

Calculation of diversity indices

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2026-01-07

```
# setup -----

library(tidyverse)

## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr      1.1.4      v readr      2.1.6
## v forcats    1.0.1      v stringr    1.6.0
## v ggplot2    4.0.1      v tibble     3.3.0
## v lubridate  1.9.4      v tidyr      1.3.2
## v purrr      1.2.0
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()     masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors

library(GET)
library(vegan)

## Loading required package: permute

resampled <- readRDS("transect_resampled.rds")
speciestraits <- readRDS("traits_imputation.rds")
source("0.functions.R")

indices <- vector(mode = "list")
indexcurves <- vector(mode = "list")

# transform traits -----

speciestraits |>
  select(-n) |>
  summarize(across(where(is.numeric), ~ boxcox(., check = T))) |>
  pivot_longer(everything(), names_to = "trait", values_to = "lambda") |>
  arrange(lambda, trait)

## Box-Cox transform with lambda = 0.1
## Box-Cox transform with lambda = -0.5
## Box-Cox transform with lambda = -0.4
## Box-Cox transform with lambda = 0.3
## Box-Cox transform with lambda = 0.2
```

```
## Box-Cox transform with lambda = 0
## Box-Cox transform with lambda = 0.8
## Box-Cox transform with lambda = 1.8
## Box-Cox transform with lambda = -0.2
## Box-Cox transform with lambda = 0

## # A tibble: 10 x 2
##   trait          lambda
##   <chr>         <dbl>
## 1 nmass_mg_g    -0.5
## 2 lma_g_m2     -0.4
## 3 offspring    -0.2
## 4 spread        0
## 5 ssd_combined_mg_mm3 0
## 6 leaf_area_mm2 0.100
## 7 diaspore_mass_mg 0.200
## 8 plant_height_m 0.300
## 9 BBRsize       0.8
## 10 persistence  1.8
```

This is a dry run check. The traits will be transformed as such:

