DateLife Workflows

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Taxon Cetacea

I. Query data

There are 198 species in the Open Tree of Life Taxonomy for the taxon Cetacea. Information on time of divergence is available for 89 of these species across 6 published and peer-reviewed chronograms. Original study citations as well as proportion of Cetacea species found across those source chronograms is shown in Table 1.

All source chronograms are fully ultrametric.

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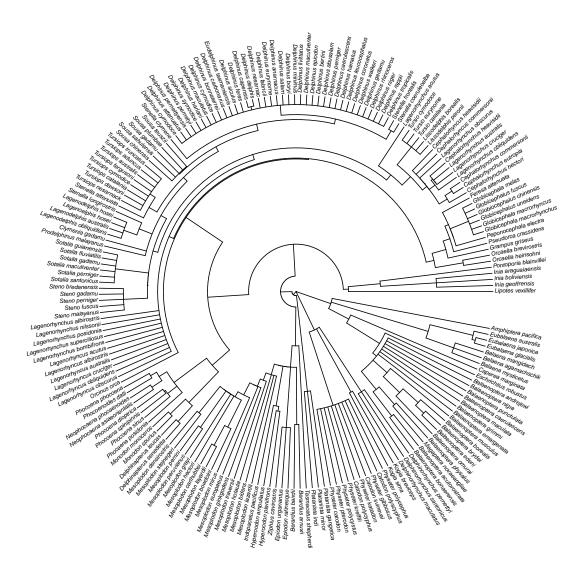


Figure 1: Cetacea Species Dated Open Tree of Life Induced Subtree. This chronogram was obtained with get_dated_otol_induced_subtree() function.

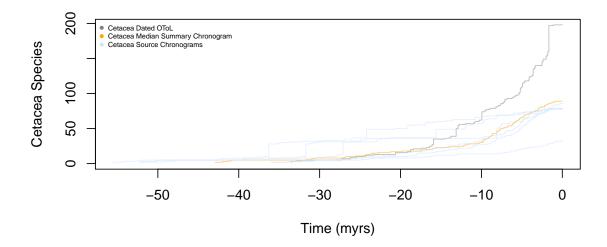


Figure 2: Cetacea lineage through time (LTT) plots from source chronograms, summary median chronogram and dated Open Tree of Life chronogram.

II. Summarize results.

II.A. Diagnosing clustering issues.

We identified some issues with chronograms coming from SDM and Median summary matrices. First, clustering algorithms used to go from a summary distance matrix to a tree return trees that are too old (generally with UPGMA algorithms) or non-ultrametric (generally with Neighbour Joining algorithms). In most studied cases, UPGMA returns fully ultrametric trees but with very old ages (we had to multiply the matrix by 0.25 to get ages approximate to source chronograms ages, however this is a number chosen at random, it was just the number that worked well). NJ returned reasonable ages, but trees are way non ultrametric, as you can see in Fig. 3 and Fig. 4.

This taxon's SDM matrix has some negative values in the following taxa: *Eubalaena japonica*, *Eubalaena glacialis*. This taxon's Median matrix has NO negative values.

II.B. Age distributions form Median and SDM summary trees.

Comparison of summary chronograms reconstructed with min and max ages.

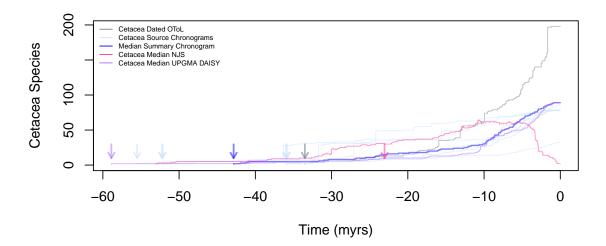


Figure 3: Cetacea lineage through time (LTT) plots from source chronograms and Median summary matrix converted to phylo with different methods (NJ and UPGMA). Clustering algorithms used often are returning non-ultrametric trees or with maximum ages that are just off (too old or too young). So we developed an alternative algorithm in datelife to go from a summary matrix to a fully ultrametric tree.

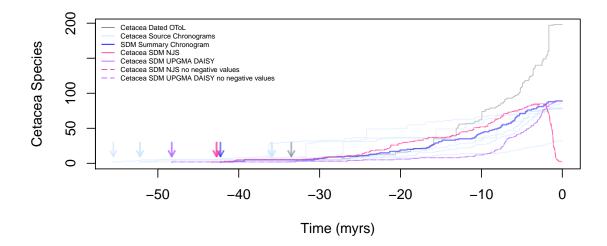


Figure 4: Cetacea lineage through time (LTT) plots from source chronograms and SDM summary matrix converted to phylo with different methods (NJ and UPGMA). As you can note, dashed lines and solid lines from trees coming out from both types of clustering algorithms implemented are mostly overlapping. This means that removing negative values does not change results from clustering algorithms much. Clustering algorithms used often are returning non-ultrametric trees or with maximum ages that are just off (too old or too young). So we developed an alternative algorithm in datelife to go from a summary matrix to a fully ultrametric tree.

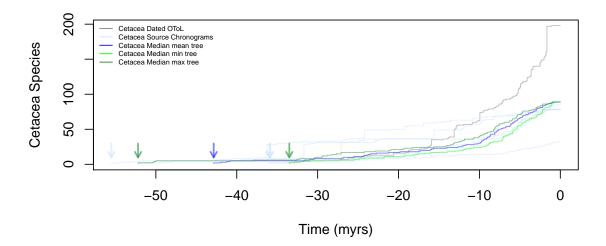


Figure 5: Cetacea lineage through time (LTT) plots from source chronograms and Median summary matrix converted to phylo with datelife algorithm.

