

Q1-R

August 15, 2024

1 Q1 - R section

In this section, I use R to perform differential expression analysis for feature selection. In my opinion, differential expression (DE) analysis is the most efficient feature selection method in this case because it not only considers the variance or mutual information of features but also how they are expressed across different classes. Therefore, I believe it will help us identify the best features (DEGs) for classification.

```
[23]: # Import needed libraries
library(limma)
library(edgeR)
```

```
[24]: # Load Data
normal_counts <- read.csv("train_normal_counts.csv")
meta_data <- read.csv("train_meta_data.csv")
```

```
[25]: head(normal_counts)
```

		DLDR_0036	DLDR_0081	DLDR_0191	DLDR_0188	DLDR_0130	DLDR_0
		<dbl>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>
A data.frame: 6 × 134	1	5.820135	6.5462994	6.6040504	6.480745	6.550016	6.569252
	2	-1.060061	0.5821648	-0.8650363	-1.083676	-1.222374	0.767254
	3	4.388400	3.7520898	4.3514891	4.361634	4.534941	4.150470
	4	4.080172	4.6451746	4.0721368	4.313540	4.370763	4.166038
	5	2.564430	3.8408991	3.1431376	3.120196	3.512952	3.757010
	6	3.552685	3.2010747	4.0374758	1.941859	2.517867	3.253653

```
[26]: dim(normal_counts)
```

1. 17396 2. 134

```
[27]: head(meta_data)
```

	Simplified_class <chr>
1	Normal
2	Advanced_fibrosis
3	Normal
4	Normal
5	Non_advanced_Fibrosis
6	Normal

A data.frame: 6 × 1

```
[28]: dim(meta_data)
```

```
1. 134 2. 1
```

```
[29]: labels <- factor(meta_data$Simplified_class)
```

```
[30]: print(labels)
```

```

[1] Normal          Advanced_fibrosis   Normal
[4] Normal          Non_advanced_Fibrosis Normal
[7] Advanced_fibrosis Non_advanced_Fibrosis Non_advanced_Fibrosis
[10] Normal          Advanced_fibrosis   Advanced_fibrosis
[13] Advanced_fibrosis Non_advanced_Fibrosis Advanced_fibrosis
[16] Advanced_fibrosis Normal          Normal
[19] Normal          Normal          Advanced_fibrosis
[22] Advanced_fibrosis Non_advanced_Fibrosis Non_advanced_Fibrosis
[25] Advanced_fibrosis Non_advanced_Fibrosis Normal
[28] Non_advanced_Fibrosis Normal          Advanced_fibrosis
[31] Advanced_fibrosis Advanced_fibrosis   Advanced_fibrosis
[34] Normal          Normal          Non_advanced_Fibrosis
[37] Non_advanced_Fibrosis Advanced_fibrosis   Advanced_fibrosis
[40] Non_advanced_Fibrosis Advanced_fibrosis   Advanced_fibrosis
[43] Advanced_fibrosis Advanced_fibrosis   Non_advanced_Fibrosis
[46] Non_advanced_Fibrosis Normal          Advanced_fibrosis
[49] Advanced_fibrosis Advanced_fibrosis   Non_advanced_Fibrosis
[52] Advanced_fibrosis Normal          Non_advanced_Fibrosis
[55] Normal          Advanced_fibrosis   Advanced_fibrosis
[58] Non_advanced_Fibrosis Normal          Normal
[61] Normal          Non_advanced_Fibrosis Advanced_fibrosis
[64] Advanced_fibrosis Normal          Normal
[67] Advanced_fibrosis Normal          Advanced_fibrosis
[70] Advanced_fibrosis Advanced_fibrosis   Advanced_fibrosis
[73] Normal          Advanced_fibrosis   Non_advanced_Fibrosis
[76] Normal          Advanced_fibrosis   Advanced_fibrosis
[79] Non_advanced_Fibrosis Non_advanced_Fibrosis Advanced_fibrosis
[82] Normal          Advanced_fibrosis   Non_advanced_Fibrosis
[85] Non_advanced_Fibrosis Advanced_fibrosis   Normal
[88] Non_advanced_Fibrosis Advanced_fibrosis   Non_advanced_Fibrosis
[91] Advanced_fibrosis Normal          Normal
[94] Advanced_fibrosis Non_advanced_Fibrosis Non_advanced_Fibrosis

```

```

[97] Non_advanced_Fibrosis Advanced_fibrosis    Advanced_fibrosis
[100] Advanced_fibrosis      Normal          Normal
[103] Non_advanced_Fibrosis Non_advanced_Fibrosis Normal
[106] Normal                Advanced_fibrosis Non_advanced_Fibrosis
[109] Non_advanced_Fibrosis Normal          Advanced_fibrosis
[112] Non_advanced_Fibrosis Normal          Normal
[115] Advanced_fibrosis      Normal          Normal
[118] Non_advanced_Fibrosis Advanced_fibrosis Non_advanced_Fibrosis
[121] Normal                Advanced_fibrosis Non_advanced_Fibrosis
[124] Non_advanced_Fibrosis Non_advanced_Fibrosis Normal
[127] Advanced_fibrosis      Non_advanced_Fibrosis Advanced_fibrosis
[130] Normal                Normal          Normal
[133] Normal                Normal
Levels: Advanced_fibrosis Non_advanced_Fibrosis Normal

