Untitled

Marcin Kosinski 25.10.2015

```
logitGD <- function(y, x, optim.method = "GDI", eps = 10e-4,</pre>
                     max.iter = 100, alpha = function(t)\{1/t\}, beta_0 = c(0,0))\{
  stopifnot(length(y) == length(x) & optim.method %in% c("GDI", "GDII", "SGDI")
            & is.numeric(c(max.iter, eps, x)) & all(c(eps, max.iter) > 0) &
               is.function(alpha))
  iter <- 0
  err <- list()
  err[[iter+1]] <- eps+1
  w_old <- beta_0
  res <-list()
  while(iter < max.iter && (abs(err[[ifelse(iter==0,1,iter)]]) > eps)){
    iter <- iter + 1
    if (optim.method == "GDI"){
      w_new <- w_old + alpha(iter)*updateWeightsGDI(y, x, w_old)</pre>
    if (optim.method == "GDII"){
      w_new <- w_old + as.vector(inverseHessianGDII(x, w_old)%*%
                                     updateWeightsGDI(y, x, w_old))
    if (optim.method == "SGDI"){
      w_new <- w_old + alpha(iter)*updateWeightsSGDI(y[iter], x[iter], w_old)</pre>
    res[[iter]] <- w_new
    err[[iter]] <- sqrt(sum((w_new - w_old)^2))
    w_old <- w_new
  return(list(steps = c(list(beta_0),res), errors = c(list(c(0,0)),err)))
}
updateWeightsGDI <- function(y, x, w_old){</pre>
  (1/length(y))*c(sum(y-p(w_old, x)), sum(x*(y-p(w_old, x))))
  \#c(sum(y-p(w_old, x)), sum(x*(y-p(w_old, x))))
}
updateWeightsSGDI <- function(y_i, x_i, w_old){</pre>
  c(y_i-p(w_old, x_i), x_i*(y_i-p(w_old, x_i)))
p <- function(w_old, x_i){</pre>
  1/(1+\exp(-w_{old}[1]-w_{old}[2]*x_{i}))
inverseHessianGDII <- function(x, w_old){</pre>
```

```
solve(
    matrix(c(
        sum(p(w_old, x)*(1-p(w_old, x))),
        sum(x*p(w_old, x)*(1-p(w_old, x))),
        sum(x*p(w_old, x)*(1-p(w_old, x))),
        sum(x*x*p(w_old, x)*(1-p(w_old, x)))
    ),
    nrow =2 )
}
```

```
# wstępna inicjalizacja parametrów
eps = 1e-5
                                          # warunek stopu.
n = length(data)
                                         # data jest listą ramek danych.
diff = eps + 1
                                         # różnice w oszacowaniach parametrów
                                         # między kolejnymi krokami.
learningRates = function(x) 1/x
                                         # długości kroku algorytmu.
beta old = numeric(0, length = k)
                                         # punkt startowy dlugosci k,
                                         # qdzie k to liczba zmiennych
                                         # objaśniających w modelu.
max.iter = 500
                                         # maksymalna liczba kroków.
                              # estymacja
i = 1
                                         # iterator kroku algorytmu.
while(i <= max.iter | diff < eps) do
  iter = ifelse(i mod n == 0, n, i mod n)# wybierz kolejny podzbiór batch.
  batch = data[[iter]]
 beta_new = beta_old - learningRates(i) * U_Batch(batch)
                                         # U Batch to częściowa funkcja
                                         # log-wiarogdności dla zaobserwowanego
                                         # zbioru `batch`
 diff = euclidean_dist(beta_new, beta_old) # odległość euklidesowa
 beta_old = beta_new
  i = i + 1
end while
return beta_new
```

```
beta_new[[i]] <- coxphSGD_batch(formula = formula, beta = beta_old,</pre>
        learningRate = learningRates(i), data = data[[ifelse(i%%n==0,n,i%%n)]])
    diff <- sqrt(sum((beta_new[[i]] - beta_old)^2))</pre>
    beta_old <- beta_new[[i]]</pre>
    i <- i + 1
  # return results
 list(Call = match.call(), epsilon = epsilon, learningRates = learningRates,
       steps = i, coefficients = c(list(beta_start), beta_new))
}
coxphSGD_batch <- function(formula, data, learningRate, beta){</pre>
  # collect times, status, variables and reorder samples
  # to make the algorithm more clear to read and track
  batchData <- prepareBatch(formula = formula, data = data)</pre>
  # calculate the log-likelihood for this batch sample
  partial_sum <- list()</pre>
  for(k in 1:nrow(batchData)) {
    # risk set for current time/observation
    risk_set <- batchData %>% filter(times >= batchData$times[k])
    nominator <- apply(risk_set[, -c(1,2)], MARGIN = 1, function(element){</pre>