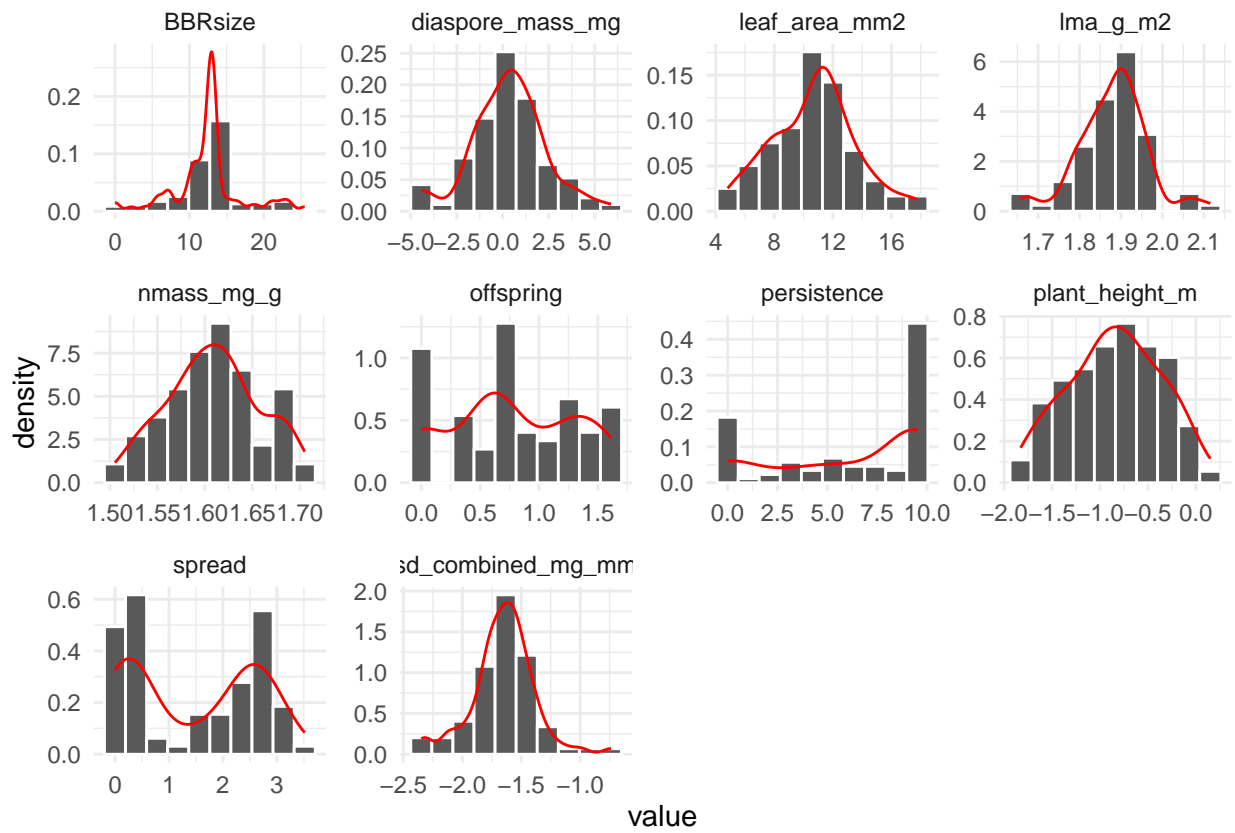
- $\sim 1/\sqrt{N}$ mass, LMA, offspring
- $\sim \log$ spread, SSD, leaf area, diaspore mass
- $\sim \sqrt{\text{height}}$
- $\sim \text{identity}$ BBR size
- $\sim \text{square}$ persistence

```
data <- speciestraits |>
  select(species, where(is.numeric)) |>
  select(-n) |>
  mutate(across(where(is.numeric), ~ boxcox(.))) |>
  as.data.frame()
```

```
## Box-Cox transform with lambda = 0.1
## Box-Cox transform with lambda = -0.5
## Box-Cox transform with lambda = -0.4
## Box-Cox transform with lambda = 0.3
## Box-Cox transform with lambda = 0.2
## Box-Cox transform with lambda = 0
## Box-Cox transform with lambda = 0.8
## Box-Cox transform with lambda = 1.8
## Box-Cox transform with lambda = -0.2
## Box-Cox transform with lambda = 0
```

```
# check trait distributions
```

```
data |>
  pivot_longer(-species, names_to = "trait", values_to = "value") |>
  histdensity(value)+
  facet_wrap(~ trait, scales = "free")+
  theme_minimal()
```



```
# overall -----

## calculate distances -----

distances <- vegdist(data[-1], method = "mahalanobis", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1

distances <- distances / max(distances)

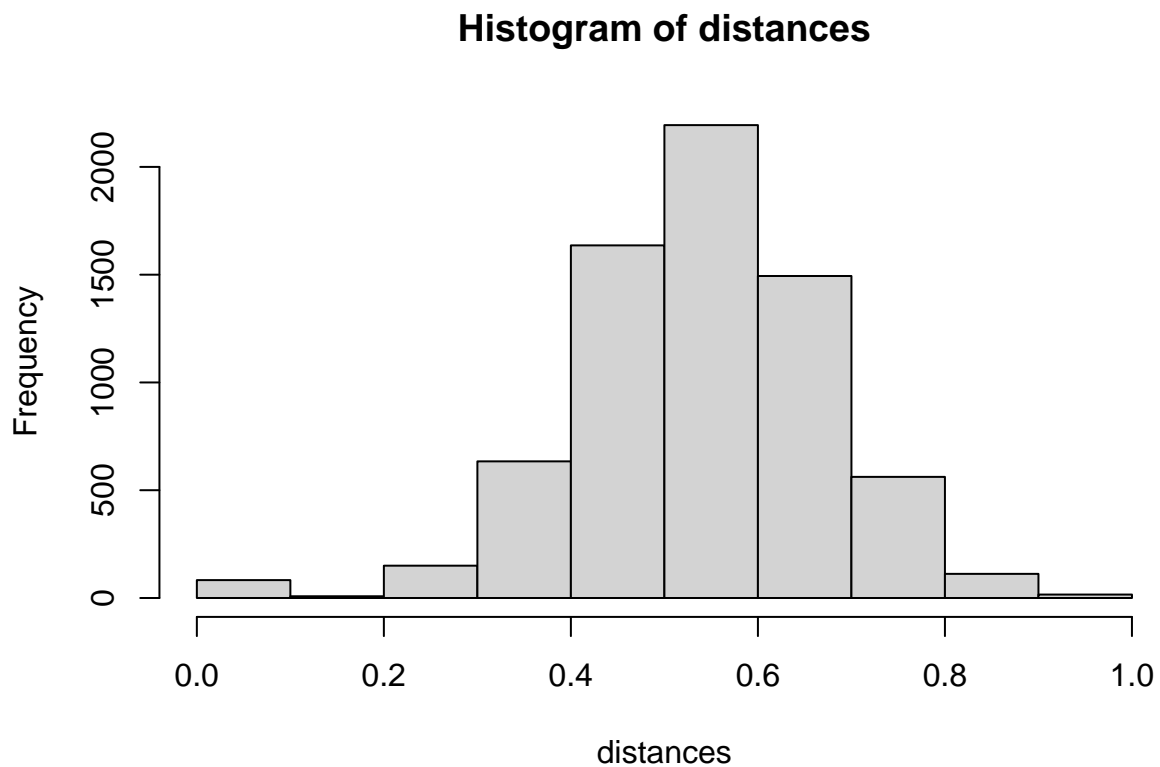
#double checking the squared euclidean property

