

Enhanced Sampling Method	CVs	RNA Systems	QM/MM	Total Timescale (μ s)	Ref.
<i>Refinements and Validations of Force Fields</i>					
M-REMD	-	UUCG-TL (10,14-mers), CCCC, GACC	MM	~7663	[1]
M-REMD	-	CCCC, GACC	MM	~920	[2]
H-REMD	-	single strands, duplexes (12-mers)	MM	~1.7	[3]
T-REMD	-	AAAA, CAAU, CCCC, GACC, UUUU	MM	264	[4]
METAD, T-REMD, RECT	$\alpha, \beta, \gamma, \epsilon, \zeta, Z_p, \chi$, distance	CC, AA, CA, AC, GACC, CCCC, AAAA	MM	~206	[5]
METAD, T-REMD, REST2	H-bonds, RMSD	GAGA-TL (8,10-mers)	MM	966	[6]
T-REMD+METAD	ϵ RMSD	GAGA-TL, UUCG-TL (8-mers)	MM	48	[7]
RECT	$\alpha, \beta, \gamma, \epsilon, \zeta, Z_p, \chi$, distance	A, C, AA, AC, CA, CC	MM	~35	[8]
RECT, T-REMD+METAD	$\alpha, \beta, \gamma, \epsilon, \zeta, Z_p, \chi$, distance, ϵ RMSD	GACC, CCCC, AAAA, CAAA, UUCG-TL (8-mer)	MM	~420	[9]
US	$\alpha, \beta, \gamma, \epsilon, \zeta, \chi$	16 dinucleotides	MM	~18	[10]
<i>Conformational Landscapes</i>					
RECT, H-REMD, T-REMD	$\alpha, \beta, \gamma, \delta, \epsilon, \zeta, Z_p, \chi$, distance	GACC	MM	~14.4	[11]
METAD	H-bonds, RGyr, RMSD, χ	GAGA-TL, UUCG-TL (10-mers)	MM	0.84	[12]
T-REMD	-	SAM-II riboswitch	MM	6	[13]
T-REMD, SMD	distances (COM)	(cgauUCUaugc) duplex (22-mer)	MM	~5	[14]
T-REMD	-	SVL loop (17-mer)	MM	~58	[15]
A-REMD	$Z_p, Z_{p'}, \chi$	U	QM/MM	~0.1	[16]
BE-METAD	H-bond, RGyr, energy	gene32 mRNA pseudoknot (32-mer)	MM	3	[17]
US	distance (COM)	add riboswitch	MM	~0.9	[18]
US	χ	duplex, single strands with 2-thiouridines (14-mers)	MM	0.144	[19]
US	χ , pseudodihedral (COM)	duplexes with CUG repeats (9,11,19-mers)	MM	~6	[20]
US - pseudo string method	distance (COM)	duplex (32-mer)	MM	~0.06	[21]
T-REMD	-	GCAA -TL (8-mer)	MM	448	[22]
T-REMD	-	pT181 RNA hairpins (22-mers)	MM	~9	[23]
RAM	H-bonds, distances	TAR (29-mer) in RNA:peptide complex	MM	0.8	[24]
REMD+US	distances, RMSD	U-singlestrand (5-mer) in RNA:protein complex	MM	3.6	[25]
US	distances	GTPase center of rRNA	MM	~1.3	[26]
RAM	$\alpha, \beta, \delta, \epsilon, \zeta$, H-bonds	UUCG-TL (14-mer)	MM	~1	[27]
T-REMD	-	theophylline aptamer (ligand-free)	MM	1.6	[28]
GaMD, TMD	RMSD	CRISPR-Cas9 RNA complex	MM	~14.6	[29]
METAD	distance (COM), stacking	PNA-RNA duplex (12-mer)	MM	0.4	[30]
T-REMD+METAD	H-bonds, RGyr	polio viral RNA hairpin (22-mer)	MM	~6.5	[31]
US	pseudodihedral (COM)	hairpin (35-mer)	MM	0.72	[32]
METAD	χ , pseudodihedral (COM)	duplexes (9,12-mers) with CAG repeats	MM	1.92	[33]
T-REMD	-	gcGCAAg-TL (8-mer)	MM	6	[34]
<i>Ion Interactions and Ion/Ligand Induced Conformational Changes</i>					
METAD	distance (COM), hydrophobic contacts, H-bonds	duplex (10-mer) with CAG repeats	MM	1.4	[35]
US+TI	distance, coordination	mononucleotides, hammerhead ribozyme	MM	~11.5	[36]
SMD	distances	preQ ₁ -III riboswitch	MM	0.1	[37]
