# Longitudinal Analysis - Assignment I

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We will use the following libraries in  ${\bf R}$  to perform our analysis:

We import our data:

```
pksm_data <- read.csv("pksm data.csv")
dim(pksm_data)

## [1] 67 27
head(pksm_data,5)

## county visit_no uniquefarmid uniquecalfid uniqobs hsize parity ageatvst
## 1 0 1 1 1 1 15 3 24</pre>
```

##		county	visit_no	unique	erarmid	uniq	[uecali1d	uniqo	DS .	nsıze	parity	y ageatvst
##	1	0	1		1		1		1	15	;	3 24
##	2	0	2		1		1		2	15	;	81
##	3	0	3		1		1		3	15	;	3 207
##	4	0	4		1		1		4	15	;	3 270
##	5	0	5		1		1		5	15	;	3 332
##		weight a	age1stde	wm_wks	strong	yles	coccidia	breed	fr	eqmilk	fed ag	gewaterfree
##	1	65		24		0	0	0			3	4
##	2	143		24		0	0	0			3	4
##	3	182		24		0	0	0			3	4
##	4	246		24		0	0	0			3	4
##	5	276		24		0	0	0			3	4
##		agefedco	oncwks ag	gefedha	aywks w	eaned	lfecalco	nsist	bcs	socia	ltype	floorraised
##	1		2		2	0	)	1	3.5		0	0
##	2		2		2	0	)	2	3.5		0	0
##	3		2		2	1		1 .	4.0		1	0
##	4		2		2	1		1 .	4.0		1	0
##	5		2		2	1		1	3.5		1	0
##		floorcle	ean bedpi	resent	tether	ed ly	ring_day	intera	cti	on		
##	1		0	0		0	18			0		
##	2		1	1		0	18			1		
##	3		0	1		0	14			1		
##	4		1	1		0	15			1		

Next we perform data Wrangling

1

#### colnames(pksm\_data)

## 5

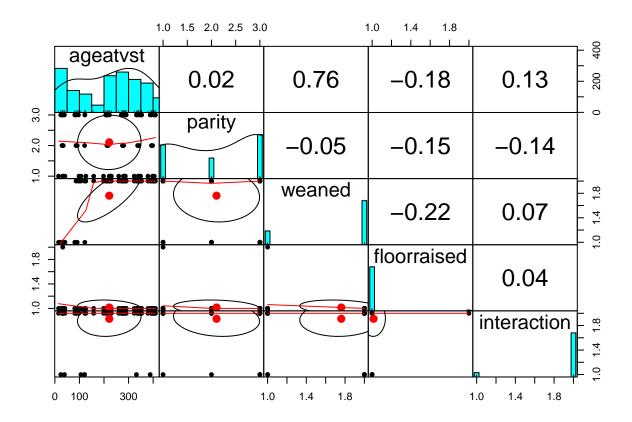
```
"uniquefarmid"
                                                              "uniquecalfid"
    [1] "county"
                          "visit_no"
                          "hsize"
                                            "parity"
                                                              "ageatvst"
##
    [5] "uniqobs"
   [9] "weight"
                          "age1stdewm_wks" "strongyles"
                                                             "coccidia"
## [13] "breed"
                          "freqmilkfed"
                                           "agewaterfree"
                                                             "agefedconcwks"
## [17] "agefedhaywks"
                          "weaned"
                                            "fecalconsist"
                                                             "bcs"
```

