

# High Fidelity DNA Polymerase Introduces Bias into 16S rRNA Gene Sequencing Results

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# Abstract

**Background.** Bias is introduced at different stages of the 16S rRNA gene sequencing workflow. Commonly studied sources of bias include preservation and DNA extraction methods. Although cycle number and high fidelity (HiFi) DNA polymerase are studied less often, they are still important sources of bias in this workflow. Here, we examine how both cycle number and HiFi DNA polymerase can change the bacterial community and introduce bias to the final obtained results.

**Methods.** We extracted DNA from fecal samples (n=4) using a PowerMag DNA extraction kit with a 10 minute bead beating step and amplified at 15, 20, 25, 30, and 35 cycles using Accuprime, Kappa, Phusion, Platinum, or Q5 HiFi DNA polymerase. Amplification of mock communities (technical replicates n=4) consisting of previously isolated whole genomes of 8 different bacteria used the same approach. The analysis initially examined the number of Operational Taxonomic Units (OTUs) for both fecal samples and mock communities. It also assessed HiFi DNA polymerase dependent differences in the Bray-Curtis index, error rate, sequence error prevalence, and chimera prevalence. Our analysis also examined chimera prevalence correlation with the number of OTUs.

**Results.** When analyzing fecal samples we observed a different number of OTUs between HiFi DNA polymerases at 35 cycles (P-value < 0.0001). Our analysis identified these HiFi dependent differences in the number of OTUs as early as 20 cycles in the mock communities (P-value = 0.002). Chimera prevalence varied by HiFi DNA polymerase and this variation persisted after chimera removal using VSEARCH. We also observed positive correlations between chimera prevalence and the number of OTUs which was not affected by chimera removal with VSEARCH.

**Conclusions.** HiFi DNA polymerase dependent differences in the number of OTUs and chimera prevalence makes comparison across studies difficult. Care should be exercised

<sup>26</sup> when choosing both HiFi DNA polymerase and cycle number to be used in 16S rRNA gene  
<sup>27</sup> sequencing studies.

## Introduction

Accounting for bias is critical in order to reach an understanding of bacterial community changes using 16S rRNA gene sequencing results. Differentiating between bias, reproducibility, and standardization is important since often times these three can be confused and used interchangeably with each other. Bias can change the observed results in a way that is reproducible and standardized. For example, if one group uses one brand of DNA extraction kit for their 16S rRNA gene sequencing, their results may be biased versus another group not using the same brand kit but within their group they can still have reproducible results (Salter et al., 2014). Our 16S rRNA gene sequencing methods are biased even when these workflows are standardized to increase reproducibility. In order to interpret specific studies within the broader context of the overall field, assessing bias at different parts of the 16S rRNA gene sequencing workflow is critical.

Many parts of the 16S rRNA gene sequencing workflow contribute bias to the results and are studied extensively. A typical 16S rRNA gene sequencing workflow can be divided into preservation, extraction, PCR, and sequencing steps. Generally, not using a preservation media and leaving samples at room temperature supports overgrowth of low abundance members of the fecal bacterial community (Amir et al., 2017). Similarly, this overgrowth can still occur if the preservation media does not adequately inhibit growth (Song et al., 2016; Luo et al., 2016). Reports have also shown that changes in specific community members might occur due to differing susceptibility to freeze thaw cycles amongst microbes (Gorzelak et al., 2015). Additionally, reagent contamination can add community members and the contribution of these contaminant members grows larger with lower biomass samples (Salter et al., 2014). Overall, biases due to either preservation or extraction tend to be smaller than the overall biological signal being measured (Song et al., 2016; Bassis et al., 2017). However, the contribution of PCR bias to this overall workflow is not well characterized since these studies use the same PCR approach while varying preservation

or extraction method.

Identifying the biases in the PCR stage of 16S rRNA gene sequencing is important because a large body of literature shows that there are a variety of steps during PCR that can change the observed results (Eckert & Kunkel, 1991; Burkardt, 2000). Many of these sources of biases are made worse as cycle number increases (Wang & Wang, 1996; Haas et al., 2011; Kebschull & Zador, 2015). For example, the selective amplification of AT-rich over GC-rich sequences can exaggerate the difference between 16S rRNA genes higher in AT versus those higher in GC (Polz & Cavanaugh, 1998). Both amplification error and non-specific amplification (e.g. incorrect amplicon size products) can also increase as cycle number increases which can drastically change commonly used diversity measures (Acinas et al., 2005; Santos et al., 2016). Additionally, chimeras can form from an aborted extension step followed by a subsequent priming error and secondary extension and will artificially increase community diversity (Haas et al., 2011). Although these differences are not necessarily dependent on primer and DNA polymerase used, there are also biases that are.

