

RiboTag Neural Development

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Libraries are loaded.

```
library(limma)
library(DESeq2)
library(hwriter)
library(RColorBrewer)
library(genefilter)
library(grid)
library(gridSVG)
source(file.path("R","hwriteSidebar.R"))
```

The dataset is loaded and genes with a count less than 1 over all samples are removed.

```
load(file.path("data","rawCounts.rda"))
load(file.path("data","expectedCounts.rda"))
load(file.path("data","Anno.rda"))

# pre-filter low count rows
Anno <- Anno[ rowSums(rawCounts) > 1, ]
expectedCounts <- expectedCounts[ rowSums(rawCounts) > 1, ]
rawCounts <- rawCounts[ rowSums(rawCounts) > 1, ]
Anno$transcript_type[is.na(Anno$transcript_type)] = "other"
```

The samples are annotated.

```

load("data/SampleList.rda")

SampleList$Population = factor(SampleList$Population,
                               levels=c("NSC", "NSC/ENB", "ENB", "LNB", "LNB/NEURON", "NEURON"))
SampleList$type = factor(SampleList$type,
                        levels=c("RIBOctr", "RNaseq", "RIBOseq"))
SampleList$Replicate = factor(SampleList$Replicate,
                             levels=c("RIBOctr", "RNaseq", "RIBOseq"))

```

1 Differential gene expression of Total RNA

Sample indices for five different pairs of total RNA datasets are extracted.

```

SamplesRNaseq = list(
  "ENB-NSC" = which(SampleList$Population %in% c("NSC", "ENB") &
                     SampleList$type == "RNaseq"),
  "LNB-ENB" = which(SampleList$Population %in% c("ENB", "LNB") &
                     SampleList$type == "RNaseq"),
  "NEURON-LNB" = which(SampleList$Population %in% c("LNB", "NEURON") &
                        SampleList$type == "RNaseq"))

DesignRNaseq = list(
  "ENB-NSC" = as.formula(~Population),
  "LNB-ENB" = as.formula(~Population),
  "NEURON-LNB" = as.formula(~Population))

for (n in names(DesignRNaseq)) {
  SamplesRNaseq[[n]] = SamplesRNaseq[[n]][
    order(SampleList[SamplesRNaseq[[n]], as.character(DesignRNaseq[[n]])[2]],
          SampleList[SamplesRNaseq[[n]], "Replicate"])]
}

```

Differential gene expression analysis is performed for each of these comparisons.

1.1 Differential gene expression in ENB-NSC

Create an output directory.

```

n = "ENB-NSC"
type="RNaseq"
Samples = SamplesRNaseq
filter = "none"
d = file.path("result", type, n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../", "...", "...", "mainmenu.html")

load(file.path("data", "ENSG2category.rda"))
source(file.path("R", "doGSEA.R"))

```

The count matrix is copied for convenience.

```

X = rawCounts[, Samples[[n]]]
Design = DesignRNaseq

```

A DESeq dataset is created that contains the respective samples.

```

dds <- DESeqDataSetFromMatrix(countData = X,
                               colData = SampleList[Samples[[n]],],
                               design = Design[[n]])

## converting counts to integer mode

## factor levels were dropped which had no samples

colData(dds)

## DataFrame with 6 rows and 6 columns
##           Name Population      Type Replicate Comment      OriginalName
##           <character> <factor> <factor> <factor> <character> <character>
## NSC_RNAseq_1 NSC_RNAseq_1     NSC   RNASEQ      NA          01_Ticry5547YFP
## NSC_RNAseq_2 NSC_RNAseq_2     NSC   RNASEQ      NA          14_Ticry8165_8166YFP
## NSC_RNAseq_3 NSC_RNAseq_3     NSC   RNASEQ      NA          02_Ticry6261YFP
## ENB_RNAseq_1 ENB_RNAseq_1    ENB   RNASEQ      NA          15_DicryS8356_8495YF
## ENB_RNAseq_2 ENB_RNAseq_2    ENB   RNASEQ      NA          08_DicryS6365YFP
## ENB_RNAseq_3 ENB_RNAseq_3    ENB   RNASEQ      NA          16_DicryS8355_8492YF

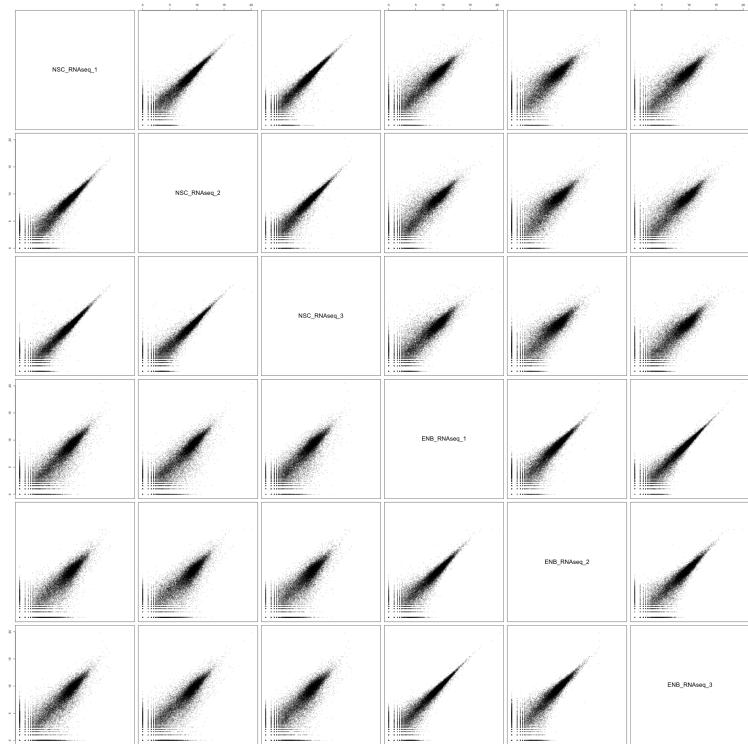
```

Log counts are computed and plotted as a pairs plot.

```

C = counts(dds)
C = log2(C+1)
lim = range(as.vector(C))
pairs(C,pch=20,cex=0.5,xlim=lim, ylim=lim,col="#00000022")

```



The standard DESeq2 workflow is used to for differential gene expression analysis. size factors are estimated using all genes. Subsequently, dispersion is estimated and p-values are computed by a Wald test.

```

dds = DESeq(dds)

## estimating size factors

## estimating dispersions

## gene-wise dispersion estimates

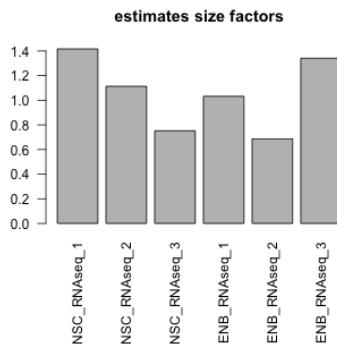
## mean-dispersion relationship

```

```
## final dispersion estimates
## fitting model and testing
res = results(dds)
```

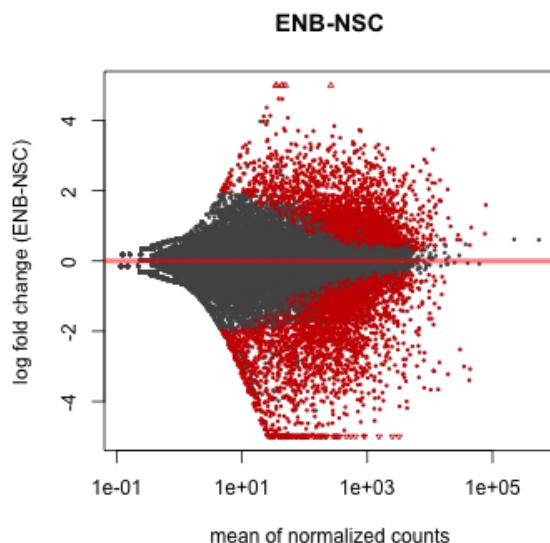
The size factors are plotted.

```
p = par(mar=c(10,4,4,2)+0.1)
barplot(sizeFactors(dds), main="estimates size factors", las=2)
par(p)
```



An MA plot shows in red the genes that are enriched by the RiboTag pull-down. For comparison, the second plot shows the genes that were used to estimate the size factors.

```
plotMA(res, ylim=c(-5,5), ylab=sprintf("log fold change (%s)", n), main=n)
```



```
load(file.path("data", "ENSGannotation.rda"))
res$symbol = ENSGannotation[row.names(res), "name"]
res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

res$significant = ""
res$significant[which(res$padj <= 0.1)] = "*"
res$significant[which(res$padj <= 0.01)] = "**"
```

The data is saved.

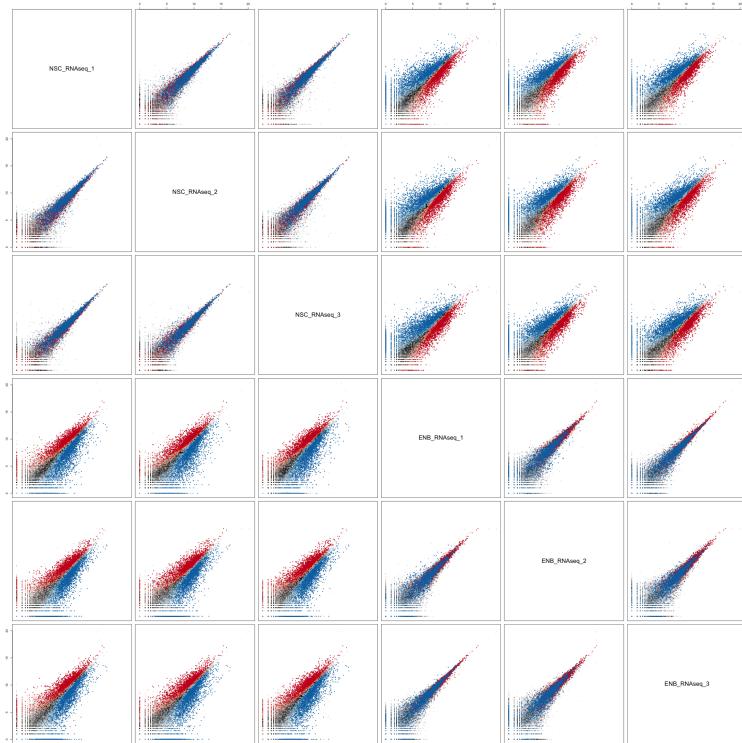
```
save(dds, file=file.path(d, "dds.rda"))
save(res, file=file.path(d, "res.rda"))
```

```

col=rep("#00000022", nrow(res))
col[which(res$padj <= 0.1 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$padj <= 0.1 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$padj <= 0.01 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$padj <= 0.01 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[4]

I = order(factor(col, levels=c("#00000022",brewer.pal(4,"RdBu")[c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[I,],pch=20,cex=0.5,xlim=lim, ylim=lim,col=col[I])

```



```

fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

```

```

write.table(res, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d,"resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Differential gene expression (%s;%s); result table",type,n),
      heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR",page=page,br=TRUE)
col = rep(NA, nrow(res))
col[which(res$significant != "" & res$log2FoldChange > 0)] = "#fbb4ae"
col[which(res$significant != "" & res$log2FoldChange < 0)] = "#b3cde3"
hwrite(as.data.frame(res), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)

```

1.2 Differential gene expression in LNB-ENB

Create an output directory.

```

n = "LNB-ENB"
type="RNAseq"
Samples = SamplesRNaseq
filter = "none"
d = file.path("result",type, n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../","..","..","mainmenu.html")

load(file.path("data","ENSG2category.rda"))
source(file.path("R","doGSEA.R"))

```

The count matrix is copied for convenience.

```

X = rawCounts[,Samples[[n]]]
Design = DesignRNaseq

```

A DESeq dataset is created that contains the respective samples.

```

dds <- DESeqDataSetFromMatrix(countData = X,
                               colData = SampleList[Samples[[n]],],
                               design = Design[[n]])

## converting counts to integer mode

## factor levels were dropped which had no samples

colData(dds)

## DataFrame with 6 rows and 6 columns
##           Name Population      Type Replicate Comment      OriginalName
##           <character>   <factor> <factor> <factor> <character>   <character>
## ENB_RNaseq_1 ENB_RNaseq_1     ENB  RNASEQ       NA 15_DicryS8356_8495YF
## ENB_RNaseq_2 ENB_RNaseq_2     ENB  RNASEQ       NA 08_DicryS6365YFP
## ENB_RNaseq_3 ENB_RNaseq_3     ENB  RNASEQ       NA 16_DicryS8355_8492YF
## LNB_RNaseq_1 LNB_RNaseq_1     LNB  RNASEQ       NA 10_Dicry06364YFP
## LNB_RNaseq_2 LNB_RNaseq_2     LNB  RNASEQ       NA 18_Dicry08355_8492YF
## LNB_RNaseq_3 LNB_RNaseq_3     LNB  RNASEQ       NA 11_Dicry06365YFP

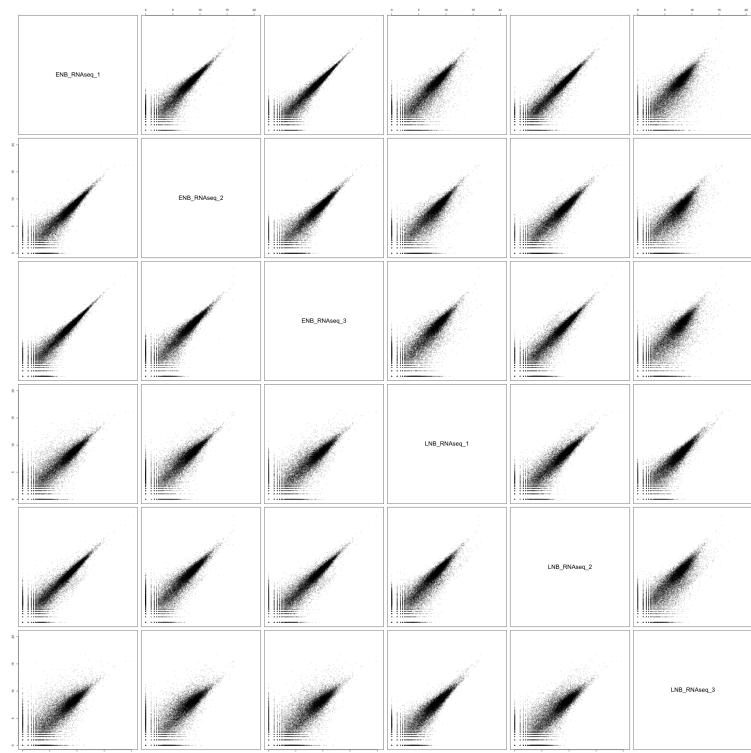
```

Log counts are computed and plotted as a pairs plot.

```

C = counts(dds)
C = log2(C+1)
lim = range(as.vector(C))
pairs(C,pch=20,cex=0.5,xlim=lim, ylim=lim,col="#00000022")

```

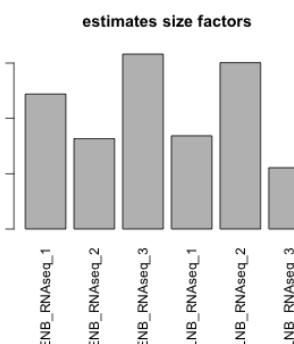


The standard DESeq2 workflow is used to for differential gene expression analysis. size factors are estimated using all genes. Subsequently, dispersion is estimated and p-values are computed by a Wald test.

```
dds = DESeq(dds)
## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
res = results(dds)
```

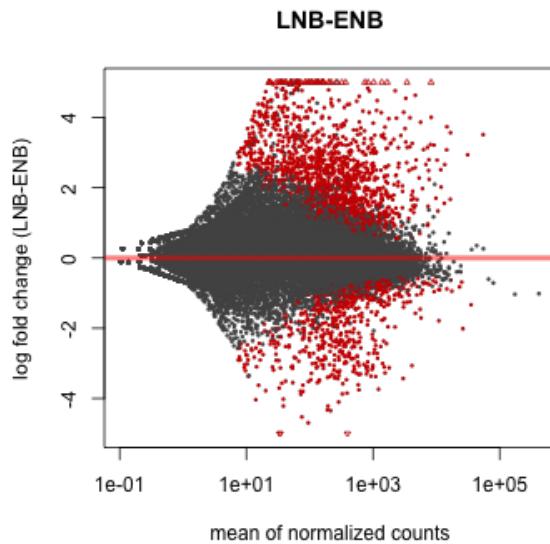
The size factors are plotted.

```
p = par(mar=c(10,4,4,2)+0.1)
barplot(sizeFactors(dds), main="estimates size factors", las=2)
par(p)
```



An MA plot shows in red the genes that are enriched by the RiboTag pull-down. For comparison, the second plot shows the genes that were used to estimate the size factors.

```
plotMA(res, ylim=c(-5,5), ylab=sprintf("log fold change (%s)",n), main=n)
```



```
load(file.path("data", "ENSGannotation.rda"))
res$symbol = ENSGannotation[row.names(res), "name"]
res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

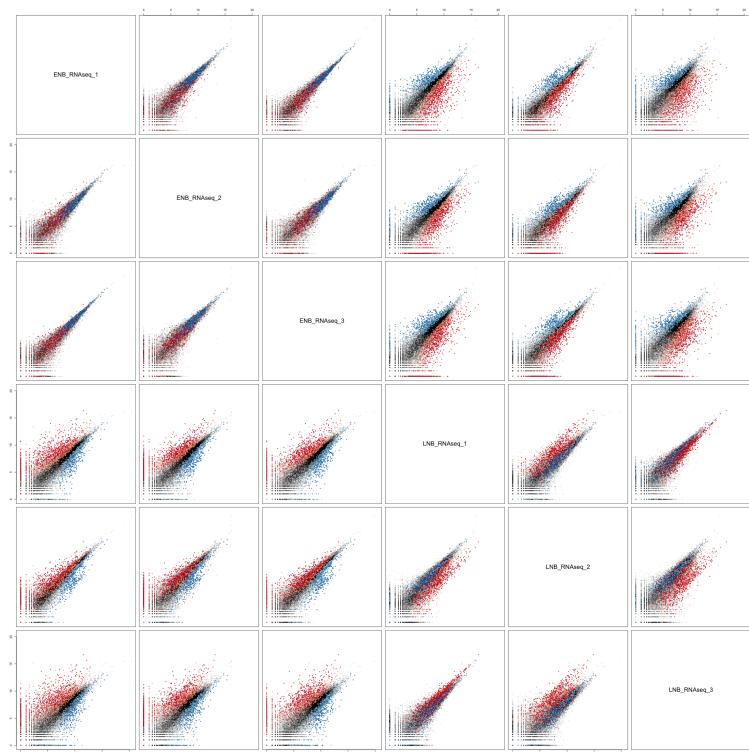
res$significant = ""
res$significant[which(res$padj <= 0.1)] = "*"
res$significant[which(res$padj <= 0.01)] = "**"
```

