

Biochemistry and Modern Applications

Review Article ISSN: 2638-7735

The Role of Human Coactosin-Like Protein in Neurodegenerative Disorders

Y Anu Shanu^{1,4,5}, Antonio Lauto^{2,3,5}, Simon J Myers^{1,3,4,5*}

Affiliation

¹Neuro-Cell Biology Laboratory, Western Sydney University, Locked Bag 1797, Penrith, NSW 1797 Australia

*Corresponding author: Simon J Myers, Neuro-Cell Biology Laboratory, Western Sydney University, Locked Bag 1797, Penrith, NSW 1797 Australia, Tel: +61-2-46203383, Fax: +61-2-46203025, Email: s.myers@uws.edu.au

Citation: Anu SY, Lauto A, Myers SJ. The Role of Human Coactosin-Like Protein in Neurodegenerative Disorders (2017) Biochemistry and

Modern Applications 1: 20-24. **Received:** Nov 27, 2017 **Accepted:** Dec 26, 2017 **Published:** Dec 31, 2017

Copyright: © 2017 Myers SJ, et al., This is an open-access article distributed under the terms of the Creative Commons Attribution License,

which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Coactosin is one of the numerous actin-binding proteins which regulate the actin cytoskeleton. Coactosin binds F-actin, and also interacts with 5-lipoxygenase, which is the first committed enzyme in leukotriene biosynthesis. Coactosin and human coactosin like protein 1 (COTL1) have the potential to play a role in the degradation or impairment of neuronal cells and their functioning. Its homology to other proteins that affect neuronal cells also contributes to this notion. The objective of this review is to explore its structural novelty, regulation and its significance in neurodegenerative diseases.

Keywords: COTL1; Coactosin; 5'Lipoxygenase; Actin polymerisation; Hereditary Sensory Neuropathy; Neurodegeneration; Cytoskeleton

Introduction

Neurodegenerative disorders include some of the most common and least understood diseases characterised by impaired sensory, motor and/or mental functioning. Many, such as Alzheimer's disease, currently have no cure and subsequently have become the focus of significant research in searching for an understanding of the molecular pathophysiology of the disease and looking for methods of prevention, treatment and cure.

Neurodegenerative disorders are categorised by the loss of structure and/or function of neurons. Neuronal cell degeneration and death as well as plaques and atypical protein assemblies are the most common causes of such loss of structure and function (Heemels 2006). Neurons are made up of two morphologically defined regions: the dendrites and the axons. Microtubules, actin filaments and neurofilaments make up the neuronal cytoskeleton and are responsible for several critical roles including the formation of axons, intracellular transport of cargoes and maintenance of its structure (Kevenaar &Hoogenraad 2015).

Coactosin is a 17 kDa actin binding protein originally isolated from *Dictyostelium discoidum* but it has since been isolated from other organisms including humans, fruit flies and mice (Poukkula et al.

2011). Both coactosin and human coactosin like protein 1 (COTL1) can bind to 5-lipoxygenase (5-LO) and filamentous actin (F-actin) (Lackie 2007) and has the potential to play a role in the degradation or impairment of neuronal cells and their functioning. Its homology to other proteins that affect neuronal cells also contributes to this notion.

Structure of COTL1

Coactosin shows structural homology to actin-depolymerising factor (ADF-H) domain. The ADF homology domain, a highly conserved protein motif, promotes cytoskeletal dynamics by facilitating processes such as endocytosis, cell migration, morphogenesis and cytokinesis (Poukkula et al. 2011), owing to its binding affinity towards filamentous actin (F-actin) and not globular actin (G-actin). ADF domain consists of five internal β -sheets, of which β 1- β 4 are antiparallel and β 4 is parallel to β 5. The sheets are surrounded by four α -helices with α 1- α 3 parallel to the β -sheets and α 4 packed perpendicular to the β 3 and β 4 sheets (Hellman et al. 2004).

