Recap general linear model

Lieven Clement

statOmics, Ghent University (https://statomics.github.io)

Contents

1	Breast cancer example						
2	Dat	Data Exploration					
	2.1	Import	7				
	2.2	Summary statistics	8				
	2.3	Visualisation	8				
	2.4	Research questions	10				
	2.5	Estimation of effect size and standard error	11				
3	Statistical Inference						
	3.1	Log transformation	15				
	3.2	Conclusion	18				
4	Ger	General Linear Model					
	4.1	Implementation in R	20				
	4.2	Assumptions	21				
	4.3	Breast cancer example	25				
5	Linear regression in matrix form						
	5.1	Scalar form	26				
	5.2	Matrix form	26				
	5.3	Least Squares (LS)	27				
	5.4	Variance Estimator?	38				
	5.5	Contrasts	38				
6	Hor	Homework: Adopt the gene analysis on log scale in matrix form!					
	6.1	Inspiration	39				
Тŀ	nie ie	part of the online course Statistical Genomics 2022 (SGA)					

1 Breast cancer example

- part of study https://doi.org/10.1093/jnci/djj052)
- Histologic grade in breast cancer clinically prognostic. Association of histologic grade on expression of KPNA2 gene that is known to be associated with poor BC prognosis.
- Population: all current and future breast cancer patients













2 Data Exploration

2.1 Import

```
library(tidyverse)
gene <- read.table("https://raw.githubusercontent.com/stat0mics/SGA21/master/data/kpna2.txt",header=TRU.head(gene)</pre>
```

```
## J grade node gene
## 1 3 1 367.8179
## 2 3 1 590.3576
## 3 1 1 346.6583
## 4 1 1 258.4455
## 5 1 0 153.8416
## 6 3 0 643.6799
```

We will transform the variable grade and node to a factor

```
gene$grade <- as.factor(gene$grade)
gene$node <- as.factor(gene$node)</pre>
```

2.2 Summary statistics

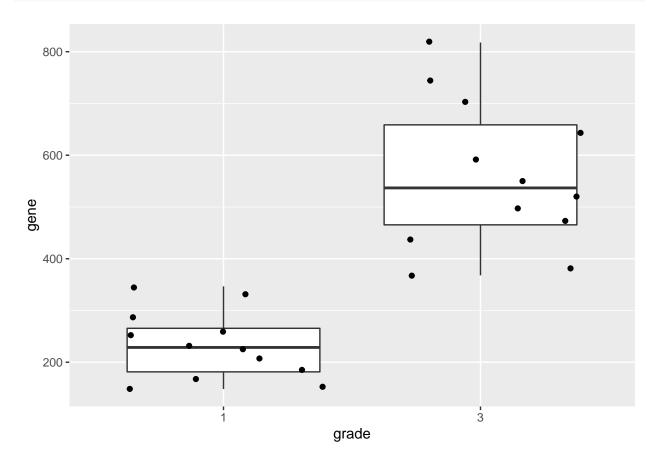
2.3 Visualisation

2 3

561. 144.

12 41.4

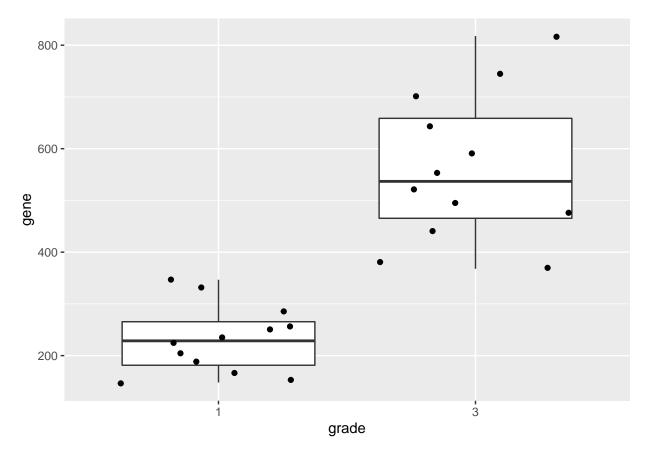
```
gene %>%
ggplot(aes(x=grade,y=gene)) +
geom_boxplot(outlier.shape=NA) +
geom_jitter()
```



We can also save the plots as objects for later use!

```
p1 <- gene %>%
    ggplot(aes(x=grade,y=gene)) +
    geom_boxplot(outlier.shape=NA) +
    geom_jitter()

p2 <- gene %>%
    ggplot(aes(sample=gene)) +
    geom_qq() +
    geom_qq_line() +
    facet_wrap(~grade)
```



p2



2.4 Research questions

Researchers want to assess the association of the histological grade on KPNA2 gene expression



2.5 Estimation of effect size and standard error

```
effectSize <- tibble(</pre>
 delta = geneSum$mean[2] - geneSum$mean[1],
  seDelta = geneSum %>%
    pull(se) %>%
    .^2 %>%
    sum %>%
    sqrt
effectSize
## # A tibble: 1 x 2
##
     delta seDelta
     <dbl>
           <dbl>
##
              45.5
## 1 329.
```

3 Statistical Inference

• Researchers want to assess the association of histological grade on KPNA2 gene expression

• Inference?



