Name

UNIVERSITY OF CAMBRIDGE INTERNATIONAL EXAMINATIONS General Certificate of Education Advanced Level

BIOLOGY 9700/06

Paper 6 Options

October/November 2004

1 hour

Candidates answer on the Question Paper. No Additional Materials are required.

READ THESE INSTRUCTIONS FIRST

Write your Centre Number, Candidate Number and Name in the spaces at the top of this page. Write in dark blue or black pen in the spaces provided on the Question Paper. You may use a soft pencil for any diagrams, graphs or rough working. Do not use staples, paper clips, highlighters, glue or correction fluid.

Answer all the questions set on one of the options.

At the end of the examination, enter the number of the option you have answered in the grid below.

INFORMATION FOR CANDIDATES

The number of marks is given in brackets [] at the end of each question or part question. The options are:

- 1 Mammalian Physiology (page 2)
- 2 Microbiology and Biotechnology (page 11)
- 3 Growth, Development and Reproduction (page 20)
- 4 Applications of Genetics (page 28)

If you have been given a label, look at the details. If any details are incorrect or missing, please fill in your correct details in the space given at the top of this page.

Stick your personal label here, if provided.

| [| | | | |
|--------------------|------|--|--|--|
| OPTION ANSW | EKED | | | |
| FOR EXAMINER'S USE | | | | |
| 1 | | | | |
| 2 | | | | |
| 3 | | | | |
| 4 | | | | |
| TOTAL | | | | |

This document consists of **35** printed pages and **1** blank page.



OPTION 1 – MAMMALIAN PHYSIOLOGY

1 (a) Fig. 1.1 shows part of the retina in a mammalian eye.

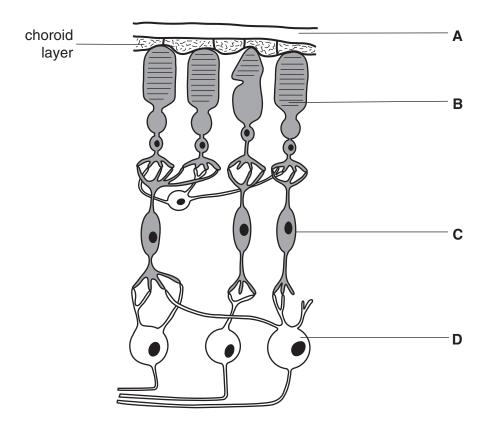


Fig. 1.1

| (i) | Name A to D. | |
|------|---|------|
| | A | |
| | В | |
| | C | |
| | D | [2] |
| (ii) | Describe the function of the choroid layer. | |
| | | |
| | | |
| | | .[2] |
| | | |

| With reference to Fig. 1.1, explain why visual acuity is greater when light is detected by cone cells than when it is detected by rod cells. |
|--|
| |
| |
| [3] |

(b) Rod cells contain the light-sensitive pigment rhodopsin. There are three different types of cone cells each containing a different pigment, known as B (blue), R (red) and G (green).

Fig. 1.2 shows the absorption spectra of these four pigments.

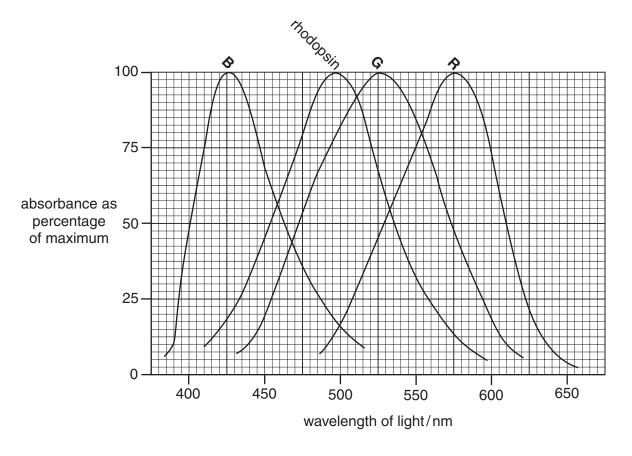


Fig. 1.2

With reference to Fig. 1.2,

| (i) | explain how the brain distinguishes red light from blue light; |
|--------------|--|
| | |
| | |
| <i>(</i> 11) | [2] |
| (11) | explain why colours cannot be seen when only rod cells are in use. |
| | |
| | [0] |
| | [2] |

| (c) | All four pigment molecules of the rod and cone cells are composed of a protein called |
|-----|--|
| | opsin and the light-sensitive compound retinal. The genes for the pigment molecules in |
| | R cones and in G cones are on the X chromosome. They are very close to each other |
| | and very similar to each other. Mutations affecting these genes are relatively common. |

