# BEStack 02B Using Batch Effects Interface Corrections: EBNPlus Tod Casasent 2018-04-25-1520

## 1 Using Batch Effects Interface Assessments

This document focuses on explaining the components of the Batch Effects Interface (BEI) involved with creating a job, loading data, and running assessments. This document will not address statistical issues or "how to spot" batch effects.

The URL for your install should be provided to you, but will likely be something like:

http://your-server.your-company.com:9999/BatchEffectsInterface/

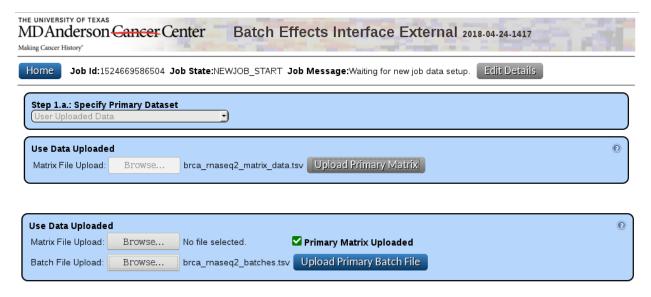
These instructions are aimed at people familiar with R and familiar with TCGA/GDC platforms and data types. They are intended to introduce the reader to producing the given assessment. These instructions will only rarely, if ever, touch on the appropriateness of the assessment algorithm or interpretation of output. See MBatch\_01\_InstallLinux at <a href="https://github.com/MD-Anderson-Bioinformatics/MBatch/tree/master/pdf">https://github.com/MD-Anderson-Bioinformatics/MBatch/tree/master/pdf</a> for instructions on downloading test data.

EBNPlus corrections uses replicates between the two datasets for training and combines the two data sets based on replicates between sets.

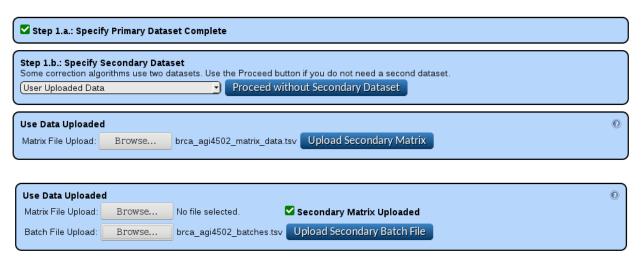
# 2 Starting a Job

See BEStack\_02A\_BEIUsingAssessments for more details about starting a job.

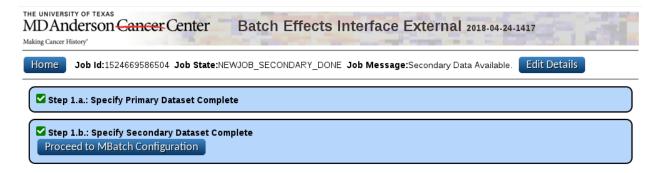
Use the "Start New Job" button and select "User Uploaded Data" for Step 1.a. From within the MATRIX\_DATA.zip archive, upload brca\_rnaseq2\_matrix\_data.tsv as the data matrix and brca\_rnaseq2\_batches.tsv as the batch matrix.



Then for Step 1.b. also select User Uploaded Data, and use brca\_agi4502\_matrix\_data.tsv for the Matrix File and brca\_agi4502\_batches.tsv for the Batch File.



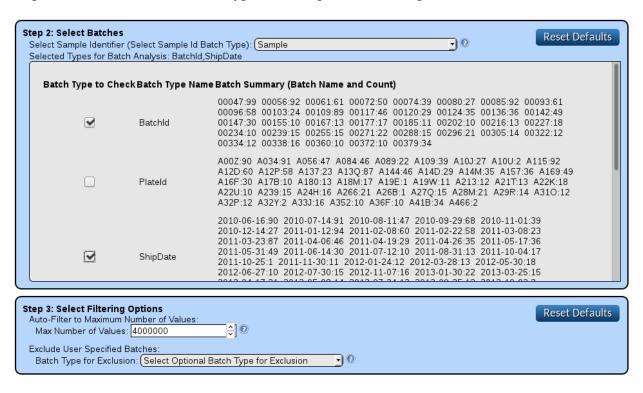
Then select Proceed to MBatch Configuration.



