

# Encoding interactions as fingerprints in MD simulations

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# Why?

Interactions are the driving force behind biological processes.

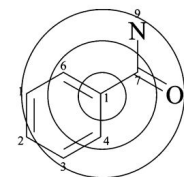
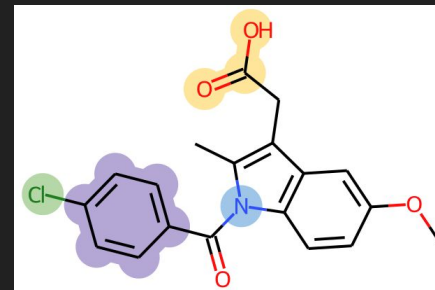
Need to automatically detect and label them, and store them in an appropriate way in order to analyse this data and extract information efficiently.



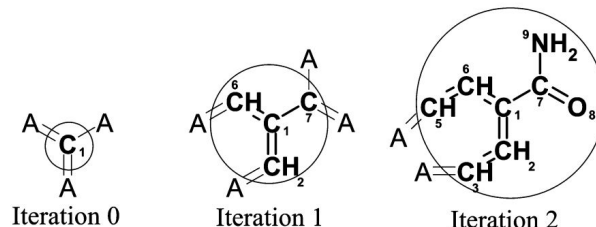
# Interaction Fingerprints

# Fingerprints in cheminformatics

- Compact representation for small molecules
- Encodes structural information as a bitvector
- Each bit defines the presence or absence of a feature (substructure)
- Different ways to define the substructures
  - predefined: substructure keys, e.g. MACCS
  - automatically generated: circular, e.g. ECFP



Considering atom 1 in benzoic acid amide



[Rogers and Hahn \(2010\) J. Chem. Inf. Model.](#)

# Similarity principle

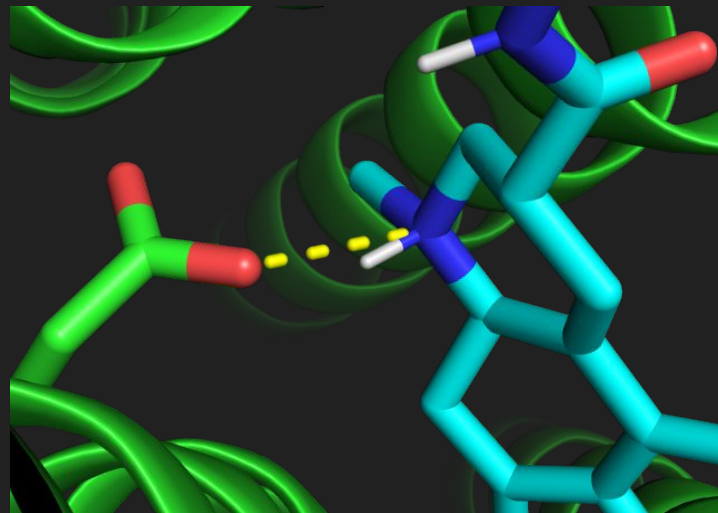
- Structurally similar molecules tend to exhibit similar properties
- Different metrics for similarity between molecules, e.g. Tanimoto

Compound 1	0	1	1	0	0	1	0	1
Compound 2	1	1	0	0	0	1	0	0

$$T_{animoto} = \frac{|A \cap B|}{|A \cup B|} = \frac{|A \cap B|}{|A| + |B| - |A \cap B|} = \frac{2}{4 + 3 - 2} = 0.4$$

# Interaction fingerprints?

- Still encodes structural information as a bitvector
- Bitvector that stores the presence/absence of interactions
- Similarity between complexes or binding modes
- Compounds with similar interaction profiles are likely to exhibit similar properties



# Flavors of IFP

- Structural IFP
- Pharmacophore IFP
- Circular IFP

Interactions usually defined based on **atom selections** and **geometrical constraints**

# Structural IFP

- Each bit position corresponds to an interaction type and residue
- Can only compare IFPs from the same protein family and pocket