One such mutation, resulting from loss of part of the chromosome, results in the two genes becoming one 'hybrid' gene, which codes for the production of a single type of pigment. This results in red-green colour blindness.

| (i) | Explain why red-green colour blindness is more common in men than in women. |
|------|---|
| | |
| | |
| | |
| | |
| | [3] |
| (ii) | Suggest which part of the $\bf R$ and $\bf G$ pigment molecules is coded for by the genes on the X chromosome, giving a reason for your answer. |
| | |
| | [1] |
| | [Total : 15] |

2 Fig. 2.1 shows the arrangement of two of the muscles in the upper arm. Fig. 2.2 is a diagrammatic representation of part of a myofibril from the biceps muscle when this muscle is relaxed.

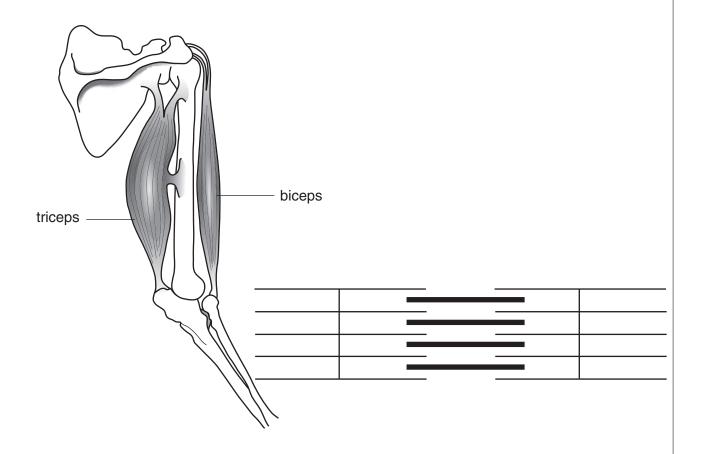


Fig. 2.1 Fig. 2.2

(a) In the space below, draw the part of the myofibril shown in Fig. 2.2 as it would appear when it is fully contracted.

| (b) | Describe the role of myosin in producing the changes between the relaxed myofibril in Fig. 2.2 and the contracted myofibril in your diagram. |
|-----|--|
| | |
| | |
| | |
| | [4] |
| (c) | With reference to Fig. 2.1, explain how the contracted myofibril of the biceps could be returned to the relaxed state shown in Fig. 2.2. |
| | |
| | [2] |
| | |
| | [Total : 9] |

3 Bile salts (bile acids) are produced in the liver and secreted in bile. They pass along the bile duct and into the duodenum, and are mostly reabsorbed as they pass through the ileum. Some, however, are converted by bacteria in the small intestine into substances which cannot be absorbed. These are lost in the faeces.

Fig. 3.1 shows the quantities of bile salts passing along these routes each day.

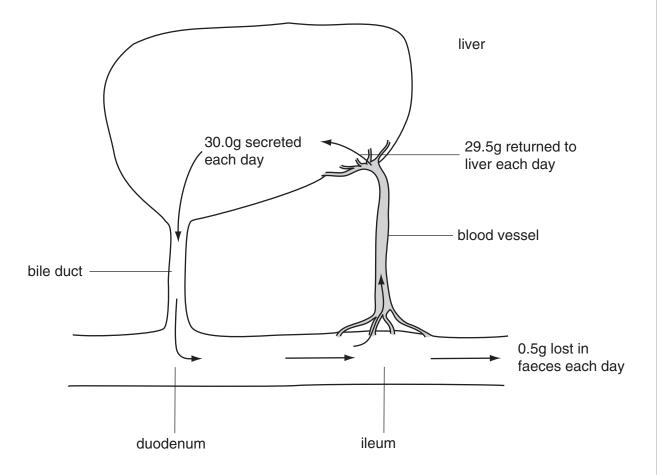


Fig. 3.1

(a) Calculate the percentage of the bile salts that are lost from the body each day. Show your working.

| | [2] |
|-----|---|
| (b) | Name the blood vessel in which the reabsorbed bile salts are returned to the liver. |
| | [1] |
| (c) | Describe how and where bile salts are produced in the liver. |
| | |
| | |
| | [0] |

| ble of bile salts in fat digestion. | (d) |
|-------------------------------------|-----|
| | |
| | |
| | |
| | |
| | |
| [3] | |
| [Total : 8] | |

[Total: 8]

4 (a) Fig. 4.1 shows a gymnast performing on the pommel horse.