14

0

```
## [21] "socialtype"
                        "floorraised"
                                         "floorclean"
                                                          "bedpresent"
## [25] "tethered"
                        "lying_day"
                                         "interaction"
pksm_data1 <- pksm_data %>%
 select(lying_day,strongyles,coccidia,breed,ageatvst,parity,weaned,floorraised,interaction)
dim(pksm_data1)
## [1] 67 9
str(pksm_data1)
## 'data.frame':
                   67 obs. of 9 variables:
## $ lying day : int 18 18 14 15 14 13 18 13 13 14 ...
## $ strongyles : int 0000000000...
## $ coccidia : int 0 0 0 0 0 200 0 100 200 400 ...
## $ breed
                : int 0000000000...
                       24 81 207 270 332 388 32 158 221 283 ...
## $ ageatvst
                : int
## $ parity
                : int 3 3 3 3 3 1 1 1 1 ...
## $ weaned
                : int 0011110111...
## $ floorraised: int 0 0 0 0 0 0 1 0 0 0 ...
## $ interaction: int 0 1 1 1 0 0 1 1 1 1 ...
pksm_data1$weaned <- as.factor(pksm_data1$weaned)</pre>
pksm_data1$interaction <- as.factor(pksm_data1$interaction)</pre>
pksm_data1$breed <- as.factor(pksm_data1$breed)</pre>
pksm_data1$floorraised <- as.factor(pksm_data1$floorraised)</pre>
str(pksm_data1)
## 'data.frame':
                   67 obs. of 9 variables:
## $ lying_day : int 18 18 14 15 14 13 18 13 13 14 ...
## $ strongyles : int 0000000000...
## $ coccidia : int 0 0 0 0 0 200 0 100 200 400 ...
## $ breed
                : Factor w/ 1 level "0": 1 1 1 1 1 1 1 1 1 1 ...
                : int 24 81 207 270 332 388 32 158 221 283 ...
## $ ageatvst
                : int 3 3 3 3 3 1 1 1 1 ...
## $ parity
                : Factor w/ 2 levels "0", "1": 1 1 2 2 2 2 1 2 2 2 ...
## $ floorraised: Factor w/ 2 levels "0","1": 1 1 1 1 1 1 2 1 1 1 ...
## $ interaction: Factor w/ 2 levels "0", "1": 1 2 2 2 1 1 2 2 2 2 ...
pksm_data1 %<>% mutate_if(is.integer,as.numeric)
str(pksm data1)
                   67 obs. of 9 variables:
## 'data.frame':
## $ lying_day : num 18 18 14 15 14 13 18 13 13 14 ...
## $ strongyles : num 0 0 0 0 0 0 0 0 0 ...
## $ coccidia : num 0 0 0 0 0 200 0 100 200 400 ...
                : Factor w/ 1 level "0": 1 1 1 1 1 1 1 1 1 ...
## $ breed
## $ ageatvst
                : num 24 81 207 270 332 388 32 158 221 283 ...
## $ parity
                : num 3 3 3 3 3 3 1 1 1 1 ...
## $ weaned
                : Factor w/ 2 levels "0", "1": 1 1 2 2 2 2 1 2 2 2 ...
## $ floorraised: Factor w/ 2 levels "0","1": 1 1 1 1 1 1 2 1 1 1 ...
## $ interaction: Factor w/ 2 levels "0", "1": 1 2 2 2 1 1 2 2 2 2 ...
Removing the response variables and variables with zero fill
pksm_data2 <- subset(pksm_data1,select = -c(lying_day,strongyles,coccidia,breed))</pre>
```

We generate scatter plots to check for multi-correlation in the data

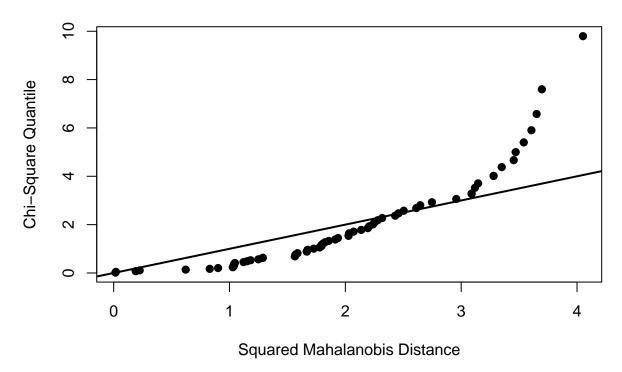


There is a high correlation between *angeatvst* and *weaned* at 0.76. The other predictor variables have statistically insignificant correlation to induce multi-correlation as they have correlations less than 0.7.