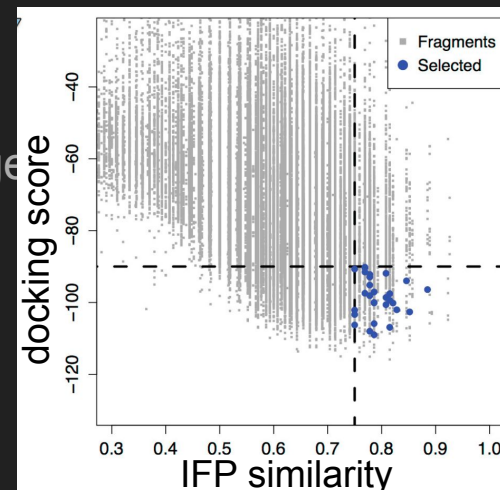
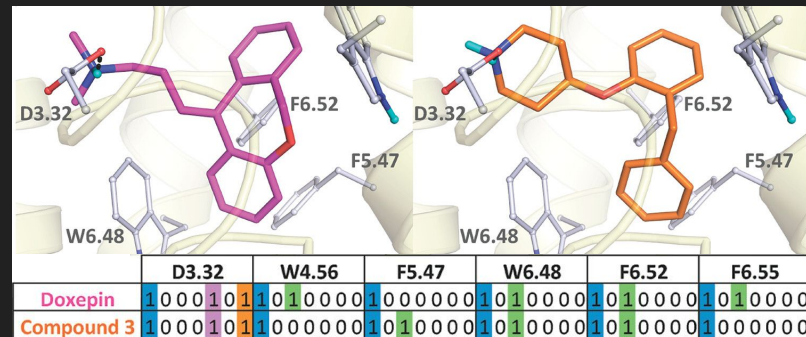
VAL87				ASP114				PHE189				SER193			
Hydrophobic	Acceptor	Donor	PiStacking	Hydrophobic	Acceptor	Donor	PiStacking	Hydrophobic	Acceptor	Donor	PiStacking	Hydrophobic	Acceptor	Donor	PiStacking
1	0	0	0	0	1	0	0	1	0	0	1	0	0	1	0



# Example use cases

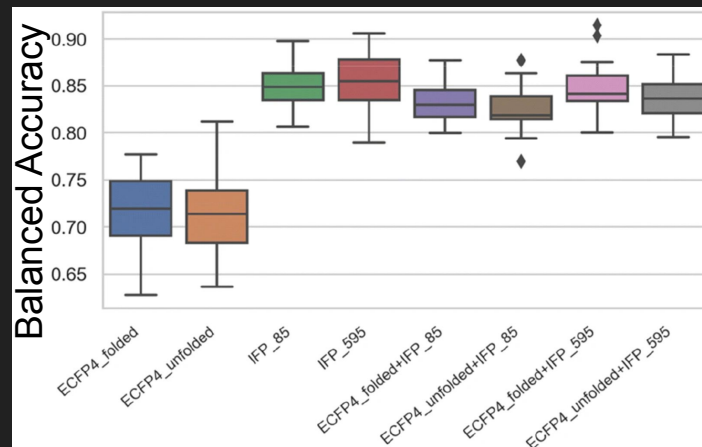
# Filtering docking poses

- Human histamine H1 receptor (H1R) GPCR
- Structure with co-crystallized ligand (**doxepin**)
- Filtered **docking poses** based on
  - Presence of ionic interaction with key residue
  - IFP-based Tanimoto similarity with reference crystal structure above threshold
  - Docking score below threshold
  - ECFP4 Tanimoto similarity with any known H1R ligand above threshold
- 19 of 26 compounds tested had affinities in  $\mu\text{M}$  to nM range



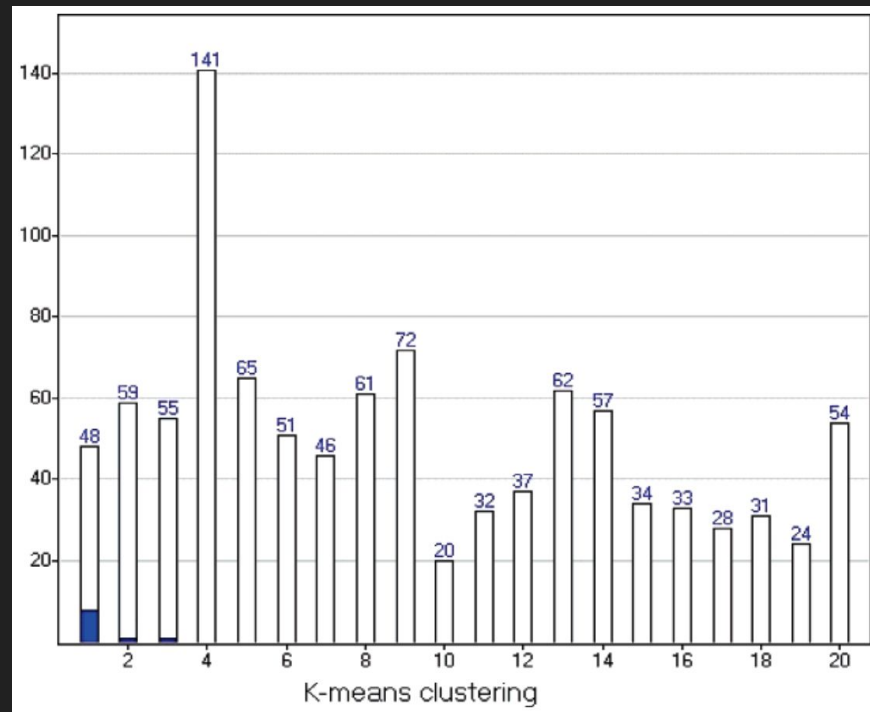
# Representation for supervised machine-learning models

