

codingRMD_MNK

MNK

2024-11-11

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1 Start

1.1 Installing and loading packages

```
if(!requireNamespace("rempsyc")) install.packages("rempsyc"); library(rempsyc)
if(!requireNamespace("flextable")) install.packages("flextable"); library(flextable)
if(!requireNamespace("broom")) install.packages("broom"); library(broom)
if(!requireNamespace("report")) install.packages("report"); library(report)
if(!requireNamespace("effectsize")) install.packages("effectsize"); library(effectsize)
if(!requireNamespace("tinytex")) install.packages("tinytex"); library(tinytex)
if(!requireNamespace("haven")) install.packages("haven"); library(haven)
if(!requireNamespace("dplyr")) install.packages("dplyr"); library(dplyr)
if(!requireNamespace("visdat")) install.packages("visdat"); library(visdat)
if(!requireNamespace("naniar")) install.packages("naniar"); library(naniar)
if(!requireNamespace("psych")) install.packages("psych"); library(psych)
if(!requireNamespace("mice")) install.packages("mice"); library(mice)
if(!requireNamespace("Hmisc")) install.packages("Hmisc"); library(Hmisc)
if(!requireNamespace("knitr")) install.packages("knitr"); library(knitr)
if(!requireNamespace("kableExtra")) install.packages("kableExtra"); library(kableExtra)
if(!requireNamespace("lavaan")) install.packages("lavaan"); library(lavaan)
if(!requireNamespace("semPlot")) install.packages("semPlot"); library(semPlot)
if(!requireNamespace("rstatix")) install.packages("rstatix"); library(rstatix)
```

1.2 Loading datafiles

1.2.1 EMC data

```
setwd("V:/Research/Dementie/Studenten/Studenten/Max/Databeheer")
dataEMC <- haven::read_sav("data_SCTQ_merged_23082024.sav") #load EMC collected data
dataEMC[dataEMC == 999] <- NA #set missing values to NA
dataEMC_copy <- dataEMC #make copy of dataset
```

1.2.2 UMCG data

```
setwd("V:/Research/Dementie/Studenten/Studenten/Max/Databeheer")
dataUMCG <- haven::read_sav("data_SC_UMCG_mnk_errorAdj.sav") #load UMCG collected data
dataUMCG[dataUMCG == 999] <- NA #set missing values to NA
dataUMCG_copy <- dataUMCG #make copy of dataset
```

2 Methods: Data preprocessing

2.1 Pre defined transformation

```
#change direction; higher scores better perception
dataEMC$TAS20_fac1_tf <- 35 - dataEMC$TAS20_fac1_Identifieren_Gevoelens
dataUMCG$TAS20_fac1_tf <- 35 - dataUMCG$TAS20_fac1_Identifieren_Gevoelens
```

2.2 Synchronizing names and compute totals

```
#adding total scores for FP subset to EMC data
dataEMC$FP_1_6_total <- dataEMC$FP_1t6_ToM + dataEMC$FP_1t6_empathy

#realising identical column names
colnames(dataUMCG)[which(colnames(dataUMCG) == "SET_UMCG_Totaal")] <- "SET_UMCG_total"

#calculating subscores of SET subset to EMC data
dataEMC$SET_UMCG_Cognitief_Totaal <- dataEMC$SET_UMCG_1 + dataEMC$SET_UMCG_2 + dataEMC$SET_UMCG_3
dataEMC$SET_UMCG_Affectief_Totaal <- dataEMC$SET_UMCG_4 + dataEMC$SET_UMCG_5 + dataEMC$SET_UMCG_6
```

2.3 Creating ID values for UMCG sample

```
dataUMCG$ID <- paste0("UMCG", seq(1, nrow(dataUMCG)))
```

2.4 Data pooling

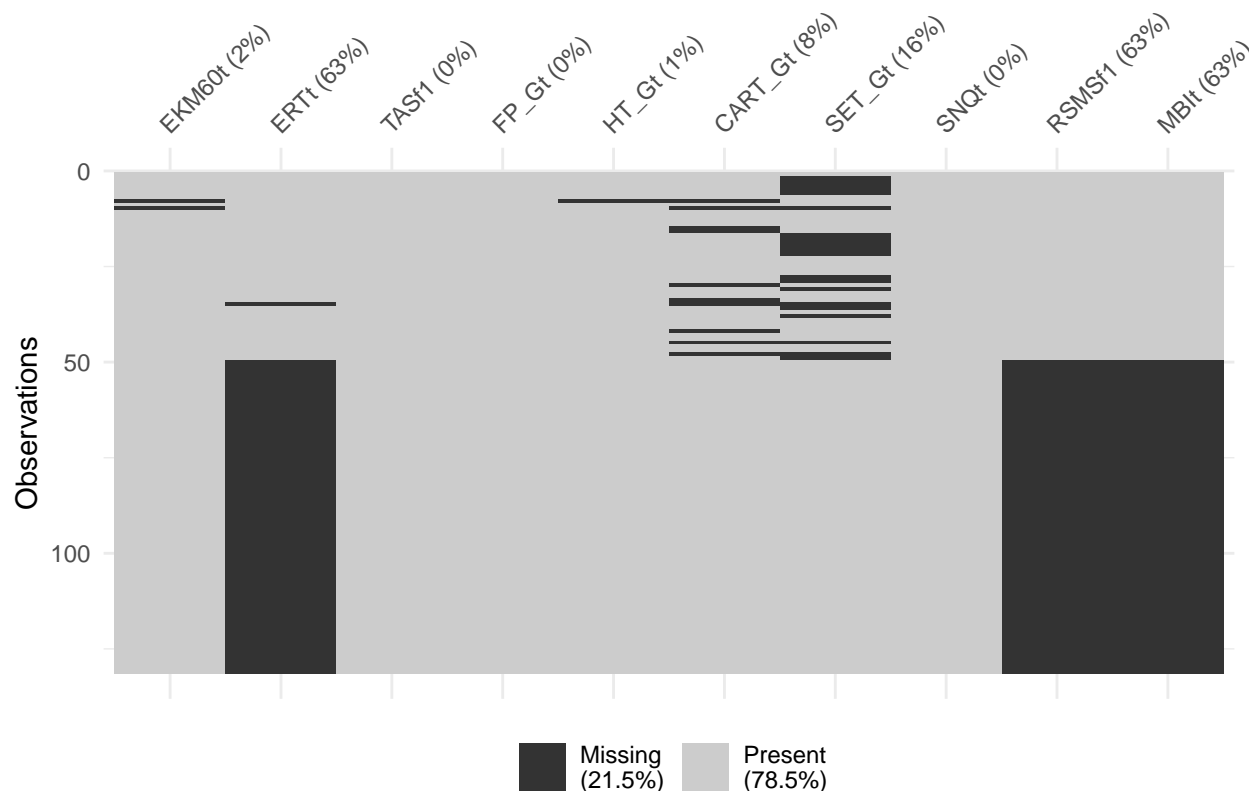
```
data_pooled <- dplyr::bind_rows(dataEMC, dataUMCG) #pooling the data
data_pooled_copy <- data_pooled #make a copy of pooled dataset
```

2.5 Renaming and var name sets

```
names_relevant_items <- c("Ekman60_total", "ERT_total", "TAS20_fac1_tf",
                          "FP_1_6_total", "hintingtask_total", "cartoons_total", "SET_UMCG_total",
                          "SNQ_NL_total", "RSMS_AMSP", "MBI_total")
names_rel_items_abbr <- c('EKM60t', 'ERTt', 'TASf1',
                          'FP_Gt', 'HT_Gt', 'CART_Gt', 'SET_Gt',
                          'SNQt', 'RSMSf1', 'MBIt')
names(data_pooled)[match(names_relevant_items, names(data_pooled))] <- names_rel_items_abbr #renaming
names_covariates <- c("age", "sex", "education_level")
```

2.6 Missing values part 1; visualisation

```
vis_miss(data_pooled[,names_rel_items_abbr])
```



15% of data is missing for SET_Gt, all these missings are in the EMC data. For these people with missings, SET was conducted with another testing protocol (solely multiple choice question, no open questions). Below, we will impute SET_Gt scores for these people based on means in the EMC sample corrected for their scores on the MC questions.

2.6.1 Imputing the SET_Gt variable some EMC missings due to different testing protocol

```
#imputing SET_Gt open question scores based on means in UMG sample conditioned on
#correctness of corresponding MC question.
data_pooled[data_pooled$centerID == "EMC" & is.na(data_pooled$SET_open_2), "SET_open_2"] <-
  as.numeric(colMeans(data_pooled[data_pooled$centerID == "EMC", "SET_open_2"], na.rm = TRUE))
data_pooled[data_pooled$centerID == "EMC" & is.na(data_pooled$SET_open_3), "SET_open_3"] <-
  as.numeric(colMeans(data_pooled[data_pooled$centerID == "EMC", "SET_open_3"], na.rm = TRUE))
data_pooled[data_pooled$centerID == "EMC" & is.na(data_pooled$SET_open_6), "SET_open_6"] <-
  as.numeric(colMeans(data_pooled[data_pooled$centerID == "EMC", "SET_open_6"], na.rm = TRUE))
data_pooled[data_pooled$centerID == "EMC" & is.na(data_pooled$SET_open_14), "SET_open_14"] <-
  as.numeric(colMeans(data_pooled[data_pooled$centerID == "EMC", "SET_open_14"], na.rm = TRUE))
data_pooled[data_pooled$centerID == "EMC" & is.na(data_pooled$SET_open_16), "SET_open_16"] <-
  as.numeric(colMeans(data_pooled[data_pooled$centerID == "EMC", "SET_open_16"], na.rm = TRUE))
data_pooled[data_pooled$centerID == "EMC" & is.na(data_pooled$SET_open_17), "SET_open_17"] <-
  as.numeric(colMeans(data_pooled[data_pooled$centerID == "EMC", "SET_open_17"], na.rm = TRUE))
```

```

data_pooled[data_pooled$centerID == "EMC", "SET_UMCG_1"] <-
  data_pooled[data_pooled$centerID == "EMC",]$SET_open_2 +
  data_pooled[data_pooled$centerID == "EMC",]$SET_mc_2
data_pooled[data_pooled$centerID == "EMC", "SET_UMCG_2"] <-
  data_pooled[data_pooled$centerID == "EMC",]$SET_open_3 +
  data_pooled[data_pooled$centerID == "EMC",]$SET_mc_3
data_pooled[data_pooled$centerID == "EMC", "SET_UMCG_3"] <-
  data_pooled[data_pooled$centerID == "EMC",]$SET_open_6 +
  data_pooled[data_pooled$centerID == "EMC",]$SET_mc_6
data_pooled[data_pooled$centerID == "EMC", "SET_UMCG_4"] <-
  data_pooled[data_pooled$centerID == "EMC",]$SET_open_14 +
  data_pooled[data_pooled$centerID == "EMC",]$SET_mc_14
data_pooled[data_pooled$centerID == "EMC", "SET_UMCG_5"] <-
  data_pooled[data_pooled$centerID == "EMC",]$SET_open_16 +
  data_pooled[data_pooled$centerID == "EMC",]$SET_mc_16
data_pooled[data_pooled$centerID == "EMC", "SET_UMCG_6"] <-
  data_pooled[data_pooled$centerID == "EMC",]$SET_open_17 +
  data_pooled[data_pooled$centerID == "EMC",]$SET_mc_17

data_pooled$SET_Gt <- rowSums(data_pooled[,c(which(names(data_pooled)=="SET_UMCG_1")
      : which(names(data_pooled)=="SET_UMCG_6"))])

```

In the above plot it is visible that we miss $\geq 63\%$ of data for four items (ERT, SET, RSMS, MBI). These items were not included in the data collection of UMCG, and therefore we assume that these missing values are missing completely at random (aka the characteristics of these participants did not influence whether this data is present/absent). Later on, data for the UMCG sample will be imputed for those four items using stochastic imputation (predicted mean matching).

The 8% missing values for the CART_Gt variable can probably be explained by lack of time (slower participants) because it was the last test in the EMC protocol. We will impute the scores using stochastic imputation (predicted mean matching).

Imputation will be continued after checks for outliers and non-normality.

3 Methods: Outlier and normality checks

3.1 Descriptives (before outlier deletion and/or transformations)

3.1.1 EMC sample

```

# Descriptives EMC before outlier deletion and/or transformations
psych::describe(data_pooled[data_pooled$centerID == "EMC",
  c(names_covariates,
    "MoCA_total",
    names_rel_items_abbr)]) %>%
  select(n, min, max, mean, median, sd, skew, kurtosis)

```

##		n	min	max	mean	median	sd	skew	kurtosis
##	age	49	21.0	79	48.10	54.00	19.28	-0.12	-1.47
##	sex	49	0.0	1	0.65	1.00	0.48	-0.62	-1.64

```
## education_level 49 2.0 7 5.71 6.00 1.12 -1.01 1.11
## MoCA_total      49 19.0 30 25.96 26.00 2.60 -0.72 0.10
## EKM60t         47 34.0 58 46.96 47.00 5.32 -0.47 0.13
## ERTt           48 34.0 76 56.85 57.50 9.67 -0.19 -0.65
## TASf1          49 5.0 28 20.08 21.00 5.37 -0.62 0.02
## FP_Gt          49 10.5 24 20.14 20.50 2.51 -1.07 2.30
## HT_Gt          48 6.0 12 11.21 11.50 1.11 -2.32 7.76
## CART_Gt        39 2.5 12 8.10 8.00 2.67 -0.30 -0.91
## SET_Gt         46 8.0 12 11.33 11.45 0.86 -1.91 3.88
## SNQt           49 11.0 21 19.06 19.00 1.63 -2.49 9.95
## RSMSf1         49 11.0 32 22.61 23.00 4.68 -0.51 -0.15
## MBI_t          49 42.0 70 54.43 55.00 7.08 0.16 -0.69
```

3.1.2 UMCG sample

```
# Descriptives UMCG before outlier deletion and/or tranformations
psych::describe(data_pooled[data_pooled$centerID == "UMCG",
                             c(names_covariates,
                                "MoCA_total",
                                names_rel_items_abbrev))] %>%
  select(n, min, max, mean, median, sd, skew, kurtosis)
```

```
## Warning in FUN(newX[, i], ...): no non-missing arguments to min; returning Inf
## Warning in FUN(newX[, i], ...): no non-missing arguments to min; returning Inf
## Warning in FUN(newX[, i], ...): no non-missing arguments to min; returning Inf
## Warning in FUN(newX[, i], ...): no non-missing arguments to min; returning Inf
```

```
## Warning in FUN(newX[, i], ...): no non-missing arguments to max; returning -Inf
## Warning in FUN(newX[, i], ...): no non-missing arguments to max; returning -Inf
## Warning in FUN(newX[, i], ...): no non-missing arguments to max; returning -Inf
## Warning in FUN(newX[, i], ...): no non-missing arguments to max; returning -Inf
```

```
##           n min  max mean median    sd skew kurtosis
## age       82 18   66 28.71    22 14.07  1.36    0.16
## sex       82  0    1  0.63     1  0.48 -0.55   -1.72
## education_level 82  3    7  6.00     6  0.61 -1.30    6.07
## MoCA_total    0 Inf -Inf   NaN    NA    NA    NA     NA
## EKM60t       82 38   56 46.99    48  4.52 -0.26   -0.88
## ERTt         0 Inf -Inf   NaN    NA    NA    NA     NA
## TASf1       82  7   28 21.00    21  4.50 -0.80    0.58
## FP_Gt       82 13   24 20.17    21  2.51 -0.81    0.21
## HT_Gt       82  6   12 11.40    12  1.00 -2.66    9.62
## CART_Gt     82  0   12  8.51     9  2.51 -0.49   -0.05
## SET_Gt      82  8   12 11.33    12  0.94 -1.38    1.28
## SNQt        82 16   22 19.38    19  1.27 -0.40   -0.21
## RSMSf1       0 Inf -Inf   NaN    NA    NA    NA     NA
## MBI_t        0 Inf -Inf   NaN    NA    NA    NA     NA
```

