## **Additional File 1**

**Table S1.** List of genes, sequences of primers, cycling conditions for PCR assays, and amplicon sizes.

Assay	Gene	Primer sequence	Primer sequence PCR program <sup>a</sup>		Reference
E. coli	uspA	F: 5'-CCGATACGCTGCCAATCAGT-3'	94°C/ 120 s, 70°C/ 60 s,	884	1
confirmation	R: 5'-ACGCAGACCGTAGGCCAGAT-3'	72°C/ 60 s (30 cycles)			
Phylogenetic	chuA _	F: 5'-GACGAACCAACGGTCAGGAT-3' R: 5'-TGCCGCCAGTACCAAAGACA-3'	94°C/ 30 s,	279	
typing (Triplex	yjaA .	F: 5'-TGAAGTGTCAGGAGACGCTG-3' 55°C/ 30 s, R: 5'-ATGGAGAATGCGTTCCTCAAC-3' 72°C/ 30 s	211	2	
method)	TspE4.C2	F: 5'-GAGTAATGTCGGGGCATTCA-3' R: 5'-CGCGCCAACAAAGTATTACG-3'	(30 cycles)	152	
Phylogenetic typing	chuA _	F: 5'-ATGGTACCGGACGAACCAAC-3' R: 5'-TGCCGCCAGTACCAAAGACA-3'	94°C/ 5 s,	288	
(Quadruplex method)	ујаА	F: 5'-CAAACGTGAAGTGTCAGGAG-3' R: 5'-AATGCGTTCCTCAACCTGTG-3'	57°C/ 20 s (30 cycles)	211	3
metriouj	TspE4.C2	F: 5'-CACTATTCGTAAGGTCATCC-3'		152	

		R: 5'-AGTTTATCGCTGCGGGTCGC-3'			
	arpA	arpA F: 5'-AACGCTATTCGCCAGCTTGC-3'		400	
	(Group F)	R: 5'-TCTCCCCATACCGTACGCTA-3'		400	
	arpA	F: 5'-GATTCCATCTTGTCAAAATATGCC-3'		301	
	(Group E)	R: 5'-GAAAAGAAAAGAATTCCCAAGAG-3'		301	
	trpA	F: 5'-AGTTTTATGCCCAGTGCGAG-3'	94°C/ 5 s,		
	(Group C)	P. F/ TOTOGOGOGOGOGOGO	59°C/ 20 s	219	
	(Group C)	R: 5'-TCTGCGCCGGTCACGCCC-3'	(30 cycles)		
	trpA	F: 5'-CGGCGATAAAGACATCTTCAC-3'	94°C/ 5 s,		
	(Internal	D. F./ CCA ACCCCCCCCCCC A A C. 2/	57°C/ 20 s	489	
	control)	R: 5'-GCAACGCGGCCTGGCGGAAG-3'	(30 cycles)		
		F: 5'-GACCCGGCACAAGCATAAGC-3'	95°C/ 30 s,		
	eae		54°C/ 90 s,	384	1
		R: 5'-CCACCTGCAGCAACAAGAGG-3'	72°C/ 90 s	301	
DEC			(30 cycles)		4, 5
DLC	F: 5'-ATAAATCGCCATTCGTTGACTAC-3'	95°C/ 30 s,	180	7, 5	
	35.71	R: 5'-AGAACGCCCACTGAGATCATC-3'	52°C/ 60 s,	150	
	stx2	F: 5'-GGCACTGTCTGAAACTGCTCC-3'	72°C/ 60 s	255	
	R: 5'-TCGCCAGTTATCTGACATTCTG-3'		(35 cycles)	233	

	fyuA	F: 5'-GTAAACAATCTTCCCGCTCGGCAT-3'	72°C/ 90 s	850	
ExPEC <sup>c</sup>		R: 5'-GGCCAGAACATTTGCTCCCTTGTT-3'	63°C/ 90 s,		6
	vat	F: 5'-TCAGGACACGTTCAGGCATTCAGT-3'	94°C/ 30 s,	1100	
			(35 cycles)		
	aggA	R: 5'-TCAACCTTGACACTTGCC-3'	72°C/ 120 s	414	
	aaa4		50°C/ 60 s,	414	
		F: 5'-ATGCATTACTTTGGGTTTAG-3'	94°C/ 60 s,		
		R: 5'-ATAGCAGTCGATTTAGCAGCC-3'	(30 cycles)		
	bfpA .		72°C/ 40 s	461	
		F: 5'-ATTGAATCTGCAATGGTGC-3'	55°C/ 40 s,		
			95°C/ 40 s,		
		R: 5'-GCCGGTCAGCCACCCTCTGAGAGTAC-3'	(35 cycles)		
	іраН		72°C/ 60 s	620	
		F: 5'-GTTCCTTGACCGCCTTTCCGATACCGTC-3'	60°C/ 60 s,		
			94°C/ 40 s,		
	st1A _	R: 5'-ATAACATCCAGCACAGGC-3'	(35 cycles)	186	
		F: 5'-TCTGTATTATCTTTCCCCTC-3'	72°C/ 120 s		
	ItA	R: 5'-CCGAATTCTGTTATATATGTC-3'	50°C/ 60 s,	696	
		F: 5'-GGCGACAGATTATACCGTGC-3'	94°C/ 60 s,	606	

