





Draft Genome Sequences of Fungi Isolated from the International Space Station during the Microbial Tracking-2 **Experiment**

Anna C. Simpson, a Camilla Urbaniak, a John R. Bateh, a Nitin K. Singh, a 📵 Jason M. Wood, a* Marilyne Debieu, b 📵 Niamh B. O'Hara, b.c DJos Houbraken, d Christopher E. Mason, e,f D Kasthuri Venkateswarana

^aJet Propulsion Laboratory, California Institute of Technology, Pasadena, California, USA

ABSTRACT As part of the Microbial Tracking-2 study, 94 fungal strains were isolated from surfaces on the International Space Station, and whole-genome sequences were assembled. Characterization of these draft genomes will allow evaluation of microgravity adaption, risks to human health and spacecraft functioning, and biotechnological applications of fungi.

ungi are potential sources of nutrients and bioactive compounds during long-term spaceflight but also could affect astronaut health through both opportunistic infections and system biofouling (1, 2). As part of a study characterizing fungal responses to the space environment, we report the draft genomes of 94 fungal strains that were isolated from the International Space Station (ISS), representing 10 ascomycetous and 1 basidiomycetous

Aspergillus species are environmental fungi and opportunistic pathogens (3). Aspergillus unquis is a member of the ISS microbiome (4) and produces industrially important compounds (5). Aureobasidium pullulans is a black fungus that was previously isolated from the ISS water filtration system (6) and Mars mission spacecraft-associated surfaces (7).

Cladosporium species are dominant fungal contaminants in indoor air (8, 9). Cladosporium sphaerospermum and Cladosporium cladosporioides were detected multiple time on the ISS, and their properties in microgravity were studied (10, 11).

Fusarium veterinarium is a recently described species within the Fusarium oxysporum complex, the species of which are ubiquitous in soil, are known human/plant pathogens (12), and were isolated both from surfaces and from infected Zinnia hybrida plants aboard the ISS (13, 14). Fusarium annulatum, which has been isolated from plant and human tissues on Earth, has not been reported previously in space (15).

Penicillium species produce important bioactive compounds and can contaminate food and cause secondary infections (16). Previously detected on the ISS or Mir (2, 11) are Penicillium citrinum, a common soil and indoor species (17), Penicillium rubens, from which penicillin was isolated (18), and Penicillium corylophilum, which is commonly found in damp buildings (19). Penicillium palitans, which has been reported in cheese (20) and also in a wide range of habitats, including Antarctica (21), has not been reported previously in space.

Rhodotorula mucilaginosa is a ubiquitous environmental (22) and human commensal yeast and opportunistic pathogen (23) that is found in aquatic and built environments, including bathrooms and dishwashers (24, 25). It is the most commonly isolated yeast on the ISS (26-28).

Citation Simpson AC, Urbaniak C, Bateh JR, Singh NK, Wood JM, Debieu M, O'Hara NB, Houbraken J, Mason CE, Venkateswaran K. 2021. Draft genome sequences of fungi isolated from the International Space Station during the Microbial Tracking-2 experiment. Microbiol Resour Announc 10:e00751-21. https://doi.org/10.1128/MRA.00751-21.

Editor Vincent Bruno, University of Maryland School of Medicine

Copyright © 2021 Simpson et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Kasthuri Venkateswaran, kjvenkat@jpl.nasa.gov.

* Present address: Jason M. Wood, Research Informatics Core, University of Illinois at Chicago, Chicago, Illinois, USA.

Received 22 July 2021 Accepted 14 August 2021 Published 16 September 2021

bBiotia, New York, New York, USA

cDepartment of Cell Biology, College of Medicine, SUNY Downstate Health Sciences University, Brooklyn, New York, USA

^dWesterdijk Fungal Biodiversity Institute, Utrecht, the Netherlands

eDepartment of Physiology and Biophysics, Weill Cornell Medicine, New York, New York, USA

WorldQuant Initiative for Quantitative Prediction, Weill Cornell Medicine, New York, New York, USA

TABLE 1 Sampling locations, genetic loci used for taxonomic analysis, and WGS assembly quality for fungal species isolated from the ISS during the Microbial Tracking-2 mission