ade4::is.euclid(as.dist(sqrt(distances)))

## [1] TRUE
```

```
#check distance distribution
```

```
hist(distances)
```



```
#pairs with distance 0
```

```
as.data.frame(as.table(distances)) |>  
  filter(Freq == 0 & Var1 != Var2)
```

```
## [1] Var1 Var2 Freq  
## <0 rows> (or 0-length row.names)
```

```
## calculate indices -----
```

```
species <- data$species
```

```
raotrait <- lapply(resampled, function (x)  
  lapply(x, function (y) qdecomp(y, distances, species)) |>  
    bind_rows(.id = "step") |>  
    mutate(step = as.numeric(step))  
) |>  
  bind_rows(.id = "site")
```

```
#scale steps in units of area
```

```

raotrait <- raotrait |>
  mutate(step = step / 100)

#add to list
indices[[1]] <- raotrait
names(indices)[1] <- "overall"

## create curve set -----

curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

indexcurves[[1]] <- curvesets
names(indexcurves)[1] <- "overall"

# aboveground -----

## calculate distances -----

distances <- vegdist(data[2:7], method = "mahalanobis", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1

distances <- distances / max(distances)

#double checking the squared euclidean property

ade4::is.euclid(as.dist(sqrt(distances)))

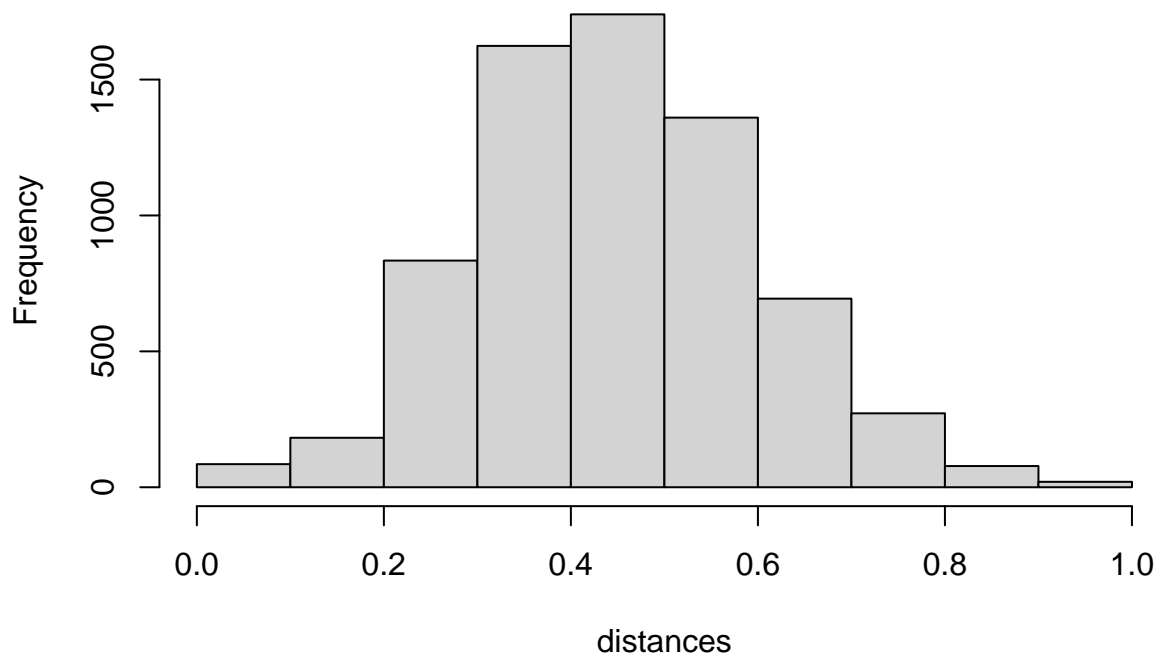
## [1] TRUE

#check distance distribution

hist(distances)

