Our view is that it is the phenomenon of the separation of hydrophobes from water and/or the desolvation of hydrophobic surfaces that is primarily driven by the highly cohesive nature of water. More specifically, it is an observed effect arising from the strong hydrogen bonding within water and, to a lesser extent, the van der Waals forces between solutes. The simple face-to-face cyclohexane dimer in water illustrates this idea perfectly. But it doesn't take much structural diversity to make things ambiguous. Is a similar face-to-face dimer of cyclohexanol driven by the hydrophobic effect? Or is the potential hydrogen bond between alcohol OH groups important? Our drawing of the line in this regard is based on the following: (1) the system in question must be in water. Systems with co-solvents were not included (mostly); (2) no systems involving covalent, or strong non-covalent interactions such as metal coordination, are discussed in detail. Reversible 13 and irreversible covalent bond14 formation and metal-ligand driven assembly are very important topic in self-assembly but are excluded here; (3) there must be both structural features in the solute and empirical evidence that desolvation of hydrophobic surfaces is key to assembly.

Finally, in this paper we have tried to summarize the progress made in the field over the last ten years. During this time-period we have attempted to be as comprehensive as possible. However, as is always the case, the topic of "molecular containers assembled through the hydrophobic effect" did not appear out of the ether. Consequently, where appropriate we have stretched back in the literature to before 2004.

2.0 Classes of self-assembling molecules

2.1 Deep-cavity cavitands

2.1.1 Investigating the properties of deep-cavity cavitands

(a) Synthesis, structure and coatings. The water-soluble deepcavity cavitands discussed here have their roots in analogous organic-soluble deep-cavity cavitands devised by the Gibb group. 16 Available in two linear steps from the respective resorcinarene, hosts such as 1 possess deeper pockets than those popularized by Cram and others, and acted as a proof-of-concept for the facile formation of relatively rigid (non-collapsing) binding pockets.

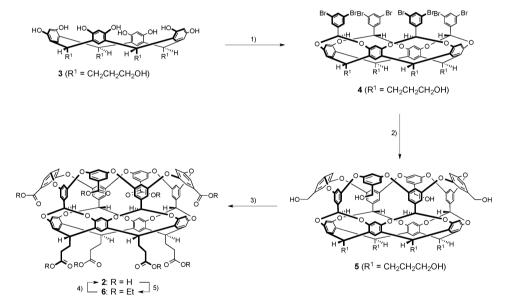
Building on this early work, the Gibb group reported the first example of a water-soluble deep-cavity cavitand (2) that binds and assembles into well-defined supramolecular entities via the hydrophobic effect; a host that is commonly called octa-acid (OA).17 The synthesis of OA 218 involves six steps from the requisite resorcinarene, only one of which requires chromatographic separation. Consequently, gram quantities of OA 2 are readily available. Briefly (Scheme 1), the resorcinarene scaffold was built up by first stereoselective bridging resorcinarene 3 with the corresponding benzal bromide (1-3-dibromo-5-(dibromomethyl)benzene) to afford the deep-cavity cavitand octabromide 4.16e,19 Subsequently, an eight-fold Ullmann ether reaction introduces rigidity to the cavity and adds functionality at the rim in the form of hydroxyl groups (5). Oxidation then gives crude OA 2.

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Esterification followed by chromatographic purification, then hydrolysis of the octa-ester 6 affords OA 2 in six linear steps.

Taking OA 2 as a representative of this class of molecule, these hosts possess a ca. 0.8 nm diameter wide and ca. 0.8 nm deep hydrophobic pocket comprised primarily of eight aromatic rings (Fig. 1). Atop the cavity, forming a rim around the portal, are four more aromatic rings orthogonal to the C4 axis of the host that 'stitch' the bowl together. The rim defined by these four rings is relatively hydrophobic and is key to the predisposition²⁰ of the molecules to assemble into supramolecular capsules. In addition, they provide diverging functionalization points for water solubilizing groups (R_1 , in this case four carboxylic acids), as well as four converging (endo) positions for groups that modulate the shape and properties of the hydrophobic pocket (R_2) . Four more functional groups for water-solubilization can be found at the termini of the pendant groups that hang down from the main bowl structure (R₃).

By virtue of its eight carboxylic acid/carboxylate groups OA 2 is soluble in tens of micromolar concentration at pH 7, and greater than one hundred millimolar concentrations at pH 9 or above. As is outlined in this section, this deep-cavity cavitand has been the most utilized and studied of this family of hosts.



Scheme 1 Synthesis of octa acid (OA) 2. Reagents are: (1) 3,5-dibrombenzal bromide, DBU, DMA; (2) 3,5-dihydroxybenzyl alcohol, K2CO3, CuO, Pyr; (3) KMnO₄, DMA, t-BuOH; (4) HCl, EtOH/CHCl₃; (5) LiOH, DMA, H₂O.

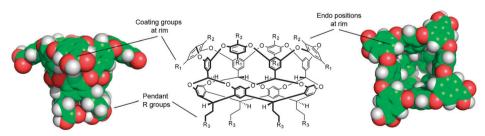
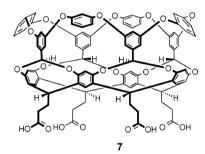


Fig. 1 Structure of octa-acid 2. The space-filling model on the left is from the same perspective as the chemical structure in the center. The space-filling model on the right looking down into the pocket of the host is from 'above' as viewed from the chemical structure.

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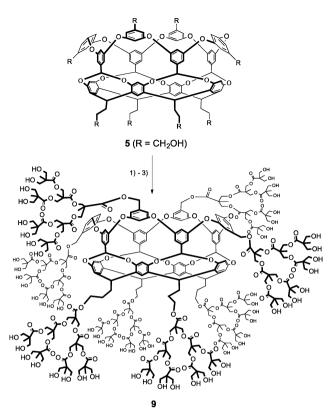
Key to this point is not only its good water solubility, but also its good behavior under a variety of conditions. Thus, whereas at above pH \approx 10 and concentrations less than 10 mM random aggregation of the host is rare, the presence of guests from as small as propane²¹ to as large as cholesterol¹⁷ trigger quantitative dimerization into monodispersed capsular complexes. We attribute the good behavior of OA 2 not only to the fact that it has eight water-solubilizing groups, but also to the fact that they are distributed in an pseudo square antiprismatic or anti-cubic array that spreads the negative charges evenly (but not uniformly) around its outer surface (eight, maximally distant points on the surface of a sphere define an anti-cube). The distortion to the anti-cube - that the upper portal face is larger than its opposing face, and the carboxylates on the rim are below the lip of the pocket - means that the charge distribution is not even; the upper portal face is a relatively hydrophobic surface and likely contributes to the predisposition of these hosts to assemble.

Before discussing the different derivatives of these watersoluble deep-cavity cavitands it is worth noting the fine balance between charge number/charge distribution and the water solubility of these hosts; a concept that applies to the design of water-soluble hosts and self-assembling subunits in general. Cavitands 7, possessing only four carboxylic acid groups, aggregates in aqueous solution and DMSO is required as a co-solvent to give interpretable NMR spectra.22 It is a similar story for host 8 (R = 3,5-benzene dicarboxylic acid), whose eight water-solubilizing groups are all on one end of the molecule. This host is too amphiphilic and also undergoes aggregation in aqueous solution.

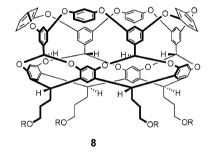


As well as these anionic hosts, neutral and cationic derivatives have also been synthesized. Examples of these are highlighted in the following sections. Here we briefly highlight some examples from these two classes for illustrative purposes.

The first example of a neutral water-soluble host was third generation (G3) dendrimer 9 (Scheme 2) from the Grayson group. ²³ The G3 host 9 was prepared by three successive rounds of coupling, and deprotection starting from the precursor compound octol 5. Thus, the eight hydroxy moieties were dendronized with an acetonide-protected anhydride and deprotected using a solid-phase acid resin to give the G1 cavitand with 16 hydroxyl groups. Two successive rounds of protectioncoupling-deprotection gave the G3 cavitand with 64 watersolubilizing hydroxyls on the termini of the dendritic chains. Three generations of dendrimer were necessary for full water solubility. Thus the corresponding G2 cavitand required at least 20% methanol solutions for dissolution. Although possessing



Scheme 2 Synthesis of dendronized cavitand 9. Conditions are: (1) acetonideprotected bis(hydroxymethyl)propanoic anhydride, DMAP; (2) Dowex acid resin; (3) repeat coupling and deprotection steps (1) and (2) twice.



the same type of properties of OA 2 (see below), guest affinities to the G3 cavitand 9 were generally lower than with OA 2. This was attributed to self-inclusion of the dendritic arms within the hydrophobic concavity.

In an alternative approach, Grayson introduced a range of surface modifications using Cu(1) catalyzed aza-alkyne cycloaddition ("click") chemistry. Thus, the octol 5 was treated with a carbodiimide in the presence of 4-pentynoic acid to give the activated core cavitand 10 with eight alkynes protruding from the exterior (Scheme 3). Subsequently, treatment of 10 with an excess of the requisite azide under "click" conditions led to an array of coated hosts (11) including two with dendrons, and several with polymer coats such as polycaprolactone (PCL), and polyethylene glycol (PEG). All of the functionalized cavitands were highly mono-disperse and were characterized by MALDI-TOF MS analysis. The PCL-grafted cavitand showed increased solubility in organic solvents, whilst the PEGylated cavitand

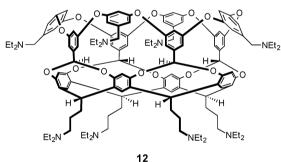
$$R = OH$$

$$R$$

Scheme 3 "Click" chemistry for diverse water-soluble cavitands. Conditions are: (1) 4-pentynoic acid, N,N'-dicyclohexylcarbodiimide, DMAP, pyr/ CH₂Cl₂; (2) various azides, Cu(ı)Br/Cu/N,N,N',N"-pentamethyldiethylenetriamine/CH₂Cl₂

showed increased solubility in polar solvents such as methanol $(>88 \text{ mg ml}^{-1})$, ethanol $(>24 \text{ mg ml}^{-1})$ and water $(>33 \text{ mg ml}^{-1})$.

The first example of water-soluble cavitands coated with cationic groups, host 12 and its octa-triethyl ammonium derivative, have been recently reported by the Ramamurthy group.²⁴ The initial studies on these hosts suggest that at low pH, 12 undergoes a degree of aggregation, but that it is better behaved as a complex with different guests. Preliminary host-guest and photochemical studies suggest properties similar to OA 2. On the other hand, preliminary work with the corresponding octa-triethyl ammonium derivative suggest a poorly behaved host at neutral pH. Interestingly, host 12 also allowed the first glimpse in the gas phase of the 1:1 and capsular complexes made by these types of hosts.²⁵



(b) Factors affecting complexation and assembly. Both the nature of the host and guest are important in the complexes formed by deep-cavity cavitands. To date the following hostguest complexes have been identified with deep-cavity cavitands: 1:1, 1:2, 2:1, 2:2, 4:2 and 6:3. We will focus first on guest effects and the formation of 1:1 and 2:1 complexes to highlight why one or the other forms.

Guest effects. The Gibb group first identified the key point about forming 1:1 complexes with OA 2 in a study of the binding of a range of cyclic and acyclic aliphatic carboxylates.²⁶ ¹H NMR showed that these amphiphilic guests bound with their hydrophobic chains in the pocket of host 2, and their polar carboxylates exposed to the free aqueous medium. Consequently, by binding in the orientation specific manner the 'upper' surface of the host defined by the portal to the pocket and its attendant rim is rendered more hydrophilic and

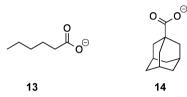


Fig. 2 Representative carboxylate guests for 1:1 complexes with OA 2.

assembly to the capsular state is switched off. A later study with related molecules confirmed these findings.²⁷

In the study of cyclic and acyclic aliphatic carboxylates,26 Isothermal Titration Calorimetry (ITC) were utilized to examine three cyclic and three acyclic carboxylates typified by hexanoate 13 and adamantane carboxylate 14 (Fig. 2). All of the examined carboxylates were found to associate with a K_a of between 10³ and $10^6 \,\mathrm{M}^{-1}$, with the weakest being hexanoate $(K_a = 3.64 \times 10^3)$ and the strongest proving to be adamantane carboxylate $(K_a = 1.14 \times 10^6)$. Butanoate, with a short propyl chain, did not bind. Apparently, the propyl chain was unable to effectively compete with the hydrophilicity of the carboxylate. A linear trend was observed for the relationship between ΔG° for complexation and the number of carbon atoms in the guest (N_c) . ΔG° values become more favorable with increasing number of N_c , with the gradients $d\Delta G^{\circ}/dN_c$ for the acyclic acids and cyclic acids respectively being similar to those observed for the transfer of hydrocarbons from water to organic solvents.

Examining how the enthalpy and entropy changes as a function of N_c revealed two trends. First, for the acyclic guests as N_c increased both the entropy and enthalpy terms for complexation became more favorable. In contrast, enthalpyentropy compensation was observed with the cyclic guests; with increasing N_c the entropy component of binding became less favorable, while the enthalpy component became more favorable. This was interpreted to arise from increasing hostguest contacts with larger guests being countered by restricted motion within the cavity.

For both series, the thermodynamics of complexation as a function of temperature displayed near perfect enthalpyentropy compensation so that ΔG° was nearly independent of T. In all cases the exothermicity of complexation increase

with T confirming that the heat capacity change $(\Delta C_{\rm p}^{\circ})$ upon complexation was negative. The obtained values were generally equal to and/or greater than those seen with cyclodextrins, perhaps reflective of the more hydrophobic pocket of the deepcavity cavitand. Expanding on this work, the same authors recently examined the binding of a series of cyclic aliphatic and aromatic carboxylates as part of the fourth statistical assessment of modeling of proteins and ligands (sampl.eyesopen.com).²⁸

Another illustrative example of the formation of 1:1 complexes is the binding of 'hydrophobic' anions to the cavity of OA 2.29 This work examined the effects of different salts on the binding of adamantane carboxylate 14. There was only a slight change in the free energy of guest binding (~ 0.5 kcal mol⁻¹) between the absence and presence of salting-out salts such as sodium fluoride. In these cases complexation was strongly endothermic and only slightly entropically promoted. In contrast to this were the series of salting-in sodium salts (chlorate, iodide, thiocyanate and perchlorate) which demonstrated a trend towards a decreasing free energy of complexation. Underlying this free energy change was a dramatic decline in the contribution to the free energy by enthalpy and a concomitant increase in the contribution made by entropy; so much so that in the presence of sodium perchlorate, guest binding was entirely entropy driven and the binding of 14 was actually endothermic.

¹H NMR showed that fundamental to the reported changes in thermodynamics with salting-in salts was anion binding to the pocket of the host competing with the complexation of 14. Subsequent NMR titrations of salt to OA 2 without 14 revealed association constants ranging from <1 M⁻¹ in the case of sodium nitrate, to 95 M⁻¹ in the case of sodium perchlorate. This first reported instance of inorganic anion binding to a hydrophobic pocket again demonstrates that like 13 or 14, very small charged guests lead to 1:1 complexes.

It is understood that anionic amphiphiles and simple anions form 1:1 complexes with OA 2 because the upper portal region of the complex remains quite hydrophilic. If on the other hand the binding of a guest renders the portal region of OA 2 hydrophobic, then there is a thermodynamic preference for a second host molecule to cap the complex and form an encapsulation complex. In fact, the first complexes reported with OA 2 were 2:1 host-guest complexes with steroids. 17 In this report a number of steroid guests were encapsulated including estradiol 15, progesterone 16 and cholesterol 17 (Fig. 3). In each case, because of the chirality of the guest the 2:1 host-guest complexes could be identified from symmetry arguments of the ¹H NMR data alone. In addition though, 2D NOESY NMR confirmed the spatial proximity of the host-host interactions

Fig. 3 Example of steroids that from 2:1 host-guest complexes with OA 2.

and the host-guest interactions from the two sets of nonequivalent hydrogens in each hemisphere of the capsule. Guests such as 15 and 16 formed very stable and structurally defined complexes, with a K_a for the former (empty capsule to full capsule) estimated to be $>1 \times 10^8 \text{ M}^{-1}$. Strong as these complexes are however, they were 'denatured' when methanol was added to the solution. It is noteworthy that the steroids such as cholesterol 17, although greatly solubilized by the host, did not form kinetically stable complexes (on the NMR time scale). In such cases, the length of guest is too large for the capsule and as a result the two hemispheres of the capsule cannot clamp down on one another and must remain partially solvated.

The stability of 2:1 host-guest capsular complexes involving OA 2 to co-solvent induced denaturation has been examined using ¹H NMR. ³⁰ In this study, a series of capsular complexes of trioxolane guests, e.g. 18-20 (Fig. 4) were investigated. The trioxolanes varied from practically insoluble to sparingly soluble in aqueous media, yet nevertheless were fully dissolved in a 2:1 solution of OA 2 in aqueous buffer. ¹H NMR demonstrated that all of these trioxolanes were readily taken up as a kinetically and thermodynamically stable 2:1 complex. The denaturation experiments of the different complexes utilized a series of per-deuterated water miscible solvents, namely: acetone-d₆, dimethylsulfoxide-d₆ (DMSO-d₆) methanol-d₄, ethanol-d₆, isopropanol-d₈, acetonitrile-d₃, and tetrahydrofuran-d₈ (THF-d₈); as well as trifluoroethanol (TFE). Overall, 18 formed the weakest of the complexes, and a 1:1 ratio of the host and guest did not form any 1:1 or 2:1 complex in 50% aqueous acetone. On the other hand, under identical conditions guest 19 formed 78% 1:1 and no 2:1 complex. Because of solubility issues with 20, only a 2:1 host-guest ratio could be examined in 50% aqueous acetone, in which case the mixture formed essentially only the 2:1 complex.

The different cyclic alkyl groups of 19 proved illuminating in so much as it was possible to observe the denaturation of the 2:1 capsular complex to form the 1:1 complex in which the adamantyl group is bound, and ultimately the breakdown of this complex with even higher equivalents of added co-solvent. Overall, for the different solvents examined, the ability to denature the complex formed by 19 was: acetonitrile- $d_3 > THF-d_8 > TFE >$ i-PrOD- d_8 > acetone- d_6 > ethanol- d_6 > methanol- $d_4 \approx DMSO-d_6$. Thus it was observed that whereas acetonitrile denatures most of the capsular complex at 30% by volume, a 70% aqueous solution of DMSO was needed to bring about the same extent of decomplexation.

In related work it has been shown that a fully fitting and rigid guest is not a prerequisite for capsule formation, but that even small straight-chain alkanes can lead to assembly.³¹ In this work, the authors investigated the binding of the alkanes

Fig. 4 Examples of encapsulated trioxolanes.

n-pentane through *n*-octadecane (C_{18}). Although, the latter guest was reportedly (vide infra) too large to form a kinetically stable complex, the C5-C17 guests did form stable complexes with assembly/disassembly rates slower than the NMR timescale. As was noted for the aforementioned steroid and trioxolane complexes, for the free host the diffusion constant (D) as determined by pulse-gradient stimulated spin-echo (PGSE) NMR experiment was $1.82 \times 10^{-6} \, \mathrm{cm}^2 \, \mathrm{s}^{-1}$, considerably larger than that for the capsular complexes ($D = 1.36 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$).

More specifically, the guests n-pentane through n-heptane were found to form stable 2:2 complexes with OA 2. These were the first examples of multiple molecules binding within the capsule formed by the host. Apparently there is too much empty space in the capsule when only one molecule is encapsulated. For the longer alkanes examined, the complexes formed were 2:1 in stoichiometry; the one exception being intermediately sized n-octane which formed both 2:1 and 2:2 capsules in a temperature dependent ratio (the quaternary complex dominating at higher temperature).

The propensity for deep-cavity cavitands to tend towards capsule formation was further investigated by comparing a series of guests of roughly equal volume but differing polarity/water solubility (Fig. 5).32 In these instances, it was shown that as the number of oxygen atoms increased in the series the stability of the corresponding capsular complexes decreased.

Competition experiments between the varying guests indicated that both tridecane (C13) 21 and dodecanol 22 bound strongly and favored 2:1 capsule formation, with the former possessing a binding affinity an order of magnitude greater than the latter. This was attributed to both the hydrophilicity of the polar hydroxyl group of 22 and its ability to hydrogen bond and be freely solvated by waters, and the loss of $C-H\cdots\pi$ interactions between the host and guest upon replacement of the terminal methyl of 21. Both tridecane 21 and dodecanol 22 adopt an extended, pole-to-pole motif within the capsule (for guest packing within the capsule formed by OA 2 see Section 2.1.1.e), and for the methylene groups at C-1 and C-11 of 22 the $\Delta\delta$ values indicated that the hydroxyl moiety was bound more deeply in one half of the capsule that the methyl group was in the other.