Fig. 4.1

Outline the roles of the gymnast's cerebellum and medulla oblongata in enabling him to

carry out this activity.

cerebellum

medulla oblongata

[4]

(b) Alzheimer's disease is a type of dementia characterised by changes in the appearance of the tissue in part of the brain.

(i) Name the part of the brain that is affected in Alzheimer's disease.

[1]

(ii) Describe the possible causes of Alzheimer's disease.

OPTION 2 – MICROBIOLOGY AND BIOTECHNOLOGY

| | OF HON 2 - WICHOBIOLOGI AND DIOTECTINOLOGI | | | | | 11 | | |
|---|--|---|-----------|--|---|-----------------------|---------------------|---|
| 1 | (a) | The number of bacteriophages in a liquid medium can be determined by serial dilution of the medium. A small volume of each dilution, 5 mm³, is plated on to agar plates that have a lawn of bacteria growing on them. When the agar plates are incubated at 25°C there are clear areas, known as plaques, in the bacterial lawn. When one or more bacteriophages infect a bacterium, each infected bacterial cell lyses, releasing many bacteriophages. The released bacteriophages then infect the surrounding bacteria, which also lyse to leave plaques. Explain why the agar plates are incubated at 25°C rather than 37°C. [2] To determine the number of bacterophages in a medium, three serial dilutions were made. Nine plates were set up with bacteria lawns. These were used to make three | | | | | | |
| | | repl | icate pla | tes, A , B and | C, for each of | f the serial dil | | ncubation, the number |
| | | ot p | laques II | n each plate is | s shown in Ta | ble 1.1. | | |
| | | | | | Table | e 1.1. | | |
| | | | | | number of | plaques in ea | nch plate | |
| | | | | dilution | replicate A | replicate B | replicate C | |
| | | | | 1.0 × 10 ⁻⁶ | 657 | 616 | 620 | |
| | | | | 1.0×10^{-7} | 68 | 64 | 66 | |
| | | | | 1.0 × 10 ⁻⁸ | 4 | 8 | 7 | |
| | | | | 1.0 × 10 | 4 | | <i>I</i> | |
| | | (i) | dilution | s of 1.0x10 ⁻⁶ d to estimate | and 1.0x10 ⁻⁸ bacteriophage | may be inace numbers. | curate or unre | aques counted at the eliable and should not |
| | | (ii) | | te the numben. Show your | | ophage partic | les per mm ³ | in the original liquid |

(c) Legionnaires' disease is caused by the bacterium *Legionella pneumophila*. Outbreaks of legionnaires' disease have occurred after people have breathed mists that come from a water source. The disease is not passed from person to person. It affects the lungs.

An outbreak of legionnaires' disease occurred in Cumbria, UK, in the summer of 2002. The cumulative total number of deaths is shown in Fig. 1.1.

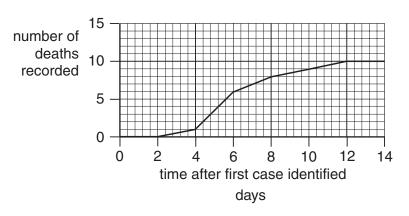


Fig. 1.1

| | | scribe the pattern in the number of deaths during the outbreak of legionnaires' ease shown in Fig. 1.1. |
|-----|------|---|
| | | |
| | | |
| | | F03 |
| | | [2] |
| (d) | (i) | Legionnaires' disease is not an infectious disease. |
| | | Explain what is meant by an infectious disease. |
| | | |
| | | |
| | | [2] |
| | (ii) | Describe the specialist structural features of laboratories working with pathogenic microorganisms that prevent the spread of infectious diseases. |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | [4] |
| | | [Total : 15] |

Question 2 begins on the next page.

2 (a) Table 2.1 shows the distinguishing characteristics of the three groups of organisms collectively known as microorganisms. Complete the table by filling in the empty boxes.