BE-METAD	distance, coordination	nucleosides, GpG dinucleotide, GC duplex (8-mer)	MM	82	[38]
GCMC-MD	distances, coordinations	BWVY pseudoknot, VS ribozyme, 23S rRNA, Mg ²⁺ riboswitch	MM	~4	[39]
US	distance (COM)	GTPase center of rRNA	MM	20.5	[40]
TI	distances	guanine riboswitch	MM	~0.6	[41]
<i>Reactivity and Catalysis</i>					
A-REMD	distances	HDV ribozyme	QM/MM	0.235	[42]
US+string method	distances	glmS riboswitch	QM/MM	~0.00017	[43]
T-REMD	-	hammerhead ribozyme	MM	5	[44]
US+string method	distances, angles	HDV ribozyme	MM, QM/MM	0.0465, 0.00019	[45]
US, TI	distances	twister ribozyme	MM	0.5	[46]
US+string method	distances	glmS riboswitch	QM/MM	~0.00017	[47]
TI	distances	group-II introns	QM/MM	~0.00001	[48]
METAD	distances	GAAA-TL, UUCG-TL (10-mers), GC duplex (14-mer)	QM/MM	~0.001	[49]
US+string method	distances	glmS riboswitch	QM/MM	0.0072	[50]
T-REMD	-	hairpin ribozyme	MM	25.6	[51]
US, TI	distances	hammerhead ribozyme	MM, QM/MM	0.11, 0.0002	[52]

References

- [1] Christina Bergonzo, Niel M Henriksen, Daniel R Roe, and Thomas E Cheatham III. Highly sampled tetranucleotide and tetraloop motifs enable evaluation of common RNA force fields. *RNA*, 21(9):1578–1590, 2015.
- [2] Christina Bergonzo and Thomas E Cheatham III. Improved force field parameters lead to a better description of RNA structure. *J. Chem. Theory Comput.*, 11(9):3969–3972, 2015.
- [3] Shun Sakuraba, Kiyoshi Asai, and Tomoshi Kameda. Predicting RNA duplex dimerization free-energy changes upon mutations using molecular dynamics simulations. *J. Phys. Chem. Lett.*, 6(21):4348–4351, 2015.
- [4] Sandro Bottaro, Alejandro Gil-Ley, and Giovanni Bussi. RNA folding pathways in stop motion. *Nucleic Acids Res.*, 44(12):5883–5891, 2016.
- [5] Alejandro Gil-Ley, Sandro Bottaro, and Giovanni Bussi. Empirical corrections to the AMBER RNA force field with target metadynamics. *J. Chem. Theory Comput.*, 12(6):2790–2798, 2016.
- [6] Petra Kührová, Robert B Best, Sandro Bottaro, Giovanni Bussi, Jiří Šponer, Michal Otyepka, and Pavel Banáš. Computer folding of RNA tetraloops: Identification of key force field deficiencies. *J. Chem. Theory Comput.*, 12(9):4534–4548, 2016.
- [7] Sandro Bottaro, Pavel Banáš, Jiří Šponer, and Giovanni Bussi. Free energy landscape of GAGA and UUCG RNA tetraloops. *J. Phys. Chem. Lett.*, 7(20):4032–4038, 2016.
- [8] A Cesari, A Gil-Ley, and G Bussi. Combining simulations and solution experiments as a paradigm for RNA force field refinement. *J. Chem. Theory Comput.*, 12(12):6192–6200, 2016.
- [9] Changwon Yang, Manho Lim, Eunae Kim, and Youngshang Pak. Predicting RNA structures via a simple van der Waals correction to an all-atom force field. *J. Chem. Theory Comput.*, 13(2):395–399, 2017.