		:		SRA	:	i		:	,			Q+C	No. of
Sample name	Fungal species	Loci used for identification	WGS accession no.	accession no.	Medium and temperature	Flight no.	Location description ^c	No. of contigs	Genome size (bp)	N _{so} (bp)	Coverage depth (×)	content (%)	filtered reads
F6_8S_P_2A	Aspergillus unguis	benA, CaM	JAGUQD000000000	SRR14342084	BA, 37°C	F6	Crew quarters	22	25,891,216	2,495,528	84.28	50.30	14,548,376
F6_85_P_4A	Aspergillus unguis	benA, CaM	JAGUQC000000000	SRR14342083	8A, 37°C	1 2	Crew quarters	19	25,892,532	2,741,542	179.59	50.30	30,999,366
F/_65_YPU	Aureobasialum pullulans	<u> </u>	JAGUPWOOOOOOO	SKK143420/2	YPD, 25-C	1 1	PPIM port I	50.2	28,552,932	171,111	17:71	50.35	800,555,51
F7_35_TFD	Aureobasidium pullulans	<u> </u>	1AG LIPYOOOOOOOO	SPN 14542075	VPD, 25 C	2 6	WILD	0,11	20,340,471	060,010	11.05	50.33	10,320,034
F7_23_1FD F7_1S_VPD	Aureobasidium pullulans	ΞE	IAGIIDADDOOOOOOO	SRR14342074	VPD 25°C	F7	Cinola	103	28,343,476	847,007	148.55	50.33	22,07,3,034
F7 2A YPD	Aureobasidium pullulans	ΞΞ	IAGIIPZOOOOOOO	SRR14342075	VPD 25°C	F7	NHO CHA	10.2	28 545 383	869 723	90 19	50.35	18 158 278
F7 1A YPD	Aureobasidium pullulans	TIS	JAGUOB000000000	SRR14342077	YPD, 25°C	F7	Cupola	66	28.555,273	793,969	102.69	50.34	20,674.178
F6 15 B 1B	Aureobasidium pullulans	LIS	JAGUOJ0000000000	SRR14342071	R2A, 25°C	. F6	Cupola	162	28.763,896	746,636	72.11	50.32	14,517,038
F6 15 P 3A	Aureobasidium pullulans	ITS	JAGUQ10000000000	SRR14342125	BA, 37°C	F6	Cupola	142	28,780,047	881,182	177.55	50.32	35,744,148
F6 45 B 1	Aureobasidium pullulans	ITS	JAGUQE000000000	SRR14342118	R2A, 25°C	F6	Dining table	140	28,772,883	884,258	97.41	50.32	19,610,724
F6_3S_1A_F	Aureobasidium pullulans	ITS	JAGUQH0000000000	SRR14342124	PDA, 25°C	F6	ARED	156	28,770,985	819,420	133.99	50.32	26,975,786
F6 35 1B F	Aureobasidium pullulans	ITS	JAGUOGOOOOOOOO	SRR14342123	PDA, 25°C	F6	ARED	184	28,720,136	734,664	106.49	50.35	21,439,104
F6 3S 1C F	Aureobasidium pullulans	ITS	JAGUOF000000000	SRR14342122	PDA, 25°C	F6	ARED	188	28,737,399	748,713	78.37	50,33	15,778,598
F8 55 2F	Cladosporium cladosporioides	191	JAGUPV0000000000	SRR14342051	PDA, 25°C	F8	Overhead 4	130	34,025,119	1.102.510	120.93	52.59	26,786,080
F8 55 3F	Cladosporium cladosporioides	TFF	IAGLIPLIOODOOOOO	SRR14342048	PDA 25°C	8	Overhead 4	132	34 027 712	965 801	122.78	52.59	27 195 854
F8 55 4F	Cladosporium cladosporioides	747	IAGUIPTOOOOOOO	SRR14342047	PDA 25°C	. 8	Overhead 4	23.4	33 871 387	634 743	11102	52.52	24 592 420
F 20 - 21	Cladosporium	111	OCCOCCOCCION IN THE PROPERTY OF THE PROPERTY O	SEDIASASISE	2.52, VOI	2 5	lab a	573	30,616,838	079 279	135.80	53.05	21,27,27,27
L4_/3_F1_F	Cidaosporiam	101	000000000000000000000000000000000000000	34714347170	LDA, 23 C	ŧ	Labo	0/0	000,010,00	6,0,0,0	133.00	50.65	01/'046'47
00	spnderospermum	, 000	000000000000000000000000000000000000000	01004041000	7010	S.	Overnead	71.0	0000	1 075	p 4 C 4	70.77	011 707 10