Table 2.1

| group of organisms | type of genetic material | presence or absence of nucleus | type of cell wall material |
|--------------------|-----------------------------|-----------------------------------|-------------------------------|
| Fungi | | present | |
| | DNA or RNA | | no cell wall |
| | DNA | | peptidoglycan |

| r | 21 |
|----|----|
| I١ | " |
| | |

| (b) | List | two ways in which the life cycle of a bacteriophage differs from that of a bacterium | |
|-----|------|--|---|
| | 1. | | |
| | 2. | | 2 |

Fig. 2.1

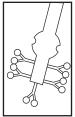
plasma membrane

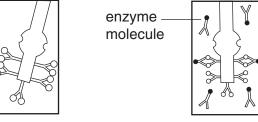
Draw and label the cell wall of a **Gram negative** bacterium in the space below.

[2]

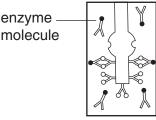
[Total: 7]

| (a) | Outline one method for the production of monoclonal antibodies. | | | | | | | | | |
|------|---|-------------------------------------|---|---|----------|------------|---|-----------------------|------------------------|-----------------|
| | | | | | | | | | | |
| | | | | | | | | | | |
| | | | | | | | | | | |
| | | | | | | | | | | |
| (b) | Monoclonal the monoclo | | | | | | | eagents. | In some (| case |
| | Suggest the | advanta | ge of co | upling the | e monocl | onal antil | body to a | a | | |
| | fluorescent r | molecule | for diag | nosis, | | | | | | |
| | | | • | • | | | • | | | |
| | | | | | | | | | | |
| | radioactive r | nolecule | for treat | ment. | | | | | | |
| | | | | | | | | | | |
| | | | | | | | | | | [3 |
| (c) | During pregiplacenta. It at this hormone monoclonal a plastic dipersonal control of the control | accumula e is the b antibodie | ates in the | ne blood some hor | stream a | ind is rel | eased in s in whicl | the uring h HCG bi | e. Detect nds to sp | ion o ecific |
| | Complete th letter of each you. | | • | | _ | | • | | | _ |
| sec | quence | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | |
| lett | er of stage | | | Α | | В | | | | 1 |
| | - | | | | | | | | |] [3 |

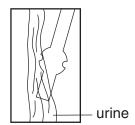




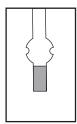
A HCG has bound to antibody on stick. All other antigens washed away.



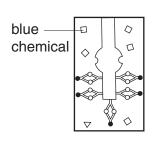
B Second antibody binds to first.



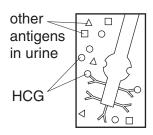
C Stick held in flowing urine.



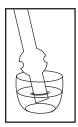
D Stick appears blue - a positive pregnancy test.



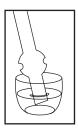
E Chemical turns blue with specific enzyme.



F HCG in urine of pregnant female binds to antibody on surface of stick.



G Stick dipped into second antibody with enzyme attached.



H Stick dipped into chemical.

Fig. 3.1

[Total: 10]

| 4 | (a) | Penicillin is produced in batch fermenters. Explain how a batch fermenter differs from a continuous fermenter. |
|---|-----|--|
| | | |
| | | |
| | | |
| | | |
| | | rea |

(b) Fig. 4.1 shows the number of reported cases, between the years 1982 and 1992, of a sexually-transmitted disease caused by a bacterium resistant to penicillin.

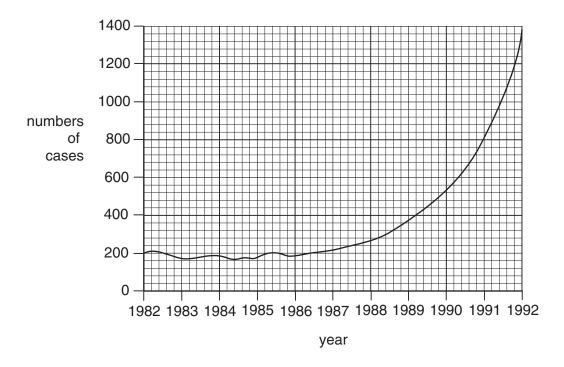


Fig. 4.1

| | | | | | | | [| [2] |
|-------|-----------|---------------|---------------|---------------|--------------|--------------|---------|-----|
| | | | | | | | | |
| | | | | | | | | |
| 1992. | | | | | | | | |
| | t wny tne | steep rise ii | i the incidei | nce of this c | ilsease occu | rred between | 1987 ar | na |

(c) Penicillin is produced as a secondary metabolite in batch fermenters. Fig. 4.2. shows the rate of penicillin production when grown in various glucose concentrations.