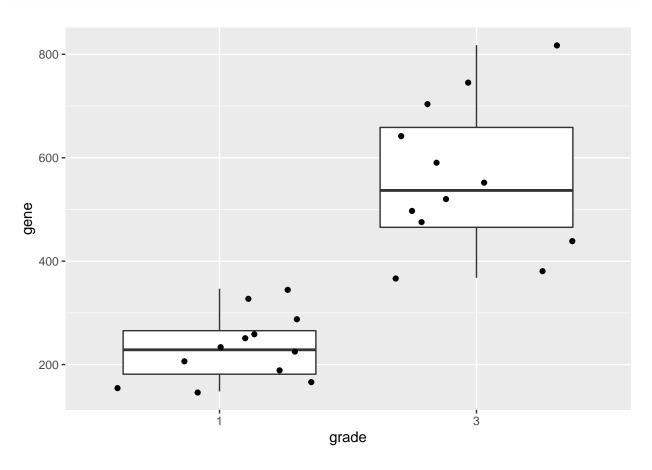
- Researchers want to assess the association of histological grade on KPNA2 gene expression
- Inference?
- testing + CI $\$ \rightarrow \$$ Assumptions
- In general we start from alternative hypothese H_A : we want to show an association
- Gene expression of grade 1 and grade 3 patients is on average different
- But, we will assess it by falsifying the opposite:
- The average KPNA2 gene expression of grade 1 and grade 3 patients is equal
- How likely is it to observe an equal or more extreme association than the one observed in the sample when the null hypothesis is true?

- When we make assumptions about the distribution of our test statistic we can quantify this probability: **p-value**.
- If the p-value is below a significance threshold α we reject the null hypothesis

We control the probability on a false positive result at the α -level (type I error)

• The p-value will only be calculated correctly if the underlying assumptions hold!

library(gridExtra) p1



p2



t.test(gene~grade,data=gene)

<dbl> <dbl> <dbl>

1 329. 45.5 7.21 0.000000376

##

```
##
   Welch Two Sample t-test
##
##
## data: gene by grade
## t = -7.2132, df = 15.384, p-value = 2.598e-06
\#\# alternative hypothesis: true difference in means between group 1 and group 3 is not equal to 0
## 95 percent confidence interval:
## -425.4218 -231.6751
## sample estimates:
## mean in group 1 mean in group 3
##
          232.5003
                          561.0487
effectSize <- effectSize %>%
  mutate(t.stat=delta/seDelta) %>%
  mutate(p.value= pt(-abs(t.stat),21.352)*2)
effectSize
## # A tibble: 1 x 4
     delta seDelta t.stat
                              p.value
```

<dbl>

- Intensities are often not normally distributed and have a mean variance relation
- Commonly log2-transformed
- Differences on log scale:

$$\log_2(B) - \log_2(A) = \log_2\frac{B}{A} = \log_2FC_{\frac{B}{A}}$$



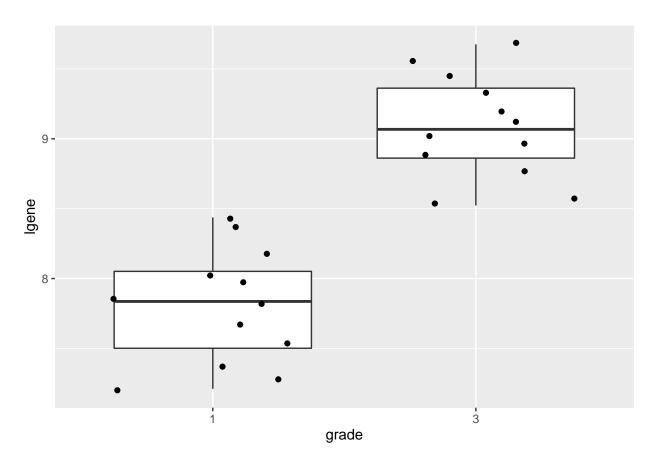
3.1 Log transformation

```
gene <- gene %>%
  mutate(lgene = log2(gene))

p1 <- gene %>%
  ggplot(aes(x=grade,y=lgene)) +
  geom_boxplot(outlier.shape=NA) +
  geom_jitter()

p2 <- gene %>%
  ggplot(aes(sample=lgene)) +
  geom_qq() +
```

```
geom_qq_line() +
facet_wrap(~grade)
p1
```