3.1.3 Pooled

```
# Descriptives pooled before outlier deletion and/or tranformations
psych::describe(data_pooled[,
                      c(names_covariates,
                        "MoCA_total",
                        names_rel_items_abbr)]) %>%
  select(n, min, max, mean, median, sd, skew, kurtosis)
```

```
##           n  min max  mean median   sd  skew kurtosis
## age       131 18.0  79 35.96   24.0 18.69  0.73   -1.02
## sex       131  0.0   1  0.64    1.0  0.48 -0.58   -1.67
## education_level 131  2.0   7  5.89    6.0  0.84 -1.48    4.09
## MoCA_total    49 19.0  30 25.96   26.0  2.60 -0.72    0.10
## EKM60t       129 34.0  58 46.98   48.0  4.81 -0.37   -0.23
## ERTt         48 34.0  76 56.85   57.5  9.67 -0.19   -0.65
## TASf1        131  5.0  28 20.66   21.0  4.84 -0.77    0.47
## FP_Gt        131 10.5  24 20.16   20.5  2.50 -0.91    1.05
## HT_Gt        130  6.0  12 11.33   12.0  1.04 -2.55    8.99
## CART_Gt      121  0.0  12  8.38    9.0  2.56 -0.44   -0.33
## SET_Gt       128  8.0  12 11.33   12.0  0.91 -1.56    2.13
## SNQt        131 11.0  22 19.26   19.0  1.42 -1.67    7.31
## RSMSf1       49 11.0  32 22.61   23.0  4.68 -0.51   -0.15
## MBit         49 42.0  70 54.43   55.0  7.08  0.16   -0.69
```

3.2 Outliers

3.2.1 EMC sample

The following code will check whether there are participants with an absolute scaled mean indicator (ASMI) Z-score above 3 solely based on EMC sample.

```
#calculating Z-scores
dataEMC_subset_scaled <- cbind(data_pooled[data_pooled$centerID == "EMC",]
                                [, c("ID", names_covariates, "MoCA_total")],
                                scale(data_pooled[data_pooled$centerID == "EMC",]
                                      [,names_rel_items_abbr]))

#check if MoCA score is below 26
moca_below26_EMC <- dataEMC_subset_scaled$MoCA_total < 26

#calculating mean Z-score of all items per participant
meanitemSS_EMC <- rowMeans(dataEMC_subset_scaled[,c(
  'EKM60t', 'ERTt', 'TASf1',
  'FP_Gt', 'CART_Gt', 'SET_Gt',
  'SNQt', 'RSMSf1', 'MBit')], na.rm = TRUE)

#scaling the vector of mean Z-scores of all items per participant
SSmeanitemSS_EMC <- scale(meanitemSS_EMC)

# #check if ASMI Z-score > 3
outlier_EMC <- dataEMC_subset_scaled[which(abs(SSmeanitemSS_EMC)>3), "ID"]
length(outlier_EMC)
```



```
## [1] 1
```

The detection method identified 'r length(outlier_UMCG)' outlier within the EMC sample.

3.2.2 UMCG sample

The following code will check whether there are participants with an absolute scaled mean indicator (ASMI) Z-score above 3 solely based on UMCG sample.

```
#calculating Z-scores
dataUMCG_subset_scaled <- cbind(data_pooled[data_pooled$centerID == "UMCG",]
                                [, c("ID", names_covariates, "MoCA_total")],
                                scale(data_pooled[data_pooled$centerID == "UMCG",]
                                      [,names_rel_items_abbr]))

#check if MoCA score is below 26
moca_below26_UMCG <- dataUMCG_subset_scaled$MoCA_total < 26

#calculating mean Z-score of all items per participant
meanitemSS_UMCG <- rowMeans(dataUMCG_subset_scaled[,c(
  'EKM60t', 'ERTt', 'TASf1',
  'FP_Gt', 'CART_Gt', 'SET_Gt',
  'SNQt', 'RSMSf1', 'MBIt')], na.rm = TRUE)

#scaling the vector of mean Z-scores of all items per participant
SSmeanitemSS_UMCG <- scale(meanitemSS_UMCG)

# #check if ASMI Z-score > 3
outlier_UMCG <- dataUMCG_subset_scaled[which(abs(SSmeanitemSS_UMCG)>3), "ID"]
length(outlier_UMCG)
```

```
## [1] 1
```

The detection method identified 'r length(outlier_UMCG)' outlier within the EMC sample.

3.2.3 Pooled sample

The following code will check whether there are participants with an absolute scaled mean indicator (ASMI) Z-score above 3 based on the pooled.

```
#calculating Z-scores
datapld_subset_scaled <- cbind(data_pooled[, c("ID",
                                                names_covariates,
                                                "MoCA_total")],
                              scale(data_pooled[,names_rel_items_abbr]))

#check if MoCA score is below 26
moca_below26_pld <- datapld_subset_scaled$MoCA_total < 26

#calculating mean Z-score of all items per participant
meanitemSS_pld <- rowMeans(datapld_subset_scaled[,c(
```

```

'EKM60t', 'ERTt', 'TASf1',
'FP_Gt', 'CART_Gt', 'SET_Gt',
'SNQ', 'RSMSf1', 'MBIt')], na.rm = TRUE)

#scaling the vector of mean Z-scores of all items per participant
SSmeanitemSS_pld <- scale(meanitemSS_pld)

# #check if ASMI Z-score > 3
outlier_pld <- datapld_subset_scaled[which(abs(SSmeanitemSS_pld)>3), "ID"]
length(outlier_pld)

## [1] 2

```

The detection method identified 'r length(outlier_pld)' outliers within the pooled sample.

The following code will check whether the 'r length(outlier_pld)' outliers detected in the pooled sample correspond to the outliers detected separately in the EMC sample and UMCg sample

```

c(outlier EMC, outlier UMCg) == outlier_pld

## [1] TRUE TRUE

```

It does. These two outliers are considered as 'not belonging to the intended population of interest' and will be deleted from the analysis dataset.

```

#deleting the outlier from the pooled-dataset
data_pooled <- data_pooled[-(which(data_pooled$ID==outlier EMC)),]
data_pooled <- data_pooled[-(which(data_pooled$ID==outlier UMCg)),]

```

3.3 Normality

3.3.1 Normality - EMC sample

```

psych::describe(data_pooled[data_pooled$centerID == "EMC",
                             c(names_covariates, "MoCA_total",
                               names_rel_items_abbrev)]) %>%
  select(n, min, max, mean, median, sd, skew, kurtosis)

```

##		n	min	max	mean	median	sd	skew	kurtosis
##	age	48	21.0	79	47.60	52.50	19.16	-0.09	-1.46
##	sex	48	0.0	1	0.65	1.00	0.48	-0.59	-1.68
##	education_level	48	2.0	7	5.75	6.00	1.10	-1.10	1.48
##	MoCA_total	48	19.0	30	26.10	26.50	2.42	-0.59	-0.08
##	EKM60t	46	34.0	58	47.24	47.50	5.02	-0.37	0.15
##	ERTt	47	34.0	76	57.21	58.00	9.44	-0.20	-0.55
##	TASf1	48	5.0	28	20.08	21.00	5.43	-0.62	-0.04
##	FP_Gt	48	10.5	24	20.23	20.50	2.46	-1.16	2.83
##	HT_Gt	47	6.0	12	11.26	12.00	1.07	-2.57	9.65
##	CART_Gt	39	2.5	12	8.10	8.00	2.67	-0.30	-0.91
##	SET_Gt	45	9.0	12	11.40	11.45	0.71	-1.52	2.17

```
## SNQt          48 16.0  21 19.23  19.00  1.13 -0.44  -0.28
## RSMSf1        48 11.0  32 22.75  23.00  4.62 -0.57   0.02
## MBI_t         48 42.0  70 54.29  54.50  7.09  0.21  -0.66
```

```
normalityCheck_EMCC <- psych::describe(data_pooled[data_pooled$centerID == "EMCC",
                                                    c(names_covariates, "MoCA_total",
                                                      names_rel_items_abbrev)]) %>%
  select(skew, kurtosis)

any(abs(normalityCheck_EMCC$skew)>3)
```

```
## [1] FALSE
```

```
any(abs(normalityCheck_EMCC$kurtosis)>10)
```

```
## [1] FALSE
```

No absolute skew>3 and no absolute kurtosis>10 is EMCC sample after deleting 1 outlier.

3.3.2 Normality - UMCG sample

```
psych::describe(data_pooled[data_pooled$centerID == "UMCG",
                              c(names_covariates, "MoCA_total",
                                names_rel_items_abbrev)]) %>%
  select(n, min, max, mean, median, sd, skew, kurtosis)
```

```
##           n min  max mean median    sd skew kurtosis
## age       81  18   66 28.70    22 14.16  1.35    0.12
## sex       81   0    1  0.63     1  0.49 -0.53   -1.74
## education_level 81   3    7  6.01     6  0.60 -1.36    6.61
## MoCA_total    0 Inf -Inf   NaN    NA    NA    NA     NA
## EKM60t       81  38   56 47.06    48  4.50 -0.29   -0.83
## ERTt         0 Inf -Inf   NaN    NA    NA    NA     NA
## TASf1       81   7   28 21.11    21  4.42 -0.83    0.79
## FP_Gt       81  14   24 20.26    21  2.40 -0.72    0.03
## HT_Gt       81   6   12 11.40    12  1.01 -2.64    9.49
## CART_Gt     81   0   12  8.55     9  2.49 -0.53    0.05
## SET_Gt      81   8   12 11.35    12  0.94 -1.44    1.50
## SNQt       81  16   22 19.41    19  1.25 -0.41   -0.12
## RSMSf1      0 Inf -Inf   NaN    NA    NA    NA     NA
## MBI_t       0 Inf -Inf   NaN    NA    NA    NA     NA
```

```
normalityCheck_UMCG <- psych::describe(data_pooled[data_pooled$centerID == "UMCG",
                                                    c(names_covariates, "MoCA_total",
                                                      names_rel_items_abbrev)]) %>%
  select(skew, kurtosis)

any(abs(normalityCheck_UMCG$skew)>3, na.rm = TRUE)
```

```
## [1] FALSE
```

```
any(abs(normalityCheck_UMCG$kurtosis)>10, na.rm = TRUE)
```

```
## [1] FALSE
```

No absolute skew>3 and no absolute kurtosis>10 is EMC sample after deleting 1 outlier.

3.3.3 Normality - pooled

```
psych::describe(data_pooled[, c(names_covariates, "MoCA_total", names_rel_items_abbrev)]) %>%
  select(n, min, max, mean, median, sd, skew, kurtosis)
```

##	n	min	max	mean	median	sd	skew	kurtosis
## age	129	18.0	79	35.74	24.0	18.56	0.74	-1.01
## sex	129	0.0	1	0.64	1.0	0.48	-0.56	-1.70
## education_level	129	2.0	7	5.91	6.0	0.83	-1.55	4.63
## MoCA_total	48	19.0	30	26.10	26.5	2.42	-0.59	-0.08
## EKM60t	127	34.0	58	47.13	48.0	4.68	-0.32	-0.30
## ERTt	47	34.0	76	57.21	58.0	9.44	-0.20	-0.55
## TASf1	129	5.0	28	20.73	21.0	4.82	-0.79	0.56
## FP_Gt	129	10.5	24	20.25	20.5	2.41	-0.90	1.23
## HT_Gt	128	6.0	12	11.34	12.0	1.03	-2.64	9.79
## CART_Gt	120	0.0	12	8.40	9.0	2.55	-0.46	-0.28
## SET_Gt	126	8.0	12	11.37	12.0	0.86	-1.53	1.99
## SNQt	129	16.0	22	19.34	19.0	1.21	-0.41	-0.11
## RSMSf1	48	11.0	32	22.75	23.0	4.62	-0.57	0.02
## MBIt	48	42.0	70	54.29	54.5	7.09	0.21	-0.66

```
normalityCheck_pld <- psych::describe(data_pooled[, c(names_covariates, "MoCA_total",
  names_rel_items_abbrev)]) %>%
  select(skew, kurtosis)
```

```
any(abs(normalityCheck_pld$skew)>3, na.rm = TRUE)
```

```
## [1] FALSE
```

```
any(abs(normalityCheck_pld$kurtosis)>10, na.rm = TRUE)
```

```
## [1] FALSE
```

No absolute skew>3 and no absolute kurtosis>10 is pooled sample after deleting 2 outliers.