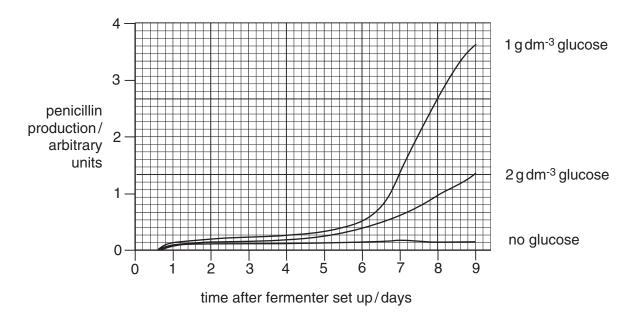


Fig. 4.2

| Describe how the glucose concentration affects penicillin production. | | | |
|---|-------------|--|--|
| | | | |
| | | | |
| | | | |
| | | | |
| | [3] | | |
| | | | |
| | [Total : 8] | | |

OPTION 3 – GROWTH, DEVELOPMENT AND REPRODUCTION

| 1 | (a) | Describe briefly the process of fertilisation in humans. |
|---|-----|---|
| | | |
| | | |
| | | |
| | | |
| | | |
| | | [4] |
| | (b) | Mice and humans share a gene for a protein ion channel that is found only in the plasma membrane (cell surface membrane) of the tails of spermatozoa. Mice homozygous for a mutation which results in an inactive ion channel are sterile. |
| | | Explain how a mutation can result in an inactive ion channel. |
| | | |
| | | |
| | | |
| | | [3] |
| | (c) | Sperm from mice homozygous for the mutation giving an inactive ion channel and from homozygous normal mice were examined. Their rate of movement was measured and their ability to fertilise eggs was tested by <i>in vitro</i> fertilisation (IVF) of intact eggs and eggs without their outer zona pellucida. The results are shown in Table 1.1. |

Table 1.1

| | sperm from homozygous normal mice | sperm from homozygous mutant mice |
|--|-----------------------------------|--------------------------------------|
| rate of movement/µm s ⁻¹ | 180 | 60 |
| successful fertilisation of intact eggs/% | 81 | 0 |
| successful fertilisation of eggs with zona pellucida removed/% | 76 | 62 |

| (i) | Explain what is meant by in vitro fertilisation. |
|-------|--|
| | |
| | |
| | [2] |
| (ii) | With reference to Table 1.1, compare the abilities of sperm from normal and mutant mice to fertilise eggs successfully <i>in vitro</i> . |
| | |
| | |
| | |
| | |
| | [4] |
| (iii) | With reference to Table 1.1, suggest an explanation for the differences you have described in (ii). |
| | |
| | |
| | [2] |
| | [Total · 15] |

| 2 | (a) | Explain briefly what is meant by <i>growth</i> of an organism. |
|---|-----|--|
| | | |
| | | |
| | | [2] |

(b) Two thousand Sitka spruce trees were planted 2.0 m apart and their growth in volume measured at five year intervals from ages 20 to 75 years.

Table 2.1 shows the mean volume of the trees per hectare from 20 to 75 years, their mean increase in volume per 5 year period and also their relative growth calculated as:

mean change in volume over 5 year period mean volume at start of 5 year period

Table 2.1

| | | 1 | |
|--------------|---|--|--|
| | | | relative growth of trees |
| age of trees | mean volume of | mean increase in | |
| / years | trees / m ³ ha ⁻¹ | volume per 5 year | mean increase in volume over 5 year period |
| | | period / m ³ ha ⁻¹ | mean volume at start of 5 year period |
| 20 | 66 | _ | _ |
| 25 | 133 | 67 | 1.02 |
| 30 | 214 | 81 | 0.61 |
| 35 | 301 | 87 | 0.41 |
| 40 | 386 | 85 | 0.28 |
| 45 | 465 | 79 | 0.20 |
| 50 | 534 | 69 | 0.15 |
| 55 | 593 | 59 | 0.11 |
| 60 | 642 | 49 | 0.08 |
| 65 | 683 | 41 | 0.06 |
| 70 | 718 | 35 | 0.05 |
| 75 | 751 | 33 | |

to 75 years; and put your answer in the space provided in Table 2.1.