The data is saved.

```
save(dds, file=file.path(d, "dds.rda"))
save(res, file=file.path(d, "res.rda"))

col=rep("#00000022", nrow(res))
col[which(res$padj <= 0.1 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$padj <= 0.1 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$padj <= 0.01 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$padj <= 0.01 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[4]

I = order(factor(col, levels=c("#00000022", brewer.pal(4, "RdBu")[c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[I,], pch=20, cex=0.5, xlim=lim, ylim=lim, col=col[I])
```



```

fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

write.table(res, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d,"resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Differential gene expression (%s;%s); result table",type,n),
       heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR", page=page, br=TRUE)
col = rep(NA, nrow(res))
col[which(res$significant != "" & res$log2FoldChange > 0)] = "#fbb4ae"
col[which(res$significant != "" & res$log2FoldChange < 0)] = "#b3cde3"
hwrite(as.data.frame(res), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)

```

1.3 Differential gene expression in NEURON-LNB

Create an output directory.

```

n = "NEURON-LNB"
type="RNAseq"
Samples = SamplesRNaseq
filter = "none"
d = file.path("result",type, n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../","..","..","mainmenu.html")

load(file.path("data","ENSG2category.rda"))
source(file.path("R","doGSEA.R"))

```

The count matrix is copied for convenience.

```
X = rawCounts[,Samples[[n]]]
Design = DesignRNAseq
```

A DESeq dataset is created that contains the respective samples.

```
dds <- DESeqDataSetFromMatrix(countData = X,
                               colData = SampleList[Samples[[n]],],
                               design = Design[[n]])

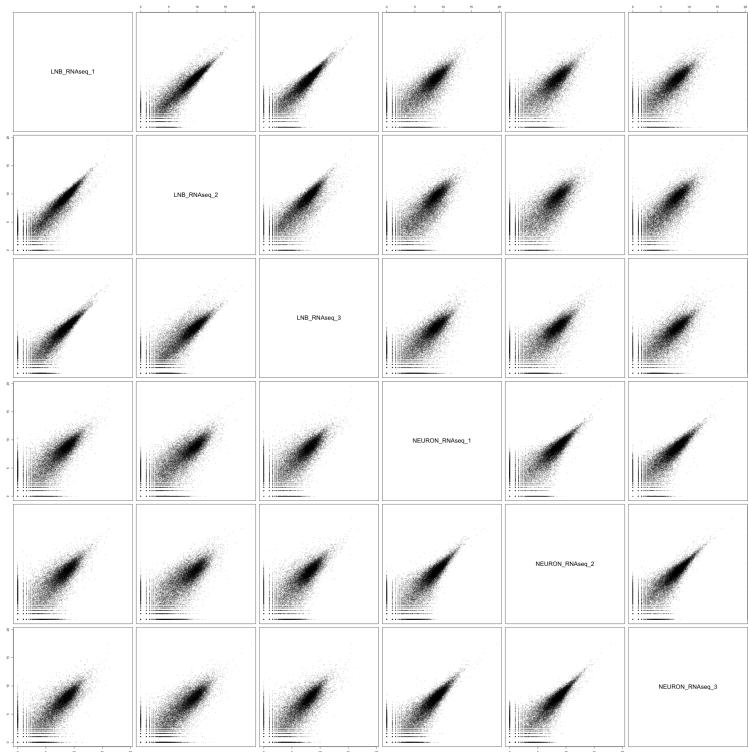
## converting counts to integer mode
## factor levels were dropped which had no samples

colData(dds)

## DataFrame with 6 rows and 6 columns
##                                     Name Population      Type Replicate   Comment
##                                     <character>    <factor> <factor> <factor> <character>
## LNB_RNAseq_1       LNB_RNAseq_1        LNB   RNAseq      NA
## LNB_RNAseq_2       LNB_RNAseq_2        LNB   RNAseq      NA
## LNB_RNAseq_3       LNB_RNAseq_3        LNB   RNAseq      NA
## NEURON_RNAseq_1   NEURON_RNAseq_1    NEURON RNAseq      NA
## NEURON_RNAseq_2   NEURON_RNAseq_2    NEURON RNAseq      NA
## NEURON_RNAseq_3   NEURON_RNAseq_3    NEURON RNAseq      NA
##                                     OriginalName
##                                     <character>
## LNB_RNAseq_1         10_Dicry06364YFP
## LNB_RNAseq_2         18_Dicry08355_8492YF
## LNB_RNAseq_3         11_Dicry06365YFP
## NEURON_RNAseq_1     09_Dxc4wpi_7490total
## NEURON_RNAseq_2     10_Dxc4wpi_7652total
## NEURON_RNAseq_3     11_Dxc4wpi_8047total
```

Log counts are computed and plotted as a pairs plot.

```
C = counts(dds)
C = log2(C+1)
lim = range(as.vector(C))
pairs(C,pch=20,cex=0.5,xlim=lim, ylim=lim,col="#00000022")
```

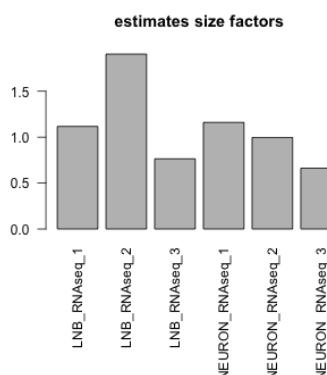


The standard DESeq2 workflow is used to for differential gene expression analysis. size factors are estimated using all genes. Subsequently, dispersion is estimated and p-values are computed by a Wald test.

```
dds = DESeq(dds)
## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
res = results(dds)
```

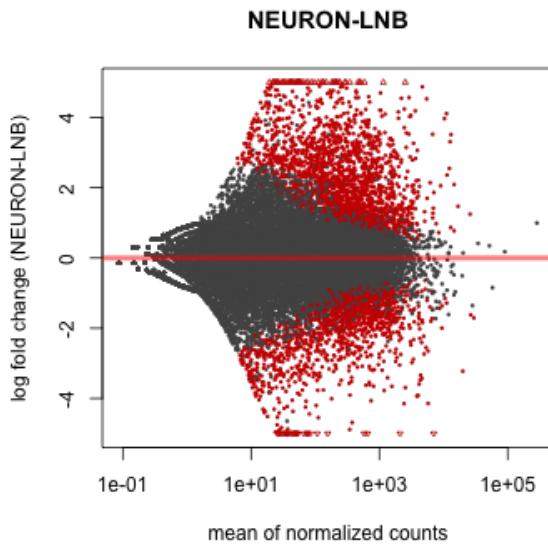
The size factors are plotted.

```
p = par(mar=c(10,4,4,2)+0.1)
barplot(sizeFactors(dds), main="estimates size factors", las=2)
par(p)
```



An MA plot shows in red the genes that are enriched by the RiboTag pull-down. For comparison, the second plot shows the genes that were used to estimate the size factors.

```
plotMA(res, ylim=c(-5,5), ylab=sprintf("log fold change (%s)",n), main=n)
```



```
load(file.path("data", "ENSGannotation.rda"))
res$symbol = ENSGannotation[row.names(res), "name"]
res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

res$significant = ""
res$significant[which(res$padj <= 0.1)] = "*"
res$significant[which(res$padj <= 0.01)] = "**"
```

The data is saved.

```
save(dds, file=file.path(d, "dds.rda"))
save(res, file=file.path(d, "res.rda"))

col=rep("#00000022", nrow(res))
col[which(res$padj <= 0.1 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$padj <= 0.1 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$padj <= 0.01 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$padj <= 0.01 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[4]

I = order(factor(col, levels=c("#00000022", brewer.pal(4, "RdBu")[c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[I,], pch=20, cex=0.5, xlim=lim, ylim=lim, col=col[I])
```



```

fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

write.table(res, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d,"resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Differential gene expression (%s;%s); result table",type,n),
       heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR", page=page, br=TRUE)
col = rep(NA, nrow(res))
col[which(res$significant != "" & res$log2FoldChange > 0)] = "#fbb4ae"
col[which(res$significant != "" & res$log2FoldChange < 0)] = "#b3cde3"
hwrite(as.data.frame(res), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)

```

2 Differential ribosome signal

Sample indices for five different pairs of ribosome tag datasets are extracted.

```

SamplesRIBOseq = list(
  "ENB-NSC" = which(SampleList$Population %in% c("NSC", "ENB") &
                     SampleList$type == "RIBOseq"),
  "LNB-ENB" = which(SampleList$Population %in% c("ENB", "LNB") &
                     SampleList$type == "RIBOseq"),
  "NEURON-LNB" = which(SampleList$Population %in% c("LNB", "NEURON") &
                        SampleList$type == "RIBOseq"))

DesignRIBOseq = list(
  "ENB-NSC" = as.formula(~Population),

```

```

"LNB-ENB" = as.formula(~Population),
"NEURON-LNB" = as.formula(~Population))

for (n in names(DesignRIBOseq)) {
  SamplesRIBOseq[[n]] = SamplesRIBOseq[[n]][
    order(SampleList[SamplesRIBOseq[[n]], as.character(DesignRIBOseq[[n]])[2]],
          SampleList[SamplesRIBOseq[[n]], "Replicate"])]
}

```

Differential gene expression analysis is performed for each of these comparisons.

2.1 Differential ribosome tag signal in ENB-NSC

Create an output directory.

```

n = "ENB-NSC"
type="RIBOseq"
Samples = SamplesRIBOseq
filter = "none"
d = file.path("result", type, n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../", "...", "...", "mainmenu.html")

load(file.path("data", "ENSG2category.rda"))
source(file.path("R", "doGSEA.R"))

```

The count matrix is copied for convenience.

```

Design = DesignRIBOseq
X = rawCounts[, Samples[[n]]]

```

A DESeq dataset is created that contains the respective samples.

```

dds <- DESeqDataSetFromMatrix(countData = X,
                               colData = SampleList[Samples[[n]], ],
                               design = Design[[n]])

## converting counts to integer mode
## factor levels were dropped which had no samples
colData(dds)

## DataFrame with 4 rows and 6 columns
##           Name Population      Type Replicate     Comment
##           <character>   <factor> <factor> <factor> <character>
## NSC_RIBOseq_1 NSC_RIBOseq_1      NSC   RIBOseq       NA
## NSC_RIBOseq_2 NSC_RIBOseq_2      NSC   RIBOseq       NA
## ENB_RIBOseq_1 ENB_RIBOseq_1      ENB   RIBOseq       NA
## ENB_RIBOseq_2 ENB_RIBOseq_2      ENB   RIBOseq       NA
##           OriginalName
##           <character>
## NSC_RIBOseq_1 10_SVZ_TICRY4d_4632_plus_
## NSC_RIBOseq_2 12_SVZ_TICRY4d_4943_plus_
## ENB_RIBOseq_1 6_SVZ_DICRY3dp_5516_plus_
## ENB_RIBOseq_2 9_SVZ_DICRY3dp_5519_plus_

```

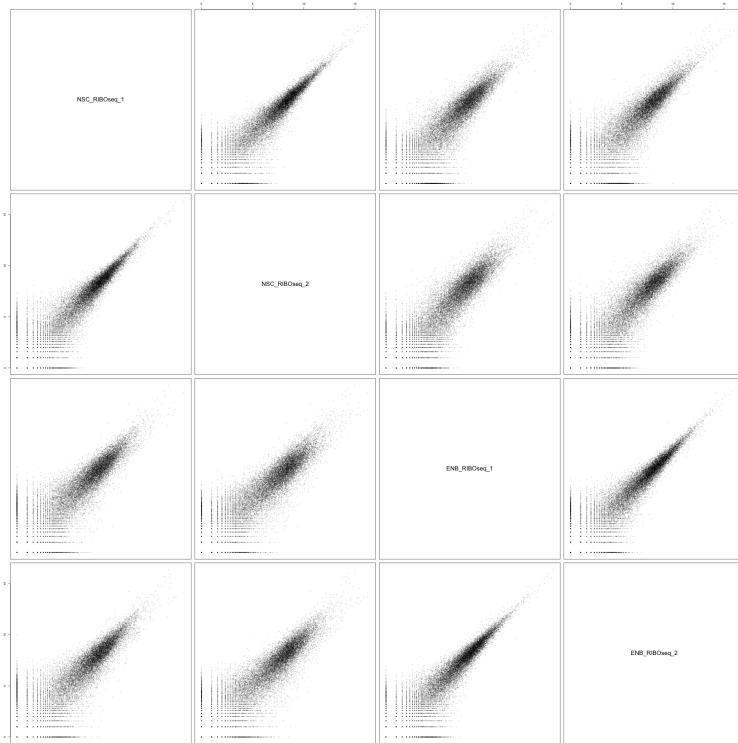
Log counts are computed and plotted as a pairs plot.

```

C = counts(dds)
C = log2(C+1)
lim = range(as.vector(C))

```

```
pairs(C,pch=20,cex=0.5,xlim=lim, ylim=lim,col="#00000022")
```

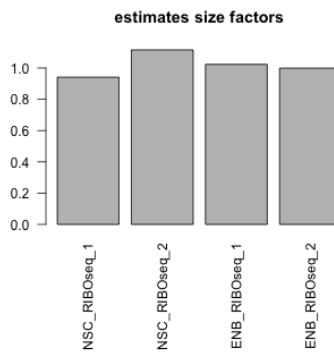


The standard DESeq2 workflow is used to for analysis differential ribosome tag signal. Size factors are estimated using all genes. Subsequently, dispersion is estimated and p-values are computed by a Wald test.

```
dds = DESeq(dds)
## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
res = results(dds)
```

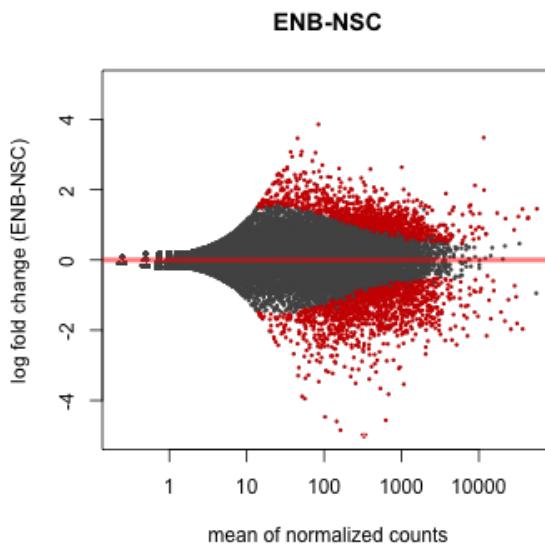
The size factors are plotted.

```
p = par(mar=c(10,4,4,2)+0.1)
barplot(sizeFactors(dds), main="estimates size factors", las=2)
par(p)
```



An MA plot shows in red the genes that are enriched by the RiboTag pull-down. For comparison, the second plot shows the genes that were used to estimate the size factors.

```
plotMA(res, ylim=c(-5,5), ylab=sprintf("log fold change (%s)",n), main=n)
```



```
load(file.path("data", "ENSGannotation.rda"))
res$symbol = ENSGannotation[row.names(res), "name"]
res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

res$significant = ""
res$significant[which(res$padj <= 0.1)] = "*"
res$significant[which(res$padj <= 0.01)] = "**"
```

The data is saved.

```
save(dds, file=file.path(d, "dds.rda"))
save(res, file=file.path(d, "res.rda"))
```

```
col=rep("#00000022", nrow(res))
col[which(res$padj <= 0.1 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$padj <= 0.1 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$padj <= 0.01 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$padj <= 0.01 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[4]

I = order(factor(col, levels=c("#00000022", brewer.pal(4, "RdBu")[c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[,], pch=20, cex=0.5, xlim=lim, ylim=lim, col=col[I])
```



```

fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

write.table(res, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d,"resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Differential ribosome signal (%s;%s); result table",type,n),
       heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR",page=page,br=TRUE)
col = rep(NA, nrow(res))
col[which(res$significant != "" & res$log2FoldChange > 0)] = "#fbb4ae"
col[which(res$significant != "" & res$log2FoldChange < 0)] = "#b3cde3"
hwrite(as.data.frame(res), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)

```

2.2 Differential ribosome tag signal in LNB-ENB

Create an output directory.

```

n = "LNB-ENB"
type="RIBOseq"
Samples = SamplesRIBOseq
filter = "none"
d = file.path("result",type, n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../","..","..","mainmenu.html")

load(file.path("data","ENSG2category.rda"))
source(file.path("R","doGSEA.R"))

```

The count matrix is copied for convenience.

```
Design = DesignRIBOseq
X = rawCounts[,Samples[[n]]]
```