The intrinsic properties to primers and DNA polymerases chosen can also introduce bias. Primers have variable region dependent binding affinities for different bacteria and depending on the primer pair do not detect specific bacteria (e.g. V1-V3 does not detect *Haemophilus influenzae* and V3-V5 does not detect *Propionibacterium acnes*) (Sze et al., 2015 (Table S4); Meisel et al., 2016). Additionally, there are multiple families of DNA polymerases that have their own error rate and proof reading capacity (Ishino & Ishino, 2014). Interestingly, the influence that these different DNA polymerases can have on the observed 16S rRNA gene sequencing results have not been well studied like some of the other previously mentioned sources of PCR-based bias.

A recent study found clear differences between normal and high fidelity (HiFi) DNA polymerase and that optimization of the PCR protocol could reduce error and chimera

80 generation (Gohl et al., 2016). This study also found that regardless of DNA polymerase,  
81 the number of Operational Taxonomic Units (OTUs) or taxa generated were not easily  
82 reduced using the authors chosen bioinformatic pipeline (Gohl et al., 2016). It is natural to  
83 extend this line of inquiry and ask if biases in the number of OTUs and chimeras are also  
84 dependent on the type of HiFi DNA polymerase. There is some reason to think that this  
85 may be the case since many of these HiFi DNA polymerases come from different families  
86 (e.g. *Taq* belongs to the family A polymerases) and may intrinsically have different error  
87 rates that cannot be completely removed with modifications (Ishino & Ishino, 2014).

88 Although bias introduced due to differences between DNA polymerase and HiFi DNA  
89 polymerase has been investigated for 16S rRNA gene sequencing, the bias caused due  
90 to differences between specific HiFi DNA polymerases has not been. This study will  
91 specifically address how HiFi DNA polymerases can bias observed bacterial community  
92 results derived from 16S rRNA gene sequencing. We will accomplish this by examining if  
93 any of five different types of HiFi DNA polymerases introduce significant biases into 16S  
94 rRNA gene surveys, if this is a cycle dependent phenomenon, and whether they can be  
95 removed using a standard bioinformatic pipeline.

## Materials & Methods

**Human and Mock Samples:** A single fecal sample was obtained from 4 individuals who were part of the Enterics Research Investigational Network (ERIN). The processing and storage of these samples were previously published (Seekatz et al., 2016). Other than confirmation that none of these individuals had a *Clostridium difficile* infection, clinical data and other types of meta data were not utilized or accessed for this study. All samples were extracted using the MOBIO™ PowerMag Microbiome RNA/DNA extraction kit (now Qiagen, MD, USA). The ZymoBIOMICS™ Microbial Community DNA Standard (Zymo, CA, USA) was used for mock communities and was made up of *Pseudomonas aeruginosa*, *Escherichia coli*, *Salmonella enterica*, *Lactobacillus fermentum*, *Enterococcus faecalis*, *Staphylococcus aureus*, *Listeria monocytogenes*, and *Bacillus subtilis* at equal genomic DNA abundance (<http://www.zymoresearch.com/microbiomics/microbial-standards/zymbiomics-microbial-community-standards>).

**PCR Protocol:** The five different HiFi DNA polymerases that were tested included AccuPrime™ (ThermoFisher, MA, USA), KAPA HIFI (Roche, IN, USA), Phusion (ThermoFisher, MA, USA), Platinum (ThermoFisher, MA, USA), and Q5 (New England Biolabs, MA, USA). The PCR cycle conditions for Platinum and Accuprime followed a previously published protocol (Kozich et al., 2013) ([https://github.com/SchlossLab/MiSeq\\_WetLab\\_SOP/blob/master/MiSeq\\_WetLab\\_SOP\\_v4.md](https://github.com/SchlossLab/MiSeq_WetLab_SOP/blob/master/MiSeq_WetLab_SOP_v4.md)). The HiFi DNA polymerase activation time was 2 minutes, unless a different activation was specified. For Kappa and Q5, a previously published protocol was used (Gohl et al., 2016). For Phusion, the company defined conditions were used except for extension time, where the Accuprime and Platinum settings were used.

