

# Reverse mapping of CustumArray oligos to original proteins

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This workflow aligns the short oligos from the CustomArray order to the full reference sequences using Bowtie2. This enables the mapping to all genes sharing the same sequence.

```
suppressPackageStartupMessages(library(knitr))
```

## Load sequences

```
LUT.dna <- read.table("data/SortedFragments_all.txt", header = TRUE, skip = 0,
  sep = "\t", stringsAsFactors = FALSE, fill = TRUE)
LUT.dna <- data.table(LUT.dna)
```

## Remove constitutive backbone sequences

```
invisible(LUT.dna[, `:=`(Sequence, gsub("aacctccagagaggcaacg", "", Sequence))])
invisible(LUT.dna[, `:=`(Sequence, gsub("cagacaagcagctaccgca", "", Sequence))])
invisible(LUT.dna[, `:=`(Sequence, toupper(Sequence))])
setkey(LUT.dna, "Sequence")
LUT.dna <- unique(LUT.dna)
LUT.dna$Names <- LUT.dna$Sequence
LUT.dna$LUTnr <- make.names(seq(nrow(LUT.dna)), unique = TRUE)
```

## Split sequences based on linker and length

```
LUT.14aaG4S <- LUT.dna[substr(LUT.dna$Sequence, 1, 14) == "GAGGCGGAGGAAGT"]
LUT.remaining <- LUT.dna[!(substr(LUT.dna$Sequence, 1, 14) == "GAGGCGGAGGAAGT")]
LUT.14aaA5 <- LUT.remaining[substr(LUT.remaining$Sequence, 1, 14) == "CTGCTGCAGCAGCC"]
LUT.remaining <- LUT.remaining[!(substr(LUT.remaining$Sequence, 1, 14) == "CTGCTGCAGCAGCC")]
LUT.22aa <- LUT.remaining[nchar(LUT.remaining$Sequence) == 70L & substr(LUT.remaining$Sequence,
  1, 2) == "CT"]
LUT.remaining <- LUT.remaining[!(nchar(LUT.remaining$Sequence) == 70L & substr(LUT.remaining$Sequence,
  1, 2) == "CT")]
LUT.14aa <- LUT.remaining[nchar(LUT.remaining$Sequence) == 46L & substr(LUT.remaining$Sequence,
  1, 2) == "CT"]
rm(LUT.remaining)
LUT.dna[LUT.dna$Sequence %in% LUT.14aaG4S$Sequence, "Structure"] <- "14aaG4S"
LUT.dna[LUT.dna$Sequence %in% LUT.14aaA5$Sequence, "Structure"] <- "14aaA5"
LUT.dna[LUT.dna$Sequence %in% LUT.22aa$Sequence, "Structure"] <- "22aa"
LUT.dna[LUT.dna$Sequence %in% LUT.14aa$Sequence, "Structure"] <- "14aa"

save(LUT.dna, file = "data/LUTdna.rda")
```

## Trim sequences

```
LUT.14aa$Sequence <- substr(LUT.14aa$Sequence, 3, 44)
LUT.14aaG4S$Sequence <- substr(LUT.14aaG4S$Sequence, 15, 56)
LUT.14aaA5$Sequence <- substr(LUT.14aaA5$Sequence, 15, 56)
LUT.22aa$Sequence <- substr(LUT.22aa$Sequence, 3, 68)
```

## Save fasta files for Bowtie alignments

```
LUT.14aa.fa <- tempfile(pattern = "LUT_14aa_", tmpdir = tmpdir(), fileext = ".fa")
LUT.14aa.seq = ShortRead(DNAStringSet(LUT.14aa$Sequence), BStringSet(LUT.14aa$LUTnr))
writeFasta(LUT.14aa.seq, LUT.14aa.fa)

LUT.14aaG4S.fa <- tempfile(pattern = "LUT_14aaG4s_", tmpdir = tmpdir(), fileext = ".fa")
LUT.14aaG4S.seq = ShortRead(DNAStringSet(LUT.14aaG4S$Sequence), BStringSet(LUT.14aaG4S$LUTnr))
writeFasta(LUT.14aaG4S.seq, LUT.14aaG4S.fa)

LUT.14aaA5.fa <- tempfile(pattern = "LUT_14aaA5_", tmpdir = tmpdir(), fileext = ".fa")
LUT.14aaA5.seq = ShortRead(DNAStringSet(LUT.14aaA5$Sequence), BStringSet(LUT.14aaA5$LUTnr))
writeFasta(LUT.14aaA5.seq, LUT.14aaA5.fa)

LUT.22aa.fa <- tempfile(pattern = "LUT_14aaA5_", tmpdir = tmpdir(), fileext = ".fa")
LUT.22aa.seq = ShortRead(DNAStringSet(LUT.22aa$Sequence), BStringSet(LUT.22aa$LUTnr))
writeFasta(LUT.22aa.seq, LUT.22aa.fa)
```