A DESeq dataset is created that contains the respective samples.

```
dds <- DESeqDataSetFromMatrix(countData = X,
                               colData = SampleList[Samples[[n]],],
                               design = Design[[n]])

## converting counts to integer mode
## factor levels were dropped which had no samples

colData(dds)

## DataFrame with 5 rows and 6 columns
##           Name Population      Type Replicate Comment
##           <character>   <factor> <factor> <factor> <character>
## ENB_RIBOseq_1 ENB_RIBOseq_1       ENB RIBOseq      NA
## ENB_RIBOseq_2 ENB_RIBOseq_2       ENB RIBOseq      NA
## LNB_RIBOseq_1 LNB_RIBOseq_1       LNB RIBOseq      NA
## LNB_RIBOseq_2 LNB_RIBOseq_2       LNB RIBOseq      NA
## LNB_RIBOseq_3 LNB_RIBOseq_3       LNB RIBOseq      NA
##           OriginalName
##           <character>
## ENB_RIBOseq_1 6_SVZ_DICRY3dp_5516_plus_
## ENB_RIBOseq_2 9_SVZ_DICRY3dp_5519_plus_
## LNB_RIBOseq_1 1_0B_DICRY3dpi_5516_plus_
## LNB_RIBOseq_2 4_0B_DICRY3dpi_5519_plus_
## LNB_RIBOseq_3 5_0B_DICRY3dpi_5524_plus_
```

Log counts are computed and plotted as a pairs plot.

```
C = counts(dds)
C = log2(C+1)
lim = range(as.vector(C))
pairs(C,pch=20,cex=0.5,xlim=lim, ylim=lim,col="#00000022")
```

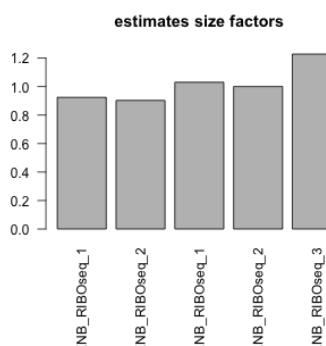


The standard DESeq2 workflow is used to for analysis differential ribosome tag signal. Size factors are estimated using all genes. Subsequently, dispersion is estimated and p-values are computed by a Wald test.

```
dds = DESeq(dds)
## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
res = results(dds)
```

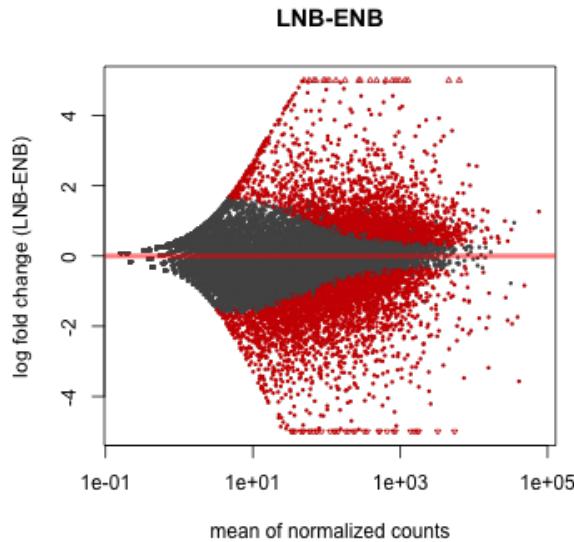
The size factors are plotted.

```
p = par(mar=c(10,4,4,2)+0.1)
barplot(sizeFactors(dds), main="estimates size factors", las=2)
par(p)
```



An MA plot shows in red the genes that are enriched by the RiboTag pull-down. For comparison, the second plot shows the genes that were used to estimate the size factors.

```
plotMA(res, ylim=c(-5,5), ylab=sprintf("log fold change (%s)",n), main=n)
```



```
load(file.path("data", "ENSGannotation.rda"))
res$symbol = ENSGannotation[row.names(res), "name"]
res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

res$significant = ""
res$significant[which(res$padj <= 0.1)] = "*"
res$significant[which(res$padj <= 0.01)] = "**"
```

The data is saved.

```
save(dds, file=file.path(d, "dds.rda"))
save(res, file=file.path(d, "res.rda"))

col=rep("#00000022", nrow(res))
col[which(res$padj <= 0.1 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$padj <= 0.1 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$padj <= 0.01 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$padj <= 0.01 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[4]

I = order(factor(col, levels=c("#00000022", brewer.pal(4, "RdBu")[c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[I,], pch=20, cex=0.5, xlim=lim, ylim=lim, col=col[I])
```



```

fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

write.table(res, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d,"resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Differential ribosome signal (%s;%s); result table",type,n),
       heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR", page=page, br=TRUE)
col = rep(NA, nrow(res))
col[which(res$significant != "" & res$log2FoldChange > 0)] = "#fbb4ae"
col[which(res$significant != "" & res$log2FoldChange < 0)] = "#b3cde3"
hwrite(as.data.frame(res), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)

```

2.3 Differential ribosome tag signal in NEURON-LNB

Create an output directory.

```

n = "NEURON-LNB"
type="RIBOseq"
Samples = SamplesRIBOseq
filter = "none"
d = file.path("result",type, n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../","..","..","mainmenu.html")

load(file.path("data","ENSG2category.rda"))
source(file.path("R","doGSEA.R"))

```

The count matrix is copied for convenience.

```
Design = DesignRIBOseq
X = rawCounts[,Samples[[n]]]
```

A DESeq dataset is created that contains the respective samples.

```
dds <- DESeqDataSetFromMatrix(countData = X,
                               colData = SampleList[Samples[[n]],],
                               design = Design[[n]])

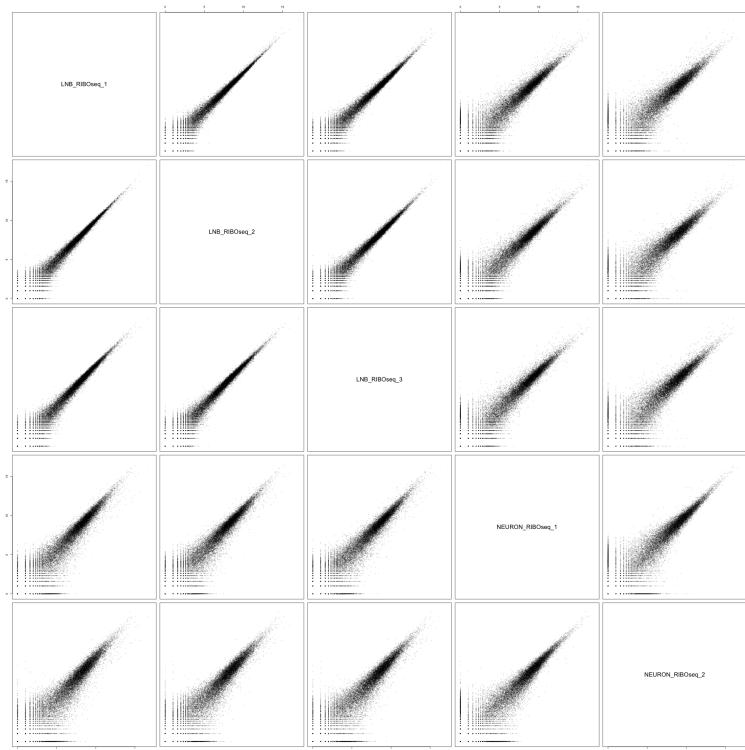
## converting counts to integer mode
## factor levels were dropped which had no samples

colData(dds)

## DataFrame with 5 rows and 6 columns
##           Name Population      Type Replicate Comment
##           <character>   <factor> <factor>  <factor>  <character>
## LNB_RIBOseq_1    LNB_RIBOseq_1     LNB  RIBOseq     NA
## LNB_RIBOseq_2    LNB_RIBOseq_2     LNB  RIBOseq     NA
## LNB_RIBOseq_3    LNB_RIBOseq_3     LNB  RIBOseq     NA
## NEURON_RIBOseq_1 NEURON_RIBOseq_1 NEURON RIBOseq     NA
## NEURON_RIBOseq_2 NEURON_RIBOseq_2 NEURON RIBOseq     NA higher 3 bias
##           OriginalName
##           <character>
## LNB_RIBOseq_1    1_0B_DICRY3dpi_5516_plus_
## LNB_RIBOseq_2    4_0B_DICRY3dpi_5519_plus_
## LNB_RIBOseq_3    5_0B_DICRY3dpi_5524_plus_
## NEURON_RIBOseq_1 05_Dxc4wpi_7155_plus_Ribo
## NEURON_RIBOseq_2 07_Dxc4wpi_7154_plus_Ribo
```

Log counts are computed and plotted as a pairs plot.

```
C = counts(dds)
C = log2(C+1)
lim = range(as.vector(C))
pairs(C,pch=20,cex=0.5,xlim=lim, ylim=lim,col="#00000022")
```

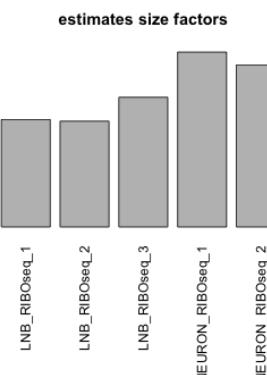


The standard DESeq2 workflow is used to for analysis differential ribosome tag signal. Size factors are estimated using all genes. Subsequently, dispersion is estimated and p-values are computed by a Wald test.

```
dds = DESeq(dds)
## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
res = results(dds)
```

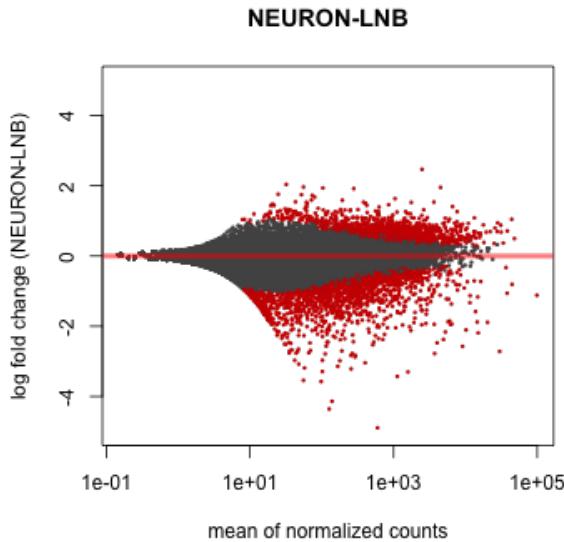
The size factors are plotted.

```
p = par(mar=c(10,4,4,2)+0.1)
barplot(sizeFactors(dds), main="estimates size factors", las=2)
par(p)
```



An MA plot shows in red the genes that are enriched by the RiboTag pull-down. For comparison, the second plot shows the genes that were used to estimate the size factors.

```
plotMA(res, ylim=c(-5,5), ylab=sprintf("log fold change (%s)",n), main=n)
```



```
load(file.path("data", "ENSGannotation.rda"))
res$symbol = ENSGannotation[row.names(res), "name"]
res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

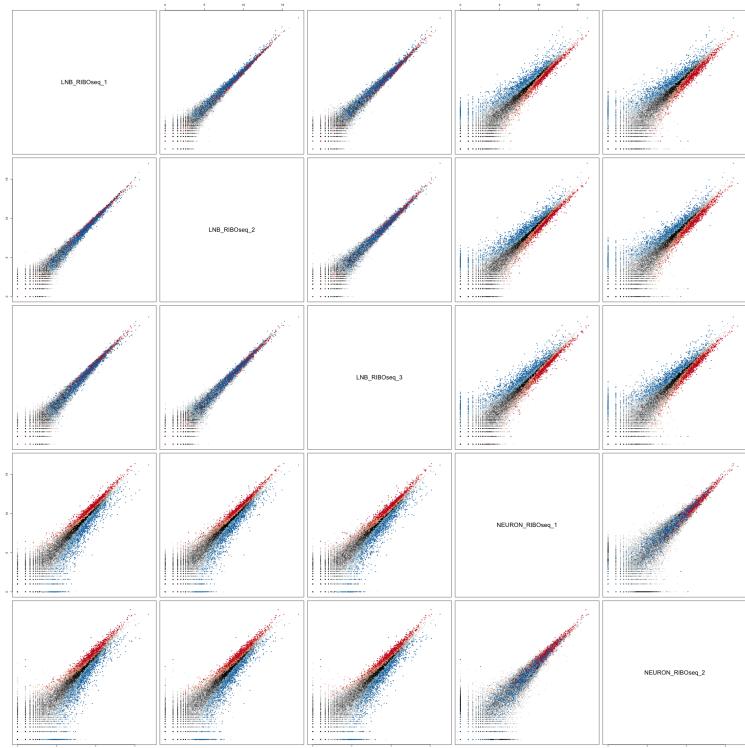
res$significant = ""
res$significant[which(res$padj <= 0.1)] = "*"
res$significant[which(res$padj <= 0.01)] = "**"
```

The data is saved.

```
save(dds, file=file.path(d, "dds.rda"))
save(res, file=file.path(d, "res.rda"))

col=rep("#00000022", nrow(res))
col[which(res$padj <= 0.1 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$padj <= 0.1 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$padj <= 0.01 & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$padj <= 0.01 & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[4]

I = order(factor(col, levels=c("#00000022", brewer.pal(4, "RdBu")[c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[I,], pch=20, cex=0.5, xlim=lim, ylim=lim, col=col[I])
```



```

fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

write.table(res, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d,"resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Differential ribosome signal (%s;%s); result table",type,n),
       heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR",page=page,br=TRUE)
col = rep(NA, nrow(res))
col[which(res$significant != "" & res$log2FoldChange > 0)] = "#fbb4ae"
col[which(res$significant != "" & res$log2FoldChange < 0)] = "#b3cde3"
hwrite(as.data.frame(res), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)

```

3 Translation Efficiency

```

SamplesTranslationEfficiency = list(
  "NSC" = which(SampleList$Population %in% c("NSC", "NSC/ENB")),
  "ENB" = which(SampleList$Population %in% c("ENB", "NSC/ENB")),
  "LNB" = which(SampleList$Population %in% c("LNB", "LNB/NEURON")),
  "NEURON" = which(SampleList$Population %in% c("NEURON", "LNB/NEURON")) )

DesignTranslationEfficiency = as.formula(~Type)

for (n in names(SamplesTranslationEfficiency)) {
  SamplesTranslationEfficiency[[n]] = SamplesTranslationEfficiency[[n]][
    order(SampleList[SamplesTranslationEfficiency[[n]], "Type"],
          SampleList[SamplesTranslationEfficiency[[n]], "Replicate"])]]
}

```

```
}
```

Translation efficiency is estimated for high abundant genes.

3.1 Filter unspecific binders in NSC

Create an output directory.

```
n = "NSC"
d = file.path("result", "TranslationEfficiency", n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../..", "mainmenu.html")
source(file.path("R", "doGSEA.R"))
```

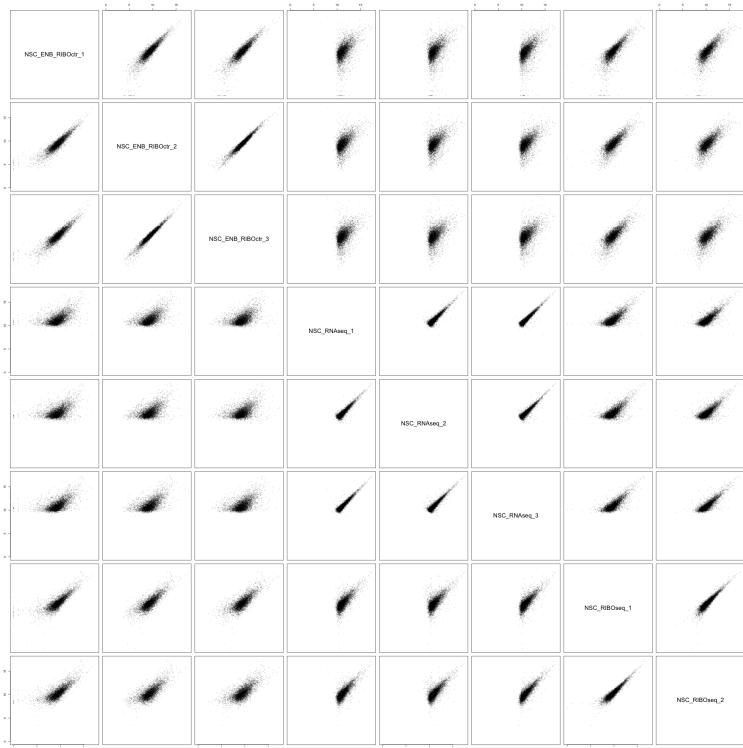
A DESeq dataset is created that contains the respective samples.

```
Design = DesignTranslationEfficiency
EC_all <- expectedCounts[, SamplesTranslationEfficiency[[n]]]
SL <- SampleList [SamplesTranslationEfficiency[[n]], ]
# dds_all <- DESeqDataSetFromMatrix(
#   countData = expectedCounts[, SamplesTranslationEfficiency[[n]]],
#   colData = SampleList [SamplesTranslationEfficiency[[n]], ],
#   design = ~ Type)
# mcols(dds_all) = Anno
# colData(dds_all)
```

Log counts are computed and plotted as a pairs plot.

