

# Calculation of diversity indices

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```
# setup -----
library(tidyverse)

## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr     1.1.4     v readr     2.1.6
## vforcats   1.0.1     v stringr   1.6.0
## v ggplot2   4.0.1     v tibble    3.3.0
## v lubridate 1.9.4     v tidyr    1.3.2
## v purrr    1.2.0

## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()   masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors

library(GET)
library(vegan)

## Loading required package: permute

resampled <- readRDS("transect_resampled.rds")
speciestraits <- readRDS("traits_imputation.rds")
source("0.functions.R")

indices <- vector(mode = "list")
indexcurves <- vector(mode = "list")

# transform traits -----
speciestraits |>
  select(-n) |>
  summarize(across(where(is.numeric), ~ boxcox(., check = T))) |>
  pivot_longer(everything(), names_to = "trait", values_to = "lambda") |>
  arrange(lambda, trait)

## Box-Cox transform with lambda = 0.1
## Box-Cox transform with lambda = -0.5
## Box-Cox transform with lambda = -0.4
## Box-Cox transform with lambda = 0.3
## Box-Cox transform with lambda = 0.2
```

```

## Box-Cox transform with lambda = 0
## Box-Cox transform with lambda = 0.8
## Box-Cox transform with lambda = 1.8
## Box-Cox transform with lambda = -0.2
## Box-Cox transform with lambda = 0

## # A tibble: 10 x 2
##   trait      lambda
##   <chr>     <dbl>
## 1 nmass_mg_g    -0.5
## 2 lma_g_m2     -0.4
## 3 offspring     -0.2
## 4 spread         0
## 5 ssd_combined_mg_mm3  0
## 6 leaf_area_mm2    0.100
## 7 diaspore_mass_mg    0.200
## 8 plant_height_m     0.300
## 9 BBRsize        0.8
## 10 persistence    1.8

```

This is a dry run check. The traits will be transformed as such:

- $\sim 1/\sqrt{N}$  mass, LMA, offspring
- $\sim \log$  spread, SSD, leaf area, diaspore mass
- $\sim \sqrt{\text{height}}$
- $\sim \text{identity}$  BBR size
- $\sim \text{square}$  persistence

```

data <- speciestraits |>
  select(species, where(is.numeric)) |>
  select(-n) |>
  mutate(across(where(is.numeric), ~ boxcox(.))) |>
  as.data.frame()

```

