

A Benchmarking Workflow for High-Throughput DIA Label-Free Quantification using a Novel High-Resolution Accurate Mass Platform

Anna Pashkova¹, Julia Kraegenbring¹, Pedro Navarro¹, Tabiwang N. Arrey¹, Eduard Denisov¹, Hamish Stewart¹, Jeff Op de Beeck², Eugen Damoc¹; ¹Thermo Fisher Scientific GmbH, Hanna-Kunath-Str. 11, 28199 Bremen, Germany; ²Thermo Fisher Scientific MSS, Gent, Belgium

ABSTRACT

Purpose

Evaluate the performance of the novel Thermo Scientific™ Orbitrap™ Astral™ mass spectrometer for Label-Free Quantitation (LFQ) applications under a variety of conditions, such as sample loads, gradient lengths, and MS acquisition parameters.

Methods

LFQ with a micropillar array-based column for highly reproducible separation of peptide digests coupled to an Orbitrap Astral mass spectrometer operated in data-independent acquisition (DIA) mode with narrow isolation windows (2Th), providing data acquisition at scan rates close to 200 Hz.

Results

Very high quantitative accuracy and precision were demonstrated for 3-proteome digest mixture loads from 50 to 500ng. The dynamic range for quantitation spanned 5 orders of magnitude, which translated into reliable quantitation of proteins within the entire range of expression (from 100s to 100,000s of copies per cell).

INTRODUCTION

Bottom-up proteomics has proven to be the most suitable technology for the high-throughput analysis of very complex biological samples, such as cell lysates or blood. As the obtained data become more and more employed in biomedical research, the challenge of analyzing the smallest sample input amounts in the shortest time remains. Data analysis software must keep up with the fast data acquisition and increasing size of raw data files. To meet all these challenges, a novel mass spectrometer has been developed by combining three mass analyzers: a quadrupole mass analyzer for precursor ion selection, an Orbitrap mass analyzer to acquire high dynamic range HRAM spectra, and the novel Astral mass analyzer to acquire high sensitivity, high dynamic range HRAM spectra at a rate of up to 200 Hz. An integrated workflow for label-free quantitative proteomic studies, based on the Orbitrap Astral mass spectrometer, is presented. Two samples containing E.coli, HeLa (human) and S.cerevisiae (yeast) digests mixed in known ratios, were analyzed in DIA mode, and the results are presented in this poster.

MATERIALS AND METHODS

Sample:

Mix A: E.coli 36%, Human 46%, Yeast 18%;
Mix B: E.coli 18%, Human 46%, Yeast 36% (3 replicates/vial for both).

For the data presented in Fig.1-5 and 8-10:

- LC: Thermo Scientific™ Vanquish™ Neo UHPLC, Direct Injection mode, 500ng sample load, Thermo Scientific™ EASY-Spray™ NG Source, Thermo Scientific™ μPAC Neo 50 cm, 20 min gradient;
- Orbitrap Astral MS Acquisition method: OTMS1 240K, max. inj. time 3ms, 200Hz Astral DIA, 2Th isolation window, max. inj. time 3ms, AGC 500%, NCE25.

For the data presented in Fig.6-7:

