

kallisto live demo

Páll Melsted

Overview

- Downloading and installing kallisto
- Ingredients for a Coelacanth analysis
 - Nikaido *et al.* Genome Research, 2013
- Indexing
- Quantification



Downloading and installing kallisto

- Download the source code and compile?

The screenshot shows the GitHub repository page for `pachterlab/kallisto`. The repository is described as "Near-optimal RNA-Seq quantification" with a link to <https://pachterlab.github.io/kallisto>. It has 445 commits, 17 branches, 8 releases, and 4 contributors. The repository is currently on the `master` branch. A table of recent commits is shown, with the latest commit by `pmelsted` titled "Uses sparse output for batch data" dated June 3.

Commit	Description	Time
<code>ext/catch</code>	add catch as an external project	2 years ago
<code>src</code>	Uses sparse output for batch data	5 months ago
<code>test</code>	add snakemake and readme to test	a year ago
<code>unit_tests</code>	Adding test	2 years ago
<code>.gitignore</code>	Reformatting using astyle	2 years ago
<code>.gitmodules</code>	remove Catch as a submodule	2 years ago
<code>.ycm_extra_conf.py</code>	first cut of EM... not normalized	2 years ago
<code>CMakeLists.txt</code>	rearrange pthread to dynamically link it	a year ago

Downloading and installing kallisto



Mac: use Homebrew science

- Install Homebrew from <http://brew.sh>

```
> brew tap homebrew/science  
> brew install kallisto
```

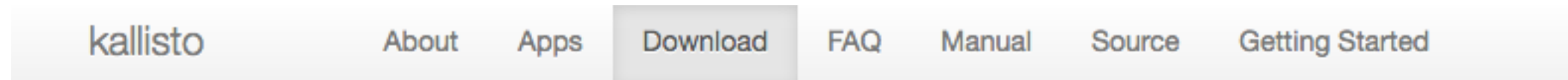
- Can also download binary, but Homebrew science adds a lot of useful tools



Downloading and installing kallisto



Linux: download the binary (works on all distributions) from <https://pachterlab.github.io/kallisto>



Download

Repository

The **kallisto** GitHub repository is [here](#).

Releases

Version	Date	Mac	Linux	Windows	Source
Release notes: v0.43.0	June 02, 2016	Mac	Linux	Windows	Source
Release notes: v0.42.5	April 04, 2016	Mac	Linux		Source

Downloading and installing kallisto



Windows: download binary and run using command line (cmd)



```
Administrator: C:\Windows\system32\cmd.exe
C:\Users\Pachter Lab Dell\Desktop\kallisto>
C:\Users\Pachter Lab Dell\Desktop\kallisto>kallisto.exe quant -i index.i
t reads_1.fastq.gz reads_2.fastq.gz

[quant] fragment length distribution will be estimated from the data
[index] k-mer length: 31
[index] number of targets: 15
[index] number of k-mers: 18,982
[index] number of equivalence classes: 22
[quant] running in paired-end mode
[quant] will process pair 1: reads_1.fastq.gz
                             reads_2.fastq.gz
[quant] finding pseudoalignments for the reads ... done
[quant] processed 10,000 reads, 10,000 reads pseudoaligned
[quant] estimated average fragment length: 178.077
[   en] quantifying the abundances ... done
[   en] the Expectation-Maximization algorithm ran for 52 rounds

C:\Users\Pachter Lab Dell\Desktop\kallisto>
```

Downloading and installing kallisto



Windows: download binary and run using command line (cmd)

```
C:\Windows\system32\cmd.exe
C:\Users\Pachter Lab Dell\Desktop\kallisto>
C:\Users\Pachter Lab Dell\Desktop\kallisto>kallisto.exe quant -i index.i
t reads_1.fastq.gz reads_2.fastq.gz

[quant] fragment length distribution will be estimated from the data
[index] k-mer length: 31
[index] number of targets: 15
[index] number of k-mers: 18,902
[index] number of equivalence classes: 22
[quant] running in paired-end mode
[quant] will process pair 1: reads_1.fastq.gz
                             reads_2.fastq.gz
[quant] finding pseudoalignments for the reads ... done
[quant] processed 10,000 reads, 10,000 reads pseudoaligned
[quant] estimated average fragment length: 178.097
[   em] quantifying the abundances ... done
[   em] the Expectation-Maximization algorithm ran for 52 rounds

C:\Users\Pachter Lab Dell\Desktop\kallisto>
```