The cycle conditions for both fecal and mock samples started at 15 and increased by 5 up to 35 cycles with amplicons used at each 5-step increase for sequencing. The PCR of

fecal DNA samples consisted of all 4 samples at 15, 20, 25, 30, and 35 cycles for each HiFi DNA polymerase (total sample n=100). The mock communities had 4 replicates at 15, 20, 25, and 35 cycles and 10 replicates for 30 cycles for all HiFi DNA polymerases (total samples n=130). No mock community sample had enough PCR product at 15 cycles for adequate 16S rRNA gene sequencing.

**Sequence Processing:** The mothur software program was used for all sequence processing steps (Schloss et al., 2009). The protocol has been previously published (Kozich et al., 2013) ([https://www.mothur.org/wiki/MiSeq\\_SOP](https://www.mothur.org/wiki/MiSeq_SOP)). Two major differences from the published protocol were the use of VSEARCH instead of UCHIME for chimera detection and the use of the OptiClust algorithm instead of average neighbor for OTU generation at 97% similarity (Edgar et al., 2011; Rognes et al., 2016; Westcott & Schloss, 2017). Sequence error was determined using the 'seq.error' command on mock samples before the 'pre.cluster' command, before chimera removal, and after chimera removal (Schloss et al., 2009; Cole et al., 2013; Rognes et al., 2016).

**Analysis Workflow:** The total number of OTUs was analyzed after sub-sampling for both the fecal and mock community samples. For fecal samples, cycle dependent affects on Bray-Curtis indices were assessed for cycle group and within individual differences from the previous cycle (e.g. 20 versus 25, 25 versus 30, etc.). These community based measures for fecal samples were analyzed at 4 different sub-sampling sequence depths (1000, 5000, 10000, and 15000) while the mock community samples were analysed at 3 levels (1000, 5000, 10000). Based on these observations we analyzed potential reasons for these differences. Analysis of the mock community of each HiFi DNA polymerase for general sequence error rate, number of sequences with an error, base substitution, and numbers of chimeras were assessed before the 'pre.cluster' command, before chimera removal, and after chimera removal. Additionally, the correlation between the number of chimeras and the number of OTUs was also assessed before the 'pre.cluster' command,



before chimera removal, and after chimera removal.

**Statistical Analysis:** All analysis was done with the R (v 3.4.4) software package (R Core Team, 2017). Data transformation and graphing was completed using the tidyverse package (v 1.2.1) and colors selected using the viridis package (v 0.4.1) (Garnier, 2017; Wickham, 2017). Differences in the total number of OTUs were analyzed using an ANOVA with a tukey post-hoc test. For the fecal samples the data was normalized to each individual by cycle number to account for the biological variation between people. Bray-Curtis distance matrices were generated using mothur after 100 sub-samplings at 1000, 5000, 10000, and 15000 sequence depth. The distance matrix data was analyzed using PERMANOVA with the vegan package (v 2.4.5) (Oksanen et al., 2017) and Kruskal-Wallis tests within R. For both error and chimera analysis, samples were tested using Kruskal-Wallis with a Dunns post-hoc test. Where applicable correction for multiple comparison utilized the Benjamini-Hochberg method (Benjamini & Hochberg, 1995).

**Reproducible Methods:** The code and analysis can be found here [https://github.com/SchlossLab/Sze\\_PCRSeqEffects\\_XXXX\\_2017](https://github.com/SchlossLab/Sze_PCRSeqEffects_XXXX_2017). The raw sequences can be found in the SRA at the following accession number SRP132931.

## Results

***The Number of OTUs are Dependent on HiFi DNA Polymerase:*** A consistent difference in the number of OTUs, that was dependent on the HiFi DNA polymerase used was observed regardless of sub-sampling depth for fecal samples [Figure 1]. Additionally, there was a trend for lower cycle numbers (15-20) to result in less differences in the number of OTUs versus higher cycle numbers (25, 30, and 35) between HiFi DNA polymerases [Figure 1]. For fecal samples, all sub-sampling levels had significant differences between HiFi DNA polymerases at 35 cycles (F-stat > 16.35, P-value < 0.0001) [Table S1]. Most of the differences observed at 35 cycles were between Platinum and other HiFi DNA polymerases, based on a Tukey post-hoc test (P-value < 0.05) [Table S2]. Differences in the number of OTUs between HiFi DNA polymerases were identifiable at earlier cycles (25 and 30) but the sub-sampling depth had to be 5000 sequences or higher (F-stat > 4.98, P-value < 0.05) [Table S1].