F8_45_2B	Fusarium annulaturn	TEF, RPBZ	JAHAFROUGOOOOO	SKK14342059	R2A, 25 C	0 0	Dining table	6/7	45,009,810	1,8/3,/02	42.45	48.32	21,480,110
F8_45_3B	Fusarium annulatum	IEF, RPBZ	JAHAPP000000000	SKK14342057	R2A, 25°C	8 1	Dining table	2/3	45,012,509	1,556,966	51.28	48.32	25,765,430
F8_45_4P	Fusarium annulatum	TEF, RPB2	JAHAPN00000000000	SRR14342055	BA, 37°C	82	Dining table	283	45,010,147	1,599,633	40.99	48.32	20,595,810
F8_45_5P	Fusarium annulatum	TEF, RPB2	JAHAPL000000000	SRR14342053	BA, 37°C	F8	Dining table	309	45,001,499	1,598,178	47.32	48.33	23,616,132
F8_45_1F	Fusarium annulatum	TEF, RPB2	JAHAPT000000000	SRR14342062	PDA, 25°C	F8	Dining table	341	44,584,936	1,556,345	51.36	48.62	25,159,054
F5_8S_1A_F	Fusarium veterinarium	TEF	JAHARR0000000000	SRR14342093	PDA, 25°C	F5	Crew quarters	861	48,079,010	325,419	58.78°	47.63	31,171,132
F5_8S_1B_F	Fusarium veterinarium	TEF	JAHARQ0000000000	SRR14342082	PDA, 25°C	F5	Crew quarters	925	47,312,867	276,643	44.14	48.09	23,238,836
F4_1A_F1_F	Penicillium citrinum	benA, CaM	JAHART0000000000	SRR14342127	PDA, 25°C	F4	Cupola	88	31,021,730	1,108,866	127.62	46.39	26,826,262
F5 1S 1A F	Penicillium corylophilum	benA, CaM	JAGUQL0000000000	SRR14342115	PDA, 25°C	F5	Cupola	53	28,229,796	1,725,911	104.56 ^d	50.20	32,206,152
F5 15 1B F	Penicillium corvlophilum	benA. CaM	JAGUOKOOOOOOOO	SRR14342104	PDA, 25°C	F5	Cupola	23	28,231,333	1,604,566	80.64°	50.20	24.805.488
F6 45 1A F	Penicillium palitans	hen A. CaM	IAHABMOOOOOOOO	SRR14342121	PDA, 25°C	. P	Dining table	798	36 471 070	281 544	57 4 ^d	47.70	23,233,174
F6 45 18 F	Penicillium palitans	benA. CaM	JAHARI 000000000	SRR14342120	PDA. 25°C	F6	Dining table	806	36.468.364	273.565	58.94°	47.70	24.107.708
F6 45 1C F	Penicillium palitans	hend CaM	IAHABKOOOOOOO	SRR14342119	PDA 25°C	F 6	Dining table	80.2	36 477 181	268 530	61 99°	47.70	25 189 046
E6 60 1 E	Donicillium politons	band Call	OCCOOL DATA PACION OCCOOL DATA	CDD14242117	2.52, VOI	2 4	DDM nort 1	808	36.467.611	268,530	64 479	07.74	040,001,02
E6 75 1A E	Posicillium palitans	ben / Call	000000000000000000000000000000000000000	CDD14242107	ראלי ארום	0 9	1.5 2	920	36 420 425	260,231	67.00d	07.74	C17 54 5C
	emember painting	לפווא, כמווא		10021-01-1110	7 7 7 7	2	Lab J	020	מהלימה לימה	100,002	90.	2:	21 //CFF/C2
1 7 7 7		44		70004041000	0	2	Overnedu	7	011111	701	p00 00	1	2000
r6_/>_IC_F	Penicillum palitans	benA, CaM	JAHAQQOOOOOOOO	SKK14342096	PDA, 25°C	ō.	Lab 3	/94	36,471,119	303,786	-60.69	4/./0	28,047,394
		7		100000000000000000000000000000000000000	0	2	overnead	0	000	0	11	1	000
F6_85_1A_F	Penicillium palitans	benA, CaM	JAHAQIOOOOOOOO	SKK14342087	PDA, 25°C	۲ ۲	Crew quarters	878	36,453,009	268,529	5/.59	47.70	23,229,736
F6_85_1C_F	Penicillium palitans	benA, CaM	JAHAQH0000000000	SRR14342086	PDA, 25°C	F6	Crew quarters	850	36,588,697	255,978	57.23	47.70	22,858,702
F8_65-1F	Penicillium palitans	benA, CaM	JAHAO 200000000000	SRR14342036	PDA, 25°C	82 1	PPM port 1	987	36,400,448	262,197	53.41	47.80	21,366,974