Requirements for running kallisto


- Reference transcriptome
- Paired end reads, FASTQ files

Coelacanth analysis

- Reference transcriptome
- Paired end reads, FASTQ files
 - Downloaded from the short read archive (SRA)
 - 19 samples
 - 6 tissues - gill, kidney, pharynx, pectoral fin, pelvic fin, muscle
 - ~3 technical replicates per tissue



Reference transcriptome

 [BLAST/BLAT](#) | [BioMart](#) | [Tools](#) | [Downloads](#) | [Help & Documentation](#) | [Blog](#) | [Mirrors](#)

Search: for


Go



e.g. **BRCA2** or rat 5:62797383-63627669 or rs699 or coronary heart disease


Browse a Genome


Ensembl is a genome browser for vertebrate genomes that supports research in comparative genomics, evolution, sequence variation and transcriptional regulation. Ensembl annotate genes, computes multiple alignments, predicts regulatory function and collects disease data. Ensembl tools include BLAST, BLAT, BioMart and the Variant Effect Predictor (VEP) for all supported species.

Favourite genomes

 **Human**
GRCh38.p7

 **Human** 
GRCh37

 **Mouse**
GRCm38.p4

 **Zebrafish**
GRCz10



[Edit favourites](#)

All genomes


[View full list of all Ensembl species](#)

Still using Human GRCh37?

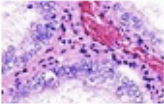
Go to

  GRCh37

Variant Effect Predictor



Gene expression in different tissues




Find SNPs and other variants for my gene

```
GTATATACATTCT  
CTTAAAGTCTT  
CTTCTAATTCT  
GTAACATTTTCC
```


Retrieve gene sequence

```
GCCTGACTTCGGGTGG  
GGGCTTGTGGCGGAGC  
GGGCTTGTGGCGGAGC  
AGGGGACAGATTGTGA  
CACCTCTGGAGCGGTT  
CCCACTCCAGCGTGGCG
```


Compare genes across species



Use my own data in Ensembl



ENCODE data in Ensembl



Reference transcriptome

e!Ensembl BLAST/BLAT | BioMart | Tools | Downloads | Help & Documentation | Blog | Mirrors

Search: for

e.g. **BRCA2** or **rat 5:62797383-63627669** or **rs699** or **coronary heart disease**

Fish

- Amazon molly
- Cave fish
- Cod
- Fugu
- Medaka
- Platyfish
- Spotted gar
- Stickleback
- Tetraodon
- Tilapia
- Zebrafish

Other vertebrates

- Coelacanth**
- Lamprey

Other chordates

- C.intestinalis
- C.savignyi

Other eukaryotes

- Caenorhabditis elegans
- Fruitfly
- Saccharomyces cerevisiae


-- Select a species --


[View full list of all Ensembl species](#)

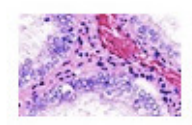
For vertebrate genomes that supports BLAT, evolution, sequence variation and BioMart, Ensembl annotate genes, computes multiple alignments, BLAT, BioMart and the Variant Effect Predictor.

Human
GRCh37

Zebrafish
GRCz10


Using Human GRCh37?
Go to 

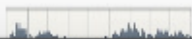
Variant Effect Predictor



Gene expression in different tissues


Find SNPs and other variants for my gene
GTATATACATTCT
CTTAAAGTCTT
CTTCTAAATTCT
GTAACATTTTCC


Retrieve gene sequence
GCCTGACTTCGGGTGG
GGGCTTGTGGCGGAGC
GGGCTTGTGGCGGAGC
AGGGGACAGATTGTGA
CACCTCTGGAGCGGTT
CCCACTCCAGCGTGGCG