```

Histogram of distances



```
#pairs with distance 0
```

```
as.data.frame(as.table(distances)) |>
  filter(Freq == 0 & Var1 != Var2)
```

```
## [1] Var1 Var2 Freq
## <0 rows> (or 0-length row.names)
```

```
## calculate indices -----
```

```
raotrait <- lapply(resampled, function (x)
  lapply(x, function (y) qdecomp(y, distances, species)) |>
  bind_rows(.id = "step") |>
  mutate(step = as.numeric(step))
) |>
  bind_rows(.id = "site")
```

```
#scale steps in units of area
```

```
raotrait <- raotrait |>
  mutate(step = step / 100)
```

```
#add to list
```

```
indices[[2]] <- raotrait
```

```

names(indices)[2] <- "aboveground"

## create curve set -----

curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

indexcurves[[2]] <- curvesets
names(indexcurves)[2] <- "aboveground"

# clonal -----

## calculate distances -----

distances <- vegdist(data[8:11], method = "mahalanobis", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1

distances <- distances / max(distances)

#double checking the squared euclidean property

ade4::is.euclid(as.dist(sqrt(distances)))

## Warning in ade4::is.euclid(as.dist(sqrt(distances))): Zero distance(s)

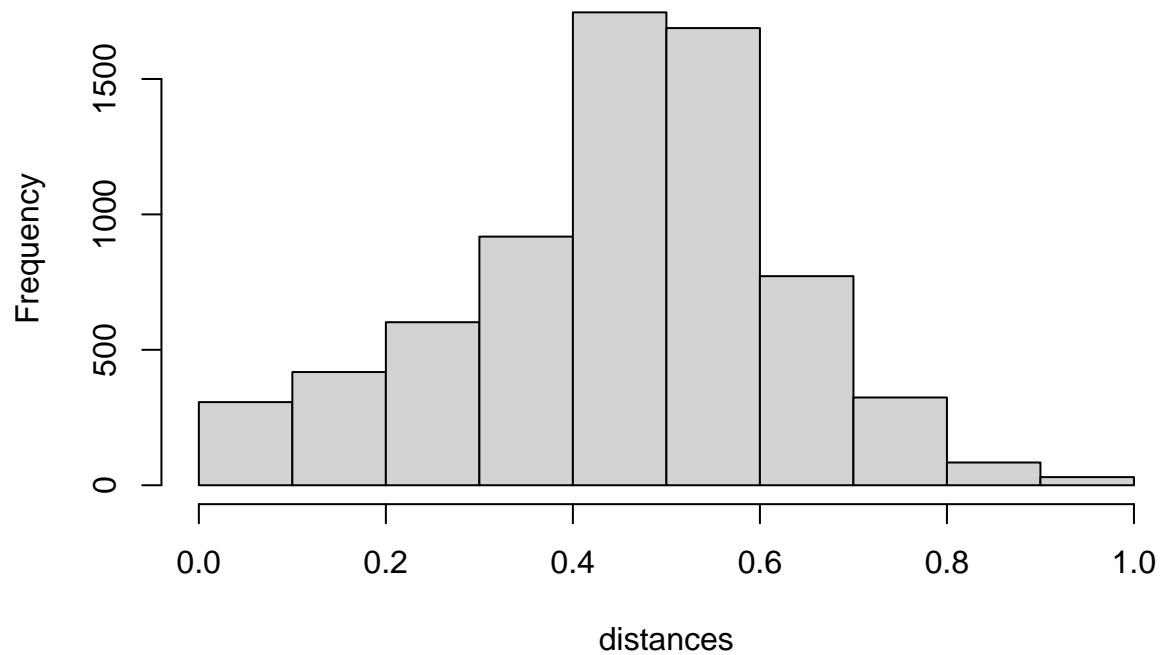
## [1] TRUE

#check distance distribution

hist(distances)

```

Histogram of distances



#pairs with distance 0

```
as.data.frame(as.table(distances)) |>
  filter(Freq == 0 & Var1 != Var2)
```

##	Var1	Var2	Freq
## 1	10	1	0
## 2	29	1	0
## 3	44	1	0
## 4	49	1	0
## 5	67	1	0
## 6	1	10	0
## 7	29	10	0
## 8	44	10	0
## 9	49	10	0
## 10	67	10	0
## 11	19	15	0
## 12	37	15	0
## 13	34	17	0
## 14	75	17	0
## 15	15	19	0
## 16	37	19	0
## 17	48	28	0
## 18	66	28	0
## 19	1	29	0

