

ggpicrust2

AUTHOR

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Objective

The purpose of this script is to integrate the annotated KEGG pathways from the KO outputs from picrust2 and then compare differences observed between Type 2 diabetic and control cohorts using DESEQ2 wrapped inside ggpicrust2.

Analysis

1. Data Importation

2. Data Wrangling

3. Data Filtering

1. Data Import

Lets begin by loading our libraries and importing our metadata and KO predictions, then use ggpicrust2 to translate the KO to KEGG pathways

```
library(readr)
```

Warning: package 'readr' was built under R version 4.2.3

```
library(ggpicrust2)  
library(tibble)
```

Warning: package 'tibble' was built under R version 4.2.3

```
library(tidyverse)
```

Warning: package 'tidyverse' was built under R version 4.2.3

Warning: package 'ggplot2' was built under R version 4.2.3

Warning: package 'tidyr' was built under R version 4.2.3

Warning: package 'purrr' was built under R version 4.2.3

Warning: package 'dplyr' was built under R version 4.2.3

Warning: package 'stringr' was built under R version 4.2.3

Warning: package 'forcats' was built under R version 4.2.3

Warning: package 'lubridate' was built under R version 4.2.3

— Attaching core tidyverse packages — tidyverse 2.0.0 —

✓ dplyr 1.1.2 ✓ purrr 1.0.1
 ✓ forcats 1.0.0 ✓ stringr 1.5.0
 ✓ ggplot2 3.4.2 ✓ tidyr 1.3.0
 ✓ lubridate 1.9.2

— Conflicts — tidyverse_conflicts() —

✗ dplyr::filter() masks stats::filter()

✗ dplyr::lag() masks stats::lag()

ℹ Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors

```
library(ggprism)
```

Warning: package 'ggprism' was built under R version 4.2.3

```
library(patchwork)
library(mosaic)
```

Registered S3 method overwritten by 'mosaic':

| | |
|----------------------------------|---------|
| method | from |
| fortify.SpatialPolygonsDataFrame | ggplot2 |

The 'mosaic' package masks several functions from core packages in order to add additional features. The original behavior of these functions should not be affected by this.

Attaching package: 'mosaic'

The following object is masked from 'package:Matrix':

mean

The following objects are masked from 'package:dplyr':

count, do, tally

The following object is masked from 'package:purrr':

cross

The following object is masked from 'package:ggplot2':

stat

The following objects are masked from 'package:stats':

```
binom.test, cor, cor.test, cov, fivenum, IQR, median, prop.test,
quantile, sd, t.test, var
```

The following objects are masked from 'package:base':

```
max, mean, min, prod, range, sample, sum
```

```
library(ggh4x)
```

Warning: package 'ggh4x' was built under R version 4.2.3

```
getwd()
```

```
[1] "K:/github/lmatz"
```

```
metadata <- read_delim("IRB_Human_Metadata.txt", delim = "\t", escape_double = FALSE, trim_ws = T
```

Rows: 12 Columns: 5

— Column specification —————

Delimiter: "\t"

chr (3): SampleID, Diabetes_Status, Sex

dbl (2): BMI, Age

• Use `spec()` to retrieve the full column specification for this data.

• Specify the column types or set `show_col_types = FALSE` to quiet this message.

```
metadata
```

A tibble: 12 × 5

| | SampleID | Diabetes_Status | Sex | BMI | Age |
|----|--------------------|-----------------|--------|-------|-------|
| | <chr> | <chr> | <chr> | <dbl> | <dbl> |
| 1 | Buffington_201_001 | T2D | Male | 31.8 | 57 |
| 2 | Buffington_201_002 | T2D | Male | 39.5 | 59 |
| 3 | Buffington_201_003 | HC | Female | 25.2 | 50 |
| 4 | Buffington_201_005 | HC | Female | 34.5 | 53 |
| 5 | Buffington_201_006 | T2D | Female | 28.7 | 62 |
| 6 | Buffington_201_007 | HC | Male | 25.8 | 52 |
| 7 | Buffington_201_008 | HC | Female | 35.8 | 54 |
| 8 | Buffington_201_009 | HC | Female | 21.1 | 56 |
| 9 | Buffington_201_010 | T2D | Female | 33.5 | 64 |
| 10 | Buffington_201_011 | HC | Female | 25 | 59 |
| 11 | Buffington_201_013 | T2D | Male | 45.4 | 67 |
| 12 | Buffington_201_014 | T2D | Female | 28.5 | 55 |

```
# Load KEGG pathway abundance
```