F8_65_2F	Penicillium palitans	benA, CaM	JAHAPD0000000000	SRR14342041	PDA, 25°C	8 1	PPM port 1	943	36,503,579	273,760	70.93	47.76	28,334,002
F8_65-3F	Penicillium palitans	benA, CaM	JAHAO Y00000000000	SKK14342035	PDA, 25°C	20	PPM port 1	66/	36,607,893	290,808	60.5	47.69	24,779,924
F8_65-4F	Penicillium palitans	benA, CaM	JAHAO X00000000000	SKK14342034	PDA, 25°C	8 i	PPM port 1	827	36,605,405	290,729	60.84	47.69	24,966,852
F8_65_5F	Penicillium palitans	benA, CaM	JAHAPC0000000000	SRR14342040	PDA, 25°C	82 82	PPM port 1	808	36,607,101	281,504	73.76	47.70	30,172,808
F8_65_6F	Penicillium rubens	benA, CaM	JAHAPB0000000000	SRR14342039	PDA, 25°C	8 1	PPM port 1	182	30,459,701	858,760	118.82	49.00	26,445,782
F8_65_7F	Penicillium rubens	benA, CaM	JAHAPA0000000000	SRR14342037	PDA, 25°C	8 1	PPM port 1	469	31,558,626	716,780	112.63	48.97	25,069,320
F6_45_B_2B	Rhodotorula mucilaginosa	IIS	JAHARI0000000000	SRR14342116	R2A, 25°C	9 1	Dining table	199	20,171,565	432,962	132.05	60.55	17,800,488
F6_85_B_1B	Rhodotorula mucilaginosa	E E	JAHAQG000000000	SRR14342085	K2A, 25°C	9 Y	Crew quarters	209	20,172,702	367,952	215.04	60.55	28,987,090
F6_85_P_5A	Khodotorula mucilaginosa	SI	JAHAQF000000000	SRR14342081	BA, 37°C	9 L	Crew quarters	199	20,077,383	389,367	212.72	59.99	28,6/4,/16
F6_85_P_5B	Khodotorula mucilaginosa	SI E	JAHAQE000000000	SRR14342080	BA, 37°C	P 7	Crew quarters	194	20,074,587	329,841	244.78	59.99	32,995,640
F0_85_P_0A	knodotorula mucilaginosa	<u> </u>	JAHAQD000000000	SKK14342079	5A, 3/ C	0 7	Crew quarters	201	20,175,089	456,573	227.88	60.55	30,717,298
F0_03_F_0B	phodotorula mucilaginosa	S E	IAHADIOOOOOOOO	SPD14342076	BA 37°C	2 α	Overhead 4	202	20,109,473	317,008	22.49	60.33	21 212 072
F8 55 5D	Rhodotorula musilaginosa	ΞĽ	IAHAPHOOOOOOOO	SBB14342045	BA 37°C	2 8	Overhead 4	105	20,109,007	319 540	215.19	60.53	2,0,515,15
F8 55 6P	Rhodotorula mucilaainosa	Ξ	IAHAPGOOOOOOO	SRR14342044	BA 37°C	2 8	Overhead 4	190	20,113,827	354 782	251.44	60.53	33 893 490
F6 15 P 1A	Rhodotorula mucilaginosa	ITS	JAHARP0000000000	SRR14342060	BA, 37°C	. Pe	Cupola	179	20,117,026	392,515	180,21	60.53	24,292,752
F6_1S_P_1B	Rhodotorula mucilaginosa	ITS	JAHARO0000000000	SRR14342049	BA, 37°C	F6	Cupola	182	20,114,730	392,620	225.92	60.53	30,453,336
F6_15_P_1C	Rhodotorula mucilaginosa	ITS	JAHARN0000000000	SRR14342038	BA, 37°C	F6	Cupola	200	20,115,701	322,180	218.74	60.53	29,485,618
F6_4S_B_2A	Rhodotorula mucilaginosa	ITS	JAHARJ000000000	SRR14342117	R2A, 25°C	F6	Dining table	172	19,998,495	334,411	207.28	60.55	27,941,724
F6_45_B_2C	Rhodotorula mucilaginosa	S	JAHARH0000000000	SRR14342114	R2A, 25°C	F6	Dining table	200	20,107,055	319,541	200.96	60.53	27,089,466
F6_45_P_3B	Rhodotorula mucilaginosa	TS	JAHARG000000000	SRR14342113	BA, 37°C	2 2	Dining table	184	20,108,582	331,731	105.50	60.53	14,221,722
F6_45_P_3C	Knodotorula mucilaginosa	SII	JAHAKFOOOOOOOO	SKK14342112	BA, 3 / ℃	Pb	Dining table	196	7/0/111/07	31/,326	150.66	60.53	20,308,150
											<u>y</u>	(Continued on next page)	next page)
											•		, n