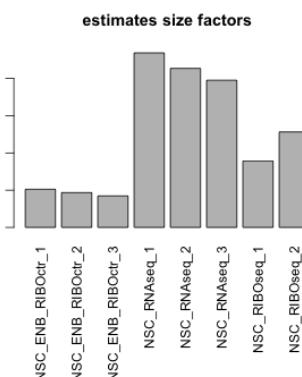
```
I = which(rowMeans(log2(EC_all[,
  SL$Type == "RNaseq"]+1)) >= 10 &
  Anno$transcript_type == "protein_coding")
EC = EC_all[I,]
# I = which(rowMeans(log2(counts(dds_all)[,
#   colData(dds_all)$Type == "RNaseq"]+1)) >= 10 &
#   mcols(dds_all)$transcript_type == "protein_coding")
# dds = dds_all[I,]

C = log2(EC+1)
Call = log2(EC_all+1)
# C = log2(counts(dds)+1)
# Call = log2(counts(dds_all)+1)
lim = range(as.vector(C))
pairs(C, pch=20, cex=0.5, col="#00000022", xlim=lim, ylim=lim)
```



The genes in the cluster with lower read counts in the control sample are used to estimate the size factors.

```
s = sizeFactors(estimateSizeFactors(DESeqDataSetFromMatrix(
  countData = round(EC), colData = SL, design = ~ Type),
  controlGenes=which(rowMeans(log2(EC[,SL$Type == "RNAseq"])+1)) >= 13)))
## converting counts to integer mode
#sizeFactors(dds) = s
p = par(mar=c(10,4,4,2)+0.1)
barplot(s, main="estimates size factors", las=2)
par(p)
```



The count data is normalized and unspecific binders are identified. Therefore, RiboControl samples are compared to total RNA samples. The genes that are significantly higher in RiboControl samples are potential unspecific binders of the tag. These genes are filtered as well.

```
D = C
m = log2(s)
for (i in 1:ncol(EC)) {
  D[,i] = D[,i] - m[i]
}
```

```

Dall = Call
for (i in 1:ncol(EC)) {
  Dall[,i] = Dall[,i] - m[i]
}

IR = which(SL$type == "RIBOseq")
IT = which(SL$type == "RNAseq")
IC = which(SL$type == "RIBOctr")

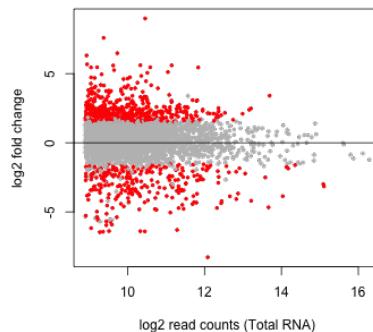
fitc = eBayes(lmFit(cbind(D[,IC[1]]-D[,IT[1]],D[,IC[2]]-D[,IT[2]])))
padjc = p.adjust(fitc$p.value[,1],method="BH")
lfcc = fitc$coefficients[,1]

plot(rowMeans(D[,SL$type == "RNAseq"]),lfcc,
  pch=20,cex=0.7,col=ifelse(padjc < 0.01, "red","gray"),
  xlab="log2 read counts (Total RNA)", ylab="log2 fold change")
abline(h=0)

UnspecBinders = which(padjc <= 0.01 & lfcc > 0)

fitc$padj = padjc

```



To estimate the translation efficiency, the RiboTag data is modeled as a function of RiboControl and total RNA.

```

fit1 = list()
for (i in 1:2) {
  D2 = data.frame("Ribo" = D[,IR[i]], "Total" = D[,IT[i]], "ctrl" = D[,IC[i]])
  fit1[[i]] = lm(Ribo ~ Total+ctrl, data=D2)
  fit1[[i]]$anova = anova(fit1[[i]])
  fit1[[i]]$expVar = fit1[[i]]$anova[, "Sum Sq", drop=FALSE]
  fit1[[i]]$expVar = fit1[[i]]$expVar/sum(fit1[[i]]$expVar)
  D2all = data.frame("Ribo" = Dall[,IR[i]], "Total" = Dall[,IT[i]], "ctrl" = Dall[,IC[i]], row.names =
  p = predict(fit1[[i]], newdata = D2all)
  fit1[[i]]$residualsAll = D2all$Ribo - p
}

R1 = fit1[[1]]$residuals
R2 = fit1[[2]]$residuals
R1all = fit1[[1]]$residualsAll
R2all = fit1[[2]]$residualsAll
R1all = R1all - shorth(R1,na.rm=TRUE)
R2all = R2all - shorth(R2,na.rm=TRUE)
R1 = R1 - shorth(R1,na.rm=TRUE)
R2 = R2 - shorth(R2,na.rm=TRUE)

```

```

D2 = data.frame("R1" = R1, "R2" = R2)
fit2 = lm(data=D2)
fit2$anova = anova(fit2)
fit2$expVar = fit2$anova[, "Sum Sq", drop=FALSE]
fit2$expVar = fit2$expVar/sum(fit2$expVar)

fita = eBayes(lmFit(cbind(R1,R2)))
padja = p.adjust(fita$p.value[,1], method="BH")
lfca = fita$coefficients[,1]

ExpVar = t(as.matrix(cbind(rep1=fita[[1]]$expVar, rep2=fita[[2]]$expVar)))
ExpVar = cbind(ExpVar[,1:2], ExpVar[,3] * fit2$expVar[1,1], ExpVar[,3] * fit2$expVar[2,1])
colnames(ExpVar) = c("total RNA", "background", "transl. eff.", "residuals")
row.names(ExpVar) = paste0("rep ", 1:2)
# barplot(ExpVar, beside=TRUE,
#         legend=TRUE, ylab="explained variance")
# barplot(t(ExpVar), beside=FALSE,
#         legend=TRUE, ylab="explained variance", xlim=c(0,4))

res = as.data.frame(Anno[row.names(EC),])
#res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

res$pval = fita$p.value[,1]
res$padj = padja
res$log2FoldChange = lfca
res$significant = ""
res$significant[which(padja <= 0.1)] = "*"
res$significant[which(padja <= 0.01)] = "**"
attr(res, "log2FC") = rowMeans(cbind(R1all, R2all))
attr(res, "log2total") = rowMeans(Dall[, IT])
attr(res, "log2FCsd") = rowSds(cbind(R1all, R2all))
attr(res, "log2TotalSD") = rowSds(Dall[, IT])

```

The R/Bioconductor package limma shrinks the standard deviation of each gene towards a global estimate of standard deviation along all genes. In the case of large outliers, this can result in spurious hits. To avoid these, genes with standard deviation larger than the absolute value of the log-fold change are not regarded as significant.

```

K = which(res$significant != "" & (fita$sigma > abs(fita$coefficients[,1])))

write.table(res[K,], sep="\t", file = file.path(d, "filteredGenes.txt"), quote=FALSE)

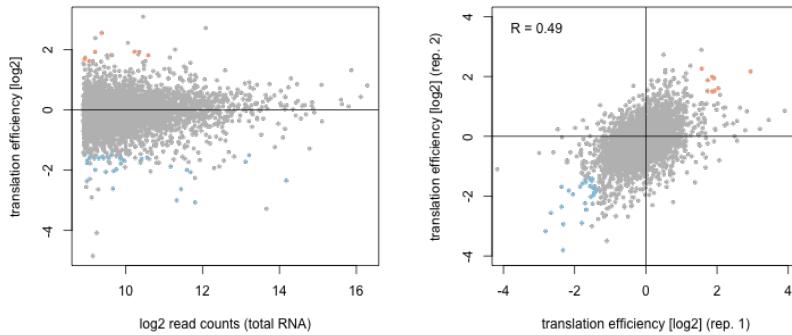
res$padj[K] = NA
res$significant[K] = ""

col=rep("gray", length(padja))
col[which(res$significant == "*" & lfca > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$significant == "*" & lfca < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$significant == "**" & lfca > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$significant == "**" & lfca < 0)] = brewer.pal(4, "RdBu")[4]
plot(rowMeans(D[,SL$type == "RNaseq"]), lfca,
      pch=20, cex=0.7, col=col,
      xlab="log2 read counts (total RNA)", ylab="translation efficiency [log2]")
abline(h=0)

plot(R1, R2,
      pch=20, cex=0.7, col=col,
      xlab="translation efficiency [log2] (rep. 1)",
      ylab="translation efficiency [log2] (rep. 2)",
      xlim=c(-4,4), ylim=c(-4,4))

```

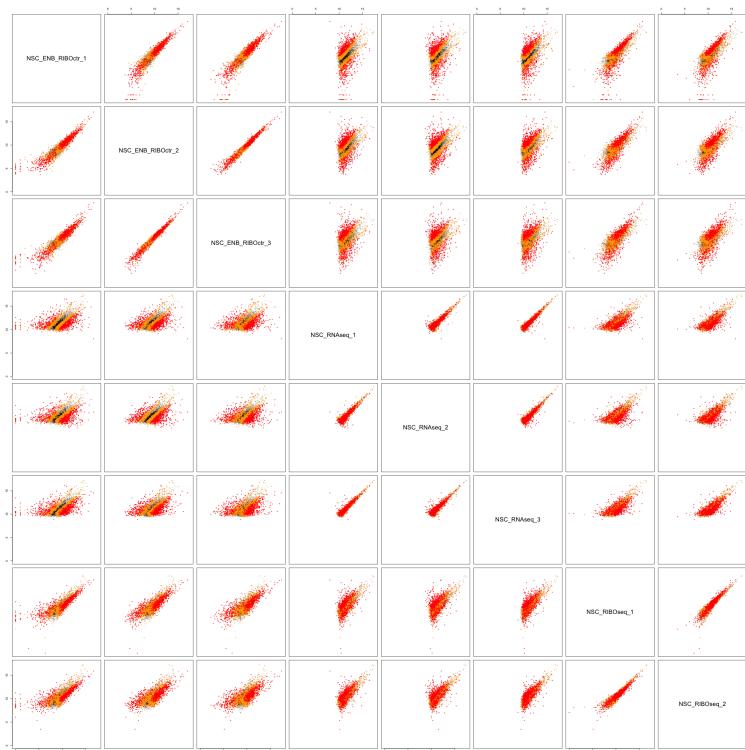
```
abline(h=0,v=0)
text(-3.8,3.8,paste0("R = ",format(cor(R1,R2,use="pairwise.complete"),digits=2)),adj = c(0,1))
```



The data is saved.

```
save(EC, file=file.path(d, "EC.rda"))
save(res, file=file.path(d, "res.rda"))
save(fitc, file=file.path(d, "fitc.rda"))
save(ExpVar, file=file.path(d, "ExpVar.rda"))

col = rep("#00000022", dim(C)[1])
col[which(fitc$padj <= 0.1)] = "orange"
col[which(fitc$padj <= 0.01)] = "red"
I = order(factor(col, levels=c("#00000022","orange","red")))
lim = range(as.vector(C))
pairs(C[I,],pch=20,cex=0.5,xlim=lim, ylim=lim,col=col[I])
```



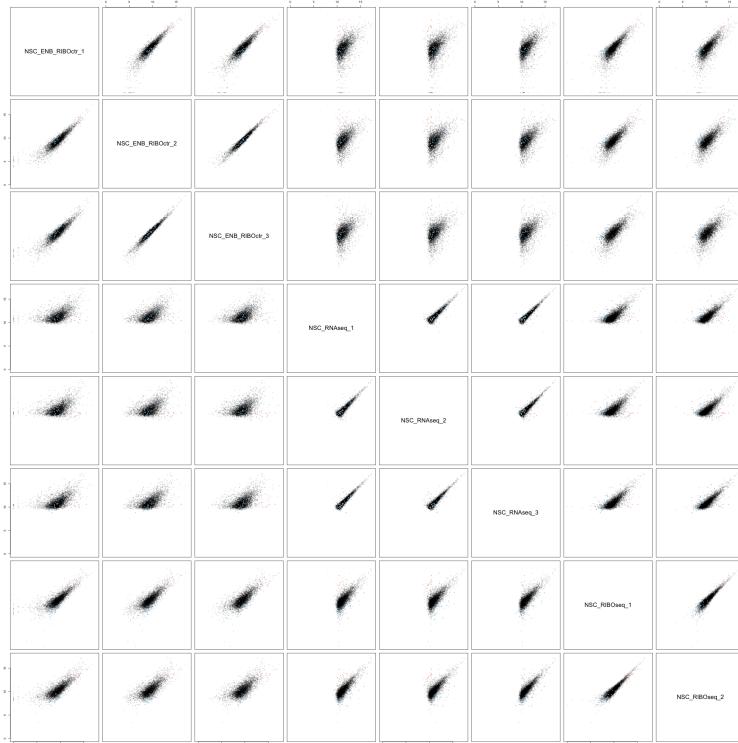
```
col=rep("#00000022", nrow(res))
col[which(res$significant == "*" & res$log2FoldChange > 0)] = brewer.pal(4,"RdBu")[2]
col[which(res$significant == "*" & res$log2FoldChange < 0)] = brewer.pal(4,"RdBu")[3]
col[which(res$significant == "***" & res$log2FoldChange > 0)] = brewer.pal(4,"RdBu")[1]
```

```

col[which(res$significant == "***" & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu") [4]

I = order(factor(col, levels=c("#00000022",brewer.pal(4,"RdBu")[c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[I,],pch=20,cex=0.5,xlim=lim, ylim=lim,col=col[I])

```



```

fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

```

```

res2 = res[order(factor(res$significant, levels=c("***", "**", "*")),
                 sign(-res$log2FoldChange),res$symbol),]
res2$UnspecBinders = ""
res2$UnspecBinders[row.names(res2) %in% names(UnspecBinders)] = "UnspecBinder"
write.table(res2, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d,"resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Translation efficiency (%s); result table",n),
       heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR",page=page,br=TRUE)
col = rep("",nrow(res))
col[which(res2$significant %in% c("***", "**") & res2$log2FoldChange > 0)] =
  brewer.pal(3, "Pastel1")[1]
col[which(res2$significant %in% c("***", "**") & res2$log2FoldChange < 0)] =
  brewer.pal(3, "Pastel1")[2]
hwrite(as.data.frame(res2), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)

```

3.2 Filter unspecific binders in ENB

Create an output directory.

```
n = "ENB"
d = file.path("result", "TranslationEfficiency", n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../..", "mainmenu.html")
source(file.path("R", "doGSEA.R"))
```

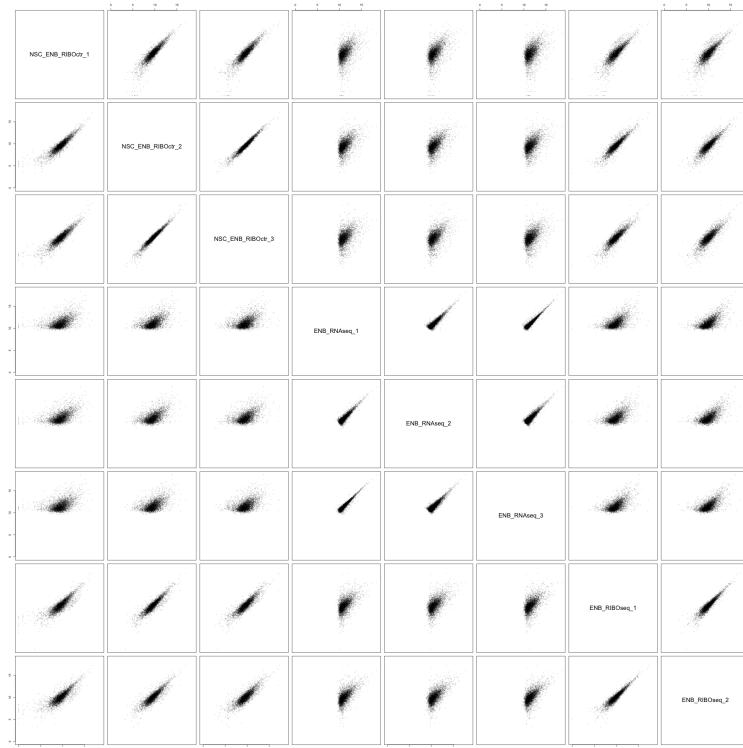
A DESeq dataset is created that contains the respective samples.

```
Design = DesignTranslationEfficiency
EC_all <- expectedCounts[, SamplesTranslationEfficiency[[n]]]
SL <- SampleList[SamplesTranslationEfficiency[[n]], ]
# dds_all <- DESeqDataSetFromMatrix(
#   countData = expectedCounts[, SamplesTranslationEfficiency[[n]]],
#   colData = SampleList[SamplesTranslationEfficiency[[n]], ],
#   design = ~ Type)
# mcols(dds_all) = Anno
# colData(dds_all)
```

Log counts are computed and plotted as a pairs plot.