We assess for Multivariate Collinearity:

```
pksm_data3 <- subset(pksm_data2,select = -c(weaned,floorraised,interaction))
mvn(pksm_data3,mvnTest = "mardia",scale = T,multivariatePlot = "qq")</pre>
```

# Chi-Square Q-Q Plot



```
## $multivariateNormality
                                                     p value Result
##
                Test
                              Statistic
## 1 Mardia Skewness 0.987858098479116
                                          0.911631475798832
## 2 Mardia Kurtosis -3.14828155517961 0.00164233421818971
                                                                 NO
## 3
                 MVN
                                   <NA>
                                                        <NA>
                                                                 NO
##
   $univariateNormality
##
             Test Variable Statistic
                                         p value Normality
## 1 Shapiro-Wilk ageatvst
                                0.9161
                                          2e-04
                                                     NO
  2 Shapiro-Wilk parity
                                        <0.001
                                                     NO
                                0.7467
##
## $Descriptives
##
             n
                     Mean
                               Std.Dev Median Min Max 25th 75th
## ageatvst 67 221.447761 129.3952465
                                          224
                                                15 409
                                                         97
                                                             338 -0.1801691
                 2.104478
                             0.8899154
                                                               3 -0.2003226
  parity
            67
##
             Kurtosis
## ageatvst -1.374055
## parity
            -1.726317
```

The chi-square Q-Q plot indicates departures from multivariate normal distribution hence the data set is not Multivariate Normal.

We perform a Ridge Regression Using lying\_day as the outcome variable:

```
pksm_data4 <- subset(pksm_data1,select = -c(strongyles,coccidia,breed))
set.seed(222)
ind <- sample(2,nrow(pksm_data4),replace=T,prob = c(0.7,0.3))</pre>
```

```
train <- pksm_data4[ind==1,]</pre>
test <- pksm_data4[ind==2,]</pre>
custom <- trainControl(method = "repeatedcv",</pre>
                          number = 10,
                         repeats=5,
                         verboseIter = F)
```

### The Ridge Model

```
set.seed(1234)
ridge <- train(lying_day~.,data=pksm_data4,</pre>
               method='glmnet',
               tuneGrid=expand.grid(alpha=0,
               lambda=seq(0.0001,1,length=5)),
               trControl=custom)
ridge
## glmnet
##
## 67 samples
## 5 predictor
##
## No pre-processing
## Resampling: Cross-Validated (10 fold, repeated 5 times)
```

```
##
##
    lambda
              RMSE
                       Rsquared
##
    0.000100 1.458879 0.5737612 1.205384
    0.250075 1.460663 0.5742970 1.208074
    0.500050 1.467757 0.5748649 1.213643
##
    0.750025 1.478545 0.5751238 1.219888
##
##
    1.000000 1.491256 0.5752932 1.227049
##
```

## Resampling results across tuning parameters:

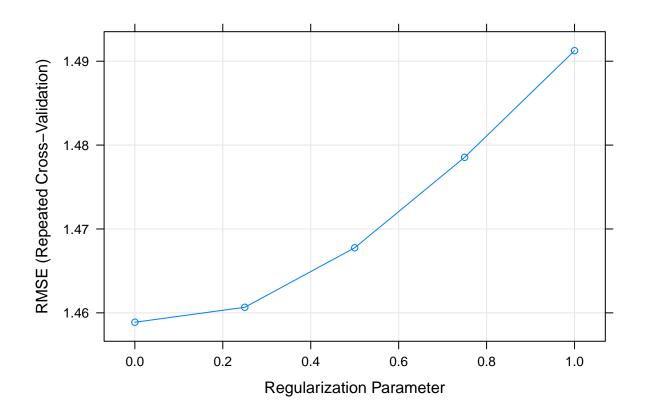
## Summary of sample sizes: 60, 61, 60, 61, 61, 60, ...

## Tuning parameter 'alpha' was held constant at a value of 0

## RMSE was used to select the optimal model using the smallest value. ## The final values used for the model were alpha = 0 and lambda = 1e-04.