This HiFi DNA polymerase dependent difference in the number of OTUs was also observed in the mock community samples [Figure 2]. Regardless if fecal or mock communities were used, the same HiFi DNA polymerases had high (Platinum) and low (Accuprime) number of OTUs and this was consistent across cycle number and sub-sampling depth [Figure 1-2 & Table S1-S4]. In contrast to the results obtained with fecal samples, differences between HiFi DNA polymerases were observed as early as 20 cycles and at as low of a sub-sampling depth as 1000 sequences in the mock community samples (F-stat = 15.82, P-value = 0.002) [Table S3]. For both cycle numbers and sub-sampling depths, the majority of differences in the number of OTUs were between Platinum and the other HiFi DNA polymerases [Table S4]. Based on these observations in fecal and mock communities, it is clear that different HiFi DNA polymerases result in a different total number of OTUs observed within a sample.

**Minimal Bray-Curtis Differences are Detected and are Dependent on both Cycle**

**Number and Sub-Sampling Depth:**

A few small differences based on sub-sampling and cycle number were detected in overall bacterial community composition. Within the same fecal sample and independent of HiFi DNA polymerases, there were differences in the community composition between 20 and 25 cycles that was dependent on sub-sampling depth (sub-sampled to 1000 = 0.51 (0.4 - 0.79) (median (IQR)), sub-sampled to 5000 = 0.43 (0.33 - 0.63), sub-sampled to 10000 = 0.4 (0.24 - 0.43)) [Figure 3A]. Further, when data was available for the mock communities, there were larger observed differences between 20 and 25 cycles (sub-sampled to 1000 = 0.88 (0.42 - 0.91)) [Figure 3B]. Additionally, these stated community differences disappear when comparing 25 to 30 cycles and do not persist past 25 cycles [Figure 3]. Although these trends are clearly noticeable, we found that there was no detectable difference in Bray-Curtis index when comparing to the previous 5-cycle increment for both fecal and mock communities after multiple comparison correction (P-value > 0.05). Using PERMANOVA to test for community differences based on cycle number within HiFi DNA polymerases, only Phusion had cycle dependent differences at 1000 and 5000 sub-sampling depths (P-value = 0.03 and 0.01). For fecal samples, Phusion was one of two HiFi DNA polymerases that had enough sequences to reach a sub-sampling depth of 1000 at 15 cycles. Overall, these data suggest that there are small HiFi DNA polymerase differences in Bray-Curtis index that are dependent on both sub-sampling depth and cycle number.

**Sequence Error is Dependent on both HiFi DNA Polymerase and Cycle Number:**

Differences in the median average per base error varied by HiFi DNA polymerase without a clear pattern across sub-sampling depth [Table S5]. The highest median average per base error rates were for the Kappa HiFi DNA polymerase [Figure 4]. This error rate was minimally affected by both the 'pre.cluster' step and chimera removal by VSEARCH [Figure 4]. The differences in the median average per base error rate between the different HiFi DNA polymerases was cycle dependent with Platinum having the largest changes

versus other HiFi DNA polymerases [Figure 4B-C and Table S6]. The total sequences with at least one error was also cycle number dependent and differences between HiFi DNA polymerases could be drastically reduced by the use of the 'pre.cluster' step [Figure S1]. These differences in sequences with at least one error were mostly due to differences in Accuprime<sup>TM</sup> and Platinum versus the other HiFi DNA polymerases [Figure S1 & Table S7 & S8]. Finally, we did not observe a HiFi DNA polymerase dependent difference on base substitution rate [Figure S2]. Although sequence error is dependent on HiFi DNA polymerase some of these error dependent differences can be corrected using existing bioinformatic approaches.