```

## Box-Cox transform with lambda = 0.1

## Box-Cox transform with lambda = -0.5

## Box-Cox transform with lambda = -0.4

## Box-Cox transform with lambda = 0.3

## Box-Cox transform with lambda = 0.2

## Box-Cox transform with lambda = 0

## Box-Cox transform with lambda = 0.8

## Box-Cox transform with lambda = 1.8

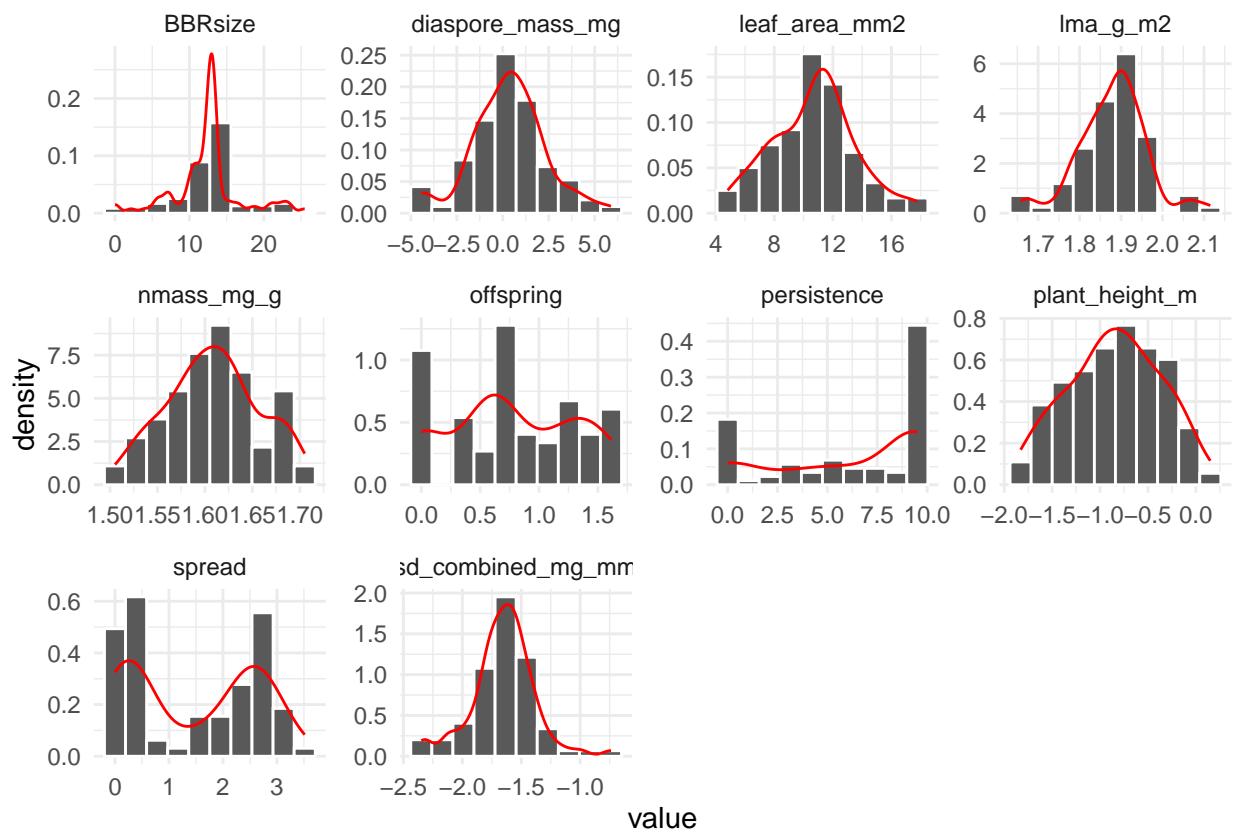
## Box-Cox transform with lambda = -0.2

## Box-Cox transform with lambda = 0

```

```
# check trait distributions

data |>
  pivot_longer(~species, names_to = "trait", values_to = "value") |>
  histdensity(value) +
  facet_wrap(~ trait, scales = "free") +
  theme_minimal()
```



```
# overall -----
## calculate distances -----

distances <- vegdist(data[-1], method = "mahalanobis", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1

distances <- distances / max(distances)

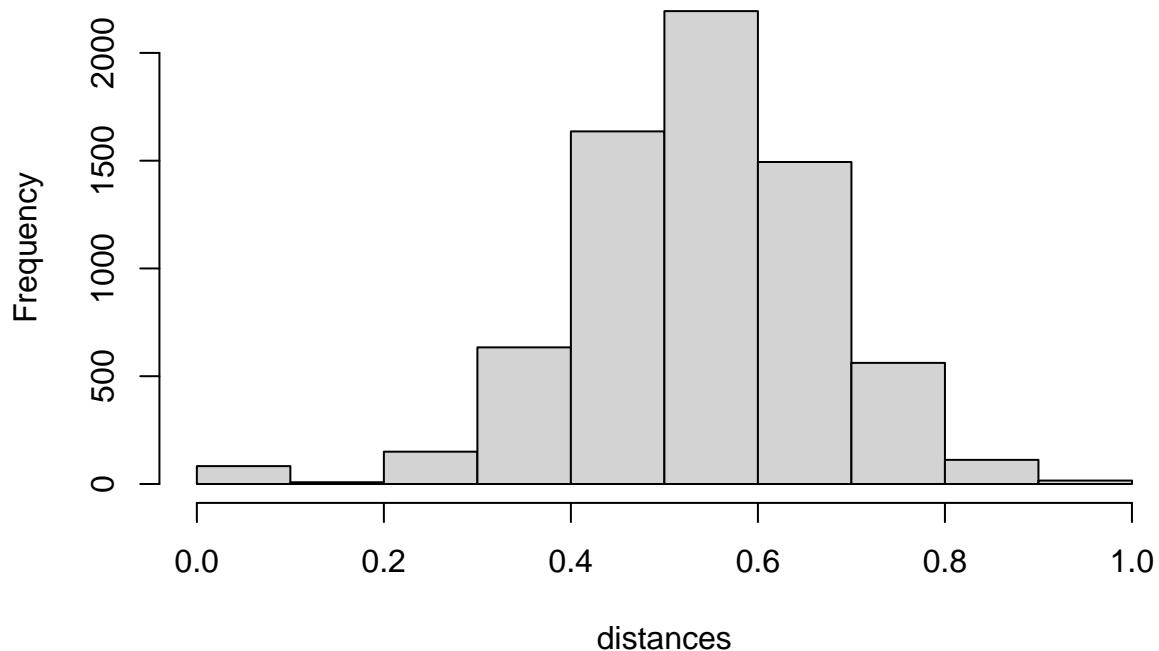
#double checking the squared euclidean property