```

Let's perform DE analysis

```

[31]: # Create a design matrix
design <- model.matrix(~0 + labels)
colnames(design) <- levels(labels)

[32]: fit <- lmFit(normal_counts, design)

[33]: contrast.matrix <- makeContrasts(
  AdvancedFibrosis_vs_Normal = `Advanced_fibrosis` - Normal,
  Fibrosis_vs_Normal = Non_advanced_Fibrosis - Normal,
  AdvancedFibrosis_vs_Fibrosis = `Advanced_fibrosis` - Non_advanced_Fibrosis,
  levels = design
)

# Apply contrasts to the fit
fit2 <- contrasts.fit(fit, contrast.matrix)

# Empirical Bayes moderation to get p-values
fit2 <- eBayes(fit2)

```

Now, we are going to extract the DEGs for each pair of classes and save them

```

[34]: # Get the top DEGs for the Advanced Fibrosis vs Normal comparison
top_genes_adv_vs_norm <- topTable(fit2, coef = "AdvancedFibrosis_vs_Normal",
  ↪adjust.method = "BH", number = Inf)

# Get the top DEGs for the Fibrosis vs Normal comparison
top_genes_fib_vs_norm <- topTable(fit2, coef = "Fibrosis_vs_Normal", adjust.
  ↪method = "BH", number = Inf)

# Get the top DEGs for the Advanced Fibrosis vs Fibrosis comparison

```

```
top_genes_adv_vs_fib <- topTable(fit2, coef = "AdvancedFibrosis_vs_Fibrosis",
  ↪adjust.method = "BH", number = Inf)

# View the top DEGs
head(top_genes_adv_vs_norm)
head(top_genes_fib_vs_norm)
head(top_genes_adv_vs_fib)
```

		logFC <dbl>	AveExpr <dbl>	t <dbl>	P.Value <dbl>	adj.P.Val <dbl>	B <dbl>
A data.frame: 6 × 6	10728	-1.3278914	1.902497	-10.235955	1.733536e-18	3.015659e-14	31.41974
	13385	1.0088994	6.051398	9.886439	1.299982e-17	9.020981e-14	29.46392
	10694	-1.3132140	3.432308	-9.855214	1.555699e-17	9.020981e-14	29.28958
	16113	-3.4233202	-1.392599	-9.734863	3.105946e-17	1.350776e-13	28.61834
	16278	-2.8530739	-0.172656	-9.680406	4.245065e-17	1.476943e-13	28.31501
	6969	0.4379282	4.207436	9.600224	6.721385e-17	1.948753e-13	27.86885

		logFC <dbl>	AveExpr <dbl>	t <dbl>	P.Value <dbl>	adj.P.Val <dbl>	B <dbl>
A data.frame: 6 × 6	13623	0.7946468	5.255822	11.73980	2.831866e-22	4.926315e-18	40.01880
	10970	0.6321228	5.361546	11.47202	1.339959e-21	1.021819e-17	38.49831
	5442	-1.4931333	2.180779	-11.38733	2.190883e-21	1.021819e-17	38.01728
	17075	0.6202635	7.265926	11.37529	2.349550e-21	1.021819e-17	37.94887
	4461	0.6782287	6.569435	11.29235	3.802820e-21	1.323077e-17	37.47774
	6563	0.6004911	5.545244	11.13147	9.676618e-21	2.805574e-17	36.56387

		logFC <dbl>	AveExpr <dbl>	t <dbl>	P.Value <dbl>	adj.P.Val <dbl>	B <dbl>
A data.frame: 6 × 6	16863	1.223013	2.9208664	9.097265	1.182040e-15	2.056276e-11	24.90970
	3296	1.594761	1.9955120	8.122841	2.755851e-13	1.021684e-09	19.70546
	673	1.485822	1.1568774	8.121636	2.774199e-13	1.021684e-09	19.69912
	14913	-1.153029	2.6115748	-8.115173	2.874679e-13	1.021684e-09	19.66515
	12060	1.675825	0.1951384	8.111304	2.936548e-13	1.021684e-09	19.64482
	3227	1.279852	4.3441375	8.016339	4.947792e-13	1.434530e-09	19.14676

```
[35]: write.csv(top_genes_adv_vs_norm, "DEGs_AdvancedFibrosis_vs_Normal.csv")
write.csv(top_genes_fib_vs_norm, "DEGs_Fibrosis_vs_Normal.csv")
write.csv(top_genes_adv_vs_fib, "DEGs_AdvancedFibrosis_vs_Fibrosis.csv")
```