- Kinase inhibitors
- More than 2,000 X-ray structures available
- 3 different binding modes emerge
- Classification task
- IFPs achieved superior predictive performance than 2D-ligand fingerprints (ECFP4)



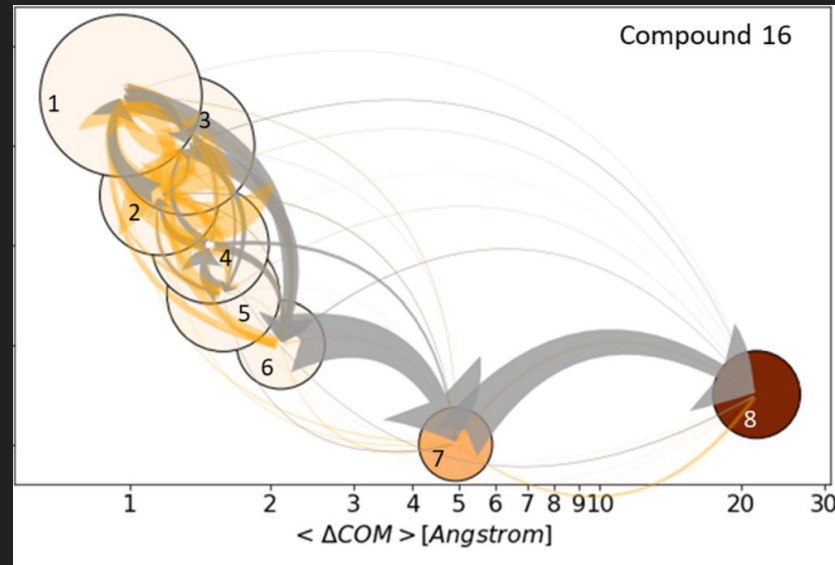
# Extracting binding modes through clustering

- For compound selection in docking
- Cluster all IFPs using k-means
- Include IFPs of active compounds
- Identify which cluster(s) the actives fall into



# Investigate ligand dissociation pathways

- Biased MD protocol (Random Acceleration MD) to analyse unbinding trajectories
- Extract clusters based on IFPs
- Characterization of the dissociation pathway and metastable states by representing the pathway as a graph



# Limitations of existing IFP implementations

- Only specific files can be read
- Only supports protein-ligand interactions
- Not configurable
- Not extensible
- Poor interoperability with other Python packages

# ProLIF

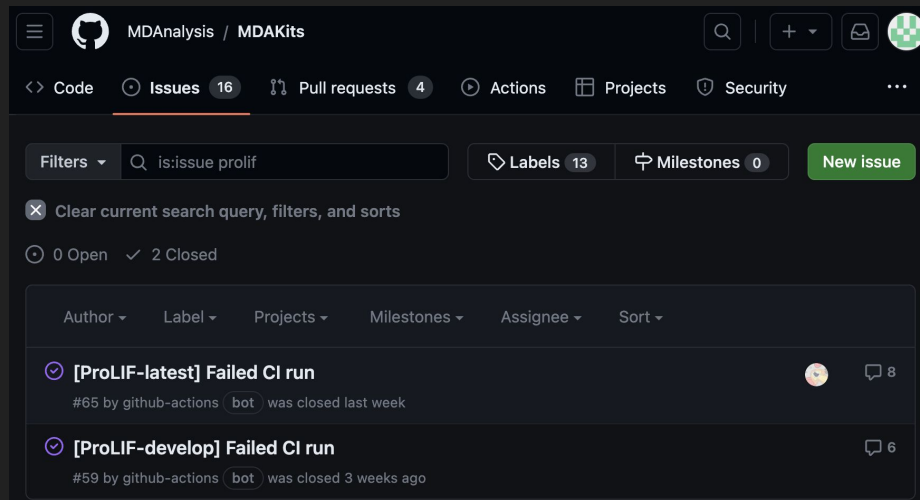


[chemosim-lab/ProLIF](https://github.com/chemosim-lab/ProLIF)

*Bouysset C. and Fiorucci S. (2021) J. Cheminform.*

# ProLIF features

- Almost any 3D file can be read
- Supports any kind of complex: protein-ligand, protein-protein...etc.
- Configurable and extensible
- Exports to other Python objects
- Jupyter-notebook centric
- Part of the MDAKits