## Build Bowtie index

```
seqs.original <- readFasta("input/DNA-lib_RetrogradeTransport.fasta")

seqs.AA <- Biostrings::translate(sread(seqs.original), genetic.code = GENETIC_CODE,
                                if.fuzzy.codon = "error")

source("functions/AAtoDNA.R")
seqs.optimized = ShortRead(DNAStringSet(sapply(seqs.AA, function(x) AAtoDNA(x,
                                species = "hsa"))), BStringSet(gsub("[ ]", "_", ShortRead::id(seqs.original)))))

bowtie.fasta <- tempfile(pattern = "bowtie_", tmpdir = tmpdir(), fileext = ".fa")

writeFasta(seqs.optimized, bowtie.fasta)

bowtie.idx <- tempfile(pattern = "IDX_bowtie_", tmpdir = tmpdir(), fileext = "")

sys.out <- system(paste("bowtie2-build", bowtie.fasta, bowtie.idx, ">&1", sep = " "),
                  intern = TRUE, ignore.stdout = FALSE)
```

## Align fragments to reference

Align 14aa sequences

```
name.bowtie <- tempfile(pattern = "bowtie_", tmpdir = tmpdir(), fileext = "")
```

```

sys.out <- system(paste("bowtie2 --non-deterministic --threads ", detectCores(),
  "--very-sensitive -f -a", " -x ", bowtie.idx, " -U ", LUT.14aa.fa, " -S ",
  name.bowtie, ".sam 2>&1", sep = ""), intern = TRUE, ignore.stdout = FALSE)

sys.out <- as.data.frame(sys.out)

colnames(sys.out) <- c("Bowtie 2 alignment to library")
invisible(sys.out[" "] <- " ")
lengthOut <- (nrow(sys.out))
knitr::kable(sys.out[1:lengthOut, ], format = "latex", booktabs = T) %>% kable_styling(latex_options = "str

```

Bowtie 2 alignment to library
44705 reads; of these:
44705 (100.00%) were unpaired; of these:
0 (0.00%) aligned 0 times
27420 (61.34%) aligned exactly 1 time
17285 (38.66%) aligned >1 times
100.00% overall alignment rate

```

command.args <- paste("view -@ ", detectCores(), " -Sb ", name.bowtie, ".sam > ",
  name.bowtie, ".bam", sep = "")
system2("/usr/local/bin/samtools", args = command.args, stdout = TRUE, stderr = TRUE)

```

```
character(0)
```

```

command.args <- paste("sort -@ ", detectCores(), " ", name.bowtie, ".bam -o ",
  name.bowtie, "_sort.bam", sep = "")
system2("/usr/local/bin/samtools", args = command.args, stdout = TRUE, stderr = TRUE)

```

```
character(0)
```

```

frag14aa.ranges <- readGAlignments(paste(name.bowtie, "_sort.bam", sep = ""),
  use.names = TRUE)
length(names(frag14aa.ranges))

```

```
[1] 75152
```

```
length(unique(names(frag14aa.ranges)))
```

```
[1] 44705
```

```
length(unique(LUT.14aa$Sequence))
```

```
[1] 44705
```

Align 14aaG4S sequences

```
name.bowtie <- tempfile(pattern = "bowtie_", tmpdir = tempdir(), fileext = "")
```

```

sys.out <- system(paste("bowtie2 --non-deterministic --threads ", detectCores(),
  "--very-sensitive -f -a", " -x ", bowtie.idx, " -U ", LUT.14aaG4S.fa, " -S ",
  name.bowtie, ".sam 2>&1", sep = ""), intern = TRUE, ignore.stdout = FALSE)

```

```
sys.out <- as.data.frame(sys.out)
```

```

colnames(sys.out) <- c("Bowtie 2 alignment to library")
invisible(sys.out[" "] <- " ")
lengthOut <- (nrow(sys.out))
knitr::kable(sys.out[1:lengthOut, ], format = "latex", booktabs = T) %>% kable_styling(latex_options = "str

