

Structural and canopy variables

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```
# setup -----

library(tidyverse)

## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr      1.1.4      v readr      2.1.6
## v forcats    1.0.1      v stringr   1.6.0
## v ggplot2    4.0.1      v tibble    3.3.0
## v lubridate  1.9.4      v tidyr     1.3.2
## v purrr      1.2.0
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()     masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors

library(GGally)
library(klaR)

## Loading required package: MASS
##
## Attaching package: 'MASS'
##
## The following object is masked from 'package:dplyr':
##
##      select

library(fclust)
library(party)

## Loading required package: grid
## Loading required package: mvtnorm
## Loading required package: modeltools
## Loading required package: stats4
## Loading required package: strucchange
## Loading required package: zoo
##
## Attaching package: 'zoo'
##
## The following objects are masked from 'package:base':
##
```

```

##      as.Date, as.Date.numeric
##
## Loading required package: sandwich
##
## Attaching package: 'strucchange'
##
## The following object is masked from 'package:stringr':
##
##      boundary
##
## Attaching package: 'party'
##
## The following object is masked from 'package:dplyr':
##
##      where

set.seed(92538)

# function for the upper triangle of pairs plot. Adapted from Solomon Kurz.

my_upper <- function(data, mapping, ...) {

  # get the x and y data to use the other code
  x <- eval_data_col(data, mapping$x)
  y <- eval_data_col(data, mapping$y)

  r <- unname(cor.test(x, y)$estimate)
  rt <- format(r, digits = 2)[1]
  tt <- as.character(rt)

  # plot the cor value
  ggally_text(
    label = tt,
    mapping = aes(),
    size = 4)+
    theme_void()
}

# import data -----

# DCP canopy indices
canopy <- read_csv("data/DCP.csv") |>
  group_by(site) |>
  summarise(LAImean = mean(LAI), LAIsd = sd(LAI)) |>
  rename(LAI = LAImean)

## Rows: 384 Columns: 10
## -- Column specification -----
## Delimiter: ","
## chr (2): site, Date
## dbl (8): Gap Fraction, Foliage Cover, Crown Cover, Crown Porosity, LAI, effe...
##
## i Use `spec()` to retrieve the full column specification for this data.

```

```
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
```

```
# TLS structural indices
structure <- read_csv("data/TLS.csv") |>
  mutate(links_norm = n_links_4_6/TopH,
         min_pc_norm = pc_min_sect/TopH,
         max_pc_norm = pc_max_sect/TopH,
         sd_perc_norm = sd_perc_clust/TopH) |>
  dplyr::select(!c("Sampling_date", "n_links_4_6", "pc_min_sect", "pc_max_sect", "sd_perc_clust"))
```

```
## Rows: 28 Columns: 11
## -- Column specification -----
## Delimiter: ","
## chr (2): site, Sampling_date
## dbl (9): pc_min_sect, pc_max_sect, sd_perc_clust, n_links_4_6, TopH, VCI, RH...
##
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
```

```
# join together
variables <- structure |>
  inner_join(canopy)
```

```
## Joining with `by = join_by(site)`
```

```
rm(structure, canopy)

# join categories
cat <- read_csv("data/classification.csv")
```

```
## Rows: 32 Columns: 2
## -- Column specification -----
## Delimiter: ","
## chr (2): site, category
##
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
```