While the other guests 23 and 24 had clearly defined NMR signals indicative of 2:1 complexation, 2-[2-(2-propoxyethoxy)ethoxy] ethanol (tri(ethylene glycol)propyl ether) 25 exhibited broadened signals which were confirmed by PGSE NMR to be due to a capsule of low stability. Furthermore, bis[2-(2-hydroxyethoxy)ethyl] ether tetra(ethylene glycol) 26 did not show any signals indicative of capsule formation and indeed was confirmed

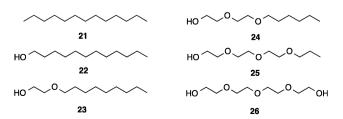
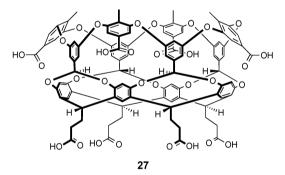


Fig. 5 Approximately isosteric guests of varying polarity.

by PGSE NMR to form only a 1:1 complex. An interesting sidenote regarding 26 is that opinion is split about whether it is hydrophilic or hydrophobic. At least by the metric of binding to OA 2 the answer is clear; it is relatively hydrophobic.

To recap, the question of whether a 1:1 or a capsular complex is formed with cavitands such as OA 2 rests in large part on the nature of the guest. If the guest is highly water soluble, or at least can adopt a binding orientation that projects a polar group to the portal of the host, then only a 1:1 complex will be formed. If on the other hand the guest binds and the net polarity of the portal/rim region of the host-guest complex is hydrophobic, then a capsular complex is preferentially formed; and the more hydrophobic the guest the stronger the capsular complex that is formed.

Host effects. In addition to the nature of the guest, the structure of the host also plays a role in the assemblies seen. This was first demonstrated with the deep-cavity cavitand TEMOA (tetra-endo-methyl octa-acid, 27). In an analogous procedure to that utilized in the synthesis of OA 2, TEMOA 27 was synthesized by the eight-fold Ullmann reaction of octabromide 4 (Scheme 1) with 3,5-dihydroxy-4-methylbenzyl alcohol. This afforded four methyl substituents in place of the corresponding protons at the hydrophobic rim, which after oxidation, esterification and purification, and hydrolysis led to host 27.33



The four endo methyl groups of 27 provide for a slightly deeper cavity with a narrower portal region. Furthermore, the methyls crenellate the rim of the cavity and consequently disrupt 2:1 capsule formation. As a result, its properties are quite different from OA 2. For example, as a function of guest size the assembly of OA 2 is straightforward: with small guests such as methane and ethane it forms 1:1 complexes (vide infra), but with bigger guests a 2:2 and then a 2:1 capsule is formed (vide supra). In contrast, diffusion NMR experiments revealed that TEMOA 27 forms 1:1 assemblies for methane, ethane, n-heptane and n-octane, a mixture of 1:1 and 2:2 complexes with propane, n-butane, and n-hexane, and only capsular assemblies for n-pentane (2:2) and the longer chain alkanes C_9 to C_{14} (2:1). Why? It would appear that because the methyl groups at the rim of 27 reduce its predisposition to dimerize, 1:1 complexes are preferentially formed even with some hydrophobic guests. This occurs with methane and ethane because they are too small and the 2:2 complexes would have too much void space, and with *n*-heptane (C_7) and *n*-octane (C_8) because in both cases two guests cannot fit in the capsule and

the 2:1 complexes again would have too much empty space. In contrast, n-pentane can form a relatively stable 2:2 complex, and the longer chain alkanes C9 to C14 form relatively stable 2:1 capsular complexes. The guests propane, *n*-butane, and *n*-hexane are intermediate between the 'islands of stability' defined by *n*-pentane, and the alkanes C_9 to C_{14} .

The unusual assembly properties of TEMOA 27 are reflected in what happens with mixtures of it and OA 2.³⁴ In this work NMR was used to probe the extent of hetero-capsule formation when these two hosts are combined, i.e., whether a guest or guests were complexed inside 2_2 , 27_2 , or 2.27. A range of *n*-alkanes $(C_5 \text{ to } C_{16})$ was used in this study.

A combination of ¹H and NOESY NMR confirmed the possibility of hetero-capsule formation, and that the ratio of the homo- and hetero-capsular complexes varied depending on the nature of the included guest. Thus 24% of the hetero-complex

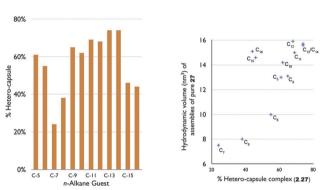


Fig. 6 Left panel: extent of hetero-capsule formation for hosts 2 and 27 in the presence of n-alkane guests. Right panel: correlation between the extent of hetero-capsule (2.27) formation and the hydrodynamic volumes of the corresponding complexes formed by 27 alone; guest size for each point are indicated

involving 2 and 27 was formed in the presence of n-heptane (C_7) , whereas 74% was formed in the presence of *n*-tridecane (C_{13}) . Overall, the extent of hetero-capsule formation was high for guests n-pentane, n-hexane and C_9 - C_{14} , and small for C_7 , C_8 , C_{15} , and C_{16} (Fig. 6). It was noted that there was a reasonable correlation between the extent of this hetero-capsule formation and the measured hydrodynamic volumes of the homo-capsular complexes formed by TEMOA 27 alone in the presence of the same guests (Fig. 6). These results were interpreted as follows. First, the guests n-heptane (C_7) and n-octane (C_8) formed limited amounts of hetero-capsular complex because they are poor templates of both homo- and hetero-capsules. As a result, it is energetically preferential for TEMOA 27 to form 1:1 complexes and allow OA 2 to from stable homo-capsular complexes. In other words, these guests promote self-sorting of OA 2. For the guests C9 through C14, the higher amounts of hetero-capsular complex formed was attributed to the more efficient packing of the (singular) guest within the capsular complexes compensating for the limited ability of 27 to assemble. The reason why the largest guests examined (C15 and C16) formed below statistical amounts of hetero-complex was not discussed.

The point that the methyl groups of deep-cavity cavitand TEMOA 27 reduce its predisposition to form a dimer capsule suggests that other assembly pathways may be more favorable to this host. Relatively recently this has indeed been shown to be the case. 35 Thus, whereas OA 2 has only been observed to assemble into dimers, TEMOA 27 can form dimeric, tetrameric and even hexameric capsules depending on the size of the guest that is encapsulated (Fig. 7a).

The different assembly states of TEMOA 27 were identified by investigating a series of homologous *n*-alkanes, from *n*-tetradecane (C_{14}) to *n*-hexacosane (C_{26}) . While the C_{14} guest bound in the typical 2:1 fashion previously observed with OA 2, (1 H and PGSE NMR), longer alkane chains (C_{17} – C_{20}) formed

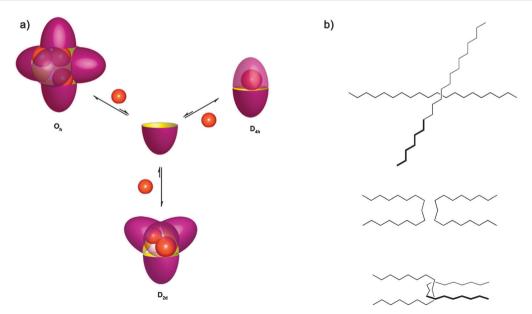
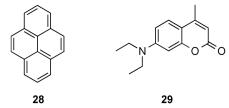


Fig. 7 (a) Different assemblies observed with TEMOA 27. (b) Different potential packing motifs within the tetramer of TEMOA 27.

pseudo $T_{\rm d}$ ($D_{\rm 2d}$) 4:2 host-guest complexes. Models suggest that the inner volume of this tetrameric host shell to be 1400–1500 Å³, i.e., this assembly is more voluminous than the familiar resorcinarene hexamer.36 In contrast, the intermediate length guests, the C₁₅ and C₁₆ alkanes, showed broad and/or poorly resolved NMR spectra suggesting that mixtures of assembly products were formed. Furthermore, although this was also the case with the C_{21} to C_{23} guests, with *n*-tetracosane (C_{24}) only one highly symmetric assembly product was observed in a 2:1 hostguest ratio. PGSE diffusion NMR examination of the host-guest signals indicated a diffusion coefficient of 9.6×10^{-7} cm² s⁻¹, which after correction for the higher working concentrations revealed a hydrodynamic volume of 48.9 nm³. In short, C₂₄ formed a nonary 6:3 host-guest complex with overall Oh symmetry and an estimated internal volume of between 3200 and 3700 Å³. The same complex was also observed for the longer guests n-pentacosane (C_{25}) and n-hexacosane (C_{26}), but unfortunately the kinetics of complex formation with even longer alkanes were too slow to investigate the formation of still larger assemblies. The precise packing arrangement of the two or three guests in the pseudo T_d and O_h capsules could not be surmised from the given data, but from the highfield positioning of the shielded guest methyl signal and the large anisotropy of the guest signals it was assumed that, to take the pseudo $T_{\rm d}$ capsule as an example, the guests adopt one of three different arrangements in which each end of a guest occupies one cavity of a host: an X-motif, a double U-motif, or a 'catenated' double U-motif (Fig. 7b).

(c) Environment inside the OA capsule. We have noted that OA 2 and TEMOA 27 are predisposed to form respectively dimeric, and dimeric, tetrameric and hexameric assemblies depending on the guests. In addition, the aforementioned studies reveal that it is the hydrophobic effect that drives the formation of these supramolecular entities. If this is indeed the case, then desolvation of the pocket and rim of the host, and of course desolvation of the guest molecules, must both be key. This leads to the question, are there any water molecules within the cavity of these capsular complexes? It is noteworthy that molecular dynamics simulations have indicated that water molecules are included within the hydrophobic concavity of free OA 2.37 These calculations suggest that on average there are 4.34 \pm 0.04 water molecules inside the hydrophobic pocket of the free host. Nature simply abhors a vacuum. Furthermore, these simulations reveal that during a wetting event the number of water molecules fluctuate, with four being the most probable (or lowest free energy) and the maximum number being seven. Overall, ΔG_{hvd} of the cavity was estimated to be -5 kcal mol⁻¹, with $\Delta H_{\text{hyd}} \approx$ $-20 \text{ kcal mol}^{-1} \text{ and } T\Delta S_{\text{hyd}} \approx 15 \text{ kcal mol}^{-1}$. This study also determined that these hydrophobic pocket waters are stabilized by their interaction with the bulk. Thus, when a hydrophobe approaches the portal of the pocket the cavity is rapidly and completely dewetted. Is there complete dewetting when a capsule is formed? The use of fluorescent probes as guests has addressed this question. For example, the ratio of the I₁ and I₃ vibronic bands in the fluorescence spectrum of pyrene 28 (Fig. 8)



Molecular probes to examine the inner environment of the OA 2 dimer.

confirm that no waters are present in the complex of 28 with OA 2.38 Furthermore, a more comprehensive study suggests that at least for sizeable guests that fill most of the cavity - this is the case regardless of the nature of the guest encapsulated.³⁹ To take one example from a series of guests investigated, coumarin-1 29, whose fluorescent quantum yield and lifetime increase with a decrease in solvent polarity, also demonstrated a low polarity environment inside the capsule. Overall these results suggest that, as would be expected considering the nature of the host, the environment inside the OA 2 dimer is benzene-like. To the authors' knowledge, a study of the environment within the TEMOA tetramer or hexamer has not been undertaken.

(d) Kinetics of capsule opening and closing. The very earliest studies with these container molecules demonstrated that for most hydrophobic guests the exchange between the free and the encapsulated state is slow on the NMR timescale; the lifetimes of many complexes are > 0.1 s. This point noted, there remains the idea that very small (atomic or diatomic) guests such as molecular oxygen might be able to enter an occupied capsule via partial opening (breathing) of the equatorial seam of the container. The first detailed analysis focusing on this possibility came from measurements of the rate constants for the quenching of excited singlet and triplet guests by oxygen.40 These guests included (Fig. 9) T₁ excited dimethyl benzil 30 (phosphorescence) and S₁ excited phenanthrene 31 (fluorescence), and covered excited-state lifetimes from 0.05 to 922 µs. Two classes of guests were observed, those that were quenched by oxygen when enclosed within the capsule, and those that were not. The latter were a mixture of singlet and triplet guests and therefore the spin state of the guest was dismissed as an explanation for why these guests were not quenched. Rather the obtained results reflected the guest-dependent accessibility of oxygen to the bound guest; if the guest had an excited lifetime of less than 5 μs it was inaccessible to oxygen.

To investigate the formation/dissociation dynamics of the encapsulation complexes formed by OA 2, the Bohne group

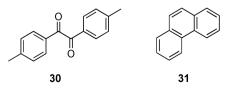


Fig. 9 Molecular probes to examine the opening and closing of the OA 2

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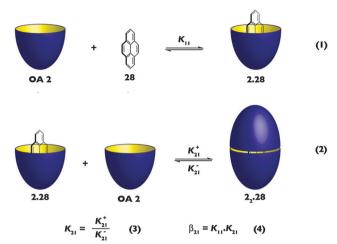


Fig. 10 Assembly states and kinetic parameters for the complex formed between OA 2 and pyrene 28.

measured the real-time kinetics of formation of the 1:1 and 2:1 host-guest complexes formed between OA 2 and pyrene 28 (Fig. 10).41 Pyrene 28 formed the expected 2:1 host-guest complex in aqueous solution, and upon encapsulation there was a marked red shift (5-6 nm) in its excitation and absorption spectra. This could not be attributed to dimerization of the guest as no excimer emission or spectral broadening was observed. Further evidence for the encapsulation of a single molecule of 28 within the OA 2 dimer was that its fluorescence lifetime was nearly three times longer than free 28 in aqueous borate buffer $(361 \pm 1 \text{ ns } vs. 130 \pm 1 \text{ ns})$, whilst the quenching rate by iodide anion was roughly four orders of magnitude slower (5.9 \pm 0.7 \times $10^5~{\rm M}^{-1}~{\rm s}^{-1}$ vs. $1.0~\pm~0.2~\times~10^9~{\rm M}^{-1}~{\rm s}^{-1}$). As the iodide quenching rate constant was similar to that observed for oxygen quenching of the triplet excited 28 within the OA 2 dimer, ³⁸ this suggested a similar mode of action, namely, a partial opening of the capsular complex allowing small guest entry and egress.

Measurements of the fluorescent intensity change of the complex as a function of added OA 2 were carried out and the observed trend fitted the expected 2:1 binding isotherm with an overall β_{21} of 3.2 \times 10¹² M⁻². Subsequent stopped-flow experiments successfully probed eqn (1) (Fig. 10), and identified

a fast reaction for the formation of the 1:1 complex with a K_{11} value of $K_1 = (4.5 \pm 0.6) \times 10^5 \text{ M}^{-1}$. Hence a K_{21} value of $K_{21} = (7 \pm 1) \times 10^6 \text{ M}^{-1}$ could be calculated (eqn (4)). The K_{11} and K_{21} values represented the first thermodynamic view of the difference between forming a 1:1 complex (desolvating half the guest and the pocket) and capping this complex (desolvating half the guest a pocket, and the two rims of the subunits). This clamping down of the two hemispheres in the second step leads to an association constant K_{21} one order of magnitude larger than the formation of the 1:1 complex, and is why K_{11} of the transient 1:1 complex could only be determined from kinetic experiments.

The lifetime of the capsular complex encapsulating 28 was also determined in this study. Thus the K_{21}^{+} and K_{21}^{-} values $((2.6 \pm 0.2) \times 10^6 \,\mathrm{M}^{-1} \,\mathrm{s}^{-1} \,\mathrm{and}\, 0.37 \pm 0.06 \,\mathrm{s}^{-1} \,\mathrm{respectively})$ were obtained from global fitting of the slow change in fluorescence intensity attributed to the second step of the assembly (eqn (2)), and how this was modulated as a function of OA 2. This gave a lifetime for the 22.28 complex of 2.7 s. While the aforementioned entrance/egress of small molecules occurred on the microsecond timescale, these results suggest that for the complex with 28 the 'breathing' of the complex was on the order of 105 times faster than the release of 28.

(e) Movement of internalized guests and guest packing motifs. The shape of guests such as steroids forces them to bind along the long Y-axis of the capsule (Fig. 11).¹⁷ Whilst their rigidity precludes their rotation around the X or Z axes of the capsule, even such rigid guests are capable of rotation around their Y-axis. The net effect of this movement is that by NMR the two hemispheres of the capsule are non-equivalent but each maintains its C_4 symmetry. In contrast, although flexible and highly mobile guests such as ethylene glycol monononyl ether 23 have been shown to bind in a similar pole-to-pole manner (Fig. 11), the two hemispheres of the capsule appear equivalent because on the NMR timescale the guest can freely tumble around the X or Z-axis of the container.³²

In an effort to gain a better understanding of the mobility of molecules within the capsule formed by OA 2, the Turro and Ramamurthy groups used a combination of NMR and EPR experiments to analyze the movement of encapsulated guests

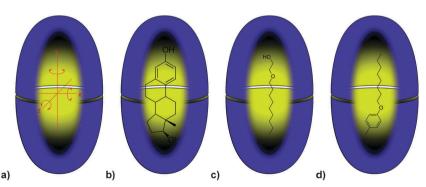


Fig. 11 (a) Axes within the capsule; (b-d) encapsulated guests illustrating predominant packing motif: (b) estradiol 15; (c) ethylene glycol monononyl ether 23; (d) 1-methyl-4-(octyloxy)benzene 32.

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within different complexes. 42 Their approach was to classify guests according to whether not they were capable of rotation around their X or Z axis, and if so, the temperature required to do so and whether or not the rotation was faster or slower than the NMR timescale. The ¹H and NOESY NMR data gathered in this study indicates that as in the case of ethylene glycol monononyl ether 23, the NMR spectra of complexes of guests such as 1-methyl-4-(octyloxy)benzene 32 (Fig. 11) show the two capsule hemispheres to be equivalent. Furthermore, the data also showed that the guest rotation around the X or Z axis needed to induce this equivalence did not involve disassembly/ reassembly of the capsule. In addition, the accompanying EPR

studies revealed that, as the guest was increased in length so

the rate of rotation around the Y-axis decreased.

Although not specifically discussed in Section 2.1.1.c, the binding of n-alkanes from C₅ to C₁₇ inside the dimer formed by OA 2 gave valuable early information about how guests pack within the capsule.31 Thus, 1H NMR investigations focusing on the spectral difference between the free and the bound state of the guest revealed pole-to-pole guest binding. Because of the tapering nature of these deep-cavity cavitands, a general rule is that the more deeply an atom resides in a host the more it is upfield shifted relative to the free state. Most simply put, at the base of the pocket the aromatic walls of the host are closest together and therefore exert the largest shift in the NMR signals of atoms located there. In contrast, at the equator the greater separation of the aromatic rings results in smaller upfield shifts. Hence an evaluation of the $\Delta\delta$ value of the free *versus* bound signals from the guest gives valuable information about the time-averaged guest-packing motif within the host dimer. For all the *n*-alkane guests examined it was the terminal methyl groups that displayed the largest $\Delta\delta$ value, with upfield shifts as high as -3.15 ppm in the case of C_{12} . In contrast the guest protons midway along the alkane chain were the least shifted, indicating they reside near the equatorial region of the capsule.

Space-filling models illustrate that C₁₂ is just slightly longer than the Y-axis of the capsule and can therefore occupy the cavity in an essentially fully extended conformation (Fig. 12a). Guests longer than this must however compress somewhat to be accommodated. Do these compressed guests adopt any distinct form? NOESY and COSY NMR experiments were also utilized to elucidate the conformational preferences of these guests, and the former revealed stronger NOE C_i-C_{i+1} and C_i-C_{i+2} correlations between the terminal methyl groups and the penultimate methylenes than for the methylenes at the center of the cavity. In addition, C_i – C_{i+3} and C_i – C_{i+4} were also observed with the longer guests. As with previous work from the Rebek group, 43 the inference from this data was that guests such as C14 adopt helical conformations within the cavity of the OA 2 dimer (Fig. 12b). Models demonstrate that the helical motif allows both for maximal C-H \cdots π interactions with the cavitand walls and for maximal packing within the cavity; two factors that counter the strain of contiguous gauche dihedrals down the length of the chain.

Two recent papers have demonstrated that other motifs are also possible. In the first, the Prabhakar and Ramamurthy groups examined the binding of phenyl-substituted hydrocarbons (Fig. 13) to the capsule formed by OA 2 using a combination of NMR and molecular dynamics (MD) simulations.44

As expected, small guests such as 33 were found to pack the cavity in an essentially fully extended manner, with the phenyl group in one hemisphere and the methyl binding to the polar region of the other. The case of 34 is very similar, but being 4 Å longer the guest must adopt a slightly compressed motif and its movement within the cavity is restricted. At room temperature the guest signals are broadened somewhat, but these are seen to sharpen as the temperature is increased. For the longer guests examined, e.g., 35, both NMR and the MD simulations demonstrate that these molecules are forced to fold into J- or U-shaped motifs. Such motifs are frequently observed in fatty acids bound to fatty acid binding proteins,45 and have also

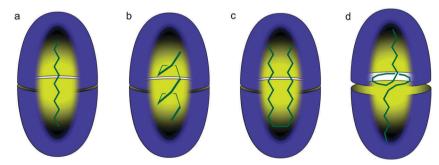


Fig. 12 Schematic representations of observed binding motifs within deep-cavity cavitand hosts.