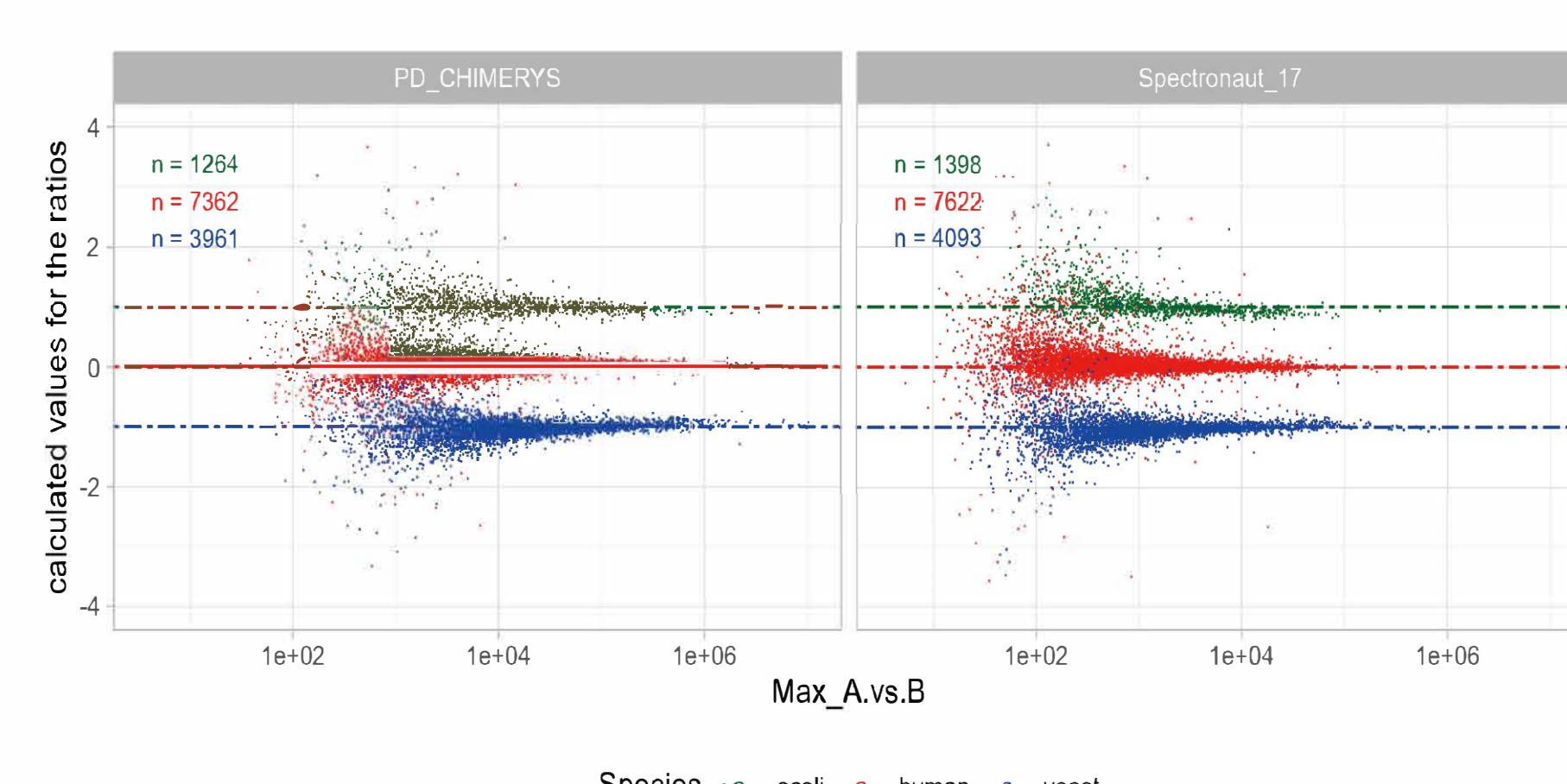
- LC: Vanquish Neo UHPLC, Trap and Elute mode, 50 to 500ng sample load, Thermo Scientific™ EASY-Spray™ NG Source, Thermo Scientific™ EASY-Spray™ PepMap™ 15cm x 150 μ m; SPD/total run time: 180/8 min, 100/14.4 min, 60/24 min;
- Orbitrap Astral MS Acquisition method: OTMS1 240K, max. inj. time 3ms, 200Hz Astral DIA, 2Th isolation window, max. inj. time 3ms, AGC 500%, NCE25.

Data Processing:

- Biognosys Spectronaut™ 17.4, directDIA, normalization by Human FASTA, Q<0.01
- Thermo Scientific™ Proteome Discoverer™ 3.1 software with Chimerys™ search algorithm (MSAID)