p2



logtest <- t.test(lgene~grade,data=gene,var.equal=TRUE)
logtest</pre>

```
##
## Two Sample t-test
##
## data: lgene by grade
## t = -8.0455, df = 22, p-value = 5.372e-08
## alternative hypothesis: true difference in means between group 1 and group 3 is not equal to 0
## 95 percent confidence interval:
## -1.610148 -0.950178
## sample estimates:
## mean in group 1 mean in group 3
##
          7.808478
                           9.088641
log2FC <- logtest$estimate[2]-logtest$estimate[1]</pre>
log2FC
## mean in group 3
          1.280163
names(log2FC) \leftarrow "g3-g1"
2<sup>1</sup>log2FC
```

g3-g1 ## 2.428664

3.2 Conclusion

There is a extremely significant association of the histological grade on the gene expression in tumor tissue. On average, the gene expression for the grade 3 patients is 2.43 times higher than the gene expression in grade 1 patients (95% CI [1.93, 3.05], $p \ll 0.001$).





The patients also differ in the their lymph node status. Hence, we have a two factorial design: grade x lymph node status!!!

Solution??



4 General Linear Model

How can we integrate multiple factors and continuous covariates in linear model.

$$y_i = \beta_0 + \beta_1 x_{i,1} + \beta_2 x_{i,2} + \beta_{12} x_{i,1} x_{i,2} + \epsilon_i,$$

with

- $x_{i,1}$ a dummy variable for histological grade: $x_{i,1} = \begin{cases} 0 & \text{grade 1} \\ 1 & \text{grade 3} \end{cases}$
- $x_{i,2}$ a dummy variable for : $x_{i,2} = \begin{cases} 0 & \text{lymph nodes were not removed} \\ 1 & \text{lymph nodes were removed} \end{cases}$
- ϵ_i ?

4.1 Implementation in R

```
lm1 <- lm(gene~grade*node,data=gene)</pre>
summary(lm1)
##
## Call:
## lm(formula = gene ~ grade * node, data = gene)
##
## Residuals:
##
             1Q Median
       Min
                                 3Q
                                        Max
## -201.748 -53.294 -6.308 46.216 277.601
##
## Coefficients:
##
              Estimate Std. Error t value Pr(>|t|)
## (Intercept) 180.51 44.37 4.068 0.0006 ***
               401.33
                           62.75 6.396 3.07e-06 ***
## grade3
## node1
               103.98
                           62.75 1.657 0.1131
## grade3:node1 -145.57
                           88.74 -1.640 0.1166
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 108.7 on 20 degrees of freedom
## Multiple R-squared: 0.7437, Adjusted R-squared: 0.7052
## F-statistic: 19.34 on 3 and 20 DF, p-value: 3.971e-06
```

4.2 Assumptions

plot(lm1)











Factor Level Combinations

4.3 Breast cancer example

- Paper: https://doi.org/10.1093/jnci/djj052
- Histologic grade in breast cancer provides clinically important prognostic information. Two factors have to be concidered: Histologic grade (grade 1 and grade 3) and lymph node status (0 vs 1). The researchers assessed gene expression of the KPNA2 gene a protein-coding gene associated with breast cancer and are mainly interested in the association of histological grade. Note, that the gene variable consists of background corrected normalized intensities obtained with a microarray platform. Upon log-transformation, they are known to be a good proxy for the log transformed concentration of gene expression product of the KPNA2 gene.
- Research questions and translate them towards model parameters (contrasts)?
- Make an R markdown file to answer the research questions

```
library(ExploreModelMatrix)
explMx <- VisualizeDesign(gene, designFormula = ~grade*node)
explMx$plotlist</pre>
```

[[1]]



You can also explore the model matrix interactively:

ExploreModelMatrix(gene,designFormula = ~grade*node)

5 Linear regression in matrix form

5.1 Scalar form

- Consider a vector of predictors $\mathbf{x} = (x_1, \dots, x_p)^T$ and
- a real-valued response Y
- then the linear regression model can be written as

$$Y = f(\mathbf{x}) + \epsilon = \beta_0 + \sum_{j=1}^p x_j \beta_j + \epsilon$$

with i.i.d. $\epsilon \sim N(0, \sigma^2)$

5.2 Matrix form

• n observations $(\mathbf{x}_1,y_1)\dots(\mathbf{x}_n,y_n)$

• Regression in matrix notation

$$\mathbf{Y} = \mathbf{X} + \\ \text{with } \mathbf{Y} = \begin{bmatrix} y_1 \\ \vdots \\ y_n \end{bmatrix}, \, \mathbf{X} = \begin{bmatrix} 1 & x_{11} & \dots & x_{1p} \\ \vdots & \vdots & & \vdots \\ 1 & x_{n1} & \dots & x_{np} \end{bmatrix}, \, = \begin{bmatrix} \beta_0 \\ \vdots \\ \beta_p \end{bmatrix} \text{ and } = \begin{bmatrix} \epsilon_1 \\ \vdots \\ \epsilon_n \end{bmatrix}$$