Compare genes across species



Use my own data in Ensembl


ENCODE data in Ensembl


Reference transcriptome

[BLAST/BLAT](#) | [BioMart](#) | [Tools](#) | [Downloads](#) | [Help & Documentation](#) | [Blog](#) | [Mirrors](#)


Coelacanth (LatCha1) ▾


**Coelacanth**
Latimeria chalumnae


[Search all categories](#) ▾ [Go](#)

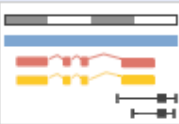
e.g. [protocadherin 1](#) or [JH126782.1:1475988-1509098](#) or [noggin](#)

Genome assembly: LatCha1 (GCA_000225785.1)

 [More information and statistics](#)


 [Download DNA sequence \(FASTA\)](#)


 [Display your data in Ensembl](#)


Example region

Comparative genomics

What can I find? Homologues, gene trees, and whole genome alignments across multiple species.

 [More about comparative analysis](#)

 [Download alignments \(EMF\)](#)

Example gene tree


What's New in Coelacanth release 86


- [Mouse Strains](#)
- [Ensembl 86 mart databases](#)
- [External reference projection](#)


[More news...](#)


Gene annotation

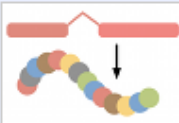
What can I find? Protein-coding and non-coding genes, splice variants, cDNA and protein sequences, non-coding RNAs.

 [More about this genebuild](#)

 [Download genes, cDNAs, ncRNA, proteins \(FASTA\)](#)


 [Update your old Ensembl IDs](#)


Example gene

Example transcript

Variation

This species currently has no variation database. However you can process your own variants using the Variant Effect Predictor:


 [Variant Effect Predictor](#)



Reference transcriptome

e!Ensembl BLAST/BLAT | BioMart | Tools | Downloads | Help & Documentation | Blog | Mirrors Search all species... [Login/Register](#)

Coelacanth (LatCha1) ▾

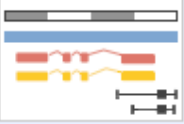
 **Coelacanth**
Latimeria chalumnae

[Search all categories](#) ▾ [Go](#)

e.g. [protocadherin 1](#) or [JH126782.1:1475988-1509098](#) or [noggin](#)

Genome assembly: LatCha1 (GCA_000225785.1)


- [More information and statistics](#)
- [Download DNA sequence \(FASTA\)](#)
- [Display your data in Ensembl](#)

 Example region

Comparative genomics

What can I find? Homologues, gene trees, and whole genome alignments across multiple species.


- [More about comparative analysis](#)
- [Download alignments \(EMF\)](#)


 Example gene tree

Gene annotation

What can I find? Protein-coding and non-coding genes, splice variants, cDNA and protein sequences, non-coding RNAs.


- [More about this genebuild](#)
- [Download genes, cDNAs, ncRNA, proteins \(FASTA\)](#)
- [Update your old Ensembl IDs](#)

 Example gene

 Example transcript

Variation

This species currently has no variation database. However you can process your own variants using the Variant Effect Predictor:


[Variant Effect Predictor](#) 

Only download the transcriptome

Reference transcriptome

e!Ensembl BLAST/BLAT | BioMart | Tools | Downloads | Help & Documentation | Blog | Mirrors Search all species... [Login/Register](#)




Coelacanth (LatCha1) ▾

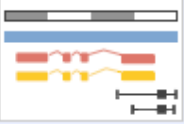
 **Coelacanth**
Latimeria chalumnae

[Search all categories](#) ▾ [Go](#)

e.g. [protocadherin 1](#) or [JH126782.1:1475988-1509098](#) or [noggin](#)



Genome assembly: LatCha1 (GCA_000225785.1)


-  [More information and statistics](#)
-  [Download DNA sequence \(FASTA\)](#)
-  [Display your data in Ensembl](#)

 [Example region](#)

Comparative genomics




What can I find? Homologues, gene trees, and whole genome alignments across multiple species.


-  [More about comparative analysis](#)
-  [Download alignments \(EMF\)](#)


 [Example gene tree](#)

Gene annotation

What can I find? Protein-coding and non-coding genes, splice variants, cDNA and protein sequences, non-coding RNAs.