		R: 5'-TGACGATTAACGAACCGGAAGGGA-3'	(30 cycles)		
	chuA	F: 5'-CTGAAACCATGACCGTTACG-3'		652	
		R: 5'-TTGTAGTAACGCACTAAACC-3'		032	
	ufc\/	F: 5'-ACATGGAGACCACGTTCACC-3'	292		
	yfcV _	R: 5'-GTAATCTGGAATGTGGTCAGG-3'		292	
		F: 5'-TAGCAAACGTTCTATTGGTGC-3'	94°C/ 30 s,		
	kpsM-K1		63°C/ 30 s,	<del>153</del>	7
		R: 5'-CATCCAGACGATAAGCATGAGCA-3'	68°C/ 3 min		
			(25 cycles)		
		F: 5'-GCGCATTTGCTGATACTGTTG-3'	94°C/ 30 s,		
	kpsM-II		<del>58°C/ 30 s,</del>	577	8
	<del>RPSIVI II</del>	R: 5'-AGGTAGTTCAGACTCACACCT-3'	72°C/ 3 min	377	
			(25 cycles)		

- <sup>a</sup>Before starting the PCR cycle, DNA was first denatured at 95°C/15 min. After completion
- 3 of the cycle, there was a final primer extension at 72°C/8 min.
- 4 bDiarrheagenic E. coli are defined by the following genes: eae+bfpA/eae (EPEC), ltA+st1A/ltA/st1A (ETEC), ipaH (EIEC),
- 5 eae+stx1+stx2/stx1/stx2 (EHEC), and aggA (EAEC).
- 6 cExtra-intestinal pathogenic *E. coli*

7 8	References in Table 1
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Table S2. Sample collection dates and locations, and phylogenetic groupings of E. coli by

## 52 triplex and quadruplex PCR assays.

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Date of sample collection	Location (no. of confirmed <i>E. coli</i> )	Phylogenetic grouping by triplex PCR (no. of isolates)  B1 (2)	Phylogenetic grouping by quadruplex PCR (no. of isolates) B1 (2)
11 June 2018	Z <sup>b</sup> (2)  J (7)  Z (2)  H <sup>c</sup> (5)	D (1)  A (6)  B1 (3)  D (5)	D (1) A (6) B1 (3) D (4) E (1)
9 July 2018	J (1) Z (3) H (2)	A (3) B1 (1) B2 (1) D (1)	A (3) B1 (1) B2 (1) D (1)
13 August 2018	J (6) Z (6) H (7)	A (9) B1 (5) D (2)	A (9) B1 (5) D (1) E (1)
10 September 2018	J (2) Z (1) H (9)	A (8) B1 (3) B2 (1)	A (7) B1 (3) B2 (1) C (1)
8 October 2018	J (4) Z (1) H (4)	A (4) B1 (3) D (2)	A (4) B1 (3) D (1) F (1)

			1
12 November	J (4)	A (7)	A (7)
2018	Z (1) H (4)	B1 (2)	B1 (2)
	11 (4)		1 (2)
	J (3)	A (3)	A (3)
10 December	Z (4)	B1 (3)	B1 (3)
2018	H (3)	D (4)	D (2)
	11 (3)	<i>D</i> (4)	F (2)
			A (3)
	1 (4)	A (3)	B1 (3)
7 January	J (4)	B1 (3)	B2 (2)
2019	Z (5)	B2 (2)	D (3)
	H (5)	D (6)	F (1)
			E (2)
4 Fobruary	J (4)	A (3)	A (3)
4 February	Z (2)	B1 (3)	B1 (3)
2019	H (2)	D (2)	F (2)
	1 (3)	A (6)	A (6)
11 March	J (3)	B1 (1)	B1 (1)
2019	Z (6)	B2 (4)	B2 (4)
	H (3)	D (1)	D (1)
			A (14)
	J (7)	A (16)	B1 (5)
8 April 2019	Z (7)	B1 (5)	D (1)
	H (10)	D (3)	F (2)
			C (2)
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<sup>53 &</sup>lt;sup>a</sup>Jabriya.

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<sup>54 &</sup>lt;sup>b</sup>Zahraa.

<sup>55 &</sup>lt;sup>c</sup>Hateen.

**Table S3.** Distribution of ExPEC according to location and date of collection of samples.

Month/Year	No. of ExPEC	Location		
Wionthy real	isolates	Jabriya	Zahraa	Hateen
14 May/2018	F	0	F	0
11 June/2018	Ţ	F	F	厚
9 July/2018	<del>[</del> =	0	1	<del>-</del>
13 August/2018	<u></u>	<del>[</del> =	2 =	<b>-</b>
10 September/2018	2	0	0	2=
8 October/2018	F	1	0	3 =
12 November/2018	3	2	1	0
10 December/2018	5	2	2	1
7 January/2019	7	2=	3 =	2
4 February/2019	4	0	2 =	2
11 March/2019	7=	2=	3	2 📁
8 April/2019	7 =	- - -	0	<b>5</b>