5.3 Least Squares (LS)

• Minimize the residual sum of squares

$$RSS(\) = \sum_{i=1}^{n} e_i^2$$
$$= \sum_{i=1}^{n} \left(y_i - \beta_0 - \sum_{j=1}^{p} x_{ij} \beta_j \right)^2$$

• or in matrix notation

$$RSS(\) = (\mathbf{Y} - \mathbf{X}\)^T (\mathbf{Y} - \mathbf{X}\)$$
$$= \|\mathbf{Y} - \mathbf{X}\ \|_2^2$$

with the L_2 -norm of a p-dim. vector $v \ \|\mathbf{v}\| = \sqrt{v_1^2 + \ldots + v_p^2} \to \hat{\ } = \mathrm{argmin}_\beta \|\mathbf{Y} - \mathbf{X}\|_2^2$

5.3.1 Minimize RSS

$$\frac{\partial RSS}{\partial} = \mathbf{0}$$

$$\frac{(\mathbf{Y} - \mathbf{X})^T (\mathbf{Y} - \mathbf{X})}{\partial} = \mathbf{0}$$

$$-2\mathbf{X}^T (\mathbf{Y} - \mathbf{X}) = \mathbf{0}$$

$$\mathbf{X}^T \mathbf{X} = \mathbf{X}^T \mathbf{Y}$$

$$= (\mathbf{X}^T \mathbf{X})^{-1} \mathbf{X}^T \mathbf{Y}$$

5.3.2 Geometric Interpretation

5.3.2.1 Toy dataset We will illustrate this on a toy dataset

```
library(tidyverse)
data <- data.frame(x=1:3,y=c(1,2,2))
data</pre>
```

x y ## 1 1 1 ## 2 2 2 ## 3 3 2 5.3.2.2 Matrix form for toy dataset We can also write this in matrix form

$$\mathbf{Y} = \mathbf{X}\beta + \epsilon$$

with

$$\mathbf{Y} = \left[\begin{array}{c} 1 \\ 2 \\ 2 \end{array} \right], \quad \mathbf{X} = \left[\begin{array}{c} 1 & 1 \\ 1 & 2 \\ 1 & 3 \end{array} \right], \quad \beta = \left[\begin{array}{c} \beta_0 \\ \beta_1 \end{array} \right] \quad \text{and} \quad \epsilon = \left[\begin{array}{c} \epsilon_1 \\ \epsilon_2 \\ \epsilon_3 \end{array} \right]$$

5.3.2.3 Classical interpretation Model fit and predictions based on the toy dataset

```
lm1 <- lm(y~x,data)
data$yhat <- lm1$fitted

data %>%
    ggplot(aes(x,y)) +
    geom_point() +
    ylim(0,4) +
    xlim(0,4) +
    stat_smooth(method = "lm", color = "red", fullrange = TRUE) +
    geom_point(aes(x=x, y =yhat), pch = 2, size = 3, color = "red") +
    geom_segment(data = data, aes(x = x, xend = x, y = y, yend = yhat), lty = 2)

## `geom_smooth()` using formula 'y ~ x'
```

Warning in max(ids, na.rm = TRUE): no non-missing arguments to max; returning
-Inf



5.3.3 Projection

There is also another picture to regression:

- Instead of plotting each observation $i=1\dots n$ as a data-point in \mathbb{R}^p with dimensions $1\dots p$ for every variable/feature that is recorded for each observation
- We can also plot \mathbf{Y} , $\hat{\mathbf{Y}}$ and each column of \mathbf{X} : \mathbf{X}_j with $j=1\dots p$ as a vector in \mathbb{R}^n with dimensions $1\dots n$ for every observation.
- In this representation linear regression can be interpreted as a projection of the vector \mathbf{Y} onto the subspace of \mathbb{R}^n that is spanned by the vectors for the predictors $\mathbf{X}_1 \dots \mathbf{X}_p$.
- The space $\mathbf{X}_1 \dots \mathbf{X}_p$ is also referred to as the column space of \mathbf{X} , the space that consists of all linear combinations of the vectors of the predictors or columns $\mathbf{X}_1 \dots \mathbf{X}_p$.