ade4:::is.euclid(as.dist(sqrt(distances)))
```

```
## [1] TRUE
```

```
#check distance distribution
hist(distances)
```

## Histogram of distances



```
#pairs with distance 0
as.data.frame(as.table(distances)) |>
  filter(Freq == 0 & Var1 != Var2)
```

```
## [1] Var1 Var2 Freq
## <0 rows> (or 0-length row.names)

## calculate indices -----
species <- data$species

raotrait <- lapply(resampled, function (x)
  lapply(x, function (y) qdecomp(y, distances, species)) |>
    bind_rows(.id = "step") |>
    mutate(step = as.numeric(step))
) |>
  bind_rows(.id = "site")

#scale steps in units of area
```

```

raotrait <- raotrait |>
  mutate(step = step / 100)

#add to list
indices[[1]] <- raotrait
names(indices)[1] <- "overall"

## create curve set -----
curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

indexcurves[[1]] <- curvesets
names(indexcurves)[1] <- "overall"

# aboveground -----
## calculate distances -----
distances <- vegdist(data[2:7], method = "mahalanobis", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1
distances <- distances / max(distances)

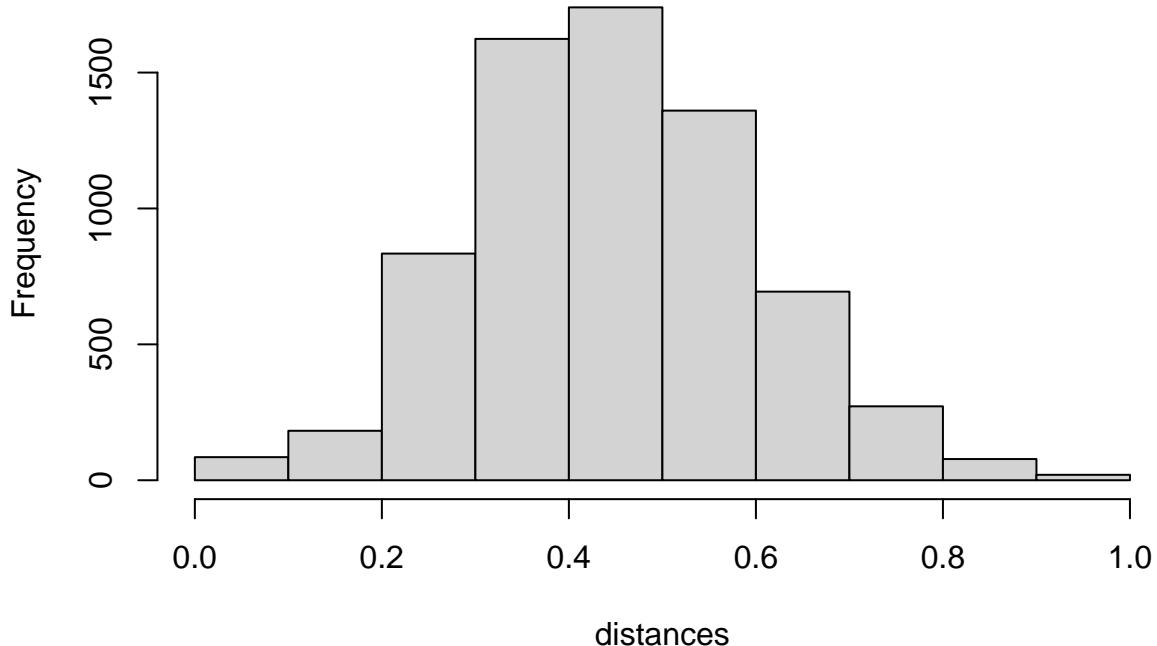
#double checking the squared euclidean property
ade4:::is.euclid(as.dist(sqrt(distances)))

## [1] TRUE

#check distance distribution
hist(distances)

```

## Histogram of distances



```
#pairs with distance 0

as.data.frame(as.table(distances)) |>
  filter(Freq == 0 & Var1 != Var2)