- [10] Asaminew H Aytenfisu, Aleksandar Spasic, Alan Grossfield, Harry A Stern, and David H Mathews. Revised RNA dihedral parameters for the AMBER force field improve RNA molecular dynamics. *J. Chem. Theory Comput.*, 13(2):900–915, 2017.
- [11] Alejandro Gil-Ley and Giovanni Bussi. Enhanced conformational sampling using replica exchange with collective-variable tempering. *J. Chem. Theory Comput.*, 11(3):1077–1085, 2015.
- [12] S Haldar, P Kührová, P Banáš, V Spiwok, J Šponer, P Hobza, and M Otyepka. Insights into stability and folding of GNRA and UNGC tetraloops revealed by microsecond molecular dynamics and well-tempered metadynamics. *J. Chem. Theory Comput.*, 11(8):3866, 2015.

- [13] Xu Xue, Wang Yongjun, and Li Zhihong. Folding of SAM-II riboswitch explored by replica-exchange molecular dynamics simulation. *J. Theor. Biol.*, 365:265–269, 2015.
- [14] HaJeung Park, Àlex L González, Ilyas Yildirim, Tuan Tran, Jeremy R Lohman, Pengfei Fang, Min Guo, and Matthew D Disney. Crystallographic and computational analyses of AUUCU repeating RNA that causes Spinocerebellar Ataxia type 10 (SCA10). *Biochemistry*, 54(24):3851, 2015.
- [15] Christina Bergonzo, Kathleen B Hall, and Thomas E Cheatham III. Stem-Loop V of Varkud Satellite RNA exhibits characteristics of the Mg^{2+} bound structure in the presence of monovalent ions. *J. Phys. Chem. B*, 119(38):12355–12364, 2015.
- [16] Brian K Radak, Melissa Romanus, Tai-Sung Lee, Haoyuan Chen, Ming Huang, Antons Treikalis, Vivekanandan Balasubramanian, Shantenu Jha, and Darrin M York. Characterization of the three-dimensional free energy manifold for the uracil ribonucleoside from asynchronous replica exchange simulations. *J. Chem. Theory Comput.*, 11(2):373–377, 2015.
- [17] Yunqiang Bian, Jian Zhang, Jun Wang, Jihua Wang, and Wei Wang. Free energy landscape and multiple folding pathways of an H-type RNA pseudoknot. *PLoS One*, 10(6):e0129089, 2015.
- [18] Francesco Di Palma, Sandro Bottaro, and Giovanni Bussi. Kissing loop interaction in adenine riboswitch: Insights from umbrella sampling simulations. *BMC Bioinformatics*, 16(Suppl 9):S6, 2015.
- [19] Aaron T Larsen, Albert C Fahrenbach, Jia Sheng, Julia Pian, and Jack W Szostak. Thermodynamic insights into 2-thiouridine-enhanced RNA hybridization. *Nucleic Acids Res.*, 43(16):7675–7687, 2015.
- [20] Ilyas Yildirim, Debayan Chakraborty, Matthew D Disney, David J Wales, and George C Schatz. Computational investigation of RNA CUG repeats responsible for myotonic dystrophy 1. *J. Chem. Theory Comput.*, 11(10):4943–4958, 2015.
- [21] Yuan-Yan Wu, Zhong-Liang Zhang, Jin-Si Zhang, Xiao-Long Zhu, and Zhi-Jie Tan. Multivalent ion-mediated nucleic acid helix-helix interactions: RNA versus DNA. *Nucleic Acids Res.*, 43(12):6156–6165, 2015.
- [22] Jacob C Miner, Alan A Chen, and Angel E García. Free-energy landscape of a hyperstable RNA tetraloop. *Proc. Natl. Acad. Sci. U.S.A.*, 113(24):6665–6670, 2016.
- [23] Melissa K Takahashi, Kyle E Watters, Paul M Gasper, Timothy R Abbott, Paul D Carlson, Alan A Chen, and Julius B Lucks. Using in-cell SHAPE-seq and simulations to probe structure–function design principles of RNA transcriptional regulators. *RNA*, 22(6):920–933, 2016.