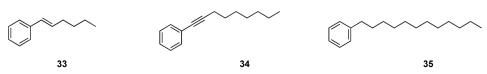


Fig. 13 Guests for probing conformational preferences within the dimer of OA 2.

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been observed with similar guests binding to cucurbiturils. 46 Overall, it was noted that the inclusion of saturation in the hydrocarbon sidechain has a pronounced effect on the conformational options available to the guest. In passing, it is perhaps worth noting that these J- and U-motifs suggest a mechanism by which the ends of very flexible guests (e.g., Fig. 12c and d) can exchange hemispheres without complex disassembly.

The second paper in question reveals a further packing motif. This study concerned a combined study of long *n*-alkanes within the capsule formed by OA 2 and the hexalenediamine-linked dimer (HOA) 36.

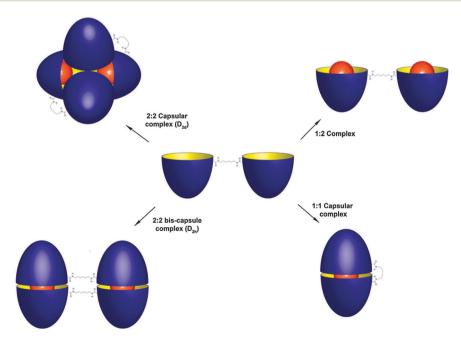
This covalently-linked dimer of OA was readily synthesized by taking advantage of a modification to the esterification of OA 2 (Scheme 1) that gave a high yield of the corresponding heptaester with one free carboxylate group at the rim of the cavitand. Two equivalents of the hepta-ester were then covalently linked using 1,6-hexanediamine and HBTU as a coupling agent to give the corresponding tetradeca-ester, subsequent hydrolysis of which gave HOA 36.47

The covalent linking of OA 2 in this manner results in a host with quite different assembly modes and subtle differences in guest packing. Regarding the former, the possible complexes/assembly

products pertinent to this discussion are: the open 1:2 hostguest complex; the 1:1 capsular complex; the 2:2 bis-capsule complex; and the 2:2 capsular complex (Scheme 4). The 2:2 bis-capsule complex possesses two essentially independent inner volumes, whereas the 2:2 capsular complex possesses a single inner space more capacious than the 2:2 bis-capsule complex because the internal volume is comprised of the binding pockets of four cavitands plus a central, pseudo tetrahedral space, defined by the four cavitands.

As extensive NMR analysis reveals OA 2 and HOA 36 have quite different properties. Earlier work had reported that for *n*-alkanes the guest-size limit of OA 2 was C₁₇, ³¹ but this proved to be largely the result of poor solubility and poor uptake and not a limiting factor of the host itself. Thus, in this more recent report a combination of heating and sonication allowed encapsulation of guests inside of the OA 2 dimer as large as *n*-hexacosane (C_{26}). As noted previously, guests up to twelve carbon atoms (dodecane) adopt extended motifs with contiguous anti dihedral angles, whereas longer guests adopt a more compressed motif. These E/C (extended/compressed) motifs cannot be readily distinguished by NMR, although helical motifs (Fig. 12b) can.

For alkane guests exceeding eighteen carbon atoms (n-octadecane) it is not possible to pack OA 2 in either of these motifs. Instead, NMR demonstrates that guests such as n-eicosane (C20) pack the cavity by adopting a U-shaped motif (Fig. 12c). This motif holds true for guests up to n-tricosane, however the largest guests seen to unequivocally bind to the capsule formed by OA 2 (C₂₄ to C₂₆) cannot adopt this U-shape. Instead, for these guests the motif appears to be akin to a spinning-top (Fig. 12d) whereby the two terminal ends of the alkane chain are nestled in the poles of each cavitand and the central portion of the hydrophobic chain sandwiched between the two aromatic rims of the host.



Scheme 4 Assemblies of HOA 36

HOA 36 was found to form a 1:1 capsular complex in the presence of short n-alkanes (C_6 , C_8 and C_{10}) as well as n-hexadecane (C_{16}), and in each case the packing motif of the guest was as is observed with OA 2. However packing of alkane chains with eighteen to twenty-three carbons displayed very different binding profiles, while packing of guests with more than twenty-three carbons did not occur as the linker prevented sufficient separation of the two hemispheres of HOA 36 to allow the guest to adopt the spinning-top motif.

For the longer alkanes (C_{18} to C_{23}), NMR experiments revealed two complexes for each guest: one with a lower energy E/C conformation and one with a higher energy U-shaped motif. Unusually, as the guest size was increased so the amount of the lower energy E/C motif increased. PGSE NMR unraveled this conundrum. Whereas the diffusion constants for the complexes with the U-shaped motif were the same as for the 1:1 complex between HOA 33 and *n*-dodecane C_{12} (*D* = 1.31 × 10⁻⁶ cm² s⁻¹), the complexes with the E/C motif had significantly smaller diffusion rates ($D = 0.95 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$) corresponding to a dimer host structure. Although it was not possible to unequivocally differentiate between the 2:2 bis-capsule complex and the 2:2 capsular complex (Scheme 4), the ¹H NMR data supported the latter. Furthermore, as the 2:2 capsular complex is necessarily more capacious than the 2:2 bis-capsule complex, this assumption fits with the guest motif changing to a more relaxed E/C motif as the guest size increases. In summary, whereas OA 2 accommodates very large guests by adopting a high energy spinning-top motif (the motif of last resort), HOA 36 accommodates such guests by switching from a 1:1 capsular complex to a more voluminous 2:2 capsular complex.

We complete this section by noting that it is not just highly flexible guests than can be persuaded to adopt unusual conformations or motifs within the confines of these supramolecular capsules. For example, a comprehensive NMR examination of piperidine derivatives 37–41 (Fig. 14) revealed that the conformational preferences of 37, and 39–41 within the capsule formed by OA 2 were similar to that seen in solutions; these guests adopt a conformation in which the OR group is exclusively equatorial. On the other hand, when 38 is encapsulated within the OA 2 dimer it adopts a 53:47 mixture of the equatorial and axial conformers that interconvert slowly on the NMR timescale. Apparently, the *n*-Pr group is just the right size to reduce the energy difference between the more stable equatorial conformer and the axial form, where as an ethyl group is too small

Fig. 14 Piperidine guests encapsulated within the OA 2 dimer, and their conformational options.

and a n-butyl group too big. Such precise control of one molecule in a homologous series suggests interesting possibilities vis- \dot{a} -vis reaction control (see Section 2.1.2.b).

(f) Communication through the capsule. As has been reviewed, when a guest is bound inside the dimer formed by OA 2 it is not completely isolated from the bulk. Thus even during occupancy, guests can be exposed to small co-guests such as molecular oxygen via capsule 'breathing'. Furthermore, as we highlight in this section, direct communication between bound guests and the bulk is also possible through the walls of the host.

The communication between molecules entrapped within the OA 2 dimer and molecules in free solution has been extensively investigated. As detailed below, to date the following systems have been explored: (1) confined excited singlet or triplet donors and a free acceptor (respectively singlet-singlet and triplet-triplet energy transfers); (2) a confined radical and one free in the bulk (spin-spin exchange); (3) an encapsulated, excited molecule and a radical in free solution; (4) internalized fluorescence resonance energy donors and external acceptors (FRET), and; (5) confined electron donors and free acceptors (electron transfer).

The Turro group has led investigations into the ability of guests inside the capsule to 'communicate' through the walls of the host with guests in bulk solution. 49 The first example of communication involved a superexchange process whereby the orbitals of a guest nitroxide interact with those of a nitroxide solute in free solution through the bond orbitals of the host. The guests utilized for this study were 42 and 43 (Fig. 15). Encapsulation of paramagnetic guest 14N labeled 42 inside the capsule lead to broadening of the ¹H NMR spectrum of the complex. Although this is good evidence of complex formation, conformation of guest internalization was sought using 14N labeled diamagnetic guest 43. This similarly sized molecule resulted in a 2:1 host-guest complex that was readily characterized by ¹H NMR. Subsequent analysis of the Electron Paramagnetic Resonance (EPR) rotational correlation times and hyperfine coupling constants for the complex between OA 2 dimer and the ¹⁴N labeled **42** confirmed encapsulation. As anticipated by the researchers, EPR studies of the capsular complex of diamagnetic ¹⁴N guest 43 demonstrated that positively charged ¹⁴N labeled solute **44** bound to the outside of the capsule, but the negatively charged 45 did not. Furthermore, using the capsular complex containing the 15N paramagnetic guest 42, EPR demonstrated a strong electron spin-electron spin interaction between the encapsulated guest and the cationic 14N solute 44. As expected however, the same experiment using the anionic 14N solute 45 indicated no interaction with the anionic capsular complex.

Building on this steady-state EPR work, the Turro and Ramamurthy groups used time-resolved EPR to gain a better understanding of superexchange between an encapsulated guest and a solute. ⁵⁰ Key to this work was the encapsulation complex formed between ¹⁵N thioxanthone guest **46** (Fig. 15) and the dimeric capsule formed by OA **2**. Steady state EPR spectroscopy demonstrated that encapsulated ¹⁵N **46** possessed the expected increased rotational correlation time and decreased hyperfine

Fig. 15 Guests and solutes used by the Turro and Ramamurthy groups to explore through-the-wall communication in the OA 2 dimer.

coupling constant (relative to free solution) indicative of restricted motion within a non-polar environment. Separately, transient triplet-triplet absorption measurements upon 14N 46 and 47 in acetonitrile revealed that upon irradiation at 355 nm their thioxanthone moieties were readily excited to the triplet state, but that in this case of ¹⁴N 46 this was rapidly quenched by the adjacent nitroxide. Furthermore, time-resolved EPR of ¹⁴N **46** in acetonitrile (355 nm, 5 ns) demonstrated that during this quenching process the spin-polarized triplet states of the thioxanthone moiety transferred their spin polarization to the nitroxide. A similar time-resolved EPR experiment with ¹⁵N guest **46** encapsulated within the OA **2** dimer revealed the same phenomenon, but that confinement resulted in a longer spin polarization lifetime for the nitroxide. Following this, an analogous experiment but with the addition of 14N solute 44 in bulk solution demonstrated that spin polarization of the incarcerated guest could be transferred to the cationic solute. Thus the time-resolved EPR kinetic traces of spin polarization for both the 15N guest 46 and the 14N solute 44 revealed a reduced lifetime of the former, and a delayed rise-time of the spin polarization of the latter. That association with the outside of the capsule is key was demonstrated in two ways: by the addition of cucurbit[8]uril that strongly sequesters 44, or by using anionic guest 45 that is repelled from the surface of the host. In both cases no superexchange between encapsulated guest and solute occurred. As excited thioxanthone 47 and benzil 30 (Fig. 9) guests led to opposite signs of spin polarization of nitroxides, it was concluded that the principle mechanism for the formation of polarized nitroxides involved the electron spin polarization transfer mechanism.51

A different viewpoint on the barrier engendered by the wall of the capsule formed by OA was obtained by examining the communication (exited spin state-radical spin transfer) between solutes 44, 45 and 48, and a range of eight internalized

guests including benzil 30 (Fig. 9), thione 49, and stilbene 50 (R = 4-Me, Fig. 15). The cationic, anionic and neutral states of the nitroxide radicals 44, 45 and 48 allowed control of the lifetime of the solute-capsule encounter complex, and in the case of the cationic solute 44 NMR signal relaxation times suggested that it specifically associates at the equator of the capsule. These results also suggested that 48 has no specific interaction with the capsule, whilst not unexpectedly 45 is repelled from it. Although the guests examined formed both 2:2 and 2:1 host-guest complexes and formed a range of excited states, quenching of the excited, entrapped guest by the nitroxide solute 44 revealed only two distinct systems: (1) triplet states that are quenched dynamically by cationic nitroxide 44; (2) triplet and excited singlet states that are quenched via a static process. Moreover, the decay curves and lifetimes of the encapsulated, excited state guests revealed that regardless of the lifetime of the former, they are quenched with almost identical rate constants. In contrast the latter group all have short ($<5 \mu s$) lifetimes. En masse, a detailed analysis of these results supported the hypothesis that if a guest relaxed more slowly than 5 µs then its relaxation is controlled by through the wall quenching as a result of diffusion controlled encounter complexes, whereas guests with lifetimes shorter than 5 µs undergo static relaxation in the cavity of the capsule.

As an alternative to this short range (Dexter type) superexchange, the Ramamurthy group have used femtosecond (fs) up-conversion to study longer range Förster resonance energy transfers (FRET) between an encapsulated donor and an cationic acceptor associating with the outer surface of the OA 2 capsule. 52 In these experiments the (bound) donors were coumarin dyes such as 51, whilst the acceptor in free solution was rhodamine 6G 52 (Fig. 15). A combination of steady-state fluorescence and ¹H NMR confirmed that the different donors examined were bound within the dry interior of the capsule, whilst fluorescence,

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¹H, and diffusion NMR spectroscopies revealed that the cationic acceptor had an affinity for the outside of the negatively charged capsule.

The accompanying time-resolved fluorescence studies involving the encapsulated coumarin dye and the externally bound 52 revealed a rapid decay of the excited state of the donor. This was attributed to an ultrafast FRET process arising from the close proximity of donor and acceptor; a hypothesis that was confirmed by the rapid rise on the acceptor emission observed in a fs up-conversion experiment. Calculations from these studies suggested that the donor-acceptor distance in these systems was a mere 13 Å.

It is not only spin and energy that can be transferred through the walls of the capsule formed by OA 2; electron transfer is also possible. In a step towards this, the Kaifer group examined the properties of encapsulated ferrocene 53 (Fig. 16).⁵³ A combination of ¹H and PGSE NMR experiments demonstrated that 53 formed a 2:1 host-guest complex with OA 2. Neither cyclic voltammetry (CV) nor square-wave voltammetry (SWV) detected any faradaic response from the encapsulated 53, and a series of experiments varying the host-guest ratio indicate that inside the OA 2 dimer the guest does not give rise to a measurable current-potential response.

Whereas ferrocene 53 is strongly bound within the host capsule, ¹H and PGSE NMR clearly showed that methyl, ethyl, and n-butyl viologen 54 (R = Me, Et and n-Bu respectively) bind to the exterior of empty OA 2 dimer. Indeed, two molecules of viologen bind to this empty capsule, suggesting that the four carboxylates at each pole are the viologen-specific binding site. That noted, this was the first example of an empty capsule formed by OA 2; all work prior, and the vast majority of subsequent work, demonstrates that an internalized guest is required for capsule formation. These facts suggest that there is also likely association of the viologens with the equatorial carboxylates of the capsule that helps to hold the empty capsule together.

CV studies of viologens 54 associated with OA 2 demonstrate the two expected reduction steps (to radical cation and then to the neutral), but with substantially reduced current levels.

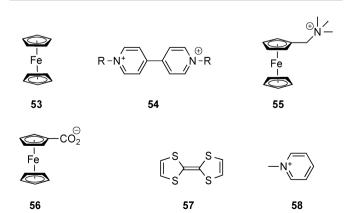


Fig. 16 Guests and solutes used to study electron transfer through the walls of the OA 2 capsule.

Furthermore, these studies revealed that the host stabilized both the reduced forms; particularly the first (cationic) reduction product. Furthermore, the rates of heterogeneous electron transfer for the viologens associated with the OA 2 dimer were significantly attenuated relative to free solution. This was attributed to the association between the anionic poles of the OA 2 capsule and the viologen increasing the average distance of maximum approach between the viologen center and the electrode surface.

Subsequently it was shown that the capsule formed by the dimerization of OA 2 around ferrocene 53 could also (externally) bind ethyl viologen (54, R = Et) to form the corresponding quinternary complex.

The Kaifer group subsequently built on this work by demonstrating cationic ferrocene derivative 55 (Fig. 16) associates strongly with the outer surface of the OA 2 capsule containing ferrocene 53, and that this quaternary complex can undergo a homogeneous electron transfer process.⁵⁴ PGSE NMR confirmed the quaternary complex, yielding very similar diffusion coefficients from signals corresponding to the capsular complex and 55. A similar situation was obtained if (E)-stilbene (50, R = H)replaced the internalized guest 53.

When the voltammetric behavior of the quaternary assembly was investigated with the encapsulated stilbene, the current levels were depressed relative to 55 alone. However, when stilbene was replaced by guest 53 a number of changes to the cyclic voltammogram were observed. On the forward scan, there was no faradaic response detected where 53 would be directly oxidized, but the oxidation of 55 was observed, albeit at higher current levels than expected. Furthermore, on the reverse scan, two peaks were observed: one corresponding to the reduction of the ferrocenium form of 55, and a second corresponding to the reduction of ferrocenium form of 53. The results demonstrate that once oxidized, the ferrocenium form of 55 accepts an electron from the internalized 53, to generate the cation (ferrocenium) form 53 inside the capsule. However, this guest is a poor one and it is released to bulk solution. This process was fully reversible and so the OA 2 capsule containing 53 was again formed upon reduction. Control experiments were also carried out with 56. In this instance, there were no changes in the CV response in the presence of the OA 2 dimer encapsulating either (E)-stilbene 50 (R = H) or 53. Furthermore, the complex of 55 with cucurbit[7]uril was determined to be a poor promoter of the electron transfer process in the system where 55 needs to associate with the outer walls of the OA 2 dimer. Clearly, there must be close association between the internalized and external ferrocene derivatives for electron transfer to occur.

A second example of redox-controlled capsular assembly/ disassembly from the Kaifer group focused on tetrathiafulvalene (TTF) 57 (Fig. 16) encapsulated inside the OA 2 dimer.⁵⁵ As anticipated, ¹H and PGSE NMR demonstrated that 57 formed the corresponding 2:1 complex. These studies were performed at 1 mM concentrations, and therefore the complex represents a much higher concentration of 57 'in water' than is usually possible.

Tetrathiafulvalene 57 and its derivatives undergo two consecutive one-electron oxidations to first generate the corresponding

radical cation and eventually the dication form at more positive potentials. However the first oxidation is usually poorly developed in water because of the low solubility of 57.

The electrochemical behavior of this OA 2 dimer complex containing 57 was analyzed by both CV and SWV. In the absence of OA 2, two shallow waves at +0.18 V and +0.48 V were observed in the forward scan of the cyclic voltammogram, and no peaks were apparent on the reverse scan. This is consistent with the behavior of 57 in aqueous solution. However, in the presence of two equivalents of OA 2, the small initial current corresponding to the first oxidation step disappeared entirely and there was a well-developed peak corresponding to the second oxidation process at +0.56 V. In the reverse scan only one small wave was observed corresponding to the second reduction to regenerate neutral 57. The SWV trace confirmed the hindered voltammetric response associated with encapsulated 57, which was attributed to the increased distance between it (versus free TTF) and the electrode surface. In contrast, the well-developed second oxidation process was attributed to the release of the cationic form of 57. The solitary, small peak observed in the reverse process was attributed to the dicationic form of 54 being unstable under basic conditions. Larger tetrathiafulvalene derivatives were also examined in this study, but they were found to not bind to the host.

More recently, electron transfer across the walls of the OA 2 capsule has also been observed by the Ramamurthy group. 56 To demonstrate this, one donor-acceptor pair ideally suited to the capsule formed by OA 2 is 4,4'-dimethylstilbene 50 (R = 4-Me, Fig. 15) and N-methyl pyridinium 58 or viologen 54 (R = Me, Fig. 16). ¹H NMR demonstrated that electron donor **50** is strongly encapsulated within the capsule, whilst diffusion NMR revealed that both 54 (R = Me) and 58 associated with the exterior of the capsule via Coulombic forces. Fluorescence spectroscopic analysis of the titration of 54 or 58 into a solution of encapsulated 50 revealed that the pyridinium solutes statically quenched the fluorescence of the encapsulated guest. Laser flash photolysis pin-pointed the cause of this quenching, namely the transient formation of the radical cation of 50. The presence of two transient absorptions (at 510 and >700 nm) with identical decay kinetics suggested that the quenching was due to electron transfer from the singlet excited state of 50 to 54 or 58. In the case of viologen 54 (R = Me) it was also possible to observe the mono-cation radical 'product' resulting from electron transfer. Further supporting electron transfer were similar fluorescence and transient absorption experiments in the presence of cucurbit[7]uril (CB7). CB7 is a strong binder of viologen 54 (R = Me), and consequently its addition restored fluorescence of the internalized 50, and quenched the observed transient absorption. Laser flash photolysis also revealed that the lifetime of the radical cation of 50 was < 20 ns in the presence of 54 (R = Me) and 4.6 μ s in the presence of 58. This difference was attributed to the fact that the single electron reduction product of the latter is neutral but mono-cationic in the former, and that the mono-cation has a relatively strong affinity with the exterior of the capsule.