3.4 Descriptives (after outlier removal)

```
# Descriptives pooled before outlier deletion and/or transformations
psych::describe(data_pooled[,
  c(names_covariates,
    "MoCA_total",
    names_rel_items_abbrev)]) %>%
  select(n, min, max, mean, median, sd, skew, kurtosis)
```

```
##           n  min max  mean median    sd  skew kurtosis
## age       129 18.0  79 35.74   24.0 18.56  0.74   -1.01
## sex       129  0.0   1  0.64    1.0  0.48 -0.56   -1.70
## education_level 129  2.0   7  5.91    6.0  0.83 -1.55    4.63
## MoCA_total    48 19.0  30 26.10   26.5  2.42 -0.59   -0.08
## EKM60t       127 34.0  58 47.13   48.0  4.68 -0.32   -0.30
## ERTt         47 34.0  76 57.21   58.0  9.44 -0.20   -0.55
## TASf1        129  5.0  28 20.73   21.0  4.82 -0.79    0.56
## FP_Gt        129 10.5  24 20.25   20.5  2.41 -0.90    1.23
## HT_Gt        128  6.0  12 11.34   12.0  1.03 -2.64    9.79
## CART_Gt       120  0.0  12  8.40    9.0  2.55 -0.46   -0.28
## SET_Gt       126  8.0  12 11.37   12.0  0.86 -1.53    1.99
## SNQt         129 16.0  22 19.34   19.0  1.21 -0.41   -0.11
## RSMSf1        48 11.0  32 22.75   23.0  4.62 -0.57    0.02
## MBIt         48 42.0  70 54.29   54.5  7.09  0.21   -0.66
```

4 Methods: Sample comparisons

Statistical comparisons – Welch’s t-tests, Chi-square test of independence, and the Mann-Whitney U test – of the EMC sample versus the UMCG sample based on the observed data (before imputation).

```
#age
scAge <- t.test(formula = age ~ centerID, data = data_pooled); scAge
```

```
##
## Welch Two Sample t-test
##
## data: age by centerID
## t = 5.9392, df = 77.565, p-value = 7.644e-08
## alternative hypothesis: true difference in means between group EMC and group UMCG is not equal to 0
## 95 percent confidence interval:
## 12.56440 25.23652
## sample estimates:
## mean in group EMC mean in group UMCG
## 47.60417 28.70370
```

```
cohens_d(formula = age ~ centerID, data = data_pooled, var.equal = FALSE)
```

```
## # A tibble: 1 x 7
##   .y. group1 group2 effsize    n1    n2 magnitude
## * <chr> <chr> <chr>    <dbl> <int> <int> <ord>
## 1 age   EMC   UMCG    1.12   48   81 large
```

```
#sex
table(data_pooled$sex, data_pooled$centerID)
```

```
##
##      EMC UMCG
## 0   17   30
## 1   31   51
```

```
scSex <- chisq.test(table(data_pooled$sex, data_pooled$centerID)); scSex
```

```
##
## Pearson's Chi-squared test with Yates' continuity correction
##
## data: table(data_pooled$sex, data_pooled$centerID)
## X-squared = 2.4514e-31, df = 1, p-value = 1
```

```
sqrt(scSex$statistic[[1]] / (nrow(data_pooled)*5))
```

```
## [1] 1.949511e-17
```

```
#education level
```

```
scEduc <- rstatix:: wilcox_test(data_pooled, education_level~centerID)
rstatix:: wilcox_effsize(data_pooled, education_level~centerID)
```

```
## # A tibble: 1 x 7
##   .y.      group1 group2 effsize    n1    n2 magnitude
## * <chr>      <chr> <chr>    <dbl> <int> <int> <ord>
## 1 education_level EMC    UMCG    0.102   48    81 small
```

```
#Ekman
```

```
scEKM <- t.test(formula = EKM60t ~ centerID, data = data_pooled); scEKM
```

```
##
## Welch Two Sample t-test
##
## data: EKM60t by centerID
## t = 0.1987, df = 85.501, p-value = 0.843
## alternative hypothesis: true difference in means between group EMC and group UMCG is not equal to 0
## 95 percent confidence interval:
## -1.597633  1.952437
## sample estimates:
## mean in group EMC mean in group UMCG
##          47.23913          47.06173
```

```
cohens_d(formula = EKM60t ~ centerID, data = data_pooled, var.equal = FALSE)
```

```
## # A tibble: 1 x 7
##   .y.      group1 group2 effsize    n1    n2 magnitude
## * <chr> <chr> <chr>    <dbl> <int> <int> <ord>
## 1 EKM60t EMC    UMCG    0.0372   46    81 negligible
```

```
#TAS
```

```
scTAS<- t.test(formula = TASf1 ~ centerID, data = data_pooled); scTAS
```

```
##
## Welch Two Sample t-test
##
```

```
## data: TASf1 by centerID
## t = -1.1121, df = 83.573, p-value = 0.2693
## alternative hypothesis: true difference in means between group EMC and group UMCG is not equal to 0
## 95 percent confidence interval:
## -2.8657047 0.8101491
## sample estimates:
## mean in group EMC mean in group UMCG
## 20.08333 21.11111
```

```
cohens_d(formula = TASf1 ~ centerID, data = data_pooled, var.equal = FALSE)
```

```
## # A tibble: 1 x 7
## .y. group1 group2 effsize n1 n2 magnitude
## * <chr> <chr> <chr> <dbl> <int> <int> <ord>
## 1 TASf1 EMC UMCG -0.208 48 81 small
```

```
#FP
```

```
scFP<- t.test(formula = FP_Gt ~ centerID, data = data_pooled); scFP
```

```
##
## Welch Two Sample t-test
##
## data: FP_Gt by centerID
## t = -0.06772, df = 96.615, p-value = 0.9461
## alternative hypothesis: true difference in means between group EMC and group UMCG is not equal to 0
## 95 percent confidence interval:
## -0.9120807 0.8518955
## sample estimates:
## mean in group EMC mean in group UMCG
## 20.22917 20.25926
```

```
cohens_d(formula = FP_Gt ~ centerID, data = data_pooled, var.equal = FALSE)
```

```
## # A tibble: 1 x 7
## .y. group1 group2 effsize n1 n2 magnitude
## * <chr> <chr> <chr> <dbl> <int> <int> <ord>
## 1 FP_Gt EMC UMCG -0.0124 48 81 negligible
```

```
#CART
```

```
scCART<- t.test(formula = CART_Gt ~ centerID, data = data_pooled); scCART
```

```
##
## Welch Two Sample t-test
##
## data: CART_Gt by centerID
## t = -0.87672, df = 70.697, p-value = 0.3836
## alternative hypothesis: true difference in means between group EMC and group UMCG is not equal to 0
## 95 percent confidence interval:
## -1.4631069 0.5694697
## sample estimates:
## mean in group EMC mean in group UMCG
## 8.102564 8.549383
```

```
cohens_d(formula = CART_Gt ~ centerID, data = data_pooled, var.equal = FALSE)
```

```
## # A tibble: 1 x 7
##   .y.      group1 group2 effsize    n1    n2 magnitude
## * <chr>   <chr>   <chr>   <dbl> <int> <int> <ord>
## 1 CART_Gt EMC     UMCG    -0.173   39    81 negligible
```

```
#SET
```

```
scSET<- t.test(formula = SET_Gt ~ centerID, data = data_pooled); scSET
```

```
##
## Welch Two Sample t-test
##
## data: SET_Gt by centerID
## t = 0.39442, df = 112.48, p-value = 0.694
## alternative hypothesis: true difference in means between group EMC and group UMCG is not equal to 0
## 95 percent confidence interval:
## -0.2356628 0.3528113
## sample estimates:
## mean in group EMC mean in group UMCG
##      11.40425      11.34568
```

```
cohens_d(formula = SET_Gt ~ centerID, data = data_pooled, var.equal = FALSE)
```

```
## # A tibble: 1 x 7
##   .y.      group1 group2 effsize    n1    n2 magnitude
## * <chr>   <chr>   <chr>   <dbl> <int> <int> <ord>
## 1 SET_Gt EMC     UMCG    0.0704   45    81 negligible
```

```
#SNQ
```

```
scSNQ<- t.test(formula = SNQt ~ centerID, data = data_pooled); scSNQ
```

```
##
## Welch Two Sample t-test
##
## data: SNQt by centerID
## t = -0.82939, df = 106.73, p-value = 0.4087
## alternative hypothesis: true difference in means between group EMC and group UMCG is not equal to 0
## 95 percent confidence interval:
## -0.6042762 0.2477947
## sample estimates:
## mean in group EMC mean in group UMCG
##      19.22917      19.40741
```

```
cohens_d(formula = SNQt ~ centerID, data = data_pooled, var.equal = FALSE)
```

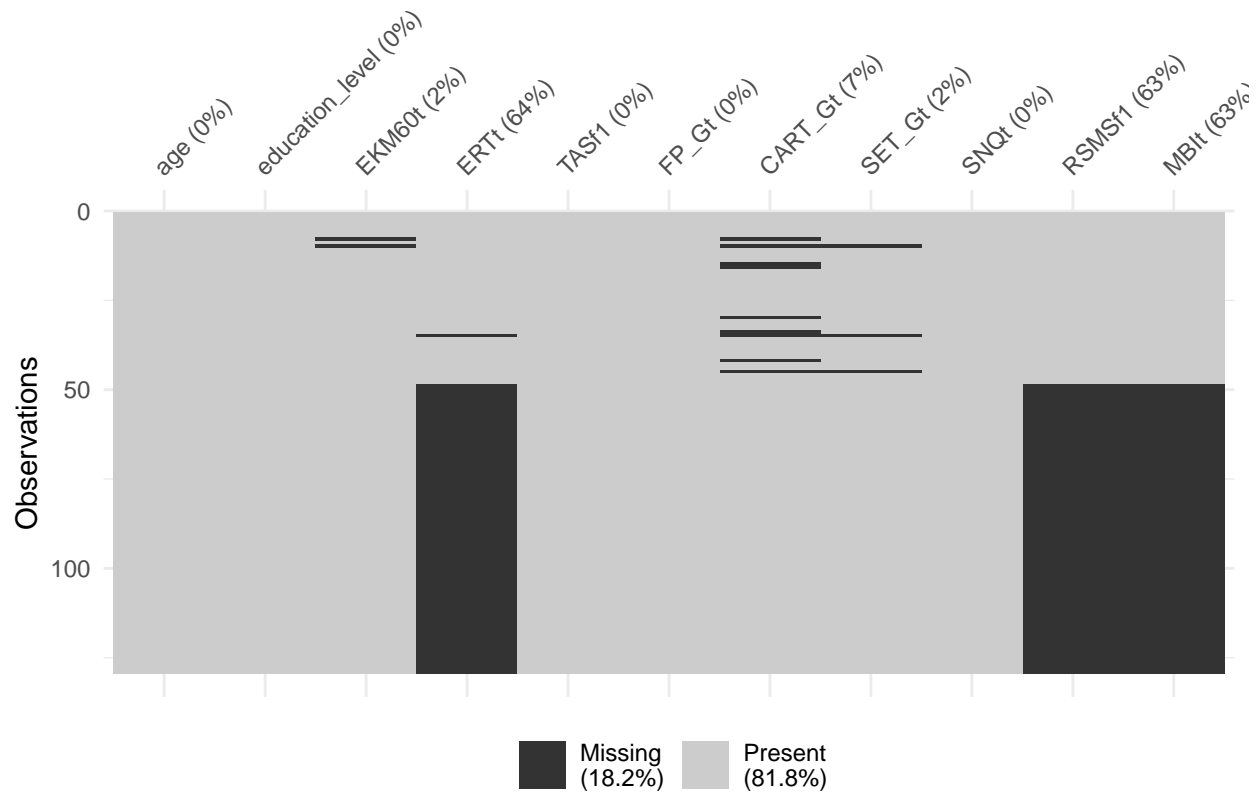
```
## # A tibble: 1 x 7
##   .y.      group1 group2 effsize    n1    n2 magnitude
## * <chr>   <chr>   <chr>   <dbl> <int> <int> <ord>
## 1 SNQt   EMC     UMCG    -0.149   48    81 negligible
```


5 Methods: Stochastic imputation

5.1 Missing data

At this point, we have the following missing values.

```
vis_miss(data_pooled[,c("age", "education_level",
                        'EKM60t', 'ERTt', 'TASf1',
                        'FP_Gt', 'CART_Gt', 'SET_Gt',
                        'SNQt', 'RSMSf1', 'MBIt')])
```



5.2 Correlations

And the following zero order paired correlation (respectively: corr, N, p-values).

```
Hmisc::rcorr(as.matrix(data_pooled[, c("age", "education_level",
                                        'EKM60t', 'ERTt', 'TASf1',
                                        'FP_Gt', 'CART_Gt', 'SET_Gt',
                                        'SNQt', 'RSMSf1', 'MBIt')]),
             type = "spearman")
```

```
##          age education_level EKM60t  ERTt TASf1 FP_Gt CART_Gt SET_Gt
## age      1.00          -0.09  -0.02 -0.47  0.06 -0.14   0.08  -0.19
```

```

## education_level -0.09          1.00  0.23  0.46  0.10  0.18    0.36  0.17
## EKM60t          -0.02          0.23  1.00  0.59  0.14  0.26    0.19  0.05
## ERTt           -0.47          0.46  0.59  1.00  0.16  0.38    0.21  0.27
## TASf1           0.06          0.10  0.14  0.16  1.00  0.06    0.01  0.04
## FP_Gt          -0.14          0.18  0.26  0.38  0.06  1.00    0.18  0.17
## CART_Gt         0.08          0.36  0.19  0.21  0.01  0.18    1.00  0.13
## SET_Gt         -0.19          0.17  0.05  0.27  0.04  0.17    0.13  1.00
## SNQt           -0.01          0.19  0.13  0.08  0.08  0.24    0.07  0.06
## RSMSf1         -0.08         -0.06  0.23  0.13  0.27  0.09    0.23 -0.06
## MBI_t          0.30          0.00  0.19  0.13  0.20 -0.01    0.30 -0.03
##
##          SNQt RSMSf1 MBI_t
## age      -0.01 -0.08 0.30
## education_level 0.19 -0.06 0.00
## EKM60t     0.13  0.23 0.19
## ERTt       0.08  0.13 0.13
## TASf1      0.08  0.27 0.20
## FP_Gt      0.24  0.09 -0.01
## CART_Gt    0.07  0.23 0.30
## SET_Gt     0.06 -0.06 -0.03
## SNQt       1.00 -0.23 0.10
## RSMSf1    -0.23  1.00 -0.09
## MBI_t     0.10 -0.09 1.00
##
## n
##          age education_level EKM60t ERTt TASf1 FP_Gt CART_Gt SET_Gt SNQt
## age      129          129    127  47   129   129    120   126  129
## education_level 129          129    127  47   129   129    120   126  129
## EKM60t    127          127    127  45   127   127    120   125  127
## ERTt      47          47     45  47    47    47     39    45   47
## TASf1     129          129    127  47   129   129    120   126  129
## FP_Gt     129          129    127  47   129   129    120   126  129
## CART_Gt   120          120    120  39   120   120    120   120  120
## SET_Gt    126          126    125  45   126   126    120   126  126
## SNQt      129          129    127  47   129   129    120   126  129
## RSMSf1    48          48     46  47    48    48     39    45   48
## MBI_t     48          48     46  47    48    48     39    45   48
##
##          RSMSf1 MBI_t
## age         48   48
## education_level 48   48
## EKM60t      46   46
## ERTt        47   47
## TASf1       48   48
## FP_Gt       48   48
## CART_Gt     39   39
## SET_Gt      45   45
## SNQt        48   48
## RSMSf1      48   48
## MBI_t       48   48
##
## P
##          age      education_level EKM60t ERTt   TASf1 FP_Gt CART_Gt
## age              0.2964          0.8498 0.0008 0.5149 0.1108 0.3629
## education_level 0.2964          0.0108 0.0011 0.2542 0.0358 0.0000
## EKM60t          0.8498 0.0108          0.0000 0.1110 0.0035 0.0347