```
variables <- cat |>
  inner_join(variables) |>
  mutate(category = as.factor(category))
```

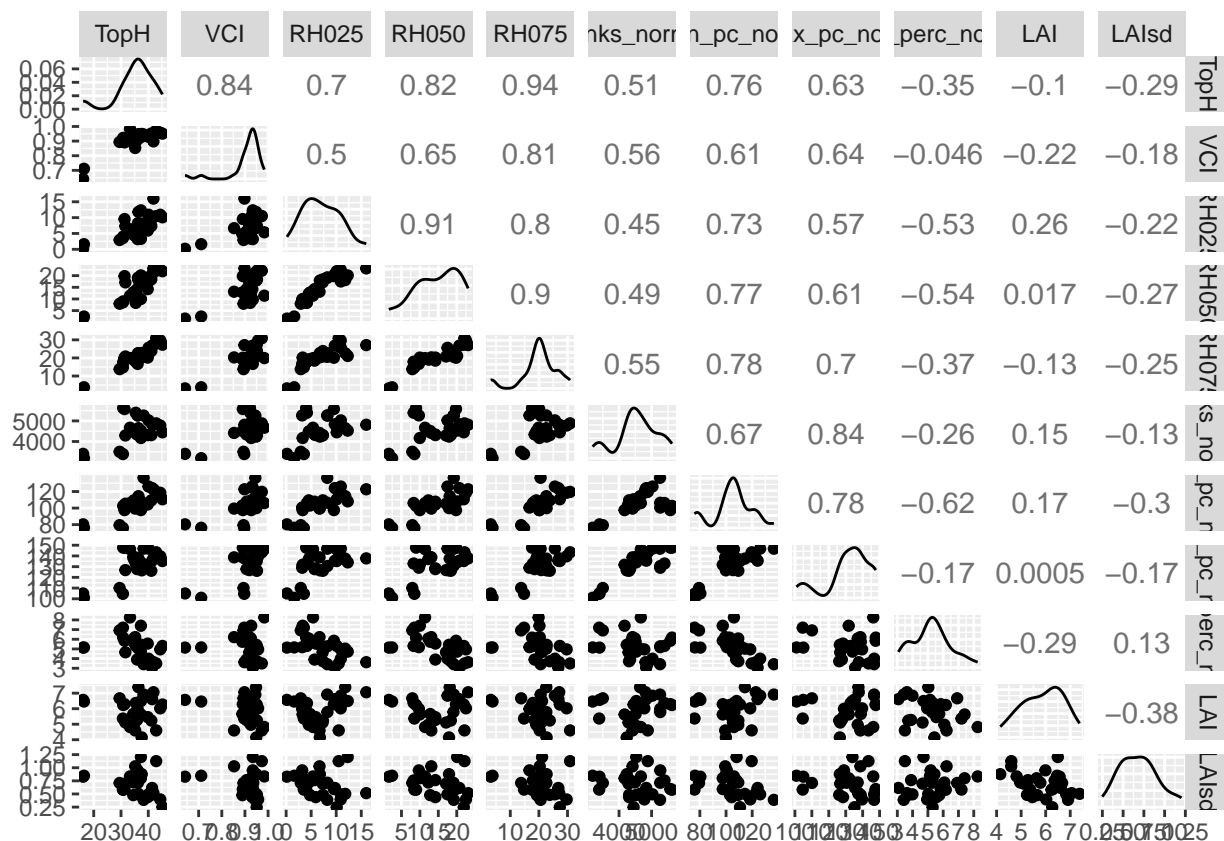
```
## Joining with `by = join_by(site)`
```

```
rm(cat)

variables |>
  dplyr::select(2:7, 12, 13) |>
  print(n = 30)
```

```
## # A tibble: 28 x 8
##   category    TopH    VCI  RH025 RH050 RH075    LAI LAIsd
##   <fct>      <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl>
## 1 Coppice    16.1 0.641 0.159  1.57  3.16  6.57 0.829
## 2 Coppice    16.4 0.711 1.61   2.46  3.79  6.46 0.848
## 3 High Forest 37.6 0.949 6.74  17.9 22.7  4.15 0.877
## 4 High Forest 31.6 0.919 7.29  17.1 20.8  5.11 0.696
## 5 Old growth 39.8 0.932 5.91  14.2 20.2  5.46 0.440
## 6 Old growth 41.1 0.952 7.34  18.4 23.8  5.24 0.483
## 7 Old growth 35.9 0.907 5.67  14.2 18.5  5.27 0.764
## 8 Old growth 34.7 0.926 4.53  12.3 19.1  4.96 0.719
## 9 Old growth 37.3 0.927 3.3    9.98 20.2  5.63 0.800
## 10 Coppice   29.5 0.894 2.93   7.95 13.8  6.67 0.710
## 11 Coppice   30.7 0.897 3.54   8.51 14.8  5.37 0.583
## 12 Old growth 33.3 0.984 5.37  11.4 19.9  4.82 0.834
## 13 Old growth 35.2 0.853 6.67  13.1 20.3  4.60 1.03
## 14 Old growth 34.5 0.889 4.78  12.8 19.3  5.66 0.478
## 15 High Forest 40.1 0.931 10.9   17.5 25.6  6.84 0.493
## 16 High Forest 38.3 0.945 8.21  19.2 20.6  6.91 0.388
## 17 High Forest 41.9 0.898 16     23.1 27.1  7.06 0.516
## 18 High Forest 36.2 0.936 8.24  19.1 22.6  5.78 0.714
## 19 Coppice   36.1 0.934 3.21   9.72 16.7  5.99 0.950
## 20 Coppice   35.4 0.923 4.15   8.31 17.9  6.37 0.758
## 21 Coppice   30.8 0.922 4.01   8.96 17.6  6.24 0.581
## 22 High Forest 31.3 0.891 9.54  19.8 20.1  6.29 0.593
## 23 Coppice   36.4 0.948 11.8   18.1 22.6  7.38 0.503
## 24 Coppice   37.3 0.934 12.3   20.1 21.1  6.49 1.20
## 25 High Forest 41.4 0.934 9.73  20.7 26.8  6.61 0.615
## 26 Old growth 45.2 0.951 10.1   21.9 27.3  6.27 0.248
## 27 Old growth 44.4 0.957 10.9   22.5 29.7  6.02 0.396
## 28 Old growth 42.9 0.973 10.5   22.1 30.9  4.60 1.13
```