5.3.3.1 Intermezzo: Projection of vector on X and Y axis

$$\mathbf{e} = \left[\begin{array}{c} e_1 \\ e_2 \end{array} \right], \mathbf{u}_1 = \left[\begin{array}{c} 1 \\ 0 \end{array} \right], \mathbf{u}_2 = \left[\begin{array}{c} 0 \\ 1 \end{array} \right]$$



1. Projection of error on x-axis

$$\begin{aligned} \mathbf{u}_1^T \mathbf{e} &= & \|\mathbf{u}_1\|_2 \|\mathbf{e}_1\|_2 \cos < \mathbf{u}_1, \mathbf{e}_1 > \\ &= & \begin{bmatrix} 1 & 0 \end{bmatrix} \begin{bmatrix} e_1 \\ e_2 \end{bmatrix} \\ &= & 1 \times e_1 + 0 \times e_2 \\ &= & e_1 \end{aligned}$$

2. Projection of error on y-axis

$$\mathbf{u}_2^T \mathbf{e} = \begin{bmatrix} 1 & 0 \end{bmatrix} \begin{bmatrix} e_1 \\ e_2 \end{bmatrix}$$
$$= 0 \times e_1 + 1 \times e_2$$
$$= e_2$$

3. Projection of error on itself

$$\begin{array}{lcl} \mathbf{e}^T\mathbf{e} & = & \left[\begin{array}{ccc} e_1 & e_2 \end{array} \right] \left[\begin{array}{c} e_1 \\ e_2 \end{array} \right] \\ & = & e_1^2 + e_2^2 \\ & = & \|e\|_2^2 \rightarrow \text{ Pythagorean theorem} \end{array}$$

5.3.3.2 Interpretation of least squares as a projection Fitted values:

$$\begin{aligned} \hat{\mathbf{Y}} &=& \mathbf{X}\hat{\boldsymbol{\beta}} \\ &=& \mathbf{X}(\mathbf{X}^T\mathbf{X})^{-1}\mathbf{X}^T\mathbf{Y} \\ &=& \mathbf{H}\mathbf{Y} \end{aligned}$$

with ${\bf H}$ the projection matrix also referred to as the hat matrix.

```
X <- model.matrix(~x,data)</pre>
Х
##
     (Intercept) x
## 1 1 1
## 2
              1 3
## 3
## attr(,"assign")
## [1] 0 1
XtX \leftarrow t(X)%*%X
               (Intercept) x
## (Intercept)
                         6 14
## x
XtXinv <- solve(t(X)%*%X)</pre>
XtXinv
               (Intercept)
## (Intercept) 2.333333 -1.0
                 -1.000000 0.5
H <- X %*% XtXinv %*% t(X)
                        2
##
## 1 0.8333333 0.3333333 -0.1666667
## 2 0.3333333 0.3333333 0.3333333
## 3 -0.1666667 0.3333333 0.8333333
```

```
Y <- data$y
Yhat <- H%*%Y
Yhat

## [,1]
## 1 1.166667
## 2 1.666667
## 3 2.166667
```

- We can also interpret the fit as the projection of the $n \times 1$ vector **Y** on the column space of the matrix **X**.
- So each column in **X** is also an $n \times 1$ vector.
- For the toy example n=3 and p=2. The other picture to linear regression is to consider X_0 , X_1 and Y as vectors in the space of the data \mathbb{R}^n , here \mathbb{R}^3 because we have three data points. So the column space of X is a plane in the three dimensional space.

$$\hat{\mathbf{Y}} = \mathbf{X}(\mathbf{X}^T \mathbf{X})^{-1} \mathbf{X}^T \mathbf{Y}$$

1. Plane spanned by column space:

```
originRn <- data.frame(X1=0, X2=0, X3=0)</pre>
data$x0 <- 1
dataRn <- data.frame(t(data))</pre>
library(plotly)
p1 <- plot_ly(
    originRn,
    x = \sim X1
    y = \sim X2
    z= ~ X3, name="origin") %>%
  add_markers(type="scatter3d") %>%
  layout(
    scene = list(
      aspectmode="cube",
      xaxis = list(range=c(-4,4)), yaxis = list(range=c(-4,4)), zaxis = list(range=c(-4,4))
      )
    )
p1 <- p1 %>%
  add_trace(
    x = c(0,1),
    y = c(0,0),
    z = c(0,0),
    mode = "lines",
    line = list(width = 5, color = "grey"),
    type="scatter3d",
    name = "obs1") %>%
  add_trace(
    x = c(0,0),
    y = c(0,1),
```

```
z = c(0,0),
    mode = "lines",
    line = list(width = 5, color = "grey"),
    type="scatter3d",
    name = "obs2") %>%
  add_trace(
   x = c(0,0),
    y = c(0,0),
    z = c(0,1),
    mode = "lines",
   line = list(width = 5, color = "grey"),
    type="scatter3d",
   name = "obs3") %>%
  add_trace(
   x = c(0,1),
    y = c(0,1),
    z = c(0,1),
   mode = "lines",
   line = list(width = 5, color = "black"),
   type="scatter3d",
   name = "X1") %>%
   add_trace(
   x = c(0,1),
   y = c(0,2),
   z = c(0,3),
   mode = "lines",
   line = list(width = 5, color = "black"),
    type="scatter3d",
    name = "X2")
р1
```