##	20	10	29	0
##	21	44	29	0
##	22	49	29	0
##	23	67	29	0
##	24	35	31	0
##	25	36	31	0
##	26	40	31	0
##	27	77	31	0
##	28	17	34	0
##	29	75	34	0
##	30	31	35	0
##	31	36	35	0
##	32	40	35	0
##	33	77	35	0
##	34	31	36	0
##	35	35	36	0
##	36	40	36	0
##	37	77	36	0
##	38	15	37	0
##	39	19	37	0
##	40	61	39	0
##	41	31	40	0
##	42	35	40	0
##	43	36	40	0
##	44	77	40	0
##	45	1	44	0
##	46	10	44	0
##	47	29	44	0
##	48	49	44	0
##	49	67	44	0
##	50	28	48	0
##	51	66	48	0
##	52	1	49	0
##	53	10	49	0
##	54	29	49	0
##	55	44	49	0
##	56	67	49	0
##	57	39	61	0
##	58	28	66	0
##	59	48	66	0
##	60	1	67	0
##	61	10	67	0
##	62	29	67	0
##	63	44	67	0
##	64	49	67	0
##	65	17	75	0
##	66	34	75	0
##	67	31	77	0
##	68	35	77	0
##	69	36	77	0
##	70	40	77	0
##	71	82	80	0
##	72	80	82	0

```

## calculate indices -----

raotrait <- lapply(resampled, function (x)
  lapply(x, function (y) qdecomp(y, distances, species)) |>
  bind_rows(.id = "step") |>
  mutate(step = as.numeric(step))
) |>
  bind_rows(.id = "site")

#scale steps in units of area

raotrait <- raotrait |>
  mutate(step = step / 100)

#add to list

indices[[3]] <- raotrait
names(indices)[3] <- "clonal"

## create curve set -----

curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

indexcurves[[3]] <- curvesets
names(indexcurves)[3] <- "clonal"

# plotting indices -----

categories <- read_csv("data/classification.csv")

## Rows: 32 Columns: 2

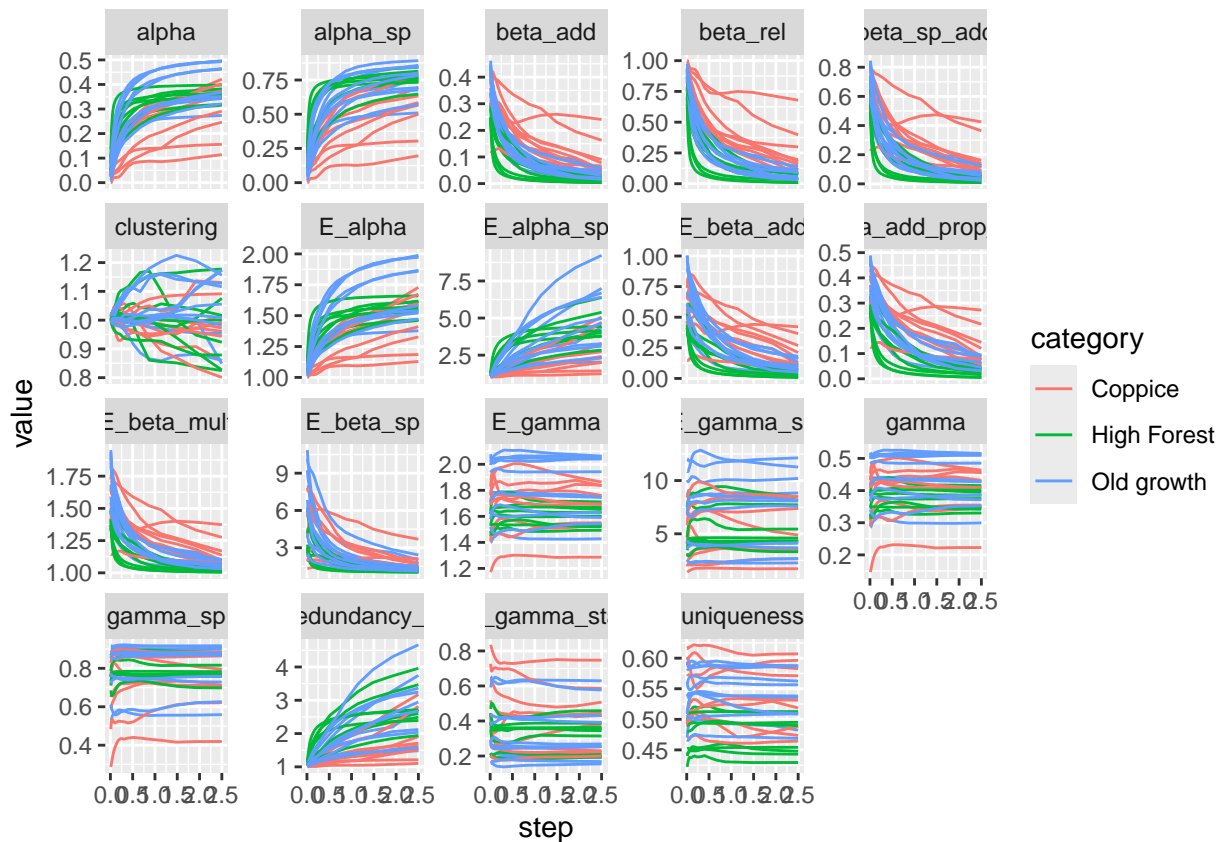
## -- Column specification -----
## Delimiter: ","
## chr (2): site, category
##
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.

indices$overall |>
  left_join(categories) |>
  pivot_longer(3:21, names_to = "index", values_to = "value") |>