Despite the similar structure, the amino acid sequence of COTL1 shows low homology to Coactosin with 33.3% sequence identity and

²Biomedical Engineering & Neuroscience (BENS), Western Sydney University, Locked Bag 1797, Penrith South DC, NSW 1797 Australia

³School of Science and Health, Western Sydney University, Locked Bag 1797, Penrith, NSW 1797 Australia

⁴Medical Sciences Research Group, School of Medicine, Western Sydney University, Locked Bag 1797, Penrith, NSW 1797 Australia

⁵School of Medicine, Western Sydney University, Locked Bag 1797, Penrith, NSW 1797 Australia



75% similarity (Liu et al. 2004). COTL1 is entirely composed of a single ADF domain and consists of five β -stranded sheets with two helices each on either side (Fig. 1A). The loop between β 3- β 4 is flexible while the other regions are held rigid (Rakonjac, 2009). Lys-75 and Lys-131, which corresponds to binding sites for F-actin and 5-LO respectively (Fig. 1B) lies close to each other suggesting overlapping binding sites, which has been suggested as a structural mechanism to prevent simultaneous binding (Liu et al 2004), which is also supported by the fact that a 5-LO-COTL1-F-Actin ternary complex has not yet been reported (Esser et al. 2010).

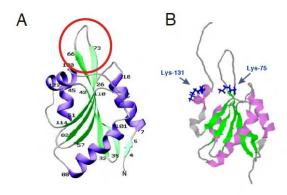


Figure 1: Structure of human COTL1.

1A shows the ribbon structure with β 1- β 5 sheets shown in green and α 1- α 4 helices in purple. The flexible β 3- β 4 loop is highlighted in red. 1B shows spatial alignment of Lys-75 and Lys-131, binding sites for Factin and 5-LO respectively (Rakonjac, 2009).

COTL1 and Actin polymerisation

COTL1 plays several functional roles in normal physiology, however it is primarily identified as an actin binding protein due to its role in the promotion of actin polymerisation. Actin treadmilling is a dynamic process where ATP bound globular monomer actin (G-Actin) is added on to the '+' end of the growing filament by ADF-H protein Profilin, promoting actin polymerisation (Fig. 2) where as depolymerisation occurs when Cofilin, another ADF-H protein, removes ADP bound G-Actin from the '-' end of the filament (Hou et al. 2013). Capping proteins are employed at the '+' end to regulate the rate of polymerisation and COTL1 counteracts the activities of the capping proteins cap32/34 (Rohrig et al. 1995). Therefore by interfering with these proteins COTL1 indirectly promotes actin polymerisation (Hou et al. 2013; Liepinsh et al. 2004; Rohrig et al. 1995). The binding follows a 1:2 stoichiometry between COTL1 and actin and is a Ca²⁺ independent process (Rakonjac, 2009).

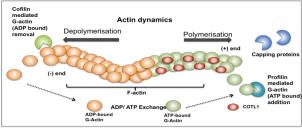


Figure 2: Schematic diagram showing the dynamic nature of F-actin.

ATP-G-actin is added on to the '+' end by Profilin, promoting actin polymerisation and ADP-G-actin is removed from the '-' end by

Cofilin resulting in depolymerisation. COTL1 binds to F-actin at a 1:2 ratio, preventing binding of capping proteins, thereby promoting actin polymerisation.

This dynamic polymerisation-depolymerisation process enables F-actin to perform its functions involving cell migration and morphogenesis. Growth cones are structures that lead axons to synaptic targets by virtue of this process (Hou et al. 2013). Actin polymerisation drives the protrusion of lamellipodia and filopodia in the growth cones of axons (Gomez & Letourneau 2013). Therefore COTL1, which functions in actin polymerisation regulation, plays a critical role in neurite extension and neural crest migration (Hou et al. 2013). COTL1 was also found to be significantly involved in T-cell migration following CD28 stimulation (Kim et al, 2014). Suppression of COTL1 prevented lamellipodial protrusion at the T cell – B cell contact site owing to impaired T cell spreading in response to TCR ligation (Kim et al, 2014).