TABLE 1 (Continued)

Sample		Loci used for	SDM	accession	Mediumand	Flight	Location	No of	Genome		Coverage	content	filtered
name	Fungal species	identification	accession no.	no.	temperature	no.	description	contigs	size (bp)	N _{so} (bp)	depth (×)	(%)	reads
F6_4S_P_4A	Rhodotorula mucilaginosa	ITS	JAHARE0000000000	SRR14342111	BA, 37°C	F6	Dining table	175	20,126,113	410,825	121.23	60.52	16,341,478
F6_4S_P_4B	Rhodotorula mucilaginosa	ITS	JAHARD0000000000	SRR14342110	BA, 37°C	F6	Dining table	206	20,115,791	297,628	230.74	60.53	31,103,222
F6_4S_P_5A	Rhodotorula mucilaginosa	ITS	JAHARC0000000000	SRR14342109	BA, 37°C	F6	Dining table	195	20,113,572	334,410	220.75	60.53	29,757,146
F6_4S_P_5B	Rhodotorula mucilaginosa	ITS	JAHARB0000000000	SRR14342108	BA, 37°C	F6	Dining table	210	20,109,987	323,162	141.10	60.53	19,019,728
F6_6S_B_1A	Rhodotorula mucilaginosa	ITS	JAHAQZ000000000	SRR14342106	R2A, 25°C	F6	PPM port 1	202	20,109,799	329,302	205.40	60.53	27,687,960
F6_6S_B_1B	Rhodotorula mucilaginosa	ITS	JAHAQY000000000	SRR14342105	R2A, 25°C	F6	PPM port 1	199	20,106,114	331,797	130.16	60.53	17,545,556
F6_6S_B_1C	Rhodotorula mucilaginosa	ITS	JAHAQX000000000	SRR14342103	R2A, 25°C	F6	PPM port 1	199	20,115,093	335,579	133.73	60.53	18,026,650
F6_6S_P_1A	Rhodotorula mucilaginosa	ITS	JAHAQW0000000000	SRR14342102	BA, 37°C	F6	PPM port 1	202	20,108,679	319,902	114.06	60.53	15,375,556
F6_6S_P_1B	Rhodotorula mucilaginosa	ITS	JAHAQV0000000000	SRR14342101	BA, 37°C	F6	PPM port 1	197	20,116,515	323,169	233.47	60.53	31,471,482
F6 6S P 1C	Rhodotorula mucilaginosa	ITS	JAHAQU0000000000	SRR14342100	BA, 37°C	F6	PPM port 1	190	20,114,955	329,677	145.50	60.53	19,613,002
F6_6S_P_2A	Rhodotorula mucilaginosa	ITS	JAHAQT000000000	SRR14342099	BA, 37°C	F6	PPM port 1	191	20,108,060	323,034	240.93	60.53	32,476,452
F6_6S_P_2B	Rhodotorula mucilaginosa	ITS	JAHAQS000000000	SRR14342098	BA, 37°C	F6	PPM port 1	194	20,117,485	323,058	209.85	60.52	28,287,446
F6_75_B_2A	Rhodotorula mucilaginosa	ITS	JAHAQP0000000000	SRR14342095	R2A, 25°C	F6	Lab 3	183	20,055,075	293,584	240.80	60.55	32,459,440
F6 75 B 2B	Rhodotoula mucilaainosa	ZII	IAHAOOOOOOOOO	SRR14342094	B2A 25°C	F6	overhead	192	20 049 052	793 667	13813	60.55	18 619 460
		2					overhead	1					
F6_75_B_2C	Rhodotorula mucilaginosa	ITS	JAHAQN0000000000	SRR14342092	R2A, 25°C	F6	Lab 3	196	20,050,303	292,236	107.06	60.55	14,431,658
F6_7S_P_6B	Rhodotorula mucilaginosa	ITS	JAHAQM0000000000	SRR14342091	BA, 37°C	F6	overhead Lab 3	164	20,065,557	369,724	234.38	60.55	31,594,198
A C O O O	Olombra of more of o	Ë	000000000000000000000000000000000000000	000000000000000000000000000000000000000	0,000	Ž	overhead	9	01000	700 110	50,100	11	230 300 50
LO_/3_L_/A	nitodotolaid macingimosa	2	מממממממקלישואר	06024501 NUC	7 /5 /20	2	Cabo	661	20,032,030	106,116	76:107	55.00	006,022,12
F6_7S_P_7B	Rhodotorula mucilaginosa	ITS	JAHAQK0000000000	SRR14342089	BA, 37°C	F6	Overnead Lab 3	198	20,056,236	284,145	169.60	60.55	22,862,414
		į				ì	overhead					;	
F6_75_P_7C	Rhodotorula mucilaginosa	ITS	JAHAQJ00000000000	SRR14342088	BA, 37°C	P6	Lab 3	190	20,058,305	320,873	233.38	60.55	31,459,534
F8 15 2B	Rhodotorula mucilaainosa	ITS	JAHAOB0000000000	SRR14342070	R2A. 25°C	82	Cupola	172	19.998.942	329.308	180.71	60.55	24.358.996
F8 15 3B	Rhodotorula mucilaginosa	ITS	JAHAQA0000000000	SRR14342069	R2A, 25°C	. &	Cupola	197	20,118,130	319,542	217.40	60.53	29,305,254
F8_35_1B	Rhodotorula mucilaginosa	ITS	JAHAPZ0000000000	SRR14342068	R2A, 25°C	84 84	ARED	197	20,118,712	325,275	190.77	60.53	25,716,020
F8_3S_2B	Rhodotorula mucilaginosa	ITS	JAHAPX0000000000	SRR14342066	R2A, 25°C	82	ARED	200	20,109,774	323,187	181.13	60.53	24,415,874
F8_3S_3B	Rhodotorula mucilaginosa	ITS	JAHAPV0000000000	SRR14342064	R2A, 25°C	F8	ARED	189	20,104,868	317,674	194.41	60.53	26,206,654
F8_45_4B	Rhodotorula mucilaginosa	ITS	JAHAPO00000000000	SRR14342056	R2A, 25°C	F8	Dining table	172	19,998,636	338,336	224.66	60.54	30,283,760
F8_4S_5B	Rhodotorula mucilaginosa	ITS	JAHAPM00000000000	SRR14342054	R2A, 25°C	F8	Dining table	172	19,999,954	323,188	197.65	60.55	26,642,384
F8_55_2B	Rhodotorula mucilaginosa	ITS	JAHAPK0000000000	SRR14342052	R2A, 25°C	F8	Overhead 4	189	20,113,164	335,288	219.00	60.53	29,520,936
F8_55_3B	Rhodotorula mucilaginosa	ITS	JAHAPJ0000000000	SRR14342050	R2A, 25°C	F8	Overhead 4	199	20,113,962	333,776	192.52	60.53	25,952,066
F8_65_1B	Rhodotorula mucilaginosa	ITS	JAHAPF0000000000	SRR14342043	R2A, 25°C	F8	PPM port 1	169	19,995,004	329,309	187.59	60.54	25,287,420
F8_65_2B	Rhodotorula mucilaginosa	ITS	JAHAPE0000000000	SRR14342042	R2A, 25°C	F8	PPM port 1	179	19,994,438	397,651	188.04	60.55	25,347,104
F8_35_1P	Rhodotorula mucilaginosa	ITS	JAHAPY0000000000	SRR14342067	BA, 37°C	F8	ARED	183	19,994,621	322,715	192.67	60.55	25,972,138
F8_3S_2P	Rhodotorula mucilaginosa	ITS	JAHAPW00000000000	SRR14342065	BA, 37°C	F8	ARED	167	19,991,553	415,431	121.35	60.54	16,358,020
F8_35_3P	Rhodotorula mucilaginosa	ITS	JAHAPU00000000000	SRR14342063	BA, 37°C	8	ARED	193	20,115,201	319,541	196.12	60.53	26,437,390
F8_45_1P	Rhodotorula mucilaginosa	ITS	JAHAPS0000000000	SRR14342061	BA, 37°C	F8	Dining table	171	19,999,629	322,714	190.79	60.55	25,717,964
F8 45 2P	Rhodotorula mucilaginosa	ITS	JAHAPQ00000000000	SRR14342058	BA, 37°C	<u>8</u> 2	Dining table	173	19,998,413	415,991	257.98	60.54	34,775,980