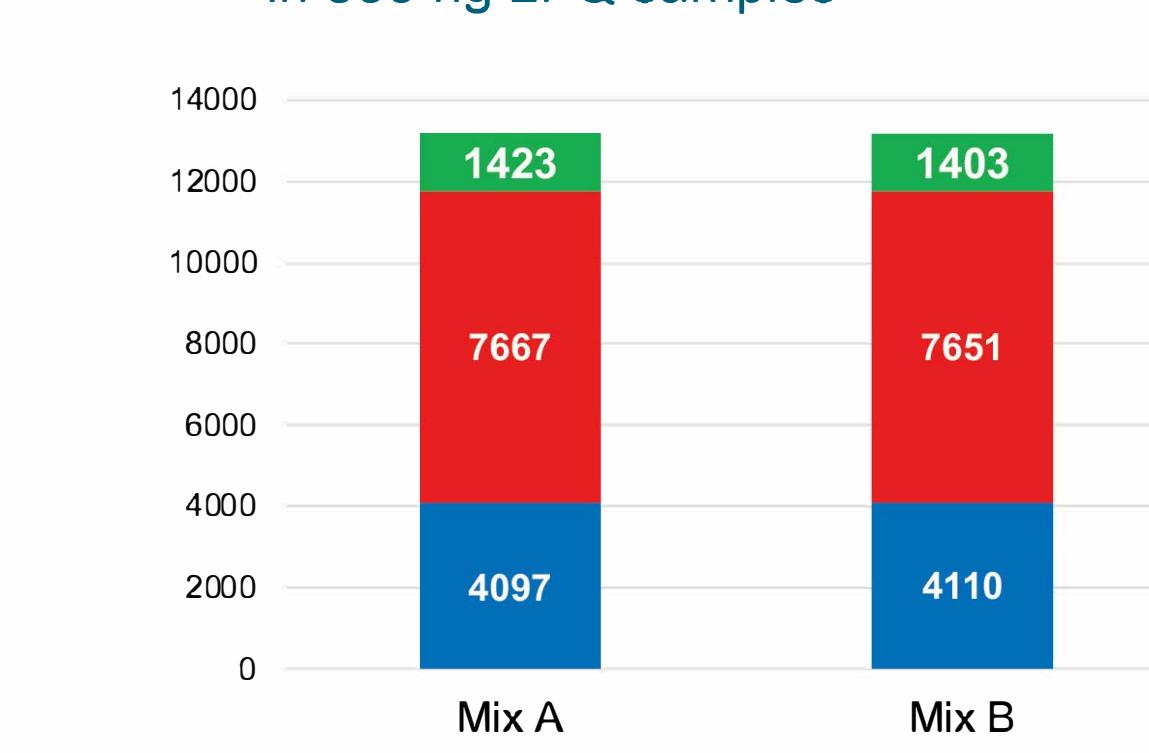
RESULTS

Figure 1 Orbitrap Astral detected log2 ratios (y-axis) distributed by the abundance (x-axis) for the 3-proteome mix samples A and B, processed with Chimerys and Spectronaut 17



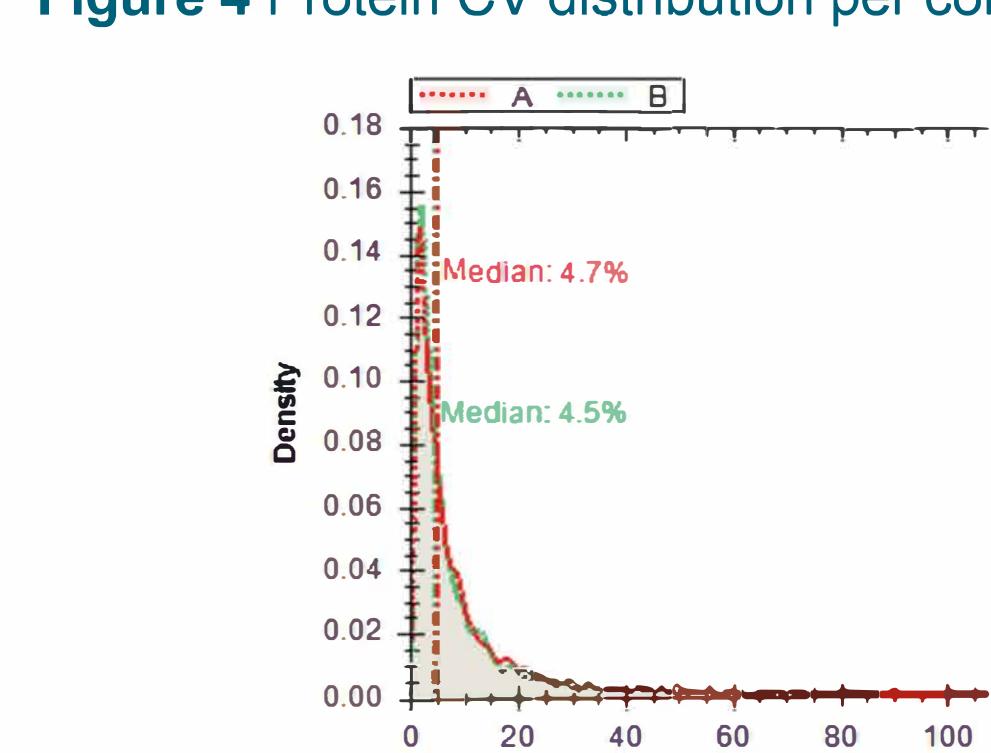
Distribution of average log2 ratios of intensities (highest detected abundance among both conditions) of the identified human, yeast, and E.coli proteins between the two 3-proteome mixtures, A and B, are presented in Fig.1. Data processed with the two search engines, Spectronaut and Chimerys, demonstrate very high correlation. The numbers of identified proteins are shown in the matching colors. Theoretical A/B ratios were 1 for human, 0.5 for yeast, and 2 for E.coli proteomes. Experimentally obtained (SN17) average ratios were 1.016 for human, 0.490 for yeast, and 1.993 for E.coli.