-  [More about this genebuild](#)
-  [Download genes, cDNAs, ncRNA, proteins \(FASTA\)](#)
-  [Update your old Ensembl IDs](#)

 [Example gene](#)

 [Example transcript](#)

Variation







This species currently has no variation database. However you can process your own variants using the Variant Effect Predictor:

 [Variant Effect Predictor](#) 

Only download the transcriptome







Reference transcriptome

Index of /pub/release-86/fasta/latimeria_chalumnae/

Name	Size	Date Modified
 [parent directory]		
 cdna/		9/26/16, 3:06:00 PM
 cds/		9/26/16, 3:06:00 PM
 dna/		9/26/16, 3:11:00 PM
 ncrna/		9/26/16, 3:11:00 PM
 pep/		9/26/16, 3:11:00 PM

Reference transcriptome

Index of /pub/release-86/fasta/latimeria_chalumnae/

Name	Size	Date Modified
 [parent directory]		
 cdna/		9/26/16, 3:06:00 PM
 cds/		9/26/16, 3:06:00 PM
 dna/		9/26/16, 3:11:00 PM
 ncrna/		9/26/16, 3:11:00 PM
 pep/		9/26/16, 3:11:00 PM

Index of /pub/release-86/fasta/latimeria_chalumnae/cdna/

Name	Size	Date Modified
 [parent directory]		
 CHECKSUMS	134 B	9/23/16, 2:47:00 PM
 Latimeria_chalumnae.LatChal.cdna.abinitio.fa.gz	33.6 MB	9/20/16, 3:05:00 PM
 Latimeria_chalumnae.LatChal.cdna.all.fa.gz	16.1 MB	9/20/16, 2:34:00 PM
 README	3.1 kB	9/20/16, 3:05:00 PM

Indexing command

- Creates an index of the transcriptome reference that kallisto will use for quantification
 - Required only once per reference transcriptome

```
> time kallisto index -i index.idx  
Latimeria_chalumnae.LatCha1.cdna.all.fa.gz
```

Indexing command

- Creates an index of the transcriptome reference that kallisto will use for quantification
- Required only once per reference transcriptome

```
> time kallisto index -i index.idx  
Latimeria_chalumnae.LatCha1.cdna.all.fa.gz
```



where to store index

Indexing command

- Creates an index of the transcriptome reference that kallisto will use for quantification
- Required only once per reference transcriptome

```
> time kallisto index -i index.idx  
Latimeria_chalumnae.LatCha1.cdna.all.fa.gz
```

where to store index



Reference transcriptome

Indexing guide

What value of k should be chosen?

- k-mer tradeoffs
 - high k - more specific
 - low k - robust to sequencing errors

Indexing guide

What value of k should be chosen?

- k-mer tradeoffs
 - high k - more specific
 - low k - robust to sequencing errors
- general guide
 - for 75bp reads, use default, $k=31$
 - for 50bp reads, use default except if known issues
 - shorter reads, lower $k=25$ or $k=21$.

Quantification

Quantification is run separately for each sample

```
> time kallisto quant -i index/index.idx  
-o results/DRR002318 -b 30 -t 4  
data/DRR002318_1.fastq.gz  
data/DRR002318_2.fastq.gz
```

Quantification

Quantification is run separately for each sample

index constructed



```
> time kallisto quant -i index/index.idx  
-o results/DRR002318 -b 30 -t 4  
data/DRR002318_1.fastq.gz  
data/DRR002318_2.fastq.gz
```

Quantification

Quantification is run separately for each sample

where to put results

index constructed

```
> time kallisto quant -i index/index.idx  
-o results/DRR002318 -b 30 -t 4  
data/DRR002318_1.fastq.gz  
data/DRR002318_2.fastq.gz
```


