

Indices based on rotated principal components

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```
# setup -----

library(tidyverse)

## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr      1.1.4      v readr      2.1.6
## v forcats    1.0.1      v stringr    1.6.0
## v ggplot2     4.0.1      v tibble     3.3.0
## v lubridate   1.9.4      v tidyr      1.3.2
## v purrr       1.2.0
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()     masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors

library(GET)
library(vegan)

## Loading required package: permute

library(psych)

##
## Attaching package: 'psych'
##
## The following object is masked from 'package:vegan':
##
##     pca
##
## The following objects are masked from 'package:ggplot2':
##
##     %+%, alpha

resampled <- readRDS("transect_resampled.rds")
speciestraits <- readRDS("traits_imputation.rds")
source("0.functions.R")

indicesaxes <- vector(mode = "list")
curvesaxes <- vector(mode = "list")
```

```
# transform traits -----
```

```
data <- speciestraits |>
  select(species, where(is.numeric)) |>
  select(-n) |>
  mutate(across(where(is.numeric), ~ boxcox(.))) |>
  as.data.frame()
```

```
## Box-Cox transform with lambda = 0.1
## Box-Cox transform with lambda = -0.5
## Box-Cox transform with lambda = -0.4
## Box-Cox transform with lambda = 0.3
## Box-Cox transform with lambda = 0.2
## Box-Cox transform with lambda = 0
## Box-Cox transform with lambda = 0.8
## Box-Cox transform with lambda = 1.8
## Box-Cox transform with lambda = -0.2
## Box-Cox transform with lambda = 0
```

```
# check dimensions -----
```

```
PCA_above <- data |>
  select(2:7) |>
  psych::principal(nfactors = 2, rotate = "varimax")
```

```
PCA_above
```

```
## Principal Components Analysis
## Call: psych::principal(r = select(data, 2:7), nfactors = 2, rotate = "varimax")
## Standardized loadings (pattern matrix) based upon correlation matrix
##
```

	RC1	RC2	h2	u2	com
## leaf_area_mm2	-0.06	0.79	0.63	0.37	1.0
## nmass_mg_g	0.79	0.32	0.73	0.27	1.3
## lma_g_m2	-0.68	0.32	0.57	0.43	1.4
## plant_height_m	0.04	0.71	0.51	0.49	1.0
## diaspore_mass_mg	0.54	-0.23	0.35	0.65	1.3
## ssd_combined_mg_mm3	-0.54	-0.30	0.38	0.62	1.6

```
##
```

	RC1	RC2
## SS loadings	1.68	1.49
## Proportion Var	0.28	0.25
## Cumulative Var	0.28	0.53
## Proportion Explained	0.53	0.47
## Cumulative Proportion	0.53	1.00

```
##
```

Mean item complexity = 1.3

Test of the hypothesis that 2 components are sufficient.

```
##
```

The root mean square of the residuals (RMSR) is 0.17

with the empirical chi square 67.8 with prob < 6.6e-14

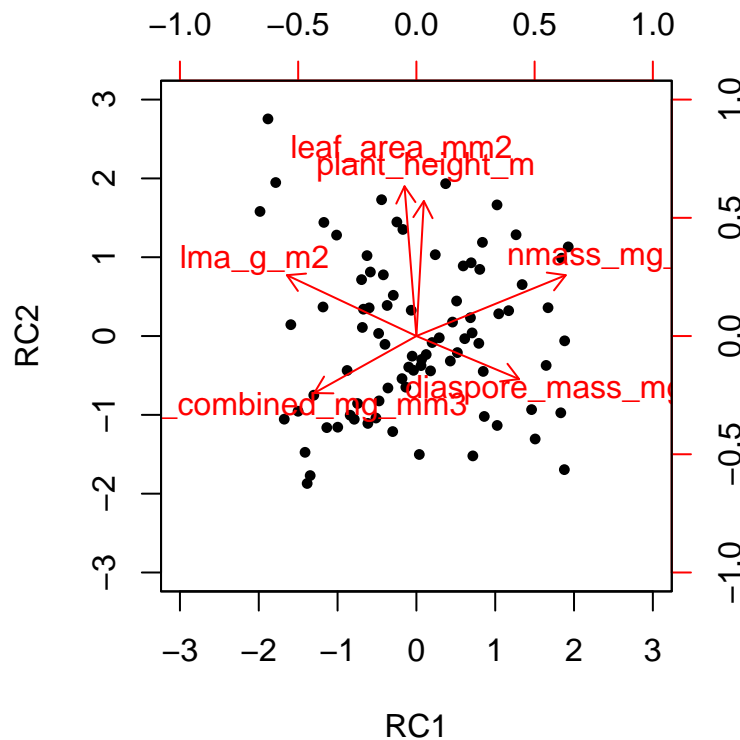
```
##
```

Fit based upon off diagonal values = 0.38