(i) calculate, showing your working, the relative growth of the trees for the period 70

With reference to Table 2.1,

| [2 |
|---|
| (ii) describe the change in relative growth with age; |
| |
| iii) describe how the data could be used to plot a curve of absolute growth rate . |
| |
| [2] [Total : 9] |

3 (a) Fig. 3.1 shows the female gametophyte of a typical flowering plant.

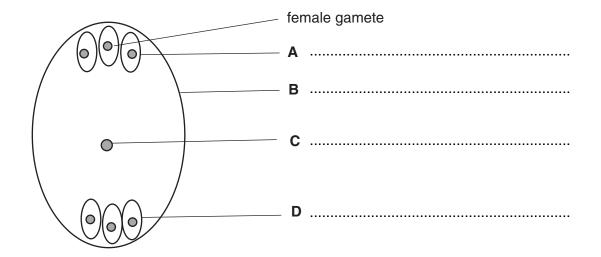


Fig. 3.1

| (1) | Name A to D. | [2] |
|------|--|-----|
| (ii) | Describe what happens at fertilisation to the female gamete and to structure C | |
| | female gamete | |
| | | |
| | | |
| | structure C | |
| | | |
| | | [4] |
| | | |

(b) The water lily, *Nuphar*, appears to have a female gametophyte containing only four haploid nuclei, as shown in Fig. 3.2.

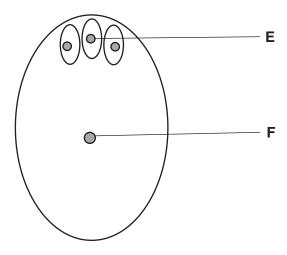


Fig. 3.2

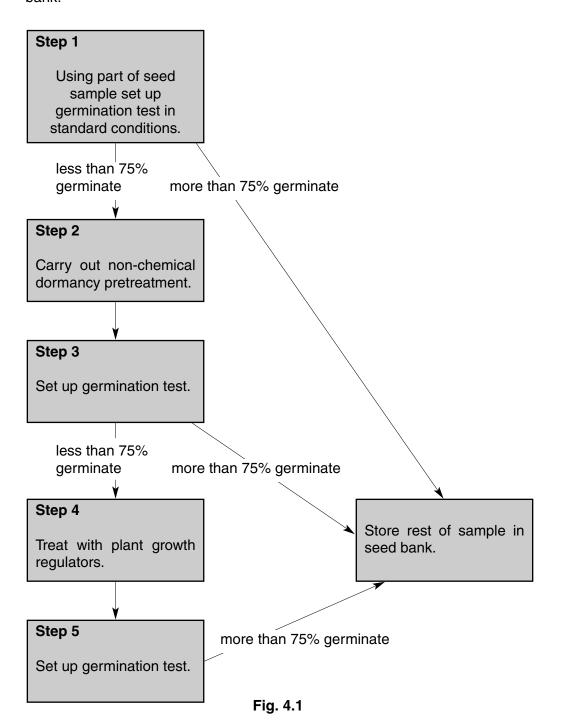
The DNA content of nuclei **E** and **F** can be measured by their fluorescence. This was measured in a number of ovules before and after fertilisation. The results are shown in Table 3.1.

Table 3.1

| | mean relative fluore unit | • |
|---------|------------------------------|---------------------|
| nucleus | before fertilisation | after fertilisation |
| E | 10.19 | 20.55 |
| F | 10.31 | 20.33 |

| illy | water | แเษ | liial | iggestion | эрогс | | loid nu | | | | |
|------|--------|-----|-------|-----------|-----------|------|---------|------|------|------|------|
| | ••••• | | | | | •••• | | | | | •••• |
| [2] | | | | | | | | | | | |
| : 8] | [Total | | | | | | | | | | |

4 (a) Fig. 4.1 shows the procedure that is followed when a sample of seeds arrives at a seed bank.



