





Draft Genome Sequences of Enterobacteriales Strains Isolated from the International Space Station

Achintya R. Bharadwaj, Robert Daudu, Altin K. Singh, Bolason M. Wood, Marilyne Debieu, Niamh B. O'Hara, D. C. Bara, Baradwaj, Altin K. Singh, Bolason M. Wood, Marilyne Debieu, Mara, Discount B. O'Hara, D. C. Baradwaj, Altin K. Singh, Bolason M. Wood, Marilyne Debieu, Maradwaj, Fathi Karouia, de Christopher E. Mason, f, De Kasthuri Venkateswarana

^aJet Propulsion Laboratory, California Institute of Technology, Pasadena, California, USA

ABSTRACT The whole-genome sequences of 26 strains isolated from the International Space Station were generated, and the strains were identified as being members of the order Enterobacteriales. Characterization of these whole-genome sequences might enable the identification of potential pathogenic bacteria that have been adapting to the space environment.

embers of the order Enterobacteriales have been found to exhibit human pathogenicity and therefore pose a health risk for people on Earth and for astronauts aboard the International Space Station (ISS) (1, 2). The latter is of concern for longduration missions, as astronauts have been shown to be immunocompromised (3). Importantly, these bacteria are able to adapt to extreme conditions such as microgravity and radiation and thus persist, necessitating the development of appropriate countermeasures to control them. Members of the order Enterobacteriales that were found on ISS surfaces were Pantoea brenneri, Pantoea agalomerans, Kalamiella piersonii, and Enterobacter bugandensis (4-6). On Earth, P. agglomerans and P. brenneri were reported to have been isolated from human infections (4). K. piersonii is a member of a novel genus in the family Erwiniaceae that has exhibited resistance to multiple clinical drugs, such as penicillin and vancomycin, allowing it to be an emerging pathogen (5). E. bugandensis was documented from blood as a causative agent of septicemia in various geological locations (7). Analyses of draft genome assemblies for these species might pave the way to identify the genetic processes responsible for potential pathogenicity, as previously reported for some of these strains (5, 6).

The strains used for whole-genome sequencing (WGS) were isolated from four different locations in the ISS across three flights and are detailed in Table 1 (8). The ISS surface samples collected and brought back to Earth were aseptically handled, suitable aliquots of the sample concentrate (100 μ l) were plated onto Reasoner's 2A (R2A) medium and incubated at 25°C for 7 days, and a single well-isolated colony was archived at -80°C until DNA extraction. DNA was extracted from cultures grown in R2A medium using the ZymoBIOMICS DNA MagBead kit according to the manufacturer's instructions.

WGS of 26 bacterial isolates from the ISS was performed using the Illumina Nextera Flex protocol for library preparation, as used in similar studies (6). The NovaSeq 6000 system with an S4 flow cell (paired-end 2 imes 150-bp reads) was used to execute paired-end sequencing. FastQC (v0.11.7) was used to validate the quality of the raw

Citation Bharadwaj AR, Daudu R, Singh NK, Wood JM, Debieu M, O'Hara NB, Karouia F, Mason CE, Venkateswaran K. 2020. Draft genome sequences of Enterobacteriales strains isolated from the International Space Station. Microbiol Resour Announc 9:e00817-20. https://doi.org/10.1128/MRA.00817-20.

Editor David Rasko, University of Maryland School of Medicine

This is a work of the U.S. Government and is not subject to copyright protection in the United States. Foreign copyrights may apply.

Address correspondence to Kasthuri Venkateswaran, kivenkat@ipl.nasa.gov.