Figure 2 Number of Proteins (1%FDR) quantified in 500 ng LFQ samples



Over 13,000 proteins were identified and quantified across the 3 proteomes from 500ng of the mixed samples A and B, triplicate runs (Fig.2). In addition to the deep proteome coverage, 87% of identified proteins had a CV less than 20% and 72% of identified proteins had a CV less than 10% (Fig.3), with a median CV 4.7% (Fig.4).

Figure 4 Protein CV distribution per condition



According to the Spectronaut data, such a high quantitation accuracy was obtained with a median value of 4 datapoints per peak (Spectronaut defines peak width as 1.7* FWHM) (Fig.5). Increasing DIA window from 2Th to 3 and 4Th raised the number of SN datapoints to 5 and 6 per peak, yet this did not bring an improvement to the average quantitation accuracy (data not shown).

Figure 3 Protein Group CV below X

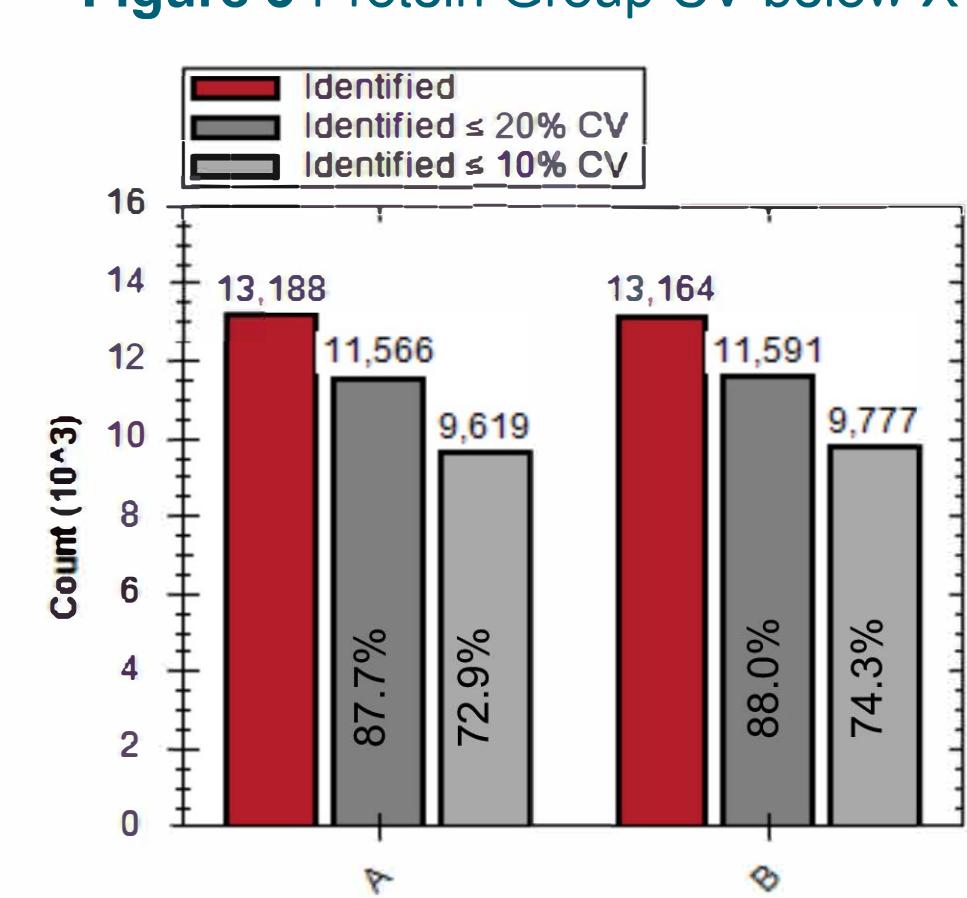


Figure 5 Datapoints per peak distribution

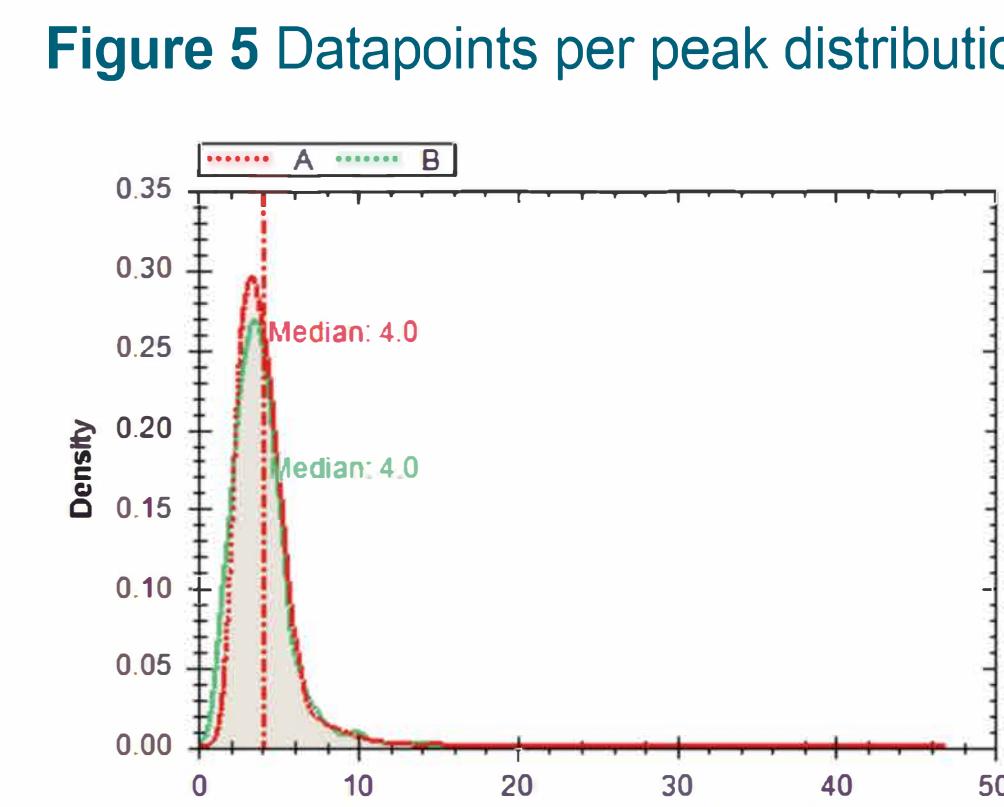
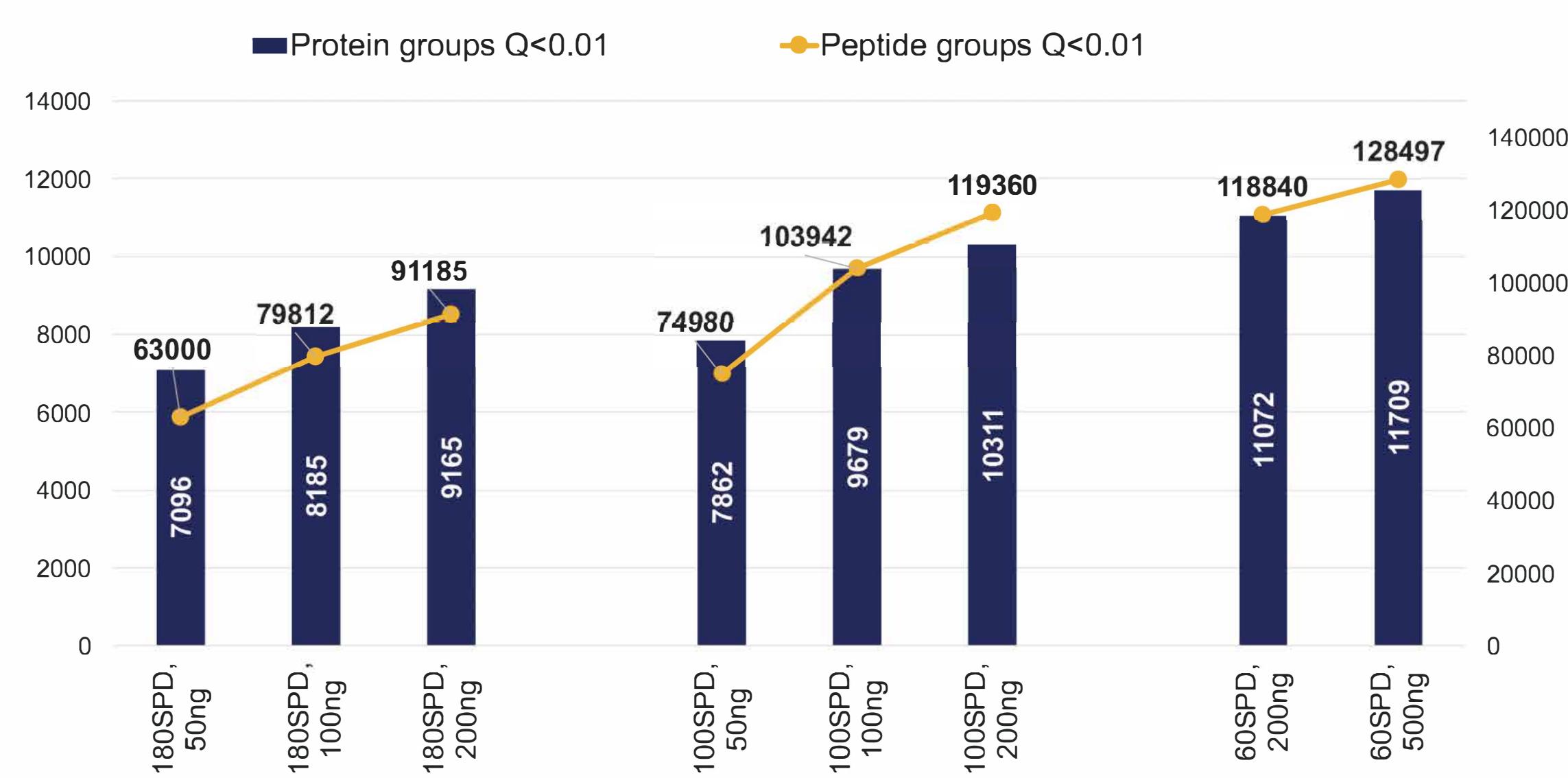