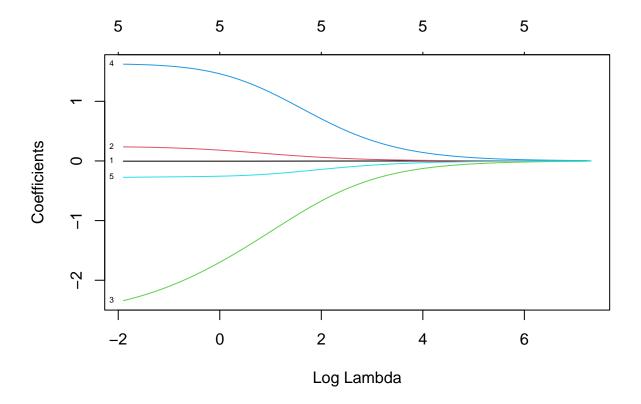
We plot the results and the coefficients of the model:

plot(ridge)



### print(ridge)

```
## glmnet
##
## 67 samples
    5 predictor
##
##
## No pre-processing
## Resampling: Cross-Validated (10 fold, repeated 5 times)
## Summary of sample sizes: 60, 61, 60, 61, 61, 60, ...
  Resampling results across tuning parameters:
##
##
     lambda
               RMSE
                         Rsquared
                                     MAE
##
     0.000100
                         0.5737612
               1.458879
                                     1.205384
##
     0.250075
               1.460663
                         0.5742970
                                     1.208074
##
     0.500050
               1.467757
                         0.5748649
                                     1.213643
##
     0.750025
               1.478545
                         0.5751238
                                     1.219888
##
     1.000000
               1.491256
                         0.5752932
                                     1.227049
## Tuning parameter 'alpha' was held constant at a value of 0
## RMSE was used to select the optimal model using the smallest value.
## The final values used for the model were alpha = 0 and lambda = 1e-04.
plot(ridge$finalModel,xvar = "lambda",label = T)
```

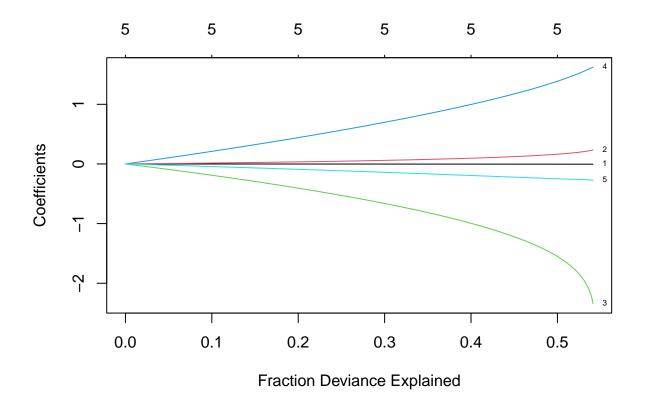


We can observe that for higher values of  $\lambda$  the RMSE increases. The RMSE sould be held lowest (1.458879) at the best value of lambda (1e-04).

For values of  $\lambda$  above 6, all the coefficients are more or less zero. As  $\lambda$  approaches zero, all the coefficients increase in value. At the top of the plot, it shows that we have all the 5 predictor variables in the model. This proves that indeed the Ridge does not shrink coefficients of the less significant variables to zero.

We perform another analysis by making the x variable the 'dev':

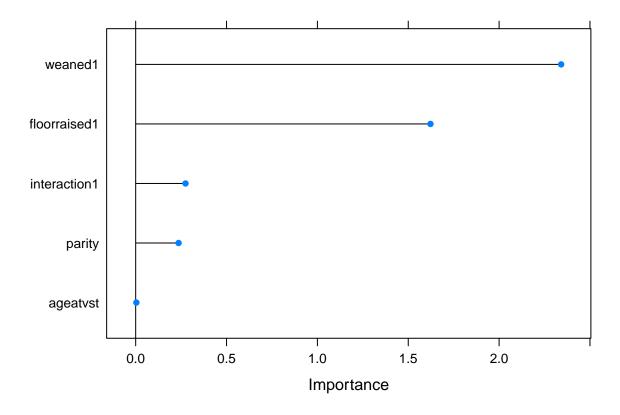
```
plot(ridge$finalModel,xvar = "dev",label = T)
```



This results on the x axis **Fraction Deviance Explained.** Beyond 40% of the deviance (variation), there is a sudden jump and the coefficients become highly inflated and therefore above 0.4 point on the deviance, over-fitting start to take place.