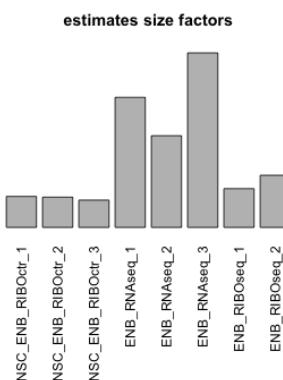
```
I = which(rowMeans(log2(EC_all[, SL$Type == "RNaseq"])+1)) >= 10 &
      Anno$transcript_type == "protein_coding")
EC = EC_all[I, ]
# I = which(rowMeans(log2(counts(dds_all)[, colData(dds_all)$Type == "RNaseq"])+1)) >= 10 &
#       mcols(dds_all)$transcript_type == "protein_coding")
# dds = dds_all[I, ]

C = log2(EC+1)
Call = log2(EC_all+1)
# C = log2(counts(dds)+1)
# Call = log2(counts(dds_all)+1)
lim = range(as.vector(C))
pairs(C, pch=20, cex=0.5, col="#00000022", xlim=lim, ylim=lim)
```



The genes in the cluster with lower read counts in the control sample are used to estimate the size factors.

```
s = sizeFactors(estimateSizeFactors(DESeqDataSetFromMatrix(
  countData = round(EC), colData = SL, design = ~ Type),
  controlGenes=which(rowMeans(log2(EC[,SL$Type == "RNAseq"])+1)) >= 13)))
## converting counts to integer mode
#sizeFactors(dds) = s
p = par(mar=c(10,4,4,2)+0.1)
barplot(s, main="estimates size factors", las=2)
par(p)
```



The count data is normalized and unspecific binders are identified. Therefore, RiboControl samples are compared to total RNA samples. The genes that are significantly higher in RiboControl samples are potential unspecific binders of the tag. These genes are filtered as well.

```
D = C
m = log2(s)
for (i in 1:ncol(EC)) {
  D[,i] = D[,i] - m[i]
}
```

```

Dall = Call
for (i in 1:ncol(EC)) {
  Dall[,i] = Dall[,i] - m[i]
}

IR = which(SL$Type == "RIBOseq")
IT = which(SL$Type == "RNAseq")
IC = which(SL$Type == "RIBOctr")

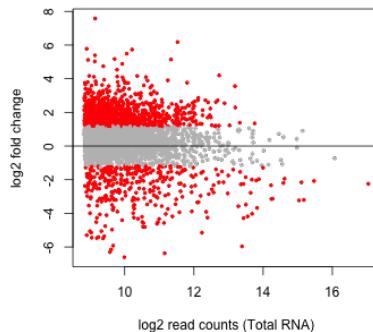
fitc = eBayes(lmFit(cbind(D[,IC[1]]-D[,IT[1]],D[,IC[2]]-D[,IT[2]])))
padjc = p.adjust(fitc$p.value[,1],method="BH")
lfcc = fitc$coefficients[,1]

plot(rowMeans(D[,SL$Type == "RNAseq"]),lfcc,
  pch=20,cex=0.7,col=ifelse(padjc < 0.01, "red","gray"),
  xlab="log2 read counts (Total RNA)", ylab="log2 fold change")
abline(h=0)

UnspecBinders = which(padjc <= 0.01 & lfcc > 0)

fitc$padj = padjc

```



To estimate the translation efficiency, the RiboTag data is modeled as a function of RiboControl and total RNA.

```

fit1 = list()
for (i in 1:2) {
  D2 = data.frame("Ribo" = D[,IR[i]], "Total" = D[,IT[i]], "ctrl" = D[,IC[i]])
  fit1[[i]] = lm(Ribo ~ Total+ctrl, data=D2)
  fit1[[i]]$anova = anova(fit1[[i]])
  fit1[[i]]$expVar = fit1[[i]]$anova[, "Sum Sq", drop=FALSE]
  fit1[[i]]$expVar = fit1[[i]]$expVar/sum(fit1[[i]]$expVar)
  D2all = data.frame("Ribo" = Dall[,IR[i]], "Total" = Dall[,IT[i]], "ctrl" = Dall[,IC[i]], row.names =
  p = predict(fit1[[i]], newdata = D2all)
  fit1[[i]]$residualsAll = D2all$Ribo - p
}

R1 = fit1[[1]]$residuals
R2 = fit1[[2]]$residuals
R1all = fit1[[1]]$residualsAll
R2all = fit1[[2]]$residualsAll
R1all = R1all - shorth(R1,na.rm=TRUE)
R2all = R2all - shorth(R2,na.rm=TRUE)
R1 = R1 - shorth(R1,na.rm=TRUE)
R2 = R2 - shorth(R2,na.rm=TRUE)

```

```

D2 = data.frame("R1" = R1, "R2" = R2)
fit2 = lm(data=D2)
fit2$anova = anova(fit2)
fit2$expVar = fit2$anova[, "Sum Sq", drop=FALSE]
fit2$expVar = fit2$expVar/sum(fit2$expVar)

fita = eBayes(lmFit(cbind(R1,R2)))
padja = p.adjust(fita$p.value[,1], method="BH")
lfca = fita$coefficients[,1]

ExpVar = t(as.matrix(cbind(rep1=fita[[1]]$expVar, rep2=fita[[2]]$expVar)))
ExpVar = cbind(ExpVar[,1:2], ExpVar[,3] * fit2$expVar[1,1], ExpVar[,3] * fit2$expVar[2,1])
colnames(ExpVar) = c("total RNA", "background", "transl. eff.", "residuals")
row.names(ExpVar) = paste0("rep ", 1:2)
# barplot(ExpVar, beside=TRUE,
#         legend=TRUE, ylab="explained variance")
# barplot(t(ExpVar), beside=FALSE,
#         legend=TRUE, ylab="explained variance", xlim=c(0,4))

res = as.data.frame(Anno[row.names(EC),])
#res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

res$pval = fita$p.value[,1]
res$padj = padja
res$log2FoldChange = lfca
res$significant = ""
res$significant[which(padja <= 0.1)] = "*"
res$significant[which(padja <= 0.01)] = "**"
attr(res, "log2FC") = rowMeans(cbind(R1all, R2all))
attr(res, "log2total") = rowMeans(Dall[, IT])
attr(res, "log2FCsd") = rowSds(cbind(R1all, R2all))
attr(res, "log2TotalSD") = rowSds(Dall[, IT])

```

The R/Bioconductor package limma shrinks the standard deviation of each gene towards a global estimate of standard deviation along all genes. In the case of large outliers, this can result in spurious hits. To avoid these, genes with standard deviation larger than the absolute value of the log-fold change are not regarded as significant.

```

K = which(res$significant != "" & (fita$sigma > abs(fita$coefficients[,1])))

write.table(res[K,], sep="\t", file = file.path(d, "filteredGenes.txt"), quote=FALSE)

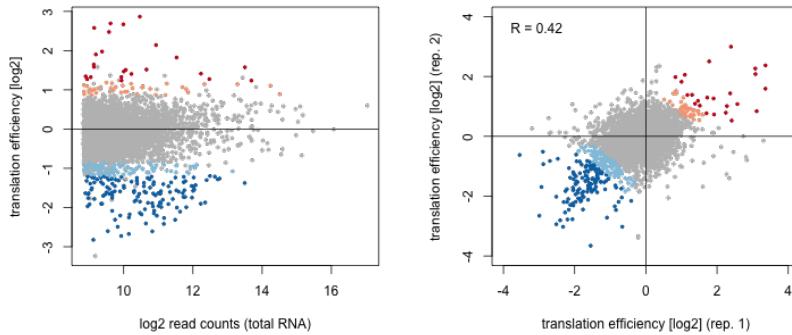
res$padj[K] = NA
res$significant[K] = ""

col=rep("gray", length(padja))
col[which(res$significant == "*" & lfca > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$significant == "*" & lfca < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$significant == "**" & lfca > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$significant == "**" & lfca < 0)] = brewer.pal(4, "RdBu")[4]
plot(rowMeans(D[,SL$type == "RNaseq"]), lfca,
      pch=20, cex=0.7, col=col,
      xlab="log2 read counts (total RNA)", ylab="translation efficiency [log2]")
abline(h=0)

plot(R1, R2,
      pch=20, cex=0.7, col=col,
      xlab="translation efficiency [log2] (rep. 1)",
      ylab="translation efficiency [log2] (rep. 2)",
      xlim=c(-4,4), ylim=c(-4,4))

```

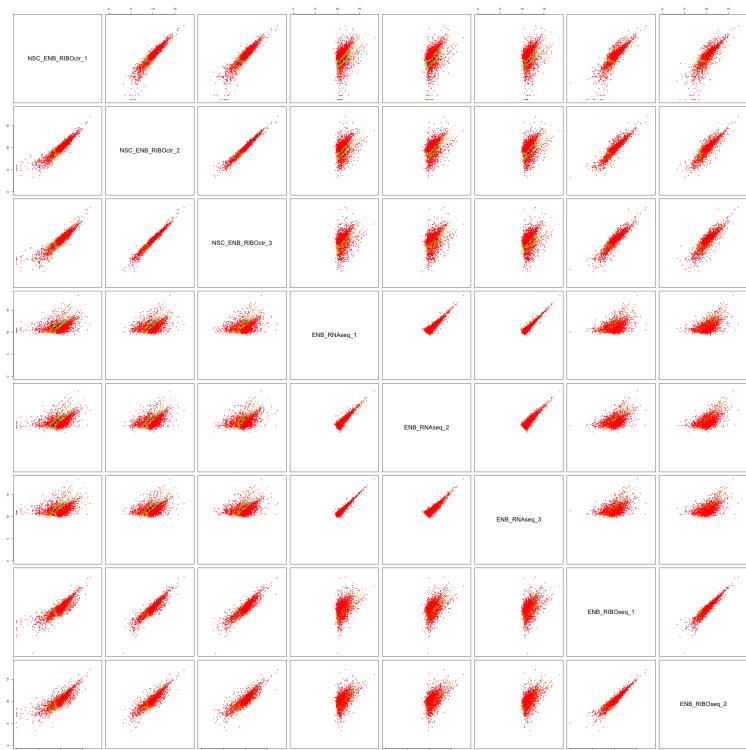
```
abline(h=0, v=0)
text(-3.8, 3.8, paste0("R = ", format(cor(R1,R2,use="pairwise.complete"), digits=2)), adj = c(0,1))
```



The data is saved.

```
save(EC, file=file.path(d, "EC.rda"))
save(res, file=file.path(d, "res.rda"))
save(fitc, file=file.path(d, "fitc.rda"))
save(ExpVar, file=file.path(d, "ExpVar.rda"))

col = rep("#00000022", dim(C)[1])
col[which(fitc$padj <= 0.1)] = "orange"
col[which(fitc$padj <= 0.01)] = "red"
I = order(factor(col, levels=c("#00000022", "orange", "red")))
lim = range(as.vector(C))
pairs(C[I,], pch=20, cex=0.5, xlim=lim, ylim=lim, col=col[I])
```



```
col=rep("#00000022", nrow(res))
col[which(res$significant == "*" & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$significant == "*" & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$significant == "***" & res$log2FoldChange > 0)] = brewer.pal(4, "RdBu")[1]
```

```

col[which(res$significant == "***" & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[4]

I = order(factor(col, levels=c("#00000022", brewer.pal(4, "RdBu")[c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[I,], pch=20, cex=0.5, xlim=lim, ylim=lim, col=col[I])

```



```

fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

```

```

res2 = res[order(factor(res$significant, levels=c("!!!", "**", "*")), 
                 sign(-res$log2FoldChange), res$symbol),]
res2$UnspecBinders = ""
res2$UnspecBinders[row.names(res2) %in% names(UnspecBinders)] = "UnspecBinder"
write.table(res2, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d, "resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Translation efficiency (%s); result table", n),
       heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR", page=page, br=TRUE)
col = rep("", nrow(res))
col[which(res2$significant %in% c("!!!", "**") & res2$log2FoldChange > 0)] =
  brewer.pal(3, "Pastel1")[1]
col[which(res2$significant %in% c("!!!", "**") & res2$log2FoldChange < 0)] =
  brewer.pal(3, "Pastel1")[2]
hwrite(as.data.frame(res2), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)

```

3.3 Filter unspecific binders in LNB

Create an output directory.

```
n = "LNB"
d = file.path("result", "TranslationEfficiency", n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../..", "mainmenu.html")
source(file.path("R", "doGSEA.R"))
```

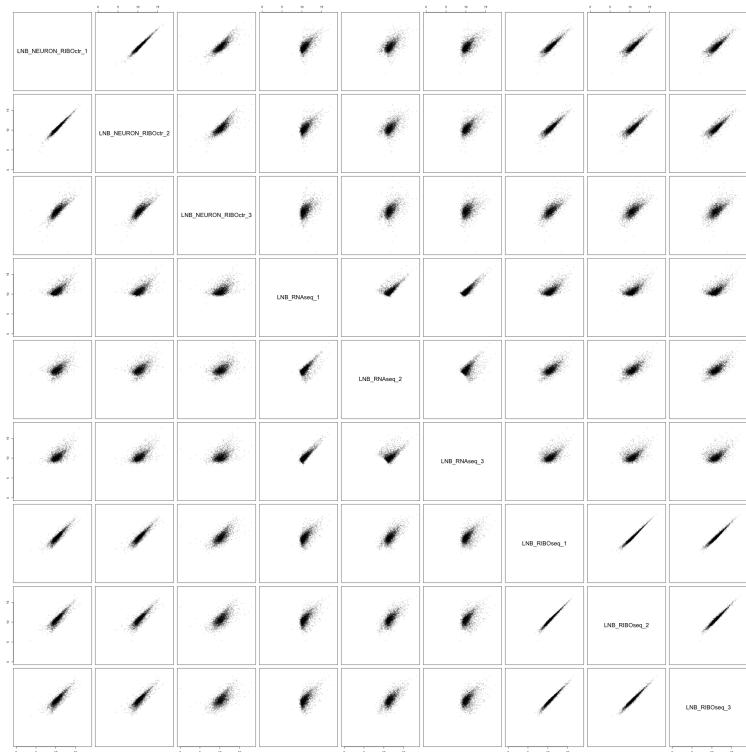
A DESeq dataset is created that contains the respective samples.

```
Design = DesignTranslationEfficiency
EC_all <- expectedCounts[, SamplesTranslationEfficiency[[n]]]
SL <- SampleList[SamplesTranslationEfficiency[[n]], ]
# dds_all <- DESeqDataSetFromMatrix(
#   countData = expectedCounts[, SamplesTranslationEfficiency[[n]]],
#   colData = SampleList[SamplesTranslationEfficiency[[n]], ],
#   design = ~ Type)
# mcols(dds_all) = Anno
# colData(dds_all)
```

Log counts are computed and plotted as a pairs plot.

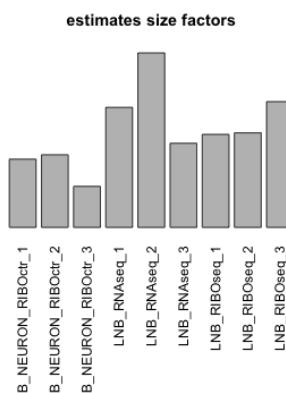
```
I = which(rowMeans(log2(EC_all[, SL$Type == "RNaseq"])+1)) >= 10 &
      Anno$transcript_type == "protein_coding")
EC = EC_all[I, ]
# I = which(rowMeans(log2(counts(dds_all)[, colData(dds_all)$Type == "RNaseq"])+1)) >= 10 &
#       mcols(dds_all)$transcript_type == "protein_coding")
# dds = dds_all[I, ]

C = log2(EC+1)
Call = log2(EC_all+1)
# C = log2(counts(dds)+1)
# Call = log2(counts(dds_all)+1)
lim = range(as.vector(C))
pairs(C, pch=20, cex=0.5, col="#00000022", xlim=lim, ylim=lim)
```



The genes in the cluster with lower read counts in the control sample are used to estimate the size factors.

```
s = sizeFactors(estimateSizeFactors(DESeqDataSetFromMatrix(
  countData = round(EC), colData = SL, design = ~ Type),
  controlGenes=which(rowMeans(log2(EC[,SL$Type == "RNAseq"]+1)) >= 13)))
## converting counts to integer mode
sizeFactors(dds) = s
p = par(mar=c(10,4,4,2)+0.1)
barplot(s, main="estimates size factors", las=2)
par(p)
```



The count data is normalized and unspecific binders are identified. Therefore, RiboControl samples are compared to total RNA samples. The genes that are significantly higher in RiboControl samples are potential unspecific binders of the tag. These genes are filtered as well.

```
D = C
m = log2(s)
for (i in 1:ncol(EC)) {
  D[,i] = D[,i] - m[i]
}
```

```

Dall = Call
for (i in 1:ncol(EC)) {
  Dall[,i] = Dall[,i] - m[i]
}

IR = which(SL$type == "RIBOseq")
IT = which(SL$type == "RNAseq")
IC = which(SL$type == "RIBOctr")

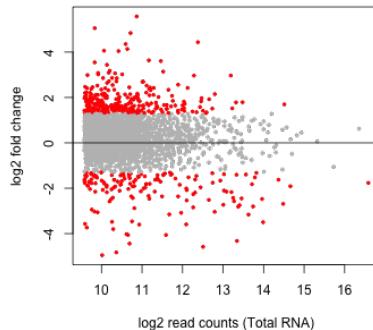
fitc = eBayes(lmFit(cbind(D[,IC[1]]-D[,IT[1]],D[,IC[2]]-D[,IT[2]])))
padjc = p.adjust(fitc$p.value[,1],method="BH")
lfcc = fitc$coefficients[,1]

plot(rowMeans(D[,SL$type == "RNAseq"]),lfcc,
  pch=20,cex=0.7,col=ifelse(padjc < 0.01, "red","gray"),
  xlab="log2 read counts (Total RNA)", ylab="log2 fold change")
abline(h=0)

UnspecBinders = which(padjc <= 0.01 & lfcc > 0)

fitc$padj = padjc

```



To estimate the translation efficiency, the RiboTag data is modeled as a function of RiboControl and total RNA.

```

fit1 = list()
for (i in 1:2) {
  D2 = data.frame("Ribo" = D[,IR[i]], "Total" = D[,IT[i]], "ctrl" = D[,IC[i]])
  fit1[[i]] = lm(Ribo ~ Total+ctrl, data=D2)
  fit1[[i]]$anova = anova(fit1[[i]])
  fit1[[i]]$expVar = fit1[[i]]$anova[, "Sum Sq", drop=FALSE]
  fit1[[i]]$expVar = fit1[[i]]$expVar/sum(fit1[[i]]$expVar)
  D2all = data.frame("Ribo" = Dall[,IR[i]], "Total" = Dall[,IT[i]], "ctrl" = Dall[,IC[i]], row.names =
  p = predict(fit1[[i]], newdata = D2all)
  fit1[[i]]$residualsAll = D2all$Ribo - p
}

R1 = fit1[[1]]$residuals
R2 = fit1[[2]]$residuals
R1all = fit1[[1]]$residualsAll
R2all = fit1[[2]]$residualsAll
R1all = R1all - shorth(R1,na.rm=TRUE)
R2all = R2all - shorth(R2,na.rm=TRUE)
R1 = R1 - shorth(R1,na.rm=TRUE)
R2 = R2 - shorth(R2,na.rm=TRUE)

```

```

D2 = data.frame("R1" = R1, "R2" = R2)
fit2 = lm(data=D2)
fit2$anova = anova(fit2)
fit2$expVar = fit2$anova[, "Sum Sq", drop=FALSE]
fit2$expVar = fit2$expVar/sum(fit2$expVar)

fita = eBayes(lmFit(cbind(R1,R2)))
padja = p.adjust(fita$p.value[,1], method="BH")
lfca = fita$coefficients[,1]

ExpVar = t(as.matrix(cbind(rep1=fita[[1]]$expVar, rep2=fita[[2]]$expVar)))
ExpVar = cbind(ExpVar[,1:2], ExpVar[,3] * fit2$expVar[1,1], ExpVar[,3] * fit2$expVar[2,1])
colnames(ExpVar) = c("total RNA", "background", "transl. eff.", "residuals")
row.names(ExpVar) = paste0("rep ", 1:2)
# barplot(ExpVar, beside=TRUE,
#         legend=TRUE, ylab="explained variance")
# barplot(t(ExpVar), beside=FALSE,
#         legend=TRUE, ylab="explained variance", xlim=c(0,4))

res = as.data.frame(Anno[row.names(EC),])
#res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

res$pval = fita$p.value[,1]
res$padj = padja
res$log2FoldChange = lfca
res$significant = ""
res$significant[which(padja <= 0.1)] = "*"
res$significant[which(padja <= 0.01)] = "**"
attr(res, "log2FC") = rowMeans(cbind(R1all, R2all))
attr(res, "log2total") = rowMeans(Dall[, IT])
attr(res, "log2FCsd") = rowSds(cbind(R1all, R2all))
attr(res, "log2TotalSD") = rowSds(Dall[, IT])