^b BA, blood agar; R2A, Reasoner's 2A agar; YPD, yeast extract-peptone-dextrose.
^c ARED, advanced resistive exercise device; WHC, waste and hygiene compartment; PMM, permanent multipurpose module.
^d Reference genome was not available; average sequencing depth was calculated from k-mer coverage.



Sample collection and fungal isolation steps were described elsewhere (26). For five flight missions, eight surfaces aboard the ISS were sampled with moistened polyester wipes (Table 1). Upon return to Earth, the wipes were agitated in sterile phosphate-buffered saline, which was concentrated using an InnovaPrep CP150 concentrating pipette, and suitable aliquots were spread onto nutrient media (Table 1). Fungal isolates were restreaked on potato-dextrose agar (PDA), and genomic DNA was extracted using the ZymoBlOMICS MagBead DNA kit according to the manufacturer's instructions. Whole-genome shotgun sequencing libraries were prepared with an Illumina Nextera DNA Flex library preparation kit (29) and were sequenced on the NovaSeq 6000 paired-end 2 \times 150-bp platform with a S4 flow cell. After quality filtering and trimming with FastQC v0.11.7 (30) and fastp v0.20.0 (31), genomes were assembled using SPAdes v3.11.1 (32). Assembly quality was assessed with QUAST v5.0.2 (33). Fastp included screening for 512 adapters; otherwise, default settings were used for all steps.

Genus-level identification was made via BLAST searches against the UNITE nuclear ribosomal internal transcribed spacer (ITS) database (34). Species identification was performed using specific loci suitable for species recognition (Table 1) (35). Homology searches were performed with extracted sequences against the NCBI nucleotide database and in-house Westerdijk Fungal Biodiversity Institute databases containing reference sequences; in case of doubt, identification was confirmed by constructing phylograms.

Data availability. The whole-genome sequences (WGSs) and raw data have been deposited in GenBank under BioProject accession number PRJNA723004. This project has also been deposited in the NASA GeneLab system (36) under project number GLDS-400. The version described in this paper is the first version.

ACKNOWLEDGMENTS

Part of the research described was carried out at the Jet Propulsion Laboratory (JPL) of the California Institute of Technology under a contract with NASA. This research was funded by a 2014 Space Biology NNH14ZTT002N award (grant 80NSSC18K0113) to Crystal Jaing and K.V., which also partially funded postdoctoral fellowships for C.U. and J.M.W. Additionally, A.C.S. was supported by grant 80NM0018D0004, funded to K.V.

We thank astronauts Colonel Jack Fischer, Colonel Mark Vande Hei, Norishige Kanai, and Alexander Gerst for collecting samples aboard the ISS, the implementation team (Fathi Karouia) at NASA Ames Research Center for coordinating this effort, and Crystal Jaing (Lawrence Livermore National Laboratory), principal investigator of the team. We thank Ryan Kemp (Zymo Corp.) for extracting the DNA and Dan Butler (Weill Cornell Medicine) for generating the shotgun sequencing. The JPL supercomputing facility staff is acknowledged, notably, Narendra J. Patel (Jimmy) and Edward Villanueva, for their continuous support in providing the best possible infrastructure for BIG-DATA analysis.

