

Tom's simple guide to writing a script

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Credit: Dr Gwen Fernandes on helping prepare slides

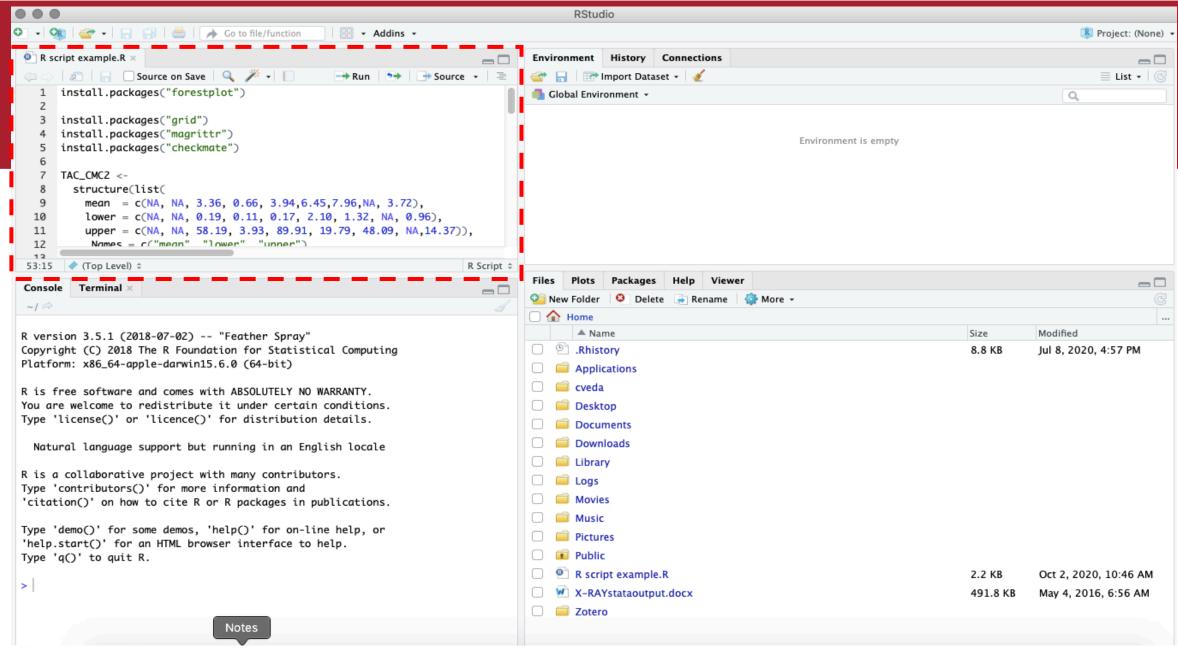


Outline

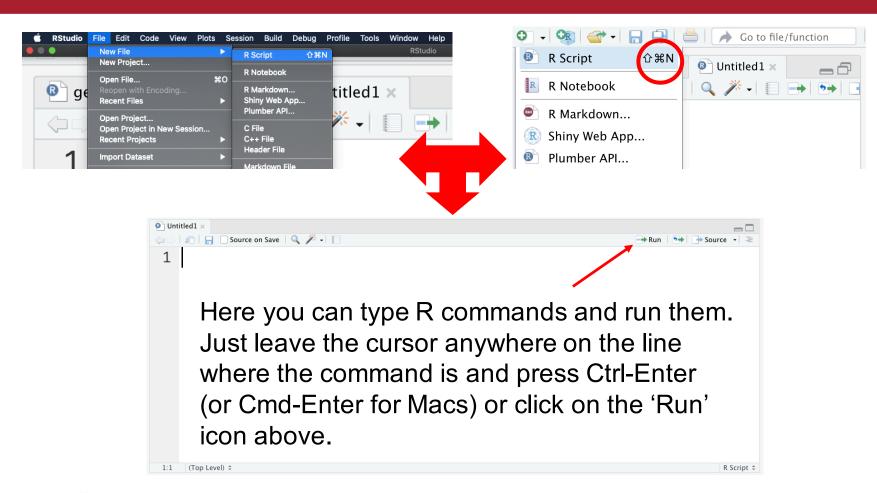
- –Making a new R script
- Commenting your scripts
- -What to think about before starting to script
- –How the top of your script should look
- -Making your code easier to read
- -Saving your scripts
- –Bad/good script comparison