- [24] Aditi N Borkar, Michael F Bardaro, Carlo Camilloni, Francesco A Aprile, Gabriele Varani, and Michele Vendruscolo. Structure of a low-population binding intermediate in protein–RNA recognition. *Proc. Natl. Acad. Sci. U.S.A.*, 113(26):7171–7176, 2016.
- [25] Lela Vukovic, Christophe Chipot, Debora L Makino, Elena Conti, and Klaus Schulten. Molecular mechanism of processive 3’ to 5’ RNA translocation in the active subunit of the RNA exosome complex. *J. Am. Chem. Soc.*, 138(12):4069–4078, 2016.
- [26] Hamed S Hayatshahi, Christina Bergonzo, and Thomas E Cheatham III. Investigating the ion dependence of the first unfolding step of GTPase-associating center ribosomal RNA. *J. Biomol. Struct. Dyn.*, pages 1–11, 2017.
- [27] Aditi N Borkar, Pramodh Vallurupalli, Carlo Camilloni, Lewis E Kay, and Michele Vendruscolo. Simultaneous NMR characterisation of multiple minima in the free energy landscape of an RNA UUCG tetraloop. *Phys. Chem. Chem. Phys.*, 19(4):2797–2804, 2017.
- [28] Becka M Warfield and Peter C Anderson. Molecular simulations and Markov state modeling reveal the structural diversity and dynamics of a theophylline-binding RNA aptamer in its unbound state. *PloS One*, 12(4):e0176229, 2017.
- [29] Giulia Palermo, Yinglong Miao, Ross C Walker, Martin Jinek, and J Andrew McCammon. CRISPR-Cas9 conformational activation as elucidated from enhanced molecular simulations. *Proc. Natl. Acad. Sci. U.S.A.*, 114(28):7260–7265, 2017.
- [30] Massimiliano Donato Verona, Vincenzo Verdolino, Ferruccio Palazzesi, and Roberto Corradini. Focus on PNA flexibility and RNA binding using molecular dynamics and metadynamics. *Sci. Rep.*, 7:42799, 2017.
- [31] Arup K Pathak and Tusar Bandyopadhyay. Water isotope effect on the thermostability of a polio viral RNA hairpin: A metadynamics study. *J. Chem. Phys.*, 146(16):165104, 2017.
- [32] Zhaoxi Sun, Xiaohui Wang, and John ZH Zhang. Protonation-dependent base flipping in the catalytic triad of a small RNA. *Chem. Phys. Lett.*, 684:239–244, 2017.
- [33] Feng Pan, Viet Hoang Man, Christopher Roland, and Celeste Sagui. Structure and dynamics of DNA and RNA double helices of CAG and GAC trinucleotide repeats. *Biophys. J.*, 113(1):19–36, 2017.
- [34] Jacob C Miner and Angel E García. Equilibrium denaturation and preferential interactions of an RNA tetraloop with urea. *J. Phys. Chem. B*, 121(15):3734–3746, 2017.

- [35] Anna Bochicchio, Giulia Rossetti, Oriana Tabarrini, Sybille Krauß, and Paolo Carloni. Molecular view of ligands specificity for CAG repeats in anti-Huntington therapy. *J. Chem. Theory Comput.*, 11(10):4911–4922, 2015.
- [36] Maria T Panteva, George M Giambasu, and Darrin M York. Comparison of structural, thermodynamic, kinetic and mass transport properties of Mg^{2+} ion models commonly used in biomolecular simulations. *J. Comput. Chem.*, 36(13):970–982, 2015.
- [37] Joseph A Liberman, Krishna C Suddala, Asaminew Aytenfisu, Dalen Chan, Ivan A Belashov, Mohammad Salim, David H Mathews, Robert C Spitale, Nils G Walter, and Joseph E Wedekind. Structural analysis of a class III preQ1 riboswitch reveals an aptamer distant from a ribosome-binding site regulated by fast dynamics. *Proc. Natl. Acad. Sci. U.S.A.*, 112(27):E3485–E3494, 2015.