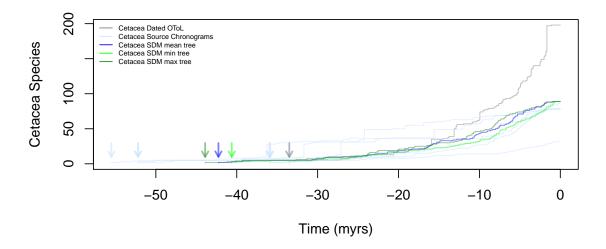


Figure 6: Cetacea lineage through time (LTT) plots from source chronograms and SDM summary matrix converted to phylo with datelife algorithm.

III. Create new data

As an example, we're gonna date the Open Tree Synthetic tree (mainly because the taxonomic tree is usually less well resolved.)

Now, let's say you like the Open Tree of Life Taxonomy and you want to stick to that tree. Dates from available studies were tested over the Open Tree of Life Synthetic tree of Cetacea and a tree was constructed, but all branch lengths are NA. We also tried each source chronogram independently, with the Dated OToL and with each other, as a form of cross validation in Table 2. This is not working perfectly yet, but we are developping new ways to use all calibrations efficiently.

Table 1: Was it successful to use each source chronogram independently as calibration (CalibN) against the Dated Open Tree of Life (dOToL) and each other (ChronoN)?

	dOToL	Chrono1	Chrono2	Chrono3	Chrono4	Chrono5	Chrono6
Calibrations1	TRUE	TRUE	FALSE	TRUE	FALSE	TRUE	TRUE
Calibrations2	TRUE	TRUE	FALSE	TRUE	FALSE	TRUE	TRUE
Calibrations3	TRUE	TRUE	FALSE	TRUE	FALSE	TRUE	TRUE
Calibrations4	TRUE	TRUE	FALSE	TRUE	FALSE	TRUE	TRUE
Calibrations5	TRUE	TRUE	FALSE	TRUE	FALSE	TRUE	TRUE
Calibrations6	TRUE	TRUE	FALSE	TRUE	FALSE	TRUE	TRUE

III. Simulate data

An alternative to generate a dated tree from a set of taxa is to take the available information and simulate into it the missing data. We will take the median and sdm summary chronograms to date the Synthetic tree of Life:

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Appendix

The following species were completely absent from the chronogram data base: Amphiptera pacifica, Balaena aqamachschik, Balaena manqidach, Balaenoptera andrejewi, Balaenoptera caerulescens, Balaenoptera emargenata, Balaenoptera grimmi, Balaenoptera maculata, Balaenoptera nigra, Balaenoptera punctulata, Catodon polycyphus, Catodon polycyphus, Catodon svineval, Cephalorhyncus commersonii, Cephalorhyncus heavisidii, Clymenia qadamu, Delphinapterus senedetta, Delphinorhynchus maculatus, Delphinorhynchus pernettyi, Delphinorhynchus santonicus, Delphinus abusalam, Delphinus anarnacus, Delphinus attenuatus, Delphinus bertini, Delphinus bonnaterrei, Delphinus boryi, Delphinus caerulescens, Delphinus carbonarius, Delphinus coronatus, Delphinus cymodice, Delphinus cymodoce, Delphinus epiodon, Delphinus eurynome, Delphinus fabricii, Delphinus feres, Delphinus gadamu, Delphinus hamatus, Delphinus harlani, Delphinus leucocephalus, Delphinus livittatus, Delphinus maculatus, Delphinus maculiventer, Delphinus minimus, Delphinus nesarnac, Delphinus niger, Delphinus pernettyensis, Delphinus pernetyi, Delphinus perniqer, Delphinus rappii, Delphinus rhinoceros, Delphinus salam, Delphinus siculus, Delphinus symodice, Delphinus walkeri, Epiodon rafinesque, Epiodon urganantus, Eudelphinus tasmaniensis, Globicephala macrorhyncus, Globicephalus fuscus, Globicephalus uneidens, Globiocephalus chinensis, Inia araquaiaensis, Inia boliviensis, Lagenodelphis australis, Lagenodelphis obliquidens, Lagenoelphis hosei, Lagenorhynchus bombifrons, Lagenorhynchus nilssonii, Lagenorhynchus posidonia, Lagenorhynchus superciliosus, Lagenorhyncus acutus, Lagenorhyncus albirostris, Lagenorhyncus australis, Lagenorhyncus cruciqer, Lagenorhyncus obliquidens, Lagenorhyncus obscurus, Mesoplodon hotaula, Mesoplodon lazardii, Monodon spurius, Neophocaena asiaeorientalis, Phocaena posidonia, Physeter gibbosus, Physeter katadon, Physeter krefftii, Physeter polycephus, Physeter polycystus, Physeter pterodon, Platanista indi, Prodelphinus malayanus, Sotalia gadamu, Sotalia maculiventer, Sotalia perniger, Sotalia santonicus, Sousa gadamu, Sousa plumbea, Sousa sahulensis, Steno fuscus, Steno qadamu, Steno malayanus, Steno perniqer, Tursio catalania, Tursio cymodoce, Tursio eurynome, Tursiops australis, Tursiops catalania, Tursiops cymodice, Tursiops dawsoni, Tursiops fergusoni, Tursiops nesarnack