```
png('plots/PCA_above.png', bg = "white", width = 7, height = 7, units = "in",
    res = 1000)
biplot(PCA_above, main = NULL)
dev.off()
```

```
## pdf
## 2
```

```
biplot(PCA_above, main = NULL)
```



```
data <- cbind(data, PCA_above$scores) |>
  rename(above1 = RC1, above2 = RC2)

PCA_clonal <- data |>
  select(8:11) |>
  psych::principal(nfactors = 2, rotate = "varimax")

PCA_clonal
```

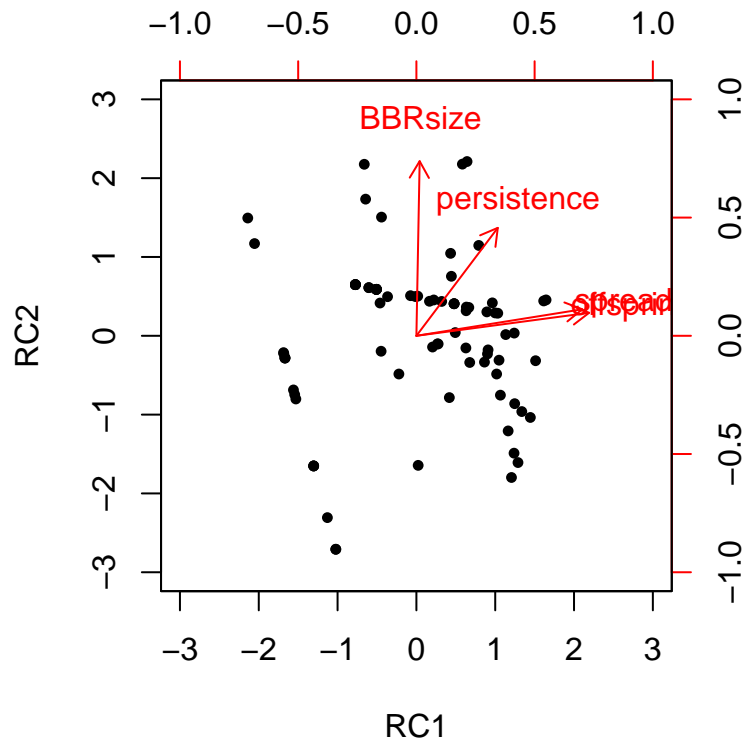
```
## Principal Components Analysis
## Call: psych::principal(r = select(data, 8:11), nfactors = 2, rotate = "varimax")
## Standardized loadings (pattern matrix) based upon correlation matrix
##           RC1  RC2  h2  u2 com
## BBRsize    0.02 0.92 0.85 0.15 1.0
```

```
## persistence 0.43 0.57 0.51 0.49 1.9
## offspring   0.91 0.12 0.85 0.15 1.0
## spread      0.88 0.14 0.79 0.21 1.1
##
##              RC1  RC2
## SS loadings    1.79 1.21
## Proportion Var  0.45 0.30
## Cumulative Var  0.45 0.75
## Proportion Explained 0.60 0.40
## Cumulative Proportion 0.60 1.00
##
## Mean item complexity = 1.2
## Test of the hypothesis that 2 components are sufficient.
##
## The root mean square of the residuals (RMSR) is 0.15
## with the empirical chi square 21.43 with prob < NA
##
## Fit based upon off diagonal values = 0.85
```