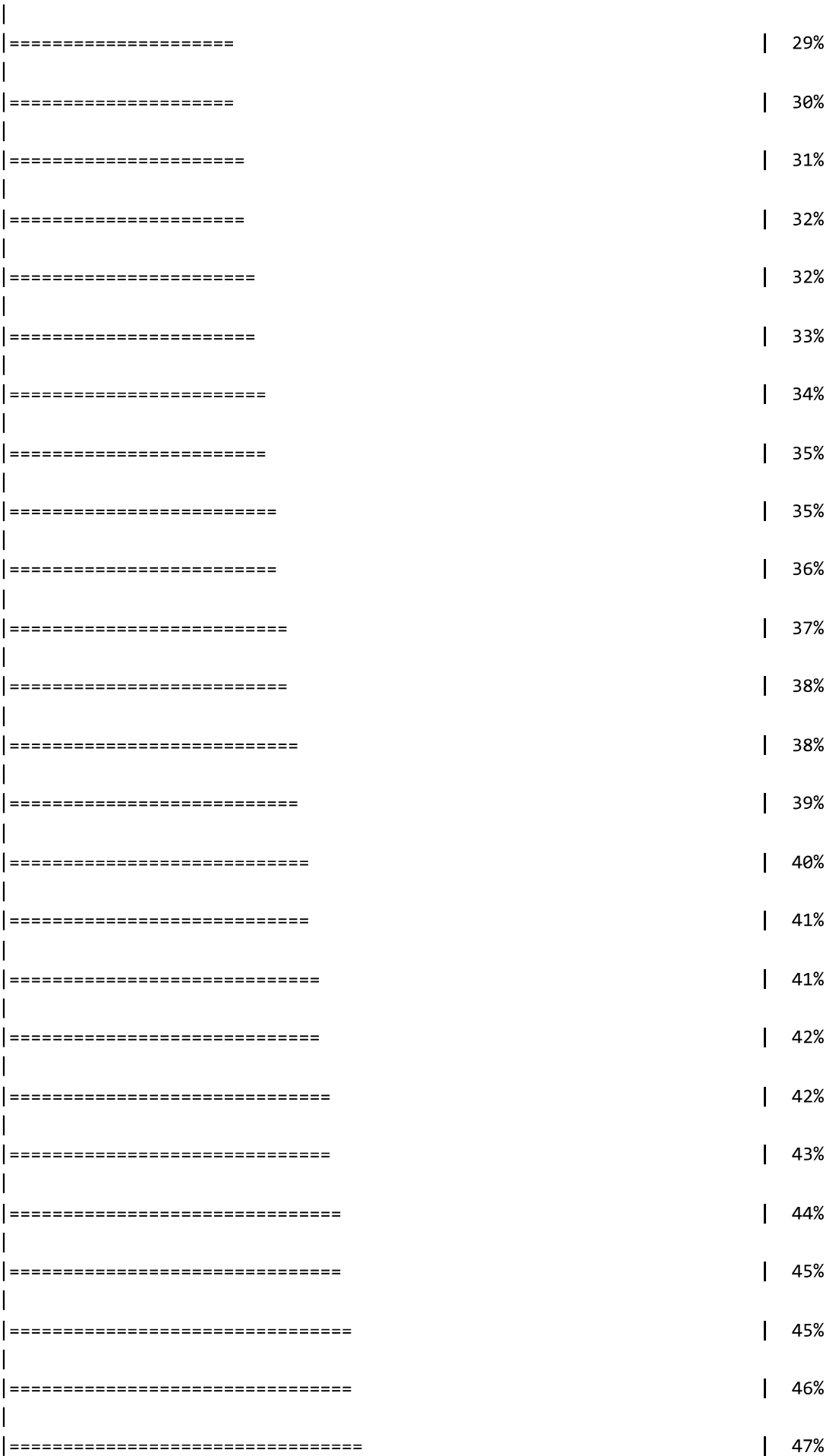
```
kegg_abundance <- ko2kegg_abundance("picrust2/KO_metagenome_out/pred_metagenome_unstrat.tsv")
```

Loading data from file...
Rows: 5565 Columns: 13— Column specification

Delimiter: "\t"
chr (1): function
dbl (12): Buffington_201_001, Buffington_201_002, Buffington_201_003, Buffin...
i Use `spec()` to retrieve the full column specification for this data.
i Specify the column types or set `show_col_types = FALSE` to quiet this message.Loading KEGG
reference data. This might take a while...
Performing KO to KEGG conversion. Please be patient, this might take a while...

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```

KO to KEGG conversion completed. Time elapsed: 16.24 seconds.

Removing KEGG pathways with zero abundance across all samples...