***Prevalence of Chimeric Sequences are HiFi DNA Polymerase Dependent and Correlate with the Number of OTUs:*** There were significant differences in the chimera prevalence based on HiFi DNA polymerase used at all levels of sub-sampling and cycle numbers (P-value < 0.05) [Table S9]. Differences in chimera prevalence between Platinum and all other HiFi DNA polymerases accounted for the majority of these differences [Table S10]. Accuprime<sup>TM</sup> had the lowest chimera prevalence of all HiFi DNA polymerases regardless of whether 'pre.cluster' or chimera removal with VSEARCH was used [Figure 5]. A positive correlation was observed between chimeric sequences and the number of OTUs for all HiFi DNA polymerases [Figure 6]. This positive correlation was strongest for Accuprime<sup>TM</sup>, Platinum, and Phusion HiFi DNA Polymerases [Figure 6]. The R<sup>2</sup> value between the number of OTUs and chimeric sequences did not change with the use of 'pre.cluster' or with the removal of chimeras using VSEARCH [Figure 6]. This data suggests that chimera prevalence depends on HiFi DNA polymerase used and confirms that the number of OTUs is dependent on the prevalence of these chimeric sequences.

## Discussion

In this study we show that the number of OTUs, error rate, and chimera prevalence depends on HiFi DNA polymerase [Figure 1-2 & 4-5]. These differences are important because many diversity metrics rely on the number of OTUs or other measures dependent on error rate and chimera prevalence as part of their metric calculations (e.g. richness). Additionally, the earlier detection of differences in total number of OTUs between HiFi DNA polymerases in the mock versus fecal samples might indicate that high biomass samples may underestimate the biases present within low biomass samples. We observed that undetected chimeras that were not identified and removed using standard bioinformatic approaches cause many of these differences. This suggests that specific diversity differences between studies can be attributed to differences in HiFi DNA polymerase used. Based on our observations metrics that measure within sample diversity depend on HiFi DNA polymerase but this may not be the case for metrics that assess between sample diversity.

There were few differences that depend on HiFi DNA polymerase for between sample diversity, as measured by the Bray-Curtis index. Using this metric our observations found no differences in the overall bacterial community composition for sub-sampling depth or cycle number used. One possible reason for this outcome was that our study did not have enough power to detect differences due to low sample number in each group. Another reason was that many of the OTUs are likely not highly abundant, allowing the Bray-Curtis index to be able to successfully down-weight chimeric OTUs (Minchin, 1987). The choice of downstream diversity metric could be an important consideration in helping to mitigate these observed HiFi DNA polymerase dependent differences in chimera prevalence. Metrics that solely use presence/absence of OTUs (e.g. Jaccard (Real & Vargas, 1996)) may be less robust to chimera prevalence and by extension total number of OTU differences in HiFi DNA Polymerases. When choosing a distance metric,

careful consideration of the biases introduced from the PCR step of the 16S rRNA gene sequencing workflow need to be taken into account. With differences in the number of OTUs and chimera prevalence depending on HiFi DNA polymerase used, it might be easier to avoid specific DNA polymerase families altogether.

Although the variation in error rate and chimera prevalence may be due to the DNA polymerase family, the highest and lowest chimera rates both belonged to a family A polymerase (Platinum and Accuprime™ respectively) (Ishino & Ishino, 2014). Additionally, based on the material safety data sheet (MSDS) the differences between the two HiFi DNA polymerases are not immediately apparent. Both HiFi DNA polymerases contain a recombinant *Taq* DNA polymerase, a *Pyrococcus* spp GB-D polymerase and a platinum *Taq* antibody. With everything else being equal, it is possible that differences in how the recombinant *Taq* was generated could be a contributing factor for the differences observed between the HiFi DNA polymerases. We are unlikely to avoid adding HiFi dependent bias to 16S rRNA gene sequencing results, however, these differences may not be large enough to mask the actual biological signal.

The majority of HiFi DNA polymerases we studied add small increases in the number of OTUs and chimera prevalence that can be masked by biological differences. The sequence error introduced by the HiFi DNA polymerase is small and likely to be smaller than the biological variation within a specific study, which would be consistent with previous findings for preservation and DNA extraction methods (Salter et al., 2014; Song et al., 2016; Luo et al., 2016). However, the chimera prevalence for some HiFi DNA polymerases (e.g. Platinum) are relatively large and might be greater than the observed biological variation within a specific study. The choice of HiFi DNA polymerase can be as important a consideration as either preservation or DNA extraction method used because similar to using different preservation methods or different DNA extraction kits, the type of HiFi DNA polymerase can add bias to the observed bacterial community. Although avoiding

HiFi DNA polymerases that yield a high number of OTUs and chimera prevalence might provide better standardization of results it does come at a cost.