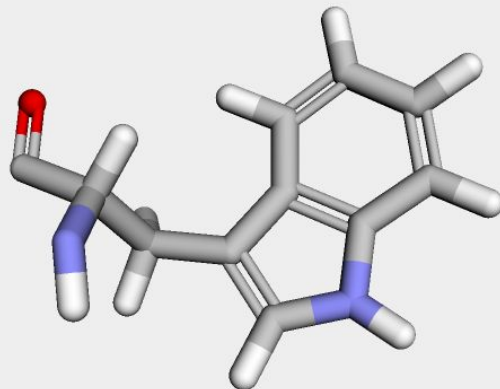
# The implementation

# Reading inputs

- Uses both MDAnalysis and RDKit
- Internally uses RDKit to represent molecules, with a few extras:
  - Fragment parent molecule into its residues
  - Residue are then available as individual molecules by indexing

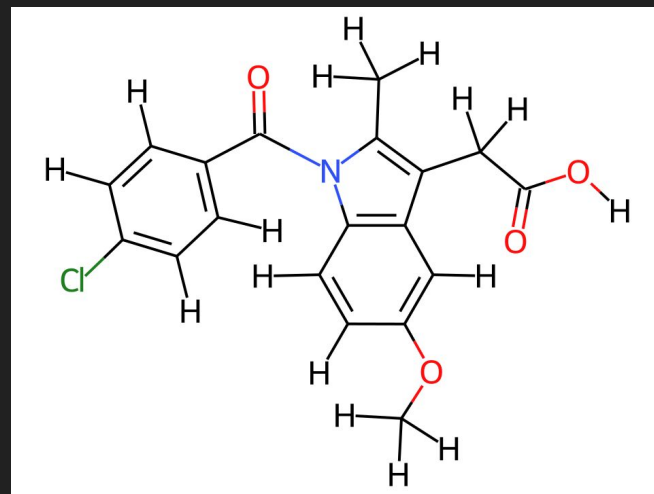
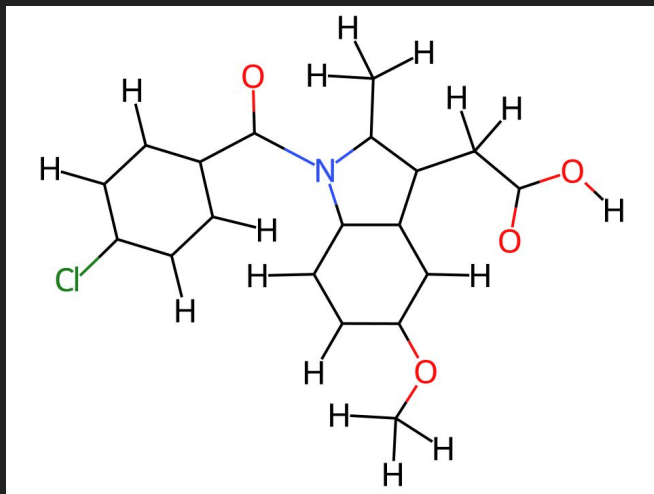
```
import prolif as plf
import MDAnalysis as mda

u = mda.Universe(TOPOLOGY)
protein_selection = u.select_atoms("protein")
protein = plf.Molecule.from_mda(protein_selection)
protein["TRP48.A"]
```



# Reading inputs - MD trajectories and PDB files

- Parse file(s) with MDAnalysis
- Convert to an RDKit mol through the converter interface
  - Bond orders and charges have to be inferred from the topology
- Requires explicit hydrogen atoms



# Fingerprint

```
import prolific as plf
```

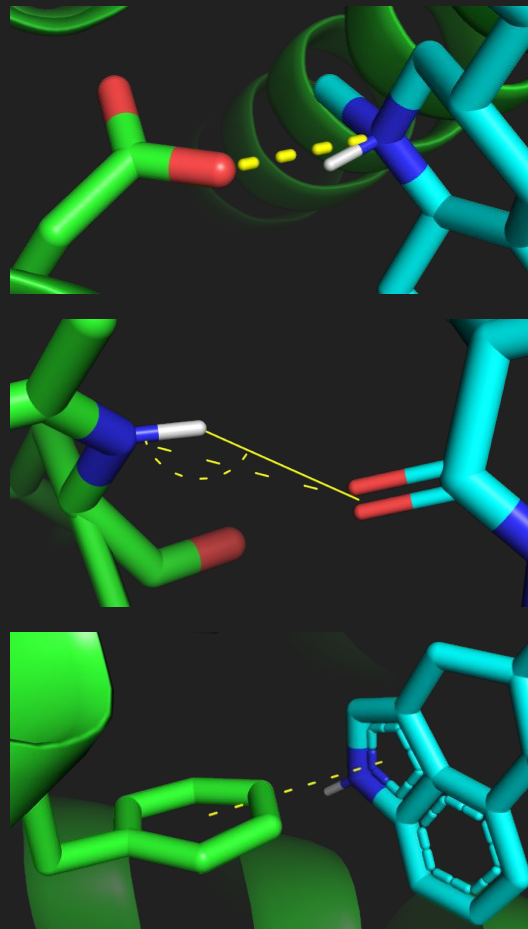
```
fp = plf.Fingerprint(  
    interactions=["HBAcceptor", "HBDonor", "Cationic", "PiStacking"],  
    parameters={"Cationic": {"distance": 4.5}},  
    count=True  
)  
fp.run(u.trajectory[:1], ligand_selection, protein_selection)  
fp.ifp[0][("LIG1.G", "ASP129.A")]
```