```
png('plots/PCA_clonal.png', bg = "white", width = 7, height = 7, units = "in",
    res = 1000)
biplot(PCA_clonal, main = NULL)
dev.off()
```

```
## pdf
## 2
```

```
biplot(PCA_clonal, main = NULL)
```



```
data <- cbind(data, PCA_clonal$scores) |>
  rename(clonal1 = RC1, clonal2 = RC2)

# aboveground-----

## aboveground RC1 -----
### calculate distances -----

distances <- data |>
  select(above1) |>
  dist(method = "euclidean", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1

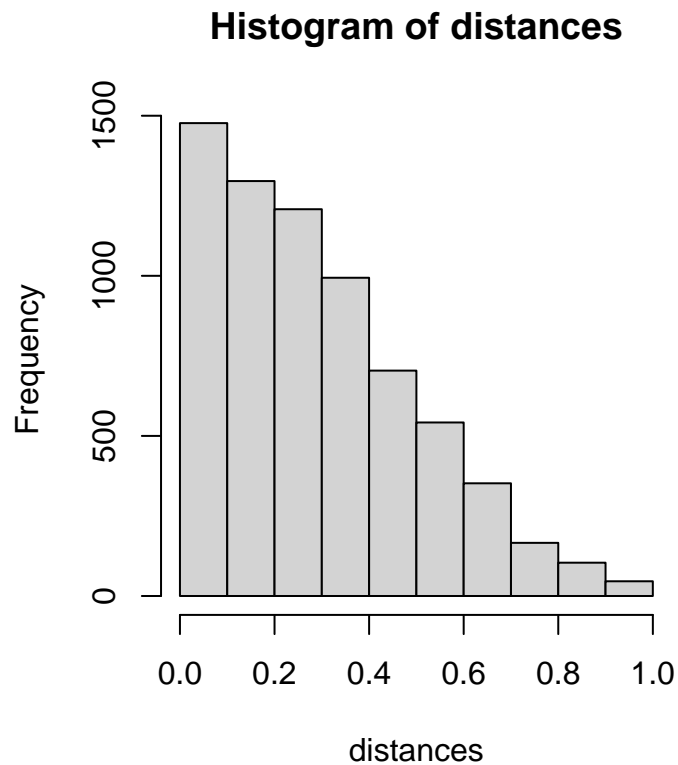
distances <- distances / max(distances)

#double checking the squared euclidean property
ade4::is.euclid(as.dist(sqrt(distances)))

## [1] TRUE
```

```
#check distance distribution
```

```
hist(distances)
```



```
#pairs with distance 0
```

```
as.data.frame(as.table(distances)) |>  
  filter(Freq == 0 & Var1 != Var2)
```

```
## [1] Var1 Var2 Freq  
## <0 rows> (or 0-length row.names)
```

```
### calculate indices -----
```

```
species <- data$species
```

```
raotrait <- lapply(resampled, function (x)  
  lapply(x, function (y) qdecomp(y, distances, species)) |>  
    bind_rows(.id = "step") |>  
    mutate(step = as.numeric(step))  
) |>  
  bind_rows(.id = "site")
```

```
#scale steps in units of area
```

```

raotrait <- raotrait |>
  mutate(step = step / 100)

#add to list
indicesaxes[[1]] <- raotrait
names(indicesaxes)[1] <- "above1"

### create curve set -----

curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

curvesaxes[[1]] <- curvesets
names(curvesaxes)[1] <- "above1"

## aboveground RC2 -----
### calculate distances -----

distances <- data |>
  select(above2) |>
  dist(method = "euclidean", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1

distances <- distances / max(distances)

#double checking the squared euclidean property

ade4::is.euclid(as.dist(sqrt(distances)))