      element * exp(element * beta)
    }) %>% rowSums()
    denominator <- apply(risk_set[, -c(1,2)], MARGIN = 1, function(element){</pre>
      exp(element * beta)
    }) %>% rowSums()
    partial_sum[[k]] <-</pre>
      batchData[k, "event"] * (batchData[k, -c(1,2)] - nominator/denominator)
  do.call(rbind, partial_sum) %>%
    colSums() -> U_batch
 return(beta + learningRate * U_batch)
checkArguments <- function(formula, data, learningRates,</pre>
                              beta 0, epsilon) {
  assert_that(is.list(data) & length(data) > 0)
  assert_that(length(unique(unlist(lapply(data, ncol)))) == 1)
  # + check names and types for every variables
  assert_that(is.function(learningRates))
  assert_that(is.numeric(epsilon))
  assert_that(is.numeric(beta_0))
    # check length of the start parameter
  if (length(beta_0) == 1) {
    beta_0 <- rep(beta_0, as.character(formula)[3] %>%
```

```
strsplit("\\+") %>%
                    unlist %>%
                    length)
  }
  return(beta_0)
x <- runif(1000)
z < -2 + 3*x
pr <- 1/(1+exp(-z))
y <- rbinom(1000,1,pr)
logitGD(y, x, optim.method = "GDI", eps = 10e-5, max.iter = 500)$steps -> GDI
logitGD(y, x, optim.method = "GDII", eps = 10e-5, max.iter = 500)$steps -> GDII
ind <- sample(length(y))</pre>
logitGD(y[ind], x[ind], optim.method = "SGDI",
        max.iter = 500, eps = 10e-5)$steps -> SGDI.1
ind2 <- sample(length(y))</pre>
logitGD(y[ind2], x[ind2], optim.method = "SGDI",
        max.iter = 500, eps = 10e-5)$steps -> SGDI.2
ind3 <- sample(length(y))</pre>
logitGD(y[ind3], x[ind3], optim.method = "SGDI",
        max.iter = 500, eps = 10e-5)$steps -> SGDI.3
ind4 <- sample(length(y))</pre>
logitGD(y[ind4], x[ind4], optim.method = "SGDI",
        max.iter = 500, eps = 10e-5)$steps -> SGDI.4
ind5 <- sample(length(y))</pre>
logitGD(y[ind5], x[ind5], optim.method = "SGDI",
        max.iter = 500, eps = 10e-5)$steps -> SGDI.5
do.call(rbind, c(GDI, GDII, SGDI.1, SGDI.2, SGDI.3, SGDI.4, SGDI.5)) -> coeffs
unlist(lapply(list(GDI, GDII, SGDI.1, SGDI.2, SGDI.3, SGDI.4, SGDI.5), length)) -> algorithm
data2viz <- cbind(as.data.frame(coeffs),</pre>
      algorithm = unlist(mapply(rep, c("GDI", "GDII", "SGDI.1", "SGDI.2", "SGDI.3", "SGDI.4", "SGDI.5")
names(data2viz)[1:2] <- c("Intercept", "X")</pre>
library(ggplot2); library(ggthemes)
ggplot(data2viz) +
  geom_point(aes(x = X, y = Intercept, col = algorithm)) +
  geom_line(aes(x = X, y = Intercept, col = algorithm,
                group = algorithm)) +
  theme_tufte(base_size = 20)
logitGD() asda graphSGD()
graphSGD(c(0,0), y, x)
graphSGD(c(3.1,2.1), y, x)
graphSGD(c(4,3), y, x)
graphSGD(c(1,2), y, x)
```

```
dataCox <- function(N, lambda, rho, x, beta, censRate){</pre>
  # real Weibull times
  u <- runif(N)
  Treal <- (-\log(u) / (lambda * exp(x %*% beta)))^(1 / rho)
  # censoring times
  Censoring <- rexp(N, censRate)</pre>
  # follow-up times and event indicators
  time <- pmin(Treal, Censoring)</pre>
  status <- as.numeric(Treal <= Censoring)</pre>
  # data set
  data.frame(id=1:N, time=time, status=status, x=x)
x <- matrix(sample(0:1, size = 40, replace = TRUE), ncol = 2)
head(dataCox(20, 3, 2, x, beta = c(2,3), 5))
  id
           time status x.1 x.2
1 1 0.04217186
                    0 0
                             1
2 2 0.04583595
                    0 0
                             1
3 3 0.03361776
                   0 1 1
4 4 0.10477472
                    0 0 0
5 5 0.22068183
                    0 0
                             0
6 6 0.12595706
                   1 0
graphSGD(c(0,0), y, x, 4561);graphSGD(c(0,0), y, x, 456)
graphSGD(c(2,1), y, x, 4561);graphSGD(c(2,1), y, x, 456);
graphSGD(c(1,0), y, x, 4561);graphSGD(c(1,0), y, x, 456);
graphSGD(c(2.1,3.1), y, x, 4561)graphSGD(c(2.1,3.1), y, x, 456)
x <- matrix(sample(0:1, size = 20000, replace = TRUE), ncol = 2)
dCox \leftarrow dataCox(10^4, lambda = 3, rho = 2, x, beta = c(1,3), censRate = 5)
vizCoxSGD(dCox)
```

```
coxphSGD <- function(formula, data, learningRates = function(x){1/x},</pre>
                     beta_0 = 0, epsilon = 1e-5) {
  checkArguments(formula, data, learningRates,