Building on this work, photo-induced electron transfer has been recently demonstrated using TiO2 as an acceptor.⁵⁷ In this work, the encapsulated donors were a series of three coumarins including guest 29 (Fig. 8) whilst for solution studies viologen 54 (R = Me) was again used as the acceptor that partitioned between bulk solution and the outer wall of the capsule. As anticipated, titration of the acceptor 54 (R = Me) into a solution of the capsular complex resulted in static quenching of the fluorescence of the donor 29, but that fluorescence could be recovered by the addition of a strong binder of 54 (R = Me) such as CB7. With this knowledge in hand, the emersion of films of TiO₂ (and as a control ZrO₂ films) into solutions of the capsular complex lead to the complex adhering to the surface of the oxides (Fourier Transform, Infra-Red Attenuated Total Reflection spectroscopy, FT-IR-ATR). Furthermore, in the case of the large band-gap ZrO2 film where electron transfer is not possible, fluorescence spectroscopy indicated that adherence to the surface did not induce capsular breakup and release of the coumarin. In contrast, adherence to the TiO2 film resulted in full quenching of the coumarin indicative of electron transfer. Finally, titration of TiO₂ into a solution of the capsular complex of OA 2 and the coumarins was achieved by using colloidal TiO₂. In this experiment the reduction in fluorescence suggested that quenching was again static rather than dynamic.

In a recent collaboration between the Burda and Ramamurthy groups the ultrafast electron transfer dynamics of these processes have been investigated using femtosecond laser spectroscopy.⁵⁸ In these studies coumarin-153 (51, Fig. 15) was used as the encapsulated donor, whilst once more viologen 54 (R = Me) and N-methyl pyridinium 58 (Fig. 16) served as the solute acceptors. The transient absorption spectra obtained using a 390 nm pump pulse confirmed the existence of the oxidized state of the internal excited guest and the reduced state of viologen 54 (R = Me) associating with the outside of the cavity. A kinetic analysis of the increase of the radical cation of 54 (R = Me) as a function of time, and the corresponding decay of the laser induced fluorescence of the excited donor 51, gave time constants of τ = 20 \pm 1 and 18 \pm 2 ps respectively. Furthermore, the recombination kinetics were measured at τ = 724 \pm 38 ps. These results were in sharp contrast with those obtained in free (30% aqueous acetonitrile) solution, where the electron transfer rate was much lower (τ = 631 \pm 50 ps) and almost identical to the recombination rate.

2.1.2 Applications for deep-cavity cavitands and their assemblies

(a) Novel separations strategies. The preferred encapsulation of one guest over another within molecular containers - differential compartmentalization - offers many intriguing possibilities vis-à-vis molecular separations. Examples of both physical and chemical-based separations have been documented with the deep-cavity cavitands discussed here. With respect to the former, the Gibb group has reported on the separation of hydrocarbon gases using dimeric capsules formed by OA 2.21 This study demonstrated (¹H NMR) that when *n*-butane was added to the headspace above a buffered solution of OA 2, the corresponding 2:2 capsular complex was formed in solution. Furthermore, these experiments revealed that it was possible to store n-butane as its 2:2 complex at ca. 200 times its nominal

solubility in pure water. The actual physical limit of solubility of this complex may be even higher than this as it was viscosity issues with the chosen sodium tetraborate buffer that restricted NMR analysis. NMR was also used to determine the stability of the 2:2 host-butane complex. As it was not possible to differentiate between the free host and guest and their 1:1 complex, an apparent K value was defined as: $K_{app} = [H_2G_2]/[H_a][G_a]$, with $[H_a] = [H_{free} + HG]$ and $[G_a] = [G_{free} + HG]$. Values for K_{app} ranged from 1400 M^{-1} in simple buffer solution to 1 × 10⁶ M^{-1} in 14 mM NaCl. A subsequent competition experiment between propane and n-butane demonstrated that the latter bound an order of magnitude more strongly ($K_{rel} = 12$), and this led to an investigation to determine whether or not the two gases could be separated. This was indeed the case. Thus, when the head-space above a solution of OA 2 was filled with a 1:1 mixture of propane and n-butane, only the latter was complexed leaving a propaneenriched gas-phase in the head space.

The Gibb group has also demonstrated an example of a chemical separation using these types of supramolecular capsules.⁵⁹ The focus of this report was on the supramolecular protection of molecules from reactive (hydrolytic) media. More specifically, pairs of difficult to separate, constitutionally isomeric esters were examined in basic media. The idea behind this work was that one guest would be preferentially bound to the capsule formed from OA 2, leaving the other molecule of the pair to reside in solution and undergo hydrolysis. Hence the supramolecular nanocapsules formed by OA 2 would bias the relative rates of ester hydrolysis and bring about kinetic resolution.⁵⁹ Two series of constitutional esters were examined: those composed of thirteen non-hydrogen atoms, and those composed of sixteen non-hydrogen atoms. The former were chosen in part because previously reported data³¹ suggested that dodecane and tridecane optimally fill the cavitand by neither being too compressed or by having too much empty space. The latter series was a test of what happens to a resolution at the limit of guest encapsulation. The obtained kinetic resolutions were highest with those composed of thirteen non-hydrogen atoms (Fig. 17, 59-63) and therefore are the primary focus of our discussion here.

The free esters were hydrolyzed in 3:7 acetone-d₆: D₂O containing 10 mM NaOH to obtain the pseudo first-order hydrolysis rates at room temperature. The methyl ester 59, and to a lesser extent the ethyl ester 60, hydrolyzed slightly faster than the others, but the relative rate (k_{rel}) between the fastest (59) and slowest (62 and 63) was only three, i.e., insufficient for kinetic resolution in itself. ¹H NMR and COSY NMR were conducted to determine the goodness of the fit of the guests in the series, and all were found to form strong 2:1 complexes within the OA 2 dimer. The relative binding constants (K_{rel}) between all the esters were then determined, with the strongest and weakest binding guests found to be 59 and 61 respectively ($K_{\rm rel}$ for esters 59-63 were respectively: 165.4 1.8 1.0 10.7 16.4). Two key factors were concluded to influence binding. First, the relative position of the ester group within the cavity presents significant steric clash as it is moved closer to the end of the molecule and hence deeper into the pole-region of the capsule. Second, this steric

Fig. 17 Constitutional isomeric esters for kinetic resolution.

clash is overcome by compensatory $-CH_3 \cdots \pi$ interactions between the methyl of the guest and the walls of the host when the C-H bonds are more polarized by a proximal electronegative atom (the sp3 O atom). Hence the methyl ester 59 binds strongest, the ethyl- and propyl-esters 60 and 61 the weakest, and the butyl and pentyl-esters 62 and 63 have intermediate affinity. It was noted in the corresponding experiments with the second set of esters that the range of binding affinities was essentially half that found in the first series, presumably because of increased steric strain in the more tightly packed cavity.

Competition experiments between pairs of esters were then carried out in the presence of the capsule formed by OA 2. As it was noted for a complex of a single ester that even after one month $\leq 5\%$ hydrolysis had occurred at room temperature, all competition experiments were carried out at 100 °C. After cooling of each sample the extent of hydrolysis was determined by NMR. When pairs of esters were competed against each other it was determined that for esters with similar K_{rel} kinetic resolution was not possible, but for those with K_{rel} an order of magnitude or more different, fair to good kinetic resolution of isomers was observed. Larger K_{rel} differences still led to very good resolutions. Thus, the best resolution was determined between 59 (the strongest binder) and 61 (the weakest). In this case when all of the latter had reacted, 84% of the former still remained. Subsequent studies in the presence and absence of host indicated that although the rate-determining step in the hydrolysis of free ester is nucleophilic attack by hydroxide, this was not the case for the encapsulated ester where instead the rate-determining step was likely capsule opening. To test this, the authors added a stronger-binding, inert guest (n-dodecane, C₁₂) as they monitored the slow hydrolysis of 61; almost immediately following injection the ester was displaced, and hydrolyzed within 10 minutes. Furthermore, using the projected kinetic data for hydrolysis of free ester at 100 °C (via Arrhenius plots), a model based on the notion that capsule opening was the rate-determining step fitted well with the kinetic traces of ester hydrolysis in the presence of a capsule. Overall, these

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studies demonstrated that kinetic resolutions can be brought about via selective molecular protection; selective binding one of the reactive molecules biases reactions that, in the absence of the capsule, are otherwise very similar.

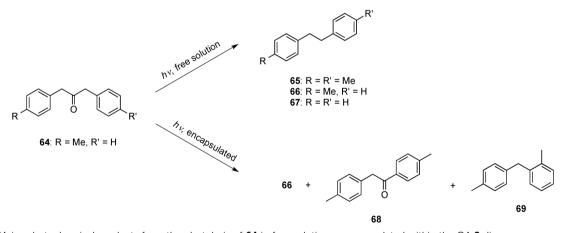
(b) As nano-scale reactors. As the last example of the previous section demonstrated, encapsulation can protect molecules from reactive external environments. The opposite of this strategy is to use the inner nano-space of capsules as yoctoliter (10⁻²⁴ L) reaction vessels; a popular topic in supramolecular chemistry,60 and one we discuss in this section.

The vast majority of chemical conversions carried out in the capsules discussed here involve OA 2 and photochemistry. A collaboration between the Gibb and Ramamurthy groups established the foundation for these studies, and from this the Ramamurthy group have built up a flourishing deep-cavity cavitand program within their field of expertise of studying photophysical and photochemical processes within confined media.

Arguably the most studied reaction within the capsule formed by OA has been the photolysis of dibenzyl ketone derivatives; a class of reaction that has shed much light on the ability of capsules formed by OA 2 to bring about unusual photochemical reactions. By way of example, 38 the photolysis of ketone 64 (Scheme 5) in aqueous solution gave, via homolytic cleavage of the C(O)-C bond and decarbonylation, products 65 (AA) 66 (AB) and 67 (BB) in the expected (statistical) 1:2:1 ratio. In contrast, irradiation of 64 encapsulated within the OA 2

dimer led to only type AB products because the radical formed from 64 are - in the timeframe of the photochemical conversion – trapped within the capsule. This exceedingly high (100%) cage effect had only previously been observed in the solid-state. More interestingly, the major products formed from encapsulated 64 were identified as rearrangement product 68 (44%), 66 (41%), and a mixture of various decarbonylated, ortho substituted derivatives such as 69 (15% combined yield). Compound 68 is believed to be formed (Fig. 18) via cleavage of the C(O)-C bond, intersystem crossing from the triplet to the singlet radical pair, rapid reorientation of the unsubstituted benzyl radical, recombination and a 1,5 H-shift. Reorientation occurs because whereas the phenylacetyl radical is anchored in place by its p-methyl group, the benzyl radical isn't. Consequently, it rotates 180° relative to rest of the complex to better pack the cavity and place the benzyl group in the narrow region at the base of the nano-space. A hydrogen shift and recombination of the radicals gives product 68.

Extending this work, the Gibb and Ramamurthy groups examined α -(n-alkyl) dibenzyl ketones 70 (Scheme 6) where R varied from Me to n-octyl. Extensive NMR analysis including ¹H studies, diffusion, and 2D experiments revealed all guests formed 2:1 host-guest complexes with three distinct packing motifs depending on the R group of the guest. For the smaller R groups (R = Me, Et, n-Pr) it was found that each phenyl ring occupied the polar regions of the container whilst the R group occupied the equatorial region (Fig. 19). In contrast, for mid-sized



Scheme 5 Major photochemical products from the photolysis of 64 in free solution or encapsulated within the OA 2 dimer

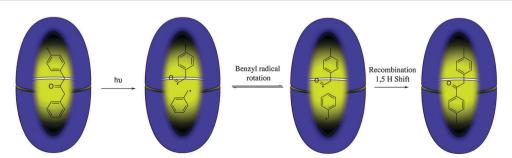
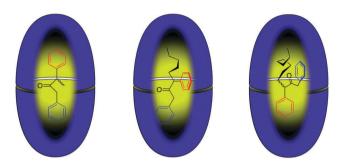


Fig. 18 Schematic of the proposed mechanism for the formation of rearranged product 68

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Scheme 6 Examples of photochemical products from the photolysis of 70 (R = various) encapsulated within the OA 2 dimer.



Fia. 19 Packing motifs observed for the different guests within the OA 2 capsule

R groups (R = n-Bu n-pentyl and n-hexyl) it was found that one phenyl ring (distal to the R group) and the R group occupied the polar regions of the container and the proximal phenyl ring occupied the equatorial region. Finally, in the case of the largest R groups examined (R = n-heptyl and n-octyl) it was found that the guest packed the cavity such that the distal phenyl ring occupies the equatorial region of the capsule.

In hexane or buffer solution the majority of products from photolysis of these eight ketones arise via Norrish Type I mechanisms and indicate no cage effect. In contrast, photolysis within the capsule revealed again a 100% cage effect with only AB type products observed. Nevertheless, the relationship between the packing of the capsule and the outcome of photolysis was complex because of the multiple chemical channels in the triplet excited-state surface. Consequently, the guests examined yielded a considerable range of products arising from both Norrish Type I and Norrish Type II (Norrish-Yang) chemistry. Prominent examples include rearranged products 71, 72 and decarbonylated product 73 via Norrish Type 1 mechanisms, and dealkylated (Norrish-Yang) products such as 74.

Although overall many products were formed, the range of products from individual host-guest complexes was much narrower than in free solution and some general conclusions regarding packing and reactivity were possible. First, smaller R groups (R = Me, Et, and n-Pr) formed substantial amounts of rearranged product 72 and only trace amounts of the other rearranged product 71. This was attributed to the faster cleavage of the more substituted C(O)-C bond of 70, and the fact that the resulting substituted benzyl radicals from the three guests were able to rotate in the cavity and generate 72. On the other hand, midsized R groups (n-butyl and n-pentyl)

give roughly equal amounts of both rearranged products 71 and 72. Although the cleavage of the more substituted C(O)-C bond of 70 would be expected to the faster, apparently the reorientation of the resulting substituted benzyl radical is difficult. Consequently, via recombination of the initially formed phenacetyl and benzyl radicals, the slower cleaving C(O)-C bond that is the first step to product 72 effectively competes. Finally, the largest R groups (n-hexyl through n-octyl) gave the most Norrish-Yang products such as 73. This was attributed to two phenomena: (1) with little space in the cavity, Type I chemistry and rearrangement processes were disfavored; (2) the conformation preference (Fig. 19) for these guests promotes γ -hydrogen abstraction and Type II chemistry.

A different perspective on this encapsulation chemistry was obtained with the series of dibenzyl ketones 75 (R = Et through n-hexyl, Scheme 7).61 With 75, the outcome of photolysis in hexanes (Scheme 7) is independent of the nature of the R group, with in all cases, a statistical mixture (1:2:1) of the AA, AB, and BB decarbonylated products 76-78. In the case of the encapsulated reactions all the products were again of the AB type (100% cage effect), with the predominant products being 77, 79, 80 and 81. In general, yields of 79 decreased and those of 77 increased as R increased in size, suggesting that with larger and larger R groups rotation of the intermediate benzyl radical became energetically more demanding and the decarbonylation pathway became more competitive. Yields of 80 and 81 were less dependent on the R group size, suggesting that the increase in yield of decarbonylated 77 is at the expense of rearrangement product 79.

One question that the photolysis of these different type of dibenzylketones does not reveal is the extent of recombination of the phenylacetyl and benzyl radicals to regenerate the starting ketone (Fig. 18). To address this question a recent study examined the photolysis of optically pure α-deoxybenzoins (Scheme 8) encapsulated within the dimer capsule formed by OA 2.62 These guests can only undergo cleavage on one side of the carbonyl group, and so photolysis of these complexes forms a narrower range of products. With 82 (R = H) only Type I reactions are possible and the rearranged product 83 (R = H) was formed quantitatively. Guest 82 (R = Et) formed mostly 83 (R = Et), but also the Type II products 84 ($R^1 = Me$) and 85. In contrast, guests 82 (R = n-pentyl and n-heptyl) gave no Type I products but only 84 (respectively $R_1 = n$ -Bu and n-pentyl) and 85. Equally important, in all cases if photolysis was kept to <20% the recovered starting

Scheme 7 Major photochemical products from the photolysis of 75 in free solution or encapsulated within the OA 2 dimer.

Scheme 8 Major photochemical products from the photolysis of 82 in free solution or encapsulated within the OA 2 dimer.

material was found to have undergone partial racemization; a phenomenon that necessitates the first step of the Type I mechanism, i.e., formation of the phenylacetyl benzyl radical pair. Two key observations were made from the data: (1) longer chain lengths led to reduced extents of racemization; (2) but even with guests 82 (R = n-pentyl and n-heptyl) which gave no Type I products, the first step of the Type I mechanisms is not suppressed. Overall these studies revealed that the reduction of free space in the complex with increasing R group significantly decreases the ability of the guest to undergo Type I chemistry and rearrangement—100% yield of 83 for 82 (R = H), 0% for 82 (R = n-pentyl and n-heptyl)—but the effect of packing coefficient on the formation of the phenylacetyl/benzyl radical pair and subsequent racemization is much smaller 75:25 er versus 88:12 er for 82 (R = H and *n*-heptyl respectively). In other words, formation of the phenylacetyl benzyl radical pair occurs with all guests, but increasing packing coefficient has more of an effect on the rearrangement reaction to form 83 than on flipping of the benzyl radical necessary for racemization.

Studies of the photophysical and photochemical properties of encapsulated stilbenes also reveals that the cavity formed by the dimerization of OA 2 has a profound effect on encapsulated guest molecules.63 In this work the authors examined a series of cis- and trans-methyl stilbenes (2-, 3- and 4-substituted) as well as dimethyl stilbenes (2,2'-, 3,3'-, and 4,4'-) **50**. (Fig. 15). NMR studies demonstrated that both cis- and trans-stilbenes readily form 2:1 host-guest complexes, with the latter more efficiently filling the nano-space of the capsule. Furthermore, the NMR shift data between the free and the bound states suggested that it was the 4,4'-disubstituted stilbene that demonstrated the greatest difference between the cis- and trans-isomers; the more rotund form of the former guest only filled the 'equatorial' region of the capsule, whereas the shape of the latter was sufficient to more efficiently fill the bulk of the interior, and in particular, allow its two methyl groups to bind deeply into the narrow 'polar' regions of the nano-space.

Fluorescence studies revealed that the stilbenes displayed weak aggregate emission in free borate solution, but welldefined monomer emission when encapsulated. Furthermore, the singlet lifetime of all the disubstituted guests was longer than the corresponding values in hexanes solution (the corresponding lifetimes of the mono-substituted stilbenes were too short to measure).

Photochemical studies revealed that the 4,4'-dimethyl stilbene 50 (R = 4-Me) was unique among all the different isomers examined. Free in hexane solution or bound in the capsule, irradiation of the other isomers led to similar cis/trans ratios in the pseudo-photostationary states (Scheme 9). However, in the case of 4,4'-dimethyl stilbene 50 its pseudo-photostationary state in hexane was 18% trans and 76% cis, but (a more slowly attained) 85% trans and 15% cis in the capsule. Furthermore, whereas irradiation in hexane led to high yields of the corresponding phenanthrene 31, only trace amounts of 31 formed over extended irradiation of the bound 4,4'-dimethyl stilbene 50. These results demonstrated that the two methyl groups of the guest are uniquely positioned to anchor the two ends of the guest in the nano-space of the capsule, inhibiting the formation of the preferred cis-isomer and so preventing the formation of the expected phenanthrene 31.

A recent related paper examined the photophysical and photochemical properties of an expanded array of encapsulated

Major photochemical products from the photolysis of 50 in free solution or encapsulated within the OA 2 dimer.

Scheme 10 Major photochemical products formed from 86 in free solution or encapsulated within the OA 2 dimer (see text)

stilbene and stilbene-like guests molecules.⁶⁴ In this work, the photostationary states of the different guests were attained via a second OA 2 capsular complex containing a triplet sensitizer. A comparison of qualitative competition studies for the encapsulation of the different cis/trans isomers, with the photostationary state ratios obtained upon photolysis suggested that the latter is biased in the capsule by the relative binding affinities of the guests. Models explaining the mechanism by which energy is transferred from the encapsulated, excited donor and the encapsulated acceptor were discussed.

A number of other photochemical transformations have been investigated within the confines of the dimeric capsule formed by OA 2. For example, the dimerization of acenaphthylene 86 in water gives 40% of the syn-dimer and 60% of the anti stereoisomer (Scheme 10). In the capsule however the syn-dimer is formed quantitatively.⁶⁵ Interestingly, when the water-soluble triplet sensitizer Eosin-Y is added to the reaction, in the case of free 86 the syn: anti ratio is 51:49, and in the case of the encapsulated 86 60:40. The authors tentatively ascribe this increase in the yield of the anti isomer in the presence of Eosin-Y to the increase in the triplet lifetime of 86 in the presence of the sensitizer. As a result, the triplet state of 86 may be able to exit the cavity and undergo reaction in free solution. Building on this concept, the general ability of the OA 2 dimer to control photo-dimerization reactions has been illustrated with methyl cinnamates, p-methyl styrene, indene, and 4,4-dimethylcyclohex-2-enone.⁶⁶

The familiar photo-Fries reaction has also been studied within the confines of the dimer capsule of OA 2.67 In organic solvents the Photo-Fries reaction can produce a bewildering array of products. For example, the naphthyl esters examined by the authors (Scheme 11) gave as many as nine products depending on the nature of the R group. However, in the confines of the

Scheme 11 Photochemical products from the photolysis of 87-89 encapsulated within the OA 2 dimer.

capsule formed by OA 2 the three esters examined, 87-89 all gave quantitative yields of their respective rearranged products 90-92. As the authors noted, such selectivity in the Photo-Fries reaction had only been observed in zeolites, and suggests little translational and rotational freedom for the excited singlet state and subsequent β-cleavage process.