We have filtered the top 200 DEGs for each pair. The choice of =200 appears to be optimized based on our greedy search, which has not been included in this notebook.

```
[36]: filtered_genes_adv_vs_norm <- top_genes_adv_vs_norm[1:200,]
filtered_genes_fib_vs_norm <- top_genes_fib_vs_norm[1:200,]
filtered_genes_adv_vs_fib <- top_genes_adv_vs_fib[1:200,]

# View filtered DEGs
head(filtered_genes_adv_vs_norm)
head(filtered_genes_fib_vs_norm)
```

```
head(filtered_genes_adv_vs_fib)
```

		logFC <dbl>	AveExpr <dbl>	t <dbl>	P.Value <dbl>	adj.P.Val <dbl>	B <dbl>
A data.frame: 6 × 6	10728	-1.3278914	1.902497	-10.235955	1.733536e-18	3.015659e-14	31.41974
	13385	1.0088994	6.051398	9.886439	1.299982e-17	9.020981e-14	29.46392
	10694	-1.3132140	3.432308	-9.855214	1.555699e-17	9.020981e-14	29.28958
	16113	-3.4233202	-1.392599	-9.734863	3.105946e-17	1.350776e-13	28.61834
	16278	-2.8530739	-0.172656	-9.680406	4.245065e-17	1.476943e-13	28.31501
	6969	0.4379282	4.207436	9.600224	6.721385e-17	1.948753e-13	27.86885
		logFC <dbl>	AveExpr <dbl>	t <dbl>	P.Value <dbl>	adj.P.Val <dbl>	B <dbl>
A data.frame: 6 × 6	13623	0.7946468	5.255822	11.73980	2.831866e-22	4.926315e-18	40.01880
	10970	0.6321228	5.361546	11.47202	1.339959e-21	1.021819e-17	38.49831
	5442	-1.4931333	2.180779	-11.38733	2.190883e-21	1.021819e-17	38.01728
	17075	0.6202635	7.265926	11.37529	2.349550e-21	1.021819e-17	37.94887
	4461	0.6782287	6.569435	11.29235	3.802820e-21	1.323077e-17	37.47774
	6563	0.6004911	5.545244	11.13147	9.676618e-21	2.805574e-17	36.56387
		logFC <dbl>	AveExpr <dbl>	t <dbl>	P.Value <dbl>	adj.P.Val <dbl>	B <dbl>
A data.frame: 6 × 6	16863	1.223013	2.9208664	9.097265	1.182040e-15	2.056276e-11	24.90970
	3296	1.594761	1.9955120	8.122841	2.755851e-13	1.021684e-09	19.70546
	673	1.485822	1.1568774	8.121636	2.774199e-13	1.021684e-09	19.69912
	14913	-1.153029	2.6115748	-8.115173	2.874679e-13	1.021684e-09	19.66515
	12060	1.675825	0.1951384	8.111304	2.936548e-13	1.021684e-09	19.64482
	3227	1.279852	4.3441375	8.016339	4.947792e-13	1.434530e-09	19.14676

```
[37]: dim(filtered_genes_adv_vs_norm)
dim(filtered_genes_fib_vs_norm)
dim(filtered_genes_adv_vs_fib)
```

```
1. 200 2. 6
```

```
1. 200 2. 6
```

```
1. 200 2. 6
```

These top 200 DEGs are biologically meaningful in addition to their role in computationally classifying the data. They are likely genes whose expression changes significantly when transitioning from one class to another. These genes are probably among the most correlated with the class labels, though they are not necessarily causal genes. The change in class labels may have a substantial impact on their expression, potentially affecting their associated pathways or other related biological processes.

```
[38]: genes_adv_vs_norm_names <- rownames(filtered_genes_adv_vs_norm)
genes_fib_vs_norm_names <- rownames(filtered_genes_fib_vs_norm)
genes_adv_vs_fib_names <- rownames(filtered_genes_adv_vs_fib)
```

then we combined the filtered DEGs to create a new feature space

```
[39]: combined_gene_names <- unique(c(genes_adv_vs_norm_names,
                                     genes_fib_vs_norm_names,
                                     genes_adv_vs_fib_names))
```

```
[40]: length(combined_gene_names)
```

527

```
[41]: common_genes <- intersect(rownames(normal_counts), combined_gene_names)
selected_normal_counts <- normal_counts[common_genes, ]
head(selected_normal_counts)
```

		DLDR_0036	DLDR_0081	DLDR_0191	DLDR_0188	DLDR_0130	DLDR_0120
		<dbl>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>
A data.frame: 6 × 134	10	4.5895546	5.4821690	5.01315395	5.0660709	4.6558168	4.2990168
	57	-0.7190239	0.5821648	-1.15454295	-1.0836762	-0.2894877	-0.0064168
	265	0.5876374	-2.2251901	-0.07654044	0.1964317	0.2723912	2.5231168
	275	1.6624052	2.5296974	1.50842206	1.9916119	1.4679420	1.9341168
	278	5.5299023	5.8462723	5.55358296	5.3592673	5.7745493	4.9626168
	297	7.5494873	7.7592284	7.61253797	8.0700567	7.4394046	7.6716168

```
[42]: dim(selected_normal_counts)
```

1. 527 2. 134

We extracted a subset from the data based on selected features. Let's save it and continue the analysis in Python Jupyter Notebook

```
[43]: write.csv(selected_normal_counts, "subset_data.csv")
```