                   beta_0, epsilon) -> beta_old # check arguments
  n <- length(data)</pre>
  diff \leftarrow epsilon + 1
  i <- 1
  beta_new <- list() # steps are saved in a list so that they can
                      # be tracked in the future
  # estimate
  while(i <= n & diff > epsilon) {
    beta_new[[i]] <- coxphSGD_batch(formula = formula, data = data[[i]],</pre>
                       learningRate = learningRates(i), beta = beta_old)
    diff <- sqrt(sum((beta_new[[i]] - beta_old)^2))</pre>
    beta_old <- beta_new[[i]]</pre>
    i <- i + 1
 }
  # return results
 list(Call = match.call(), coefficients = beta_new, epsilon = epsilon,
       learningRates = learningRates, steps = i)
}
coxphSGD_batch <- function(formula, data, learningRate, beta){</pre>
  # collect times, status, variables and reorder samples
  \# to make the algorithm more clear to read and track
  batchData <- prepareBatch(formula = formula, data = data)</pre>
  # calculate the log-likelihood for this batch sample
  partial_sum <- list()</pre>
  for(k in 1:nrow(batchData)) {
    # risk set for current time/observation
    risk_set <- batchData %>% filter(times <= batchData$times[k])</pre>
    nominator <- apply(risk_set[, -c(1,2)], MARGIN = 1, function(element){</pre>
      element * exp(element * beta)
    }) %>% rowSums()
    denominator <- apply(risk_set[, -c(1,2)], MARGIN = 1, function(element){</pre>
      exp(element * beta)
    }) %>% rowSums()
    partial_sum[[k]] <-</pre>
      batchData[k, "event"] * (batchData[k, -c(1,2)] - nominator/denominator)
  do.call(rbind, partial_sum) %>%
    colSums() -> U_batch
 return(beta + learningRate * U_batch)
}
```

```
prepareBatch <- function(formula, data) {</pre>
  # Parameter identification as in `survival::coxph()`.
  Call <- match.call()</pre>
  indx <- match(c("formula", "data"),</pre>
                 names(Call), nomatch = 0)
  if (indx[1] == 0)
      stop("A formula argument is required")
  temp <- Call[c(1, indx)]</pre>
  temp[[1]] <- as.name("model.frame")</pre>
  mf <- eval(temp, parent.frame())</pre>
  Y <- model.extract(mf, "response")</pre>
  if (!inherits(Y, "Surv"))
      stop("Response must be a survival object")
  type <- attr(Y, "type")</pre>
  if (type != "right" && type != "counting")
      stop(paste("Cox model doesn't support \"", type, "\" survival data",
          sep = "")
  # collect times, status, variables and reorder samples
  # to make the algorithm more clear to read and track
  cbind(event = unclass(Y)[,2], # 1 indicates event, 0 indicates cens
        times = unclass(Y)[,1],
        mf[, -1]) %>%
    arrange(times)
}
```

```
logitGD <- function(y, x, optim.method = "GDI", eps = 10e-4,</pre>
                    max.iter = 100, alpha = function(t)\{1/t\}, beta_0 = c(0,0))\{
  stopifnot(length(y) == length(x) & optim.method %in% c("GDI", "GDII", "SGDI")
            & is.numeric(c(max.iter, eps, x)) & all(c(eps, max.iter) > 0) &
              is.function(alpha))
  iter <- 0
  err <- list()
  err[[iter+1]] <- eps+1
  w_old <- beta_0
  res <-list()
  while(iter < max.iter && (abs(err[[ifelse(iter==0,1,iter)]]) > eps)){
    iter <- iter + 1
    if (optim.method == "GDI"){
      w_new <- w_old + alpha(iter)*updateWeightsGDI(y, x, w_old)</pre>
    if (optim.method == "GDII"){
      w new <- w old + as.vector(inverseHessianGDII(x, w old)%*%
                                    updateWeightsGDI(y, x, w_old))
```

```
if (optim.method == "SGDI"){
      w_new <- w_old + alpha(iter)*updateWeightsSGDI(y[iter], x[iter], w_old)</pre>
    res[[iter]] <- w_new
    err[[iter]] <- sqrt(sum((w_new - w_old)^2))</pre>
    w_old <- w_new
  }
  return(list(steps = c(list(beta_0),res), errors = c(list(c(0,0)),err)))
}
updateWeightsGDI <- function(y, x, w old){
  \#(1/length(y))*c(sum(y-p(w_old, x)), sum(x*(y-p(w_old, x))))
  c(sum(y-p(w_old, x)), sum(x*(y-p(w_old, x))))
}
updateWeightsSGDI <- function(y_i, x_i, w_old){</pre>
  c(y_i-p(w_old, x_i), x_i*(y_i-p(w_old, x_i)))
p <- function(w_old, x_i){</pre>
  1/(1+exp(-w_old[1]-w_old[2]*x_i))
inverseHessianGDII <- function(x, w_old){</pre>
  solve(
    matrix(c(