With reference to Fig. 4.1,

| (i) | describe the conditions that might be used for germinating seeds in steps ${\bf 1},{\bf 3}$ and ${\bf 5};$ |
|-------|---|
| | |
| | |
| | |
| | [3] |
| (ii) | describe briefly two different treatments that might be used to break seed dormancy in step 2 ; |
| | treatment 1 |
| | |
| | treatment 2 |
| | [2] |
| (iii) | name one plant growth regulator that might be used to break seed dormancy in step 4 and explain how it does so. |
| | plant growth regulator |
| | explanation |
| | |
| | [3] |
| | [Total : 8] |

OPTION 4 – APPLICATIONS OF GENETICS

| 1 | (a) | mer | mbrane (cell surface memb | | nat is found only in the plasma tozoa. Mice homozygous for rile. |
|---|------|----------------------|---|---|---|
| | | Ехр | lain how a mutation can re | esult in an inactive ion chann | el. |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | [3] |
| | (b) | hon thei of in | nozygous normal mice wer r ability to fertilise | re examined. Their rate of n eggs was tested by <i>i</i> | nactive ion channel and from novement was measured and in vitro fertilisation (IVF) a. The results are shown in |
| | | | | Table 1.1 | |
| | | | - | | |
| ı | | | | sperm from homozygous normal mice | sperm from homozygous mutant mice |
| | rate | of r | movement/µm s ⁻¹ | 180 | 60 |
| | | cess s/% | sful fertilisation of intact | 81 | 0 |
| | | | oful fertilisation of eggs na pellucida removed/% | 76 | 62 |
| | | (i) | Explain what is meant by | in vitro fertilisation. | |
| | | | | | |
| | | | | | |
| | | (ii) | With reference to Table 1 mice to fertilise eggs succ | .1, compare the abilities of sp | perm from normal and mutant |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |

| (iii) | With reference to Table 1.1, suggest an explanation for the differences you have described in (ii). |
|-------|--|
| | |
| | |
| | [2] |

- (c) The DNA coding for the ion channel was analysed by a process similar to genetic fingerprinting. DNA from three different mice was used:
 - **A** a mouse homozygous for the normal ion channel;
 - **B** a heterozygous mouse;
 - **C** a mouse homozygous for the mutation giving an inactive ion channel.

The resulting genetic fingerprint is shown in Fig. 1.1.

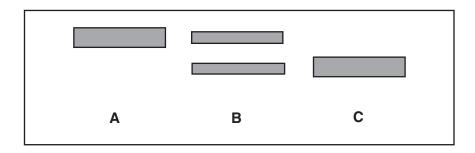


Fig. 1.1

With reference to Fig. 1.1, explain

| (i) | the difference in the position of the bands from mouse A and mouse C in the genetic fingerprint; |
|-----|--|
| | |
| | |
| | [3] |
| ii) | the presence of two bands in the genetic fingerprint of mouse ${\bf B}$. |
| | |
| | [1] |
| | [Total : 15] |

| 2 | (a) | disease of rice called rice blast. Non-sticky, hybrid rices are more resistant to rice blast. |
|---|-----|---|
| | | Explain briefly how sticky rice might be selectively bred to show resistance to rice blast. |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | 1/1 |

(b) In 1998 and 1999 a large number of farmers in Yunnan Province planted both sticky rice and hybrid rice alone (in monoculture) and in mixed planting. In the mixed plantings, single rows of sticky rice (S) were planted between rows of hybrid rice (R) in the pattern shown in Fig. 2.1.

RRSRR RRSRR RRSRR RRSRR RRSRR RRSRR RRSRR RRSRR

Fig. 2.1

The severity of infection by rice blast in mixed planting was then compared with that in monoculture. The genetic diversity of the rice blast fungus populations was also found. The results are shown in Table 2.1.

Table 2.1

| | rice grown in monoculture | rice grown in mixed planting |
|---|------------------------------|--|
| mean percentage of stems of sticky rice dying from rice blast | 20.0 | 1.2 |
| mean percentage of stems of hybrid rice dying from rice blast | 2.3 | 1.0 |
| genetic diversity of rice blast fungus populations | one or a few strains | many strains with no single predominant strain |

With reference to Table 2.1, explain

| (i) | the difference in infection by rice blast between monoculture and mixed | planting; |
|------|---|-------------|
| | | |
| | | [2] |
| (ii) | the difference in the genetic diversity of rice blast fungus populations. | |
| | | |
| | | [2] |
| | | [Total : 8] |

3 Drylands are among the most threatened environments in the world. Seeds from dryland plants are being collected and stored. At intervals, samples of seeds from storage are germinated, plants grown and a new generation of seeds collected for storage.