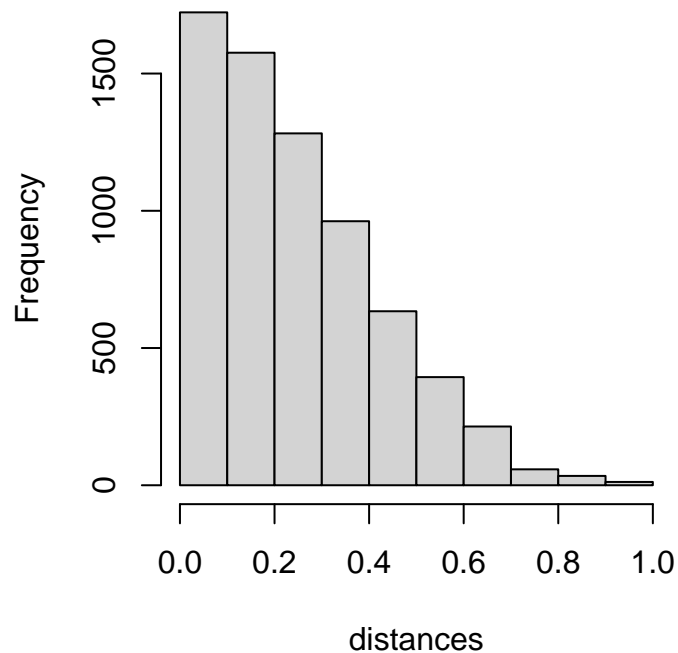
## [1] TRUE

#check distance distribution

hist(distances)

```

Histogram of distances



```
#pairs with distance 0
```

```
as.data.frame(as.table(distances)) |>
  filter(Freq == 0 & Var1 != Var2)
```

```
## [1] Var1 Var2 Freq
## <0 rows> (or 0-length row.names)
```

```
### calculate indices -----
```

```
species <- data$species
```

```
raotrait <- lapply(resampled, function (x)
  lapply(x, function (y) qdecomp(y, distances, species)) |>
    bind_rows(.id = "step") |>
    mutate(step = as.numeric(step))
) |>
  bind_rows(.id = "site")
```

```
#scale steps in units of area
```

```
raotrait <- raotrait |>
  mutate(step = step / 100)
```

```
#add to list
```



```

indicesaxes[[2]] <- raotrait
names(indicesaxes)[2] <- "above2"

### create curve set -----

curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

curvesaxes[[2]] <- curvesets
names(curvesaxes)[2] <- "above2"

# clonal -----

## clonal RC1 -----
### calculate distances -----

distances <- data |>
  select(clonal1) |>
  dist(method = "euclidean", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1

distances <- distances / max(distances)

#double checking the squared euclidean property

ade4::is.euclid(as.dist(sqrt(distances)))

## Warning in ade4::is.euclid(as.dist(sqrt(distances))): Zero distance(s)

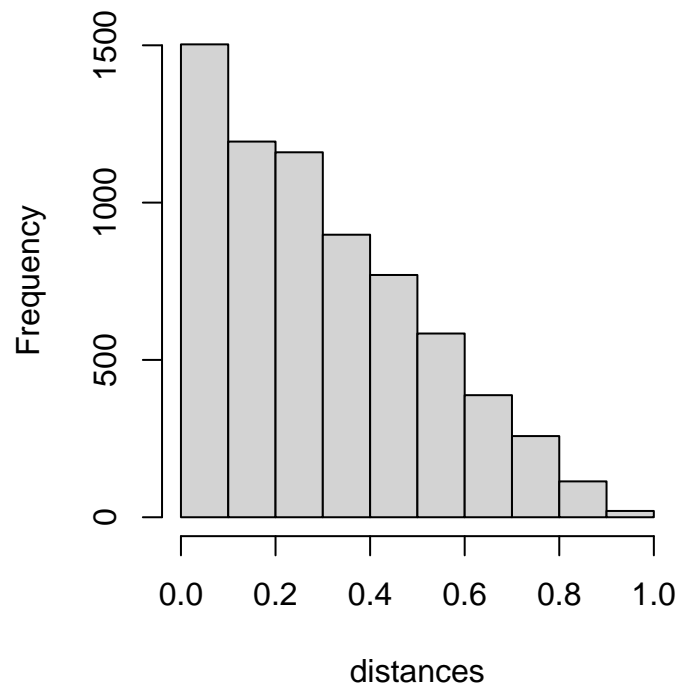
## [1] TRUE

#check distance distribution

hist(distances)

```

Histogram of distances



```
#pairs with distance 0
```

```
as.data.frame(as.table(distances)) |>  
  filter(Freq == 0 & Var1 != Var2)
```