      sum(p(w_old, x)*(1-p(w_old, x))),
      sum(x*p(w_old, x)*(1-p(w_old, x))),
      sum(x*p(w_old, x)*(1-p(w_old, x))),
      sum(x*x*p(w_old, x)*(1-p(w_old, x)))
    ),
    nrow = 2)
  )
}
set.seed(1283)
x <- runif(10000)
z < -2 + 3*x
pr <- 1/(1+exp(-z))
y <- rbinom(10000,1,pr)
global_loglog <- function(beta1, beta2, xX, yY){</pre>
  sum(yY*(beta1+beta2*xX)-log(1+exp(beta1+beta2*xX)))
calculate_outer <- function(x, y){</pre>
  ## contours
  outer_res <- outer(seq(0,4, length = 100),</pre>
                      seq(0,5, length = 100),
```

```
Vectorize( function(beta1,beta2){
                        global_loglog(beta1, beta2, xX = x, yY = y)
  )
  outer_res_melted <- melt(outer_res)</pre>
  outer_res_melted$Var1 <- as.factor(outer_res_melted$Var1)</pre>
  levels(outer_res_melted$Var1) <- as.character(seq(0,4, length = 100))</pre>
  outer_res_melted$Var2 <- as.factor(outer_res_melted$Var2)</pre>
  levels(outer_res_melted$Var2) <- as.character(seq(0,5, length = 100))</pre>
  outer_res_melted$Var1 <- as.numeric(as.character(outer_res_melted$Var1))</pre>
  outer_res_melted$Var2 <- as.numeric(as.character(outer_res_melted$Var2))</pre>
  return(outer_res_melted)
library(ggplot2); library(ggthemes); library(reshape2)
graphSGD <- function(beta, y, x, seed = 4561, outerBounds = calculate_outer(x,y)){</pre>
  set.seed(seed)
  beta <- rev(beta)</pre>
 logitGD(y, x, optim.method = "GDI", beta 0 = beta,
          eps = 10e-4, max.iter = 10000,
          alpha = function(t) \{1/(1000*sqrt(t))\})$steps -> GDI.S
  logitGD(y, x, optim.method = "GDII", beta_0 = beta,
          eps = 10e-4, max.iter = 5000)$steps -> GDII
  ind2 <- sample(length(y))</pre>
  logitGD(y[ind2], x[ind2], optim.method = "SGDI", beta_0 = beta,
          max.iter = 10000, eps = 10e-4,
          alpha = function(t){1/sqrt(t)})$steps -> SGDI.1.S
  ind3 <- sample(length(y))</pre>
  logitGD(y[ind3], x[ind3], optim.method = "SGDI", beta_0 = beta,
          max.iter = 10000, eps = 10e-4,
          alpha = function(t){5/sqrt(t)})$steps -> SGDI.5.S
  ind4 <- sample(length(y))</pre>
  logitGD(y[ind4], x[ind4], optim.method = "SGDI", beta_0 = beta,
          max.iter = 10000, eps = 10e-4,
          alpha = function(t){6/sqrt(t)})$steps -> SGDI.6.S
  do.call(rbind, c(GDI.S, GDII, SGDI.1.S, SGDI.5.S, SGDI.6.S)) -> coeffs
  unlist(lapply(list(GDI.S, GDII, SGDI.1.S, SGDI.5.S, SGDI.6.S),
                length)) -> algorithm
  data2viz <- cbind(as.data.frame(coeffs),</pre>
  algorithm = unlist(mapply(rep,
                               c(paste("GDI", length(GDI.S), "steps"),
                               paste("GDII", length(GDII), "steps"),
                               paste("SGDI.1", length(SGDI.1.S), "steps"),
```

```
paste("SGDI.5", length(SGDI.5.S), "steps"),
                               paste("SGDI.6", length(SGDI.6.S), "steps")),
                             algorithm)))
  names(data2viz)[1:2] <- c("Intercept", "X")</pre>
  data2viz$algorithm <- factor(data2viz$algorithm, levels = rev(levels(data2viz$algorithm)))</pre>
  beta[2] -> XX
  beta[1] -> YY
  ggplot()+
    geom_path(aes(x = data2viz$X,
                  y = data2viz$Intercept,
                  col = data2viz$algorithm,
                   group = data2viz$algorithm), size = 1) +
    geom_point(aes(as.vector(round(coefficients(glm(y~x,
                           family = 'binomial')), 2)[2]),
                    as.vector(round(coefficients(glm(y~x,
                           family = 'binomial')), 2)[1])),
               col = "black", size = 4, shape = 15) +
    geom_point(aes(x=XX, y=YY),
               col = "black", size = 4, shape = 17) +
    theme_bw(base_size = 20) +
    theme(panel.border = element_blank(),
          legend.key = element_blank()) +
    scale_colour_brewer(palette="Set1", name = 'Algorithm') +
    xlab('X') +
    ylab('Intercept') -> pl_g
 return(pl_g)
}
full_cox_loglik <- function(beta1, beta2, x1, x2, censored){</pre>
  sum(rev(censored)*(beta1*rev(x1) + beta2*rev(x2) -
                        log(cumsum(exp(beta1*rev(x1) + beta2*rev(x2))))))
}
calculate_outer_cox <- function(x1, x2, censored){</pre>