Quantification

Quantification is run separately for each sample

where to put results

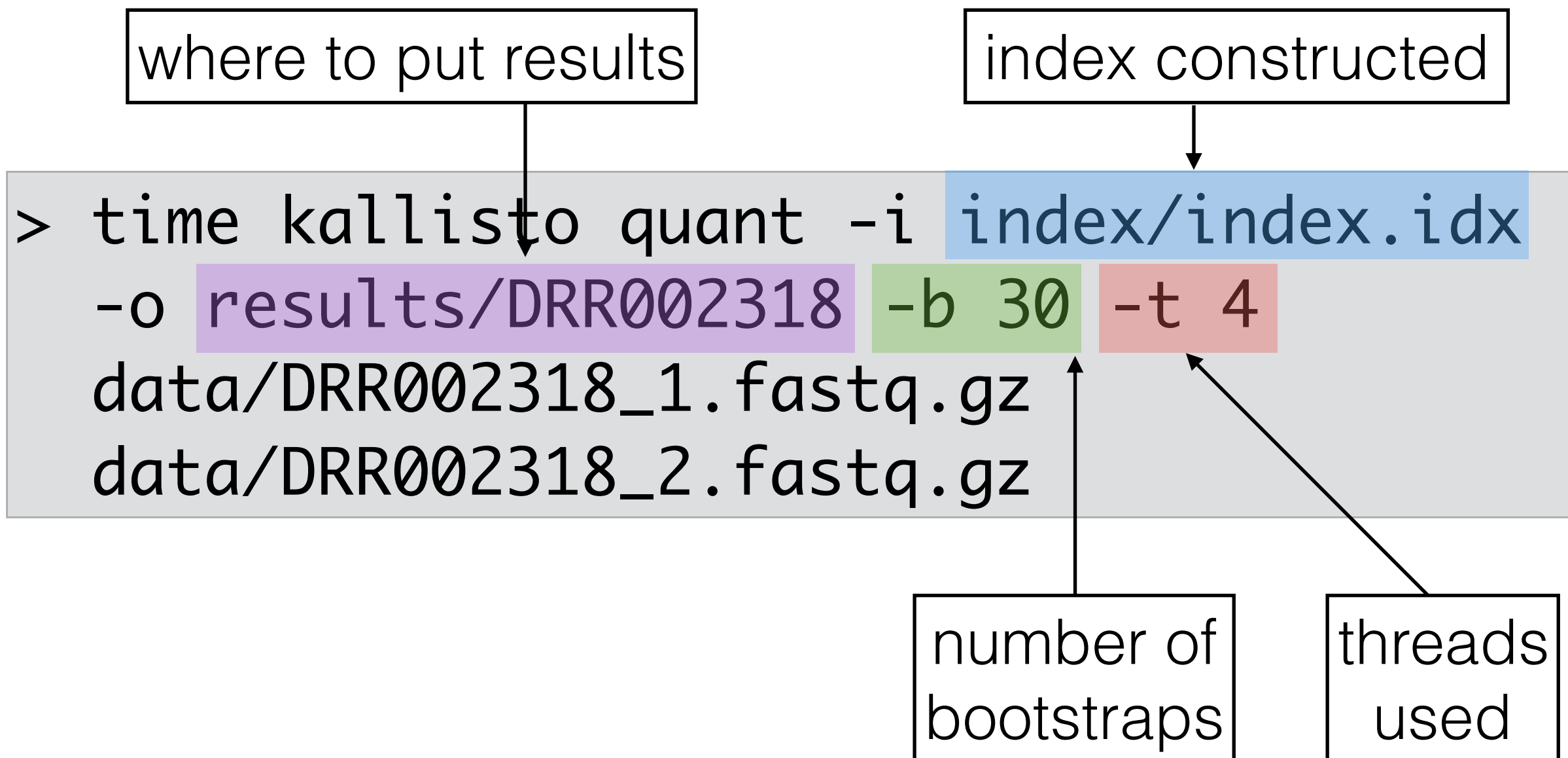
index constructed

```
> time kallisto quant -i index/index.idx  
-o results/DRR002318 -b 30 -t 4  
data/DRR002318_1.fastq.gz  
data/DRR002318_2.fastq.gz
```