```

```
## ERTt      0.0008 0.0011      0.0000      0.2912 0.0093 0.1951
## TASf1     0.5149 0.2542      0.1110 0.2912      0.4645 0.9166
## FP_Gt     0.1108 0.0358      0.0035 0.0093 0.4645      0.0544
## CART_Gt   0.3629 0.0000      0.0347 0.1951 0.9166 0.0544
## SET_Gt    0.0340 0.0645      0.6146 0.0738 0.6720 0.0605 0.1560
## SNQt      0.9125 0.0280      0.1441 0.5789 0.3492 0.0073 0.4617
## RSMSf1    0.5954 0.7063      0.1205 0.3964 0.0635 0.5606 0.1509
## MBit      0.0353 0.9846      0.2060 0.3663 0.1678 0.9730 0.0651
##          SET_Gt SNQt  RSMSf1 MBit
## age      0.0340 0.9125 0.5954 0.0353
## education_level 0.0645 0.0280 0.7063 0.9846
## EKM60t    0.6146 0.1441 0.1205 0.2060
## ERTt      0.0738 0.5789 0.3964 0.3663
## TASf1     0.6720 0.3492 0.0635 0.1678
## FP_Gt     0.0605 0.0073 0.5606 0.9730
## CART_Gt   0.1560 0.4617 0.1509 0.0651
## SET_Gt          0.5168 0.7084 0.8448
## SNQt      0.5168      0.1213 0.5163
## RSMSf1    0.7084 0.1213      0.5422
## MBit      0.8448 0.5163 0.5422
```

5.3 Imputation predictor selection

5.3.1 Predictor selection ERTt

```
matcor <- Hmisc::rcorr(as.matrix(data_pooled[, c("age", "education_level",
        'EKM60t', 'ERTt', 'TASf1',
        'FP_Gt', 'CART_Gt', 'SET_Gt',
        'SNQt', 'RSMSf1', 'MBit')])),
        type = "spearman")

#check whether sex is also a significant predictor
sexERTt <- lm(ERTt ~ sex, data = data_pooled)
summary(sexERTt)$coefficients["sex", "Pr(>|t|)"] #Not significant; p = 0.218

## [1] 0.2181282
```

```
#selecting predictors based on significant correlation
sigpred_ERTt <- matcor$r["ERTt",matcor$p["ERTt",]<.05]; sigpred_ERTt
```

```
##          age education_level      EKM60t      <NA>      FP_Gt
##      -0.4733335      0.4620705      0.5866877      NA      0.3753067
```

5.3.2 Predictor selection RSMSf1

Lets check the correlations of RSMSf1 with other relevant variables.

```
#Check whether sex is also a significant predictor
sexRSMSf1 <- lm(RSMSf1 ~ sex, data = data_pooled)
summary(sexRSMSf1)$coefficients["sex", "Pr(>|t|)"] #Not significant, p = 0.0867
```

```
## [1] 0.08671293
```

```
#Check numerical correlations
```

```
sigpred_RSMSf1 <- matcor$r["RSMSf1",matcor$P["RSMSf1",]<.05];  
sigpred_RSMSf1 #less than three predictors
```

```
## [1] NA
```

```
# increment alpha with .05
```

```
sigpred_RSMSf1 <- matcor$r["RSMSf1",matcor$P["RSMSf1",]<.1]  
#adds sex to the predictors  
sigpred_RSMSf1$sex <- summary(sexRSMSf1)$coefficients["sex", "Pr(>|t|)"]
```

```
## Warning in sigpred_RSMSf1$sex <- summary(sexRSMSf1)$coefficients["sex", :  
## Coercing LHS to a list
```

```
sigpred_RSMSf1 #less than three predictors
```

```
## $TASf1  
## [1] 0.2700051  
##  
## $<NA>  
## [1] NA  
##  
## $sex  
## [1] 0.08671293
```

```
# increment alpha with .05
```

```
sigpred_RSMSf1 <- matcor$r["RSMSf1",matcor$P["RSMSf1",]<.15]  
#adds sex to the predictors  
sigpred_RSMSf1$sex <- summary(sexRSMSf1)$coefficients["sex", "Pr(>|t|)"]
```

```
## Warning in sigpred_RSMSf1$sex <- summary(sexRSMSf1)$coefficients["sex", :  
## Coercing LHS to a list
```

```
sigpred_RSMSf1 # 4 predictors
```

```
## $EKM60t  
## [1] 0.2321768  
##  
## $TASf1  
## [1] 0.2700051  
##  
## $SNQt  
## [1] -0.2266534  
##  
## $<NA>  
## [1] NA  
##  
## $sex  
## [1] 0.08671293
```

5.3.3 Predictor selection MBIIt

Lets check the correlations of MBIIt with other relevant variables.

```
#Check whether sex is also a significant predictor
sexMBIt <- lm(MBIIt ~ sex, data = data_pooled)
summary(sexMBIt)$coefficients["sex", "Pr(>|t|)"] #Not significant, p = 0.3339
```

```
## [1] 0.3339226
```

```
#Check numerical correlations
sigpred_MBIIt <- matcor$r["MBIt",matcor$P["MBIt"],<.05];
sigpred_MBIIt # only one sign predictor
```

```
##      age      <NA>
## 0.3045774      NA
```

```
# increment alpha with .05
sigpred_MBIIt <- matcor$r["MBIt",matcor$P["MBIt"],<.1];
sigpred_MBIIt # only 2 predictors
```

```
##      age  CART_Gt      <NA>
## 0.3045774 0.2983429      NA
```

```
# increment alpha with .05
sigpred_MBIIt <- matcor$r["MBIt",matcor$P["MBIt"],<.15];
sigpred_MBIIt # only 2 predictors
```

```
##      age  CART_Gt      <NA>
## 0.3045774 0.2983429      NA
```

```
# increment alpha with .05
sigpred_MBIIt <- matcor$r["MBIt",matcor$P["MBIt"],<.2];
sigpred_MBIIt # 3 predictors
```

```
##      age  TASf1  CART_Gt      <NA>
## 0.3045774 0.2023403 0.2983429      NA
```

5.4 Imputation of ERTt, RSMSf, MBIIt

```
data_pooled_subset <- data_pooled[,c("ID", "centerID", "sex", "age", "education_level",
                                     'EKM60t', 'ERTt', 'TASf1',
                                     'FP_Gt', 'CART_Gt', 'SET_Gt',
                                     'SNQt', 'RSMSf1', 'MBIt')]

predictor_matrix <- make.predictorMatrix(data_pooled_subset)
predictor_matrix[] <- 0
```

```

#assign the significant predictors to the prediction matrix
predictor_matrix["ERTt", ] <- colnames(predictor_matrix) %in%
  names(sigpred_ERTt)[!is.na(names(sigpred_ERTt))]
predictor_matrix["RSMSf1", ] <- colnames(predictor_matrix) %in%
  names(sigpred_RSMSf1)[!is.na(names(sigpred_RSMSf1))]
predictor_matrix["MBIt", ] <- colnames(predictor_matrix) %in%
  names(sigpred_MBIt)[!is.na(names(sigpred_MBIt))]

#assign imputation methods
imputation_methods <- make.method(data_pooled_subset)
imputation_methods[] <- "" # Set all methods to "" initially
# Use "pmm" for ERTt, ESMf1, and MBIt
imputation_methods[c("ERTt", "RSMSf1", "MBIt")] <- "pmm"

#imputation
data_pooled_subset_imp_s1 <- mice(data_pooled_subset,
                                method = imputation_methods,
                                predictorMatrix = predictor_matrix,
                                m = 1,
                                seed = 2)

```

```

##
## iter imp variable
## 1 1 ERTt RSMSf1 MBIt
## 2 1 ERTt RSMSf1 MBIt
## 3 1 ERTt RSMSf1 MBIt
## 4 1 ERTt RSMSf1 MBIt
## 5 1 ERTt RSMSf1 MBIt

```

```

data_pooled_subset_imp <- complete(data_pooled_subset_imp_s1, 1)

```

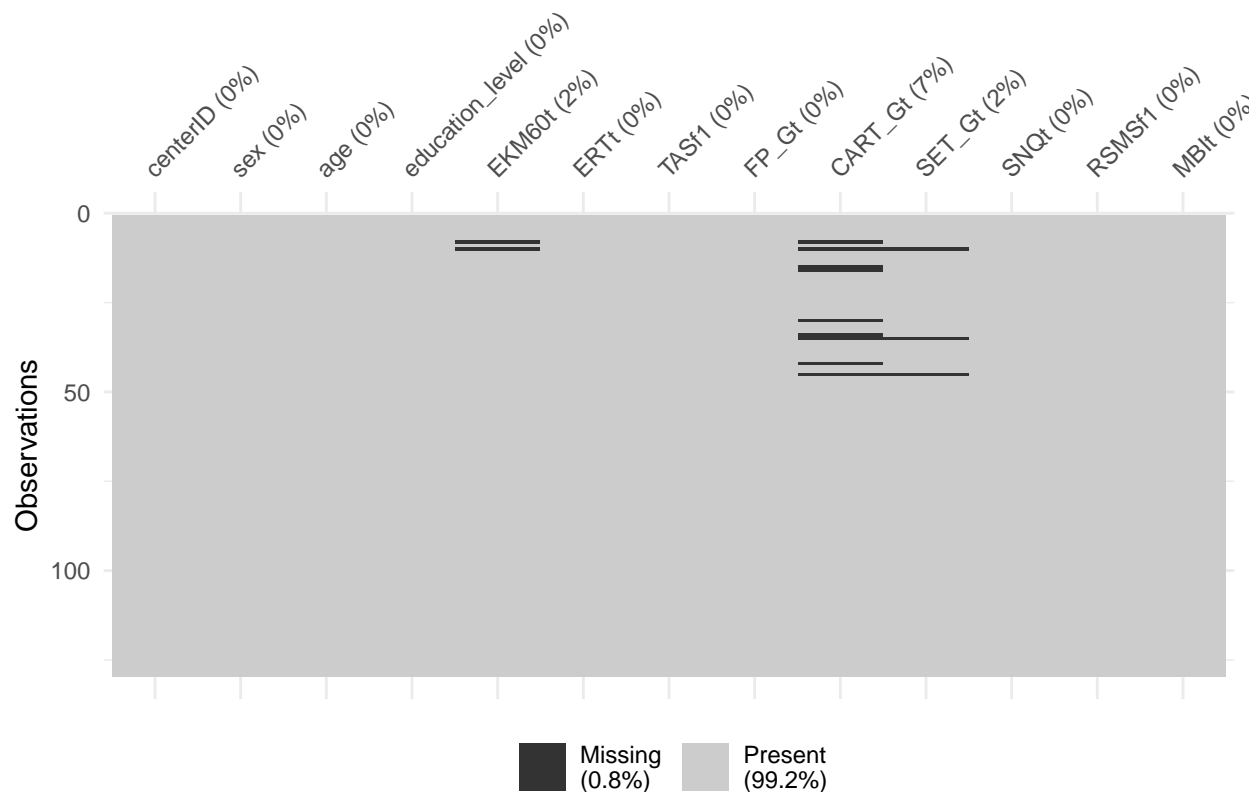
5.5 Mid evaluation of missing data

At this point, we have the remaining missing data:

```

vis_miss(data_pooled_subset_imp[, -1])

```



We will impute the remaining missing data, again with a stochastic approach based on relations in the available data using predicted mean matching.

5.6 Imputation predictor selection part 2

5.6.1 Predictor selection CART_Gt

Selecting related predictors for imputing CART_Gt.

```
matcor2 <- Hmisc:: rcorr(as.matrix(data_pooled_subset_imp[, c("age", "education_level",
  'EKM60t', 'ERTt', 'TASf1',
  'FP_Gt', 'CART_Gt', 'SET_Gt',
  'SNQt', 'RSMSf1', 'MBIt')])),
  type = "spearman")
```

#Check whether sex is also a significant predictor

```
sexCART_Gt <- lm(CART_Gt ~ sex, data = data_pooled_subset_imp)
summary(sexCART_Gt)$coefficients["sex", "Pr(>|t|)"] #Significant
```

```
## [1] 0.0003290575
```

```
sigpred_CART_Gt <- matcor2$r["CART_Gt",matcor2$P["CART_Gt",]<.05]
# add sex since it is significant
sigpred_CART_Gt$sex <- summary(sexCART_Gt)$coefficients["sex", "Pr(>|t|)"]
```

```
## Warning in sigpred_CART_Gt$sex <- summary(sexCART_Gt)$coefficients["sex", :
## Coercing LHS to a list
```

```
sigpred_CART_Gt #significant correlations of CART_Gt
```

```
## $education_level
## [1] 0.3649806
##
## $EKM60t
## [1] 0.1930374
##
## $<NA>
## [1] NA
##
## $RSMSf1
## [1] 0.187745
##
## $MBIt
## [1] 0.2487448
##
## $sex
## [1] 0.0003290575
```

5.6.2 Predictor selection SET_Gt

Selecting related predictors for imputing SET_Gt.

```
#Check whether sex is also a significant predictor
sexSET_Gt <- lm(SET_Gt ~ sex, data = data_pooled_subset_imp)
summary(sexSET_Gt)$coefficients["sex", "Pr(>|t|)"] #Not significant
```

```
## [1] 0.1178248
```

```
#< 3 predictors selected
sigpred_SET_Gt <- matcor2$r["SET_Gt",matcor2$p["SET_Gt",]<.05]; sigpred_SET_Gt
```

```
##          age          <NA>
## -0.1890881          NA
```

```
# increment alpha with .05
#Selected predictors
sigpred_SET_Gt <- matcor2$r["SET_Gt",matcor2$p["SET_Gt",]<.1]; sigpred_SET_Gt
```

```
##          age education_level          ERTt          FP_Gt          <NA>
##    -0.1890881      0.1651829      0.1639694      0.1676832          NA
```

5.6.3 Predictor selection EKM60t

Lets check the correlations of EKM60t with other relevant variables.


```
#Check whether sex is also a significant predictor
sexEKM60t <- lm(EKM60t ~ sex, data = data_pooled_subset_imp)
summary(sexEKM60t)$coefficients["sex", "Pr(>|t|)"] #Not significant
```

```
## [1] 0.2253531
```

```
#significant correlations of EKM60t
sigpred_EKM60t <- matcor2$r["EKM60t",matcor2$p["EKM60t"],<.05]; sigpred_EKM60t
```

```
## education_level      <NA>      ERTt      FP_Gt      CART_Gt
##      0.2253768      NA      0.5872751      0.2574957      0.1930374
##      RSMSf1
##      0.2370015
```

5.6.4 Predictor selection ERTt (second round)

Lets check the correlations of ERTt with other relevant variables.

```
#Check whether sex is also a significant predictor
sexERTt_2nd <- lm(ERTt ~ sex, data = data_pooled_subset_imp)
summary(sexERTt_2nd)$coefficients["sex", "Pr(>|t|)"] #Not significant
```

```
## [1] 0.5954151
```

```
#significant correlations of ERTt
sigpred_ERTt_2nd <- matcor2$r["ERTt",matcor2$p["ERTt"],<.05]; sigpred_ERTt_2nd
```

```
##      age education_level      EKM60t      <NA>      FP_Gt
##      -0.3284992      0.2868658      0.5872751      NA      0.2750760
```