```

The R/Bioconductor package limma shrinks the standard deviation of each gene towards a global estimate of standard deviation along all genes. In the case of large outliers, this can result in spurious hits. To avoid these, genes with standard deviation larger than the absolute value of the log-fold change are not regarded as significant.

```

K = which(res$significant != "" & (fita$sigma > abs(fita$coefficients[,1])))

write.table(res[K,], sep="\t", file = file.path(d, "filteredGenes.txt"), quote=FALSE)

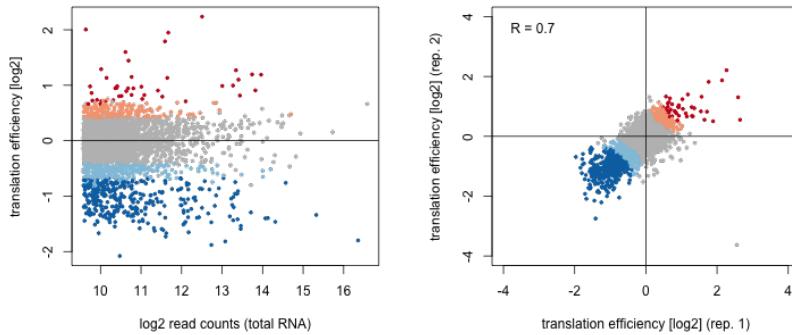
res$padj[K] = NA
res$significant[K] = ""

col=rep("gray", length(padja))
col[which(res$significant == "*" & lfca > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$significant == "*" & lfca < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$significant == "**" & lfca > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$significant == "**" & lfca < 0)] = brewer.pal(4, "RdBu")[4]
plot(rowMeans(D[,SL$type == "RNaseq"]), lfca,
      pch=20, cex=0.7, col=col,
      xlab="log2 read counts (total RNA)", ylab="translation efficiency [log2]")
abline(h=0)

plot(R1, R2,
      pch=20, cex=0.7, col=col,
      xlab="translation efficiency [log2] (rep. 1)",
      ylab="translation efficiency [log2] (rep. 2)",
      xlim=c(-4,4), ylim=c(-4,4))

```

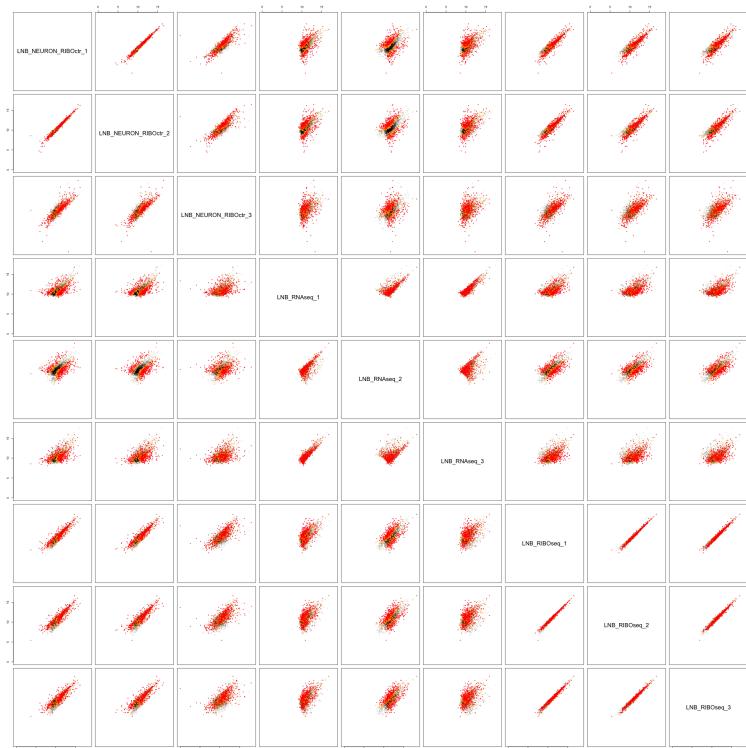
```
abline(h=0,v=0)
text(-3.8,3.8,paste0("R = ",format(cor(R1,R2,use="pairwise.complete"),digits=2)),adj = c(0,1))
```



The data is saved.

```
save(EC, file=file.path(d, "EC.rda"))
save(res, file=file.path(d, "res.rda"))
save(fitc, file=file.path(d, "fitc.rda"))
save(ExpVar, file=file.path(d, "ExpVar.rda"))

col = rep("#00000022", dim(C)[1])
col[which(fitc$padj <= 0.1)] = "orange"
col[which(fitc$padj <= 0.01)] = "red"
I = order(factor(col, levels=c("#00000022","orange","red")))
lim = range(as.vector(C))
pairs(C[I,],pch=20,cex=0.5,xlim=lim, ylim=lim,col=col[I])
```



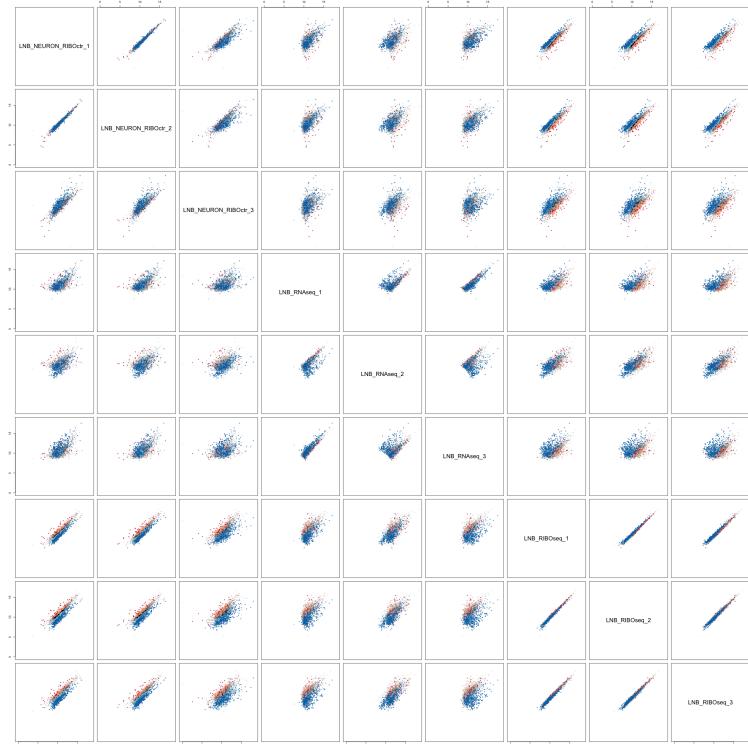
```
col=rep("#00000022", nrow(res))
col[which(res$significant == "*" & res$log2FoldChange > 0)] = brewer.pal(4,"RdBu")[2]
col[which(res$significant == "*" & res$log2FoldChange < 0)] = brewer.pal(4,"RdBu")[3]
col[which(res$significant == "***" & res$log2FoldChange > 0)] = brewer.pal(4,"RdBu")[1]
```

```

col[which(res$significant == "***" & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu")[4]

I = order(factor(col, levels=c("#00000022", brewer.pal(4, "RdBu")[c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[I,], pch=20, cex=0.5, xlim=lim, ylim=lim, col=col[I])

```



```

fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

```

```

res2 = res[order(factor(res$significant, levels=c("***", "**", "*"))),
           sign(-res$log2FoldChange), res$symbol,]
res2$UnspecBinders = ""
res2$UnspecBinders[row.names(res2) %in% names(UnspecBinders)] = "UnspecBinder"
write.table(res2, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d, "resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Translation efficiency (%s); result table", n),
       heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR", page=page, br=TRUE)
col = rep("", nrow(res))
col[which(res2$significant %in% c("***", "**")) & res2$log2FoldChange > 0] =
  brewer.pal(3, "Pastel1")[1]
col[which(res2$significant %in% c("***", "**")) & res2$log2FoldChange < 0] =
  brewer.pal(3, "Pastel1")[2]
hwrite(as.data.frame(res2), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)

```

3.4 Filter unspecific binders in NEURON

Create an output directory.

```
n = "NEURON"
d = file.path("result", "TranslationEfficiency", n)
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../..", "mainmenu.html")
source(file.path("R", "doGSEA.R"))
```

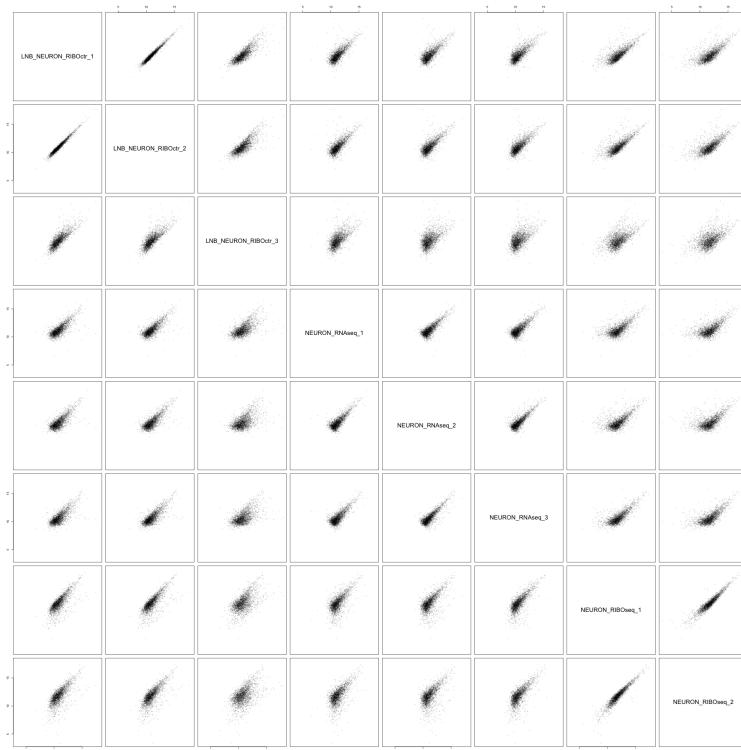
A DESeq dataset is created that contains the respective samples.

```
Design = DesignTranslationEfficiency
EC_all <- expectedCounts[, SamplesTranslationEfficiency[[n]]]
SL <- SampleList[SamplesTranslationEfficiency[[n]], ]
# dds_all <- DESeqDataSetFromMatrix(
#   countData = expectedCounts[, SamplesTranslationEfficiency[[n]]],
#   colData = SampleList[SamplesTranslationEfficiency[[n]], ],
#   design = ~ Type)
# mcols(dds_all) = Anno
# colData(dds_all)
```

Log counts are computed and plotted as a pairs plot.

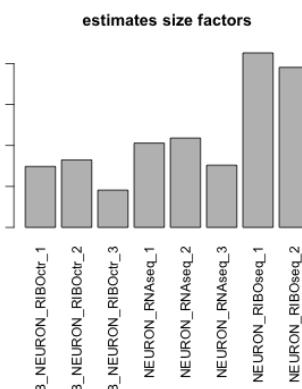
```
I = which(rowMeans(log2(EC_all[, SL$Type == "RNaseq"])+1)) >= 10 &
      Anno$transcript_type == "protein_coding")
EC = EC_all[I, ]
# I = which(rowMeans(log2(counts(dds_all)[, colData(dds_all)$Type == "RNaseq"])+1)) >= 10 &
#       mcols(dds_all)$transcript_type == "protein_coding")
# dds = dds_all[I, ]

C = log2(EC+1)
Call = log2(EC_all+1)
# C = log2(counts(dds)+1)
# Call = log2(counts(dds_all)+1)
lim = range(as.vector(C))
pairs(C, pch=20, cex=0.5, col="#00000022", xlim=lim, ylim=lim)
```



The genes in the cluster with lower read counts in the control sample are used to estimate the size factors.

```
s = sizeFactors(estimateSizeFactors(DESeqDataSetFromMatrix(
  countData = round(EC), colData = SL, design = ~ Type),
  controlGenes=which(rowMeans(log2(EC[,SL$Type == "RNaseq"]+1)) >= 13)))
## converting counts to integer mode
#sizeFactors(dds) = s
p = par(mar=c(10,4,4,2)+0.1)
barplot(s, main="estimates size factors", las=2)
par(p)
```



The count data is normalized and unspecific binders are identified. Therefore, RiboControl samples are compared to total RNA samples. The genes that are significantly higher in RiboControl samples are potential unspecific binders of the tag. These genes are filtered as well.

```
D = C
m = log2(s)
for (i in 1:ncol(EC)) {
  D[,i] = D[,i] - m[i]
}
```

```

Dall = Call
for (i in 1:ncol(EC)) {
  Dall[,i] = Dall[,i] - m[i]
}

IR = which(SL$type == "RIBOseq")
IT = which(SL$type == "RNAseq")
IC = which(SL$type == "RIBOctr")

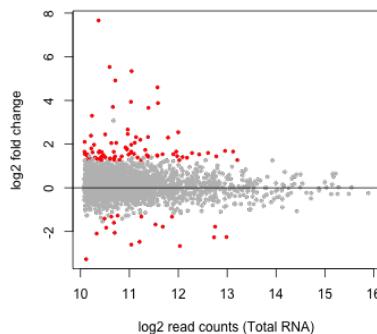
fitc = eBayes(lmFit(cbind(D[,IC[1]]-D[,IT[1]],D[,IC[2]]-D[,IT[2]])))
padjc = p.adjust(fitc$p.value[,1],method="BH")
lfcc = fitc$coefficients[,1]

plot(rowMeans(D[,SL$type == "RNAseq"]),lfcc,
  pch=20,cex=0.7,col=ifelse(padjc < 0.01, "red","gray"),
  xlab="log2 read counts (Total RNA)", ylab="log2 fold change")
abline(h=0)

UnspecBinders = which(padjc <= 0.01 & lfcc > 0)

fitc$padj = padjc

```



To estimate the translation efficiency, the RiboTag data is modeled as a function of RiboControl and total RNA.

```

fit1 = list()
for (i in 1:2) {
  D2 = data.frame("Ribo" = D[,IR[i]], "Total" = D[,IT[i]], "ctrl" = D[,IC[i]])
  fit1[[i]] = lm(Ribo ~ Total+ctrl, data=D2)
  fit1[[i]]$anova = anova(fit1[[i]])
  fit1[[i]]$expVar = fit1[[i]]$anova[, "Sum Sq", drop=FALSE]
  fit1[[i]]$expVar = fit1[[i]]$expVar/sum(fit1[[i]]$expVar)
  D2all = data.frame("Ribo" = Dall[,IR[i]], "Total" = Dall[,IT[i]], "ctrl" = Dall[,IC[i]], row.names =
  p = predict(fit1[[i]], newdata = D2all)
  fit1[[i]]$residualsAll = D2all$Ribo - p
}

R1 = fit1[[1]]$residuals
R2 = fit1[[2]]$residuals
R1all = fit1[[1]]$residualsAll
R2all = fit1[[2]]$residualsAll
R1all = R1all - shorth(R1,na.rm=TRUE)
R2all = R2all - shorth(R2,na.rm=TRUE)
R1 = R1 - shorth(R1,na.rm=TRUE)
R2 = R2 - shorth(R2,na.rm=TRUE)

```

```

D2 = data.frame("R1" = R1, "R2" = R2)
fit2 = lm(data=D2)
fit2$anova = anova(fit2)
fit2$expVar = fit2$anova[, "Sum Sq", drop=FALSE]
fit2$expVar = fit2$expVar/sum(fit2$expVar)

fita = eBayes(lmFit(cbind(R1,R2)))
padja = p.adjust(fita$p.value[,1], method="BH")
lfca = fita$coefficients[,1]

ExpVar = t(as.matrix(cbind(rep1=fita[[1]]$expVar, rep2=fita[[2]]$expVar)))
ExpVar = cbind(ExpVar[,1:2], ExpVar[,3] * fit2$expVar[1,1], ExpVar[,3] * fit2$expVar[2,1])
colnames(ExpVar) = c("total RNA", "background", "transl. eff.", "residuals")
row.names(ExpVar) = paste0("rep ", 1:2)
# barplot(ExpVar, beside=TRUE,
#         legend=TRUE, ylab="explained variance")
# barplot(t(ExpVar), beside=FALSE,
#         legend=TRUE, ylab="explained variance", xlim=c(0,4))

res = as.data.frame(Anno[row.names(EC),])
#res$symbol[is.na(res$symbol)] = row.names(res)[is.na(res$symbol)]

res$pval = fita$p.value[,1]
res$padj = padja
res$log2FoldChange = lfca
res$significant = ""
res$significant[which(padja <= 0.1)] = "*"
res$significant[which(padja <= 0.01)] = "**"
attr(res, "log2FC") = rowMeans(cbind(R1all, R2all))
attr(res, "log2total") = rowMeans(Dall[, IT])
attr(res, "log2FCsd") = rowSds(cbind(R1all, R2all))
attr(res, "log2TotalSD") = rowSds(Dall[, IT])