  ## contours
  outer_res <- outer(seq(-1,3, length = 100),
           seq(0,4, length = 100),
           Vectorize( function(beta1,beta2){
             full_cox_loglik(beta1, beta2, x1 = x1, x2 = x2, censored = censored)
           } )
  outer_res_melted <- melt(outer_res)</pre>
  outer_res_melted$Var1 <- as.factor(outer_res_melted$Var1)</pre>
  levels(outer_res_melted$Var1) <- as.character(seq(-1,3, length = 100))</pre>
  outer_res_melted$Var2 <- as.factor(outer_res_melted$Var2)</pre>
  levels(outer_res_melted$Var2) <- as.character(seq(0,4, length = 100))</pre>
  outer_res_melted$Var1 <- as.numeric(as.character(outer_res_melted$Var1))</pre>
  outer_res_melted$Var2 <- as.numeric(as.character(outer_res_melted$Var2))</pre>
  return(outer_res_melted)
```

```
simulateCoxSGD <- function(dCox = dCox, learningRates = function(x){1/x},</pre>
                      epsilon = 1e-03, beta_0 = c(0,0), max.iter = 100){
 sample(1:90, size = 10^4, replace = TRUE) -> group
 split(dCox, group) -> dCox_splitted
 coxphSGD(Surv(time, status)~x.1+x.2, data = dCox_splitted,
           epsilon = epsilon, learningRates = learningRates,
           beta 0 = beta 0, max.iter = max.iter*90) -> estimates
 sample(1:60, size = 10^4, replace = TRUE) -> group
 split(dCox, group) -> dCox_splitted
 coxphSGD(Surv(time, status)~x.1+x.2, data = dCox_splitted,
           epsilon = epsilon, learningRates = learningRates,
          beta_0 = beta_0, max.iter = max.iter*60) -> estimates2
 sample(1:120, size = 10^4, replace = TRUE) -> group
 split(dCox, group) -> dCox_splitted
 coxphSGD(Surv(time, status)~x.1+x.2, data = dCox_splitted,
           epsilon = epsilon, learningRates = learningRates,
          beta_0 = beta_0, max.iter = max.iter*120) -> estimates3
 sample(1:200, size = 10^4, replace = TRUE) -> group
 split(dCox, group) -> dCox_splitted
 coxphSGD(Surv(time, status)~x.1+x.2, data = dCox_splitted,
           epsilon = epsilon, learningRates = learningRates,
           beta_0 = beta_0, max.iter = max.iter*200) -> estimates4
 sample(1:30, size = 10^4, replace = TRUE) -> group
 split(dCox, group) -> dCox_splitted
 coxphSGD(Surv(time, status)~x.1+x.2, data = dCox_splitted,
           epsilon = epsilon, learningRates = learningRates,
           beta_0 = beta_0, max.iter = max.iter*30) -> estimates5
 sample(1:10, size = 10^4, replace = TRUE) -> group
 split(dCox, group) -> dCox_splitted
 coxphSGD(Surv(time, status)~x.1+x.2, data = dCox_splitted,
           epsilon = epsilon, learningRates = learningRates,
          beta_0 = beta_0, max.iter = max.iter*10) -> estimates6
 t(simplify2array(estimates$coefficients)) %>%
    as.data.frame() -> df1
 t(simplify2array(estimates2$coefficients)) %>%
    as.data.frame() -> df2
 t(simplify2array(estimates3$coefficients)) %>%
    as.data.frame() -> df3
 t(simplify2array(estimates4$coefficients)) %>%
    as.data.frame() -> df4
 t(simplify2array(estimates5$coefficients)) %>%
    as.data.frame() -> df5
 t(simplify2array(estimates6$coefficients)) %>%
```

```
as.data.frame() -> df6
  df1 %>%
    mutate(version = paste("90 batches,", nrow(df1), " steps")) %>%
   bind_rows(df2 %>%
                mutate(version = paste("60 batches,", nrow(df2), " steps"))) %>%
   bind_rows(df3 %>%
                mutate(version = paste("120 batches,", nrow(df3), " steps"))) %>%
   bind rows(df4 %>%
                mutate(version = paste("200 batches,", nrow(df4), " steps"))) %>%
   bind rows(df5 %>%
                mutate(version = paste("30 batches,", nrow(df5), " steps"))) %>%
    bind_rows(df6 %>%
                mutate(version = paste("10 batches,", nrow(df6), " steps"))) -> d2ggplot
 return(list(d2ggplot = d2ggplot, est1 = estimates, est2 = estimates2,
              est3 = estimates3, est4 = estimates4, est5 = estimates5))
simulateCoxSGD(dCox, learningRates = function(x){1/(100*sqrt(x))},
               max.iter = 10, epsilon = 1e-5) -> d2ggplot
d2ggplot -> backpack
d2ggplot <- d2ggplot$d2ggplot
beta_0 = c(0,0)
solution = c(1,3)
pdf(file = "b_0_0_iter_10_e-5_100sqrt_878.pdf", width = 10, height = 10)
ggplot() +
  stat contour(aes(x=outerCox$Var1,
                  y=outerCox$Var2,