- [38] Richard A Cunha and Giovanni Bussi. Unraveling Mg^{2+} -RNA binding with atomistic molecular dynamics. *RNA*, 23(5):628–638, 2017.
- [39] Justin A Lemkul, Sirish Kaushik Lakkaraju, and Alexander D MacKerell Jr. Characterization of Mg^{2+} distributions around RNA in solution. *ACS omega*, 1(4):680–688, 2016.
- [40] Hamed S Hayatshahi, Daniel R Roe, Rodrigo Galindo-Murillo, Kathleen B Hall, and Thomas E Cheatham III. Computational assessment of potassium and magnesium ion binding to a buried pocket in GTPase-associating center. *RNA*, 121(3):451–462, 2017.
- [41] Guodong Hu, Aijing Ma, and Jihua Wang. Ligand selectivity mechanism and conformational changes in guanine riboswitch by molecular dynamics simulations and free energy calculations. *J. Chem. Inf. Model.*, 57(4):918–928, 2017.
- [42] Brian K Radak, Tai-Sung Lee, Michael E Harris, and Darrin M York. Assessment of metal-assisted nucleophile activation in the hepatitis delta virus ribozyme from molecular simulation and 3D-RISM. *RNA*, 21(9):1566–1577, 2015.
- [43] Sixue Zhang, Abir Ganguly, Puja Goyal, Jamie L Bingaman, Philip C Bevilacqua, and Sharon Hammes-Schiffer. Role of the active site guanine in the *glms* ribozyme self-cleavage mechanism: Quantum mechanical/molecular mechanical free energy simulations. *J. Am. Chem. Soc.*, 137(2):784–798, 2015.
- [44] Jacob B Swadling, David W Wright, James L Suter, and Peter V Coveney. Structure, dynamics, and function of the hammerhead ribozyme in bulk water and at a clay mineral surface from replica exchange molecular dynamics. *Langmuir*, 31(8):2493–2501, 2015.

- [45] Pallavi Thaplyal, Abir Ganguly, Sharon Hammes-Schiffer, and Philip C Bevilacqua. Inverse thio effects in the hepatitis delta virus ribozyme reveal that the reaction pathway is controlled by metal ion charge density. *Biochemistry*, 54(12):2160–2175, 2015.
- [46] Colin S Gaines and Darrin M York. Ribozyme catalysis with a twist: Active state of the twister ribozyme in solution predicted from molecular simulation. *J. Am. Chem. Soc.*, 138(9):3058–3065, 2016.
- [47] Sixue Zhang, David R Stevens, Puja Goyal, Jamie L Bingaman, Philip C Bevilacqua, and Sharon Hammes-Schiffer. Assessing the potential effects of active site Mg^{2+} ions in the *glms* ribozyme–cofactor complex. *J. Phys. Chem. Lett.*, 7(19):3984–3988, 2016.
- [48] Lorenzo Casalino, Giulia Palermo, Ursula Rothlisberger, and Alessandra Magistrato. Who activates the nucleophile in ribozyme catalysis? An answer from the splicing mechanism of group II introns. *J. Am. Chem. Soc.*, 138(33):10374–10377, 2016.
- [49] Vojtěch Mlýnský and Giovanni Bussi. Understanding in-line probing experiments by modeling cleavage of nonreactive RNA nucleotides. *RNA*, 23(5):712–720, 2017.
- [50] Jamie L Bingaman, Sixue Zhang, David R Stevens, Neela H Yennawar, Sharon Hammes-Schiffer, and Philip C Bevilacqua. The GlcN6P cofactor plays multiple catalytic roles in the *glms* ribozyme. *Nat. Chem. Biol.*, 13(4):439–445, 2017.
- [51] C Schuabb, N Kumar, S Pataraia, D Marx, and R Winter. Pressure modulates the self-cleavage step of the hairpin ribozyme. *Nat. Commun.*, 8:14661, 2017.
- [52] Haoyuan Chen, Timothy J Giese, Barbara L Golden, and Darrin M York. Divalent metal ion activation of a guanine general base in the hammerhead ribozyme: Insights from molecular simulations. *Biochemistry*, 56(24):2985–2994, 2017.