REFERENCES

- Romsdahl J, Blachowicz A, Chiang AJ, Chiang YM, Masonjones S, Yaegashi J, Countryman S, Karouia F, Kalkum M, Stajich JE, Venkateswaran K, Wang CCC. 2019. International Space Station conditions alter genomics, proteomics, and metabolomics in *Aspergillus nidulans*. Appl Microbiol Biotechnol 103:1363–1377. https://doi.org/10.1007/s00253-018-9525-0.
- Klintworth R, Reher H, Viktorov A, Bohle D. 1999. Biological induced corrosion of materials II: new test methods and experiences from MIR station. Acta Astronaut 44:569–578. https://doi.org/10.1016/s0094-5765(99)00069-7.
- Hogan LH, Klein BS, Levitz SM. 1996. Virulence factors of medically important fungi. Clin Microbiol Rev 9:469–488. https://doi.org/10.1128/CMR.9.4.469.
- Mora M, Wink L, Kogler I, Mahnert A, Rettberg P, Schwendner P, Demets R, Cockell C, Alekhova T, Klingl A, Krause R, Zolotariof A, Alexandrova A, Moissl-Eichinger C. 2019. Space Station conditions are selective but do not alter microbial characteristics relevant to human health. Nat Commun 10:3990. https://doi.org/10.1038/s41467-019-11682-z.
- 5. Molina G, Contesini F, De Melo R, Sato H, Pastore G. 2016. β -Glucosidase from *Aspergillus*, p 155–169. *In* Gupta V (ed), New and future developments in microbial biotechnology and bioengineering. Elsevier, Amsterdam, Netherlands.
- La Duc MT, Sumner R, Pierson D, Venkat P, Venkateswaran K. 2004. Evidence of pathogenic microbes in the International Space Station drinking

- water: reason for concern? Habitation (Elmsford) 10:39–48. https://doi.org/10.3727/154296604774808883.
- La Duc MT, Nicholson W, Kern R, Venkateswaran K. 2003. Microbial characterization of the Mars Odyssey spacecraft and its encapsulation facility. Environ Microbiol 5:977–985. https://doi.org/10.1046/j.1462-2920.2003.00496.x.
- Bensch K, Groenewald J, Meijer M, Dijksterhuis J, Jurjević Ž, Andersen B, Houbraken J, Crous PW, Samson R. 2018. *Cladosporium* species in indoor environments. Stud Mycol 89:177–301. https://doi.org/10.1016/ j.simyco.2018.03.002.
- Aihara M, Tanaka T, Takatori K. 2001. Cladosporium as the main fungal contaminant of locations in dwelling environments. Biocontrol Sci 6:49–52. https://doi .org/10.4265/bio.6.49.
- Singh NK, Blachowicz A, Romsdahl J, Wang C, Torok T, Venkateswaran K. 2017. Draft genome sequences of several fungal strains selected for exposure to microgravity at the International Space Station. Genome Announc 5:e01602-16. https://doi.org/10.1128/genomeA.01602-16.
- Satoh K, Yamazaki T, Nakayama T, Umeda Y, Alshahni MM, Makimura M, Makimura K. 2016. Characterization of fungi isolated from the equipment used in the International Space Station or Space Shuttle. Microbiol Immunol 60:295–302. https://doi.org/10.1111/1348-0421.12375.