```

---

Bowtie 2 alignment to library
15792 reads; of these:
15792 (100.00%) were unpaired; of these:
0 (0.00%) aligned 0 times
9150 (57.94%) aligned exactly 1 time
6642 (42.06%) aligned >1 times
100.00% overall alignment rate

---

```
command.args <- paste("view -@ ", detectCores(), " -Sb ", name.bowtie, ".sam > ",
  name.bowtie, ".bam", sep = "")
system2("/usr/local/bin/samtools", args = command.args, stdout = TRUE, stderr = TRUE)
```

```
character(0)
```

```
command.args <- paste("sort -@ ", detectCores(), " ", name.bowtie, ".bam -o ",
  name.bowtie, "_sort.bam", sep = "")
system2("/usr/local/bin/samtools", args = command.args, stdout = TRUE, stderr = TRUE)
```

```
character(0)
```

```
frag14aaG4S.ranges <- readGAlignments(paste(name.bowtie, "_sort.bam", sep = ""),
  use.names = TRUE)
length(names(frag14aaG4S.ranges))
```

```
[1] 27778
```

```
length(unique(names(frag14aaG4S.ranges)))
```

```
[1] 15792
```

```
length(unique(LUT.14aaG4S$Sequence))
```

```
[1] 15792
```

Align 14aaA5 sequences

```
name.bowtie <- tempfile(pattern = "bowtie_", tmpdir = tempdir(), fileext = "")
```

```
sys.out <- system(paste("bowtie2 --non-deterministic --threads ", detectCores(),
  " --very-sensitive -f -a", " -x ", bowtie.idx, " -U ", LUT.14aaA5.fa, " -S ",
  name.bowtie, ".sam 2>&1", sep = ""), intern = TRUE, ignore.stdout = FALSE)
```

```
sys.out <- as.data.frame(sys.out)
```

```
colnames(sys.out) <- c("Bowtie 2 alignment to library")
```

```
invisible(sys.out[" "] <- " ")
```

```
lengthOut <- (nrow(sys.out))
```

```
knitr::kable(sys.out[1:lengthOut, ], format = "latex", booktabs = T) %>% kable_styling(latex_options = "str
```

---

Bowtie 2 alignment to library
15792 reads; of these:
15792 (100.00%) were unpaired; of these:
0 (0.00%) aligned 0 times
9150 (57.94%) aligned exactly 1 time
6642 (42.06%) aligned >1 times
100.00% overall alignment rate

---

```
command.args <- paste("view -@ ", detectCores(), " -Sb ", name.bowtie, ".sam > ",
  name.bowtie, ".bam", sep = "")
system2("/usr/local/bin/samtools", args = command.args, stdout = TRUE, stderr = TRUE)
```

```
character(0)
```

```
command.args <- paste("sort -@ ", detectCores(), " ", name.bowtie, ".bam -o ",
  name.bowtie, "_sort.bam", sep = "")
system2("/usr/local/bin/samtools", args = command.args, stdout = TRUE, stderr = TRUE)
```

```
character(0)
```

```
frag14aaA5.ranges <- readGAlignments(paste(name.bowtie, "_sort.bam", sep = ""),
  use.names = TRUE)
length(names(frag14aaA5.ranges))
```

```
[1] 27778
```

```
length(unique(names(frag14aaA5.ranges)))
```

```
[1] 15792
```

```
length(unique(LUT.14aaA5$Sequence))
```

```
[1] 15792
```

```
Align 22aa sequences
```

```
name.bowtie <- tempfile(pattern = "bowtie_", tmpdir = tempdir(), fileext = "")
```

```
sys.out <- system(paste("bowtie2 --non-deterministic --threads ", detectCores(),
  " --very-sensitive -f -a", " -x ", bowtie.idx, " -U ", LUT.22aa.fa, " -S ",
  name.bowtie, ".sam 2>&1", sep = ""), intern = TRUE, ignore.stdout = FALSE)
```

```
sys.out <- as.data.frame(sys.out)
```

```
colnames(sys.out) <- c("Bowtie 2 alignment to library")
```

```
invisible(sys.out[" "] <- " ")
```

```
lengthOut <- (nrow(sys.out))
```

```
knitr::kable(sys.out[1:lengthOut, ], format = "latex", booktabs = T) %>% kable_styling(latex_options = "str
```