A fascinating example of the control of a photochemical reaction is the selective oxidation of olefins by singlet oxygen within the OA 2 capsule.⁶⁸ Alkenes with multiple allylic positions normally react indiscriminately with singlet oxygen via addition to the double bond and simultaneous abstraction of an allylic hydrogen. However, when alkenes such as 1-methyl cyclohexene were exposed to solutions of OA 2 a 2:2 hostguest complex was spontaneously formed in which each guest adopted a specific orientation with its methyl group filling the narrow polar region of the inner space (Fig. 20). As a result, only the C-3 allylic position is accessible via partial opening of the capsule (see Section 2.1.1.d).

In this system singlet oxygen was generated using two sensitizers: water-soluble Rose Bengal (Fig. 20), and water insoluble **Review Article**

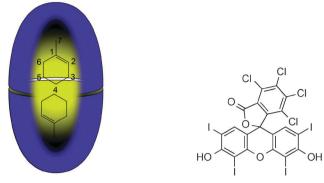


Fig. 20 Left: orientation of 1-methyl cyclohexene with the capsule formed by OA 2. Right: structure of Rose Bengal.

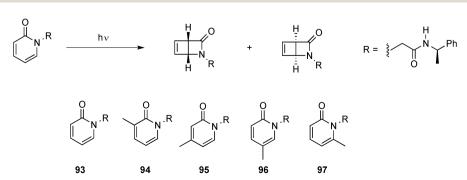
dimethyl benzil (30, Fig. 9). The latter was itself made watersoluble by encapsulation within an OA 2 capsule. Although mixing of the capsular complexes of substrate and sensitizer 30 did not result in the formation of any hetero-guest complex, irradiation at 500 nm successfully oxidized the encapsulated substrate to give a 95% yield of the product peroxide arising from C-3 abstraction. Elucidation of the mechanism for this conversion began with an examination of the encapsulated dimethyl benzil 30 on its own. Thus, lifetime measurements of singlet oxidation generated by this capsule were 5 and 41 μs in water and D₂O respectively, indicating that singlet oxygen spends most of its time in free solution. Next, the rate constant for oxygen induced quenching of the triplet excited state of encapsulated 30 was determined. Two different approaches gave very similar results; the rate was approximately one order of magnitude slower than diffusion, but faster than throughthe-wall energy transfer observed with hemicarcerands. 69 These results suggest that energy transfer from the encapsulated (triplet) guest to oxygen involves partial opening of the container to allow oxygen to enter the capsule, a mechanism facilitated by the relatively long lifetime of the excited sensitizer. Indeed, as was discussed in Section 2.1.1.d, sensitizers with much shorter lifetimes were only slightly affected by the presence of oxygen in solution. Based on this information, the overall mechanism proposed involved: (1) generation of triplet encapsulated 30, (2) capsule breathing to allow contact between oxygen and excited 30 and subsequent energy transfer; (3) exit of singlet oxygen from the capsule into bulk solution; and (4) entry of singlet oxygen

into the capsule containing the alkene and regioselective oxidation.

Two papers detailing chiral photochemistry within the capsule formed by OA 2 have also been reported. In the first, the 4e disrotatory cyclization of pyridones was examined (Scheme 12). At its most stereochemically simple level this reaction produces two enantiomers, but the addition of a chiral R group to the N-atom leads instead to two diastereomeric products. Two related questions raised by such conversions are: (1) even though the chirality of the R group is quite distant from the developing stereocenters, can packing within the capsule formed by OA 2 promote high diastereoselective reaction? (2) How do other groups around the pyridine ring influence reaction outcome?

The described research focused on the five pyridones 93-97. NMR revealed that all formed 2:1 host-guest complexes in the presence of excess OA 2, with the methyl group of 96 exhibiting the largest shift from the free state indicating that it anchors deeply into the polar region of one of the hemispheres of the capsule. These studies also revealed that the molecules adopt a pole-to-pole orientation with free rotation around the long polar axis, but no flipping of the guest on this axis (Section 2.1.1.e). Presumably, in the dry environment of the capsule a hydrogen bond can exist between the pyridine carbonyl and the N-H of the chiral amide sidechain (Fig. 21). Photochemical reaction of these pyridones in free acetonitrile solution resulted in very low diastereoselectivity (diastereoselectivity ratio (dr) < 52:48), presumably because the chiral center in each example cannot transmit its stereochemical information over the relatively long distance between it and the developing stereocenters. On the other hand, within the capsule good to excellent diastereoselectivity was observed, especially at 278 K. Thus dr values ranged from $\sim 60:40$ in the case of 93, to as high as 96:4 in the case of 94; contrary to what was observed in solution, the confining effect of the capsule did allow the stereochemical information of the chiral center to be transmitted to the developing centers.

A second paper detailing chiral photochemistry in the capsule formed by OA 2 concerned the encapsulation of tropolone ethers such as 98 and cyclohexadienones such as 99. The former undergoes a 4e disrotatory cyclization to form [3.2.0]bicycles, whilst the latter undergoes an oxa-di-π-methane rearrangement to form [3.1.0]bicycles (Scheme 13). NMR analysis revealed that both guests form 2:1 host-guest complexes with OA 2 dimer,



Scheme 12 The 4e disrotatory cyclization of chiral pyridones 93-97

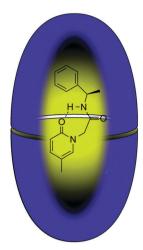


Fig. 21 Proposed binding motif of pyridine 96

with their respective 7- and 6-member rings occupying one hemisphere, and their chiral side chains occupying the other. In free solution both molecules gave little or no diastereoselectivity (dr \sim 50:50), but as anticipated, higher diastereoselectivity when encapsulated within the host dimers. Thus tropolone ether 98 and cyclohexadienone 99 gave diastereomeric ratios of their respective compounds of $\sim 68:32$ (at 278 K).

It is not only stereochemical information within a molecule that can be enhanced or biased within the confines of containers, but conformation information too. Cyclohexylphenyl ketones possess established conformational-dependent photochemistry in solution, and were therefore prime targets to examine within the nano-scape of the OA 2 dimer.⁷⁰ Cyclohexylphenyl ketones undergo Type I and Type II Norrish photochemistry (Scheme 14). The Norrish type I reaction is independent of the conformational preference of the cyclohexyl group, whereas the Norrish Type II reaction that occurs in these compounds is dependent on the position of the C-3 hydrogens in relation to the excited phenylacyl carbonyl chromophore. Thus, only when the phenylacyl group is axial are these hydrogens on the same side of the 6-membered ring within reach of the carbonyl group. Two cyclohexylphenyl ketones were examined: 100 (R = H in Scheme 14) and 100 (R = Me), both of which formed the expected 2:1 host-guest complexes with OA 2. A comparison of the photochemistry of these two ketones in free acetonitrile and encapsulated within the capsule revealed that 100 (R = H) gave mostly the Type II cyclobutanol product in free solution and only the Type I products when encapsulated, whereas 100 (R = Me) gave

98

$$hv$$
 hv
 hv

Scheme 13 The photochemistry of tropolone ethers such as 98 and cyclohexadienones such as 99.

Scheme 14 Type I and Type II Norrish photochemistry of cyclohexylphenyl ketones 100 (R = H) and 100 (R = Me)

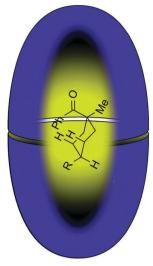


Fig. 22 Proposed binding motif of cyclohexylphenyl ketones based on reaction outcome and computer models.

exclusively the cyclobutanol in free solution and the Type I products when encapsulated. The initial conclusion of these results could be that within the capsule the conformation preference of the 6-membered ring is with an equatorial phenylacyl group whose carbonyl cannot react with the C-3 hydrogens and so Type II chemistry is switched off. However, ¹H DQF COSY NMR analysis revealed that in both free solution and within the capsule it is the conformer with the axially positioned phenacyl group that is preferred. Why no Type II chemistry? Molecular dynamics simulations of the complexes formed by 100 (R = H) and 100 (R = Me) reveal that although the phenylacyl group is oriented axially, its carbonyl group is turned away from the 6-membered ring. Thus, assuming a similar packing of the excited state, the carbonyl group is seldom in the position to react with the C-3 hydrogen atoms and form Type II products (Fig. 22).

The behavior of carbenes generated within the OA 2 host has recently been investigated (Scheme 15).⁷¹ A range of carbenes generated from diazirines 101-104 were studied, with initial investigations focused on the types of complex formed between guests 101-104 and OA 2. From a combination of ¹H and diffusion NMR studies the authors concluded that 101 formed a 2:2 host-guest complex, but that guests 102-104 formed strong (ITC) 1:1 complexes.

In free solution, irradiation of 101-104 gave predominantly ketone 106 and alcohol 107 in a combined yield of > 90%. In contrast, irradiation of the corresponding complexes generated different products depending on the guest. In the case of 101, the 2:2 complex led to an 89% yield of azine 105 (R = H). On the other hand, the 1:1 complex of 102 in OA gave a 92% yield of insertion products 108 and 109 (R = OH) and 8% 106 (R = OH), while the host-guest complexes with 103 and 104 gave 55 and 63% of 106 (R = Cl and Br respectively), and a combined 45% and 37% of the insertion products 108 and 109 (R = Cl and Br respectively). The difference between the 1:1 complex of 102, and the 1:1 complexes of 103 and 104 was rationalized with the

Scheme 15 Diazirines reactivity within OA 2.

aid of molecular dynamics simulations. Thus, the preferred orientation of 102 in the cavity of OA is with the diazirine group pointing down into the base of the pocket of the host. The corresponding carbene has little reaction options but to therefore undergo insertion reactions. On the other hand, the adamantanes 103 and 104 were predicted to bind with the diazirine group pointing up. Such an orientation allows the resulting carbene to readily react with water and generate a significant amount of the ketone 106.

Whilst compartmentalization can radically alter photochemistry it can also stop photophysical or photochemical processes normally observed in solution. An excellent example comes in the form of the entrapment of naphthalene 110, anthracene 111, and tetracene 112 (Fig. 23) within the dimer formed by OA 2.38 The smallest of these guests, naphthalene 110, shows only monomer emission in aqueous solution at 10⁻⁴ M, but at the same concentration in the presence of the host shows a red-shifted monomer and an excimer emission. NMR spectroscopy reveals why this is so: the guest forms a 2:2 complex with OA 2, and with two guests inside the capsule they experience an effective concentration of 3 M. Contrastingly, in solution tetracene 112 normally undergoes facile photo-induced dimerization and shows both monomer and excimer emission. The capsular complex of this guest however only demonstrates monomer emission and does not undergo guest dimerization

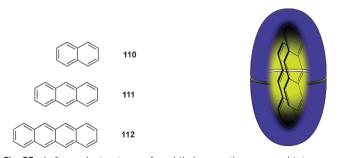


Fig. 23 Left panel: structures of naphthalene, anthracene and tetracene (110-112). Right panel: proposed binding motif of anthracene 111 based on NMR evidence and photolysis experiments.

because the size and shape of the guest is such that only one can fit within the capsule formed by OA 2. Between these two examples is the interesting case of anthracene 111. Irradiation of solutions of 111 normally leads to rapid and quantitative formation of anthracene dimer, and as a result of the speed and efficiency of this reaction only the weakest of excimer emissions is normally observed along with monomer emission. NMR reveals that this guest forms a 2:2 complex with OA 2, but rather than promoting dimerization, none is observed. However, there is a strong excimer emission band in the fluorescence spectrum of the complex. Close scrutiny of the ¹H NMR spectrum of the complex solved this puzzle. The guests are held out of register with respect to each other (Fig. 23), and in this relative position the two pairs of 9,10 atoms cannot approach each other. As a result, extensive irradiation leads to excimer emission but no dimerization. As might be anticipated, this unusual situation has been shown to be independent of the coating of the cavitand.²²

The capsules formed by OA 2 also allow thicketones to phosphoresce at room temperature.⁷² Phosphorescence of this class of molecule is normally minimal at best because they demonstrate strong diffusion limited self-quenching as well as diffusion controlled oxygen quenching. Furthermore, singlet oxygen generated by this process reacts with thioketones irreversibly. To examine the effects of encapsulation on thioketones three guests were chosen: camphorthione 113, fenchthione 114 (Fig. 24) and adamantanethione 49 (Fig. 15). Guests 113 and 49 are illustrative (Fig. 24, right).

¹H, NOESY and diffusion NMR analysis revealed that these two guests have subtly different binding motifs within the capsule. Thus camphorthione 113 binds preferentially with its thioketone groups buried in the deep, 'polar' region of the nanospace, whereas adamantanethione 49 binds in the opposite manner with the two thioketone groups of the guests at the equatorial region of the capsule (Fig. 24).

Guest 113 demonstrated phosphorescence at a capsule concentration of 10⁻⁴ M, and possessed a relatively long lifetime in the capsule (conc. 10^{-5} M), exceeding its lifetime at infinite dilution in perfluorodimethylcyclohexane. Its enhanced fluorescence and lifetime suggests that no self-quenching of the guests

occurs, even though there are two per capsule with an effective molar concentration in the nano-space. This was attributed to the preferred orientation of the guest within the capsule that inhibits self-quenching and protects the guest from dissolved oxygen. Indeed, when the rate constant for oxygen quenching was measured it was found to be two orders of magnitude lower than in common solvents. Considering the orientation of adamantanethione 49 within the capsule it may be anticipated that the complex demonstrates no phosphorescence. However, this is not the case. Although weaker than the complex with 113, the adamantanethione 49 possessed phosphorescence much greater than that observed at infinite dilution. Interestingly the lifetime of the phosphorescence from this complex was shorter than that at infinite dilution. Apparently the two guests are oriented relative to one another such that they can partially self-quench and reduce their lifetimes, but cannot adopt the relative orientations attainable in free solution that bring about nearly complete self-quenching. Overall, the ability of the capsule formed by OA 2 to enhance the phosphorescence of these guests (and in capsules formed by related cavitands²²) was found to be superior to that of comparably sized, but more open, cucurbit[7]uril.

The photophysics and photochemistry discussed thus far has assumed that the capsule formed by OA 2 is an inert container akin to a nano-scale round-bottomed flask. However, very recent results have demonstrated for the first time that this is not actually the case.⁷³ Thus in their excited state the capsules formed by OA 2 and related cavitand 8 have been shown to be good triplet energy donors to ground state guests, and in their ground state have been shown to be electron donors to excited state species. To investigate the possibility that these hosts can act as triplet sensitizers guests such as 115 - that form distinct products depending on the type of excitation - were included inside the capsule (Scheme 16). Guest 115 formed a strong, structurally well-defined 2:2 host-guest complex with OA 2, and irradiation of this complex led smoothly to a quantitative yield of the triplet product 116. Guest 115 did likewise with the capsule formed by 8, but in this instance the transformation to 116 was considerably slower than with OA 2. With either host there was no evidence of the singlet product 117.

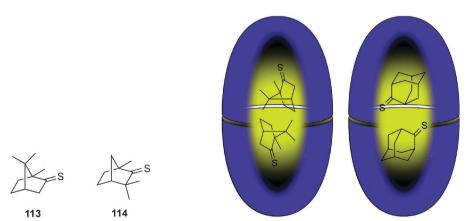


Fig. 24 Left panel: structures of camphorthione 113 and fenchthione 114. Right panel: proposed binding motif of camphorthione 113 and adamantanethione 49 based on NMR evidence and photophysics.

Scheme 16 Singlet and triplet photochemistry of 115

Benzoate groups are known triplet sensitizers, and so these results strongly support the notion that the benzoate groups of the hosts are excited to the singlet state, undergo intersystem crossing, and transfer their energy to the encapsulated guest; in the case of OA 2, the more local benzoate groups at the rim of the host are more efficient at transferring their energy than the more distal benzoates in the pendant groups of 8.

The OA 2 capsule is also capable of donating electrons to excited electron acceptors. To take one example, the fluorescence of acridinium 118 was quenched upon the addition of OA 2, a quenching that was shown to be static rather than dynamic, with a sub-nanosecond rate constant. Since water-soluble cationic guests are known to associate with the outside of the host (Section 2.1.1.f), and considering the higher S₁ energies of OA 2 versus 118, the observed fluorescence quenching was unlikely to arise from singlet-singlet energy transfer. Considering the likely oxidation potentials of OA 2 and 8, the authors attributed the loss of fluorescence of excited guests such as 118 to electron transfer.

There are also rare examples of reactions occurring between the OA 2 capsule and an encapsulated guest. In this regard it is worth highlighting a paper that, although on the edge of the purview of this review, is of significant note⁷⁴ Specifically, it has recently been noted that the 1 and 2-azidoadamantanes 119 and 120 (Fig. 25) bind to OA 2 to form the corresponding 1:1 complexes. Irradiation of these complexes results in the generation of the corresponding nitrene complexes, and this reactive guest then inserts into one of the four benzal hydrogens of the host to generate 'cavitands' such as 121 (Fig. 25).

And on the topic of photochemical insertion into the capsule formed by OA 2 we turn to the last example on the topic of photoreactions in these cavitand dimers, the release of encapsulated guests via a photo-trigger mechanism.⁷⁵ In this work, a series of p-methoxyphenacyl esters 122 (R = various) were encapsulated with the OA 2 dimer. In each case the corresponding 2:1 host-guest complex was formed. ¹H NMR analysis demonstrated that irradiation of these complexes fragmented the guest which in turn led to the breakdown of the supramolecular capsule. Each guest produced between three and five products, primarily the p-methoxy acetophenone and the corresponding carboxylic acid (Scheme 17), but also in some cases the insertion product where the p-methoxy acetophenone moiety inserted into a benzal C-H bond of the host (cf. 121). The wide range of products observed suggested to the authors that more than one mechanism for the decomposition of the guest might be operational. Because the host is a good triplet sensitizer (vide supra) it was suggested that the triplet excited state of the bound guest was central to its decomposition. However, further work is required to fully understand this system.

(c) As tools for probing fluorescent protein chromophores. The chromophores of fluorescent proteins exhibit markedly different fluorescence properties when sequestered inside a β-barrel of a protein compared to those observed in free solution.

121

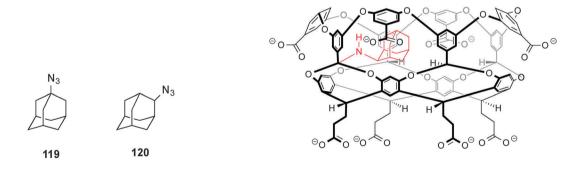


Fig. 25 Left panel: structures of azidoadamantanes 119 and 120. Right panel: structure of insertion product 121 from reaction between OA 2 and a bound nitrene.

Scheme 17 Primary photochemical outcome of 122 encapsulated within the dimer of OA 2

The effect of sequestration is generally attributed to the control of the overall shape of the 4-(p-hydroxybenzylidene)imidazolidin-5-one chromophore. Mimics of this chromophore, e.g. 123 $(R_1 \text{ and } R_2 = \text{various})$ have been investigated to examine how changes to the dihedral angle of the benzylidene single and double bond influence their fluorescence properties. To gain further insight, a group of researchers led by Tolbert examined the encapsulation of 123 (R_1 and R_2 = various) by the OA 2 dimer capsule.76 As anticipated, the para-alkyl derivatives exhibited strong shielding of both the para-alkyl and N-alkyl group signals, suggesting that groups in this position are anchored at the polar region of the capsule. However, neither para- nor meta-substituents were efficient at ameliorating internal conversion of the guest. In contrast, ortho-substituents led to a one order of magnitude increase in the fluorescence quantum yield relative to benzene solution.

$$R_1$$
 $N \sim N - R_2$

All of the 2:1 complexes of the guests examined were biased towards their trans-isomer, with photo-stationary states as high as 94:6 for the trans to cis ratio. As has been discussed above, this bias for alkene geometry reflects a more efficient filling of the cavity of the trans isomer. Interestingly, contrary to what has been observed with stilbenes, the propensity for cis to trans isomerization did not correlate with the internal conversion efficiency. Thus, the presence of an ortho-substituent in 123 attenuated internal conversion without inhibiting isomerization. The conclusion of the authors was therefore that within the capsule formed by OA 2 the ortho-substituent prevents rotation around the benzylidene single bond, decreasing internal conversion, and allowing either fluorescence or double bond isomerization. In contrast, in solution the combination of a double bond isomerization with efficient single bond rotation allows both efficient isomerization and efficient internal conversion.