number of
bootstraps

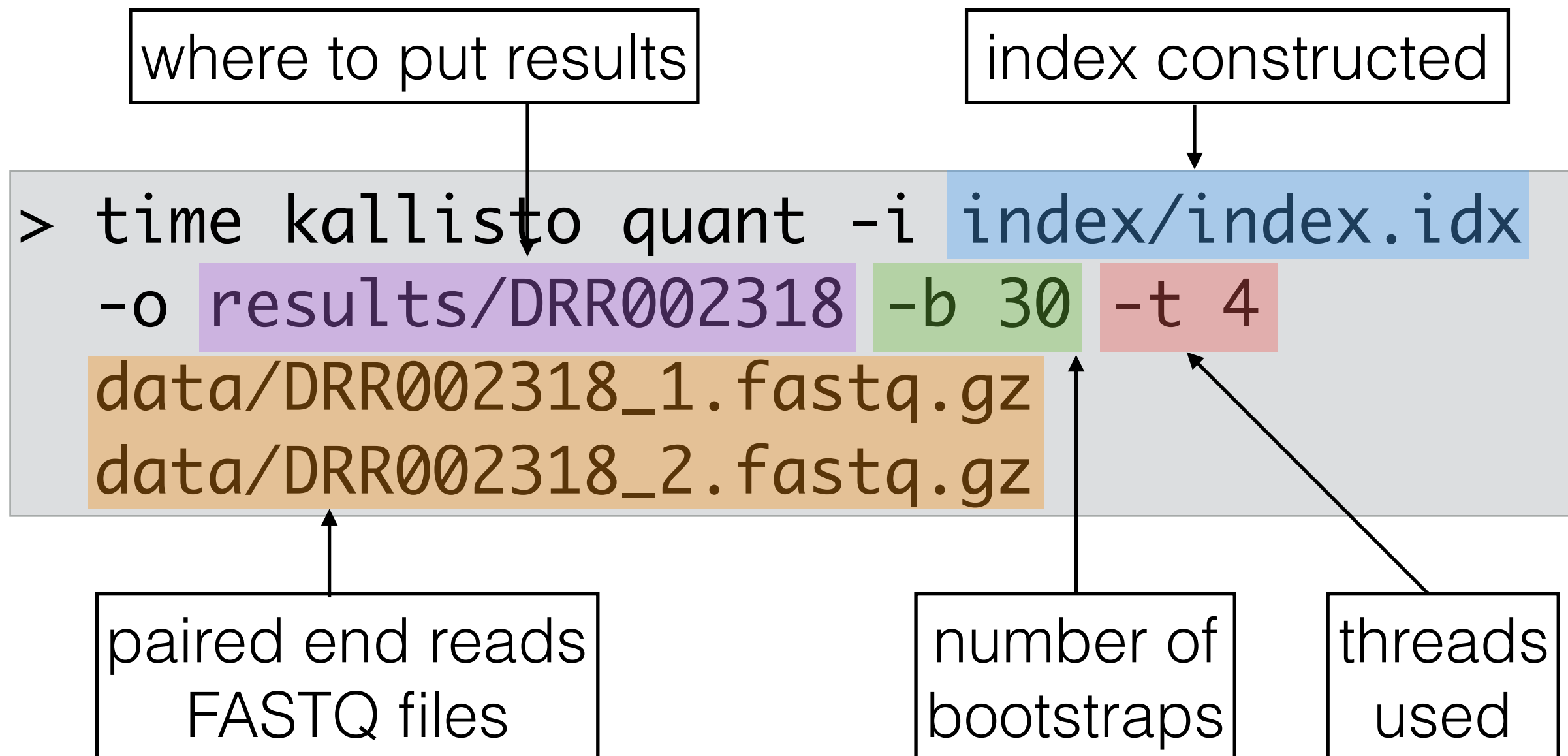
Quantification

Quantification is run separately for each sample



Quantification

Quantification is run separately for each sample



Results

- The output is stored in the specified directory
- **abundance.h5** - HDF5 compressed file, not human readable
 - contains the quantifications, all bootstraps and other information
- **abundance.tsv** - Tab Separated file
 - abundances and counts for each transcript
- **run_info.json** - JSON formatted file
 - information about the run