100%  1/1 [00:02<00:00, 2.03s/it]

```
{'HBDonor': ({'indices': {'ligand': (13, 52), 'protein': (8,)},  
              'parent_indices': {'ligand': (13, 52), 'protein': (1489,)},  
              'distance': 2.7534162113079534,  
              'DHA_angle': 158.09471525304645},),  
 'Cationic': ({'indices': {'ligand': (13,), 'protein': (8,)},  
               'parent_indices': {'ligand': (13,), 'protein': (1489,)},  
               'distance': 2.7534162113079534},  
               {'indices': {'ligand': (13,), 'protein': (9,)},  
                'parent_indices': {'ligand': (13,), 'protein': (1490,)},  
                'distance': 3.9755628587508895})})
```

# Interactions

- Currently 15 interactions implemented
- 3 base interaction classes:
  - **Distance**
    - Hydrophobic, Cationic, Anionic, MetalDonor, MetalAcceptor
  - **SingleAngle**
    - HBDonor, HBAcceptor
  - **DoubleAngle**
    - XBDonor, XBAcceptor
- Others:
  - PiStacking, EdgeToFace and FaceToFace
  - PiCation, CationPi
  - VdWContact



# Example: defining a new interaction

```
class ExampleInteraction(Interaction):
    def __init__(
        self,
        smarts_acceptor: str = "[#7,#8]",
        smarts_donor: str = "[#7,#8]-[H]",
        distance_threshold: float = 3.5,
    ) → None:
        """
        Validate, transform and store the required parameters.
        """

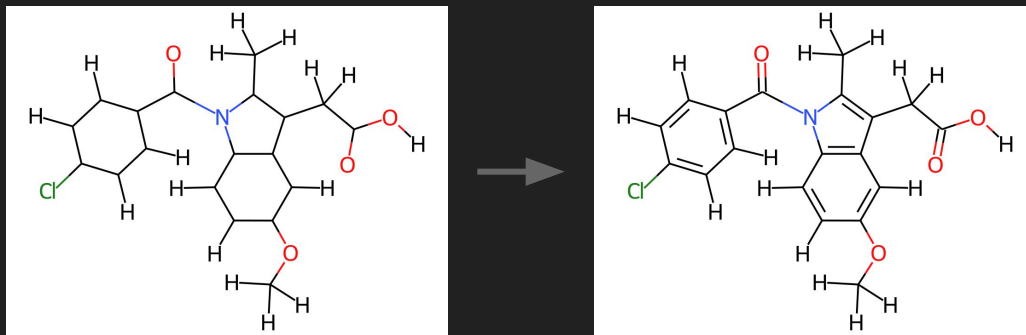
    def detect(
        self, ligand_residue: Residue, protein_residue: Residue
    ) → Iterator[dict]:
        """Implementation of the interaction detection."""
        ...
        if distance ≤ self.distance_threshold:
            yield self.metadata(
                ligand_residue,
                protein_residue,
                ligand_atom_indices,
                protein_atom_indices,
                distance=distance,
            )
```