```
#pairs plot
variables[3:13] |>
  ggpairs(upper = list(continuous = my_upper))
```



```
# standardization
# FLM test is non-parametric, same results either way
# but standardization gives more meaningful coefficients and is required for other steps here

variables <- variables |>
  mutate(across(where(is.numeric), scale))|>
  mutate(across(where(is.numeric), as.vector))

# random forest -----

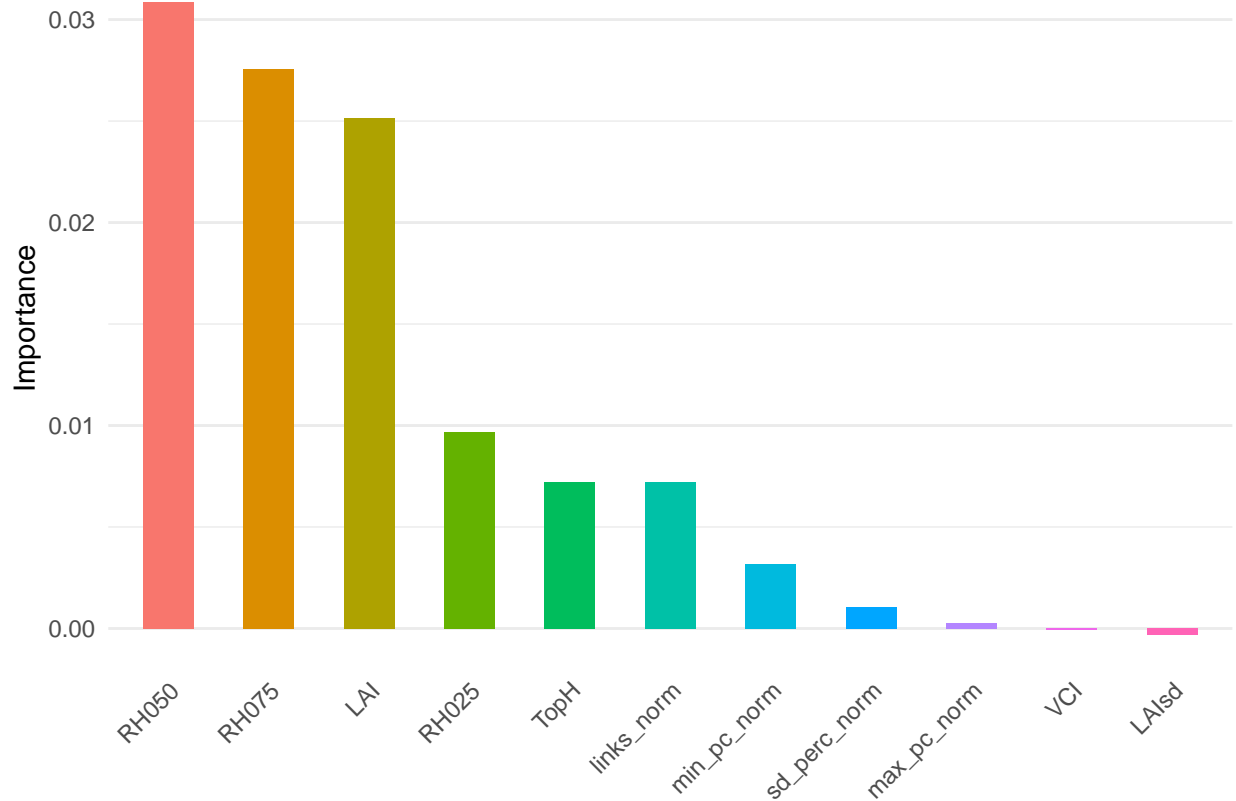
forest <- cforest(category ~ ., data = variables[, -1],
  control = cforest_control(mtry = 3, ntree = 5000, mincriterion = 0)) #mtry default fr