Making a new R script

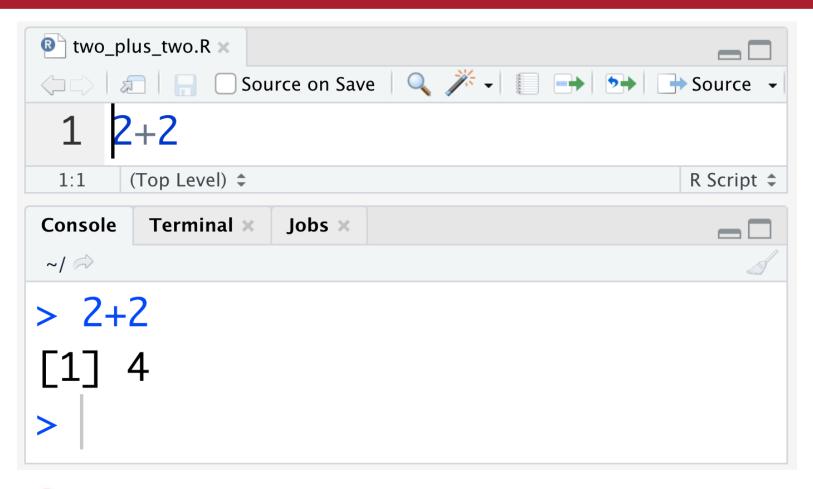
- -The usual R Studio screen has four windows:
 - >CONSOLE
 - >WORKPLACE AND HISTORY
 - >FILES, PLOTS, PACKAGES AND HELP
 - ➤ R SCRIPT AND DATA VIEW (this is where you keep a record of your work. For Stata users, this would be like your do-file, for SPSS users it is like the syntax and for SAS users, the SAS program)



Making a new R script



Making a new R script



Commenting

- -Code with a # before it does not get run
- –This is useful for making your scripts much easier to read!
- Comment on WHY and WHAT (to start with)
 - Start with many comments! Also, use comments to split up the script to make it clearer

RStudio sections

```
13 * # ---- Part C of script ----

14

15 * # ---- Part D of script ----

16

17

18 | Part D of script

Part D of script

R Script $
```

3 steps before starting

1. Why am I writing this script?

2. What do I need to write this script?

3. How am I going to write this script?

Step 1: Why am I writing this script?

- –Think about it and write it down!
 - ➤e.g. clean a dataset OR assess association between x and y
- Give the script a good name, write a descriptive title + add a couple of lines that describe the purpose of the script

```
@ generate_pack_years.R x | Source on Save | Note of Save | Note o
```

Step 2: What do I need to write this script?

- -Datasets!
- -Using only base R can make things difficult... Packages!
- -Packages are made by others and are there to make your life easier

For example, reading data into R can be tricky depending on how the data is stored, but packages can make this easier!

Reading data into R

Depending on what form the data is in, you have to use different functions to read in the data. Data you get may be in:

- –excel spreadsheets
- -comma seperated value (csv) files
- -tab separated value (tsv) files
- -spss files
- -stata files
- -images (e.g. .png files)

```
11 # load packages
12 library(haven)
13
14 # read in data
15 df <- read_dta("my_data.dta")</pre>
```

These need different functions to read them in, some of which are only available with certain packages.

Step 3: How am I going to write this script?

- -Linked to why you are writing it and what you need to write it!
- –Write out each step

```
17 # structure of the script:
18 # 1. Extract the smoking variables
19 # 2. Exclude individuals with withdrawn
20 # consent and too much missing data
21 # 3. Generate pack years variable
22 # 4. Check for outliers
23 # 5. Write out a table with identifiers
24 # and pack years variables
```

Top of the script

```
generate_pack_years.R ×
🗀 🗎 📗 🗌 Source on Save 🔍 🎢 🗸 📗
 2 # Generating pack years in ALSPAC
 3-# -----
 5 # This script extracts smoking variables from
 6 # the mothers within ALSPAC and uses this
 7 # to generate pack years.
 8 # Authors: Thomas Battram, Gwen Fernandes.
 9 # Date: 2020/02/15
10
11 # load packages
12 library(haven)
13
14 # read in data
15 df <- read_dta("my_data.dta")</pre>
16
17 # structure of the script:
18 # 1. Extract the smoking variables
19 # 2. Exclude individuals with withdrawn consent and too much missing data
20 # 3. Generate pack years variable
21 # 4. Check for outliers
22 # 5. Write out a table with identifiers and pack years variables
6:17 (Untitled) $
                                                                          R Script $
```

Make your code easy to read

1.Use a consistent style when writing code

```
a_{var} \leftarrow c(1, 2, 3)

a_{var} \leftarrow c(1, 2, 3)

a_{var} \leftarrow c(1, 2, 3)
```

2.Use spaces appropriately

```
x \leftarrow c(1,2,51,124,4124)

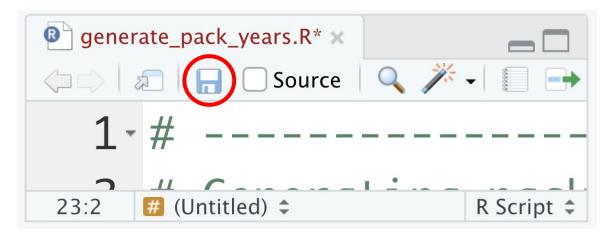
x \leftarrow c(1, 2, 51, 124, 4124)

x \leftarrow c(1, 2, 51, 124, 4124)
```

3.Use indents appropriatelya) No long lines of code!