## [1] Var1 Var2 Freq
## <0 rows> (or 0-length row.names)

## calculate indices -----
raotrait <- lapply(resampled, function (x)
  lapply(x, function (y) qdecomp(y, distances, species)) |>
    bind_rows(.id = "step") |>
    mutate(step = as.numeric(step))
) |>
  bind_rows(.id = "site")

#scale steps in units of area

raotrait <- raotrait |>
  mutate(step = step / 100)

#add to list

indices[[2]] <- raotrait
```

```

names(indices)[2] <- "aboveground"

## create curve set -----
curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

indexcurves[[2]] <- curvesets
names(indexcurves)[2] <- "aboveground"

# clonal -----
## calculate distances -----
distances <- vegdist(data[8:11], method = "mahalanobis", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1
distances <- distances / max(distances)

#double checking the squared euclidean property
ade4::is.euclid(as.dist(sqrt(distances)))

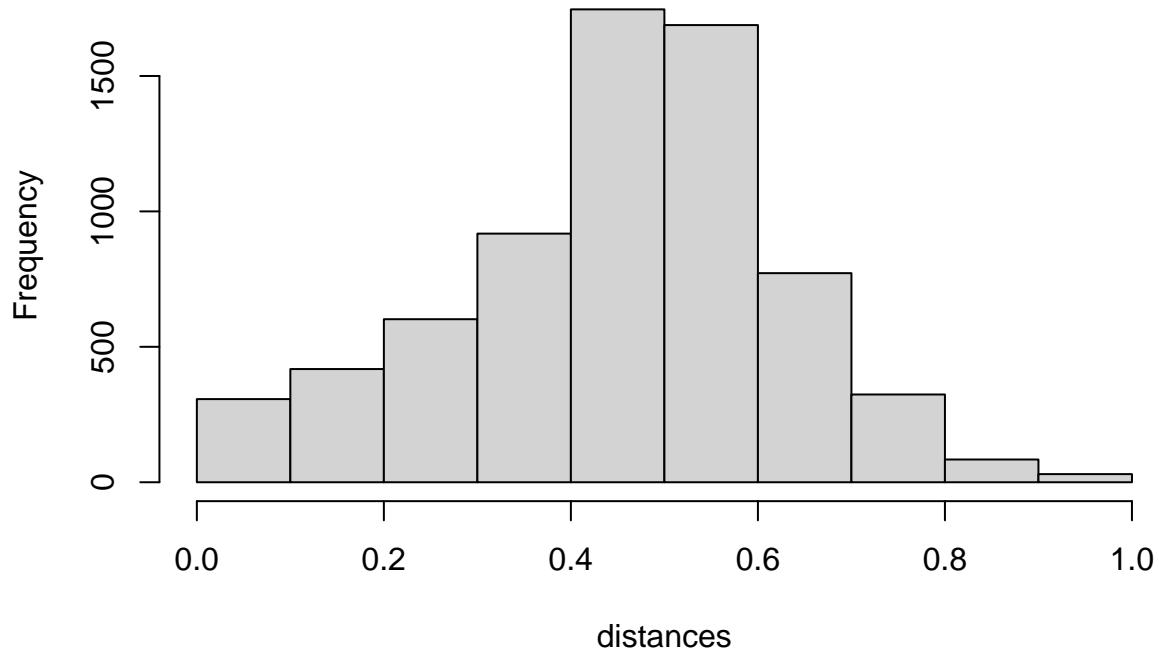
## Warning in ade4::is.euclid(as.dist(sqrt(distances))): Zero distance(s)

## [1] TRUE

#check distance distribution
hist(distances)

```

## Histogram of distances



```
#pairs with distance 0  
  
as.data.frame(as.table(distances)) |>  
  filter(Freq == 0 & Var1 != Var2)
```

```
##      Var1 Var2 Freq  
## 1      10    1    0  
## 2      29    1    0  
## 3      44    1    0  
## 4      49    1    0  
## 5      67    1    0  
## 6      1   10    0  
## 7      29   10    0  
## 8      44   10    0  
## 9      49   10    0  
## 10     67   10    0  
## 11     19   15    0  
## 12     37   15    0  
## 13     34   17    0  
## 14     75   17    0  
## 15     15   19    0  
## 16     37   19    0  
## 17     48   28    0  
## 18     66   28    0  
## 19      1   29    0
```