#variable importance from RF model
importance <- varimp(forest, conditional=T)

importance <- tibble(variable = names(importance), importance = importance) |>
  mutate(variable = fct_reorder(variable, importance, .desc = T)) |>
  arrange(variable)

#plot IV
importance |>
  ggplot(aes(x = variable, y = importance, fill = variable))+
  geom_col(width = 0.5)+
  theme_minimal()+
  theme(legend.position = "none",
```

```
axis.text.x = element_text(angle = 45, hjust = 1),
panel.grid.major.x = element_blank()+
labs(x = NULL, y = "Importance")
```



```
ggsave("plots/importance.png", width = 190, height = 100, units = "mm", bg = 'white',
scale = 0.8, dpi = 1000)
ggsave('plots/FIG1.eps', width = 190, height = 100, units = "mm", bg = 'white',
scale = 0.8, dpi = 1000)
```

```
sel_var <- variables |>
  dplyr::select(site, category, RH050, LAI)
```

```
saveRDS(sel_var, "selected_variables.rds")
```

```
# clustering -----
```

```
# fuzzy k-means
```

```
fuzzy <- FKM(sel_var[-c(1, 2)], k = 3)
```

```
#add to df
```

```
sel_var <- sel_var |>
  mutate(cluster_fuzzy = fuzzy$clus[,1],
membership = fuzzy$clus[,2])
```

```
#assign the most plausible labels
```

```

contingency_fuzzy <- as.matrix(table(sel_var$category, sel_var$cluster_fuzzy))
assignment_fuzzy <- clue::solve_LSAP(t(contingency_fuzzy), maximum = T)

sel_var <- sel_var |>
  mutate(cluster_fuzzy = assignment_fuzzy[cluster_fuzzy]) |>
  mutate(cluster_fuzzy = factor(cluster_fuzzy, labels = levels(category)))

#check agreement

table(sel_var$category, sel_var$cluster_fuzzy)

```

```

##
##           Coppice High Forest Old growth
## Coppice           7           2           0
## High Forest       0           6           2
## Old growth        1           2           8

```

```

#fuzzy Rand Index
RI.F(VC = sel_var$category, U = fuzzy$U)

```

```

## [1] 0.6613095

```

```

sink("other results/external_validation.txt")
"Fuzzy Rand Index"
RI.F(VC = sel_var$category, U = fuzzy$U)
sink()

```

```

#perform PCA after clustering, only for 2D plotting

```

```

pca_results <- sel_var |>
  dplyr::select(RH050, LAI) |>
  prcomp(scale. = T, rank. = 2)

```

