

Cambridge International Examinations

Cambridge International Advanced Subsidiary and Advanced Level

CANDIDATE NAME				
CENTRE NUMBER		CANDIDATE NUMBER		
BIOLOGY Paper 5 Planning, Analysis and E	-valuation	Oct	tober/Nove	0/52
		36.	1 hour	
Candidates answer on the Quest	ion Paper.			

READ THESE INSTRUCTIONS FIRST

No Additional Materials are required.

Write your Centre number, candidate number and name on all the work you hand in.

Write in dark blue or black pen.

You may use an HB pencil for any diagrams or graphs.

Do not use staples, paper clips, glue or correction fluid.

DO NOT WRITE IN ANY BARCODES.

Answer all questions.

Electronic calculators may be used.

You may lose marks if you do not show your working or if you do not use appropriate units.

At the end of the examination, fasten all your work securely together.

The number of marks is given in brackets [] at the end of each question or part question.



1 (a) The opening and closing of stomata involves the movement of potassium ions into and out of guard cells. Opening and closing of stomata is influenced by a number of environmental factors, for example light and temperature.

A student investigated the effect of potassium chloride (KC1) on the opening of stomata.

The student was provided with:

- 500 cm³ of 250 mmol dm⁻³ KC*l* solution
- freshly picked leaves from a plant that had been kept in the dark and a high concentration of carbon dioxide for an hour. This ensured that all the stomata were closed.

Strips of leaf tissue were obtained by cutting a leaf into sections as shown in Fig. 1.1.

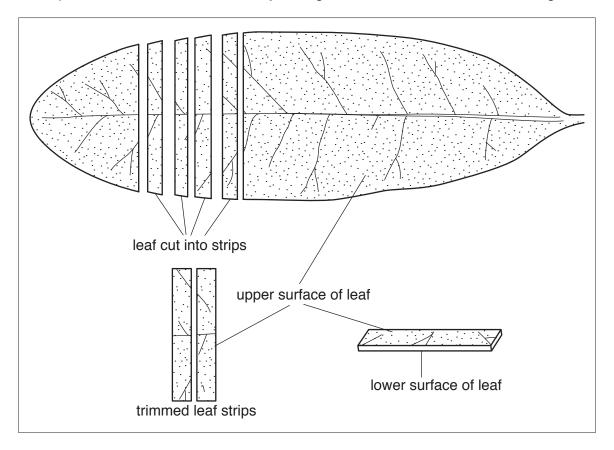


Fig. 1.1

The student floated three strips of leaf tissue in each of a range of buffered potassium chloride solutions for 2 hours and then recorded the number of open stomata.

(i)	Identify the independent and dependent variables in this investigation.
	independent variable
	dependent variable
	[2]
(ii)	The student used the $250\mathrm{mmoldm^{-3}}$ KC l solution to make $100\mathrm{cm^3}$ of four other concentrations by reducing the concentration by $50\mathrm{mmoldm^{-3}}$ each time.
	Describe a procedure that the student could use to prepare these four concentrations.
	[3]
(b) (i)	Suggest a hypothesis that the student could test about the effect of KCl on the opening and closing of stomata.
	[1]

(11)	concentrations of KCl on the opening of stomata.
	The description of your method should be detailed enough for another person to follow and should not repeat the details from (a)(ii) of how to dilute the 250 mmol dm ⁻³ solution of KC <i>l</i> .

(c) The student also tested the hypothesis:

The more light the wider the stomata open.

- Eight leaves from young plants that had been kept in the dark for 24 hours were covered by metal foil.
- A fluorescent lamp of fixed intensity was placed 10 cm from the plant. The metal foil was removed from the leaves.
- Two leaves were removed at the start of the experiment and three epidermal strips were made from each leaf. An epidermal strip is made by peeling the epidermis from a leaf as a single layer.
- The diameter of the stomatal aperture of five of the stomata with the widest aperture on each strip was measured.
- · At one hour intervals two more leaves were removed and the same procedure repeated.

Fig. 1.2 shows stomata at different stages of opening.

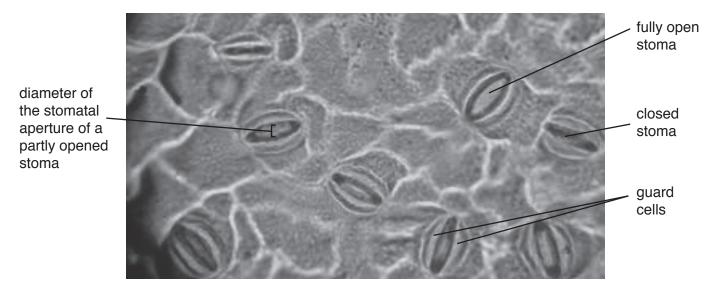


Fig. 1.2

Outline how the student could find the actual diameter of a stomatal aperture.
[2]

Table 1.1 shows the results of the student's experiment.