```
## 20 10 29 0
## 21 44 29 0
## 22 49 29 0
## 23 67 29 0
## 24 35 31 0
## 25 36 31 0
## 26 40 31 0
## 27 77 31 0
## 28 17 34 0
## 29 75 34 0
## 30 31 35 0
## 31 36 35 0
## 32 40 35 0
## 33 77 35 0
## 34 31 36 0
## 35 35 36 0
## 36 40 36 0
## 37 77 36 0
## 38 15 37 0
## 39 19 37 0
## 40 61 39 0
## 41 31 40 0
## 42 35 40 0
## 43 36 40 0
## 44 77 40 0
## 45 1 44 0
## 46 10 44 0
## 47 29 44 0
## 48 49 44 0
## 49 67 44 0
## 50 28 48 0
## 51 66 48 0
## 52 1 49 0
## 53 10 49 0
## 54 29 49 0
## 55 44 49 0
## 56 67 49 0
## 57 39 61 0
## 58 28 66 0
## 59 48 66 0
## 60 1 67 0
## 61 10 67 0
## 62 29 67 0
## 63 44 67 0
## 64 49 67 0
## 65 17 75 0
## 66 34 75 0
## 67 31 77 0
## 68 35 77 0
## 69 36 77 0
## 70 40 77 0
## 71 82 80 0
## 72 80 82 0
```

```

## calculate indices ----

raotrait <- lapply(resampled, function (x)
  lapply(x, function (y) qdecomp(y, distances, species)) |>
    bind_rows(.id = "step") |>
    mutate(step = as.numeric(step))
) |>
  bind_rows(.id = "site")

#scale steps in units of area

raotrait <- raotrait |>
  mutate(step = step / 100)

#add to list

indices[[3]] <- raotrait
names(indices)[3] <- "clonal"

## create curve set ----

curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

indexcurves[[3]] <- curvesets
names(indexcurves)[3] <- "clonal"

# plotting indices ----
categories <- read_csv("data/classification.csv")

## Rows: 32 Columns: 2

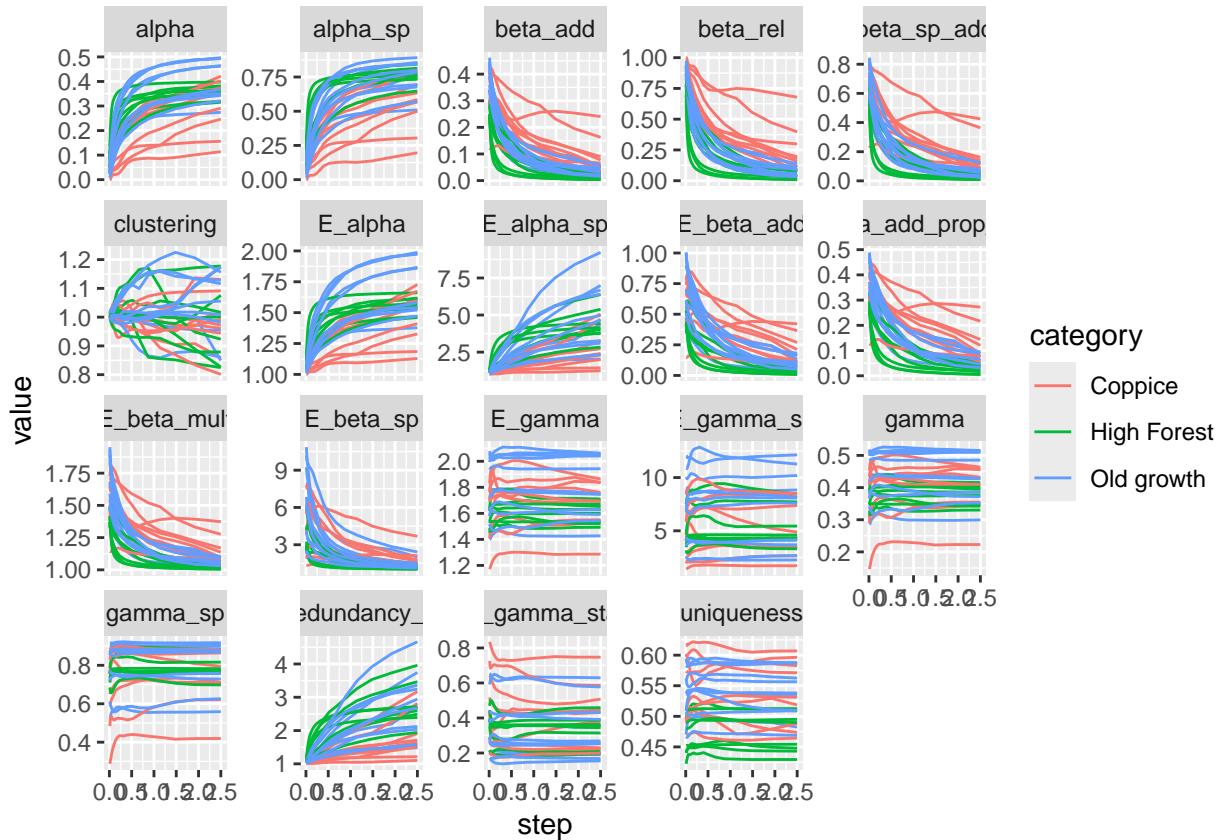
## -- Column specification -----
## Delimiter: ","
## chr (2): site, category
##
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.