Bowtie 2 alignment to library
16054 reads; of these:
16054 (100.00%) were unpaired; of these:
0 (0.00%) aligned 0 times
8730 (54.38%) aligned exactly 1 time
7324 (45.62%) aligned >1 times
100.00% overall alignment rate

```
command.args <- paste("view -@ ", detectCores(), " -Sb ", name.bowtie, ".sam > ",
  name.bowtie, ".bam", sep = "")
system2("/usr/local/bin/samtools", args = command.args, stdout = TRUE, stderr = TRUE)
```

```
character(0)
```

```
command.args <- paste("sort -@ ", detectCores(), " ", name.bowtie, ".bam -o ",
  name.bowtie, "_sort.bam", sep = "")
system2("/usr/local/bin/samtools", args = command.args, stdout = TRUE, stderr = TRUE)
```

```
character(0)
```

```
frag22aa.ranges <- readGAlignments(paste(name.bowtie, "_sort.bam", sep = ""),
  use.names = TRUE)
length(names(frag22aa.ranges))
```

```
[1] 29665
```

```
length(unique(names(frag22aa.ranges)))
```

```
[1] 16054
```

```
length(unique(LUT.22aa$Sequence))
```

```
[1] 16054
```

## Merge and annotate aligned sequences

```
mcols(frag14aa.ranges)$structure <- "14aa"
mcols(frag22aa.ranges)$structure <- "22aa"
mcols(frag14aaA5.ranges)$structure <- "14aaA5"
mcols(frag14aaG4S.ranges)$structure <- "14aaG4S"
allFragments.ranges <- append(frag14aa.ranges, frag22aa.ranges)
allFragments.ranges <- append(allFragments.ranges, frag14aaA5.ranges)
allFragments.ranges <- append(allFragments.ranges, frag14aaG4S.ranges)

mcols(allFragments.ranges)$LUTnr <- names(allFragments.ranges)
setkey(LUT.dna, LUTnr)
mcols(allFragments.ranges)$Sequence <- LUT.dna[mcols(allFragments.ranges)$LUTnr]$Sequence

save(allFragments.ranges, file = "data/alignedLibraries.rda")

devtools::session_info()
```

Session info -----

```
setting  value
version  R version 3.4.2 (2017-09-28)
system   x86_64, linux-gnu
ui        X11
language (EN)
collate   en_US.UTF-8
tz        UTC
date      2020-10-29
```

Packages -----

package	* version	date	source
acepack	1.4.1	2016-10-29	CRAN (R 3.4.2)
ade4	1.7-8	2017-08-09	CRAN (R 3.4.2)
backports	1.1.1	2017-09-25	CRAN (R 3.4.2)
base	* 3.4.2	2017-10-06	local
base64enc	0.1-3	2015-07-28	CRAN (R 3.4.2)
Biobase	* 2.36.2	2017-11-29	Bioconductor
BiocGenerics	* 0.22.1	2017-11-29	Bioconductor
BiocParallel	* 1.10.1	2017-11-29	Bioconductor
Biostrings	* 2.44.2	2017-11-29	Bioconductor
bitops	1.0-6	2013-08-17	CRAN (R 3.4.2)
checkmate	1.8.4	2017-09-25	CRAN (R 3.4.2)