```
##   Var1 Var2 Freq  
## 1   10   1    0  
## 2   29   1    0  
## 3   44   1    0  
## 4   49   1    0  
## 5   67   1    0  
## 6    1  10    0  
## 7   29  10    0  
## 8   44  10    0  
## 9   49  10    0  
## 10  67  10    0  
## 11  19  15    0  
## 12  37  15    0  
## 13  34  17    0  
## 14  75  17    0  
## 15  15  19    0  
## 16  37  19    0  
## 17  48  28    0  
## 18  66  28    0  
## 19   1  29    0
```

##	20	10	29	0
##	21	44	29	0
##	22	49	29	0
##	23	67	29	0
##	24	35	31	0
##	25	36	31	0
##	26	40	31	0
##	27	77	31	0
##	28	17	34	0
##	29	75	34	0
##	30	31	35	0
##	31	36	35	0
##	32	40	35	0
##	33	77	35	0
##	34	31	36	0
##	35	35	36	0
##	36	40	36	0
##	37	77	36	0
##	38	15	37	0
##	39	19	37	0
##	40	61	39	0
##	41	31	40	0
##	42	35	40	0
##	43	36	40	0
##	44	77	40	0
##	45	1	44	0
##	46	10	44	0
##	47	29	44	0
##	48	49	44	0
##	49	67	44	0
##	50	28	48	0
##	51	66	48	0
##	52	1	49	0
##	53	10	49	0
##	54	29	49	0
##	55	44	49	0
##	56	67	49	0
##	57	39	61	0
##	58	28	66	0
##	59	48	66	0
##	60	1	67	0
##	61	10	67	0
##	62	29	67	0
##	63	44	67	0
##	64	49	67	0
##	65	17	75	0
##	66	34	75	0
##	67	31	77	0
##	68	35	77	0
##	69	36	77	0
##	70	40	77	0
##	71	82	80	0
##	72	80	82	0

```

### calculate indices -----

species <- data$species

raotrait <- lapply(resampled, function (x)
  lapply(x, function (y) qdecomp(y, distances, species)) |>
  bind_rows(.id = "step") |>
  mutate(step = as.numeric(step))
) |>
  bind_rows(.id = "site")

#scale steps in units of area

raotrait <- raotrait |>
  mutate(step = step / 100)

#add to list
indicesaxes[[3]] <- raotrait
names(indicesaxes)[3] <- "clonal1"

### create curve set -----

curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

curvesaxes[[3]] <- curvesets
names(curvesaxes)[3] <- "clonal1"

## clonal RC2 -----
### calculate distances -----

distances <- data |>
  select(clonal2) |>
  dist(method = "euclidean", diag = T, upper = T) |>
  as.matrix()

#scaling to 0-1

distances <- distances / max(distances)

#double checking the squared euclidean property

ade4::is.euclid(as.dist(sqrt(distances)))