```

sel_var <- cbind(sel_var, pca_results$x)

```

```

#plot PCA

```

```

#adapted from ggbiplot source code

```

```

#variance explained

```

```

expl_var <- round(pca_results$sdev^2/sum(pca_results$sdev^2)*100, 2)

```

```

labels <- str_c(c("PC1 ", "PC2 "),
  str_c("(", expl_var, "%"))

```

```

#how much to shift variable labels

```

```

lbl_shift <- 0.1

```

```

#loadings and labels

```

```

loadings_tbl <- as_tibble(pca_results$rotation, rownames = "var") |>
  mutate(xlab = PC1 + lbl_shift,
    ylab = PC2 + c(lbl_shift, -lbl_shift))

```

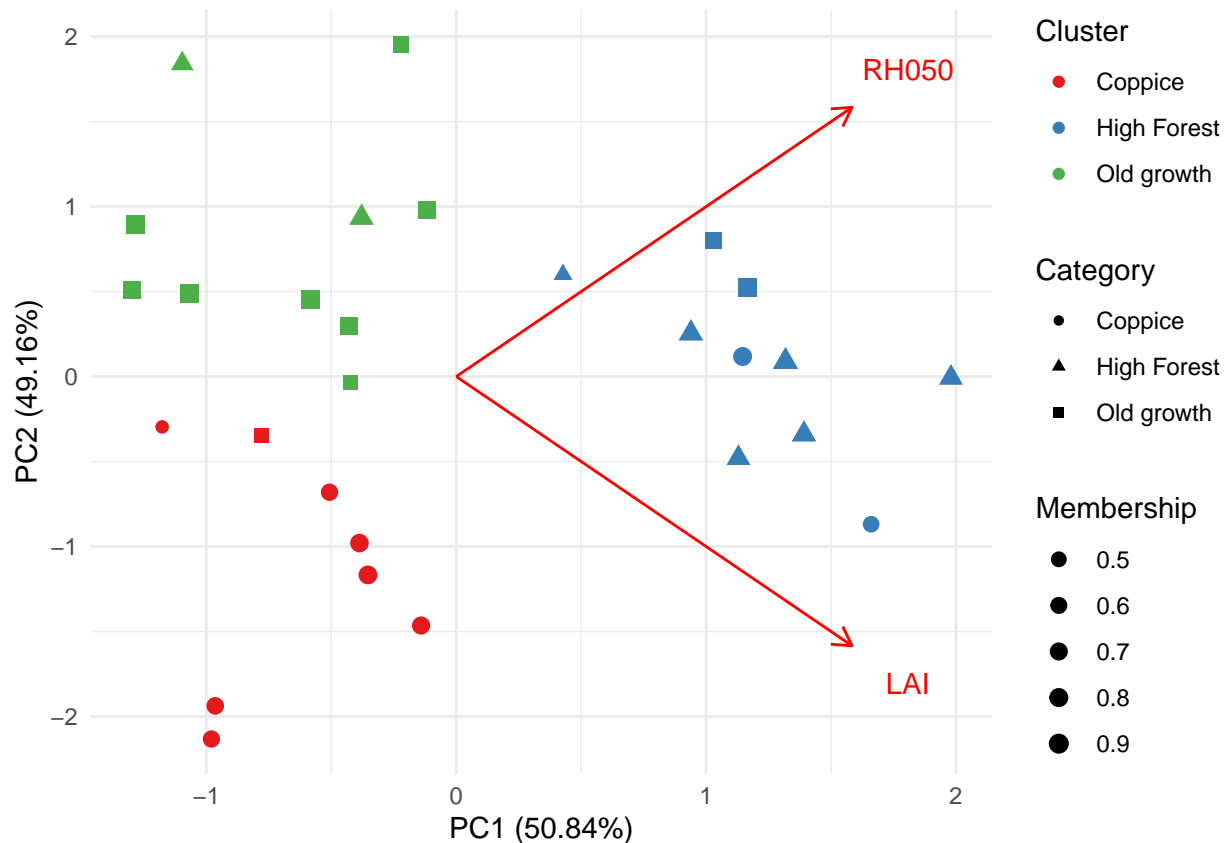
```

#default scaler from ggbiplot
scaler <- min(max(abs(sel_var$PC1))/max(abs(loadings_tbl$PC1)),
              max(abs(sel_var$PC2))/max(abs(loadings_tbl$PC2)))*0.8

loadings_tbl <- loadings_tbl |>
  mutate(across(PC1:ylab, ~ .x * scaler))

#clustering + PCA plot
sel_var |>
  ggplot()+
  geom_point(aes(x = PC1, y = PC2, color = cluster_fuzzy,
                 shape = category, size = membership)) +
  geom_segment(data = loadings_tbl, mapping = aes(x = 0, y = 0, xend = PC1,
                                                  yend = PC2),
              arrow = grid::arrow(length = grid::unit(8, "points")), colour = "red",
              linewidth = 0.5)+
  geom_text(data = loadings_tbl, aes(x = xlab, y = ylab, label = var), colour = "red")+
  scale_size_area(max_size = 3) +
  scale_color_brewer(palette = "Set1")+
  theme_minimal() +
  labs(color = "Cluster", shape = "Category", size = "Membership",
       x = labels[1], y = labels[2])

```