                   z=outerCox$value),
               bins = 40, alpha = 0.25) +
  geom_path(aes(d2ggplot$V1, d2ggplot$V2, group = d2ggplot$version,
                colour = d2ggplot$version), size = 1) +
  theme_bw(base_size = 20) +
  theme(panel.border = element_blank(),
        legend.key = element_blank(), legend.position = "top") +
  scale_colour_brewer(palette="Dark2", name = 'Algorithm \n & Steps') +
  geom_point(aes(x = beta_0[1], y = beta_0[2]), col = "black", size = 4, shape = 17) +
  geom_point(aes(x = solution[1], y = solution[2]), col = "black", size = 4, shape = 15) +
  xlab("X1") + ylab("X2") +
  guides(col = guide_legend(ncol = 3))
dev.off()
extractSurvival <- function(cohorts){</pre>
  survivalData <- list()</pre>
  for(i in cohorts){
    get(pasteO(i, ".clinical"), envir = .GlobalEnv) %>%
                select(patient.bcr_patient_barcode,
                             patient.vital_status,
                             patient.days_to_last_followup,
                             patient.days_to_death ) %>%
```

```
mutate(bcr_patient_barcode = toupper(patient.bcr_patient_barcode),
                       patient.vital status = ifelse(patient.vital status %>%
                                                  as.character() == "dead",1,0),
                   barcode = patient.bcr_patient_barcode %>%
                                     as.character(),
                 times = ifelse( !is.na(patient.days_to_last_followup),
                      patient.days_to_last_followup %>%
                        as.character() %>%
                        as.numeric(),
               patient.days to death %>%
                        as.character() %>%
                        as.numeric() )
                     ) %>%
   filter(!is.na(times)) -> survivalData[[i]]
  do.call(rbind,survivalData) %>%
    select(bcr_patient_barcode, patient.vital_status, times) %>%
    unique
}
extractMutations <- function(cohorts, prc){</pre>
  mutationsData <- list()</pre>
  for(i in cohorts){
    get(paste0(i, ".mutations"), envir = .GlobalEnv) %>%
      select(Hugo_Symbol, bcr_patient_barcode) %>%
      filter(nchar(bcr_patient_barcode)==15) %>%
      filter(substr(bcr_patient_barcode, 14, 15)=="01") %>%
      unique -> mutationsData[[i]]
  do.call(rbind,mutationsData) %>% unique -> mutationsData
  mutationsData %>%
    group_by(Hugo_Symbol) %>%
    summarise(count = n()) %>%
    arrange(desc(count)) %>%
    mutate(count_prc = count/length(unique(mutationsData$bcr_patient_barcode))) %>%
    filter_(paste0("count_prc > ",prc)) %>%
    select(Hugo_Symbol) %>%
    unlist -> topGenes
  mutationsData %>%
    filter(Hugo_Symbol %in% topGenes) -> mutationsData_top
  mutationsData_top %>%
    dplyr::group_by(bcr_patient_barcode) %>%
    dplyr::summarise(count = n()) %>%
    group_by(count) %>%
    summarise(total = n()) %>%
```

```
arrange(desc(count))
#
#
    mutationsData_top %>%
      spread(Huqo_Symbol, bcr_patient_barcode) -> mutationsData_top_sp
  as.data.table(mutationsData_top) -> mutationsData_top_DT
  dcast.data.table(mutationsData_top_DT, bcr_patient_barcode ~ Hugo_Symbol , fill = 0) %>%
    as.data.frame -> mutationsData top dcasted
  mutationsData_top_dcasted[,-1][mutationsData_top_dcasted[,-1] != "0"] <- 1</pre>
  mutationsData_top_dcasted -> result
  names(result) <- gsub(names(result),pattern = "-", replacement = "")</pre>
  result
extractCohortIntersection <- function(){</pre>
  data(package = "RTCGA.mutations")$results[,3] %>%
    gsub(".mutations", "", x = .) -> mutations_data
  data(package = "RTCGA.clinical")$results[,3] %>%
    gsub(".clinical", "", x = .) -> clinical_data
  intersect(mutations_data, clinical_data)
}
prepareCoxDataSplit <- function(mutationsData, survivalData, groups, seed = 4561){</pre>
  mutationsData %>%
  mutate(bcr_patient_barcode = substr(bcr_patient_barcode,1,12)) %>%
  left_join(survivalData,
            by = "bcr_patient_barcode") -> coxData
  coxData <- coxData[, -c(1,2)]</pre>
  coxData %>%
    filter(times > 0) %>%
    filter(!is.na(times)) -> coxData
  apply(coxData[,-c(1092, 1093)], MARGIN = 2, function(x){
    as.numeric(as.character(x))