Table 1.1

time / min					diar	neter	of sto	mata	l aper	ture /	μm				
0 (control)	0.5	0.1	0.2	0.3	0.4	0.1	0.5	0.2	0.3	0.3	0.1	0.2	0.2	0.2	0.4
60	0.9	1.1	1.0	1.3	1.2	1.8	1.5	0.8	0.2	1.3	1.1	0.8	1.0	1.9	0.9
120	1.9	2.4	2.6	2.6	2.5	2.2	2.8	2.4	2.4	3.9	2.6	2.3	2.5	2.2	2.7
180	4.1	4.8	4.2	4.0	5.7	4.7	3.9	4.1	5.5	4.5	4.3	4.0	3.1	4.1	4.3

(ii) On Table 1.1, draw circles around two values that are anomalous.

[1]

(iii) The student calculated the mean diameter of the stomatal apertures and the rate at which the diameter of the stomatal apertures increased.

Table 1.2 shows some of these calculations.

Table 1.2

time/min	mean diameter of stomatal apertures /μm	rate of increase of diameter of the stomatal apertures /μm min ⁻¹
0	0.3	
60	1.2	0.015
120	2.5	0.022
180	4.6	

Complete Table 1.2 by calculating the rate of increase of the diameter of the stomatal apertures between 120 minutes and 180 minutes.

Space for working

[1]

	(iv)	The experimental procedure described in (c) could be criticised for poor technique in obtaining results.
		Suggest how the procedure could be modified to improve the quality of these results.
		[3]
(d)	The	experimental procedure used in (c) is not completely valid for the stated hypothesis:
		The more light the wider the stomata open.
	Sug	gest how this hypothesis could be modified to match the procedure described in (c).
		[1]
		[Total: 19]

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2 Cereal crops are often sprayed with selective herbicides which can reduce the population of the local wildlife. One method of helping to conserve wildlife is to leave a 6 m strip, called a headland, around fields where cereal crops are grown. The headland is not sprayed with any herbicides.

An investigation was carried out into the effect on the butterfly populations of leaving headlands unsprayed.

- Two groups of 20 fields growing the same cereal crop were studied.
- The headlands of one group of 20 fields were left unsprayed by herbicide.
- The headlands of the other group of 20 fields were sprayed with herbicide.
- The total number of each species of butterfly was counted in each group of 20 fields.
- A chi-squared (χ^2) test was used to find out if the differences in the butterfly populations were significant.

(a)	Stat	ate two variables that were standardised in this investigation.					
	1						
	2		[1]				
(b)	(i)	State a reason why the chi-squared (χ^2) populations.	test is suitable to use for comparing the butterfly				
			[1]				
	(ii)	State a null hypothesis for the chi-square	red test for this investigation.				
			[1]				
	(iii)	Table 2.1 shows the results for one of the	ne species of butterfly, species Q.				
		Complete Table 2.1 to calculate the value	ue of χ^2 using the equation below.				
		$\chi^2 = \sum \frac{(O - E)^2}{E}$	O = observed result E = expected result v = degrees of freedom				
		v = c - 1	c = number of classes				

Table 2.1

species Q	0	E	$(O - E)^2$	$\frac{(O-E)^2}{E}$
number on headland sprayed with herbicide	3			
number on headland not sprayed with herbicide	37			

$$\chi^2 =$$
[3]

Table 2.2 shows some chi-squared (χ^2) values.

Table 2.2

dograps of freedom	p <						
degrees of freedom	0.10	0.05	0.01	0.001			
1	2.71	3.84	6.64	10.83			
2	4.61	5.99	9.21	13.82			
3	6.25	7.82	11.34	16.27			

Table 2.3 shows the results of the investigation and the significance of the results from chi-squared tests for the other species of butterflies counted.

Table 2.3

butterfly species	number of each species on headland sprayed with herbicide	number of each species on headland not sprayed with herbicide	significance of chi-squared test
R	1	17	p< 0.001
S	38	56	not significant
Т	13	29	p< 0.05
U	59	93	p< 0.05
V	0	11	p< 0.01
W	23	52	p< 0.01

c)	With reference to Table 2.3 and your calculation for species Q , state three conclusions that can be drawn from these results about the effect of herbicides on the species of butterfly studied.
	[3]
	[Total: 11]

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