cluster	2.0.6	2017-03-16	CRAN (R 3.4.2)
colorspace	1.3-2	2016-12-14	CRAN (R 3.4.2)
compiler	3.4.2	2017-10-06	local
data.table	* 1.10.4-2	2017-10-12	url
datasets	* 3.4.2	2017-10-06	local
DelayedArray	* 0.2.7	2017-11-29	Bioconductor
devtools	* 1.13.3	2017-08-02	CRAN (R 3.4.2)
digest	0.6.12	2017-01-27	CRAN (R 3.4.2)
evaluate	0.10.1	2017-06-24	CRAN (R 3.4.2)
foreign	0.8-69	2017-06-21	CRAN (R 3.4.2)
formatR	1.5	2017-04-25	CRAN (R 3.4.2)
Formula	* 1.2-2	2017-07-10	CRAN (R 3.4.2)
GeneGA	* 1.26.0	2017-11-29	Bioconductor
GenomeInfoDb	* 1.12.3	2017-11-29	Bioconductor
GenomeInfoDbData	0.99.0	2017-11-29	Bioconductor
GenomicAlignments	* 1.12.2	2017-11-29	Bioconductor
GenomicRanges	* 1.28.6	2017-11-29	Bioconductor
ggplot2	* 2.2.1	2016-12-30	CRAN (R 3.4.2)
graphics	* 3.4.2	2017-10-06	local
grDevices	* 3.4.2	2017-10-06	local
grid	3.4.2	2017-10-06	local
gridExtra	2.3	2017-09-09	CRAN (R 3.4.2)
gtable	0.2.0	2016-02-26	CRAN (R 3.4.2)
hash	* 2.2.6	2013-02-21	CRAN (R 3.4.2)
Hmisc	* 4.0-3	2017-05-02	CRAN (R 3.4.2)
hms	0.3	2016-11-22	CRAN (R 3.4.2)
htmlTable	1.9	2017-01-26	CRAN (R 3.4.2)
htmltools	0.3.6	2017-04-28	CRAN (R 3.4.2)
htmlwidgets	0.9	2017-07-10	CRAN (R 3.4.2)
httr	1.3.1	2017-08-20	CRAN (R 3.4.2)
hwriter	1.3.2	2014-09-10	CRAN (R 3.4.2)
IRanges	* 2.10.5	2017-11-29	Bioconductor
kableExtra	* 0.5.2	2017-09-15	url
knitr	* 1.17	2017-08-10	CRAN (R 3.4.2)
lattice	* 0.20-35	2017-03-25	CRAN (R 3.4.2)
latticeExtra	0.6-28	2016-02-09	CRAN (R 3.4.2)
lazyeval	0.2.0	2016-06-12	CRAN (R 3.4.2)
magrittr	1.5	2014-11-22	CRAN (R 3.4.2)
Matrix	1.2-11	2017-08-21	url
matrixStats	* 0.52.2	2017-04-14	CRAN (R 3.4.2)
memoise	1.1.0	2017-04-21	CRAN (R 3.4.2)
methods	* 3.4.2	2017-10-06	local
munsell	0.4.3	2016-02-13	CRAN (R 3.4.2)
nnet	7.3-12	2016-02-02	CRAN (R 3.4.2)
parallel	* 3.4.2	2017-10-06	local
plyr	1.8.4	2016-06-08	CRAN (R 3.4.2)
R6	2.2.2	2017-06-17	CRAN (R 3.4.2)
RColorBrewer	1.1-2	2014-12-07	CRAN (R 3.4.2)
Rcpp	0.12.13	2017-09-28	url
RCurl	1.95-4.8	2016-03-01	CRAN (R 3.4.2)
readr	1.1.1	2017-05-16	CRAN (R 3.4.2)
rlang	0.1.2	2017-08-09	CRAN (R 3.4.2)
rmarkdown	1.6	2017-06-15	url
rpart	4.1-11	2017-04-21	CRAN (R 3.4.2)
rprojroot	1.2	2017-01-16	CRAN (R 3.4.2)
Rsamtools	* 1.28.0	2017-11-29	Bioconductor
rvest	0.3.2	2016-06-17	CRAN (R 3.4.2)

S4Vectors	* 0.14.7	2017-11-29	Bioconductor
scales	0.5.0	2017-08-24	CRAN (R 3.4.2)
seqinr	* 3.4-5	2017-08-01	CRAN (R 3.4.2)
ShortRead	* 1.34.2	2017-11-29	Bioconductor
splines	3.4.2	2017-10-06	local
stats	* 3.4.2	2017-10-06	local
stats4	* 3.4.2	2017-10-06	local
stringi	1.1.5	2017-04-07	url
stringr	1.2.0	2017-02-18	CRAN (R 3.4.2)
SummarizedExperiment	* 1.6.5	2017-11-29	Bioconductor
survival	* 2.41-3	2017-04-04	CRAN (R 3.4.2)
tibble	1.3.4	2017-08-22	CRAN (R 3.4.2)
tools	3.4.2	2017-10-06	local
utils	* 3.4.2	2017-10-06	local
withr	2.0.0	2017-07-28	url
xml2	1.1.1	2017-01-24	CRAN (R 3.4.2)
XVector	* 0.16.0	2017-11-29	Bioconductor
yaml	2.1.14	2016-11-12	CRAN (R 3.4.2)
zlibbioc	1.22.0	2017-11-29	Bioconductor