5.7 Imputation of CART_Gt, SET_Gt, EKM60t, ERTt (2nd imputation round)

```
data_pooled_subset_imp2 <- data_pooled_subset_imp

predictor_matrix2 <- make.predictorMatrix(data_pooled_subset_imp)
predictor_matrix2[] <- 0

#assign the significant predictors to the prediction matrix
predictor_matrix2["CART_Gt", ] <-
  colnames(predictor_matrix2) %in% names(sigpred_CART_Gt)[!is.na(names(sigpred_CART_Gt))]
predictor_matrix2["SET_Gt", ] <-
  colnames(predictor_matrix2) %in% names(sigpred_SET_Gt)[!is.na(names(sigpred_SET_Gt))]
predictor_matrix2["EKM60t", ] <-
  colnames(predictor_matrix2) %in% names(sigpred_EKM60t)[!is.na(names(sigpred_EKM60t))]
predictor_matrix2["ERTt", ] <-
  colnames(predictor_matrix2) %in% names(sigpred_ERTt_2nd)[!is.na(names(sigpred_ERTt_2nd))]
```

```

#assign imputation methods
imputation_methods2 <- make.method(data_pooled_subset_imp)
imputation_methods2[] <- "" # Set all methods to "" initially
imputation_methods2[c("CART_Gt", "SET_Gt", "EKM60t", "ERTt")] <- "pmm" # Use "pmm" for ERTt

#imputation
data_pooled_subset_imp2_s1 <- mice(data_pooled_subset_imp,
                                   method = imputation_methods2,
                                   predictorMatrix = predictor_matrix2,
                                   m = 1,
                                   seed = 2)

```

```

##
## iter imp variable
## 1 1 EKM60t CART_Gt SET_Gt
## 2 1 EKM60t CART_Gt SET_Gt
## 3 1 EKM60t CART_Gt SET_Gt
## 4 1 EKM60t CART_Gt SET_Gt
## 5 1 EKM60t CART_Gt SET_Gt

```

```

data_pooled_subset_imp2 <- complete(data_pooled_subset_imp2_s1, 1)

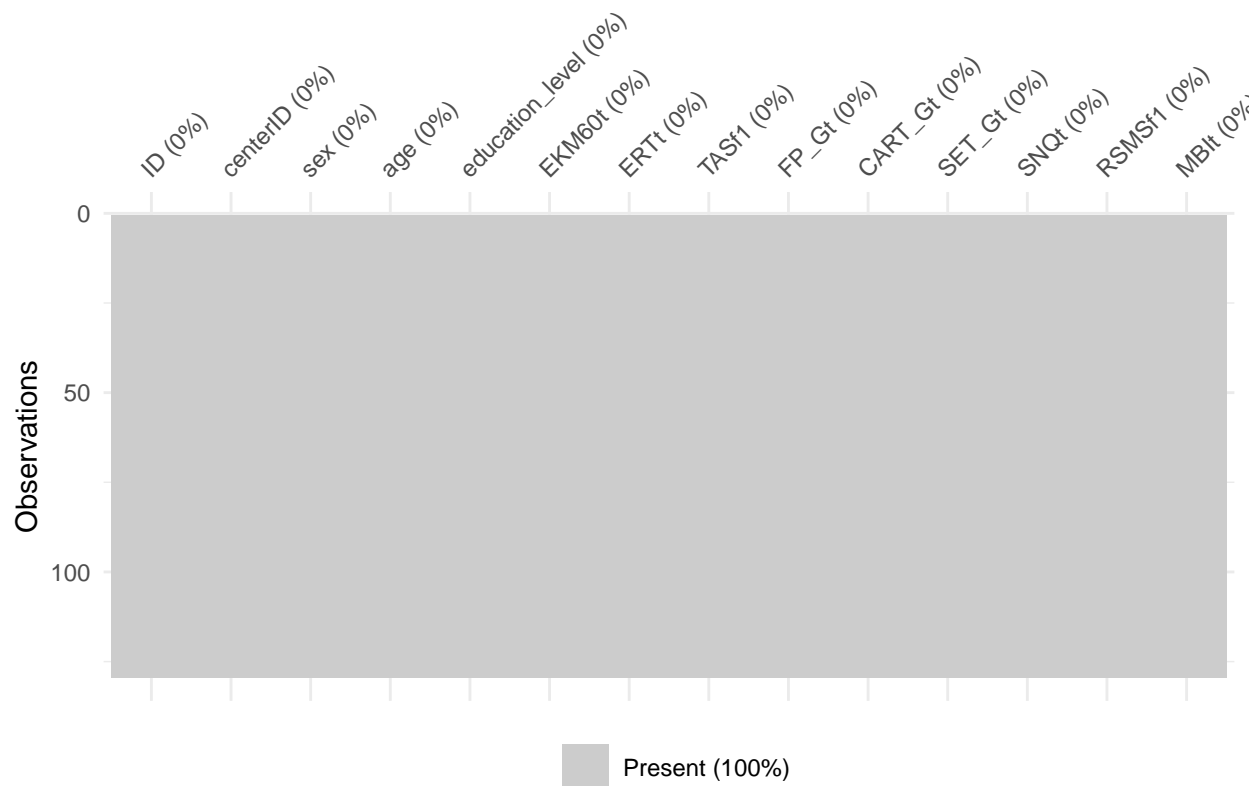
```

5.7.1 Check after imputation part 2

```

vis_miss(data_pooled_subset_imp2) #missing data check

```



No missing values anymore

6 Methods: Final dataset

6.1 Characteristics

```
data_final <- data_pooled_subset_imp2
data_final <- merge(data_final, data_pooled[,c('ID', 'TQ_total')], by = "ID", all.x = TRUE)
colnames(data_final)[colnames(data_final)=="TQ_total"] <- "TQ"

colnames(data_final) <- c("ID", "centerID", "sex", "age", "education_level",
                          "EKM", "ERT", "TASf1",
                          "FP", "CART", "SET",
                          "SNQ", "RSMSf1", "MBI",
                          "TQ")

descriptives_data_final <- psych::describe(data_final[,-1]) %>%
  select(n, min, max, mean, median, sd, skew, kurtosis); descriptives_data_final
```

##		n	min	max	mean	median	sd	skew	kurtosis
##	centerID*	129	1.0	2	1.63	2.0	0.49	-0.52	-1.74
##	sex	129	0.0	1	0.64	1.0	0.48	-0.56	-1.70
##	age	129	18.0	79	35.74	24.0	18.56	0.74	-1.01

```
## education_level 129 2.0 7 5.91 6.0 0.83 -1.55 4.63
## EKM 129 34.0 58 47.12 48.0 4.67 -0.32 -0.33
## ERT 129 34.0 76 59.64 61.0 8.87 -0.42 -0.36
## TASf1 129 5.0 28 20.73 21.0 4.82 -0.79 0.56
## FP 129 10.5 24 20.25 20.5 2.41 -0.90 1.23
## CART 129 0.0 12 8.35 8.5 2.52 -0.42 -0.31
## SET 129 8.0 12 11.35 12.0 0.88 -1.51 1.84
## SNQ 129 16.0 22 19.34 19.0 1.21 -0.41 -0.11
## RSMSf1 129 11.0 32 23.21 24.0 4.86 -0.70 0.37
## MBI 129 42.0 70 51.88 52.0 7.03 0.34 -0.76
## TQ 48 10.0 18 14.65 15.0 2.23 -0.20 -1.00
```

```
corr_data_final <- Hmisc::rcorr(as.matrix(data_final[, -c(1:2)]), type = "spearman")
corr_data_final
```

```
##          sex  age education_level  EKM  ERT TASf1  FP  CART  SET
## sex      1.00 -0.07              -0.17  0.12  0.06 -0.03 -0.10 -0.35 -0.17
## age     -0.07  1.00              -0.09 -0.01 -0.33  0.06 -0.14  0.02 -0.21
## education_level -0.17 -0.09          1.00  0.21  0.29  0.10  0.18  0.40  0.20
## EKM      0.12 -0.01              0.21  1.00  0.59  0.15  0.24  0.20  0.06
## ERT      0.06 -0.33              0.29  0.59  1.00  0.07  0.28  0.14  0.17
## TASf1    -0.03  0.06              0.10  0.15  0.07  1.00  0.06  0.02  0.03
## FP      -0.10 -0.14              0.18  0.24  0.28  0.06  1.00  0.22  0.19
## CART     -0.35  0.02              0.40  0.20  0.14  0.02  0.22  1.00  0.17
## SET     -0.17 -0.21              0.20  0.06  0.17  0.03  0.19  0.17  1.00
## SNQ      0.03 -0.01              0.19  0.11  0.10  0.08  0.24  0.10  0.07
## RSMSf1   -0.34  0.03              0.11  0.24  0.15  0.07  0.09  0.19  0.06
## MBI      0.02  0.43              0.03  0.03 -0.08  0.16 -0.06  0.19 -0.01
## TQ      0.12 -0.08              0.30  0.42  0.30  0.07  0.27  0.18  0.17
##          SNQ RSMSf1  MBI  TQ
## sex      0.03 -0.34  0.02  0.12
## age     -0.01  0.03  0.43 -0.08
## education_level 0.19  0.11  0.03  0.30
## EKM      0.11  0.24  0.03  0.42
## ERT      0.10  0.15 -0.08  0.30
## TASf1     0.08  0.07  0.16  0.07
## FP       0.24  0.09 -0.06  0.27
## CART     0.10  0.19  0.19  0.18
## SET      0.07  0.06 -0.01  0.17
## SNQ      1.00 -0.17 -0.02 -0.08
## RSMSf1   -0.17  1.00  0.01  0.17
## MBI     -0.02  0.01  1.00 -0.08
## TQ     -0.08  0.17 -0.08  1.00
##
## n
##          sex age education_level EKM ERT TASf1  FP  CART  SET SNQ RSMSf1
## sex      129 129              129 129 129  129 129  129 129 129
## age      129 129              129 129 129  129 129  129 129 129
## education_level 129 129          129 129 129  129 129  129 129 129
## EKM      129 129              129 129 129  129 129  129 129 129
## ERT      129 129              129 129 129  129 129  129 129 129
## TASf1     129 129              129 129 129  129 129  129 129 129
## FP       129 129              129 129 129  129 129  129 129 129
## CART     129 129              129 129 129  129 129  129 129 129
```

```

## SET      129 129      129 129 129 129 129 129 129 129
## SNQ      129 129      129 129 129 129 129 129 129 129
## RSMSf1   129 129      129 129 129 129 129 129 129 129
## MBI      129 129      129 129 129 129 129 129 129 129
## TQ       48 48        48 48 48 48 48 48 48 48
##          MBI TQ
## sex      129 48
## age      129 48
## education_level 129 48
## EKM      129 48
## ERT      129 48
## TASf1    129 48
## FP       129 48
## CART     129 48
## SET      129 48
## SNQ      129 48
## RSMSf1   129 48
## MBI      129 48
## TQ       48 48
##
## P
##          sex      age      education_level EKM      ERT      TASf1 FP
## sex          0.4536 0.0550      0.1789 0.5163 0.7604 0.2429
## age          0.4536      0.2964      0.8972 0.0001 0.5149 0.1108
## education_level 0.0550 0.2964      0.0188 0.0010 0.2542 0.0358
## EKM          0.1789 0.8972 0.0188      0.0000 0.0836 0.0059
## ERT          0.5163 0.0001 0.0010      0.0000      0.4495 0.0016
## TASf1        0.7604 0.5149 0.2542      0.0836 0.4495      0.4645
## FP          0.2429 0.1108 0.0358      0.0059 0.0016 0.4645
## CART         0.0000 0.7981 0.0000      0.0254 0.1095 0.8046 0.0141
## SET          0.0603 0.0185 0.0240      0.4845 0.0529 0.7024 0.0356
## SNQ          0.7112 0.9125 0.0280      0.2184 0.2523 0.3492 0.0073
## RSMSf1       0.0000 0.7135 0.2174      0.0055 0.0920 0.4428 0.3364
## MBI          0.8016 0.0000 0.7219      0.6962 0.3393 0.0753 0.5015
## TQ          0.4198 0.5811 0.0383      0.0030 0.0395 0.6353 0.0641
##          CART SET      SNQ      RSMSf1 MBI      TQ
## sex          0.0000 0.0603 0.7112 0.0000 0.8016 0.4198
## age          0.7981 0.0185 0.9125 0.7135 0.0000 0.5811
## education_level 0.0000 0.0240 0.0280 0.2174 0.7219 0.0383
## EKM          0.0254 0.4845 0.2184 0.0055 0.6962 0.0030
## ERT          0.1095 0.0529 0.2523 0.0920 0.3393 0.0395
## TASf1        0.8046 0.7024 0.3492 0.4428 0.0753 0.6353
## FP          0.0141 0.0356 0.0073 0.3364 0.5015 0.0641
## CART         0.0577 0.2384 0.0354 0.0340 0.2223
## SET          0.0577      0.4363 0.4961 0.8714 0.2501
## SNQ          0.2384 0.4363      0.0513 0.7897 0.5986
## RSMSf1       0.0354 0.4961 0.0513      0.8673 0.2444
## MBI          0.0340 0.8714 0.7897 0.8673      0.5859
## TQ          0.2223 0.2501 0.5986 0.2444 0.5859

```

```
corr_data_final_r <- round(as.data.frame(corr_data_final$r), 3); corr_data_final_r
```

```

##          sex      age education_level      EKM      ERT TASf1      FP
## sex      1.000 -0.067      -0.169 0.119 0.058 -0.027 -0.104