```
ggsave("plots/cluster_fuzzy_select.png", width = 190, height = 117,
       units = "mm", bg = 'white', scale = 1, dpi = 1000)
ggsave('plots/FIG2.eps', width = 190, height = 117,
       units = "mm", bg = 'white', scale = 1)
```

```
#summary statistics by cluster
sel_var |>
  group_by(cluster_fuzzy) |>
  summarise(across(3:5, mean))
```

```
## # A tibble: 3 x 4
##   cluster_fuzzy RH050    LAI membership
##   <fct>         <dbl> <dbl>      <dbl>
## 1 Coppice      -1.26  0.328    0.791
## 2 High Forest  0.910  0.813    0.855
## 3 Old growth   0.100 -1.08    0.817
```

```
# LDA -----

# vector of classes
class <- variables[[2]]

# standardize variables
vars <- variables[3:13]

# variable selection, criterion "ability to separate"
step <- stepclass(x = vars, grouping = class, method = "lda", direction = "both",
                  criterion = "AS", improvement = 0.1)
```

```
## `stepwise classification', using 10-fold cross-validated ability to separate of method lda'.
## 28 observations of 11 variables in 3 classes; direction: both
## stop criterion: improvement less than 10%.
```

```
## ability to separate: 0.3394; in: "LAI"; variables (1): LAI
## ability to separate: 0.58293; in: "RH050"; variables (2): LAI, RH050
##
## hr.elapsed min.elapsed sec.elapsed
##      0.000      0.000      0.161
```

```
wilks <- greedy.wilks(X = vars, grouping = class, niveau = 1) # keep going until all variables selected
```

```
write_csv(format(wilks$results, digits = 2), file = "other results/wilks_selection.csv")
```

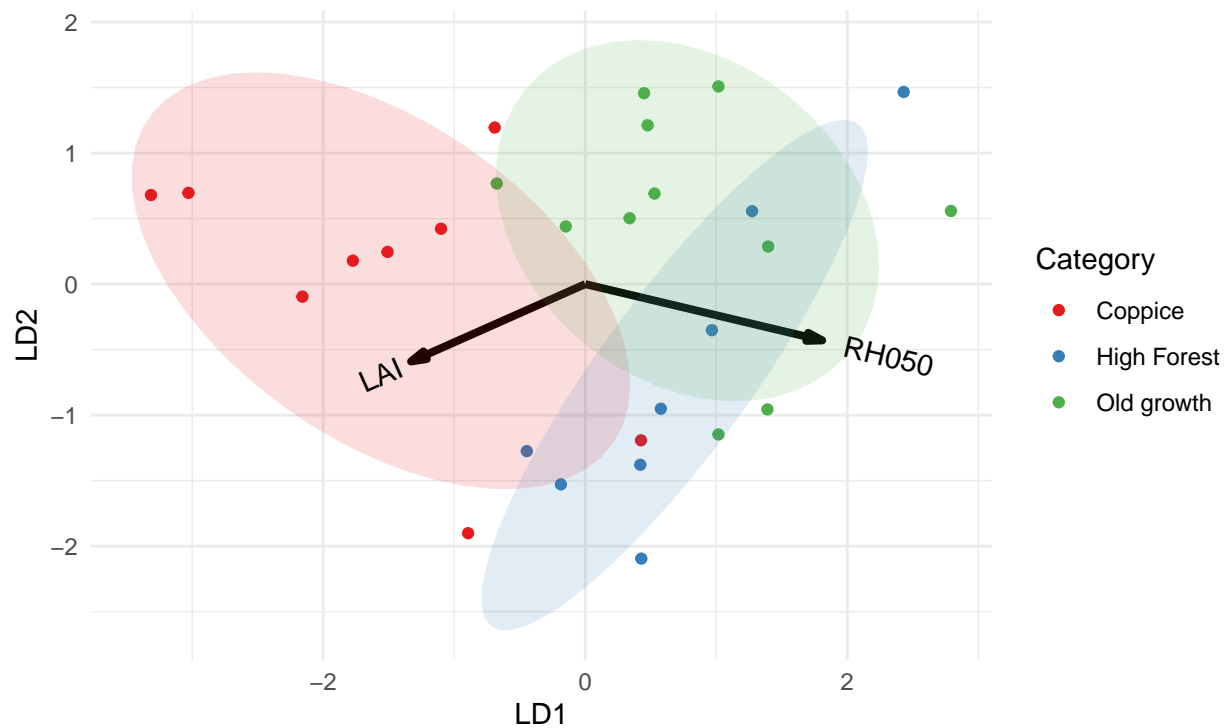
```
# LDA with selected variables
```