KEGG abundance calculation completed successfully.

differential pathway analysis

Using the kegg_abundance and metadata, run DESeq2 to obtain the sig. KO assoc. with Type 2 diabetes

```

deseq2_daa_results_df<- pathway_daa(abundance = kegg_abundance,
                                     metadata = metadata,
                                     group = "Diabetes_Status",
                                     p.adjust ="BH",
                                     daa_method = "DESeq2",
                                     select = NULL,
                                     reference = "HC")

```

Sample names extracted.

Identifying matching columns in metadata...

Matching columns identified: SampleID . This is important for ensuring data consistency.

Using all columns in abundance.

Converting abundance to a matrix...

Reordering metadata...

Converting metadata to a matrix and data frame...

Extracting group information...

Running DESeq2. Note: DESeq2 is only suitable for comparison between two groups.

Performing pairwise comparisons with DESeq2...

converting counts to integer mode

using pre-existing size factors

estimating dispersions

gene-wise dispersion estimates

mean-dispersion relationship

final dispersion estimates

fitting model and testing

Compiling DESeq2 results...

DESeq2 analysis complete.

```
deseq2_daa_results_df<-deseq2_daa_results_df%>%filter(!is.na(p_values)) #14
```

```
deseq2_daa_results_df%>%filter(p_values<0.05) #14
```

| | feature | method | group1 | group2 | p_values | adj_method | p_adjust |
|----|---------|--------|--------|--------|----------------------|------------|-------------|
| 1 | ko00311 | DESeq2 | T2D | HC | 0.0154634285344572 | BH | 0.243846373 |
| 2 | ko05215 | DESeq2 | T2D | HC | 0.00932532551594288 | BH | 0.173790157 |
| 3 | ko05211 | DESeq2 | T2D | HC | 0.0103302717505706 | BH | 0.176475476 |
| 4 | ko05130 | DESeq2 | T2D | HC | 0.00708270225776833 | BH | 0.173790157 |
| 5 | ko04914 | DESeq2 | T2D | HC | 0.00932532551594288 | BH | 0.173790157 |
| 6 | ko05143 | DESeq2 | T2D | HC | 0.0386912989347241 | BH | 0.520024327 |
| 7 | ko00400 | DESeq2 | T2D | HC | 0.0405872645834935 | BH | 0.520024327 |
| 8 | ko05131 | DESeq2 | T2D | HC | 0.00194346578891148 | BH | 0.079682097 |
| 9 | ko00340 | DESeq2 | T2D | HC | 0.00451037851803533 | BH | 0.154104599 |
| 10 | ko04612 | DESeq2 | T2D | HC | 0.00932532551594288 | BH | 0.173790157 |
| 11 | ko04621 | DESeq2 | T2D | HC | 0.00801484379440754 | BH | 0.173790157 |
| 12 | ko05014 | DESeq2 | T2D | HC | 0.0491542850008098 | BH | 0.555480616 |
| 13 | ko05100 | DESeq2 | T2D | HC | 0.000143460544632947 | BH | 0.007352353 |
| 14 | ko04144 | DESeq2 | T2D | HC | 0.032209896154432 | BH | 0.471644908 |

```
deseq2_daa_results_df%>%filter(p_adjust<0.05) #4
```

| | feature | method | group1 | group2 | p_values | adj_method | p_adjust |
|---|---------|--------|--------|--------|----------------------|------------|--------------|
| 1 | ko00592 | DESeq2 | T2D | HC | 9.66578157555687e-05 | BH | 0.0066049507 |
| 2 | ko00624 | DESeq2 | T2D | HC | 3.33156296678632e-05 | BH | 0.0034148520 |
| 3 | ko05150 | DESeq2 | T2D | HC | 5.88462099103448e-07 | BH | 0.0001206347 |
| 4 | ko05100 | DESeq2 | T2D | HC | 0.000143460544632947 | BH | 0.0073523529 |

Annotate the significant KO that have annotations

```
# Annotate pathway results using KO to KEGG conversion
```

```
deseq2_daa_annotated_sub_method_results_df <- pathway_annotation(pathway = "KO", daa_results_df =
```

Starting pathway annotation...

DAA results data frame is not null. Proceeding...

KO to KEGG is set to TRUE. Proceeding with KEGG pathway annotations...

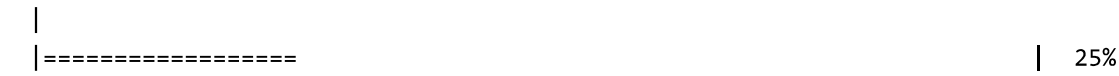
We are connecting to the KEGG database to get the latest results, please wait patiently.

Processing pathways individually...



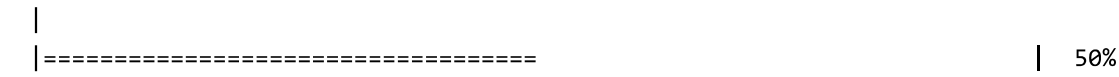
Beginning annotation for pathway 1 of 4...