Methods can be standardized but they commonly contain bias that is reproducible and may miss important associations. Bias can be easily reproduced and can be found in every step of the 16S rRNA gene sequencing workflow (Salter et al., 2014; Gohl et al., 2016; Luo et al., 2016; Amir et al., 2017). This study shows that specific diversity metrics used to measure the microbial community consistently vary based on HiFi DNA polymerase. Standardizing multiple workflows to one specific HiFi DNA polymerase could be detrimental since some HiFi DNA polymerases may work better in certain situations over others. Arguably, the degree of workflow standardization across studies and research group needs to be approached on a study by study basis and not every project needs to use the exact same approach. All aspects of the 16S rRNA gene sequencing workflow need to be customized for the specific microbial community that will be sampled. Although a diversity of approaches may make reproducibility and replicability more difficult it will help to avoid systematic biases from occurring due to widespread standardization of approaches.

## Conclusion

Our observations fill a gap in knowledge on the bias introduced to 16S rRNA gene sequencing results due to differences in HiFi DNA polymerases. We found that the number of OTUs and the chimera prevalence is dependent on both HiFi DNA polymerase and cycle number chosen. Care should be taken when choosing a HiFi DNA polymerase for 16S rRNA gene surveys because their intrinsic differences can change the number of OTUs observed and influence diversity based metrics that do not down-weight rare observations. Knowing the inherent bias associated with different HiFi DNA polymerases allows for better interpretation of the relationship between an individual study to their respective field of research.



## Acknowledgements

The authors would like to thank all the study participants in ERIN whose samples were utilized. We would also like to thank Judy Opp and April Cockburn for their effort in sequencing the samples as part of the Microbiome Core Facility at the University of Michigan. Salary support for Marc A. Sze came from the Canadian Institute of Health Research and NIH grant UL1TR002240. Salary support for Patrick D. Schloss came from NIH grants P30DK034933 and 1R01CA215574.

## References

- Acinas SG., Sarma-Rupavtarm R., Klepac-Ceraj V., Polz MF. 2005. PCR-induced sequence artifacts and bias: Insights from comparison of two 16S rRNA clone libraries constructed from the same sample. *Applied and Environmental Microbiology* 71:8966–8969. DOI: 10.1128/aem.71.12.8966-8969.2005.
- Amir A., McDonald D., Navas-Molina JA., Debelius J., Morton JT., Hyde E., Robbins-Pianka A., Knight R. 2017. Correcting for microbial blooms in fecal samples during room-temperature shipping. *mSystems* 2:e00199–16. DOI: 10.1128/msystems.00199-16.
- Bassis CM., Nicholas M. Moore., Lolans K., Seekatz AM., Weinstein RA., Young VB., Hayden MK. 2017. Comparison of stool versus rectal swab samples and storage conditions on bacterial community profiles. *BMC Microbiology* 17. DOI: 10.1186/s12866-017-0983-9.
- Benjamini Y., Hochberg Y. 1995. Controlling the false discovery rate: A practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society. Series B (Methodological)* 57:289–300.
- Burkardt H-J. 2000. Standardization and quality control of PCR analyses. *Clinical Chemistry and Laboratory Medicine* 38. DOI: 10.1515/cclm.2000.014.
- Cole JR., Wang Q., Fish JA., Chai B., McGarrell DM., Sun Y., Brown CT., Porras-Alfaro A., Kuske CR., Tiedje JM. 2013. Ribosomal database project: Data and tools for high throughput rRNA analysis. *Nucleic Acids Research* 42:D633–D642. DOI: 10.1093/nar/gkt1244.
- Eckert KA., Kunkel TA. 1991. DNA polymerase fidelity and the polymerase chain reaction. *Genome Research* 1:17–24. DOI: 10.1101/gr.1.1.17.
- Edgar RC., Haas BJ., Clemente JC., Quince C., Knight R. 2011. UCHIME improves

345 sensitivity and speed of chimera detection. *Bioinformatics* 27:2194–2200. DOI:  
 346 10.1093/bioinformatics/btr381.