Received 15 July 2020 Accepted 13 August 2020 **Published** 10 September 2020

^bBiotia, New York, New York, USA

Department of Cell Biology, College of Medicine, SUNY Downstate Health Sciences University, Brooklyn, New York, USA

^dBlue Marble Space Institute of Science, Exobiology Branch, NASA Ames Research Center, Moffett Field, California, USA

eSpace Research Within Reach, San Francisco, California, USA

WorldQuant Initiative for Quantitative Prediction, Weill Cornell Medicine, New York, New York, USA

⁹Department of Physiology and Biophysics, Weill Cornell Medicine, New York, New York, USA

TABLE 1 Metadata and genome statistics for Enterobacter, Kalamiella, and Pantoea strains isolated from various ISS environmental surfaces during the Microbial Tracking 1 flight project

.												
Sample	Nearest species GenBank	GenBank	Raw sequence	Flight(s) or	Sampling	No. of	Genome		Median	No. of quality-	No. of raw	0+C
name	identity ^a	accession no.	accession no.	facility ⁶	location	$contigs^d$	size (bp)	N ₅₀ (bp)	coverage (x)	controlled reads	reads (×10 ⁶)	content (%)
IF2SW-B4	E. bugandensis	JABWOY0000000000	SRR11885007	F1-2	WHC	36	4,892,220	511,556	657.59	34,787,506	17.5	55.9
IFACSW-B2	E. bugandensis	JABWOX000000000	SRR11885006	F1	Z	40	4,892,163	481,191	662.95	35,107,666	17.6	55.9
IFACSW-B4	E. bugandensis	JABWOW0000000000	SRR11885005	F1	FC	35	4,892,584	808,304	006.70	31,965,988	16.0	55.9
IFACSW-B5	E. bugandensis	JABWOV00000000000	SRR11885004	F1	FC.	37	4,891,741	481,191	672.32	35,130,172	17.7	55.9
IFACSW-P1	E. bugandensis	JABWOU0000000000	SRR11885003	F1	FC.	36	4,891,763	808,252	640.18	35,538,578	17.9	55.9
IF2SW-F2	E. bugandensis	JACBPD0000000000	SRR12071883	F1-2	WHC	25	4,892,159	511,419	547.77	29,062,046	14.5	55.9
IF2SW-F3	E. bugandensis	JACBPE0000000000	SRR12071879	F1-2	WHC	22	4,892,298		518.30	27,421,928	13.7	55.9
F3-6B(4)	K. piersonii	JACBPM00000000000	SRR12071882	F3-6	PMM	39	4,850,268	503,530	313.39	17,045,882	8.5	57.1
F3-6B(5)	K. piersonii	JACBPN0000000000	SRR12071881	F3-6	PMM	50	4,850,704	310,993	411.16	24,040,576	12.0	57.1
IIIF_BACT_A	K. piersonii	JACBPO0000000000	SRR12071880	F3-6	PMM	42	4,849,373	503,411	441.96	22,790,000	11.4	57.1
FJII-L5-SW-P2	P. agglomerans	JACBPL0000000000	SRR12071872	JPL SAF II	Cleanroom floor	26	4,861,660	445,707	413.84	23,353,202	11.7	55.1
IF5SW-B1	P. brenneri	JABWPM0000000000	SRR11885013	F1-5	N1-04	108	5,022,545	216,403	487.50	27,782,700	14.0	55.9
IF5SW-B2	P. brenneri	JABWPL0000000000	SRR11885012	F1-5	N1-04	107	5,023,215	216,403	372.32	20,371,906	10.2	55.9
IFACSW-B3	P. brenneri	JABWPK0000000000	SRR11885002	F1	FC	108	5,023,154	216,403	549.11	31,932,976	16.0	55.9
IF5SW-P1	P. brenneri	JABWPJ0000000000	SRR11885001	F1-5	N1-04	106	5,023,034	216,403	534.38	32,864,806	16.5	55.9
IF5SW-P2	P. brenneri	JABWP10000000000	SRR11885000	F1-5	N1-04	106	5,023,268	216,131	586.61	37,186,174	18.7	55.9
IFACSW-P2	P. brenneri	JABWPH0000000000	SRR11884999	F1	FC	108	5,023,383	216,403	553.13	34,845,798	17.5	55.9
IIF5SW-B1	P. brenneri	JABWPG000000000	SRR11884998	F1-5	N1-04	106	5,022,674	216,137	436.61	26,223,526	13.2	55.9
IIF5SW-B2	P. brenneri	JABWPF0000000000	SRR11884997	F1-5	N1-04	106	5,023,080	216,403	460.71	27,740,334	13.9	55.9
IIF5SW-B5	P. brenneri	JABWPE0000000000	SRR11884996	F1-5	N1-04	111	5,021,866	176,626	366.96	22,097,600	11.1	55.9
IIF5SW-P1	P. brenneri	JACBPF0000000000	SRR12071878	F2-5	N1-04	75	5,020,903	176,626	444.64	26,701,110	13.4	55.9
IIF5SW-P2	P. brenneri	JACBPG0000000000	SRR12071877	F2-5	N1-04	75	5,022,945	216,403	451.34	26,675,346	13.4	55.9
IIF5SW-P3	P. brenneri	JACBPH0000000000	SRR12071876	F2-5	N1-04	75	5,021,950	216,403	459.38	26,806,520	13.5	55.9
IIF5SW-P4	P. brenneri	JACBP10000000000	SRR12071875	F2-5	N1-04	75	5,023,133	216,403	586.61	35,389,420	17.8	55.9
IIF5SW-P5	P. brenneri	JACBPJ0000000000	SRR12071874	F2-5	N1-04	75	5,023,087	216,131	510.27	29,693,196	14.9	55.9
IIFCSG-B1	P. brenneri	JACBPK0000000000	SRR12071873	CRV2	CRV-FC	74	5,022,524	216,403	345.54	20,467,790	10.3	55.9