Annotated pathway 1 of 4. Time taken: 1.79 seconds.



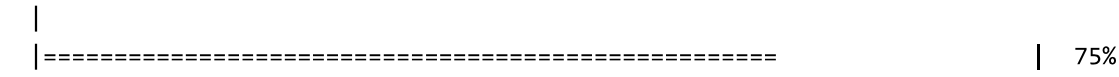
Beginning annotation for pathway 2 of 4...

Annotated pathway 2 of 4. Time taken: 0.23 seconds.



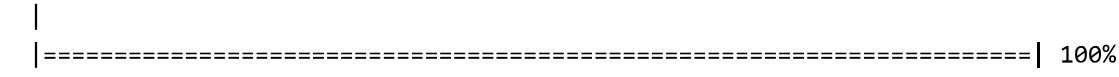
Beginning annotation for pathway 3 of 4...

Annotated pathway 3 of 4. Time taken: 0.24 seconds.



Beginning annotation for pathway 4 of 4...

Annotated pathway 4 of 4. Time taken: 0.2 seconds.



Pathway annotation completed.

Returning DAA results filtered annotation data frame...

```
deseq2_daa_annotated_sub_method_results_df
```

| | feature | method | group1 | group2 | p_values | adj_method | p_adjust |
|----|---------|--------|--------|--------|----------------------|------------|--------------|
| 45 | ko00592 | DESeq2 | T2D | HC | 9.66578157555687e-05 | BH | 0.0066049507 |

| | | | | | | | |
|-----|---------|--------|-----|----|----------------------|----|--------------|
| 58 | ko00624 | DESeq2 | T2D | HC | 3.33156296678632e-05 | BH | 0.0034148520 |
| 140 | ko05150 | DESeq2 | T2D | HC | 5.88462099103448e-07 | BH | 0.0001206347 |
| 175 | ko05100 | DESeq2 | T2D | HC | 0.000143460544632947 | BH | 0.0073523529 |

pathway_name

| | |
|-----|---|
| 45 | alpha-Linolenic acid metabolism |
| 58 | Polycyclic aromatic hydrocarbon degradation |
| 140 | Staphylococcus aureus infection |
| 175 | Bacterial invasion of epithelial cells |

pathway_description

45

<NA>

58

<NA>

140

Staphylococcus aureus can cause multiple forms of infections ranging from superficial skin infections to food poisoning and life-threatening infections. The organism has several ways to divert the effectiveness of the immune system: secreting immune modulating proteins that inhibit complement activation and neutrophil chemotaxis or lysis, modulating the sensitivity to cationic antimicrobial peptides (such as defensin) by increasing the positive net charge of its cytoplasmic membrane, and expression of superantigens that prevent development of a normal immune response or cause an emetic response when ingested.

175 Many pathogenic bacteria can invade phagocytic and non-phagocytic cells and colonize them intracellularly, then become disseminated to other cells. Invasive bacteria induce their own uptake by non-phagocytic host cells (e.g. epithelial cells) using two mechanisms referred to as zipper model and trigger model. Listeria, Staphylococcus, Streptococcus, and Yersinia are examples of bacteria that enter using the zipper model. These bacteria express proteins on their surfaces that interact with cellular receptors, initiating signalling cascades that result in close apposition of the cellular membrane around the entering bacteria. Shigella and Salmonella are the examples of bacteria entering cells using the trigger model. These bacteria use type III secretion systems to inject protein effectors that interact with the actin cytoskeleton.

pathway_class

| | |
|-----|---|
| 45 | Metabolism; Lipid metabolism |
| 58 | Metabolism; Xenobiotics biodegradation and metabolism |
| 140 | Human Diseases; Infectious disease: bacterial |
| 175 | Human Diseases; Infectious disease: bacterial |

pathway_map

| | |
|-----|---|
| 45 | alpha-Linolenic acid metabolism |
| 58 | Polycyclic aromatic hydrocarbon degradation |
| 140 | Staphylococcus aureus infection |
| 175 | Bacterial invasion of epithelial cells |

Generate the pathway error plot

```
# Generate pathway error bar plot
# Please change Group to metadata$your_group_column if you are not using example dataset
p <- pathway_errorbar(abundance = kegg_abundance,
                      daa_results_df = deseq2_daa_annotated_sub_method_results_df,
                      Group = metadata$Diabetes_Status,
                      p_values_threshold = 0.25,
                      order = "pathway_class",
```

```
select = NULL,  
ko_to_kegg = T,  
p_value_bar = TRUE,  
colors = NULL,  
x_lab = "pathway_name")
```

Registered S3 method overwritten by 'GGally':
method from
+.gg ggplot2

p