```

```
## age -0.067 1.000 -0.093 -0.011 -0.328 0.058 -0.141
## education_level -0.169 -0.093 1.000 0.207 0.287 0.101 0.185
## EKM 0.119 -0.011 0.207 1.000 0.593 0.153 0.241
## ERT 0.058 -0.328 0.287 0.593 1.000 0.067 0.275
## TASf1 -0.027 0.058 0.101 0.153 0.067 1.000 0.065
## FP -0.104 -0.141 0.185 0.241 0.275 0.065 1.000
## CART -0.346 0.023 0.404 0.197 0.142 0.022 0.216
## SET -0.166 -0.207 0.199 0.062 0.171 0.034 0.185
## SNQ 0.033 -0.010 0.193 0.109 0.102 0.083 0.235
## RSMSf1 -0.342 0.033 0.109 0.243 0.149 0.068 0.085
## MBI 0.022 0.426 0.032 0.035 -0.085 0.157 -0.060
## TQ 0.119 -0.082 0.300 0.419 0.298 0.070 0.269
## CART SET SNQ RSMSf1 MBI TQ
## sex -0.346 -0.166 0.033 -0.342 0.022 0.119
## age 0.023 -0.207 -0.010 0.033 0.426 -0.082
## education_level 0.404 0.199 0.193 0.109 0.032 0.300
## EKM 0.197 0.062 0.109 0.243 0.035 0.419
## ERT 0.142 0.171 0.102 0.149 -0.085 0.298
## TASf1 0.022 0.034 0.083 0.068 0.157 0.070
## FP 0.216 0.185 0.235 0.085 -0.060 0.269
## CART 1.000 0.168 0.105 0.185 0.187 0.179
## SET 0.168 1.000 0.069 0.060 -0.014 0.169
## SNQ 0.105 0.069 1.000 -0.172 -0.024 -0.078
## RSMSf1 0.185 0.060 -0.172 1.000 0.015 0.171
## MBI 0.187 -0.014 -0.024 0.015 1.000 -0.081
## TQ 0.179 0.169 -0.078 0.171 -0.081 1.000
```

```
corr_data_final_P <- round(as.data.frame(corr_data_final$P), 3); corr_data_final_P
```

```
## sex age education_level EKM ERT TASf1 FP CART SET
## sex NA 0.454 0.055 0.179 0.516 0.760 0.243 0.000 0.060
## age 0.454 NA 0.296 0.897 0.000 0.515 0.111 0.798 0.019
## education_level 0.055 0.296 NA 0.019 0.001 0.254 0.036 0.000 0.024
## EKM 0.179 0.897 0.019 NA 0.000 0.084 0.006 0.025 0.485
## ERT 0.516 0.000 0.001 0.000 NA 0.449 0.002 0.110 0.053
## TASf1 0.760 0.515 0.254 0.084 0.449 NA 0.465 0.805 0.702
## FP 0.243 0.111 0.036 0.006 0.002 0.465 NA 0.014 0.036
## CART 0.000 0.798 0.000 0.025 0.110 0.805 0.014 NA 0.058
## SET 0.060 0.019 0.024 0.485 0.053 0.702 0.036 0.058 NA
## SNQ 0.711 0.913 0.028 0.218 0.252 0.349 0.007 0.238 0.436
## RSMSf1 0.000 0.714 0.217 0.005 0.092 0.443 0.336 0.035 0.496
## MBI 0.802 0.000 0.722 0.696 0.339 0.075 0.501 0.034 0.871
## TQ 0.420 0.581 0.038 0.003 0.040 0.635 0.064 0.222 0.250
## SNQ RSMSf1 MBI TQ
## sex 0.711 0.000 0.802 0.420
## age 0.913 0.714 0.000 0.581
## education_level 0.028 0.217 0.722 0.038
## EKM 0.218 0.005 0.696 0.003
## ERT 0.252 0.092 0.339 0.040
## TASf1 0.349 0.443 0.075 0.635
## FP 0.007 0.336 0.501 0.064
## CART 0.238 0.035 0.034 0.222
## SET 0.436 0.496 0.871 0.250
## SNQ NA 0.051 0.790 0.599
```

```
## RSMSf1      0.051      NA 0.867 0.244
## MBI         0.790 0.867      NA 0.586
## TQ          0.599 0.244 0.586      NA
```

```
#knitr::kable(corr_data_final_r, format = "latex", booktabs = TRUE, caption = "Correlation matrix final")
#knitr::kable(corr_data_final_P, format = "latex", booktabs = TRUE, caption = "P-values corresponding t
```

6.2 Z-scores

```
names_items_analysis <- c("EKM", "ERT", "TASf1", "CART", "FP", "SET", "SNQ", "MBI", "RSMSf1")
data_final_Z <- cbind(data_final[, c("ID", "centerID", names_covariates)],
                      round(scale(data_final[, c(names_items_analysis, 'TQ')]), 4))
```

7 Methods: Analyses

7.1 CFA models

```
#full 3 factor model
m1_f3fm <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR
f2_U ~~ f3_BR
'

#two factor model: f1 = f2
m2_2fm_1eq2 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

# Allow factors to be correlated
f1_P ~~ 1*f2_U # Fix correlation between f1_P and f2_U at 1
f1_P ~~ f3_BR
f2_U ~~ f3_BR
'

#model 3: two factor model: f1 = f3 (CART)
m3_2fm_1eq3 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
```

```

f3_BR =~ RSMSf1 + SNQ + MBI

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ 1*f3_BR # Fix correlation between f1_P and f3_BR at 1
f2_U ~~ f3_BR
'

#model 4: two factor model: f2 = f3 (CART)
m4_2fm_2eq3 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR
f2_U ~~ 1*f3_BR # Fix correlation between f2_U and f3_BR at 1
'

#model 5: one factor model: f1 = f2 = f3 (CART)
m5_1fm <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

# Fixing all interfactor correlations at 1
f1_P ~~ 1*f2_U
f1_P ~~ 1*f3_BR
f2_U ~~ 1*f3_BR
'

#model 6: 3 one factor model: independent factors (CART)
m6_3x1fm <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

# Fixing all interfactor correlations at 0
f1_P ~~ 0*f2_U
f1_P ~~ 0*f3_BR
f2_U ~~ 0*f3_BR
'

# #model 6.3alt: 3 factor model; BR independent
# m6_3fm_BRindep <- '
# # Defining the factors (latent variables)
# f1_P =~ EKM + ERT + TASf1
# f2_U =~ SET + FP + CART
# f3_BR =~ RSMSf1 + SNQ + MBI

```



```
# # Fixing 2 interfactor correlations at 0
# f1_P ~~ f2_U
# f1_P ~~ 0*f3_BR
# f2_U ~~ 0*f3_BR
# '
```

7.2 CFA EMC data (N=49)

```
#model 1; EMC data
fit_m1 EMC <- cfa(model = m1_f3fm, data = data_final_Z[data_final_Z$centerID == "EMC",])
```

```
## Warning: lavaan->lav_object_post_check():
## some estimated lv variances are negative
```

```
summ_m1 EMC <- summary(fit_m1 EMC, standardized = TRUE, fit.measures = TRUE)
fm_m1 EMC <- fitMeasures(fit_m1 EMC, c("npar", "chisq", "df", "pvalue", "aic",
                                         "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
```

```
#model 2; EMC data
fit_m2 EMC <- cfa(model = m2_2fm_1eq2, data = data_final_Z[data_final_Z$centerID == "EMC",])
```

```
## Warning: lavaan->lav_start_check_cov():
## starting values imply a correlation larger than 1; variables involved are:
## f1_P f2_U
```

```
## Warning: lavaan->lav_object_post_check():
## some estimated lv variances are negative
```

```
summ_m2 EMC <- summary(fit_m2 EMC, standardized = TRUE, fit.measures = TRUE)
fm_m2 EMC <- fitMeasures(fit_m2 EMC, c("npar", "chisq", "df", "pvalue", "aic",
                                         "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
```

```
#model 3; EMC data
fit_m3 EMC <- cfa(model = m3_2fm_1eq3, data = data_final_Z[data_final_Z$centerID == "EMC",])
```

```
## Warning: lavaan->lav_start_check_cov():
## starting values imply a correlation larger than 1; variables involved are:
## f1_P f3_BR
```

```
## Warning: lavaan->lav_object_post_check():
## some estimated lv variances are negative
```

```
summ_m3 EMC <- summary(fit_m3 EMC, standardized = TRUE, fit.measures = TRUE)
fm_m3 EMC <- fitMeasures(fit_m3 EMC, c("npar", "chisq", "df", "pvalue", "aic",
                                         "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
```

```

#model 4; EMC data
fit_m4_EMC <- cfa(model = m4_2fm_2eq3, data = data_final_Z[data_final_Z$centerID == "EMC",])

## Warning: lavaan->lav_start_check_cov():
##   starting values imply a correlation larger than 1; variables involved are:
##   f2_U f3_BR

## Warning: lavaan->lav_object_post_check():
##   some estimated lv variances are negative

summ_m4_EMC <- summary(fit_m4_EMC, standardized = TRUE, fit.measures = TRUE)
fm_m4_EMC <- fitMeasures(fit_m4_EMC, c("npar", "chisq", "df", "pvalue", "aic",
                                       "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))

#model 5; EMC data
fit_m5_EMC <- cfa(model = m5_1fm, data = data_final_Z[data_final_Z$centerID == "EMC",])

## Warning: lavaan->lav_start_check_cov():
##   starting values imply a correlation larger than 1; variables involved are:
##   f1_P f2_U

## Warning: lavaan->lav_start_check_cov():
##   starting values imply a correlation larger than 1; variables involved are:
##   f1_P f3_BR

## Warning: lavaan->lav_object_post_check():
##   some estimated lv variances are negative

summ_m5_EMC <- summary(fit_m5_EMC, standardized = TRUE, fit.measures = TRUE)
fm_m5_EMC <- fitMeasures(fit_m5_EMC, c("npar", "chisq", "df", "pvalue", "aic",
                                       "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))

#model 6; EMC data
fit_m6_EMC <- cfa(model = m6_3x1fm, data = data_final_Z[data_final_Z$centerID == "EMC",])

## Warning: lavaan->lav_object_post_check():
##   some estimated ov variances are negative

summ_m6_EMC <- summary(fit_m6_EMC, standardized = TRUE, fit.measures = TRUE)
fm_m6_EMC <- fitMeasures(fit_m6_EMC, c("npar", "chisq", "df", "pvalue", "aic",
                                       "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))

```

7.3 CFA pooled data (N=128)

```

#model 1; pooled data
fit_m1_pld <- cfa(model = m1_f3fm, data = data_final_Z, std.lv = FALSE)

## Warning: lavaan->lav_object_post_check():
##   some estimated lv variances are negative

```

```
summ_m1_pld <- summary(fit_m1_pld, standardized = TRUE, fit.measures = TRUE)
fm_m1_pld <- fitMeasures(fit_m1_pld, c("npar", "chisq", "df", "pvalue", "aic",
                                         "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
```

```
#model 2; pooled data
fit_m2_pld <- cfa(model = m2_2fm_1eq2, data = data_final_Z)
```

```
## Warning: lavaan->lav_start_check_cov():
##   starting values imply a correlation larger than 1; variables involved are:
##   f1_P f2_U
```

```
## Warning: lavaan->lav_object_post_check():
##   some estimated lv variances are negative
```

```
summ_m2_pld <- summary(fit_m2_pld, standardized = TRUE, fit.measures = TRUE)
fm_m2_pld <- fitMeasures(fit_m2_pld, c("npar", "chisq", "df", "pvalue", "aic",
                                         "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
```

```
#model 3; pooled data
fit_m3_pld <- cfa(model = m3_2fm_1eq3, data = data_final_Z)
```

```
## Warning: lavaan->lav_start_check_cov():
##   starting values imply a correlation larger than 1; variables involved are:
##   f1_P f3_BR
```

```
## Warning: lavaan->lav_object_post_check():
##   some estimated lv variances are negative
```

```
summ_m3_pld <- summary(fit_m3_pld, standardized = TRUE, fit.measures = TRUE)
fm_m3_pld <- fitMeasures(fit_m3_pld, c("npar", "chisq", "df", "pvalue", "aic",
                                         "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
```

```
#model 4; pooled data
fit_m4_pld <- cfa(model = m4_2fm_2eq3, data = data_final_Z)
```

```
## Warning: lavaan->lav_start_check_cov():
##   starting values imply a correlation larger than 1; variables involved are:
##   f2_U f3_BR
```

```
## Warning: lavaan->lav_object_post_check():
##   some estimated lv variances are negative
```

```
summ_m4_pld <- summary(fit_m4_pld, standardized = TRUE, fit.measures = TRUE)
fm_m4_pld <- fitMeasures(fit_m4_pld, c("npar", "chisq", "df", "pvalue", "aic",
                                         "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
```

```
#model 5; pooled data
fit_m5_pld <- cfa(model = m5_1fm, data = data_final_Z)
```

```
## Warning: lavaan->lav_start_check_cov():
##   starting values imply a correlation larger than 1; variables involved are:
##   f1_P f2_U

## Warning: lavaan->lav_start_check_cov():
##   starting values imply a correlation larger than 1; variables involved are:
##   f1_P f3_BR

## Warning: lavaan->lav_object_post_check():
##   some estimated lv variances are negative
```

```
summ_m5_pld <- summary(fit_m5_pld, standardized = TRUE, fit.measures = TRUE)
fm_m5_pld <- fitMeasures(fit_m5_pld, c("npar", "chisq", "df", "pvalue", "aic",
                                       "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
```

```
#model 6; pooled data
fit_m6_pld <- cfa(model = m6_3x1fm, data = data_final_Z)
summ_m6_pld <- summary(fit_m6_pld, standardized = TRUE, fit.measures = TRUE)
fm_m6_pld <- fitMeasures(fit_m6_pld, c("npar", "chisq", "df", "pvalue", "aic",
                                       "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
```

7.4 SEM models

```
#3 paths model
#full 3 factor model
sem1 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

#regression
TQ ~ f1_P + f2_U + f3_BR

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR
f2_U ~~ f3_BR
'

#2 path models
#path P->TQ=0
sem2 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

#regression
TQ ~ 0*f1_P + f2_U + f3_BR
```

```

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR
f2_U ~~ f3_BR
'

#path U->TQ=0
sem3 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

#regression
TQ ~ f1_P + 0*f2_U + f3_BR

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR
f2_U ~~ f3_BR
'

#path BR->TQ=0
sem4 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

#regression
TQ ~ f1_P + f2_U + 0*f3_BR

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR
f2_U ~~ f3_BR
'

#1 path models
#only path P
sem5 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

#regression
TQ ~ f1_P + 0*f2_U + 0*f3_BR

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR

```

```

f2_U ~~ f3_BR
'

#only path U
sem6 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

#regression
TQ ~ 0*f1_P + f2_U + 0*f3_BR

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR
f2_U ~~ f3_BR
'

#only path BR
sem7 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

#regression
TQ ~ 0*f1_P + 0*f2_U + f3_BR

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR
f2_U ~~ f3_BR
'

#no path model (baseline)
sem8 <- '
# Defining the factors (latent variables)
f1_P =~ EKM + ERT + TASf1
f2_U =~ SET + FP + CART
f3_BR =~ RSMSf1 + SNQ + MBI

#regression
TQ ~ 0*f1_P + 0*f2_U + 0*f3_BR

# Allow factors to be correlated
f1_P ~~ f2_U
f1_P ~~ f3_BR
f2_U ~~ f3_BR
'