  ) \rightarrow coxData[,-c(1092, 1093)]
  set.seed(seed)
  sample(groups, replace = TRUE, size = 6085) -> groups
  split(coxData, groups) #coxData_split
}
prepareForumlaSGD <- function(coxData){</pre>
  as.formula(paste("Surv(times, patient.vital_status) ~ ",
                   paste(names(coxData[[1]])[-c(1092, 1093)],
                          collapse="+"), collapse = ""))
}
```

```
full_cox_loglik_matrix <- function(beta, x, censored){</pre>
  order(x$times) -> order2
  x[order2, ] -> xORD
  censored[order2] -> censORD
  sum(censORD*(beta%*%x[, -which(names(x)=='times')] -
                       log(cumsum(exp(beta1*rev(x1) + beta2*rev(x2))))))
}
library(dplyr)
Attaching package: 'dplyr'
The following objects are masked from 'package:stats':
    filter, lag
The following objects are masked from 'package:base':
    intersect, setdiff, setequal, union
library(RTCGA.clinical)
Loading required package: RTCGA
Loading required package: knitr
Welcome to the RTCGA (version: 1.1.10).
library(RTCGA.mutations)
library(data.table)
Attaching package: 'data.table'
The following objects are masked from 'package:dplyr':
    between, last
library(coxphSGD)
Loading required package: survival
Attaching package: 'coxphSGD'
The following object is masked _by_ '.GlobalEnv':
    dataCox
```

Do analizy badającej wpływ występowania mutacji genów na czas przeżycia wykorzystano dane kliniczne i dane o występujących u pacjentów mutacjach genetycznych. Starano się wykorzystać dane ze wszystkich 38 dostępnych kohort nowotworowych z badania *The Cancer Genome Atlas* (TCGA), jednak nie dla wszystkich kohort umieszczono w badaniu dane o mutacjach. Częśc wspólną nazw dla kohort zawierających zarówno dane kliniczne oraz dane o mutacjach wygenerowaną dzięki wywołaniu

(extractCohortIntersection() -> cohorts)

```
[1] "ACC"
                 "BLCA"
                             "BRCA"
                                          "CESC"
                                                      "CHOL"
                                                                  "COAD"
[7] "COADREAD"
                 "DLBC"
                             "ESCA"
                                          "GBM"
                                                      "GBMLGG"
                                                                  "HNSC"
[13] "KICH"
                 "KIPAN"
                             "KIRC"
                                          "KIRP"
                                                      "LAML"
                                                                  "LGG"
[19] "LIHC"
                             "LUSC"
                                          "עס"
                                                                  "PCPG"
                 "LUAD"
                                                      "PAAD"
[25] "PRAD"
                 "READ"
                             "SARC"
                                          "SKCM"
                                                      "STAD"
                                                                  "STES"
[31] "TGCT"
                             "UCEC"
                                          "UCS"
                                                      "UVM"
                 "THCA"
```

Następnie dla tak otrzymanych 35 kohort nowotworowych uzyskano dane o statusie pacjenta (śmierć bądź cenzurowanie) oraz jego czasie spędzonym pod obseracją dzięki funkcji

head(extractSurvival(cohorts) -> survivalData)

	bcr_patient_barcode	<pre>patient.vital_status</pre>	times
ACC.1	TCGA-OR-A5J1	1	1355
ACC.2	TCGA-OR-A5J2	1	1677
ACC.3	TCGA-OR-A5J3	0	1942
ACC.4	TCGA-OR-A5J4	1	423
ACC.5	TCGA-OR-A5J5	1	365
ACC.6	TCGA-OR-A5J6	0	2428

Dane o mutacjach występujących wśród tkanek nowotworowych kolejnych pacjentów uzyskano za pomocą

```
extractMutations(cohorts, 0.02) -> mutationsData
```

Using 'bcr_patient_barcode' as value column. Use 'value.var' to override

```
mutationsData[1:6, c(1,4,56,100,207,801)]
```

```
bcr_patient_barcode A2ML1 ALMS1 ATP2B2 CNTNAP4 PLEC
1
      TCGA-02-0003-01
                            0
