S.pombe - Transformation (LiAOc)

From: Hyun Soo Kim, Keogh lab, AECOM

- 1. Grow *S.pombe* cells in EMM or YES to 1 x 10⁷ cells/ml (OD_{λ 595} \approx 0.5)
- 2. Pellet 5 ml cells per transformation (3000 rpm / 3 min) (pellet volume 30-50µl)
- 3. Resuspend cells with 1ml of ddH_2O and transfer to microfuge tube. Flash spin (10K, 1 sec) and wash with 500µl LiTE (0.1M LiAc/1X TE)
- 4. Resuspend pellet in 100µl LiTE
- 5. Add 20μg <u>carrier DNA</u> (2μl of 10μg/μl ssDNA) and ≤10μl transformation <u>DNA</u> (e.g PCR product) and mix gently (by pipetting or weak vortexing)
- 6. Incubate RT°C / 10 min
- 7. Add 260µl PLATE (40% PEG/0.1M LiAc/1X TE) and mix gently by pipetting
- 8. Incubate <u>30-60 min</u> at <u>30°C</u> (or RT°C for temperature sensitive strains)
- 9. Add 43µl DMSO; mix gently by pipetting or vortexing
- 10. Heat shock: 42°C for 5 min
- 11. Cool down samples for 1-2 minutes / RT°C. Pellet and wash once with 1ml ddH₂O.
- 12. Pellet by centrifugation, discard supernatant and resuspend in 200μl water. Plate and incubate as appropriate. Expect ≥ 100 colonies.

Solutions -

10X TE (pH 7.5): 100mM Tris-HCl (pH 7.5) 10mM EDTA

1M LiAc (pH 7.5): 102.02g LiAc to 800ml ddH_2O . Adjust pH (10% acetic acid), make to 1L and autoclave 1M Tris HCl (pH 7.5): 121.1g Tris to 800ml ddH_2O . Adjust pH with HCl, make to 1L and autoclave

50% PEG 4000 (PEG3350): Dissolve 250g of PEG3350 in 350ml of ddH_2O (heat to >50°C with stirring). Make to 500ml when completely dissolved and sterilize by autoclaving.

Original: Lithium Acetate Procedure II

(from Nurse Fission Yeast Handbook)

- 1. Grow fission yeast cells in MM to 1x107 cells/ml.
- 2. Pellet 50ml of cells per transformation.
- 3. Wash cells in 50ml sterile water. Transfer to eppis in 1ml water. Wash in 1ml of LiAc-TE.
- 4. Resuspend in LiAc-TE at 2x109 cells/ml (1/200 original volume).
- 5. Mix 100µl cells with 2µl carrier DNA at 10mg/ml and up to 10µl of DNA; mix gently.
- 6. Incubate at RT for 10 min.
- 7. Add 260ul of 40% PEG/LiAc-TE; mix gently.
- 8. Incubate 30-60 min at 29°C-30°C, or lower for temp. sensitive strains.
- 9. Add 43µl pre-warmed DMSO; mix gently.
- 10. Heat shock at 42°C for 5 min.
- 11. Pellet and wash once with 1ml water.
- 12. Pellet and resuspend in 500µl water and plate 250µl in duplicate.