More recently, the range of chromophores encapsulated within the confines of the OA 2 dimer was expanded considerably. 77 Specifically, the choice of R₁ groups was expanded to include a range of ortho-, meta-, and para-substituents covering the gamut of common electron withdrawing and donating groups. With the exception of cases where the guest had an ortho ethyl group, all the chromophores formed the expected 2:1 host-guest complexes. In each case fluorescence quantum yields were measured and fitted (linear regression analysis) to an equation based on a linear free energy relationship involving a Hammett substituent constant and a density functional theory derived volume change from replacing an H-atom in the parent with the substituent. An analysis of the resulting correlation factors from the data obtained with the chromophores in free solution revealed that electronic effects dominated the fluorescence of the different molecules in the case of meta- or para-substituents, but that steric factors as large as 12% were apparent with orthosubstituents. On the other hand, the effect of R₂ was negligible.

In addition, this analysis also revealed that there is substantial negative charge transfer to the aryl ring in the transition state for decay.

In contrast, a similar analysis for the data obtained from the chromophores encapsulated within the OA 2 dimer revealed that the remote R2 group did have a significant effect, approaching that of ortho-substituents, but that the effect of R2 decreased in the order: ortho > meta > para for the position of the R_1 group. Unusually, this analysis also revealed that as R₁ was kept constant and R₂ increased in size there was a non-monotonic relationship with quantum yield; the quantum yield increased for methyl through propyl, and then decreased with larger R2 groups. This was attributed to conformational locking of the guest with increasing size, and then an over-crowding of the capsule. Comparing the steric effects of the ortho-, meta-, and parasubstituents, guest conformational locking was more apparent with the ortho- and meta-substituents; the para-substituents could not prevent aryl group rotation. Finally, a comparison of the data from benzene with that of the encapsulated guest demonstrated that internal conversion within the capsule must require more charge transfer back to the ground state; an observation that may reflect a larger amount of twist in the guest when bound within the capsule.

(d) Cavitands and capsules on surfaces. A range of cavitands has been investigated on solid surfaces. For example, with a long-term goal of examining photochemistry on inert solid supports, the photophysical properties of complexes formed by OA 2, and cavitands 7 and 8 have been studied on silica surfaces.²² Whereas the fluorescence of 30 (Fig. 9) and phosphorescence of 113 (Fig. 24) are weak at best when adhered to silica, the corresponding adhered cavitand complexes both showed strong phosphorescence, suggesting that they remained intact upon binding to the silica. Similarly, when adsorbed onto silica the capsular complex of anthracene 111 (Fig. 23) showed strong excimer emission consistent with the stable 2:2 capsular complex observed in solution.⁷⁸ Perhaps the strongest evidence for successful capsule formation on silica came from guest 64 (Scheme 5). As has been discussed, in solution irradiation of the OA 2 complex with 64 produces a high yield of rearranged product 68, and likewise complexes of this guest with OA 2, 7 and 8 absorbed onto silica gave comparable yields of this benzyl phenyl ketone.

The unusual optical and electronic properties of gold nanoparticles have led to much interest in their potential biomedical and opto-electronic applications. With this in mind, the Ramamurthy group has synthesized a number of amine-coated and thiol-footed cavitands for functionalizing gold nano-particles.⁷⁹ Nano-particles coated with the former did not behave well and will therefore not be discussed here, but within this thiol-group several cavitands were studied in aqueous solution. Hosts 124 and 125 are representative examples.

Gold particles were synthesized with 124 on their surface and characterized with a combination of Transmission Electron Microscopy (TEM), UV-vis absorption, IR, ¹H NMR, Dynamic Light Scattering (DLS), and Thermogravimetric Analysis (TGA).

All of these techniques indicated successful inclusion of the cavitands onto the gold surface. Regarding host-guest complexation, although hampered by the expected signal broadening NMR studies confirmed that guests could form 1:1 host-guest complexes with the bound cavitands. Furthermore, with guests such as 30 (Fig. 9) NMR suggested that capsular complexes were formed on the surface of the gold. Taking this one step further, a mixture of gold nano-particles coated with 124, guest 30, and excess cavitand 125 in free solution, suggested a degree of hetero-capsule formation on the surface of the nano-particle. Apparently, the hydrophobic effect is in operation at the surface of these gold nano-particles and that capsular complexes similar to the ones constituting the bulk of this review can be formed.

2.2 Velcrands

Velcrands are resorcinarene-based cavitands that, because of the two-atom bridging between pairs of opposing resorcinarene oxygen atoms, can exist as either in a 'flat' (C_{2y}) kite form, or a deeper concave (C_{4v}) vase form (Scheme 18). The rather flat nature of the kite form means that it readily undergoes selfassembly to form the corresponding dimer devoid of binding properties. On the other hand, the vase form has a rich record of host-guest chemistry that goes back some three decades.⁸⁰ One prominent thrust of the Rebek group has been the development of novel water-soluble velcrands. The majority of this work has focused on solution studies, but as we discuss below some amphiphilic velcrands have also been studied having been imbedded into micelles or membranes. We highlight these studies where they are carried out in conjunction with solution work. Readers should note in passing though that papers describing micelles composed of amphiphilic velcrands,81 and papers centered solely on micelle bound velcrands⁸² have also appeared in the literature.

An early example of a water-soluble velcrand 126 (Scheme 19) was synthesized in three steps from the octa-nitro derivative 127; itself available from bridging the corresponding resorcinarene 128 with 1,2-difluoro-4,5-dinitrobenzene.83

The carboxylate appended host 126 was shown to exist only in the vase form under aqueous conditions, in large part because water molecules can bridge the benzimidazole groups and help seam the host together. In addition though, the 'free' host was found to contain a single adventitious molecule of THF bound to the cavity that resisted removal even by heating to 100 °C under high vacuum. This uninvited guest was however displaced in the presence of suitable alternative guests.

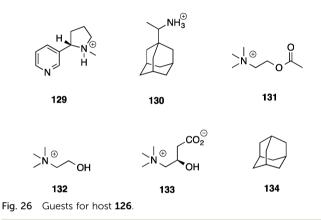
In passing it is worth noting that in further work,84 it was shown that when the solvent in the final step for the preparation of 126 (Scheme 19) was switched from THF/H2O to EtOH/H₂O, the THF-free host could be isolated. However, ¹H and DOSY NMR, as well as electrospray MS demonstrated that this 'free' host existed in the aqueous and gas phase as the kite-form dimer. The dimer was however quite weak; one equivalent of THF was sufficient to form the monomeric vase form. Consequently, all studies on 126 focused on the THF-containing monomeric host.

Tetraalkylammonium ions as large as tetraethylammonium were found to bind to host 126, but larger guests such as tetrapropylammonium did not. To take one successful example, tetramethylammonium bound with an association constant of $K_a = 3800 \text{ M}^{-1}$. Binding of these guests was slow on the ¹H NMR timescale, and 2D NOESY experiments revealed that the egression rate of bound tetramethylammonium was 8.2 s⁻¹. This corresponds to a relatively high energy-barrier of 16 kcal mol⁻¹. Binding was not just a question of size, shape and charge complementarity. For example, primary and secondary ammoniums of similar size to the aforementioned

Scheme 18 C_{2v} kite (left) and C_{4v} vase (right) forms of velcrands.

HO OH
$$O_2N$$
 O_2N O

Scheme 19 Synthesis of tetra-acid velcrand 126. Reagents are: (1) 1,2-difluoro-4,5-dinitrobenzene Et₃N, DMF; (2) SnCl₂, HCl; (3) ethyl 3-ethoxy-3iminopropanoate; (4) NaOH, THF/H2O



tetraalkylammoniums were rejected by the host, presumably because they have much stronger hydration shells. However, larger ammoniums that could simultaneously present a relatively hydrophobic moiety for binding to the pocket and an ammonium group for the rim carboxylates, were found to bind strongly. Examples include S-nicotinium 129 and rimantadium 130 (Fig. 26). Acetylcholine 131 and choline 132 were also found to bind strongly to the host ($K_a = 14600$ and 25 900 M⁻¹ respectively), but the related L-carnitine 133 was found to only weakly bind $(K_a = 150 \text{ M}^{-1})$. This was attributed to the fact that the carboxylate of the guest is in close proximity to the negatively charged upper rim of the host. Although ITC data suggested that Coulombic forces were important in these complexations, the hydrophobic effect also plays a role. Thus, host 126 could readily extract adamantane 134 from the solid state into a D2O solution.

Preliminary investigations of the effects of buffer and salts (the Hofmeister effect) on the solubility of host 126 revealed complex and poorly understood physicochemical properties.

Trembleau and Rebek also used host 126 to demonstrate the first helical motif of an encapsulated guest. 43a A 2 mM solution of 126 and dodecyl phosphocholine (DPC, 135, Fig. 27) resulted in an ¹H NMR spectra showing individual signals for each methylene unit of the hydrocarbon chain and the terminal methyl, with upfield shifts for the proton signals of eight carbons as high as $\Delta \delta = 5$ ppm. However, the internal environment of 126 can only accommodate at most six carbons in an extended motif. As the 2 mM concentration of DPC is above its critical micelle concentration (CMC) of 1 mM, analogous experiments were conducted with sodium dodecyl sulfate (SDS, 136, Fig. 27) below its \sim 8 mM CMC. These produced similar results, with far upfield shifted signals for the methylene units and terminal methyl group. A comparison of the NMR of free and bound 135 and 136 revealed little difference between the guests indicating they bind in a similar fashion.

This data can only be readily explained by assuming the alkyl chain coils up inside the pocket of the host. This preference for the coiled motif was attributed by the authors to the release of "ordered" water molecules from the surface of the hydrocarbon chain and from the interior of the cavity. Additionally, in the coiled form more C-H bonds of the guest can form attractive interactions with the aromatic π -surfaces of the benzene moieties that line the cavity. Finally, the authors noted size complementarity exists between the dodecyl chain (126 Å^3) and the inside of the cavity (225 Å^3), a close to ideal packing coefficient of 0.56. These factors combine to overcome

Fig. 27 Helix-adopting guests for host 126.

the roughly 2.5–3.0 kcal mol⁻¹ of straining *gauche* interactions that this conformation imposes on the coiled chain.

Further examination of the NMR spectra of these complexes reveals slightly broadened bound guest peaks in the upfield region. Since the exchange process must overcome a 15–20 kcal mol⁻¹ barrier afforded by the breaking and refolding of the seam of hydrogen (deuterium) bonds that enforces the vase form of the receptor, and as the helical guest motif within the cavity should produce diastereomeric complexes, the broadening of the proton signals was attributed to time-averaging of the signals from multiple conformations.

In related work the same group looked at the interaction between SDS 136 and water soluble cavitand 126 as a function of increasing amounts of surfactant.85 As just discussed, with one equivalent of 136 the guest forms a kinetically stable complex with host 126. However, in the presence of a slight excess of guest (ca. 1.3 equiv.) the exchange rate becomes rapid relative to the NMR timescale and the upfield resonances diminish as the chemical shifts relating to the free and bound species average. With a concentration of 10 mM 136, i.e., above the CMC, the host NMR signals broaden somewhat, whilst at very high concentrations of 136 (20-40 mM) a new set of broad signals appeared. Considering the amphiphilic nature of the host, the authors hypothesized that the host was being included into micelles, and to test this diffusion (DOSY) NMR measurements were performed. The first of these indicated that host 126 was monomeric between 0.2 mM-4 mM, and in the presence of one equivalent of 136 a similarly sized 1:1 complex was observed. Further DOSY measurements on 136 alone indicated a decreasing diffusion coefficient with increasing concentration; below 3.7 mM ($D = 4.2 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$) the surfactant was essentially monomeric, whilst above the CMC there existed an equilibrium between micelle and monomer. Finally, at 40 mM SDS ($D = 1.3 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$) the apparent hydrodynamic radius of the micelle was calculated to be 15.4 Å. For the mixture of 40 mM surfactant and 1 mM 126 the diffusion coefficient of the latter was noted to have decreased to just $0.8 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$; a corresponding hydrodynamic radius of approximately 25 Å. These data indicate each cavitand is associated within a micelle composed of approximately 60 amphiphiles. Incorporated into the surfactant micelle, there was no evidence of lipid tail binding into the pocket of the host. Furthermore, the host retained the ability to bind guests from the exterior aqueous solution. For example, acetylcholine bound with an affinity of $K_a \approx 1.7 \times 10^4$.

The Rebek group have also reported on a comprehensive analysis of the binding of acyclic and cyclic hydrocarbons to host 126. ^{84,86} As just discussed, in the case of DPC 135 and SDS 136 the guest adopts a helical packing motif to maximize the number of methylene groups shielded from the outer aqueous environment. For small n-alkanes, NMR illustrated well that they do not bind in quite the same way. For the smallest guests (C_3-C_5) peaks are broad, presumably because of relatively fast exchange. However, even the larger guests (C_6-C_{12}) gave broad and/or clustered peaks on the NMR spectra indicative of coiled motifs that were rapidly tumbling within the cavity. In addition,

Fig. 28 Examples of aromatic guests for host 126.

with guest lengths too long to be accommodated within the cavity (C_9 or larger) binding was quite weak and competing THF binding prevented full complexation. Branched alkanes also bound, but with slower tumbling and broad ill-defined NMR signals.

Entropic costs with confinement were smaller for cyclic guests, and these proved to be generally stronger guests than the acyclic counterparts. Overall competitive binding studies revealed that cyclohexane was the best guest.

Small aromatic guests on the other hand are rigid and therefore cannot adopt differing conformations to optimally fill the cavity space. Consequently, these showed only weak affinity to the cavity. Larger aromatic guests did however bind. 4,4′-Dimethylbiphenyl 137 (Fig. 28) was the longest guest bound, and anthracene (111), stilbene 50 (R = H), and 1,4-di-*tert*-butylbenzene 138 also bound efficiently.

The insolubility of these aliphatic and aromatic alkanes in water precluded quantification of the thermodynamics for binding. Consequently, a series of neutral, amphiphilic guests were also chosen as guests to study by ITC. Guests included a number of cyclic and adamantyl alcohols and ketones which all bound with the neutral hydrophobic moiety inside the cavity and the polar head group at the cavity rim. Binding constants were typically $>10^5\,\mathrm{M}^{-1}$. Complexation of these guests was noted to be strongly entropically driven with only a small contribution made by enthalpy. This latter point was attributed to the absence of cation- π interactions. As expected for complexations driven by the hydrophobic effect, complexation was associated with a negative heat capacity change $\left(\Delta C_p^\circ\right)$.

Two dimensional EXSY NMR experiments were used to determine the exchange rates of the various guests. For the different guests examined egression rates varied from 1.7 to 14.6 s⁻¹, which correspond to free energies of activation ΔG^{\ddagger} from 17.2 to 16.0 kcal mol⁻¹. The similarity of these values, coupled with the concentration independence of the guest exchange rates, suggests that host 126 must fully unfold for a guest to vacate the cavity.

Velcrand **126** has been shown to act as a phase-transfer catalysts for transporting hydrophobic species into aqueous solution to bring about their reaction.⁸⁷ The reaction in question was between organic-phase maleimides (**139**) and a water-soluble thiol (Scheme 20).

139: R = various

Scheme 20 Reaction between a water-soluble thiol to a maleimide 139

Host 126 readily transferred a number of N-substituted maleimides by complexation to the R group of the substrate, whereupon the resulting complex rapidly reacted with the thiol. Owing to the charge on each product, the resulting products were poorer guests than their corresponding starting materials. Hence product inhibition was avoided and efficient catalysis seen at levels as low as 0.5 mol%. Variations on the nature of the R group in 139 highlighted several important factors necessary for efficient and rapid transfer and reaction. First, if the starting material was a poor guest and poorly soluble in water then it would undergo only very slow reaction. Second, if 139 was partially soluble in water then although the presence of 126 accelerated the reaction, the fast background rate meant low relative rate acceleration. Third, the rate of the overall process was determined by the transfer rates of the maleimide into the aqueous layer, and this in turn was linked to the binding affinity of the R group. Good complementarity led to high relative rates (assuming 139 was insoluble in water). Finally, with selected pairs of maleimides significant reaction selectivities could be achieved.

Kaifer and Rebek have studied the electrochemistry of ferrocene derivatives bound to the hydrophobic pocket of cavitand 126.88 This work showed that a 1.0 mM solution of 126 increased the solubility of ferrocene 53 (Fig. 16) from $50 \mu M$ in 0.1 M NaCl to fully soluble at like salt concentrations. As expected, NMR analysis of the mixture indicated guest inclusion within the hydrophobic pocket; an association constant of $\sim 9 \times 10^4 \text{ M}^{-1}$ was estimated based on the fact that with a 1:1 stoichiometry of host and guest there was no free guest observed by NMR, and the assumption this corresponded to at least 90% guest complexation. Similar studies with partially watersoluble guests 55 (Fig. 16) and ferrocenemethanol 140 allowed association constants to be accurately determined. Thus, 55 bound with an affinity of $K_a = 1300 \text{ M}^{-1}$, whilst 140 had an association constant of $K_a = 4800 \text{ M}^{-1}$. In all cases NMR demonstrated slow exchanging systems with the expected insertion of the ferrocene moiety into the hydrophobic cavity, and where present, the polar group exposed to the exterior environment.



The included guests were also investigated using cyclic voltammetry (CV). Of particular note is that in the presence of equimolar amounts of host 126, the voltammogram was essentially flat with no observed faradaic response. In the case of excess guest 55 and 140 (53 did not possess sufficient solubility) a cyclic voltammetric response was observed. Complete encapsulation of these guests within the dimer formed by OA 2 (Section 2.1.1.f) was previously noted to completely switch off their electrochemistry,⁵³ but this was not anticipated in the open 1:1 complex with 126. Precisely why 126 has such an effect on these guests is yet to be determined.

Fig. 29 A gated cavitand 141 and tetra-ammonium cavitand 142.

Host 141 (Fig. 29) is an interesting variation on 126. The aryl groups of tetra-acid 141 effectively act as revolving doors to the cavity and modulate both the thermodynamics and kinetics of guest binding. 89 For example, the presence of the doors means that 141 has a preference for cyclopentane over cyclohexane. For 126 the reverse was true. Correspondingly, in a competition study between these two hosts, 141 preferentially bound cyclopentane (7.5:1), but in a similar study with cyclohexane 126 outcompeted 141 by two to one. Additionally, the residency time of guests within host 141 are much longer than for host 126. Thus, EXSY NMR revealed that for cyclohexane as guest entry and egression rates are approximately two orders of magnitude slower in the case of 141.

We close on the topic of ethyl-footed velcrands with a passing note concerning tetra-ammonium cavitand 142 (Fig. 29).90 The bromide salt of this cavitand had good solubility in water, and existed solely in the kite form in aqueous solution. In contrast to 126 however, the addition of THF did not trigger the formation of the vase form. In fact, formation of the vase form required substantial amounts of a water-miscible co-solvent. Consequently, its hosting properties could not be investigated in water but in 3:1 D₂O:DMSO solutions instead.

An alternative family of water-soluble velcrands is shown in Scheme 21. The formation of carbohydrate-conjugated cavitand 143 utilized tetrahydroxy resorcinarene 3,91 and was based on earlier related examples of water-soluble velcrands. 92 The tetrakis (β-D-glucosyl) moieties of 143 was added by a late-stage azidealkyne "click" reaction between a lower-rim functionalized tetrazido cavitand and a propargyl glycoside. As with other hosts of this type, the eight amide groups at the rim help to stabilize the vase conformation. NMR evidence suggested that host 143 exists in a kite-conformation in order to minimize solvent-exposed hydrophobic surfaces. However, addition of guests induced vase conformation with concomitant complexation.

NMR studies revealed that this cavitand bound a range of water-soluble guests containing hydrophobic alkane surfaces

$$HO \qquad OH \qquad Et(O)CHN \qquad NHC(O)Et$$

$$HO \qquad R \qquad R \qquad OH \qquad I) - 8) \qquad OH \qquad R \qquad HO \qquad OH \qquad NHC(O)Et \qquad III \qquad IIII \qquad III \qquad III \qquad III \qquad III \qquad III \qquad III \qquad I$$

Scheme 21 Synthesis of tetrakis(β-p)glucosyl velcrand 143. Reagents are: (1) 1,2-difluoro-4,5-dinitrobenzene Et₃N, DMF; (2) propionyl chloride, Et₃N, THF; (3) Ra-Ni, H₂, THF. (4) Propionyl chloride, Et₃N, THF, -45 °C. (5) K₂CO₃, MeOH. (6) MsCl, Et₃N, THF, 0 °C. (7) NaN₃, DMF, 70 °C. (8) propargyl glycoside, Cu(ı)I, TBTA, THF, 40 °C. (9) K2CO3, MeOH

Examples of aromatic guests for host 143.