Seeds from plants that only grow in one place in the world, from endangered species and from plants used by local people may be stored in the country of origin or in the Millennium Seed Bank in the UK.

| (a) | Explain why dryland seeds are being collected and stored. |
|-----|---|
| | |
| | |
| | |
| | [3] |
| (b) | Describe the conditions in which seeds are stored in a seed bank. |
| | |
| | |
| | [2] |
| (c) | Explain why, at intervals, samples of seeds are germinated, plants grown and a new generation of seeds collected for storage. |
| | |
| | |
| | [2] |
| | [Total : 7] |

Question 4 begins on the next page.

| | | lain the genetic basis of Down's syndrome in humans. | | | | |
|-----|------------|--|--------------|----------------------------------|----------------------------|--------------|
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| (b) | Sta | te what is meant by <i>genetic screening</i> . | | | | |
| | | | | | | |
| | | | | | | |
| (c) | syn wee | study into diagnosis of Down's syndrome, the way in which drome were initially diagnosed was recorded. All had eks of pregnancy by one of four different tests, A , B , C or onbers. | been | scree | ned b | efore |
| | | number of cases detected by each test, as well as t sed by the tests, and so detected only at or after birth, is | | | | |
| | | Table 4.1 | | | | |
| | | | | | | |
| | | Screening test | Α | В | С | D |
| | | Screening test number of cases of Down's syndrome detected before 24 weeks of pregnancy | A 32 | B 36 | C 42 | D 61 |
| nu | mbe | number of cases of Down's syndrome detected | | 36 | | |
| nu | mbe | number of cases of Down's syndrome detected before 24 weeks of pregnancy | | 36 1 | 42 | |
| nu | | number of cases of Down's syndrome detected before 24 weeks of pregnancy r of cases of Down's syndrome detected at or after birth | 32 | 36 1! 32 | 42 52 23 | 61 |
| nu | Witl | number of cases of Down's syndrome detected before 24 weeks of pregnancy r of cases of Down's syndrome detected at or after birth total number of cases of Down's syndrome n reference to Table 4.1. calculate the percentage of cases of Down's syndrome | 32 | 36 18 32 were c | 42 52 23 | 61 ed bet |
| | Witl | number of cases of Down's syndrome detected before 24 weeks of pregnancy r of cases of Down's syndrome detected at or after birth total number of cases of Down's syndrome n reference to Table 4.1. calculate the percentage of cases of Down's syndrome | 32 | 36 15 32 were c | 42 52 23 | 61 ed bef |
| | Witl | number of cases of Down's syndrome detected before 24 weeks of pregnancy r of cases of Down's syndrome detected at or after birth total number of cases of Down's syndrome reference to Table 4.1. calculate the percentage of cases of Down's syndrome 24 weeks of pregnancy. Show your working; describe the differences in the success of the differences of | 32 that v | 36 15 32 were c | 42 52 23 detector | ed bef |
| | Witl | number of cases of Down's syndrome detected before 24 weeks of pregnancy r of cases of Down's syndrome detected at or after birth total number of cases of Down's syndrome reference to Table 4.1. calculate the percentage of cases of Down's syndrome 24 weeks of pregnancy. Show your working; describe the differences in the success of the difficulty diagnosing Down's syndrome before birth; | 32 that v | 36 15 32 were comments screen | 42 52 23 detecte | ed bef |
| | Witl | number of cases of Down's syndrome detected before 24 weeks of pregnancy r of cases of Down's syndrome detected at or after birth total number of cases of Down's syndrome reference to Table 4.1. calculate the percentage of cases of Down's syndrome 24 weeks of pregnancy. Show your working; describe the differences in the success of the difficial diagnosing Down's syndrome before birth; | 32 that v | 36 15 32 were comments screen | 42 52 23 detecte | ed bef |

| (iii) | suggest one way of making use of tests A , B , C and D that would increase the percentage of cases of Down's syndrome detected before birth. | |
|-------|---|--|
| | [1] | |
| | [Total : 10] | |

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