# Example:

## HBAcceptor

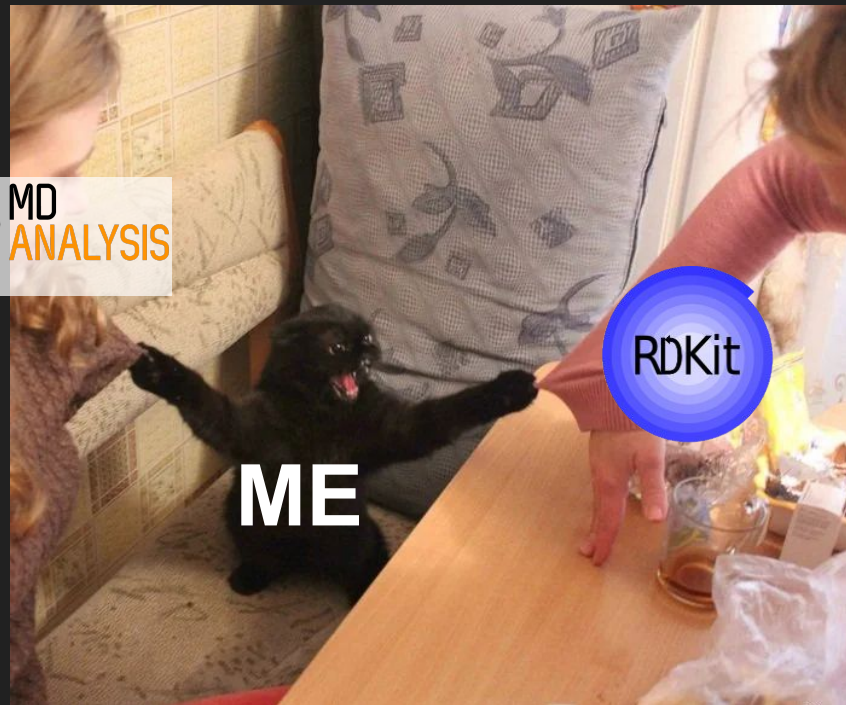
```
class HBAcceptor(SingleAngle):
    def __init__(
        self,
        acceptor: str = (
            "[#7&!$([nX3])&!$([NX3]-*=[O,N,P,S])&!$([NX3]-[a])&!$([Nv4&+1]),"
            "O&!$([OX2](C)C=O)&!$(O(~a)~a)&!$(O=N-*)&!$([O-]-N=O),o+O,"
            "F&$(F-[#6])&!$(F-[#6][F,Cl,Br,I])]"
        ),
        donor: str = "[$([O,S;+0]),$([N;v3,v4&+1]),n+0]-[H]",
        distance: float = 3.5,
        DHA_angle: tuple[int, int] = (130, 180),
    ):
        super().__init__(
            lig_pattern=acceptor,
            prot_pattern=donor,
            distance=distance,
            angle=DHA_angle,
            distance_atom="P1",
            metadata_mapping={"angle": "DHA_angle"},
        )
```

# Interoperability between MDAnalysis and RDKit









# Summary



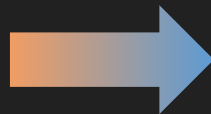
# Motivation

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	Knows chemistry	Reads MD trajectories
RDKit		
MDAnalysis		

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# From the Universe to RDKit



# Requirements

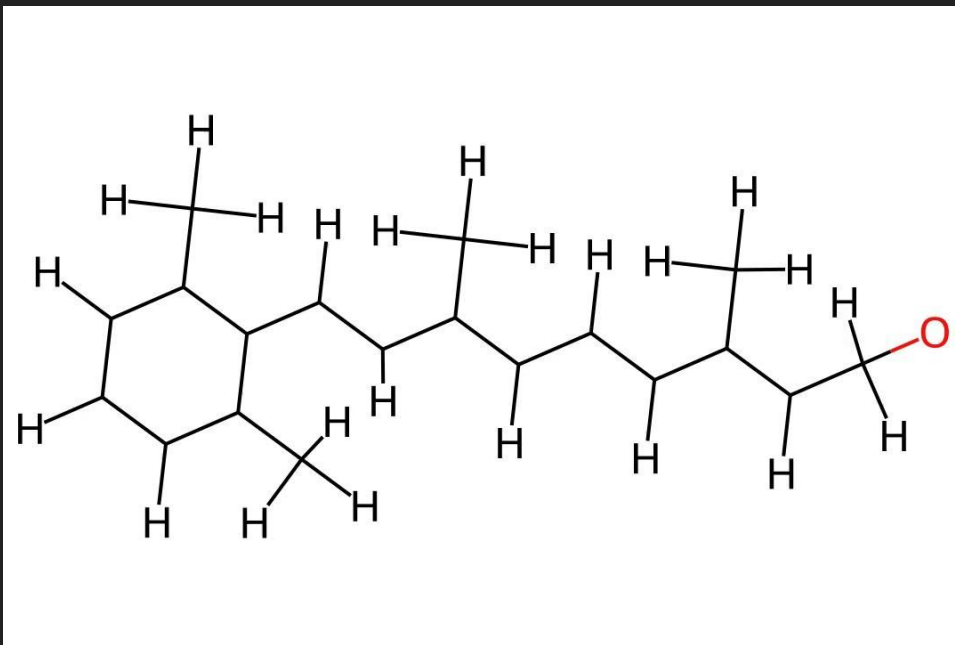
- Elements\*
- Bonds\*
- **Explicit hydrogens**

