ContinuumElectrostatics

User Manual

Last update: June 16, 2014

1 Introduction

Proteins contain residues, cofactors and ligands that bind or release protons depending on the

current pH and the interactions with their molecular environment. These titratable residues,

cofactors and ligands will be referred to as sites. The titration of proteins is often difficult

to study experimentally because the available methods, such as calorimetry, cannot determine

protonation states of individual sites. The knowledge of these individual protonation states is

crucial for understanding of many important processes, for example enzyme catalysis.

The Continuum Electrostatics module extends the functionality of the pDynamo library with

a Poisson-Boltzmann continuum electrostatic model that allows for calculations of protonation

states of individual sites. The module provides an interface between pDynamo and the external

solver of the Poisson-Boltzmann equation, MEAD. MEAD is a program developed by Donald

Bashford and extended by Timm Essigke and Thomas Ullmann. The electrostatic energy terms

obtained with MEAD can be used to calculate energies of all possible protonation states of the

protein of interest. However, analytic evaluation of protonation state energies is only possible

for proteins with only a few titratable sites. The Continuum Electrostatics module also provides

an interface to the GMCT program by Matthias Ullmann and Thomas Ullmann that can be

used to sample protonation state energies using a Monte Carlo method. The GMCT interface

allows for studying the titration of larger proteins.

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2 Copying

The module is distributed under the CeCILL Free Software License, which is a French equivalent of the GNU General Public License. For details, see the files Licence_CeCILL_V2-en.txt (or Licence_CeCILL_V2-fr.txt for the French version).

3 Installation and configuration

Before the installation of the Continuum Electrostatics module, it is necessary to have:

- pDynamo 1.8.0
- Python 2.7
- GCC (any version should be fine)
- Extended MEAD 2.3.0
- GMCT 1.2.3

Extended MEAD and GMCT can be found on the website of Thomas Ullmann:

http://www.bisb.uni-bayreuth.de/People/ullmannt/index.php?name=software

Download the two packages and follow their respective installation instructions. The ContinuumElectrostatics module requires for its functioning two programs from the MEAD package, namely my_2diel_solver and my_3diel_solver, and the GMCT's main program, gmct.

In the next step, check out the latest source code of the module. Note that for checking out the source code you should have Subversion installed as well.

svn checkout

http://pdynamo-extensions.googlecode.com/svn/trunk/ContinuumElectrostatics/

Some parts of the module implementing the state vector are written in C and therefore have to be compiled before use. In the future, I plan to shift some more parts of the code from Python to C.

Go to the subdirectory extensions/csource and edit the Makefile. Only the uppermost line has to be changed. It defines the directory where you have installed pDynamo. After editing, close the file and type in make to compile the C object file.

Go to the subdirectory extensions/pyrex and again edit the Makefile. Change the line starting from "INC2" to the location of your pDynamo installation. Close the file and type in make. It should generate a dynamically linked library StateVector.so in the ContinuumElectrostatics subdirectory.

At this point, the installation is complete.

Before using the module, you have to set the environment variable

PDYNAMO_CONTINUUMELECTROSTATICS to the directory where you have checked out the source code. Also, you have to add this directory to the PYTHONPATH variable. For example, you can do it this way (in Bash):

export

PDYNAMO_CONTINUUMELECTROSTATICS=/home/mikolaj/devel/ContinuumElectrostatics export PYTHONPATH=\$PYTHONPATH:\$PDYNAMO_CONTINUUMELECTROSTATICS

4 Usage

After the installation, it may be worth looking at some of the test cases in the subdirectory tests. I will explain the functioning of the module based on the test case "sites2". This test uses a trivial polypeptide with only two titratable sites, histidine and glutamate. The test "histidine" uses only one site. The other tests use real-life proteins.

In the first step, prepare CHARMM topology (psf) and coordinate (crd) files.

During the initial setup in CHARMM, all titratable residues in the protein should be set to their standard protonation states at pH = 7, i.e. aspartates and glutamates deprotonated, histidines doubly protonated, other residues protonated.

5 References

MEAD website:

http://stjuderesearch.org/site/lab/bashford/

Extended MEAD website:

http://www.bisb.uni-bayreuth.de/People/ullmannt/index.php?name=extended-mead

Doctoral thesis of Timm Essigke:

https://epub.uni-bayreuth.de/655/

6 Test cases

7 To-do list

- Move parts of WriteJobFiles to the instance class
- Rename variables containing filenames so that they start from "file"
- Coordinates from the FPT file should have their own data structure
- Efficiency improvements during writing job files
- Use arrays instead of lists for interactions (Real1DArray or SymmetricMatrix)
- Have a column of ETA (Estimated Time for Accomplishment) in MEAD calculations
- Optionally convert kcal/mol (MEAD units) to kJ/mol (pDynamo units)
- Make use of the pqr2SolvAccVol program to speed up the calculations a little bit
- The function calculating Gmicro should be written in C