Role of COTL1 in Leukotriene synthesis

Both coactosin and COTL1 can bind to 5-LO to promote the formation of leukotrienes (LTs), lipid mediators that elicits inflammatory responses in allergic reactions, asthma and atherosclerosis (Funk, 2005). Since inflammatory reactions are part of cell defence mechanisms as well as chronic pathologies, it is imperative that regulation of 5-LO is a key step in maintaining the balance.

Any insult resulting in an increase in intracellular Ca²⁺ leads to the association of COTL1 to 5-LO in a 1:1 molar stoichiometry, prompting the translocation of the 5-LO-COTL1 complex to the perinuclear area (Fig. 3), where it docks to nuclear membrane phosphatidylcholine (PC) with the help of 5-LO activating protein (FLAP) (Esser et al. 2010; Liepinsh et al. 2004).

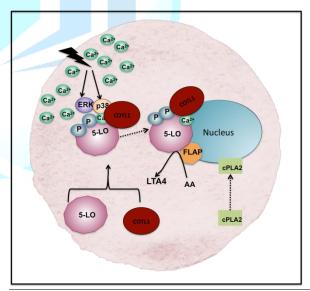


Figure 3: Schematic diagram showing 5-LO activation, translocation and LTA₄ synthesis in the cell.

A cellular stimuli leading to high intracellular Ca^{2+} or activation by MAP kinases ERK or p38 is thought to induce association of COTL1 with 5-LO. FLAP assists the translocation of the 5-LO- COTL1 complex to the nuclear membrane, where it converts AA to LTA4 (Adapted from Basavarajappa et al. 2014 and Rakonjac, 2009).

Citation: Anu SY, Lauto A, Myers SJ. The Role of Human Coactosin-Like Protein in Neurodegenerative Disorders (2017) Biochemistry and Modern Applications 1: 20-24.



The enzyme cytosolic phospholipase A2 (cPLA2), which is also translocated from cytosol to perinuclear area, releases arachidonic acid (AA) from PC, which is accessed by the 5-LO complex and converted into LTA₄ (Basavarajappa et al. 2014). 5-LO catalyses two initial steps in LT formation, the oxygenation of arachidonic acid to 5(S)hydroperoxy-6,8,11,14-eicosatetraenoic acid (5-HPETE) and the dehydration into epoxide. Thus, although 5-LO appears to be the initiator of the LT pathway, complete cellular 5-LO activity in presence of Ca2+, including the nuclear translocation, was displayed only in presence of COTL1 and was absent in a COTL1 knockdown model (Basavarajappa et al. 2014). LTA₄ undergoes further enzymatic modifications to form either LTB₄ or cistenyl leukotrienes LTC₄, LTD₄ or LTE4 (Rakonjac, 2009). Both LTB4 and cistenyl leukotrienes have been found to induce monocyte chemo attractant protein 1 (MCP-1), a chemokine heavily involved in neuroinflammatory responses (Huang et al, 2004; Ichiyama et al, 2005). Taken together, these data invariably suggest the significance of 5-LO-COTL1 association in neuroinflammation and potentially neurodegeneration. In addition to increased intracellular Ca2+, other stress stimuli causing an increase in p38 and ERK MAP kinase activation can subsequently phosphorylate 5-LO leading to its activation and ensuing LTA₄ production (Rådmark and Samuelsson, 2009).

Interaction of COTL1 with Organelles

There is a lack of research conducted on the direct interaction of coactosin and COTL1 on organelles outside of the cytoskeleton. However, there is an understanding on how cofilin, of which coactosin shares homology, interacts with mitochondria.