```
selected <- vars[step[["model"]][["nr"]]]
```

```
lda <- lda(x = selected, grouping = class)
```

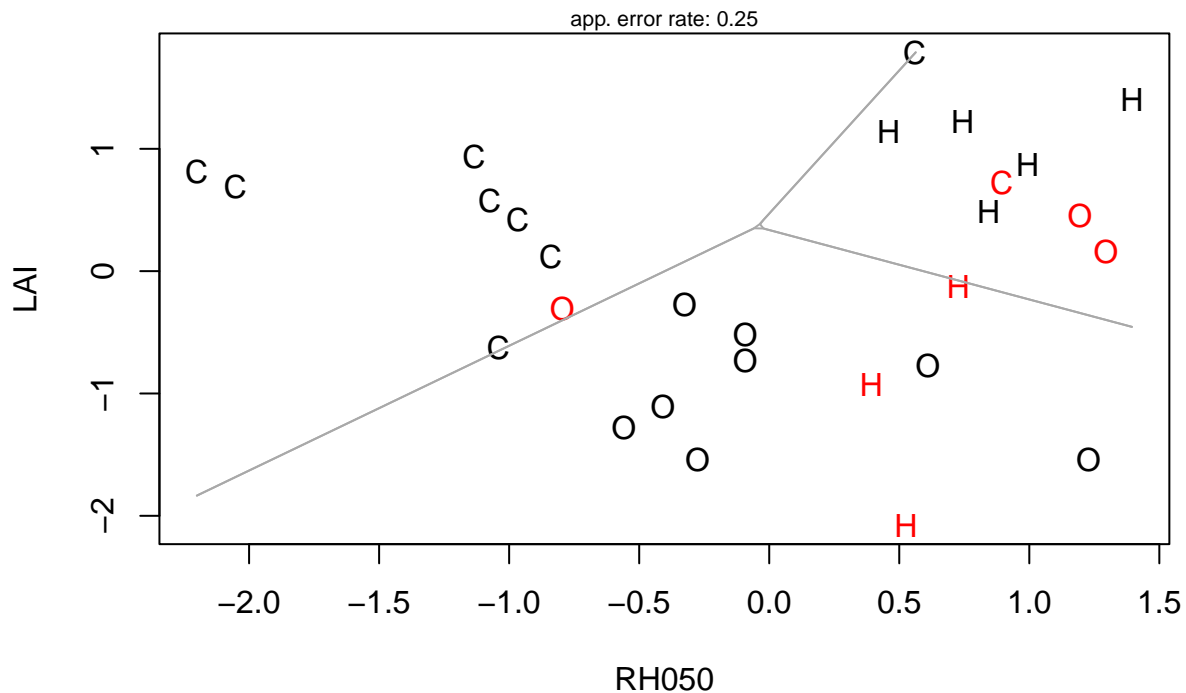
```
ggbiplot::ggbiplot(lda, groups = class, ellipse = T, varname.size = 4,
                   varname.adjust = 1.5,
                   ellipse.linewidth = NA, ellipse.alpha = 0.15, scale = 1)+
```

```
labs(x = "LD1", y = "LD2", color = "Category", fill = "Category")+
theme_minimal()+
scale_color_brewer(palette = "Set1")+
scale_fill_brewer(palette = "Set1")
```



```
ggsave("plots/LDA.png", width = 190, height = 117,
        units = "mm", bg = 'white', scale = 1.1, dpi = 1000)

drawparti(x = selected[[1]], y = selected[[2]], grouping = class, method = "lda",
          xlab = names(selected)[1], ylab = names(selected)[2], imageplot = F, col.mean = NULL)
```