```

The R/Bioconductor package limma shrinks the standard deviation of each gene towards a global estimate of standard deviation along all genes. In the case of large outliers, this can result in spurious hits. To avoid these, genes with standard deviation larger than the absolute value of the log-fold change are not regarded as significant.

```

K = which(res$significant != "" & (fita$sigma > abs(fita$coefficients[,1])))

write.table(res[K,], sep="\t", file = file.path(d, "filteredGenes.txt"), quote=FALSE)

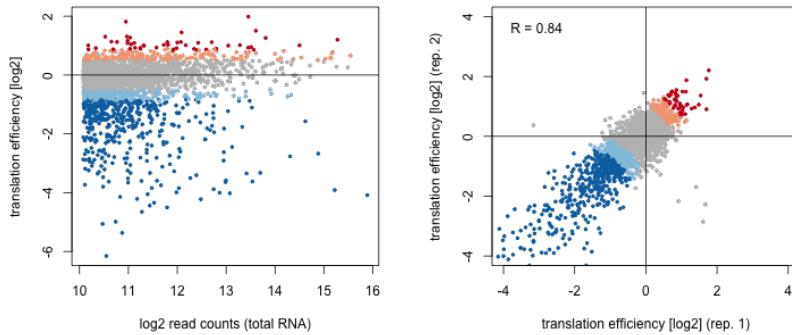
res$padj[K] = NA
res$significant[K] = ""

col=rep("gray", length(padja))
col[which(res$significant == "*" & lfca > 0)] = brewer.pal(4, "RdBu")[2]
col[which(res$significant == "*" & lfca < 0)] = brewer.pal(4, "RdBu")[3]
col[which(res$significant == "**" & lfca > 0)] = brewer.pal(4, "RdBu")[1]
col[which(res$significant == "**" & lfca < 0)] = brewer.pal(4, "RdBu")[4]
plot(rowMeans(D[,SL$type == "RNaseq"]), lfca,
      pch=20, cex=0.7, col=col,
      xlab="log2 read counts (total RNA)", ylab="translation efficiency [log2]")
abline(h=0)

plot(R1, R2,
      pch=20, cex=0.7, col=col,
      xlab="translation efficiency [log2] (rep. 1)",
      ylab="translation efficiency [log2] (rep. 2)",
      xlim=c(-4,4), ylim=c(-4,4))

```

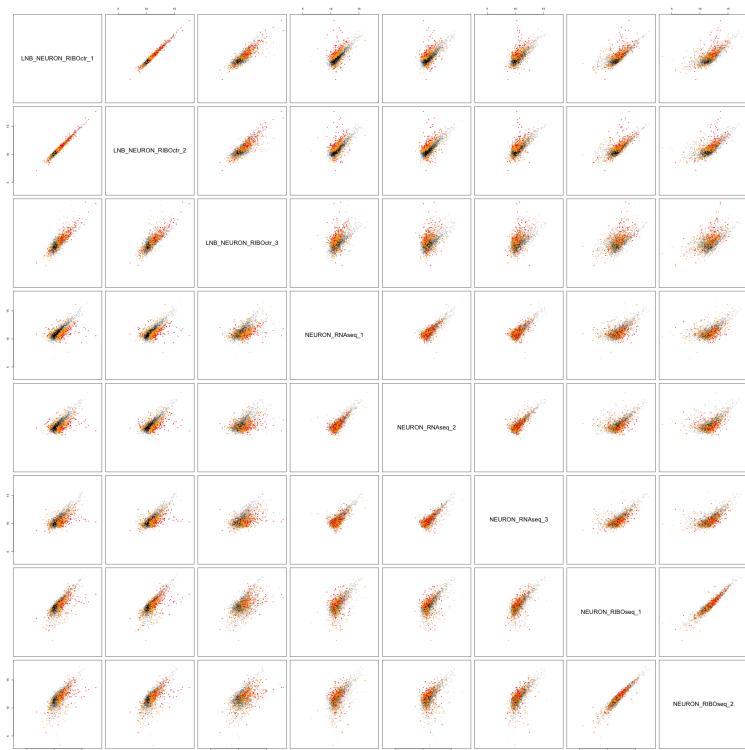
```
abline(h=0,v=0)
text(-3.8,3.8,paste0("R = ",format(cor(R1,R2,use="pairwise.complete"),digits=2)),adj = c(0,1))
```



The data is saved.

```
save(EC, file=file.path(d, "EC.rda"))
save(res, file=file.path(d, "res.rda"))
save(fitc, file=file.path(d, "fitc.rda"))
save(ExpVar, file=file.path(d, "ExpVar.rda"))

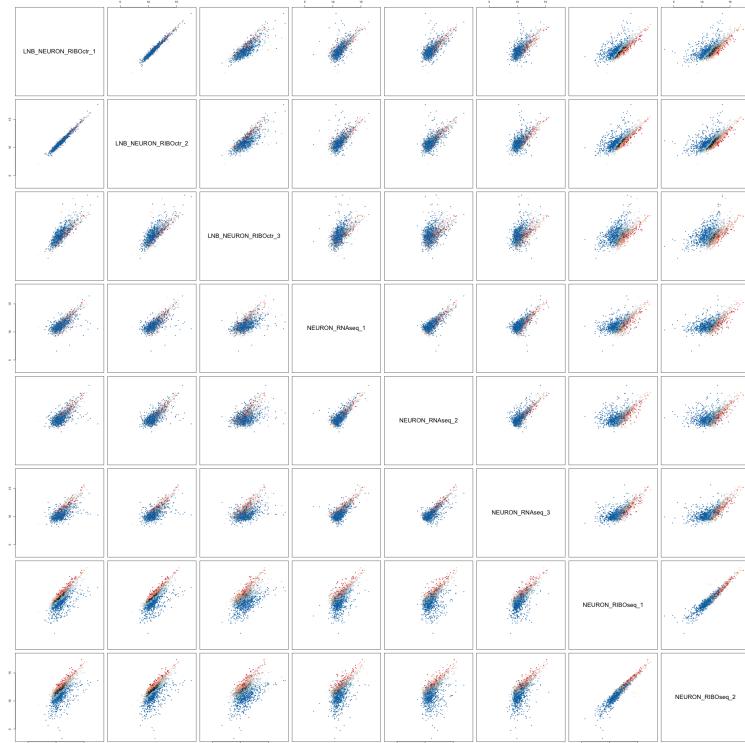
col = rep("#00000022", dim(C)[1])
col[which(fitc$padj <= 0.1)] = "orange"
col[which(fitc$padj <= 0.01)] = "red"
I = order(factor(col, levels=c("#00000022","orange","red")))
lim = range(as.vector(C))
pairs(C[I,],pch=20,cex=0.5,xlim=lim, ylim=lim,col=col[I])
```



```
col=rep("#00000022", nrow(res))
col[which(res$significant == "*" & res$log2FoldChange > 0)] = brewer.pal(4,"RdBu")[2]
col[which(res$significant == "*" & res$log2FoldChange < 0)] = brewer.pal(4,"RdBu")[3]
col[which(res$significant == "***" & res$log2FoldChange > 0)] = brewer.pal(4,"RdBu")[1]
```

```
col[which(res$significant == "***" & res$log2FoldChange < 0)] = brewer.pal(4, "RdBu") [4]

I = order(factor(col, levels=c("#00000022", brewer.pal(4, "RdBu") [c(2,3,1,4)])))
lim = range(as.vector(C))
pairs(C[I,], pch=20, cex=0.5, xlim=lim, ylim=lim, col=col[I])
```



```
fgup = row.names(res)[which(res$significant != "" & res$log2FoldChange > 0)]
fgdown = row.names(res)[which(res$significant != "" & res$log2FoldChange < 0)]
bg = row.names(res)[which(!is.na(res$padj))]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)
```

```
res2 = res[order(factor(res$significant, levels=c("!!!", "**", "*")), 
                 sign(-res$log2FoldChange), res$symbol),]
res2$UnspecBinders = ""
res2$UnspecBinders[row.names(res2) %in% names(UnspecBinders)] = "UnspecBinder"
write.table(res2, file=file.path(d, "resulttable.txt"), sep="\t", quote=FALSE)
page = openPage(file.path(d, "resulttable.html"), link.css="hwriter.css")
hwriteSidebar(page=page, link=mainmenu)
hwrite(sprintf("Translation efficiency (%s); result table", n),
       heading=1, page=page)
hwrite("** = 1% FDR; * = 10% FDR", page=page, br=TRUE)
col = rep("", nrow(res))
col[which(res2$significant %in% c("!!!", "**") & res2$log2FoldChange > 0)] =
  brewer.pal(3, "Pastel1")[1]
col[which(res2$significant %in% c("!!!", "**") & res2$log2FoldChange < 0)] =
  brewer.pal(3, "Pastel1")[2]
hwrite(as.data.frame(res2), row.bgcolor=col, page=page)
closePage(page, splash=FALSE)
```

3.5 Compare neural differentiation stages

Create an output directory.

```
d = file.path("result", "CompareStages", "")
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../..", "mainmenu.html")

library(grid)
source(file.path("R", "myballoonplot.R"))
load(file.path("data", "ENSG2category.rda"))
source(file.path("R", "doGSEA.R"))

log2FC = log2total = log2FCsd = log2totalSD = list()
S = names(SamplesTranslationEfficiency)
TransEff = list()
for (s in S) {
  load(file.path("result", "TranslationEfficiency", s, "res.rda"))
  TransEff[[s]] = res
  a = attributes(res)
  log2FC[[s]] = a$log2FC
  log2total[[s]] = a$log2total
  log2FCsd[[s]] = a$log2FCsd
  log2totalSD[[s]] = a$log2totalSD
}
log2FC = do.call(cbind, log2FC)
log2total = do.call(cbind, log2total)
log2FCsd = do.call(cbind, log2FCsd)
log2totalSD = do.call(cbind, log2totalSD)

for (s in names(TransEff)) {
  TransEff[[s]]$ENSG = row.names(TransEff[[s]])
  TransEff[[s]] = TransEff[[s]][,c("ENSG", "symbol", "log2FoldChange",
                                 "pval", "padj", "significant")]
  colnames(TransEff[[s]])[3:6] = paste(s, colnames(TransEff[[s]])[3:6], sep=".")
}

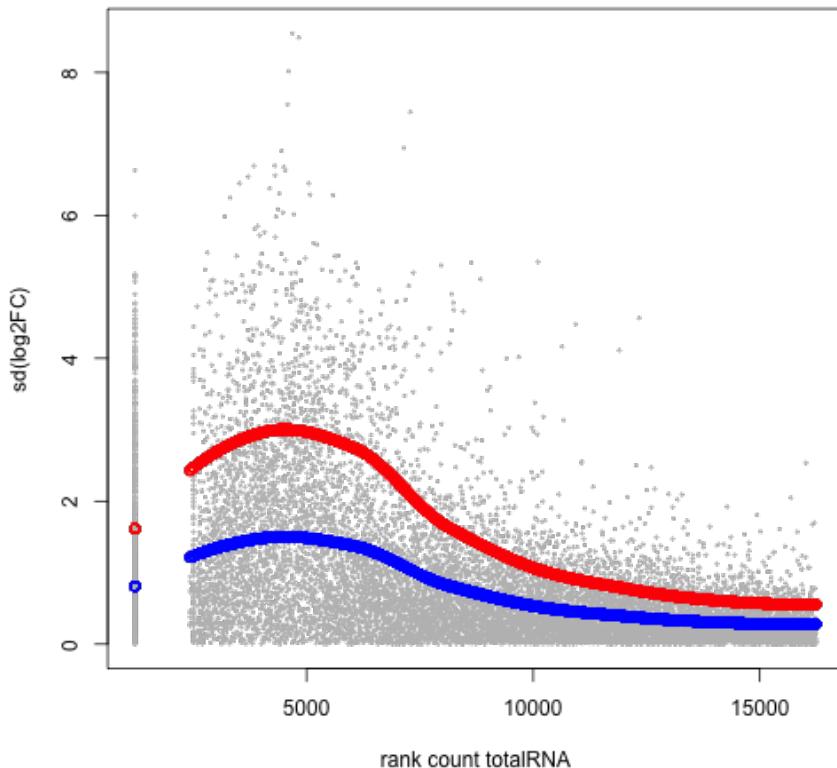
X = merge(TransEff[[1]], TransEff[[2]], by=c("ENSG", "symbol"),
          suffixes=c("", ""))
Y = merge(TransEff[[3]], TransEff[[4]], by=c("ENSG", "symbol"),
          suffixes=c("", ""))
Z = merge(X, Y, by=c("ENSG", "symbol"), suffixes=c("", ""), all=TRUE)
write.table(Z, file = file.path(d, "TableS4-TranslationEfficiency.txt"),
            sep="\t", row.names=FALSE, quote=FALSE)

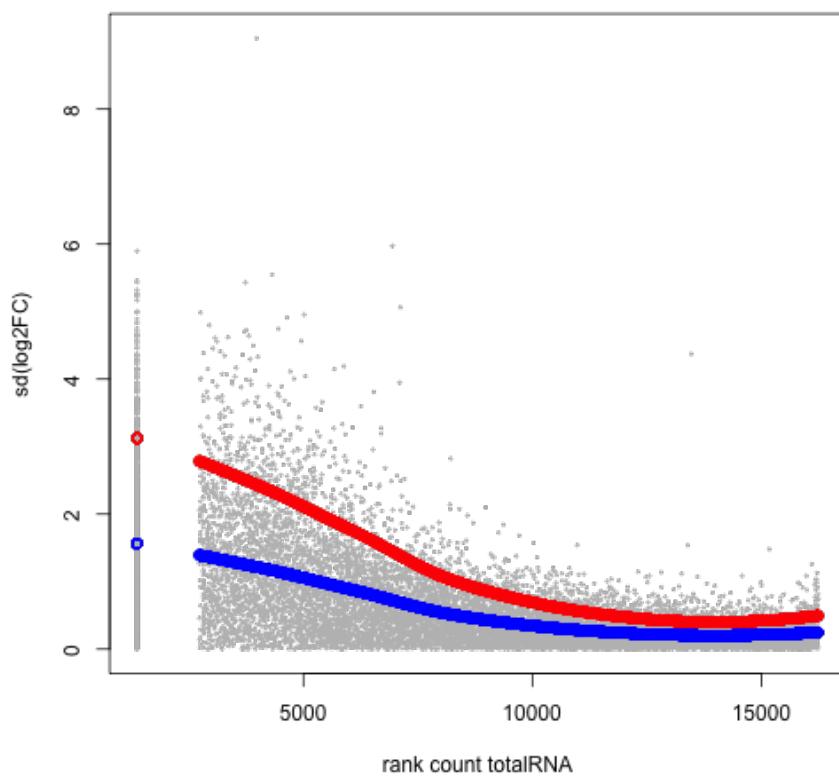
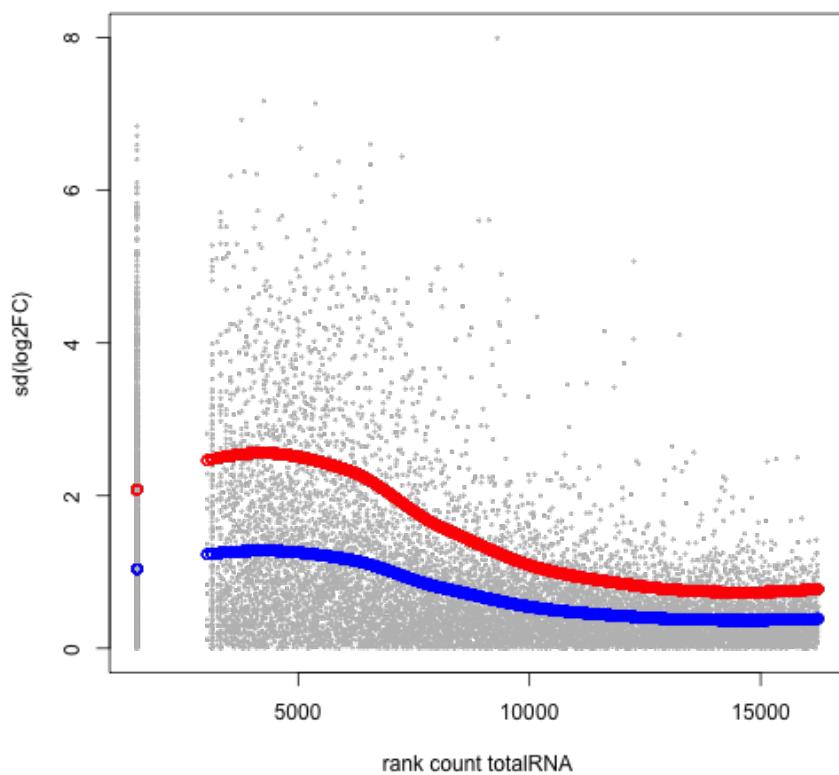
I = Anno$transcript_type == "protein_coding"
log2FC = log2FC[I,]
log2total = log2total[I,]
log2FCsd = log2FCsd[I,]
log2totalSD = log2totalSD[I,]
log2totalRank = log2total
for (s in S) {
  log2totalRank[,s] = rank(log2total[,s])
}

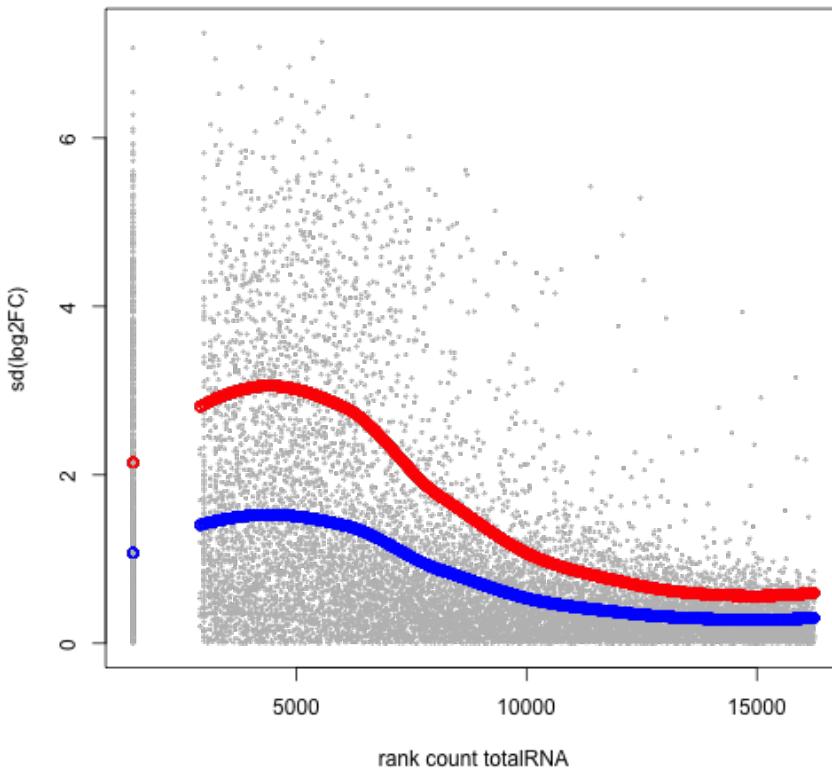
log2totalQ = log2total
Q = apply(log2totalQ, 2, quantile, probs=seq(0,1,length.out = 11))
q = apply(Q, 1, mean)
for (s in S) {
  log2totalQ[,s] = approx(x = Q[,s], q, xout=log2totalQ[,s], rule = 2)$y
```

```
}
```

```
for (s in S) {
  plot(log2totalRank[,s],log2FCsd[,s],pch=20,cex=0.3,col="grey",
    xlab="rank count totalRNA", ylab="sd(log2FC)")
  fit = loess(log2FCsd[,s] ~ rank(log2total[,s]))
  points(log2totalRank[,s],fit$fitted,col="blue")
  points(log2totalRank[,s],2*fit$fitted,col="red")
  SD = fit$fitted
  J = which(log2FCsd[,s] > 2*fit$fitted)
  SD[J] = log2FCsd[J,s]
  log2FCsd[,s] = log2FCsd[,s] / SD
}
```