Cofilin plays a role in the regulation of mitochondrial action in apoptosis (Li et al, 2013). Apoptotic cell death is characterised by a distinct change in cellular architecture, demonstrated by blebbing of plasma membrane, externalization of phosphatidylserine, nuclear condensation and finally DNA fragmentation and release of cellular contents into circulation. Apoptosis occurs in three stages. The first is initiation in which a stress occurs that causes activation of one of various pathways to trigger a death signal that once sensed by the appropriate receptor leads to the second stage. The second stage is the commitment stage. Once a cell reaches this point the cell cannot reverse the process and will proceed to death. During this stage in apoptosis, apoptotic proteins interact with the mitochondria to permeabilise it to allow effectors, such as cytochrome c, to leave. Dephosphorylated Cofilin (Ser3) translocates from the cytosol to the mitochondria, and opens pores in the mitochondria to allow cytochrome c out, thereby playing a role in the commitment stage of apoptotic cell death (Li et al, 2013). Cytochrome c then forms a complex with activating factor-1 to cleave caspase-9, which activates Caspase-3. This step is considered part of the third stage of cell death, the execution stage, as it cleaves proteins that are essential to normal cell functioning. This leads to apoptosis resulting in cell death. It has also been shown that knockdown of cofilin leads to a reduction in the release of cytochrome c and hence apoptosis and cell death (Li et al. 2013; Li et al. 2015; Taha, Mullen & Obeid 2007).

Interaction of COTL1 with Sphingolipids

Sphingolipids are a class of lipids that contain a long chain sphingoid base (for example sphingosine). They can be found in mammalian plasma membrane and have been shown to play a role in a variety of cell signaling events including cell growth, cell death, cell differentiation and stress responses (Shayman 2000; Taha, Mullen & Obeid 2007).

Ceramides are a class of sphingolipids with a fatty acid linked to the amide group of the long-chain base. They serve as an intermediate for more complex sphingolipids such as phosphosphingolipids and glucosphingolipids. It has been proposed that ceramides play a role in signaling for programmed cell death based on two primary observations. The first is that agonists that induce apoptosis, such as CD95 and chemotherapeutic drugs, raise cellular levels of ceramide. The second is the artificial addition of cell permanent ceramides leads to an apoptotic response (de Chaves 2006).

There is evidence indicating the possibility that sphingolipids may play a role in neurodegenerative disorders. An increased level of ceramides has indeed been detected in the brain of Alzheimer's patients in early stages along with a reduced level of sulfatides in their brain grey and white matter and cerebrospinal fluid (Han et al, 2002). The elevated ceramide levels most likely arose from the breakdown of the sulfatides and can therefore be a potential biomarker for Alzheimer's (de Chaves 2006).

Due to the role of sphingolipids in the signalling of programmed cell death as well as its potential involvement in neurodegenerative disorders it is possible that COTL1 may interact with this biomolecule. An increase in COTL1 in endoplasmic reticulum fraction was in fact reported in lymphocytes isolated from Hereditary Sensory Neuropathy Type 1(HSN1) patients expressing V144D mutation in their serine palmitoyltransferase (SPT) long chain subunit 1 (SPTLC1) (Stimpson et al, 2016a). This increase in COTL1 was also further established in C133W, C133Y and V144D mutants in a neuronal cell model for HSN1 (Stimpson et al, 2016). SPT is a key enzyme involved in sphingolipid synthesis. The disease is characterised by degeneration of the dorsal root ganglion and presents with clinical onset from the second or third decades of the patient life (Stimpson et al, 2016b). Although the actual mechanism of COTL1 regulation is yet to be elucidated, its upregulation could be instigated by the increased oxidative stress upon the cellular cytoskeletal system, which can cause actin remodelling and potential axonal retraction in the neuron (Stimpson et al, 2016a). Increased COTL1 might also be triggering inflammatory pathways owing to its 5-LO binding property and subsequent LTA4 synthesis as described earlier. Whether COTL1 has any direct effects on sphingolipid metabolism or vice versa needs to be further investigated.