# **3** Configuring Assessments

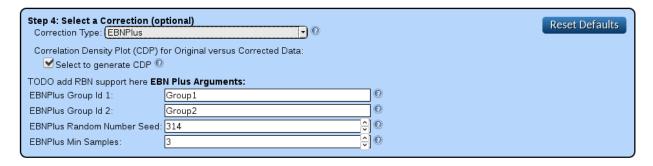
See BEStack\_02A\_BEIUsingAssessmentsExternal for more details about Configuring Assessments.

Below, we have selected Sample as the Sample Identifier, and selected BatchId as well as ShipDate as the assessment batch types. For Step 3, we have kept the defaults.

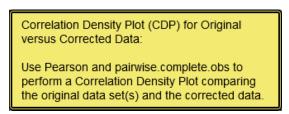


#### 3.1 Step 4

In Step 4, we begin by selecting EBNPlus as the optional Correction Type.



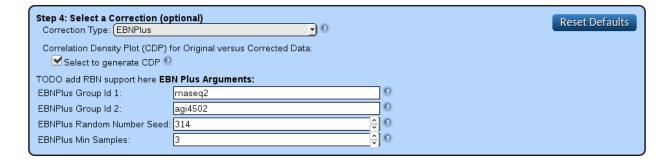
Generating a Correlation Density Plot is selected automatically. (In a future version, this option will appear for any assessment, rather than just for certain correction options.)



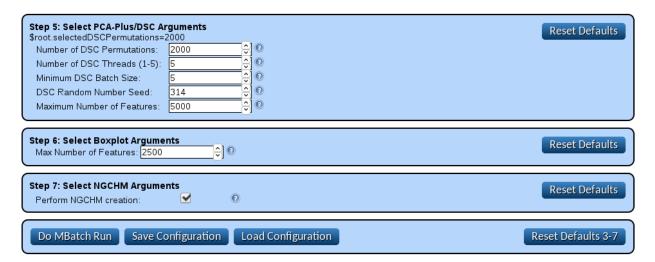
Tooltip Text for Correlation Density Plot

The Group Ids must be alphanumeric values without spaces but with underscores allowed. The random number seed is an integer used as a seed. The minimum number of samples is an integer and means any row (gene) with less than this number of samples is dropped.

Here, we have selected maseq2 as Group Id 1 and agi4502 as Group Id 2.

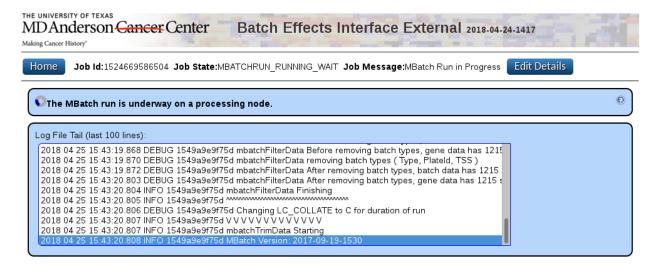


We accept the default values for the rest of the assessment settings, and hit the "Do MBatch Run" button.

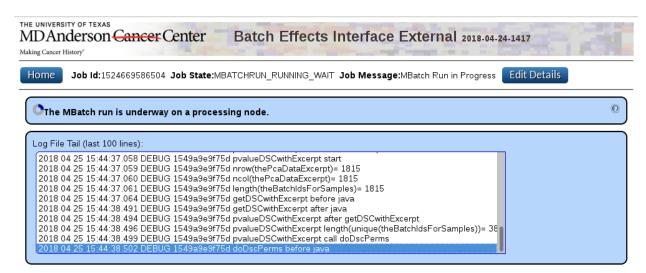


#### 3.2 Do MBatch Run

See BEStack\_02A\_BEIUsingAssessments for more details about running and monitoring a run.

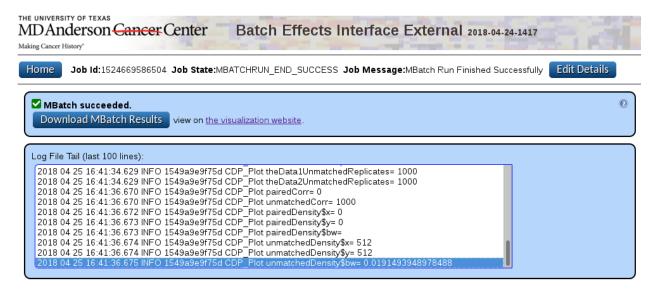


The DSC Permutations step will take some time—30 minutes or more with the full run taking several hours.