347 Garnier S. 2017. *Viridis: Default color maps from 'matplotlib'*.

348 Gohl DM., Vangay P., Garbe J., MacLean A., Hauge A., Becker A., Gould TJ., Clayton  
 349 JB., Johnson TJ., Hunter R., Knights D., Beckman KB. 2016. Systematic improvement  
 350 of amplicon marker gene methods for increased accuracy in microbiome studies. *Nature*  
 351 *Biotechnology* 34:942–949. DOI: 10.1038/nbt.3601.

352 Gorzelak MA., Gill SK., Tasnim N., Ahmadi-Vand Z., Jay M., Gibson DL. 2015. Methods for  
 353 improving human gut microbiome data by reducing variability through sample processing  
 354 and storage of stool. *PLOS ONE* 10:e0134802. DOI: 10.1371/journal.pone.0134802.

355 Haas BJ., Gevers D., Earl AM., Feldgarden M., Ward DV., Giannoukos G., Ciulla D.,  
 356 Tabbaa D., Highlander SK., Sodergren E., Methe B., DeSantis TZ., Petrosino JF.,  
 357 Knight R., and BWB. 2011. Chimeric 16S rRNA sequence formation and detection in  
 358 sanger and 454-pyrosequenced PCR amplicons. *Genome Research* 21:494–504. DOI:  
 359 10.1101/gr.112730.110.

360 Ishino S., Ishino Y. 2014. DNA polymerases as useful reagents for biotechnology â  
 361 the history of developmental research in the field. *Frontiers in Microbiology* 5. DOI:  
 362 10.3389/fmicb.2014.00465.

363 Kebschull JM., Zador AM. 2015. Sources of PCR-induced distortions in high-throughput  
 364 sequencing data sets. *Nucleic Acids Research:gkv717*. DOI: 10.1093/nar/gkv717.

365 Kozich JJ., Westcott SL., Baxter NT., Highlander SK., Schloss PD. 2013. Development of  
 366 a dual-index sequencing strategy and curation pipeline for analyzing amplicon sequence  
 367 data on the MiSeq illumina sequencing platform. *Applied and Environmental Microbiology*

79:5112–5120. DOI: 10.1128/aem.01043-13.

Luo T., Srinivasan U., Ramadugu K., Shedden KA., Neiswanger K., Trumble E., Li JJ., McNeil DW., Crout RJ., Weyant RJ., Marazita ML., Foxman B. 2016. Effects of specimen collection methodologies and storage conditions on the short-term stability of oral microbiome taxonomy. *Applied and Environmental Microbiology* 82:5519–5529. DOI: 10.1128/aem.01132-16.

Meisel JS., Hannigan GD., Tyldsley AS., SanMiguel AJ., Hodgkinson BP., Zheng Q., Grice EA. 2016. Skin microbiome surveys are strongly influenced by experimental design. *Journal of Investigative Dermatology* 136:947–956. DOI: 10.1016/j.jid.2016.01.016.

Minchin PR. 1987. An evaluation of the relative robustness of techniques for ecological ordination. *Vegetatio* 69:89–107. DOI: 10.1007/bf00038690.

Oksanen J., Blanchet FG., Friendly M., Kindt R., Legendre P., McGlinn D., Minchin PR., O'Hara RB., Simpson GL., Solymos P., Stevens MHH., Szoecs E., Wagner H. 2017. *Vegan: Community ecology package*.

Polz MF., Cavanaugh CM. 1998. Bias in template-to-product ratios in multitemplate PCR. *Applied and Environmental Microbiology* 64:3724–3730.

R Core Team. 2017. *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing.

Real R., Vargas JM. 1996. The probabilistic basis of jaccards index of similarity. *Systematic Biology* 45:380–385. DOI: 10.1093/sysbio/45.3.380.

Rognes T., Flouri T., Nichols B., Quince C., Mahé F. 2016. VSEARCH: A versatile open source tool for metagenomics. *PeerJ* 4:e2584. DOI: 10.7717/peerj.2584.

Salter SJ., Cox MJ., Turek EM., Calus ST., Cookson WO., Moffatt MF., Turner P.,

391 Parkhill J., Loman NJ., Walker AW. 2014. Reagent and laboratory contamination  
 392 can critically impact sequence-based microbiome analyses. *BMC Biology* 12. DOI:  
 393 10.1186/s12915-014-0087-z.