Currently there are very few other published reports on sphingolipid and COTL1 interaction. There is however one report that includes the interaction of a ceramide with coflilin-1 and F-actin in the stimulation of mouse embryonic stem cell migration (Park et al. 2013). The report indicates that the ceramide promoted the interaction between cofilin and F-actin to enhance mouse embryonic stem cell migration. This serves to further implicate a potential interaction between sphingolipids and COTL1

The Role of COTL1 in Neurodegeneration

In Alzheimer's and other neurodegenerative disorders, inflammatory processes (such as the ones that COTL1 help to regulate) play a crucial role. 5-LO plays an important role in inflammatory responses, which are triggered by the presence of plaques such as in Alzheimer's patients. This results in activation of microglia and astrocytes leading to neuron cell degeneration and death. This can however cause worsening of the disease rather than healing it. Furthermore it has been shown that downregulation of 5-LO improves the memory and synaptic functioning in animal Alzheimer's models. Also there has been some epidemiological evidence that may suggest that anti-inflammatory treatment can have a positive effect in preventing or minimising the effect of Alzheimer's (Czapski et al. 2016).

Coactosin and COTL1 have ADF/cofilin homology. This family of proteins is essential in the formation of neurites via the organisation of

Citation: Anu SY, Lauto A, Myers SJ. The Role of Human Coactosin-Like Protein in Neurodegenerative Disorders (2017) Biochemistry and Modern Applications 1: 20-24.



actin filaments (Sainath & Gallo 2015). For example cofilin increases actin depolymerisation and facilitates actin filament turnover as a way to regulate actin dynamics. Rods form spontaneously in neurons that overexpress ADF/cofilin, disrupting the microtubules. This does not cause cell death but does cause degeneration leading to a loss of synapses resulting in neurodegeneration (Minamide et al. 2000).

Cofilin aggregates and actin bundles have been observed in Alzheimer's brains. In low ATP environment (such as one caused by mitochondrial dysfunction or oxidative stress) there tends to be a higher ADP-actin and dephosphorylated cofilin concentration. Under these conditions the cofilin-actin complex readily forms into rods (neuropil thread structures). Rods that form as a result to this low ATP condition usually disappear shortly after, however in neurites with irreversible mitochondrial damage these rods become more permanent and can lead to loss of synaptic connections without loss of cells as mentioned above. This could be an explanation for the similar conditions that have been reported to occur in the early stages of neurodegenerative conditions (Maloney & Bamburg 2007; Whiteman et al. 2009).

Smith-Magenis syndrome (SMS) is caused by deletion of the short arm of chromosome 17. Its symptoms include neuro-behavioural abnormalities and congenital heart defects. The COTL1 gene is mapped to SMS common deletion region. This may indicate COTL1's involvement in the disease. The region also overlaps with a region associated with primitive neuro-ectodermal tumours. This suggests that COTL1 plays a role in DNA rearrangements of somatic cells (Nakatsura et al. 2002). Upregulated COTL1 expression has been identified in small cell lung cancer tissue thus suggesting it may be a biomarker or therapeutic target in these cancer patients (Jeong et al. 2010).

Conclusion

Neurodegenerative disorders are those that affect the normal functioning of the brain and its components. This occurs as a result of degradation of neurons including the formation of plaques and/or atypical protein assemblies. Despite being very common, very few of these disorders have a cure. As a result there is a large amount of research dedicated to understanding and finding cures or even early detection markers for these various diseases. One such example is the research conducted on the effect of COTL1 on neurodegeneration.

COTL1 being an actin binding protein plays a role in the maintenance of neuron structure, through its interaction with the cytoskeleton, and hence can potentially be involved in the loss of proper neuron structure leading to neurodegeneration. It is also involved in leukotriene synthesis for inflammatory responses, which can be triggered by plaques in patients suffering from these neurodegenerative disorders, and potentially worsens the condition. COTL1 shares homology with the ADF/cofilin family of proteins that play a role in mitochondrial apoptosis. This apoptotic effect can lead to unnecessary cell death in a low ATP environment and hence cause a large, unwarranted loss of neurons and/or the formation of neuropil thread structures, which can also lead to neurodegeneration. Furthermore sphingolipids also play a role in apoptosis and other important cell signalling events. The interaction between them and COTL1 may also be important in the triggering or control of neurodegeneration.