and part of Strains were retrieved from the whole-genome sequence of the queried genome and analyzed with BLAST against type strains for all 16S rRNA sequences in the NCBI database. The bacterial species identity was determined when the queried sequence showed >97.5% similarity to the 165 rRNA gene sequence of the type strain (E. bugandersis DSM 29888T, K. piersonii DSM 108198T, P. agglomerans DSM 34931, or P. brenneri DSM 242321). The whole-genome sequence of the nearest neighbor was further selected for ANI evaluation. The ANI value for all strain comparisons was 99%.

" Hyphenated designations indicate the flight number followed by the location; for example, F1-2 indicates flight 1 and location 2. JPL, Jet Propulsion Laboratory; SAF, Spacecraft Assembly Facility; CRV, commercial

· WHC, waste and hygiene compartment; PMM, permanent multipurpose module; FC, field control (a sampling wipe was exposed to the air for 120 s at the center of node 2); N1-O4, node 1 overhead 4. resupply vehicle.

^dContigs that were less than 200 nucleotides long were not analyzed.

Volume 9 Issue 37 e00817-20 mra.asm.org **2**



sequencing data (9). Adapter trimming and quality filtering were carried out using the software fastp (v0.20.0) to perform quality control (10). The cleaned sequences were assembled using SPAdes (v3.11.1) (11). The N_{50} values, numbers of contigs, and total genome lengths were generated using QUAST (v5.0.2) and used to assess the quality of the final assembly (12). The average nucleotide identity (ANI) values were calculated by comparing all strains to their respective type strains, and their taxonomic affiliations and genome statistics are given in Table 1 (13). The NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (v.4.11 and v.4.12) was used for genome annotation. Default parameters were used for all software.

Data availability. This WGS project was deposited in DDBJ/ENA/GenBank (accession numbers are given in Table 1 [BioProject accession no. PRJNA635942]) and also deposited in the NASA GeneLab database (accession no. GLDS-302 and GLDS-311). The versions described in this paper are the first versions.

ACKNOWLEDGMENTS

We thank astronauts Captain Terry Virts and Commander Jeffrey Williams for collecting samples aboard the ISS and the Implementation Team at NASA Ames Research Center for coordinating this effort. We thank Ryan Kemp (Zymo Corp.) for extracting DNA and Dan Butler (Weill Cornell Medicine) for shotgun sequencing using the NovaSeq platform.