We assess variable importance plot:  $Variable\ Importance\ Plot\ -\ Scale\ =\ False$ 

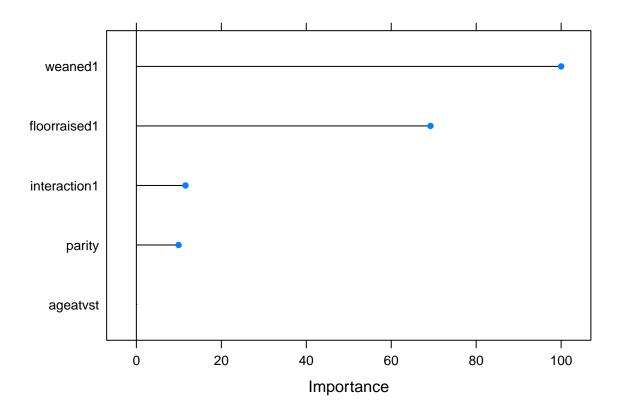
plot(varImp(ridge,scale = F))



we an ed 1 is the most important variable followed by floor aised. while the least ones are at the bottom, with low coefficient estimates.

 ${\it Variable\ Importance\ Plot\ -\ Scale\ =\ True}$ 

plot(varImp(ridge,scale = T))



The scale has changed to be between 0 and 100

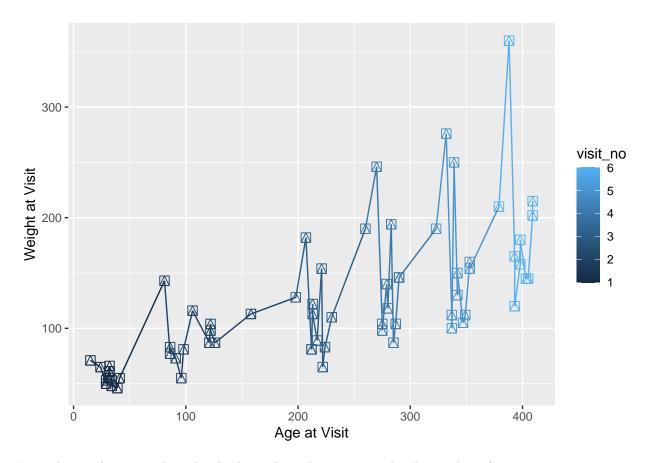
### $The\ OLS\ model$

```
fit <- lm(lying_day~.,data = pksm_data4)
model_parameters(fit)</pre>
```

			Coefficient		•	•		95% CI	١	t(61)	I	p	)	
								_						
##	(Intercept)			16.52		0.87		[14.78,	18.27]		18.93		< .001	
##	ageatvst			-3.42e-03		2.22e-03	-	[-0.01,	0.00]		-1.54		0.128	
##	parity			0.24		0.21	-	[-0.18,	0.67]		1.15		0.254	
##	weaned [1]			-2.59		0.67	-	[-3.93,	-1.25]		-3.86		< .001	
##	${\tt floorraised}$	[1]		1.63		1.56	-	[-1.50,	4.75]		1.04		0.302	
##	interaction	[1]		-0.29		0.65		[-1.59,	1.02]	$\perp$	-0.44		0.663	

The Graph of Weight against Age at Visit given the Number of Visits:

ggplot(data=pksm\_data,aes(x=ageatvst,y=weight,colour=visit\_no))+geom\_point(size=3,shape=14)+geom\_line()



From the graph, it is evident that both weight and age increase by the number of visits.