Currently there exists little research on the direct effect of COTL1 on neurodegeneration but what little there is does show promise in there being a link between the two. Subsequently more research needs to be conducted to further understand the connection, if any, between COTL1 and neurodegeneration.

References

- Basavarajappa D, Wan M, Lukic A, Steinhilber D, Samuelsson B, et al. Roles of coactosin-like protein (CLP) and 5lipoxygenase-activating protein (FLAP) in cellular leukotriene biosynthesis (2014) Proc Natl Acad Sci USA 111): 11371-11376.
- Czapski GA, Czubowicz K, Strosznajder JB, Strosznajder RP.
 The Lipoxygenases: Their Regulation and Implication in Alzheimer's Disease (2016) Neurochemical Research 41: 243-257.
- de Chaves EIP. Sphingolipids in apoptosis, survival and regeneration in the nervous system (2006) Biochimica et Biophysica Acta (BBA) – Biomembranes 1758: 1995-2015.
- Esser JR, M Hofmann, B Fischer, L Provost, Schneider G et al. Coactosin-like protein functions as stabilizing chaperone for 5lipoxygenase: role of tryptophan 102 (2010) Biochemical Journal 425: 265-274.
- Funk CD. Leukotriene modifiers as potential therapeutics for cardiovascular disease (2005) Nat Rev Drug Discov 4: 664-672.
- Han XDMH, McKeel Jr DW, Kelley J, Morris JC. Substantial sulfatide deficiency and ceramide elevation in very early Alzheimer's disease: potential role in disease pathogenesis (2002) J Neurochem 82: 809-818.
- Heemels MT. Neurodegeneration (2006) Nature 443: 767.
- Hellman M, Paavilainen VO, Naumanen P, Lappalainen P, Annila A, et al. Solution structure of coactosin reveals structural homology to ADF/cofilin family proteins (2004) FEBS Letters 576: 91-96.
- 9. Hou X, Katahira T, Ohashi K, Mizuno K, Sugiyama S, et al. Coactosin accelerates cell dynamism by promoting actin polymerization (2013) Developmental Biology 379: 53-63.
- Huang L, Zhao A, Wong F, Ayala JM, Struthers M, et al. Leukotriene B4 strongly increases monocyte chemoattractant protein-1 in human monocytes (2004) Arterioscler Thromb Vasc Biol 24: 1783-1788.
- Ichiyama T, Hasegawa M, Ueno Y, Makata H, Matsubara T, et al. Cysteinyl leukotrienes induce monocyte chemoattractant protein 1 in human monocytes/macrophages (2005) Clin Exp Allergy 35: 1214-1219.
- Jeong H-C, Kim G-I, Ho S-H, Lee K-H, Ko J-J, et al. Proteomic Analysis of Human Small Cell Lung Cancer Tissues: Up-Regulation of Coactosin-Like Protein-1 (2010) Journal of Proteomic Research 10: 269-276.
- Kevenaar JT, Hoogenraad CC. The axonal cytoskeleton: from organization to function (2015) Frontiers in molecular neuroscience 8: 44.
- Kim J, Shapiro MJ, Bamidele AO, Gurel P, Thapa P, et al. Coactosin-like 1 antagonizes cofilin to promote lamellipodial protrusion at the immune synapse (2014) PLoS One 9: e85090.
- 15. Lackie JM. The Dictionary of Cell and Molecular Biology (2007) Academic Press, Burlington.
- Li GB, Cheng Q, Liu L, Zhou T, Shan,CY, et al. Mitochondrial translocation of cofilin is required for allyl isothiocyanatemediated cell death via ROCK1/PTEN/PI3K signaling pathway (2013) Cell Communication and Signalling 11: 50.
- Li G, Zhou J, Budhraja A, Hu X, Chen Y, et al. Mitochondrial translocation and interaction of cofilin and Drp1 are required for erucin-induced mitochondrial fission and apoptosis (2015) Oncotarget 6: 1834-1849.