394 Santos QMB-d los., Schroeder JL., Blakemore O., Moses J., Haffey M., Sloan W., Pinto AJ.  
 395 2016. The impact of sampling, PCR, and sequencing replication on discerning changes in  
 396 drinking water bacterial community over diurnal time-scales. *Water Research* 90:216–224.  
 397 DOI: 10.1016/j.watres.2015.12.010.

398 Schloss PD., Westcott SL., Ryabin T., Hall JR., Hartmann M., Hollister EB., Lesniewski  
 399 RA., Oakley BB., Parks DH., Robinson CJ., Sahl JW., Stres B., Thallinger GG., Horn  
 400 DJV., Weber CF. 2009. Introducing mothur: Open-source, platform-independent,  
 401 community-supported software for describing and comparing microbial communities.  
 402 *Applied and Environmental Microbiology* 75:7537–7541. DOI: 10.1128/aem.01541-09.

403 Seekatz AM., Rao K., Santhosh K., Young VB. 2016. Dynamics of the fecal microbiome in  
 404 patients with recurrent and nonrecurrent clostridium difficile infection. *Genome Medicine* 8.  
 405 DOI: 10.1186/s13073-016-0298-8.

406 Song SJ., Amir A., Metcalf JL., Amato KR., Xu ZZ., Humphrey G., Knight R. 2016.  
 407 Preservation methods differ in fecal microbiome stability, affecting suitability for field  
 408 studies. *mSystems* 11:e00021–16. DOI: 10.1128/msystems.00021-16.

409 Sze MA., Dimitriu PA., Suzuki M., McDonough JE., Campbell JD., Brothers JF.,  
 410 Erb-Downward JR., Huffnagle GB., Hayashi S., Elliott WM., Cooper J., Sin DD., Lenburg  
 411 ME., Spira A., Mohn WW., Hogg JC. 2015. Host response to the lung microbiome in  
 412 chronic obstructive pulmonary disease. *American Journal of Respiratory and Critical Care*  
 413 *Medicine* 192:438–445. DOI: 10.1164/rccm.201502-0223oc.

414 Wang GCY., Wang Y. 1996. The frequency of chimeric molecules as a consequence of

- 415 PCR co-amplification of 16S rRNA genes from different bacterial species. *Microbiology*  
416 142:1107–1114. DOI: 10.1099/13500872-142-5-1107.
- 417 Westcott SL., Schloss PD. 2017. OptiClust, an improved method for assigning  
418 amplicon-based sequence data to operational taxonomic units. *mSphere* 2:e00073–17.  
419 DOI: 10.1128/mspheredirect.00073-17.
- 420 Wickham H. 2017. *Tidyverse: Easily install and load 'tidyverse' packages*.

**Figure 1: Total Number of OTUs in Fecal Samples by Number of Cycles.** The x-axis represents the different number of cycles used and the y-axis is the number of OTUs. The legend represents the color code for the 5 different HiFi DNA polymerases used. The range in the number of OTUs detected in the different fecal samples increased as cycle number increased. This range was also larger for specific HiFi DNA polymerases.

**Figure 2: Total Number of OTUs in Mock Samples by Number of Cycles.** The x-axis represents the different number of cycles used and the y-axis is the number of OTUs. The legend represents the color code for the 5 different HiFi DNA polymerases used. The dotted black line represents the number of OTUs detected when only the references sequences are clustered. The range in the number of OTUs detected in the Mock samples increased as cycle number increased. This range was also larger for specific HiFi DNA polymerases.

**Figure 3: Bray-Curtis Community Differences by Five-Cycle Intervals.** A) within person differences based on the next 5-cycle PCR interval in fecal samples. B) Within replicate difference based on the next 5-cycle PCR interval in Mock samples.

**Figure 4: HiFi DNA Polymerase Error Rate in Mock Samples.** The error bars represent the 75% interquartile range of the median error rate.

**Figure 5: HiFi DNA Polymerase Chimera Prevalence in Mock Samples.** A) Percentage of chimeric sequences without the removal of chimeras with VSEARCH. C) Percentage of chimeric sequences with the removal of chimeras with VSEARCH. C) The total percent of chimeric sequences removed with VSEARCH by cycle number. The error bars represent the 75% interquartile range of the median.

**Figure 6: The Correlation between Number of OTUs and Chimeras in Mock Samples.**

445 **Figure S1: HiFi DNA Polymerase Nucleotide Substitutions in Mock Samples.**