Figure 6 Number of proteins and peptides identified and quantified with Orbitrap Astral MS for different sample loads and gradient lengths



In Fig. 6, the unsurpassed quantitation performance of the new Orbitrap Astral MS is demonstrated for 3-proteome mixture with sample loads from 50 to 500 ng, for 3 different high throughput methods (180, 100 and 60 SPD). Quantitation accuracy was equally high throughout all studied conditions (Fig. 7).

Figure 7 Quantitation accuracy of Orbitrap Astral MS LFQ data acquired for different sample loads and different high throughput methods

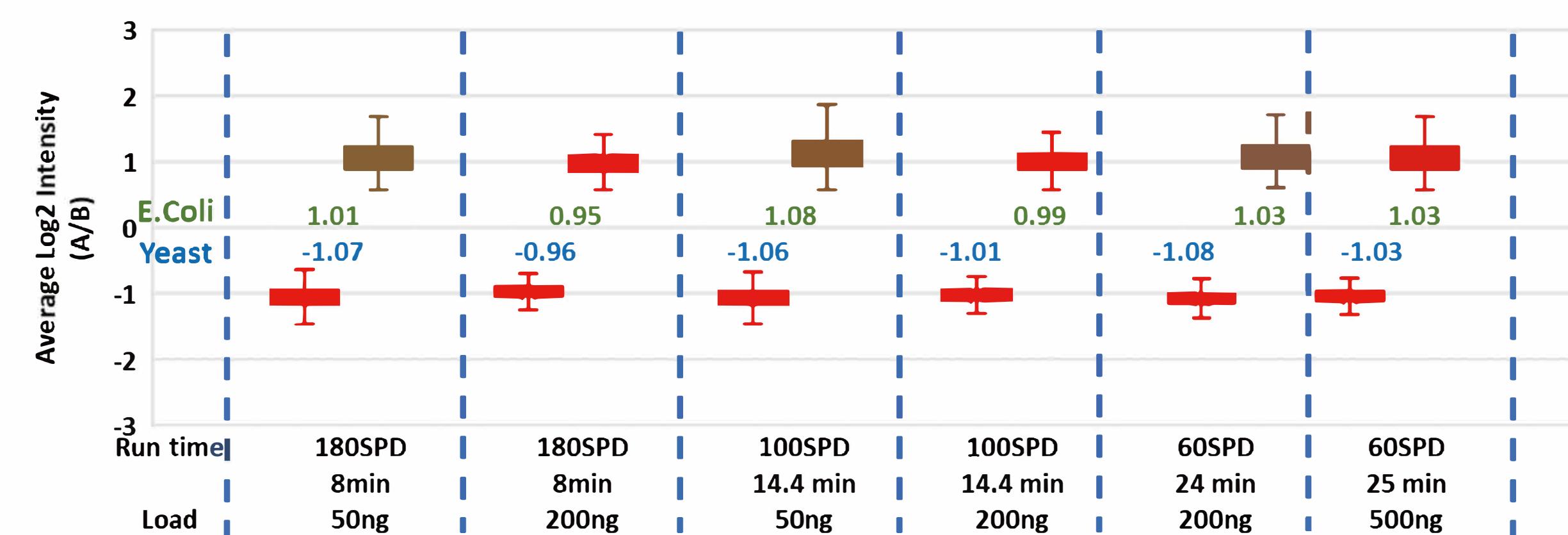
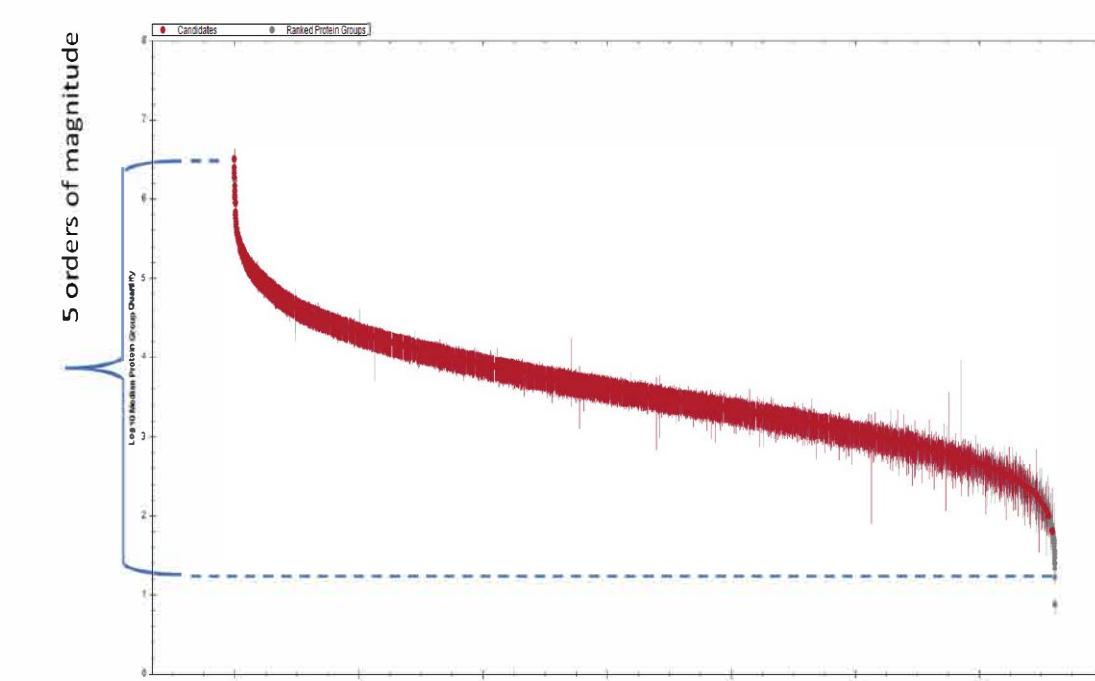
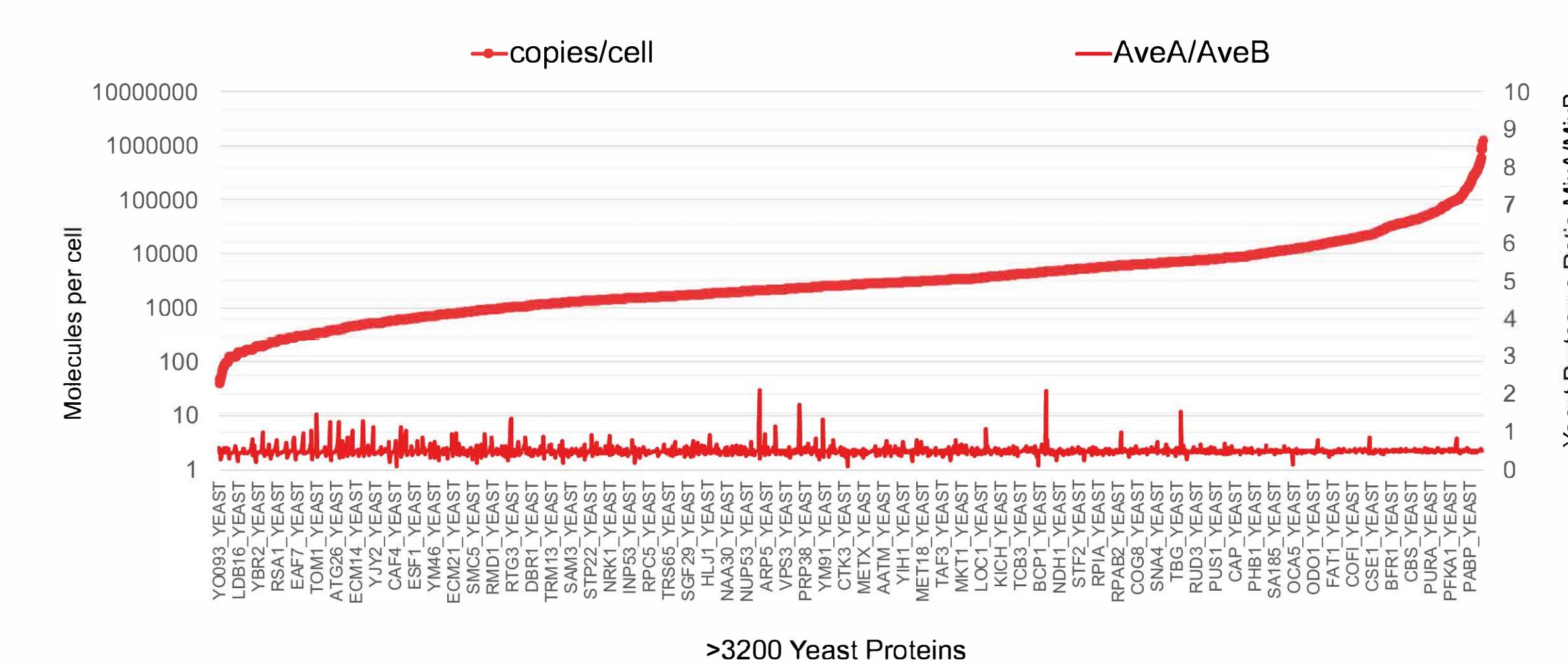