```

7.5 SEM EMC data (N=47)

```
#3 paths model
fit_sem1 <- sem(model = sem1, data = data_final_Z[data_final_Z$centerID == 'EMC',])
```

```
## Warning: lavaan->lav_object_post_check():
##      some estimated lv variances are negative
```

```
summ_sem1 <- summary(fit_sem1, standardized = TRUE, fit.measures = TRUE)
fm_sem1 <- fitMeasures(fit_sem1, c("npar", "chisq", "df", "pvalue", "aic",
                                   "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
stcoef_sem1 <- standardizedSolution(fit_sem1)[10:12,1:7]
```

```
#2 path models
fit_sem2 <- sem(model = sem2, data = data_final_Z[data_final_Z$centerID == 'EMC',])
```

```
## Warning: lavaan->lav_object_post_check():
##      some estimated lv variances are negative
```

```
summ_sem2 <- summary(fit_sem2, standardized = TRUE, fit.measures = TRUE)
fm_sem2 <- fitMeasures(fit_sem2, c("npar", "chisq", "df", "pvalue", "aic",
                                   "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
stcoef_sem2 <- standardizedSolution(fit_sem2)[10:12,1:7]
```

```
fit_sem3 <- sem(model = sem3, data = data_final_Z[data_final_Z$centerID == 'EMC',])
```

```
## Warning: lavaan->lav_object_post_check():
##      some estimated lv variances are negative
```

```
summ_sem3 <- summary(fit_sem3, standardized = TRUE, fit.measures = TRUE)
fm_sem3 <- fitMeasures(fit_sem3, c("npar", "chisq", "df", "pvalue", "aic",
                                   "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
stcoef_sem3 <- standardizedSolution(fit_sem3)[10:12,1:7]
```

```
fit_sem4 <- sem(model = sem4, data = data_final_Z[data_final_Z$centerID == 'EMC',])
```

```
## Warning: lavaan->lav_object_post_check():
##      some estimated lv variances are negative
```

```
summ_sem4 <- summary(fit_sem4, standardized = TRUE, fit.measures = TRUE)
fm_sem4 <- fitMeasures(fit_sem4, c("npar", "chisq", "df", "pvalue", "aic",
                                   "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
stcoef_sem4 <- standardizedSolution(fit_sem4)[10:12,1:7]
```

```
#1 path models
fit_sem5 <- sem(model = sem5, data = data_final_Z[data_final_Z$centerID == 'EMC',])
```

```
## Warning: lavaan->lav_object_post_check():
##      some estimated lv variances are negative

summ_sem5 <- summary(fit_sem5, standardized = TRUE, fit.measures = TRUE)
fm_sem5 <- fitMeasures(fit_sem5, c("npar", "chisq", "df", "pvalue", "aic",
                                   "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
stcoef_sem5 <- standardizedSolution(fit_sem5)[10:12,1:7]

fit_sem6 <- sem(model = sem6, data = data_final_Z[data_final_Z$centerID == 'EMC'],)
```

```
## Warning: lavaan->lav_object_post_check():
##      some estimated lv variances are negative
```

```
summ_sem6 <- summary(fit_sem6, standardized = TRUE, fit.measures = TRUE)
fm_sem6 <- fitMeasures(fit_sem6, c("npar", "chisq", "df", "pvalue", "aic",
                                   "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
stcoef_sem6 <- standardizedSolution(fit_sem6)[10:12,1:7]

fit_sem7 <- sem(model = sem7, data = data_final_Z[data_final_Z$centerID == 'EMC'],)
```

```
## Warning: lavaan->lav_object_post_check():
##      some estimated lv variances are negative
```

```
summ_sem7 <- summary(fit_sem7, standardized = TRUE, fit.measures = TRUE)
fm_sem7 <- fitMeasures(fit_sem7, c("npar", "chisq", "df", "pvalue", "aic",
                                   "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
stcoef_sem7 <- standardizedSolution(fit_sem7)[10:12,1:7]

#no path model
fit_sem8 <- sem(model = sem8, data = data_final_Z[data_final_Z$centerID == 'EMC'],)
```

```
## Warning: lavaan->lav_object_post_check():
##      some estimated lv variances are negative
```

```
summ_sem8 <- summary(fit_sem8, standardized = TRUE, fit.measures = TRUE)
fm_sem8 <- fitMeasures(fit_sem8, c("npar", "chisq", "df", "pvalue", "aic",
                                   "srmr", "cfi", "ifi", "nfi", "rmsea", "bic"))
stcoef_sem8 <- standardizedSolution(fit_sem8)[10:12,1:7]
```

8 Results

8.1 CFA EMC data

8.1.1 Fit indices


```

results_CFA EMC <- data.frame(round(rbind(fm_m1 EMC, fm_m2 EMC, fm_m3 EMC,
                                         fm_m4 EMC, fm_m5 EMC, fm_m6 EMC
                                         ), 3))

modelNames_CFA <- c("Full three-factor",
                    "Two-factor: P = U",
                    "Two-factor: P = BR",
                    "Two-factor: U = BR",
                    "One-factor: P = U = BR",
                    "Independent three factor")
rownames(results_CFA EMC) <- NULL
results_CFA EMC <- cbind(Model = modelNames_CFA, results_CFA EMC)
results_CFA EMC

```

```

##           Model npar  chisq df pvalue      aic  srmr   cfi   ifi
## 1      Full three-factor   21 26.107 24  0.348 1199.566 0.080 0.961 0.968
## 2      Two-factor: P = U   20 40.181 25  0.028 1211.639 0.325 0.718 0.766
## 3      Two-factor: P = BR   20 36.784 25  0.061 1208.242 0.266 0.781 0.818
## 4      Two-factor: U = BR   20 47.378 25  0.004 1218.836 0.409 0.584 0.655
## 5      One-factor: P = U = BR 18 47.650 27  0.008 1215.108 0.442 0.616 0.671
## 6 Independent three factor 18 46.143 27  0.012 1213.601 0.169 0.644 0.695
##      nfi rmsea      bic
## 1 0.709 0.043 1238.861
## 2 0.553 0.112 1249.063
## 3 0.590 0.099 1245.666
## 4 0.472 0.137 1256.260
## 5 0.469 0.126 1248.790
## 6 0.486 0.122 1247.283

```

8.1.2 Model comparisson

```
anova(fit_m1 EMC, fit_m2 EMC)
```

```

##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_m1 EMC 24 1199.6 1238.9 26.108
## fit_m2 EMC 25 1211.6 1249.1 40.181      14.074 0.52189      1 0.0001758 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

```
anova(fit_m1 EMC, fit_m3 EMC)
```

```

##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_m1 EMC 24 1199.6 1238.9 26.108
## fit_m3 EMC 25 1208.2 1245.7 36.784      10.676 0.44899      1 0.001085 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

```
anova(fit_m1 EMC, fit_m4 EMC)
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff    RMSEA Df diff Pr(>Chisq)
## fit_m1 EMC 24 1199.6 1238.9 26.108
## fit_m4 EMC 25 1218.8 1256.3 47.378      21.27 0.64984      1 3.989e-06 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
anova(fit_m1 EMC, fit_m5 EMC)
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff    RMSEA Df diff Pr(>Chisq)
## fit_m1 EMC 24 1199.6 1238.9 26.108
## fit_m5 EMC 27 1215.1 1248.8 47.650      21.543 0.35885      3 8.12e-05 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
#anova(fit_m1 EMC, fit_m6 EMC)
```

8.1.3 Favoured model

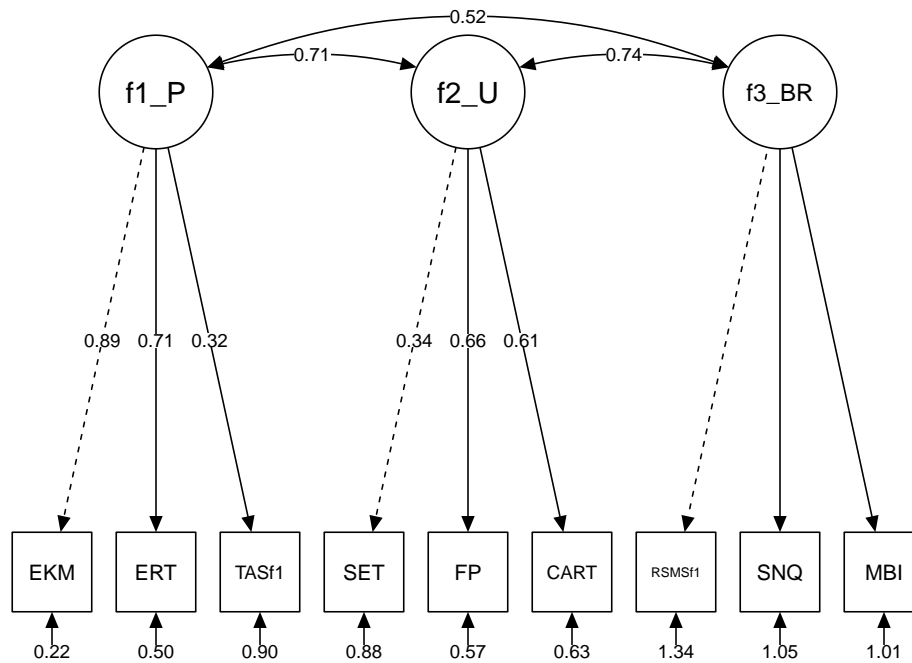
```
stcoefs_fav_CFA EMC <- standardizedSolution(fit_m1 EMC)
nice_table(stcoefs_fav_CFA EMC)
```

```
## Warning: fonts used in 'flextable' are ignored because the 'pdflatex' engine is
## used and not 'xelatex' or 'lualatex'. You can avoid this warning by using the
## 'set_flextable_defaults(fonts_ignore=TRUE)' command or use a compatible engine
## by defining 'latex_engine: xelatex' in the YAML header of the R Markdown
## document.
```

lhs	op	rhs	est.std	se	z	p	ci.lower	ci.upper
f1_P	=~	EKM	0.89	0.10	8.73	< .001***	0.69	1.08
f1_P	=~	ERT	0.71	0.11	6.68	< .001***	0.50	0.92
f1_P	=~	TASf1	0.32	0.14	2.21	.027*	0.04	0.60
f2_U	=~	SET	0.34	0.16	2.21	.027*	0.04	0.65
f2_U	=~	FP	0.66	0.13	5.08	< .001***	0.40	0.91
f2_U	=~	CART	0.61	0.13	4.64	< .001***	0.35	0.86
f3_BR	=~	RSMSf1						
f3_BR	=~	SNQ						

lhs	op	rhs	est.	std	se	z	p	ci.lower	ci.upper
f3_BR	=~	MBI							
f1_P	~~	f2_U	0.71	0.15	4.63	< .001***		0.41	1.02
f1_P	~~	f3_BR	0.52	0.44	1.19	.235		-0.34	1.39
f2_U	~~	f3_BR	0.74	0.59	1.25	.210		-0.42	1.89
EKM	~~	EKM	0.22	0.18	1.21	.227		-0.14	0.57
ERT	~~	ERT	0.50	0.15	3.30	.001***		0.20	0.79
TASf1	~~	TASf1	0.90	0.09	9.77	< .001***		0.72	1.08
SET	~~	SET	0.88	0.11	8.24	< .001***		0.67	1.09
FP	~~	FP	0.57	0.17	3.38	.001***		0.24	0.90
CART	~~	CART	0.63	0.16	3.98	< .001***		0.32	0.94
RSMSf1	~~	RSMSf1	1.34	0.53	2.52	.012*		0.30	2.38
SNQ	~~	SNQ	1.05	0.07	16.03	< .001***		0.92	1.18
MBI	~~	MBI	1.01	0.03	34.63	< .001***		0.96	1.07
f1_P	~~	f1_P	1.00	0.00				1.00	1.00
f2_U	~~	f2_U	1.00	0.00				1.00	1.00
f3_BR	~~	f3_BR							

```
#print(nice_table(stcoefs_fav_CFA EMC), preview = "docx") #Word format output
semPaths(fit_m1 EMC,
  whatLabels = "std", # Display standardized estimates
  layout = "tree2", # Choose a layout, "tree" is one option
  style = "lisrel", # Style for the diagram
  nCharNodes = 0, # Full variable names
  sizeMan = 7, # Size of manifest variables
  sizeLat = 10, # Size of latent variables
  edge.label.cex = 0.75, # Adjust the size of the edge labels
  label.cex = 0.8, # Adjust the size of the node labels
  residuals = TRUE, # Show residuals
  intercepts = FALSE, # Hide intercepts
  thresholds = FALSE,
  fade = FALSE,
  edge.color = "black"
)
```



8.2 CFA pooled data

8.2.1 Fit indices

```
results_CFA_pld <- data.frame(round(rbind(fm_m1_pld, fm_m2_pld, fm_m3_pld,
                                         fm_m4_pld, fm_m5_pld, fm_m6_pld), 3))
modelNames_CFA <- c("Full three-factor",
                    "Two-factor: P = U",
                    "Two-factor: P = BR",
                    "Two-factor: U = BR",
                    "One-factor: P = U = BR",
                    "Independent three factor")
rownames(results_CFA_pld) <- NULL
results_CFA_pld <- cbind(Model = modelNames_CFA, results_CFA_pld)
results_CFA_pld
```

##	Model	npar	chisq	df	pvalue	aic	srmr	cfi	ifi
## 1	Full three-factor	21	23.099	24	0.514	3203.035	0.052	1.000	1.007
## 2	Two-factor: P = U	20	62.850	25	0.000	3240.786	0.305	0.661	0.692
## 3	Two-factor: P = BR	20	56.667	25	0.000	3234.604	0.249	0.717	0.742
## 4	Two-factor: U = BR	20	71.317	25	0.000	3249.254	0.303	0.586	0.623
## 5	One-factor: P = U = BR	18	77.097	27	0.000	3251.034	0.395	0.552	0.585
## 6	Independent three factor	18	63.122	27	0.000	3237.059	0.126	0.677	0.701

```
##      nfi rmsea      bic
## 1 0.844 0.000 3263.091
## 2 0.575 0.108 3297.983
## 3 0.617 0.099 3291.800
## 4 0.517 0.120 3306.450
## 5 0.478 0.120 3302.510
## 6 0.573 0.102 3288.536
```

8.2.2 Model comparisson

```
anova(fit_m1_pld, fit_m2_pld)
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_m1_pld 24 3203.0 3263.1 23.099
## fit_m2_pld 25 3240.8 3298.0 62.850      39.751 0.54809      1 2.885e-10 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
anova(fit_m1_pld, fit_m3_pld)
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_m1_pld 24 3203.0 3263.1 23.099
## fit_m3_pld 25 3234.6 3291.8 56.667      33.569 0.50246      1 6.88e-09 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
anova(fit_m1_pld, fit_m4_pld)
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_m1_pld 24 3203.0 3263.1 23.099
## fit_m4_pld 25 3249.3 3306.5 71.317      48.219 0.60501      1 3.812e-12 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
anova(fit_m1_pld, fit_m5_pld)
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
```

```
## fit_m1_pld 24 3203 3263.1 23.099
## fit_m5_pld 27 3251 3302.5 77.097      53.998 0.36301      3 1.123e-11 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
anova(fit_m1_pld, fit_m6_pld)
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_m1_pld 24 3203.0 3263.1 23.099
## fit_m6_pld 27 3237.1 3288.5 63.122      40.024 0.3093      3 1.053e-08 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
# anova(fit_m2_pld, fit_m5_pld)
# anova(fit_m3_pld, fit_m5_pld)
# anova(fit_m4_pld, fit_m5_pld)
```