Citation: Anu SY, Lauto A, Myers SJ. The Role of Human Coactosin-Like Protein in Neurodegenerative Disorders (2017) Biochemistry and Modern Applications 1: 20-24.

Anu SY. Biochemistry and Modern Applications, 2017 PDF: 107, 1:1



- Liepinsh E, Rakonjac M, Boissonneault V, Provost P, Samuelsson B, et al. Letter to the Editor: NMR structure of human coactosin-like protein (2004) Journal of Biomolecular NMR 30: 353-356.
- Liu L, Wei Z, Wang Y, Wan M, Cheng Z et al. Crystal structure of human coactosin-like protein (2004) Journal of Molecular Biology 344: 317-323.
- Maloney MT, Bamburg JR. Cofilin-mediated neurodegeneration in Alzheimer's disease and other amyloidopathies (2007) Molecular Neurobiology 36: 201-204.
- Minamide LS, Striegl AM, Boyle JA, Meberg PJ, Bamburg JR. Neurodegenerative stimuli induce persistent ADF/cofilin-actin rods that disrupt distal neurite function (2000) Nature Cell Biology 2: 628-636.
- Nakatsura T, Senju S, Ito M, Nishimura Y, Itoh K. Cellular and humoral immune responses to a human pancreatic cancer antigen, coactosin-like protein, originally defined by the SEREX method (2002) European Journal of Immunology 32: 826-826.
- Park SS, Kim MO, Yun SP, Ryu JM, Park JH, et al. C(16)-Ceramide-induced F-actin regulation stimulates mouse embryonic stem cell migration: involvement of N-WASP/Cdc42/Arp2/3 complex and cofilin-1/α-actinin (2013) Biochimica et Biophysica acta 1831: 350-360.
- Poukkula M, Kremneva E, Serlachius M, Lappalainen P. Actindepolymerizing factor homology domain: a conserved fold performing diverse roles in cytoskeletal dynamics (2011) Cytoskeleton 68: 471-490.
- Rådmark O, Samuelsson B. 5-Lipoxygenase: mechanisms of regulation (2009) J Lipid Res 50: S40–S45.

- Rakonjac M. Studies on Coactosin-Like Protein Interaction with 5-Lipoxygenase, (2009) karolinska Institute 38605
- Rohrig U, Gersich G, Morozova L, Schleicher M, Wegner A. Coactosin interferes with the capping of actin filaments (1995) FEBS Letters 374: 284-286.
- Sainath R, Gallo G. Cytoskeletal and signaling mechanisms of neurite formation (2015) Cell and tissue research 59: 67-278.
- Shayman JA. Sphingolipids (2000) Kidney International 58: 11-26.
- Stimpson SE, Lauto A, Coorssen JR, Myers SJ. Isolation and Identification of ER Associated Proteins with Unique Expression Changes Specific to the V144D SPTLC1 Mutations in HSN-I (2016) Biochem Anal Biochem 5: 248.
- Stimpson SE, Shanu A, Coorssen JR. Myers SJ. Identifying Unique Protein Altera- tions Caused by SPTLC1 Mutations in a Transfected Neuronal Cell Model (2016) World Journal of Neuroscience 6: 325-347.
- Taha TA, Mullen TD, Obeid LM. A house divided: ceramide, sphingosine, and sphingosine-1-phosphate in programmed cell death (2006) Biochimica et Biophysica acta 1758: 2027-2036.
- 33. Whiteman IT, Gervasio OL, Cellen KM, Guillemin GJ, Jeong EV, et al. Activated actin-depolymerizing factor/cofilin sequesters phosphorylated microtubule-associated protein during the assembly of alzheimer-like neuritic cytoskeletal striations (2009) The Journal of Neuroscience 29: 12994-13005.