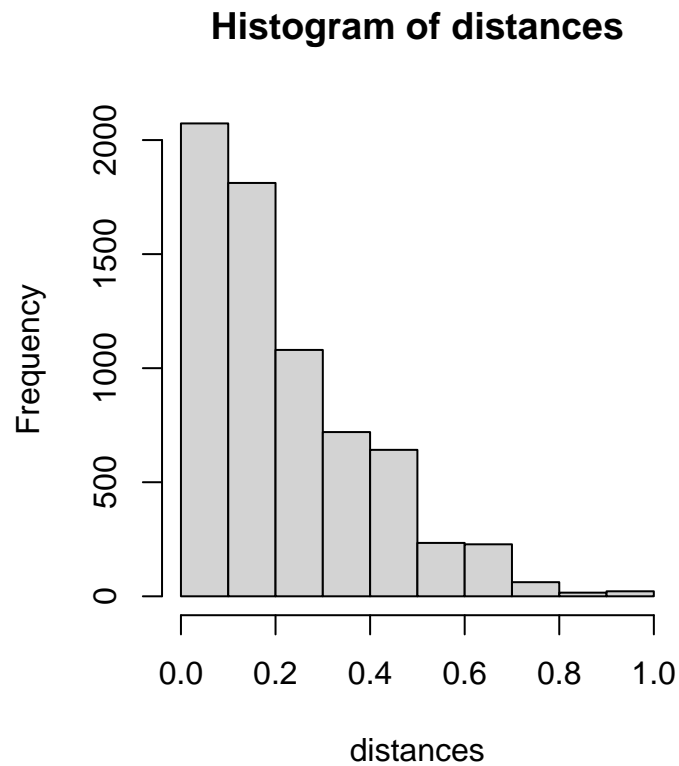
```

```
## Warning in ade4::is.euclid(as.dist(sqrt(distances))): Zero distance(s)
```

```
## [1] TRUE
```

```
#check distance distribution
```

```
hist(distances)
```



```
#pairs with distance 0
```

```
as.data.frame(as.table(distances)) |>  
  filter(Freq == 0 & Var1 != Var2)
```

```
##   Var1 Var2 Freq  
## 1   10   1    0  
## 2   29   1    0  
## 3   44   1    0  
## 4   49   1    0  
## 5   67   1    0  
## 6    1  10    0  
## 7   29  10    0  
## 8   44  10    0  
## 9   49  10    0  
## 10  67  10    0  
## 11  19  15    0
```

##	12	37	15	0
##	13	34	17	0
##	14	75	17	0
##	15	15	19	0
##	16	37	19	0
##	17	48	28	0
##	18	66	28	0
##	19	1	29	0
##	20	10	29	0
##	21	44	29	0
##	22	49	29	0
##	23	67	29	0
##	24	35	31	0
##	25	36	31	0
##	26	40	31	0
##	27	77	31	0
##	28	17	34	0
##	29	75	34	0
##	30	31	35	0
##	31	36	35	0
##	32	40	35	0
##	33	77	35	0
##	34	31	36	0
##	35	35	36	0
##	36	40	36	0
##	37	77	36	0
##	38	15	37	0
##	39	19	37	0
##	40	61	39	0
##	41	31	40	0
##	42	35	40	0
##	43	36	40	0
##	44	77	40	0
##	45	1	44	0
##	46	10	44	0
##	47	29	44	0
##	48	49	44	0
##	49	67	44	0
##	50	28	48	0
##	51	66	48	0
##	52	1	49	0
##	53	10	49	0
##	54	29	49	0
##	55	44	49	0
##	56	67	49	0
##	57	39	61	0
##	58	28	66	0
##	59	48	66	0
##	60	1	67	0
##	61	10	67	0
##	62	29	67	0
##	63	44	67	0
##	64	49	67	0
##	65	17	75	0

```
## 66  34  75  0
## 67  31  77  0
## 68  35  77  0
## 69  36  77  0
## 70  40  77  0
## 71  82  80  0
## 72  80  82  0
```

```
### calculate indices -----

species <- data$species

raotrait <- lapply(resampled, function (x)
  lapply(x, function (y) qdecomp(y, distances, species)) |>
  bind_rows(.id = "step") |>
  mutate(step = as.numeric(step))
) |>
  bind_rows(.id = "site")

#scale steps in units of area

raotrait <- raotrait |>
  mutate(step = step / 100)

#add to list
indicesaxes[[4]] <- raotrait
names(indicesaxes)[4] <- "clonal2"

### create curve set -----

curvesets <- vector(mode = "list", length = sum(map_lgl(raotrait, is.numeric))-1)

for (i in seq_along(curvesets)){
  column <- colnames(raotrait)[i+2]
  curves <- raotrait |>
    select(site, step, {{column}}) |>
    pivot_wider(names_from = site, values_from = 3) |>
    filter(if_all(where(is.numeric), ~ !is.na(.)))
  curvesets[[i]] <- curve_set(obs = as.data.frame(curves[-1]), r = curves[[1]])
  names(curvesets)[i] <- column
}

curvesaxes[[4]] <- curvesets
names(curvesaxes)[4] <- "clonal2"

# save results -----

saveRDS(indicesaxes, "indices_axes.rds")
saveRDS(curvesaxes, "curves_axes.rds")
```