indices$overall |>
  left_join(categories) |>
  pivot_longer(3:21, names_to = "index", values_to = "value") |>

```

```
ggplot(aes(x = step, y = value, group = site, colour = category))+  
  geom_line() +  
  facet_wrap(~index, scales = "free_y")
```

## Joining with `by = join\_by(site)`



```
# save results -----  
  
saveRDS(indices, "indices.rds")  
saveRDS(indexcurves, "curves.rds")  
  
# session info -----  
  
sessionInfo()  
  
## R version 4.5.2 (2025-10-31)  
## Platform: x86_64-redhat-linux-gnu  
## Running under: Nohara Linux 43 (KDE Plasma Desktop Edition)  
##  
## Matrix products: default  
## BLAS/LAPACK: FlexiBLAS OPENBLAS-OPENMP; LAPACK version 3.12.1  
##  
## locale:  
## [1] LC_CTYPE=it_IT.UTF-8          LC_NUMERIC=C
```

```

## [3] LC_TIME=it_IT.utf8      LC_COLLATE=it_IT.UTF-8
## [5] LC_MONETARY=it_IT.utf8   LC_MESSAGES=it_IT.UTF-8
## [7] LC_PAPER=it_IT.utf8     LC_NAME=C
## [9] LC_ADDRESS=C            LC_TELEPHONE=C
## [11] LC_MEASUREMENT=it_IT.utf8 LC_IDENTIFICATION=C
##
## time zone: Europe/Rome
## tzcode source: system (glibc)
##
## attached base packages:
## [1] stats      graphics grDevices datasets  utils      methods   base
##
## other attached packages:
## [1] vegan_2.7-2    permute_0.9-8   GET_1.0-7      lubridate_1.9.4
## [5] forcats_1.0.1  stringr_1.6.0   dplyr_1.1.4    purrr_1.2.0
## [9] readr_2.1.6    tidyr_1.3.2    tibble_3.3.0   ggplot2_4.0.1
## [13] tidyverse_2.0.0
##
## loaded via a namespace (and not attached):
## [1] utf8_1.2.6        generics_0.1.4   stringi_1.8.7   lattice_0.22-7
## [5] hms_1.1.4         digest_0.6.39    magrittr_2.0.4   evaluate_1.0.5
## [9] grid_4.5.2        timechange_0.3.0 RColorBrewer_1.1-3 fastmap_1.2.0
## [13] Matrix_1.7-4     gridExtra_2.3    mgcv_1.9-4     viridisLite_0.4.2
## [17] scales_1.4.0     CoprManager_0.5.7 ade4_1.7-23    cli_3.6.5
## [21] crayon_1.5.3     rlang_1.1.6     bit64_4.6.0-1  splines_4.5.2
## [25] withr_3.0.2      yaml_2.3.12    tools_4.5.2    parallel_4.5.2
## [29] tzdb_0.5.0       vctrs_0.6.5    R6_2.6.1      lifecycle_1.0.4
## [33] bit_4.6.0        vroom_1.6.7   MASS_7.3-65   cluster_2.1.8.1
## [37] pkgconfig_2.0.3   pillar_1.11.1  gtable_0.3.6   Rcpp_1.1.0
## [41] glue_1.8.0        xfun_0.55     tidyselect_1.2.1 rstudioapi_0.17.1
## [45] knitr_1.50       farver_2.1.2   nlme_3.1-168  htmltools_0.5.9
## [49] labeling_0.4.3   rmarkdown_2.30 compiler_4.5.2 S7_0.2.1

```