Infers bond orders and formal charges from there

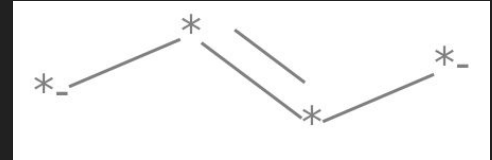
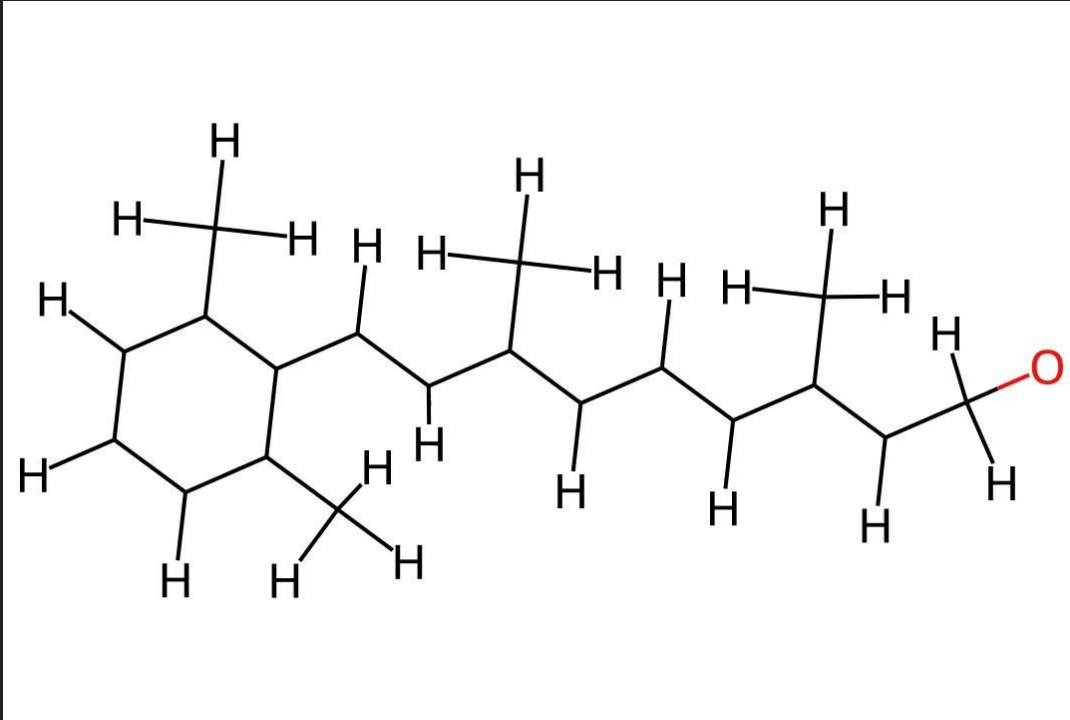
\* can be guessed by MDAnalysis

# Inferring bond orders and charges

```
For each atom:  
  Compare expected vs current valence  
  If all valence electrons are paired:  
    Continue to next atom  
  Else if atom exceeds expected valence:  
    Assign (+) charge  
  Else:  
    If a neighbor can accept a double  
    or triple bond:  
      Increase bond order  
    Else:  
      Assign (-) charge
```



# Order dependency of the algorithm



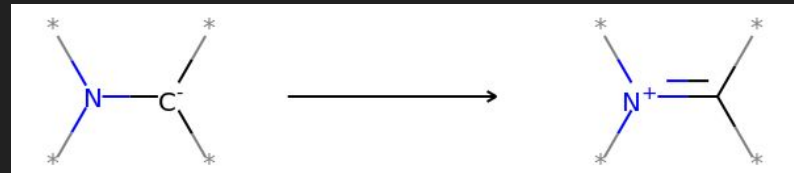
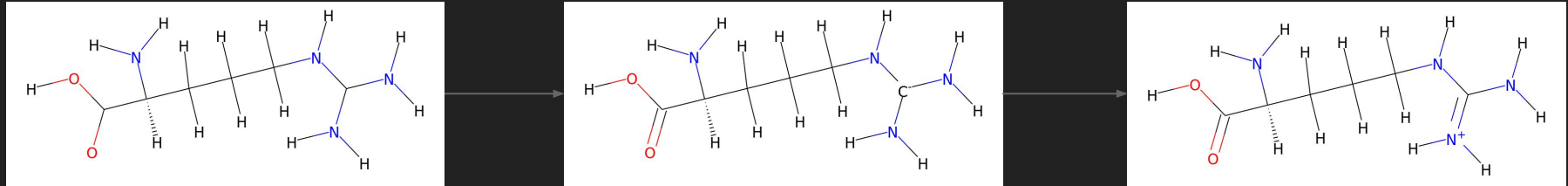
# Logical pitfalls