                                   1
                                           0
                                                    0
                                                         0
2
      TCGA-02-0033-01
                            0
                                   0
                                           0
                                                    0
                                                         0
3
      TCGA-02-0047-01
                             0
                                   0
                                           0
                                                    0
                                                         0
4
      TCGA-02-0055-01
                            0
                                   0
                                           0
                                                    0
                                                         0
5
      TCGA-02-2470-01
                             0
                                   0
                                           0
                                                    0
                                                         0
6
                                                    0
      TCGA-02-2483-01
                             0
                                   0
                                           1
                                                         0
```

gdzie wybrano jedynie te geny, których mutacja dotyczyła co najmniej 2% pacjentów mających zarówno dane kliniczne jak i dane o występujących mutacjach w genach.

Dla tak otrzymanych dwóch zbiorów danych połączono dla pacjentów informacje kliniczne z informacjami o mutacjach dzięki przypisanym do pacjentów i ich próbek kodów bcr_patient_barcode, by ostatecznie podzielić zbiór pacjentów na 100 losowo utworzonych grup.

```
prepareCoxDataSplit(mutationsData,survivalData, groups = 100) -> coxData_split
head(coxData_split[[1]][c(1,10), c(210,302,356,898,911,1092:1093)])
```

```
COL14A1 DOCK9 FASN SEMA5A SHPRH patient.vital_status times 81 0 0 0 0 0 0 1 7 1068 1 0 0 0 1 0 1171
```

Niezbędną formułę modelu potrzebną do sprezycowania, które geny (a pozostało ich 1091) należy uwzględnić w modelu uzyskano dzięki pomocniczej funkcji

```
prepareForumlaSGD(coxData_split) -> formulaSGD
```

Ostatecznie dla 6085 pacjentów, którzy posiadali informacje o występujących mutacjach, oraz dla których odnotowano komplet i poprawność danych klinicznych dotyczących statusu i obserwowanego czasu przeżycia wyliczono współczynniki modelu proporcjonalnych hazardów Coxa z wykorzystaniem stochastycznego spadku gradientu do estymacji. Model dopasowano wielokrotnie z różnymi ciągami odpowiadającymi za długość kroku algorytmu, dodatkowo badano różną ilość epok w algorytmie. Dla tak powstałych kilku modeli wybrano ten, który dla swoich współczynników dawał największą wartość funkcji częściowej log-wiarogodności dla niewykorzystanej do uczenia próbki, zawierającej 2 ostatnie zaobserwowane podzbiory obserwacji.

Niemożliwe było sprawdzenie założeń modelu dotyczących proporcjonalności hazardu, gdyż zakładano napływającą postać danych (stąd podział danych na 100 grup). Dla takiej postaci pojawiania się danych ciężko także mówić o jakiejkolwiek diagnostyce poprawności dopasowania modelu i dokładności otrzymanych wpsółczynników. Nie stworzono teorii pozwalającej badać istotność statystyczną otrzymanych współczynników w modelu, jednak założono, że współczynniki dostatecznie odległe od 0 można uznać za istotnie wpływające na czas życia pacjenta. Współczynniki dodatnie oznaczają zwiększenie hazardu pacjenta posiadającego mutację w danym genie w stosunku do pacjentów nie posiadających mutacji w danym genie. Współczynniki ujemne oznaczają zmniejszenie hazardu pacjenta posiadającego mutację w danym genie w stosunku do pacjentów nie posiadających mutacji w danym genie. Wzrost proporcji hazardu można otrzymać dla danego genu poprzez obłożenie współczynnika funkcją wykładniczą o wykładniku e.

Wyniki estymacji dla genów zawierających największe co do modułu współczynniki można znaleźć w Tabeli 1.