```

```
ggplot(aes(x = step, y = value, group = site, colour = category))+
  geom_line()+
  facet_wrap(~index, scales = "free_y")
```

```
## Joining with `by = join_by(site)`
```



```
# save results -----
saveRDS(indices, "indices.rds")
saveRDS(indexcurves, "curves.rds")

# session info -----
sessionInfo()
```

```
## R version 4.5.2 (2025-10-31)
## Platform: x86_64-redhat-linux-gnu
## Running under: Nobara Linux 43 (KDE Plasma Desktop Edition)
##
## Matrix products: default
## BLAS/LAPACK: FlexiBLAS OPENBLAS-OPENMP; LAPACK version 3.12.1
##
## locale:
## [1] LC_CTYPE=it_IT.UTF-8      LC_NUMERIC=C
```

```

## [3] LC_TIME=it_IT.utf8      LC_COLLATE=it_IT.UTF-8
## [5] LC_MONETARY=it_IT.utf8   LC_MESSAGES=it_IT.UTF-8
## [7] LC_PAPER=it_IT.utf8     LC_NAME=C
## [9] LC_ADDRESS=C            LC_TELEPHONE=C
## [11] LC_MEASUREMENT=it_IT.utf8 LC_IDENTIFICATION=C
##
## time zone: Europe/Rome
## tzcode source: system (glibc)
##
## attached base packages:
## [1] stats      graphics  grDevices  datasets  utils      methods    base
##
## other attached packages:
## [1] vegan_2.7-2      permute_0.9-8    GET_1.0-7        lubridate_1.9.4
## [5] forcats_1.0.1    stringr_1.6.0    dplyr_1.1.4      purrr_1.2.0
## [9] readr_2.1.6      tidyr_1.3.2      tibble_3.3.0     ggplot2_4.0.1
## [13] tidyverse_2.0.0
##
## loaded via a namespace (and not attached):
## [1] utf8_1.2.6      generics_0.1.4   stringi_1.8.7    lattice_0.22-7
## [5] hms_1.1.4       digest_0.6.39    magrittr_2.0.4    evaluate_1.0.5
## [9] grid_4.5.2      timechange_0.3.0 RColorBrewer_1.1-3 fastmap_1.2.0
## [13] Matrix_1.7-4    gridExtra_2.3    mgcv_1.9-4        viridisLite_0.4.2
## [17] scales_1.4.0    CoprManager_0.5.7 ade4_1.7-23        cli_3.6.5
## [21] crayon_1.5.3    rlang_1.1.6      bit64_4.6.0-1     splines_4.5.2
## [25] withr_3.0.2     yaml_2.3.12      tools_4.5.2       parallel_4.5.2
## [29] tzdb_0.5.0      vctrs_0.6.5      R6_2.6.1          lifecycle_1.0.4
## [33] bit_4.6.0       vroom_1.6.7      MASS_7.3-65       cluster_2.1.8.1
## [37] pkgconfig_2.0.3 pillar_1.11.1     gtable_0.3.6      Rcpp_1.1.0
## [41] glue_1.8.0      xfun_0.55        tidyselect_1.2.1  rstudioapi_0.17.1
## [45] knitr_1.50      farver_2.1.2     nlme_3.1-168      htmltools_0.5.9
## [49] labeling_0.4.3  rmarkdown_2.30   compiler_4.5.2    S7_0.2.1

```