2. Vector of Y:

Actual values of **Y**:

data\$y

[1] 1 2 2

$$\mathbf{Y} = \left[\begin{array}{c} 1 \\ 2 \\ 2 \end{array} \right]$$

```
p2 <- p1 %>%
  add_trace(
    x = c(0,Y[1]),
    y = c(0,Y[2]),
    z = c(0,Y[3]),
    mode = "lines",
    line = list(width = 5, color = "red"),
    type="scatter3d",
```

```
name = "Y")
p2
```

3. Projection of Y onto column space

Actual values of fitted values $\hat{\mathbf{Y}}$:

```
data$yhat
```

[1] 1.166667 1.666667 2.166667

$$\mathbf{Y} = \left[\begin{array}{c} 1.1666667 \\ 1.6666667 \\ 2.1666667 \end{array} \right]$$

```
p2 <- p2 %>%
  add_trace(
    x = c(0, Yhat[1]),
    y = c(0, Yhat[2]),
    z = c(0, Yhat[3]),
    mode = "lines",
    line = list(width = 5, color = "orange"),
    type="scatter3d",
    name="Yhat") %>%
    add_trace(
    x = c(Y[1], Yhat[1]),
    y = c(Y[2], Yhat[2]),
    z = c(Y[3], Yhat[3]),
    mode = "lines",
    line = list(width = 5, color = "red", dash="dash"),
    type="scatter3d",
    name="Y -> Yhat"
p2
```

Y is projected in the column space of X! spanned by the columns.

5.3.3.3 How does this projection works?

$$\begin{aligned} \hat{\mathbf{Y}} &= & \mathbf{X}(\mathbf{X}^T \mathbf{X})^{-1} \mathbf{X}^T \mathbf{Y} \\ &= & \mathbf{X}(\mathbf{X}^T \mathbf{X})^{-1/2} (\mathbf{X}^T \mathbf{X})^{-1/2} \mathbf{X}^T \mathbf{Y} \\ &= & \mathbf{U} \mathbf{U}^T \mathbf{Y} \end{aligned}$$

- **U** is a new orthonormal basis in \mathbb{R}^2 , a subspace of \mathbb{R}^3
- The space spanned by U and X is the column space of X, e.g. it contains all possible linear combinantions of X. $\mathbf{U}^t \mathbf{Y}$ is the projection of Y on this new orthonormal basis

```
eigenXtX <- eigen(XtX)
XtXinvSqrt <- eigenXtX$vectors %*%diag(1/eigenXtX$values^.5)%*%t(eigenXtX$vectors)
U <- X %*% XtXinvSqrt

• U orthonormal basis</pre>
```

```
## [,1] [,2]
## 1 0.9116067 -0.04802616
## 2 0.3881706 0.42738380
## 3 -0.1352655 0.90279376

t(U)%*%U

## [,1] [,2]
## [1,] 1.000000e+00 2.915205e-16
## [2,] 2.915205e-16 1.000000e+00

• UU<sup>T</sup> equals projection matrix
```

```
U%*%t(U)
```

```
## 1 2 3
## 1 0.8333333 0.3333333 -0.1666667
## 2 0.3333333 0.3333333 0.3333333
## 3 -0.1666667 0.3333333 0.8333333
```

Η

```
## 1 2 3
## 1 0.833333 0.333333 -0.1666667
## 2 0.3333333 0.333333 0.3333333
## 3 -0.1666667 0.3333333 0.8333333
```

```
p3 <- p1 %>%
  add_trace(
    x = c(0,U[1,1]),
    y = c(0,U[2,1]),
    z = c(0,U[3,1]),
    mode = "lines",
    line = list(width = 5, color = "blue"),
    type="scatter3d",
    name = "U1") %>%
  add_trace(
    x = c(0,U[1,2]),
    y = c(0,U[2,2]),
    z = c(0,U[3,2]),
    mode = "lines",
  line = list(width = 5, color = "blue"),
```

```
type="scatter3d",
name = "U2")
p3
```