mra.asm.org 5

- Laurence MH, Summerell BA, Burgess LW, Liew EC. 2014. Genealogical concordance phylogenetic species recognition in the *Fusarium oxysporum* species complex. Fungal Biol 118:374–384. https://doi.org/10.1016/j.funbio. 2014.02.002.
- Urbaniak C, van Dam P, Zaborin A, Zaborina O, Gilbert JA, Torok T, Wang CCC, Venkateswaran K. 2019. Genomic characterization and virulence potential of two Fusarium oxysporum isolates cultured from the International Space Station. mSystems 4:e00345-18. https://doi.org/10.1128/mSystems.00345-18.
- Schuerger AC, Amaradasa BS, Dufault NS, Hummerick ME, Richards JT, Khodadad CL, Smith TM, Massa GD. 2021. Fusarium oxysporum as an opportunistic fungal pathogen on Zinnia hybrida plants grown on board the International Space Station. Astrobiology https://doi.org/10.1089/ast.2020.2399.
- Khazaal FAK, Ameen MKM, Ali HA. 2019. Pathogenic fungi accompanied with sudden decline syndrome (wilting disease) of date palm tree (*Phoenix dactylifera* L.). Basrah J Sci 37:376–397.
- Visagie CM, Houbraken J, Frisvad JC, Hong SB, Klaassen CH, Perrone G, Seifert KA, Varga J, Yaguchi T, Samson RA. 2014. Identification and nomenclature of the genus *Penicillium*. Stud Mycol 78:343–371. https://doi.org/10.1016/j.simyco.2014.09.001.
- 17. Houbraken J, Frisvad JC, Samson R. 2011. Taxonomy of *Penicillium* section *Citrina*. Stud Mycol 70:53–138. https://doi.org/10.3114/sim.2011.70.02.
- Boruta T. 2018. Uncovering the repertoire of fungal secondary metabolites: from Fleming's laboratory to the International Space Station. Bioengineered 9: 12–16. https://doi.org/10.1080/21655979.2017.1341022.
- McMullin DR, Nsiama TK, Miller JD. 2014. Secondary metabolites from Penicillium corylophilum isolated from damp buildings. Mycologia 106: 621–628. https://doi.org/10.3852/13-265.
- 20. Kure C, Abeln E, Holst-Jensen A, Skaar I. 2002. Differentiation of *Penicillium commune* and *Penicillium palitans* isolates from cheese and indoor environments of cheese factories using M13 fingerprinting. Food Microbiol 19:151–157. https://doi.org/10.1006/fmic.2001.0473.
- de Menezes GCA, Porto BA, Amorim SS, Zani CL, de Almeida Alves TM, Junior PAS, Murta SMF, Simões JC, Cota BB, Rosa CA, Rosa LH. 2020. Fungi in glacial ice of Antarctica: diversity, distribution and bioprospecting of bioactive compounds. Extremophiles 24:367–376. https://doi.org/10 .1007/s00792-020-01161-5.
- Libkind D, Brizzio S, van Broock M. 2004. Rhodotorula mucilaginosa, a carotenoid producing yeast strain from a Patagonian high-altitude lake. Folia Microbiol (Praha) 49:19–25. https://doi.org/10.1007/BF02931640.
- Wirth F, Goldani LZ. 2012. Epidemiology of *Rhodotorula*: an emerging pathogen. Interdiscip Perspect Infect Dis 2012:465717. https://doi.org/10 .1155/2012/465717.
- Gümral R, Özhak-Baysan B, Tümgör A, Saraçlı MA, Yıldıran ŞT, Ilkit M, Zupančič J, Novak-Babič M, Gunde-Cimerman N, Zalar P, de Hoog GS. 2016. Dishwashers provide a selective extreme environment for human-opportunistic yeast-like fungi. Fungal Diversity 76:1–9. https://doi.org/10.1007/s13225-015-0327-8.

- Rai S, Singh DK, Kumar A. 2021. Microbial, environmental and anthropogenic factors influencing the indoor microbiome of the built environment. J Basic Microbiol 61:267–292. https://doi.org/10.1002/jobm.202000575.
- Checinska Sielaff A, Urbaniak C, Mohan GBM, Stepanov VG, Tran Q, Wood JM, Minich J, McDonald D, Mayer T, Knight R, Karouia F, Fox GE, Venkateswaran K. 2019. Characterization of the total and viable bacterial and fungal communities associated with the International Space Station surfaces. Microbiome 7:50. https://doi.org/10.1186/s40168-019-0666-x.
- Daudu R, Parker CW, Singh NK, Wood JM, Debieu M, O'Hara NB, Mason CE, Venkateswaran K. 2020. Draft genome sequences of *Rhodotorula mucilaginosa* strains isolated from the International Space Station. Microbiol Resour Announc 9:e00570-20. https://doi.org/10.1128/MRA.00570-20.
- Sugita T, Yamazaki T, Cho O, Furukawa S, Mukai C. 2021. The skin mycobiome of an astronaut during a 1-year stay on the International Space Station. Med Mycol 59:106–109. https://doi.org/10.1093/mmy/myaa067.
- Be NA, Avila-Herrera A, Allen JE, Singh N, Checinska Sielaff A, Jaing C, Venkateswaran K. 2017. Whole metagenome profiles of particulates collected from the International Space Station. Microbiome 5:81. https://doi .org/10.1186/s40168-017-0292-4.
- 30. Andrews S. 2011. FastQC: a quality control tool for high throughput sequence data. Babraham Institute, Cambridge, UK. https://www.bioinformatics.babraham.ac.uk/projects/fastqc.
- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. https://doi.org/10.1093/ bioinformatics/bty560.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10.1093/bioinformatics/btt086.
- Nilsson RH, Larsson K-H, Taylor AFS, Bengtsson-Palme J, Jeppesen TS, Schigel D, Kennedy P, Picard K, Glöckner FO, Tedersoo L, Saar I, Köljalg U, Abarenkov K. 2019. The UNITE database for molecular identification of fungi: handling dark taxa and parallel taxonomic classifications. Nucleic Acids Res 47:D259–D264. https://doi.org/10.1093/nar/gky1022.
- Samson RA, Houbraken J, Thrane U, Frisvad J, Andersen B. 2019. Food and indoor fungi, 2nd ed. Westerdijk Fungal Diversity Institute, Utrecht, The Netherlands.
- 36. Ray S, Gebre S, Fogle H, Berrios DC, Tran PB, Galazka JM, Costes SV. 2019. GeneLab: omics database for spaceflight experiments. Bioinformatics 35: 1753–1759. https://doi.org/10.1093/bioinformatics/bty884.