```
png("plots/LDA_parti.png", bg = "white", width = 190, height = 117, units = "mm",
    pointsize = 12, res = 1000)
drawpart(x = selected[[1]], y = selected[[2]], grouping = class, method = "lda",
    xlab = names(selected)[1], ylab = names(selected)[2], imageplot = F, col.mean = NULL)
title("Partition plot")
dev.off()
```

```
## pdf
## 2
```

```
# session info -----
sessionInfo()
```

```
## R version 4.5.2 (2025-10-31)
## Platform: x86_64-redhat-linux-gnu
## Running under: Nobara Linux 43 (KDE Plasma Desktop Edition)
##
## Matrix products: default
## BLAS/LAPACK: FlexiBLAS OPENBLAS-OPENMP; LAPACK version 3.12.1
##
## locale:
## [1] LC_CTYPE=it_IT.UTF-8      LC_NUMERIC=C
## [3] LC_TIME=it_IT.utf8       LC_COLLATE=it_IT.UTF-8
## [5] LC_MONETARY=it_IT.utf8   LC_MESSAGES=it_IT.UTF-8
```

```

## [7] LC_PAPER=it_IT.utf8      LC_NAME=C
## [9] LC_ADDRESS=C               LC_TELEPHONE=C
## [11] LC_MEASUREMENT=it_IT.utf8 LC_IDENTIFICATION=C
##
## time zone: Europe/Rome
## tzcode source: system (glibc)
##
## attached base packages:
## [1] stats4      grid      stats      graphics  grDevices datasets  utils
## [8] methods     base
##
## other attached packages:
## [1] party_1.3-18      strucchange_1.5-4 sandwich_3.1-1    zoo_1.8-15
## [5] modeltools_0.2-24 mvtnorm_1.3-3     fclust_2.1.3      klaR_1.7-3
## [9] MASS_7.3-65       GGally_2.4.0      lubridate_1.9.4    forcats_1.0.1
## [13] stringr_1.6.0     dplyr_1.1.4       purrr_1.2.0        readr_2.1.6
## [17] tidyr_1.3.2       tibble_3.3.0      ggplot2_4.0.1      tidyverse_2.0.0
##
## loaded via a namespace (and not attached):
## [1] tidyselect_1.2.1  libcoin_1.0-10    farver_2.1.2       S7_0.2.1
## [5] fastmap_1.2.0     combinat_0.0-8    TH.data_1.1-5      promises_1.5.0
## [9] labelled_2.16.0   digest_0.6.39     timechange_0.3.0   mime_0.13
## [13] lifecycle_1.0.4   cluster_2.1.8.1   survival_3.8-3     magrittr_2.0.4
## [17] compiler_4.5.2     rlang_1.1.6       tools_4.5.2        ggbiplot_0.6.2
## [21] utf8_1.2.6        yaml_2.3.12       knitr_1.50          labeling_0.4.3
## [25] bit_4.6.0         RColorBrewer_1.1-3 multcomp_1.4-29     miniUI_0.1.2
## [29] withr_3.0.2       xtable_1.8-4      scales_1.4.0        cli_3.6.5
## [33] rmarkdown_2.30    crayon_1.5.3      ragg_1.5.0          generics_0.1.4
## [37] otel_0.2.0        rstudioapi_0.17.1 tzdb_0.5.0          splines_4.5.2
## [41] parallel_4.5.2    CoprManager_0.5.7 matrixStats_1.5.0   vctrs_0.6.5
## [45] Matrix_1.7-4      hms_1.1.4         bit64_4.6.0-1       clue_0.3-66
## [49] systemfonts_1.3.1 glue_1.8.0         ggstats_0.12.0      codetools_0.2-20
## [53] stringi_1.8.7     gtable_0.3.6      questionr_0.8.1     later_1.4.4
## [57] pillar_1.11.1     htmltools_0.5.9   R6_2.6.1            textshaping_1.0.4
## [61] vroom_1.6.7       evaluate_1.0.5    shiny_1.12.1        lattice_0.22-7
## [65] haven_2.5.5       highr_0.11        httpuv_1.6.16       Rcpp_1.1.0
## [69] xfun_0.55         coin_1.4-3        pkgconfig_2.0.3

```