For each atom:

Compare expected vs current valence

If all valence electrons are paired:

Continue to next atom



# Benchmark

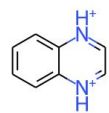
- ChEMBL 33
- 2.2M unique molecules
- Add hydrogens and remove bond orders and charges
- Apply bond order inferring algorithm
- Check whether we match the original molecule / a resonance form
- 99.2% accurate



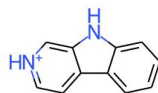
[github.com/MDAnalysis/RDKitConverter-benchmark](https://github.com/MDAnalysis/RDKitConverter-benchmark)



# Common failing scaffolds



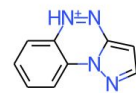
810



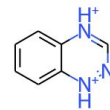
330



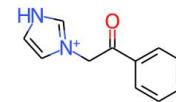
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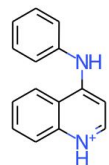
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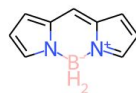
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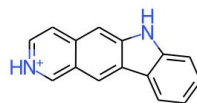
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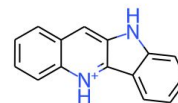
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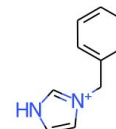
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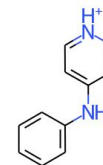
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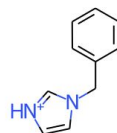
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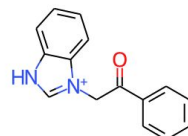
117



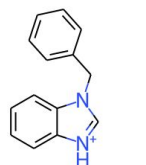
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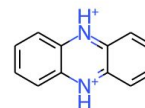
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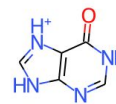
110



91



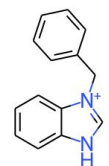
91



88



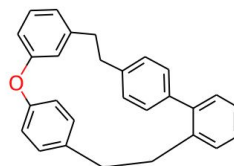
86



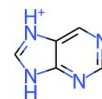
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81



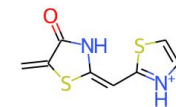
80



76



73

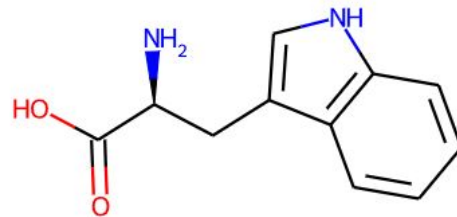


73

# Interoperability features

- Reading from an RDKit molecule
- Converting to an RDKit molecule
- Selection commands
  - aromaticity
  - SMARTS query

```
from rdkit import Chem  
tryptophan = Chem.MolFromSequence("W")  
tryptophan
```



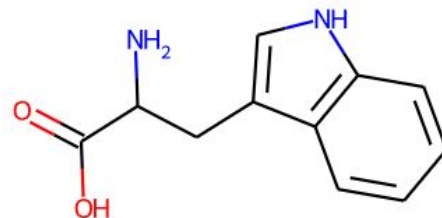
```
import MDAnalysis as mda  
u = mda.Universe(tryptophan)  
u.select_atoms("aromatic and element N")
```

<AtomGroup with 1 atom>

```
u.select_atoms("smarts n")
```

<AtomGroup with 1 atom>

```
u.atoms.convert_to.rdkit()
```



Live demo!

# Conclusion

- Can generate IFPs from MD trajectories and more
- Works with any combination of ligand, protein, DNA or RNA molecules
- Configurable and extensible
- Integrates nicely with the typical Python data-science/cheminformatics stack
- Includes many visualisation options



[prolif.readthedocs.io](https://prolif.readthedocs.io)



[github.com/chemosim-lab/ProLIF](https://github.com/chemosim-lab/ProLIF)

# Future work

## ProLIF

- CLI
- More plots
- Rewrite parts of the code in Cython for performance
- More interaction types
- More fingerprint types

## MDAnalysis RDKitConverter

- Add RDKit's own code for bond order inferring (`rdDetermineBonds`)
- Add RDKit's template matching `AssignBondOrdersFromTemplate`
- Fix problems revealed by the benchmark

# Acknowledgments



## PhD Supervisor

- Sébastien Fiorucci

## ProLIF contributors

- Code: Pavel Polishchuk, René Hamburger
- Conda packaging: Hadrien Mary
- Docs: Irfan Alibay, Muhammad Radifar

## MDAnalysis

- GSoC mentors: Richard Gowers, Irfan Alibay, Fiona Naughton
- RDKitConverter benchmark: Oliver Beckstein, Irfan Alibay
- MDAKits: Irfan Alibay