Saving scripts

- It's important to save scripts as you go and you can always come back to them and send them to other people.
- –Give the script a good name and save it regularly!



CTRL (or CMD) + S

Summary

- 3 questions to think about before writing a script:
 - 1. Why am I writing this script?
 - 2. What do I need to write this script?
 - 3. How am I going to write this script?
 - –Comment your code a lot (using #)
 - Make your code easy to read
 - -Save your scripts regularly

Bad Script

- 1. No title
- 2. No description
- 3. No sections
- No comments at all!
- 5. Spacing could potentially be improved

```
setwd("a_directory")
    data_cveda <- read.delim("data_cveda.txt")
     data_cveda
     names(data cveda)
     data cveda$ageband
     data_cveda$child<- data_cveda$ageband==1
     summary
     mean(data cveda$p sdq tot, na.rm=TRUE)
     sd(data_cveda$p_sdq_tot,na.rm=TRUE)
     median(data_cveda$p_sdq_tot, na.rm=TRUE)
     IQR(data_cveda$p_sdq_tot,
                                  na.rm=TRUE)
     mad(data_cveda$p_sdq_tot,na.rm=TRUE)
     min(data_cveda$p_sdq_tot,na.rm=TRUE)
     max(data_cveda$p_sdq_tot, na.rm=TRUE)
     range(data_cveda$p_sdq_tot, na.rm=TRUE)
    summary(data_cveda$p_sdq_tot, na.rm=TRUE)
20
     d<- data_cveda[data_cveda$ageband==1,]</pre>
    mean(d$p_sdq_tot, na.rm=TRUE)
    sd(d$p_sdq_tot, na.rm=TRUE)
    median(d$p_sdq_tot, na.rm=TRUE)
    IQR(d$p_sdq_tot,na.rm=TRUE)
    mad(d$p_sdq_tot, na.rm=TRUE)
    min(d$p_sdq_tot, na.rm=TRUE)
    max(d$p_sdq_tot, na.rm=TRUE)
     range(d$p sdq tot, na.rm=TRUE)
     summary(d$p_sdq_tot, na.rm=TRUE)
31
32
     cor(data_cveda$p_sdq_emotion, data_cveda$p_sdq_conduct, use= "pairwise.complete.obs")
     table(data_cveda$depanx_1stdeg)
     table(data_cveda$depanx_1stdeg, data_cveda$sex)
```

Good Script

```
# cVEDA: Association between parental punishment and behaviour
       This script assesses the association between parental corporal punishment
    # and total sdq (Strength and Difficulty Questionnaire) score within the cveda cohort
     # Date: 2020-10-21
     # Set directory and import data and save as data_cveda
     setwd("a_directory")
    data_cveda <- read.delim("data_cveda.txt")</pre>
13
    # Examine the data and add a variable for children
     # list the variables in data_cveda
    names(data_cveda)
     ### The names can be used to refer to the relevant variables by using the $ symbol.
    data_cveda$ageband
     ### Add which individuals in the dataset are children or in ageband=1
    # Code below gives you the number of children included in the study.
    # An additional T/F column is added to the data_cveda data frame ###
    data_cveda$child <- data_cveda$ageband == 1
     summary(data_cveda$child)
    # 1918 children in the dataset
    # Examine the statistical qualities of the data
    mean(data_cveda$p_sdq_tot, na.rm = TRUE)
    sd(data_cveda$p_sdq_tot. na_rm__
    median(data_cveda$p_sdq_tot, na.rm = TRUE)
     IQR(data_cveda$p_sdq_tot, na.rm = TRUE) # interquartile range
    mad(data_cveda$p_sdq_tot, na.rm = TRUE) # mean absolute deviation
    min(data_cveda$p_sdq_tot, na.rm = TRUE) # minimum value
      v(data_cveda$p_sdq_tot, na.rm = TRUE) # maximum value
     range(data cveda$p sdq tot, na.rm = TRUE)
```

Good Script

Any Questions?