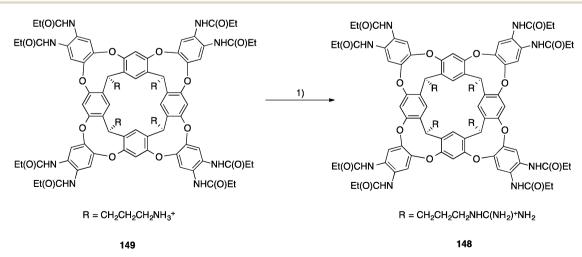
including: cylcohexanesulfonate 144, (1R)-(—)-10-camphorsulfonic acid 145, quinuclidine hydrochloride 146, and adamantylamine hydrochloride 147 (Fig. 30). EXSY NMR experiments revealed for **146** a guest dissociation barrier of 13.6 kcal mol⁻¹ at 37 °C. However, under strongly basic conditions the neutral form of this guest did not bind. Similar studies have also been performed for a velcrand with tetraethylene glycol pendant groups possessing pH-independent water solubility.93

When exposed to SDS micelles host 143 was successfully incorporated (DOSY NMR). Within the micelle the host adopted the vase form and could bind small molecules such as 146. Other molecules that did not bind to the host in free solution were also bound by 143 when it itself was within the micelle,

the interpretation being that the absence of any dimeric kite form in the micelle enhanced the binding properties of the host. Guest 146 also dissociated more slowly from the micelleassociated caviplex, with a barrier of 14.4 kcal mol⁻¹ at 37 °C.

Interestingly, the binding properties of 143 remained even after dissolution in human serum. Furthermore, in a subsequent report it was noted that this host could bind and detect guests 146 and 147 (a FDA approved drug for influenza and Parkinson's disease) in spiked samples of human urine. 94 In lyophilized human urine reconstituted with deuterium oxide, 143 existed as a soluble dimer velcrand, i.e., no endogenous constituents of human urine can occupy the binding site of the host and enforce the vase form. Unfortunately, the detection limit for this technique was in the millimolar range and therefore of limited application.

The Rebek group also showed the ability of a guanidinium-footed cavitand 148 to incorporate in dodecylphosphocholine (DPC, 135) micelles. 95 The host was prepared in one step from tetra-amine 149 (Scheme 22), itself available in ten steps from commercially available materials. 92b Cavitand 148 was soluble up to 2.0 mM in D2O and adopted the characteristic non-binding kite form. In contrast, the vase form was only observed in solutions of acetone.



Scheme 22 Synthesis of the octa-amide, guanidinium footed velcrand 148. Reagents are: (1) 1-H-pyrazole-1-carboxamidine hydrochloride, i-Pr₂NEt, CHCl₃.

In aqueous solution, 148 modestly bound a variety of cationic and neutral guests, in each case with the nonpolar group of the guest buried deep within the pocket. One possibility for the relatively weak binding was the lack of any unoccupied vase form of 148. Thus, for a guest to bind to the host it must first overcome the dimerization of the kite form of the host. Although functioning as a cation and neutral guest binder, the addition of anionic species induced precipitation of the host.

When 148 was included in a solution containing dodecylphosphocholine (DPC, 135) micelles (20 mM), the NMR spectrum showed signals characteristic of the vase form and an additional upfield signal at -0.4 ppm. ROESY NMR studies showed resonance between this peak and the trimethylammonium group of 135 and not the nonpolar tail group, suggesting that the host was located near the surface of the micelle. Approximately 25% of the host was occupied by lipid. Experiments below the critical micelle concentration of 135 showed no appreciable binding between 148 and either terminal of the phospholipid, suggesting that preorganization of the vase form by the micelle enhances the binding ability of 148. As such, 148 serves as both a guest to the micelle and host to the lipid head-group.

Molecular dynamics simulations on the incorporation of 148 into a 65-molecule DPC micelle were then carried out. The simulation began with 148 at the interface, but over time it submerged itself into the micelle to the point that its hydrophobic portions were surrounded by alkyl chains of 135 whilst its open-end remained accessible to bulk solvent. Subsequent to this, as observed by NMR, an ammonium group from the phospholipid bound into the pocket of the cavitand.

Binding studies using the micelle-incorporated host demonstrated that all of the guests tested in aqueous solution still bound to 148. In addition, guest binding was much stronger, with as much as a 230-fold increase in K_a . Van't Hoff plots for the case of 146 binding to the cavitand, in both free solution and within the micelle, revealed a favorable enthalpy of binding in both cases ($\Delta H = -3.0$ and -8.1 kcal mol⁻¹ respectively). The difference between the ΔH for complexation in water *versus* the micelle was tentatively attributed to the greater preorganization of the host in the latter. These studies also revealed a small entropic penalty of $\Delta S = -8.4$ and -9.7 cal K⁻¹ mol⁻¹ in water and DPC micelles respectively. Interestingly, other related cavitands usually give strongly positive ΔS values in water. Because the NMR spectra of 148 showed very well resolved signals, this difference was tentatively attributed to a lack of aggregation of host 143. The more negative entropy of formation in the presence of the micelle is, as the authors noted, consistent with the reduction of the degrees of freedom of the micelle-bound complex.

A variation on velcrand 149 is the chiral derivative 150.96 This host possesses a unique benzimidazole bridge with an appended carboxylic acid group that dangles into its binding pocket. Consequently it can form a hydrogen bond/salt bridge with amine guests that bind to the pocket. In the absence of a suitable guest 150 adopts a kite form, but the addition of a suitable binder such as quinuclidine hydrochloride 146 results in the vase form. The association constant of this guest is over two orders of magnitude larger than that between 146 and 149.

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This corresponds to an enhancement of -2.7 kcal mol⁻¹, a stabilization attributed in large part to the buried hydrogen bond between host and guest. A more rigorous determination using a free-energy cycle gave a value of this hydrogen bond of -3.0 kcal mol⁻¹. EXSY NMR studies suggested that the introverted carboxylic acid also affected the rate of guest exchange. An energy barrier for guest egression from 150 was measure at 19 kcal mol⁻¹ in the case of guest **146**, a full 2 kcal mol⁻¹ more than for host 149. Regardless, this relatively high barrier strongly supports the common guest exchange mechanism seen with these type of velcrands: such a mechanism requires a change from the vase to kite form costs (ca. 10-12 kcal mol^{-1}), plus the rupture of the seam of hydrogen bonds (5-7 kcal mol⁻¹).

The Hooley, Cheng, and Rebek groups have examined the immobilization of cavitands on gold substrates for protein recognition.⁹⁷ This study centered round cavitand 151, which by virtue of its thioether-functionalized feet could be readily appended onto gold surfaces. A combination of Surface Plasmon Resonance (SPR) and Surface Plasmon Resonance Imaging (SPRi) was used to study the binding properties of this host.

Et(O)CHN

O

R

R

O

NHC(O)Et

NHC(O)Et

NHC(O)Et

NHC(O)Et

R =
$$(CH_2)_{10}S(CH_2)_9CH_3$$

151

The host was first deposited as a self-assembled monolayer (SAM) onto a cleaned gold electrode by immersing the electrode in a 0.5 mM solution of 151 in n-butanol containing a 10:1 ratio of 151 and either N-octadecanethiol (ODT) or 11-mercaptoundecanoic Review Article

Fig. 31 Examples of aromatic guests for host 151

acid (MUA). The ODT or MUA were needed as fillers to enhance the predominance of the vase form of 151 and completely inhibit CV current; the SAM of 151 alone incompletely filled the gold chip and resulted in limited insulation of the electrode surface.

Focusing first on the NMe₃⁺-tagged fluorescein guest 152 (Fig. 31), it was noted that the strongest SPR response was for the SAM composed of 151 alone, but that sufficient although more modest signal could be obtained with the mixed SAM composed of 151 and ODT. Unfortunately, tests revealed that the SAM composed of just ODT also had a strong response to 152. Consequently, attention turned to the MUA filler, and as expected, monolayers of MUA alone showed no affinity for 152 whereas the mixed monolayer with 151 and MAU did.

A number of additional ammonium guests were bound to the immobilized 151, and although changes in resonance angle upon binding were modest, they were significant. To take this one step further, the authors then turned to guest binding detection using SPRi, which, because of its micro-array compatibility makes the approach amenable to large-scale screening of targets in the fields of drug discovery. Briefly, a similar sample preparation led to successful surface deposition in defined arrays, and these could be used in label-free guest binding determinations.

Finally, the group turned to protein recognition by SPR. For this they first demonstrated that the mixed monolayer with 151 and MAU successfully bound biotin derivative 153. Subsequent addition of a solution of streptavidin induced an increase in resonance angle indicative of immobilization of the streptavidin at the surface. Cross-checks demonstrate that 153 was essential for streptavidin recognition, and that no signal was observed in the absence of the cavitand. In combination these experiments proved that neither the guest nor the protein can be immobilized by MUA alone, and biosensing required the complete set of specific non-covalent interactions involving the cavitand, the biotin guest 153, and streptavidin.

Most recently the Rebek group have utilized velcrands to form capsules that form in water. As we will see, whilst hydrogen bonding is important in their formation, the hydrophobic effect also plays a key role in their formation. This program is based on velcrand 154, a host that is based on an early example of a velcrand capsule devised by De Mendoza.98 The dimeric capsule of 154 is held together by sixteen hydrogen bonds between the hydrogen bond acceptors and donor groups of the benzimidazolone rim, whilst its water-solubility arises from the eight pyridinium pendant groups.

Host 154 is synthesized in eight steps and exists in both the kite and vase forms at low concentrations. The first guests reported that confirmed capsule formation were n-undecane through *n*-tetradecane (C_{11} - C_{14}). NMR demonstrated that the guest n-undecane fits comfortably in an extended conformation, while the longer alkanes adopt gauche conformations within the capsule. Absolute binding affinities could not be determined because of the very low water solubility of these guests, but competitive studies revealed a relative order for binding affinity to be: $C_{11} > C_{12} > C_{13} > C_{14}$.

$$R_{1}$$
 R_{1} R_{1

A series of rigid molecules with subtly differing lengths, including stilbene 50 (Fig. 15) and viologen 54 (R = Me, Fig. 16), were used to probe both the length of the inner-space and the guest preferences of the 154 dimer. The initially report data suggested that uncharged guests bound more strongly that isosteric pyridinium salts.

The photophysical behavior of encapsulated stilbene derivative 50 also provided indirect evidence of the robustness of the seam of hydrogen bonds at the capsule equator. The fluorescence of 50 was strong in mesitylene but completely quenched upon encapsulation within the dimer formed by 154. This supports the idea that the capsule must be quite robust in shape, for if it were dynamic or ill-defined the host could adapt to accommodate

a planar stilbene guest and allow fluorescence. In reality, the

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seam of hydrogen bonds stoutly defines the shape of the innerspace such that the rings of a stilbene guest cannot be coplanar while inside and its fluorescence is quenched.

In a follow-up communication, the short *n*-alkanes (C₇ through C_{11}), and the appetite stimulant peptide ghrelin were respectively encapsulated and complexed by 154.99 The smallest guest n-alkanes heptane, octane, and nonane all formed the corresponding 1:1 complex with the host. These rotated rapidly in the internal hydrophobic environment created by the monomeric host and their corresponding ¹H NMR signals were broad and the spectra highly symmetrized. Interestingly, n-decane formed both a 1:1 and a 2:1 complex with the host. Tell-tail characteristics of the two complexes were: in the 1:1 complex the guest exhibited ¹H NMR signals over a narrow range indicative of gauche interactions and an overall compressed guest motif; in the 2:1 complex the same signals for the bound guest were spread out over a wide ppm range indicating an extended motif in a symmetrical environment. It was noted that the ratio of the two complexes was temperature dependent, with the capsular assembly predominating at elevated temperatures and exclusively so above 340 K.

Exposure to amphiphiles such as octanoic acid also resulted in a 1:1 complex, with as anticipated, the alkane chain imbedded in the host cavity. Similarly the peptide ghrelin - a 28 amino-acid peptide that contains an octanoyl ester moiety at the third (serine) residue - was also bound in a 1:1 manner. Similar results were obtained for rat ghrelin, a truncated 10 amino acid peptide. The binding constant of both ghrelin and rat-ghrelin were too large to determine by NMR, but ITC titrations with ratghrelin showed $K_a = 2.5 \times 10^5 \text{ M}^{-1}$, and both favorable enthalpy and entropy of binding.

Per-methylation of the benzamidazolones groups of host 154 leads to 155, a velcrand which exhibited water solubility up to 10 mM in the kite form. Host 155 was expected to have more flexible walls because whereas the vase form of 154 is stitched together by hydrogen bonds to "bridging" water molecules, this is not possible with 155. Correspondingly, the vase form of 155 could potentially hold wide guests (or wider guest motifs). It transpires that this is indeed the case.

Brief sonication and/or heating with a series of *n*-alkanes (C₆ to C₁₄) gave characteristic spectra associated with the vase form, but with noticeably compressed signal ranges for the bound-guest. Hence for the smallest guests this range spanned only about 0.6 ppm, whilst for C₁₃ and C₁₄ this range was still a relatively modest 2 ppm. As expected, for the short n-alkanes (C₆ to C₁₀) the furthest upfield shifted signals were those of the terminal methyl protons; time-averaged as they tumbled quickly on the NMR time-scale with the one end pointed to the solvent exterior and the other nestled deep within the pocket.

In contrast were the longer alkanes (C12 to C14) whose terminal methyls were unexpectedly furthest downfield on the ¹H NMR. Typically terminal methyl groups reside the deepest within the cavitand pocket and as such are shielded the most by the aromatic walls of the host. Hence they have the largest $\Delta\delta$ values with upfield shifts of 4.0 ppm or more. However, in the

case of these guests within 155 it was the central methylene protons that possessed the largest upfield shift, demonstrating that the mid-section of these guests is closest to the base of the pocket. Correspondingly, the terminal methyl group signals were the furthest downfield shifted of the bound guest region in the NMR. In combination, this data indicates that the *n*-alkanes C₁₂-C₁₄ adopt U-shaped motifs within the binding pocket of 155. Because of the small range of signals in the NMR spectra, these guests likely "writhe" in the defined space and adopt many low energy variations on the idealized U-shaped motif.

To probe this type of binding further the authors turned to a series of n-alcohols. NMR investigations confirmed that the terminal methyls reside deep within the cavity for n-octanol and *n*-nonanol (-3.4 ppm, $\Delta\delta = -4.2$ ppm), but as the carbon chain length increases the terminal methyl becomes less and less shielded. With n-decyl (C_{10}) and n-undecyl alcohols (C_{11}) the methyl signals are still the furthest upfield but have shifted significantly downfield to -3.1 ppm and -2.6 ppm, respectively, indicating the methyl is in the wider more capacious interior of the cavity. The guest adopts a J-shaped motif. For the longest homologue, tetradecanol (C14), the bound methyl signal appeared at only -0.3 ppm, suggesting that this longer guest adopts a U-shaped motif. Supporting this ¹H NMR information, COSY NMR indicated the carbon chains adopt helical and/or folded conformations within the cavity as the terminal methyl is pushed further and further out of the base of the pocket. Among other groups tested, α , ω diols also exhibited similar folding patterns.

Amphiphilic cogs

The Shionoya group has developed two water-soluble self-assemblies driven by the hydrophobic effect. The requisite amphiphiles were based on pyridine containing tris-monodentate ligands 156 (Fig. 32), which had previously been shown to self-assemble in organic media to form either cages and capsules depending on the stoichiometry of Hg²⁺ and whether or not a cryptand was also present. 100 The cages (Hg₆156₄) required no templating guest, whilst the

156:
$$R_1 = R_2 = R_3 = \text{lone pair}$$
157: $R_1 = R_2 = \text{lone pair}$
158: $R_1 = \text{lone pair}$, $R_2 = \text{lone pair}$
158: $R_1 = \text{lone pair}$, $R_2 = R_3 = \text{Me}$

159: $R_1 = R_2 = R_3 = Me$

Fig. 32 Amphiphilic cogs 156-159

Fig. 33 Guests and templates for the assembly of amphiphilic cogs 160-164.

fluorescently active capsules (Hg₆156₈) were templated by an encapsulated cryptand guest.

As a stepping-stone to working in pure water, the amphiphile 156 was shown to self-assemble in aqueous methanol to form box-shaped hexamers (156₆).¹⁰¹ Suggestive of the importance of the hydrophobic effect in this assembly, water was required to form the hexamer; the subunit was monomeric in pure methanol. Optimal assembly required the encapsulation of two templating molecules and with tribromomesitylene 160 (Fig. 33) as guest a crystal structure of the assembly was obtained. As is so often the case with templated assemblies, the nature of the template had a large influence on the success of assembly. Thus, hexamethylbenzene 161 and mesitylene 162 were shown by NMR to be good templates for this process, whereas 1,3-5trichlorobenzene 163 proved too small and hexabromobenzene 164 too big to template assembly.

Furthermore, in a subsequent publication it was revealed that when adamantane 134 was used as a template, the product formed was a tetrameric assembly 1564 with the template encapsulated within. 102 X-ray crystallography, NMR and MS were all used to characterize the assembly. Similar self-assembled inclusion complexes were also formed upon addition of pseudospherical species such as Me₄Si or CBr₄, to a solution of **156**.

Methylation of 156 gave the N-methylpyridinium amphiphiles 157-159, 103 of which 157 was insoluble in pure water whereas 158 and 159 were fully water-soluble. NMR and X-ray crystallography demonstrated that cog 158 had the predisposition to self-assembled into box-shaped hexamer 1586, and when a suitable guest molecule such as 160 was present the assembly encapsulated the guest to form the corresponding complex.

The ¹H NMR spectrum of 158 in deuterated methanol was remarkably simple showing monomeric 158 with C_{2v} symmetry, whereas the spectrum in D₂O indicated several upfield shifted proton signals indicating an aggregate. Examination of the non-equivalent N-methyl and p-tolyl signals indicated a highly symmetric box-shaped container was exclusively formed. The formation of the hexameric aggregate was confirmed by PGSE NMR measurement (log D = -9.9; $D = 1.26 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$) consistent with a 2 nm box. X-ray crystallography revealed that the stability of the assembly was in part due to interactions between electron-rich pyridyl groups sandwiched between pairs of electron-deficient pyridiniums.

The box-shaped aggregate of 1586 was shown to be remarkably stable; heating to 353 K produced noticeably sharp NMR spectra with no signs of dissociated monomer in solution. By contrast, heating aggregate 1566 in CD3OD: D2O (3:1) afforded primarily the monomeric species at only 333 K. Additionally, the critical concentration required for capsular assembly was

shown to be significantly lower than 1 µM: a 1 mM solution of 158 in MeOH showed characteristic UV absorption at 265 and 287 nm corresponding to the monomeric species, while an increasing fraction of H₂O caused a significant lessening of the absorption bands; examination of the concentration effect on absorbance in pure H₂O showed no hypochromism at all in the concentration range from 1 µM to 100 µM.

The inner space formed by 158_6 is 0.8 nm \times 0.8 nm and as such can comfortably house two copies of the guest 160 in its internal cavity; addition of water insoluble 160 to a D2O solution containing 1586 resulted in rapid internalization of the guests and downfield shifts of the ¹H NMR signals of the methyl groups from δ = 2.64 ppm (in CD₃OD) to δ = 3.49 ppm in D₂O.

In contrast to the properties of C_{2v} symmetric 158, C_{3h} subunit 159 with three pyridinium groups formed a tetrameric assembly around a templating adamantane 134 molecule. In the absence of the template 159 actually forms a 1:9 ratio of the tetramer and hexamer, but the addition of template 134 leads to the quantitative formation of the tetramer.

3.0 Conclusions and outlook

This review has discussed three classes of molecules that have been used by researchers to investigate self-assembly in aqueous solution. The use of resorcinarene-based hosts dominates the area, but all approaches demonstrate the importance of preorganization of arrays of rigid (aromatic) moieties to define a specific form predisposed to bind, fold, or assemble. Studying these molecules and their properties is: (1) teaching us how to control supramolecular chemistry in water; (2) shedding light on some dark corners of aqueous solutions chemistry, and; (3) illustrating novel applications that necessitate water being the solvent.

Regarding the first of these, supramolecular chemists are familiar with the concept of a functional group driving and/or directing binding or assembly. Thus the directionality of a hydrogen bond - a notion derived from the variance on ΔH° on the angle between donor and acceptor - may help drive a molecule to form an assembly of specific form (constitution, and relative position and orientation of subunits). The molecules discussed in this review suggest modifications to this concept, namely that: directionality can be free energy based rather than enthalpically based, and that the definition of a "functional group" can be expanded from a list useful to synthetic chemists, to include moieties useful to supramolecular chemists. In all the examples highlighted here, careful crafting of structure resulted in functional groups (to use the just expanded definition) that bind, fold, or assemble... in water.

The second and third points listed above are for the future; but we are confident much can be learned from aqueous supramolecular chemistry. Continued investigations involving the aforementioned families of molecules, and potentially new classes of molecules that are yet to be developed, will shed more light on the mysteries of water and undoubtedly engender new and useful applications. The details are certainly murky, but the bigger picture seems quite clear.