8.2.3 Favoured model

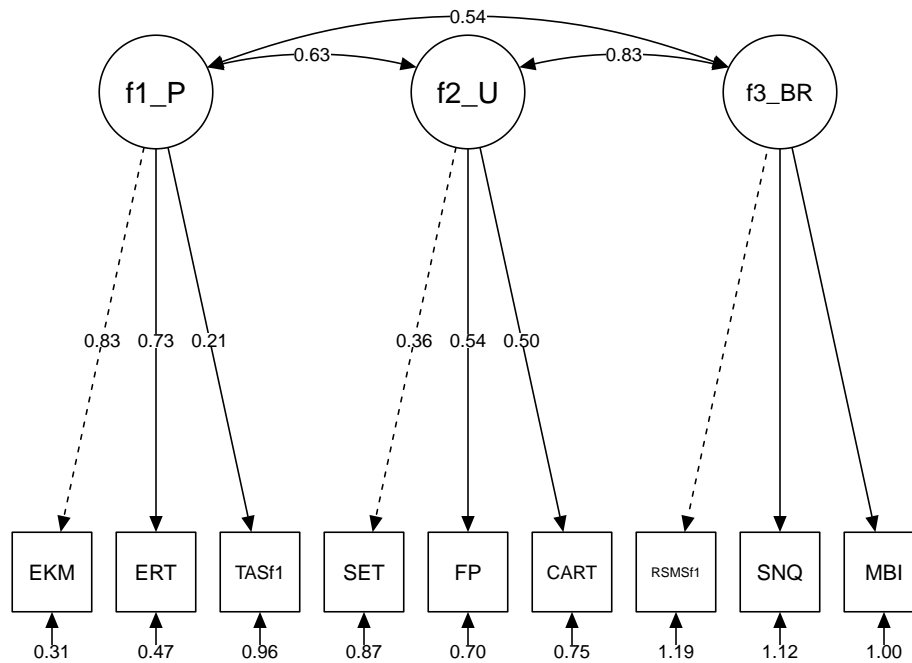
```
stcoefs_fav_CFA_pld <- standardizedSolution(fit_m1_pld)
stcoefs_fav_CFA_pld_uns <- parameterEstimates(fit_m1_pld, standardized = TRUE)
nice_table(stcoefs_fav_CFA_pld)
```

```
## Warning: fonts used in 'flextable' are ignored because the 'pdflatex' engine is
## used and not 'xelatex' or 'lualatex'. You can avoid this warning by using the
## 'set_flextable_defaults(fonts_ignore=TRUE)' command or use a compatible engine
## by defining 'latex_engine: xelatex' in the YAML header of the R Markdown
## document.
```

lhs	op	rhs	est.std	se	z	p	ci.lower	ci.upper
f1_P	==	EKM	0.83	0.08	10.20	< .001***	0.67	0.99
f1_P	==	ERT	0.73	0.08	9.16	< .001***	0.57	0.88
f1_P	==	TASf1	0.21	0.10	2.17	.030*	0.02	0.40
f2_U	==	SET	0.36	0.10	3.52	< .001***	0.16	0.57
f2_U	==	FP	0.54	0.10	5.45	< .001***	0.35	0.74
f2_U	==	CART	0.50	0.10	5.05	< .001***	0.31	0.70
f3_BR	==	RSMSf1						
f3_BR	==	SNQ						
f3_BR	==	MBI						
f1_P	~~	f2_U	0.63	0.12	5.00	< .001***	0.38	0.87

lhs	op	rhs	est.std	se	z	p	ci.lower	ci.upper
f1_P	~~	f3_BR	0.54	0.25	2.16	.031*	0.05	1.02
f2_U	~~	f3_BR	0.83	0.35	2.35	.019*	0.14	1.52
EKM	~~	EKM	0.31	0.14	2.24	.025*	0.04	0.57
ERT	~~	ERT	0.47	0.12	4.11	< .001***	0.25	0.70
TASf1	~~	TASf1	0.96	0.04	23.99	< .001***	0.88	1.03
SET	~~	SET	0.87	0.08	11.54	< .001***	0.72	1.02
FP	~~	FP	0.70	0.11	6.51	< .001***	0.49	0.92
CART	~~	CART	0.75	0.10	7.51	< .001***	0.55	0.94
RSMSf1	~~	RSMSf1	1.19	0.15	8.18	< .001***	0.90	1.47
SNQ	~~	SNQ	1.12	0.09	12.30	< .001***	0.95	1.30
MBI	~~	MBI	1.00	0.01	117.24	< .001***	0.99	1.02
f1_P	~~	f1_P	1.00	0.00			1.00	1.00
f2_U	~~	f2_U	1.00	0.00			1.00	1.00
f3_BR	~~	f3_BR						

```
#print(nice_table(stcoefs_fav_CFA_pld), preview = "docx") #Word format output
semPaths(fit_m1_pld,
  whatLabels = "std", # Display standardized estimates
  layout = "tree2", # Choose a layout, "tree" is one option
  style = "lisrel", # Style for the diagram
  nCharNodes = 0, # Full variable names
  sizeMan = 7, # Size of manifest variables
  sizeLat = 10, # Size of latent variables
  edge.label.cex = 0.75, # Adjust the size of the edge labels
  label.cex = 0.8, # Adjust the size of the node labels
  residuals = TRUE, # Show residuals
  intercepts = FALSE, # Hide intercepts
  thresholds = FALSE,
  fade = FALSE,
  edge.color = "black"
)
```



8.3 SEM EMC data

8.3.1 Fit indices

```
results_SEM <- data.frame(round(rbind(fm_sem1, fm_sem2, fm_sem3, fm_sem4,
                                     fm_sem5, fm_sem6, fm_sem7, fm_sem8), 3))
modelNames_SEM <- c("3p", "2p: U & BR", "2p: P & BR", "2p: P & U",
                    "1p: P", "1p: U", "1p: BR", "0p")
rownames(results_SEM) <- NULL
results_SEM <- cbind(Model = modelNames_SEM, results_SEM)
#results_SEM
results_SEM[,c("Model", "npar", "df", "chisq", "pvalue", "aic", "srmr", "cfi", "ifi")]
```

##	Model	npar	df	chisq	pvalue	aic	srmr	cfi	ifi
## 1	3p	25	30	29.976	0.467	1332.234	0.081	1.000	1.000
## 2	2p: U & BR	24	31	31.796	0.427	1332.053	0.080	0.987	0.989
## 3	2p: P & BR	24	31	29.977	0.519	1330.234	0.081	1.000	1.014
## 4	2p: P & U	24	31	30.384	0.498	1330.641	0.081	1.000	1.008
## 5	1p: P	23	32	30.390	0.548	1328.647	0.081	1.000	1.022
## 6	1p: U	23	32	32.932	0.421	1331.189	0.082	0.984	0.987
## 7	1p: BR	23	32	32.233	0.455	1330.490	0.085	0.996	0.997
## 8	0p	22	33	40.516	0.173	1336.773	0.119	0.873	0.894


```
#print(nice_table(results_SEM[,c("Model", "npar", "df", "chisq", "pvalue", "aic", "srmr", "cfi", "ifi")
```

8.3.2 Model comparisson

```
anova(fit_sem1, fit_sem2) #not significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC   BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_sem1 30 1332.2 1379 29.976
## fit_sem2 31 1332.0 1377 31.796      1.8199 0.1307      1      0.1773
```

```
anova(fit_sem1, fit_sem3) #not significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC   BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_sem1 30 1332.2 1379.0 29.976
## fit_sem3 31 1330.2 1375.1 29.977 0.00021022      0      1      0.9884
```

```
anova(fit_sem1, fit_sem4) #not significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC   BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_sem1 30 1332.2 1379.0 29.976
## fit_sem4 31 1330.6 1375.5 30.384      0.40759      0      1      0.5232
```

```
#So: models 2-4 are not fitting sign than model 1
```

```
anova(fit_sem1, fit_sem5) #not significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC   BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_sem1 30 1332.2 1379.0 29.976
## fit_sem5 32 1328.7 1371.7 30.390      0.41326      0      2      0.8133
```

```
anova(fit_sem1, fit_sem6) #not significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC   BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_sem1 30 1332.2 1379.0 29.976
## fit_sem6 32 1331.2 1374.2 32.932      2.9556 0.099768      2      0.2281
```

```
anova(fit_sem1, fit_sem7) #not significant
```

```
##  
## Chi-Squared Difference Test  
##  
##           Df      AIC      BIC  Chisq Chisq diff    RMSEA Df diff Pr(>Chisq)  
## fit_sem1 30 1332.2 1379.0 29.976  
## fit_sem7 32 1330.5 1373.5 32.233      2.2566 0.051699      2      0.3236
```

#So: models 5-7 are not fitting sign than model 1

```
anova(fit_sem2, fit_sem6) #not significant
```

```
##  
## Chi-Squared Difference Test  
##  
##           Df      AIC      BIC  Chisq Chisq diff    RMSEA Df diff Pr(>Chisq)  
## fit_sem2 31 1332.0 1377.0 31.796  
## fit_sem6 32 1331.2 1374.2 32.932      1.1357 0.053162      1      0.2866
```

```
anova(fit_sem2, fit_sem7) #not significant
```

```
##  
## Chi-Squared Difference Test  
##  
##           Df      AIC      BIC  Chisq Chisq diff    RMSEA Df diff Pr(>Chisq)  
## fit_sem2 31 1332.0 1377.0 31.796  
## fit_sem7 32 1330.5 1373.5 32.233      0.43669      0      1      0.5087
```

#So: models 6-7 are not fitting sign than model 2

```
anova(fit_sem3, fit_sem5) #not significant
```

```
##  
## Chi-Squared Difference Test  
##  
##           Df      AIC      BIC  Chisq Chisq diff    RMSEA Df diff Pr(>Chisq)  
## fit_sem3 31 1330.2 1375.1 29.977  
## fit_sem5 32 1328.7 1371.7 30.390      0.41305      0      1      0.5204
```

```
anova(fit_sem3, fit_sem7) #not significant
```

```
##  
## Chi-Squared Difference Test  
##  
##           Df      AIC      BIC  Chisq Chisq diff    RMSEA Df diff Pr(>Chisq)  
## fit_sem3 31 1330.2 1375.1 29.977  
## fit_sem7 32 1330.5 1373.5 32.233      2.2564 0.16179      1      0.1331
```

```
#So: models 5 & 7 are not fitting sign than model 3
```

```
anova(fit_sem4, fit_sem5) #not significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff RMSEA Df diff Pr(>Chisq)
## fit_sem4 31 1330.6 1375.5 30.384
## fit_sem5 32 1328.7 1371.7 30.390  0.0056785      0      1      0.9399
```

```
anova(fit_sem4, fit_sem6) #not significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_sem4 31 1330.6 1375.5 30.384
## fit_sem6 32 1331.2 1374.2 32.932      2.548 0.17958      1      0.1104
```

```
#So: models 5-6 are not fitting sign than model 4
```

```
anova(fit_sem5, fit_sem8) #significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_sem5 32 1328.7 1371.7 30.390
## fit_sem8 33 1336.8 1377.9 40.516      10.126 0.43603      1  0.001462 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
anova(fit_sem6, fit_sem8) #significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
## fit_sem6 32 1331.2 1374.2 32.932
## fit_sem8 33 1336.8 1377.9 40.516      7.5838 0.37035      1  0.00589 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
anova(fit_sem7, fit_sem8) #significant
```

```
##
## Chi-Squared Difference Test
##
##           Df      AIC      BIC  Chisq Chisq diff  RMSEA Df diff Pr(>Chisq)
```

```
## fit_sem7 32 1330.5 1373.5 32.233
## fit_sem8 33 1336.8 1377.9 40.516      8.2827 0.38952      1 0.004002 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

#So, model 8 fits significantly worse than models 5-7

#Out of models 5-7, model 5 is in favour based on the fit indices

8.3.3 Favoured model

```
stcoefs_fav_SEM <- standardizedSolution(fit_sem5)
#parameterestimates(fit_sem5, standardized = TRUE)[-10] #Unstandardized and standardized
nice_table(stcoefs_fav_SEM)
```

```
## Warning: fonts used in 'flextable' are ignored because the 'pdflatex' engine is
## used and not 'xelatex' or 'lualatex'. You can avoid this warning by using the
## 'set_flextable_defaults(fonts_ignore=TRUE)' command or use a compatible engine
## by defining 'latex_engine: xelatex' in the YAML header of the R Markdown
## document.
```

lhs	op	rhs	est.std	se	z	p	ci.lower	ci.upper
f1_P	=~	EKM	0.93	0.09	10.87	< .001***	0.76	1.09
f1_P	=~	ERT	0.68	0.10	6.76	< .001***	0.48	0.88
f1_P	=~	TASf1	0.31	0.14	2.24	.025*	0.04	0.59
f2_U	=~	SET	0.34	0.16	2.22	.027*	0.04	0.65
f2_U	=~	FP	0.65	0.13	5.05	< .001***	0.40	0.91
f2_U	=~	CART	0.61	0.13	4.66	< .001***	0.35	0.87
f3_BR	=~	RSMSf1						
f3_BR	=~	SNQ						
f3_BR	=~	MBI						
TQ	~	f1_P	0.47	0.13	3.74	< .001***	0.22	0.71
TQ	~	f2_U	0.00	0.00			0.00	0.00
TQ	~	f3_BR						
f1_P	~~	f2_U	0.69	0.15	4.58	< .001***	0.39	0.98
f1_P	~~	f3_BR	0.49	0.42	1.15	.250	-0.34	1.31
f2_U	~~	f3_BR	0.71	0.59	1.22	.224	-0.44	1.86
EKM	~~	EKM	0.14	0.16	0.92	.359	-0.16	0.45
ERT	~~	ERT	0.54	0.14	3.97	< .001***	0.27	0.81
TASf1	~~	TASf1	0.90	0.09	10.18	< .001***	0.73	1.07
SET	~~	SET	0.88	0.11	8.21	< .001***	0.67	1.09

lhs	op	rhs	est.std	se	z	p	ci.lower	ci.upper
FP	~~	FP	0.57	0.17	3.41	.001***	0.24	0.90
CART	~~	CART	0.63	0.16	3.94	< .001***	0.32	0.94
RSMSf1	~~	RSMSf1	1.37	0.61	2.26	.024*	0.18	2.56
SNQ	~~	SNQ	1.05	0.06	16.66	< .001***	0.92	1.17
MBI	~~	MBI	1.01	0.03	36.93	< .001***	0.96	1.07
TQ	~~	TQ	0.78	0.12	6.65	< .001***	0.55	1.01
f1_P	~~	f1_P	1.00	0.00			1.00	1.00
f2_U	~~	f2_U	1.00	0.00			1.00	1.00
f3_BR	~~	f3_BR						

```
#print(nice_table(stcoefs_fav_SEM), preview = "docx") #Word format output
semPaths(fit_sem5,
  whatLabels = "std", # Display standardized estimates
  layout = "tree2", # Choose a layout, "tree" is one option
  style = "lisrel", # Style for the diagram
  nCharNodes = 0, # Full variable names
  sizeMan = 7, # Size of manifest variables
  sizeLat = 10, # Size of latent variables
  edge.label.cex = 0.75, # Adjust the size of the edge labels
  label.cex = 0.8, # Adjust the size of the node labels
  residuals = TRUE, # Show residuals
  intercepts = FALSE, # Hide intercepts
  thresholds = FALSE,
  fade = FALSE,
  edge.color = "black"
)
```