- $\mathbf{U}^T \mathbf{Y}$ is the projection of \mathbf{Y} in the space spanned by \mathbf{U} .
- Indeed $\mathbf{U}_1^T \mathbf{Y}$

```
p4 <- p3 %>%
  add_trace(
    x = c(0, Y[1]),
   y = c(0, Y[2]),
   z = c(0, Y[3]),
   mode = "lines",
    line = list(width = 5, color = "red"),
    type="scatter3d",
    name = "Y") %>%
  add_trace(
    x = c(0,U[1,1]*(U[,1]%*%Y)),
    y = c(0,U[2,1]*(U[,1]%*%Y)),
    z = c(0,U[3,1]*(U[,1]%*%Y)),
    mode = "lines",
    line = list(width = 5, color = "red",dash="dash"),
    type="scatter3d",
    name="Y -> U1") %>% add_trace(
   x = c(Y[1],U[1,1]*(U[,1]%*%Y)),
   y = c(Y[2],U[2,1]*(U[,1]%*%Y)),
    z = c(Y[3],U[3,1]*(U[,1]%*%Y)),
    mode = "lines",
    line = list(width = 5, color = "red", dash="dash"),
    type="scatter3d",
    name="Y -> U1")
р4
```

• and $\mathbf{U}_2^T \mathbf{Y}$

```
p5 <- p4 %>%
  add_trace(
   x = c(0,U[1,2]*(U[,2]%*%Y)),
    y = c(0,U[2,2]*(U[,2]%*%Y)),
    z = c(0,U[3,2]*(U[,2]%*%Y)),
    mode = "lines",
    line = list(width = 5, color = "red",dash="dash"),
    type="scatter3d",
    name="Y -> U2") %>% add_trace(
    x = c(Y[1],U[1,2]*(U[,2]%*%Y)),
   y = c(Y[2],U[2,2]*(U[,2]%*%Y)),
    z = c(Y[3],U[3,2]*(U[,2]%*%Y)),
    mode = "lines",
    line = list(width = 5, color = "red", dash="dash"),
    type="scatter3d",
    name="Y -> U2")
p5
```

• $\hat{\mathbf{Y}}$ is the resulting vector that lies in the plane spanned by \mathbf{U}_1 and \mathbf{U}_2 and thus also in the column space of \mathbf{X} .

```
p6 <- p5 %>%
  add_trace(
    x = c(0, Yhat[1]),
    y = c(0, Yhat[2]),
    z = c(0, Yhat[3]),
    mode = "lines",
    line = list(width = 5, color = "orange"),
    type="scatter3d",
    name = "Yhat") %>%
  add_trace(
    x = c(Y[1], Yhat[1]),
    y = c(Y[2], Yhat[2]),
    z = c(Y[3], Yhat[3]),
    mode = "lines",
    line = list(width = 5, color = "maroon2"),
    type="scatter3d",
    name = "e") %>%
  add_trace(
    x = c(U[1,1]*(U[,1]%*%Y),Yhat[1]),
    y = c(U[2,1]*(U[,1]%*%Y),Yhat[2]),
    z = c(U[3,1]*(U[,1]%*%Y),Yhat[3]),
    mode = "lines",
    line = list(width = 5, color = "orange", dash="dash"),
    type="scatter3d",
    name = "Y -> U") %>%
  add_trace(
    x = c(U[1,2]*(U[,2]%*%Y),Yhat[1]),
    y = c(U[2,2]*(U[,2]%*%Y),Yhat[2]),
    z = c(U[3,2]*(U[,2]%*%Y),Yhat[3]),
    mode = "lines",
    line = list(width = 5, color = "orange", dash="dash"),
    type="scatter3d",
    name = "Y \rightarrow U")
p6
```

5.3.4 Error

Note, that it is also clear from the equation in the derivation of the least squares solution that the residual is orthogonal on the column space:

$$-2\mathbf{X}^T(\mathbf{Y} - \mathbf{X}\boldsymbol{\beta}) = 0$$

5.4 Variance Estimator?

$$\hat{\Sigma}_{-} = \operatorname{var} \left[(\mathbf{X}^{T} \mathbf{X})^{-1} \mathbf{X}^{T} \mathbf{Y} \right]$$

$$= (\mathbf{X}^{T} \mathbf{X})^{-1} \mathbf{X}^{T} \operatorname{var} \left[\mathbf{Y} \right] \mathbf{X} (\mathbf{X}^{T} \mathbf{X})^{-1}$$

$$= (\mathbf{X}^{T} \mathbf{X})^{-1} \mathbf{X}^{T} (\mathbf{I} \sigma^{2}) \mathbf{X} (\mathbf{X}^{T} \mathbf{X})^{-1}$$

$$= (\mathbf{X}^{T} \mathbf{X})^{-1} \mathbf{X}^{T} \mathbf{I} \quad \mathbf{X} (\mathbf{X}^{T} \mathbf{X})^{-1} \sigma^{2}$$

$$= (\mathbf{X}^{T} \mathbf{X})^{-1} \mathbf{X}^{T} \mathbf{X} (\mathbf{X}^{T} \mathbf{X})^{-1} \sigma^{2}$$

$$= (\mathbf{X}^{T} \mathbf{X})^{-1} \sigma^{2}$$

5.5 Contrasts

Hypotheses often involve linear combinations of the model parameters! e.g.