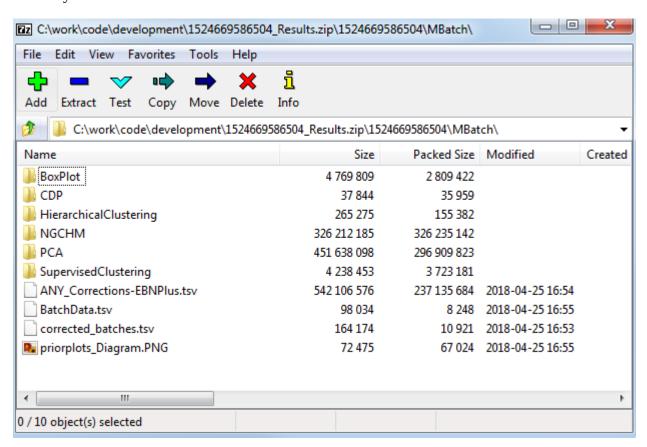


## 3.3 Finished Job

After the job has finished, use the Download option to get the corrected data.



Click the Download MBatch Results button. Open or unzip the archive and enter the MBatch directory.



The ANY\_Corrections-EBNPlus.tsv file contains the corrected data. Looking at an excerpt from that file below, you see the group ids have been added to the end of the sample ids (with a period to separate them).

	TCGA-A1-A0SB-01A-	TCGA-A1-A0SD-01A-	TCGA-A1-A0SD-01A-	TCGA-A1-A0SE-01A-	
	11R-A144-07.rnaseq2	11R-A115-07.agi4502	11R-A115-07.rnaseq2	11R-A084-07.agi4502	
A1BG	2.4365111687448295	4.565452148506589	3.7312694095439234	3.708911067971168	
A2BP1	0.7439255863822336	2.4835734270886842	-1.3208886816920007	-0.6581895449438544	
A2M	8.545488308256441	6.629607302478645	8.137917647366036	7.434465827947289	
A2ML1	0.787232177552283	1.6477352958044498	1.6447986673920711	1.4887545672394458	
A4GALT	4.748263228165531	4.799944822217981	4.886308815099057	4.550275954972794	
A4GNT	0.1884566850556865	0.3153918401926371	-0.06336625921993128	0.17228602035088397	
AAAS	4.452459243976453	4.3829614276156645	3.995812362559631	4.096645218343809	
AACS	6.252403781207319	4.691106228333074	4.818314878643724	5.068015002849296	
AADAC	-2.3617543340468186	0.2346797540826212	-2.3617543340468186	0.1719909368074798	

The corrected\_batches.tsv contains the combined batch files. Looking at an excerpt from that file below, you see the group ids have been added to the end of the sample ids (with a period to separate them).

Sample	Type	BatchId	PlateId	ShipDate	TSS	EBNPlu
						S
TCGA-XX-A899-01A-	1	372	A36F	1/29/2014	XX - Spectrum Health	rnaseq2
11R-A36F-07.rnaseq2						
TCGA-XX-A89A-01A-	1	372	A36F	1/29/2014	XX - Spectrum Health	rnaseq2
11R-A36F-07.rnaseq2						
TCGA-Z7-A8R5-01A-	1	379	A41B	5/28/2014	Z7 - John Wayne	rnaseq2
42R-A41B-07.rnaseq2					Cancer Center	
TCGA-Z7-A8R6-01A-	1	379	A41B	5/28/2014	Z7 - John Wayne	rnaseq2
11R-A41B-07.rnaseq2					Cancer Center	
TCGA-A1-A0SD-01A-	1	85	A115	1/12/2011	A1 - UCSF	agi4502
11R-A115-07.agi4502						
TCGA-A1-A0SE-01A-	1	72	A084	9/29/2010	A1 - UCSF	agi4502
11R-A084-07.agi4502						
TCGA-A1-A0SH-01A-	1	72	A084	9/29/2010	A1 - UCSF	agi4502
11R-A084-07.agi4502						
TCGA-A1-A0SJ-01A-	1	72	A084	9/29/2010	A1 - UCSF	agi4502
11R-A084-07.agi4502						