Part of the research described in this publication was carried out at the Jet Propulsion Laboratory (JPL), California Institute of Technology, under a contract with NASA. Government sponsorship is acknowledged. This research was funded by 2012 Space Biology project NNH12ZTT001N grant 19-12829-26 under task order NNN13D111T awarded to K.V.; this also funded a postdoctoral fellowship for J.M.W., a JPL graduate fellowship for R.D., and a subcontract to Biotia, Inc.

REFERENCES

- Christoff AP, Sereia AFR, Cruz GNF, Bastiani DC, Silva VL, Hernandes C, Nascente APM, Reis AA, Viessi RG, Marques ASP, Braga BS, Raduan TPL, Martino MDV, Menezes FG, Oliveira LFV. 2020. One year cross-sectional study in adult and neonatal intensive care units reveals the bacterial and antimicrobial resistance genes profiles in patients and hospital surfaces. PLoS One 15:e0234127. https://doi.org/10.1371/journal.pone.0234127.
- Wong WC, Oubre C, Mehta SK, Ott CM, Pierson DL. 2017. Preventing infectious diseases in spacecraft and space habitats, p 3–17. In Hurst CJ (ed), Modeling the transmission and prevention of infectious disease. Springer, Cham, Switzerland. https://doi.org/10.1007/978-3 -319-60616-3_1.
- Taylor PW. 2015. Impact of space flight on bacterial virulence and antibiotic susceptibility. Infect Drug Resist 8:249–262. https://doi.org/10 .2147/IDR.S67275.
- Cruz AT, Cazacu AC, Allen CH. 2007. Pantoea agglomerans, a plant pathogen causing human disease. J Clin Microbiol 45:1989–1992. https://doi.org/10 .1128/JCM.00632-07.
- Singh NK, Wood JM, Mhatre SS, Venkateswaran K. 2019. Metagenome to phenome approach enables isolation and genomics characterization of *Kalamiella piersonii* gen. nov., sp. nov. from the International Space Station. Appl Microbiol Biotechnol 103:4483–4497. https://doi.org/10 .1007/s00253-019-09813-z.
- Singh NK, Bezdan D, Checinska Sielaff A, Wheeler K, Mason CE, Venkateswaran K. 2018. Multi-drug resistant *Enterobacter bugandensis* species isolated from the International Space Station and comparative genomic analyses with human pathogenic strains. BMC Microbiol 18:175. https://doi.org/10.1186/s12866-018-1325-2.

- Doijad S, Imirzalioglu C, Yao Y, Pati NB, Falgenhauer L, Hain T, Foesel BU, Abt B, Overmann J, Mirambo MM, Mshana SE, Chakraborty T. 2016. Enterobacter bugandensis sp. nov., isolated from neonatal blood. Int J Syst Evol Microbiol 66:968–974. https://doi.org/10.1099/ijsem.0.000821.
- Checinska Sielaff A, Urbaniak C, Mohan GBM, Stepanov VG, Tran Q, Wood JM, Minich J, McDonald D, Mayer T, Knight R, Karouia F, Fox GE, Venkateswaran K. 2019. Characterization of the total and viable bacterial and fungal communities associated with the International Space Station surfaces. Microbiome 7:50. https://doi.org/10.1186/s40168-019-0666-x.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. http://www.bioinformatics.babraham.ac.uk/projects/ fastqc.
- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. https://doi.org/10.1093/ bioinformatics/bty560.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10.1093/bioinformatics/btt086.
- Yoon S-H, Ha S, Lim J, Kwon S, Chun J. 2017. A large-scale evaluation of algorithms to calculate average nucleotide identity. Antonie Van Leeuwenhoek 110:1281–1286. https://doi.org/10.1007/s10482-017-0844-4.

Volume 9 Issue 37 e00817-20 mra.asm.org **3**