•
$$H_0: \log_2 FC_{g3n1-g1n1} = \beta_{g3} + \hat{\beta}_{g3n1} = 0$$
 – "grade3+grade3:node1 = 0"

• Let

$$\beta = \left[\begin{array}{c} \beta_0 \\ \beta_{g3} \\ \beta_{n1} \\ \beta_{g3:n1} \end{array} \right]$$

• we can write that contrast using a contrast matrix:

$$\mathbf{L} = \begin{bmatrix} 0 \\ 1 \\ 0 \\ 1 \end{bmatrix} \to \mathbf{L}^T \beta$$

• Then the variance becomes:

$$\mathrm{var}_{\mathbf{L}^T\hat{\beta}} = \mathbf{L}^T \Sigma_{\hat{\beta}} \mathbf{L}$$

6 Homework: Adopt the gene analysis on log scale in matrix form!

- 1. Study the solution of the exercise to understand the analysis in R
- 2. Calculate
- model parameters and contrasts of interest
- standard errors, standard errors on contrasts
- t-test statistics on the model parameters and contrasts of interest
- 3. Compare your results with the output of the lm(.) function

6.1 Inspiration

Tip: details on the implementation can be found in the book of Faraway (chapter 2). https://people.bath. ac.uk/jjf23/book/

• Design matrix

```
X <- model.matrix(~grade*node,data=gene)</pre>
```

• Transpose of a matrix: use function t(.)

t(X)

```
##
                1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24
## (Intercept) 1 1 1 1 1 1 1 1 1
                                  1
                                     1
                                         1
                                            1
                                              1
                                                  1
                                                    1
                                                        1
                                                           1
                                                              1
                                                                 1
                                                                    1
## grade3
               1 1 0 0 0 1 1 0 1
                                         1
                                            0
                                               0
                                                  0
                                                    1
                                                        0
                                                           1
                                   1
                                      0
                1 1 1 1 0 0 1 1 0
## node1
                                  0
                                     0
                                        0
                                           0
                                               0
                                                  0
                                                     1
                                                        1
                                                           0
                                                              1
                                                                 1
                                                                    1
                                                                             0
## grade3:node1 1 1 0 0 0 0 1 0 0 0 0
                                           0
                                              0
                                                  0
                                                     1
                                                        0
                                                           0
                                                              0
## attr(,"assign")
## [1] 0 1 2 3
## attr(,"contrasts")
## attr(,"contrasts")$grade
## [1] "contr.treatment"
## attr(,"contrasts")$node
## [1] "contr.treatment"
```

• Matrix product %*% operator

t(X)%*%X

```
##
                 (Intercept) grade3 node1 grade3:node1
## (Intercept)
                          24
                                  12
                                        12
## grade3
                          12
                                  12
                                         6
                                                       6
## node1
                          12
                                   6
                                        12
                                                       6
## grade3:node1
                            6
                                         6
```

• Degrees of freedom of a model?

$$df = n - p$$

```
summary(lm1)
```

```
##
## Call:
## lm(formula = y ~ x, data = data)
##
## Residuals:
## 1 2 3
```

```
## -0.1667 0.3333 -0.1667
##
## Coefficients:
##
              Estimate Std. Error t value Pr(>|t|)
## (Intercept)
                0.6667
                           0.6236
                                     1.069
                                              0.479
## x
                 0.5000
                            0.2887
                                     1.732
                                              0.333
## Residual standard error: 0.4082 on 1 degrees of freedom
## Multiple R-squared: 0.75, Adjusted R-squared:
                3 on 1 and 1 DF, p-value: 0.3333
## F-statistic:
dfRes <- (nrow(X)-ncol(X))</pre>
dfRes
```

[1] 20

• Variance estimator: MSE

$$\hat{\sigma}^2 = \frac{\sum\limits_{i=1}^n \epsilon_i^2}{n-p}$$

- Invert matrix: use function solve(.)
- Diagonal elements of a matrix: use function diag(.)

t(X)%*%X

##		(Intercept)	grade3	node1	<pre>grade3:node1</pre>
##	(Intercept)	24	12	12	6
##	grade3	12	12	6	6
##	node1	12	6	12	6
##	<pre>grade3:node1</pre>	6	6	6	6

diag(t(X)%*%X)