Figure 8 Ranked Protein Groups



Dynamic range for signal intensities for the identified proteins spans over 5 orders of magnitude. The ranked protein groups curve shown in this figure was obtained with 500ng load and 20 min gradient. For the 50ng load, the dynamic range is still close to 5 orders. With such a depth, proteins with the expression level as low as 100 copies per cell can be reliably quantified (Fig.9-10).

Figure 9 LFQ accuracy for Yeast proteome over the protein expression scale



For the Yeast proteome, the abundance of proteins ranges from fewer than 50 to more than 10(6) molecules per cell [1]. With Orbitrap Astral MS, nearly the entire Yeast proteome could be reliably quantified using a 20 min LC gradient and 500ng load of the mixed 3-proteome samples containing 18%/36% yeast. High quantification accuracy was observed throughout the entire expression range.

Figure 10 An example of label-free quantitation for a low-abundant protein MSN2_YEAST (Zinc finger protein MSN2)

Zinc finger protein MSN2 is a positive transcriptional factor that acts as a component of the stress responsive system. Recognizes and binds to the stress response element (STRE) which is involved in the response to various forms of stress (heat, oxidative, osmotic, etc.). Present with 125 molecules/cell in log phase SD medium. This protein was quantified with high accuracy and precision in all 6 runs, 20 min LC gradient, with 500ng total sample, containing 18%/36% yeast.

CONCLUSIONS

The new Orbitrap Astral mass spectrometer ensures ultra-fast 200 Hz data acquisition rates while delivering high sensitivity and dynamic range, spanning at least 5 orders of magnitude. With such performance, proteins with the expression level from 100s to 100,000s of copies per cell can be reliably quantified, opening new frontiers for life sciences research. For LFQ applications, accurate and precise quantitation could be achieved with 6 datapoints per peak (mean value). Run-to-run precision of quantitation was high with less than 1% error, and quantitation based on single-peptide IDs was found to be as reliable as for 2 and more peptides.

REFERENCES

- Ghaemmaghami, S., Huh, W.K., Bower, K. et al. Global analysis of protein expression in yeast. *Nature* **425**, 737–741 (2003). <https://doi.org/10.1038/nature02046>

TRADEMARKS/LICENSING

© 2023 Thermo Fisher Scientific Inc. All rights reserved. Chimerys is a trademark of MSAID. Spectronaut is a trademark of Biognosys AG. All other trademarks are the property of Thermo Fisher Scientific and its subsidiaries unless otherwise specified.