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References

- 1 C. Reichardt and T. Welton, Solvents and Solvent Effects in Organic Chemistry, VCH, New York, 4th edn, 2011.
- 2 G. V. Oshovsky, D. N. Reinhoudt and W. Verboom, Angew. Chem., 2007, 46(14), 2366-2393.
- 3 R. M. Lynden-Bell, S. C. Morris, J. D. Barrow, J. L. Finney and R. L. J. Harper, Water and Life, CRC Press, Boca Raton, 2010.
- 4 (a) G. Michal and D. Schomburg, Biochemical Pathways: An Atlas of Biochemistry and Molecular Biology, John Wiley and Sons, New York, 2nd edn, 2012; (b) P. Ball, Chem. Rev., 2008, 108, 74-108.
- 5 (a) J. Maddox, J. Randi and W. W. Stewart, Nature, 1988, 334, 287-290; (b) E. Davenas, F. Beauvais, J. Amara, M. Oberbaum, B. Robinzon, A. Miadonna, A. Tedeschi, B. Pomeranz, P. Fortner, P. Belon, J. Sainte-Laudy, B. Poitevin and J. Benveniste, *Nature*, 1988, 333, 816–818; (c) D. L. Rousseau, Am. Sci., 1992, 80, 54–63; (d) E. R. Lippincott, R. R. Stromberg, W. H. Grant and G. L. Cessac, Science, 1969, 164(3887),
- 6 (a) K. Ovchinnikova and G. H. Pollack, Langmuir, 2009, 25, 542-547; (b) H. R. Corti and A. J. Colussi, Langmuir, 2009, 25(11), 6587-6589.
- 7 G. N. Clark, G. L. Hura, J. Teixeira, A. K. Soper and T. Head-Gordon, Proc. Natl. Acad. Sci. U. S. A., 2010, 107(32), 14003-14007.
- 8 P. B. Petersen, J. C. Johnson, K. P. Knutsen and R. J. Saykally, Chem. Phys. Lett., 2004, 397(1-3), 46-50.
- 9 P. B. Petersen and R. J. Saykally, Annu. Rev. Phys. Chem., 2006, 57, 333-364.
- 10 (a) K. Lum, D. Chandler and J. D. Weeks, J. Phys. Chem. B, 1999, 103(22), 4570-4577; (b) D. Chandler, Nat. Biotechnol., 2007, 445, 831-832.
- 11 (a) J. N. Israelachvili, Intermolecular & Surface Forces, Academic Press, San Diego, 2011; (b) L. Wang, R. A. Friesner and B. J. Berne, J. Phys. Chem. B, 2010, 114, 7294-7301.
- 12 (a) R. Baron, P. Setny and J. A. McCammon, J. Am. Chem. Soc., 2010, 132(34), 12091–12097; (b) P. Setny, R. Baron and J. A. McCammon, J. Chem. Theory Comput., 2010, 6,

- 2866-2871; (c) J. Ewell, B. C. Gibb and S. W. Rick, J. Phys. Chem. B, 2008, 112, 10272-10279.
- 13 (a) S. J. Rowan, S. J. Cantrill, G. R. L. Cousins, J. K. M. Sanders and J. F. Stoddart, Angew. Chem., Int. Ed., 2002, 41, 898-952; (b) M. E. Belowich and J. F. Stoddart, Chem. Soc. Rev., 2012, 41, 2003–2014; (c) J. Sun, J. L. Bennett, T. J. Emge and R. Warmuth, J. Am. Chem. Soc., 2011, 133(10), 3268-3271; (d) D. Xu and R. Warmuth, J. Am. Chem. Soc., 2008, 130(24), 7520-7521; (e) X. Liu and R. Warmuth, J. Am. Chem. Soc., 2006, 128(43), 14120-14127.
- 14 (a) R. Warmuth, Carcerands and hemicarcerands, John Wiley & Sons Ltd., 2012, pp. 917-954; (b) A. Jasat and J. C. Sherman, Chem. Rev., 1999, 99, 932-967.
- 15 (a) M. Fujita, M. Tominaga, A. Hori and B. Therrien, Acc. Chem. Res., 2005, 38(4), 369-378; (b) J. K. Klosterman, Y. Yamauchi and M. Fujita, Chem. Soc. Rev., 2009, 38(6), 1714-1725; (c) M. Yoshizawa, J. K. Klosterman and M. Fujita, Angew. Chem., 2009, 48, 3418-3438; (d) M. D. Pluth and K. N. Raymond, Chem. Soc. Rev., 2007, 36, 161-171; (e) J. R. Nitschke, Acc. Chem. Res., 2007, 40(2), 103–112.
- 16 (a) C. L. D. Gibb, E. D. Stevens and B. C. Gibb, J. Am. Chem. Soc., 2001, 123(24), 5849-5850; (b) C. L. D. Gibb, X. Li and B. C. Gibb, Proc. Natl. Acad. Sci. U. S. A., 2002, 99, 4857–4862; (c) Z. R. Laughrey, C. L. D. Gibb, T. Senechal and B. C. Gibb, Chem. - Eur. J., 2003, 9(1), 130-139; (d) H. Xi, C. L. D. Gibb and B. C. Gibb, J. Org. Chem., 1999, **64**, 9286–9288; (e) H. Xi, C. L. D. Gibb, E. D. Stevens and B. C. Gibb, Chem. Commun., 1998, 1743-1744.
- 17 C. L. D. Gibb and B. C. Gibb, J. Am. Chem. Soc., 2004, **126**(37), 11408-11409.
- 18 S. Liu, S. E. Whisenhunt-Ioup, C. L. D. Gibb and B. C. Gibb, Supramol. Chem., 2011, 23(6), 480-485.
- 19 J. O. Green, J. H. Baird and B. C. Gibb, Org. Lett., 2000, 2(24), 3845-3848.
- 20 S. J. Rowan, D. G. Hamilton, P. A. Brady and J. K. M. Sanders, J. Am. Chem. Soc., 1997, 119, 2578–2579.
- 21 C. L. D. Gibb and B. C. Gibb, J. Am. Chem. Soc., 2006, 128(51), 16498-16499.
- 22 E. Ramasamy, N. Jayaraj, M. Porel and V. Ramamurthy, Langmuir, 2012, 28(1), 10-16.
- 23 (a) M. D. Giles, S. Liu, R. L. Emanuel, B. C. Gibb and S. M. Grayson, J. Am. Chem. Soc., 2008, 130(44), 14430-14431; (b) M. D. Giles, S. Liu, R. L. Emanuel, B. C. Gibb and S. M. Grayson, Isr. J. Chem., 2009, (49), 31-40; (c) S. M. Grayson and B. C. Gibb, Soft Matter, 2010, 6, 1377-1382.
- 24 (a) R. Kulasekharan and V. Ramamurthy, Org. Lett., 2011, 13(19), 5092-5095; (b) Y. Ishida, R. Kulasekharan, T. Shimada, S. Takagi and V. Ramamurthy, Langmuir, 2013, **29**(6), 1748–1753; (c) Y. Ishida, R. Kulasekharan, T. Shimada, V. Ramamurthy and S. Takagi, J. Phys. Chem. C, 2014, **118**(19), 10198–10203.
- 25 J. P. Da Silva, R. Kulasekharan, C. Cordeiro, S. Jockusch, N. J. Turro and V. Ramamurthy, Org. Lett., 2012, 14(2), 560-563.
- 26 H. Sun, C. L. D. Gibb and B. C. Gibb, Supramol. Chem., 2008, **20**(1&2), 141–147.

27 N. Jayaraj, Y. Zhao, A. Parthasarathy, M. Porel, R. S. H. Liu and V. Ramamurthy, Langmuir, 2009, 25(18), 10575-10586.

Review Article

- 28 C. L. D. Gibb and B. C. Gibb, J. Comput.-Aided Mol. Des., 2014, 28(4), 319-325, DOI: 10.1007/s10822-013-9690-2.
- 29 C. L. D. Gibb and B. C. Gibb, J. Am. Chem. Soc., 2011, **133**(19), 7344-7347.
- 30 S. Liu and B. C. Gibb, Chem. Commun., 2011, 47(12), 3574-3576.
- 31 C. L. D. Gibb and B. C. Gibb, Chem. Commun., 2007, 1635-1637.
- 32 C. L. D. Gibb and B. C. Gibb, Tetrahedron, 2009, 65(35), 7240-7248.
- 33 H. Gan, C. J. Benjamin and B. C. Gibb, J. Am. Chem. Soc., 2011, 133(13), 4770-4773.
- 34 H. Gan and B. C. Gibb, Chem. Commun., 2012, 48(11), 1656-1658.
- 35 H. Gan and B. C. Gibb, Chem. Commun., 2013, 49(14), 1395-1397.
- 36 (a) L. R. MacGillivray and J. L. Atwood, Nature, 1997, 389, 469-472; (b) K. Rissanen, Angew. Chem., Int. Ed., 2005, 44, 3652-3654.
- 37 J. Ewell, B. C. Gibb and S. W. Rick, J. Phys. Chem. B, 2008, 112(33), 10272-10279.
- 38 L. S. Kaanumalle, C. L. D. Gibb, B. C. Gibb and V. Ramamurthy, J. Am. Chem. Soc., 2004, 126(44), 14366-14367.
- 39 M. Porel, N. Jayaraj, L. S. Kaanumalle, M. V. S. N. Maddipatla, A. Parthasarathy and V. Ramamurthy, Langmuir, 2009, 25(6), 3473-3481.
- 40 N. Jayaraj, S. Jockusch, L. S. Kaanumalle, N. J. Turro and V. Ramamurthy, Can. J. Chem., 2011, 89(2), 203-213.
- 41 H. Tang, C. S. de Oliveira, G. Sonntag, C. L. D. Gibb, B. C. Gibb and C. Bohne, J. Am. Chem. Soc., 2012, 134(12), 5544-5547.
- 42 R. Kulasekharan, N. Jayaraj, M. Porel, R. Choudhury, A. K. Sundaresan, A. Parthasarathy, M. F. Ottaviani, S. Jockusch, N. J. Turro and V. Ramamurthy, Langmuir, 2010, 26(10), 6943-6953.
- 43 (a) L. Trembleau and J. Rebek, Jr., Science, 2003, 301(5637), 1219-1220; (b) A. Scarso, L. Trembleau and J. Rebek, Jr., J. Am. Chem. Soc., 2004, **126**, 13512–13518; (c) D. Ajami and J. Rebek, Jr., J. Am. Chem. Soc., 2006, 128(47), 15038–15039.
- 44 R. Choudhury, A. Barman, R. Prabhakar and V. Ramamurthy, J. Phys. Chem. B, 2013, 117(1), 398-407.
- 45 M. Furuhashi and G. S. Hotamisligil, Nat. Rev. Drug Discovery, 2008, 7, 489-503.
- 46 Y. H. Ko, H. Kim, Y. Kim and K. Kim, Angew. Chem., Int. Ed., 2008, 47, 4106-4109.
- 47 S. Liu, D. H. Russell, N. F. Zinnel and B. C. Gibb, J. Am. Chem. Soc., 2013, 135(11), 4314-4324.
- 48 M. Porel, N. Jayaraj, S. Raghothama and V. Ramamurthy, Org. Lett., 2010, 12(20), 4544-4547.
- 49 J. Y.-C. Chen, N. Jayaraj, S. Jockusch, M. F. Ottaviani, V. Ramamurthy and N. J. Turro, J. Am. Chem. Soc., 2008, **130**, 7206-7207.
- 50 S. Jockusch, O. Zeika, N. Jayaraj, V. Ramamurthy and N. J. Turro, J. Phys. Chem. Lett., 2010, 1(18), 2628–2632.

- 51 S. Jockusch, M. Porel, V. Ramamurthy and N. J. Turro, J. Phys. Chem. Lett., 2011, 2(22), 2877-2880.
- 52 S. Gupta, A. Adhikari, A. K. Mandal, K. Bhattacharyya and V. Ramamurthy, J. Phys. Chem. C, 2011, 115(19), 9593-9600.
- 53 D. Podkoscielny, I. Philip, C. L. D. Gibb, B. C. Gibb and A. E. Kaifer, Chem. - Eur. J., 2008, 14(15), 4704-4710.
- 54 D. Podkoscielny, S. Gadde and A. E. Kaifer, J. Am. Chem. Soc., 2009, 131(36), 12876-12877.
- 55 Y. Qiu, S. Yi and A. E. Kaifer, Org. Lett., 2011, 13(7), 1770-1773.
- 56 M. Porel, S. Jockusch, A. Parthasarathy, V. J. Rao, N. J. Turro and V. Ramamurthy, Chem. Commun., 2012, 48(21), 2710-2712.
- 57 M. Porel, A. Klimczak, M. Freitag, E. Galoppini and V. Ramamurthy, Langmuir, 2012, 28(7), 3355-3359.
- 58 M. Porel, C.-H. Chuang, C. Burda and V. Ramamurthy, J. Am. Chem. Soc., 2012, 134(36), 14718-14721.
- 59 S. Liu, H. Gan, A. T. Hermann, S. W. Rick and B. C. Gibb, Nat. Chem., 2010, 2(10), 847-852.
- 60 D. M. Vriezema, M. Comellas Aragones, J. A. Elemans, J. J. Cornelissen, A. E. Rowan and R. J. Nolte, Chem. Rev., 2005, **105**(4), 1445–1489.
- 61 (a) A. K. Sundaresan and V. Ramamurthy, Org. Lett., 2007, 9(18), 3575–3578; (b) A. K. Sundaresan and V. Ramamurthy, Photochem. Photobiol. Sci., 2008, 7(12), 1555-1564.
- 62 R. Kulasekharan, M. V. S. N. Maddipatla, A. Parthasarathy and V. Ramamurthy, J. Org. Chem., 2013, 78(3), 942-949.
- 63 (a) A. Parthasarathy, L. S. Kaanumalle and V. Ramamurthy, Org. Lett., 2007, 9(24), 5059-5062; (b) A. Parthasarathy and V. Ramamurthy, Photochem. Photobiol. Sci., 2011, 10(9), 1455-1462.
- 64 S. R. Samanta, A. Parthasarathy and V. Ramamurthy, Photochem. Photobiol. Sci., 2012, 11(11), 1652-1660.
- 65 L. S. Kaanumalle and V. Ramamurthy, Chem. Commun., 2007, 1062-1064.
- 66 A. Parthasarathy, S. R. Samanta and V. Ramamurthy, Res. Chem. Intermed., 2013, 39(1), 73-87.
- 67 L. S. Kaanumalle, C. L. D. Gibb, B. C. Gibb and V. Ramamurthy, Org. Biomol. Chem., 2007, 5(2), 236-238.
- 68 A. Natarajan, L. S. Kaanumalle, S. Jockusch, C. L. D. Gibb, B. C. Gibb, N. J. Turro and V. Ramamurthy, J. Am. Chem. Soc., 2007, 129, 4132-4133.
- 69 (a) F. Pina, A. J. Parola, E. Ferreira, M. Maestri, N. Armaroli, R. Ballardini and V. Balzani, J. Phys. Chem., 1995, 99, 12701-12703; (b) I. Place, A. Farran, K. Deshayes and P. Piotrowiak, J. Am. Chem. Soc., 1998, 120, 12626-12633.
- 70 R. Kulasekharan, R. Choudhury, R. Prabhakar and V. Ramamurthy, Chem. Commun., 2011, 47(10), 2841-2843.
- 71 S. Gupta, R. Choudhury, D. Krois, G. Wagner, U. H. Brinker and V. Ramamurthy, Org. Lett., 2011, 13(22), 6074-6077.
- 72 N. Jayaraj, M. V. S. N. Maddipatla, R. Prabhakar, S. Jockusch, N. J. Turro and V. Ramamurthy, J. Phys. Chem. B, 2010, **114**(45), 14320–14328.
- 73 P. Jagadesan, B. Mondal, A. Parthasarathy, V. J. Rao and V. Ramamurthy, Org. Lett., 2013, 15(6), 1326-1329.
- 74 R. Choudhury, S. Gupta, J. P. Da Silva and V. Ramamurthy, J. Org. Chem., 2013, 78(5), 1824–1832.

- 75 N. Jayaraj, P. Jagadesan, S. R. Samanta, J. P. Da Silva and V. Ramamurthy, Org. Lett., 2013, 15(17), 4374-4377.
- 76 A. Baldridge, S. R. Samanta, N. Jayaraj, V. Ramamurthy and L. M. Tolbert, J. Am. Chem. Soc., 2010, 132(5), 1498-1499.
- 77 A. Baldridge, S. R. Samanta, N. Jayaraj, V. Ramamurthy and L. M. Tolbert, J. Am. Chem. Soc., 2011, 133(4), 712-715.
- 78 L. S. Kaanumalle, C. L. D. Gibb, B. C. Gibb and V. Ramamurthy, J. Am. Chem. Soc., 2005, 127(11), 3674-3675.
- 79 (a) S. R. Samanta, R. Kulasekharan, R. Choudhury, P. Jagadesan, N. Jayaraj and V. Ramamurthy, Langmuir, 2012, 28(32), 11920-11928; (b) B. Mondal, N. Kamatham, S. R. Samanta, P. Jagadesan, J. He and V. Ramamurthy, Langmuir, 2013, 29(41), 12703-12709.
- 80 (a) J. R. Moran, S. Karbach and D. J. Cram, J. Am. Chem. Soc., 1982, 104(21), 5826-5828; (b) J. R. Moran, J. L. Ericson, E. Dalcanale, J. A. Bryant, C. B. Knobler and D. J. Cram, J. Am. Chem. Soc., 1991, 113, 5707-5714.
- 81 J. Kubitschke, S. Javor and J. Rebek, Jr., Chem. Commun., 2012, 48(74), 9251-9253.
- 82 M. P. Schramm, R. J. Hooley and J. Rebek, Jr., J. Am. Chem. Soc., 2007, 129(31), 9773-9779.
- 83 (a) F. Hof, L. Trembleau, E. C. Ullrich and J. Rebek, Jr., Angew. Chem., Int. Ed., 2003, 42, 3150-3153; (b) S. M. Biros, E. C. Ullrich, F. Hof, L. Trembleau and J. Rebek, Jr., J. Am. Chem. Soc., 2004, 126(9), 2870-2876.
- 84 R. J. Hooley, H. J. V. Anda and J. Rebek, Jr., J. Am. Chem. Soc., 2007, 129, 13464-13473.
- 85 L. Trembleau and J. Rebek, Jr., Chem. Commun., 2004, 58-59.
- 86 R. J. Hooley, S. M. Biros and J. Rebek, Jr., Chem. Commun., 2006, 509-510.
- R. J. Hooley, S. M. Biros and J. Rebek, Jr., Angew. Chem., 2006, 45(21), 3517-3519.

- 88 D. Podkoscielny, R. J. Hooley, J. Rebek, Jr. and A. E. Kaifer, Org. Lett., 2008, 10(13), 2865-2868.
- 89 R. J. Hooley, H. J. Van Anda and J. Rebek, Jr., J. Am. Chem. Soc., 2006, 128(12), 3894-3895.
- 90 C. H. Haas, S. M. Biros and J. Rebek, Jr., Chem. Commun., 2005, 6044-6045.
- 91 D. A. Ryan and J. Rebek, J. Am. Chem. Soc., 2011, 133(49), 19653-19655.
- 92 (a) T. Haino, D. M. Rudkevich and J. Rebek, Jr., J. Am. Chem. Soc., 1999, 121, 11253-11254; (b) T. Haino, D. M. Rudkevich, A. Shivanyuk, K. Rissanen and J. Rebek, Jr., Chemistry, 2000, 6(20), 3797-3805.
- 93 A. Lledo and J. Rebek, Jr., Chem. Commun., 2010, 46(45), 8630-8632.
- 94 D. A. Ryan and J. Rebek, Analyst, 2013, 138(4), 1008-1010.
- 95 S. Javor and J. Rebek, Jr., J. Am. Chem. Soc., 2011, 133(43), 17473-17478.
- 96 S. M. Butterfield and J. Rebek, Jr., J. Am. Chem. Soc., 2006, 128(48), 15366-15367.
- 97 Y. Liu, T. Taira, M. C. Young, D. Ajami, J. Rebek, Q. Cheng and R. J. Hooley, Langmuir, 2012, 28(2), 1391-1398.
- 98 M. H. Ebbing, M. J. Villa, J. M. Valpuesta, P. Prados and J. de Mendoza, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**(8), 4962-4966.
- 99 K. D. Zhang, D. Ajami, J. V. Gavette and J. Rebek, Jr., Chem. Commun., 2014, 50(38), 4895-4897.
- 100 K. Harano, S. Hiraoka and M. Shionoya, J. Am. Chem. Soc., 2007, 129(17), 5300-5301.
- 101 S. Hiraoka, K. Harano, M. Shiro and M. Shionoya, J. Am. Chem. Soc., 2008, 130(44), 14368-14369.
- 102 S. Hiraoka, K. Harano, T. Nakamura, M. Shiro and M. Shionoya, Angew. Chem., 2009, 48(38), 7006-7009.
- 103 S. Hiraoka, T. Nakamura, M. Shiro and M. Shionoya, J. Am. Chem. Soc., 2010, 132(38), 13223-13225.