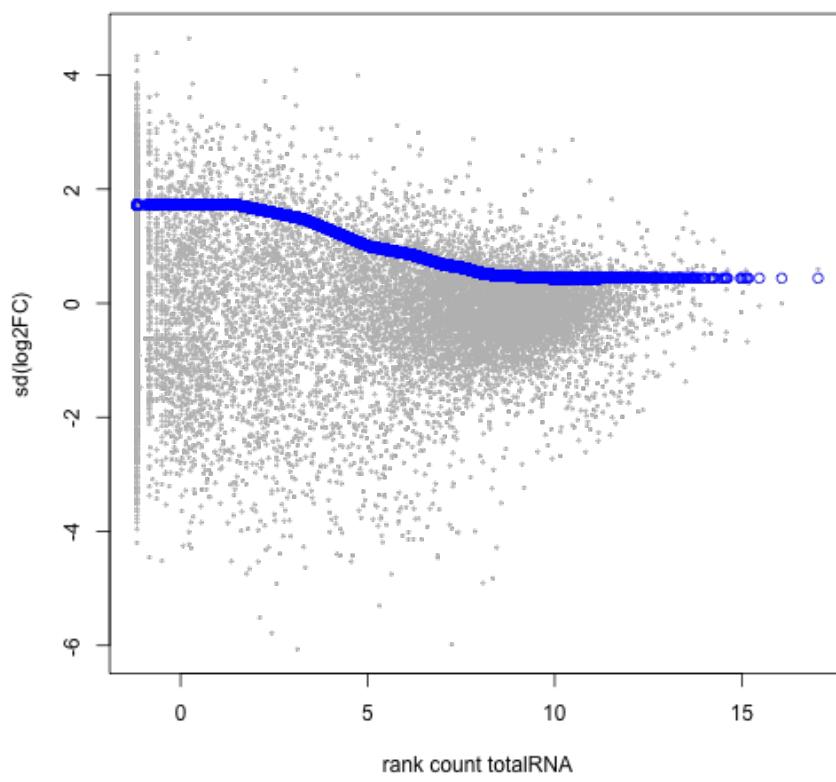
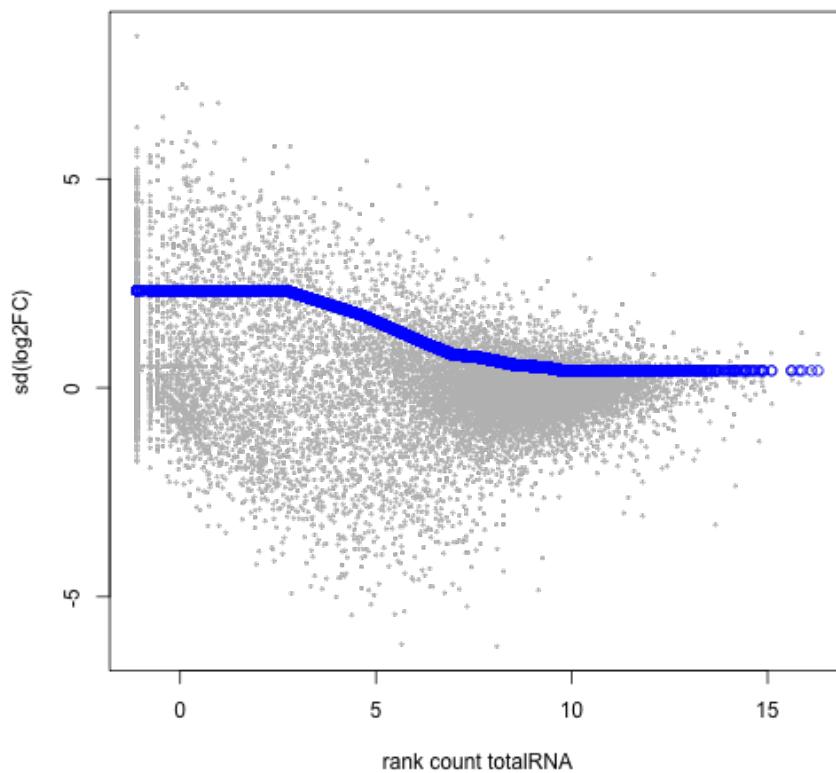
```

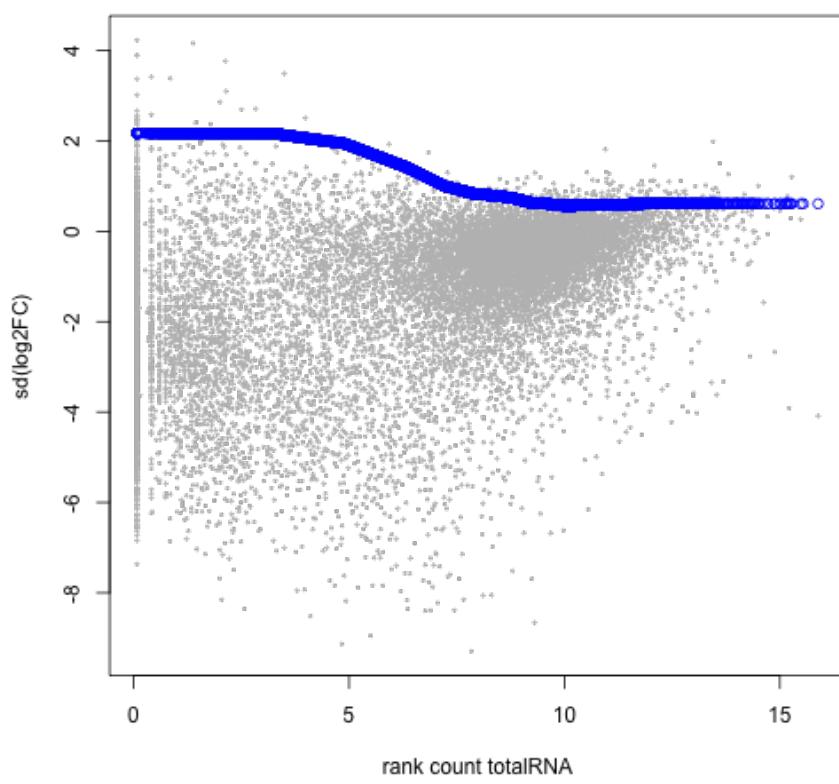
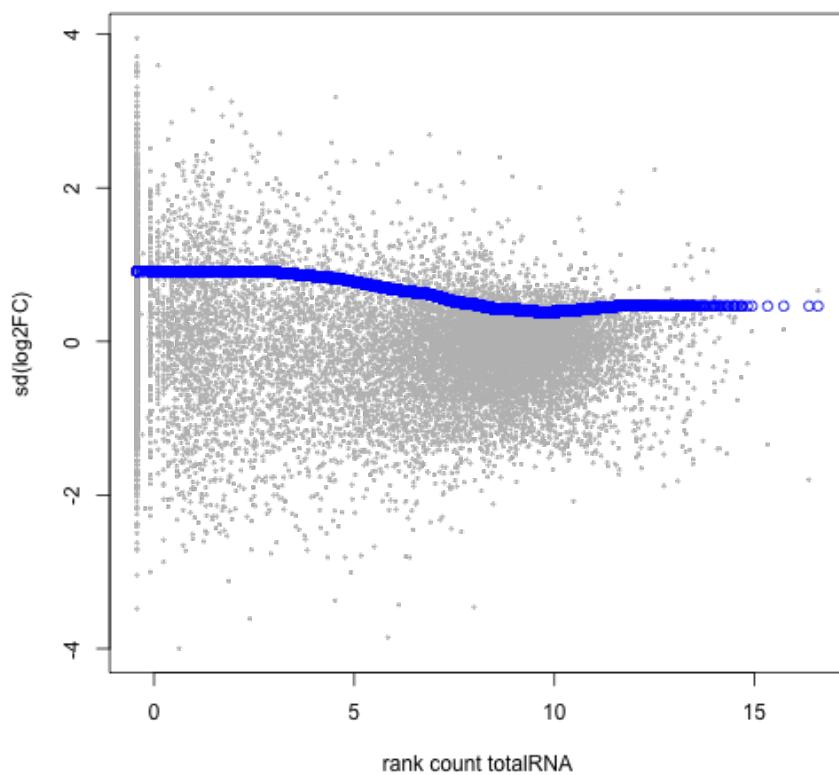
for (s in S) {
  b = cut(log2totalRank[,s],breaks=15)
#  M = tapply(log2FC[,s], b, median)
  MD = tapply(log2FC[,s], b, mad)
  x = tapply(log2total[,s], b, mean)
#  M[1:3] = M[4]
  MD[1:3] = MD[4]
#  Mnew = approx(x=x,y=M,xout=log2total[,s],rule = 2)
  MDnew = approx(x=x,y=MD,xout=log2total[,s],rule = 2)
  plot(log2total[,s],log2FC[,s],pch=20,cex=0.3,col="grey",
    xlab="rank count totalRNA", ylab="sd(log2FC)")
#  points(Mnew$x,Mnew$y,col="blue")
  points(MDnew$x,MDnew$y,col="blue")

  log2FC[,s] = log2FC[,s] / MDnew$y
#  log2FC[,s] =
#    (log2FC[,s] - Mnew$y) / MDnew$y

#  plot(log2FC[Annoftranscript_type == "protein_coding",s],
#  log2total[Annoftranscript_type == "protein_coding",s])
}

```

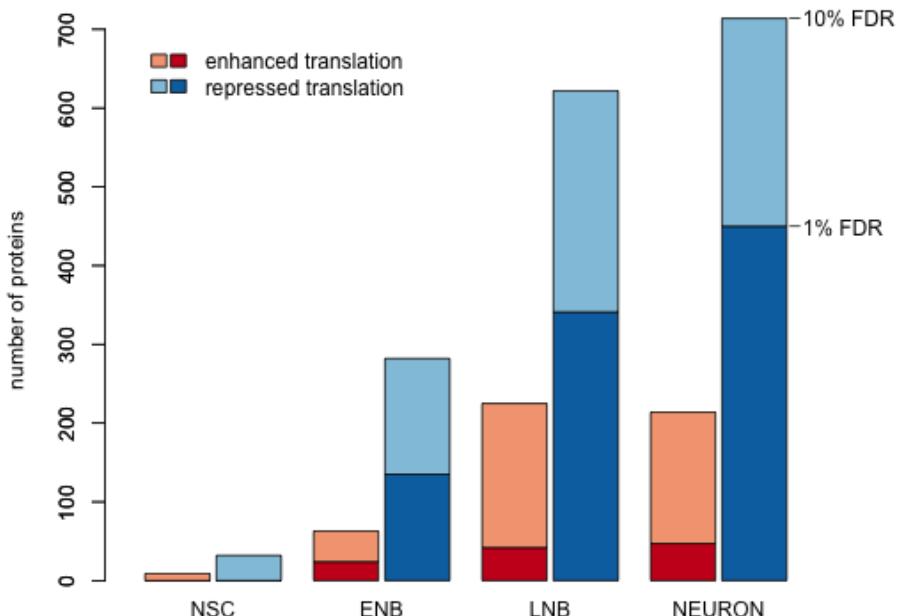




```

p = par(xpd=NA,mar=c(1.5,4,1.5,2.5))
H = matrix(NA, nrow=4, ncol=4)
colnames(H) = S
row.names(H) = c("highSigUp","sigUp","sigDown","highSigDown")
n = rep(NA,4)
names(n) = S
for (s in S) {
  load(file.path("result","TranslationEfficiency",s,"res.rda"))
  H["highSigUp",s] = sum(res$significant == "***" & res$log2FoldChange > 0,na.rm=TRUE)
  H["sigUp",s] = sum(res$significant == "**" & res$log2FoldChange > 0,na.rm=TRUE)
  H["sigDown",s] = sum(res$significant == "*" & res$log2FoldChange < 0,na.rm=TRUE)
  H["highSigDown",s] = sum(res$significant == "***" & res$log2FoldChange < 0,na.rm=TRUE)
  n[s] = sum(is.finite(res$padj))
}
col=brewer.pal(4,"RdBu")
ylim=c(0,max(H[c(1,4),]+H[c(2:3),]))
bp1 = barplot(H[1:2,],col=col[1:2],space=c(0.2,1.6,1.6,1.6),names.arg = rep("",4),
               xlim=c(0,10.4),ylim=ylim,ylab="number of proteins")
bp2 = barplot(H[4:3,],col=col[4:3],space=c(1.3,1.6,1.6,1.6),names.arg = rep("",4),add=TRUE)
text((bp1+bp2)/2,rep(-0.05*ylim[2],4),S)
legend(x=0.0,y=0.97*ylim[2],fill=col[c(2:3)],c("", ""),bty = "n")
legend(x=0.3,y=0.97*ylim[2],fill=col[c(1,4)],c("enhanced translation","repressed translation"),bty = "n")
x = bp2[4]+0.5
y = H["highSigDown", "NEURON"]
lines(c(x+0.05,x+0.2),c(y,y))
text(x+0.25,y = y,"1% FDR",adj = c(0,0.5))
y = H["sigDown", "NEURON"] + H["highSigDown", "NEURON"]
lines(c(x+0.05,x+0.2),c(y,y))
text(x+0.25,y = y,"10% FDR",adj = c(0,0.5))

```



```

for (i in 1:4) {
  H[,i] = H[,i] / n[i]
}
H = H*100

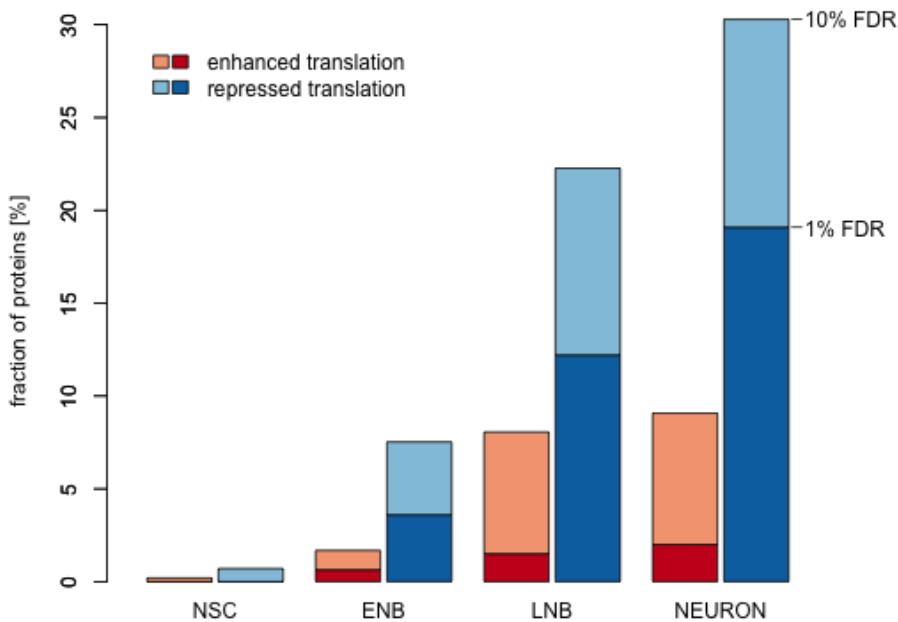
ylim=c(0,max(H[c(1,4),]+H[c(2:3),]))

```

```

bp1 = barplot(H[1:2,],col=col[1:2],space=c(0.2,1.6,1.6,1.6),names.arg = rep("",4),
              xlim=c(0,10.4),ylim=ylim,ylab="fraction of proteins [%]")
bp2 = barplot(H[4:3,],col=col[4:3],space=c(1.3,1.6,1.6,1.6),names.arg = rep("",4),add=TRUE)
text((bp1+bp2)/2,rep(-0.05*ylim[2],4),S)
legend(x=0.0,y=0.97*ylim[2],fill=col[c(2:3)],c("", ""),bty = "n")
legend(x=0.3,y=0.97*ylim[2],fill=col[c(1,4)],c("enhanced translation","repressed translation"),bty = "n")
x = bp2[4]+0.5
y = H["highSigDown", "NEURON"]
lines(c(x+0.05,x+0.2),c(y,y))
text(x+0.25,y = y,"1% FDR",adj = c(0,0.5))
y = H["sigDown", "NEURON"] + H["highSigDown", "NEURON"]
lines(c(x+0.05,x+0.2),c(y,y))
text(x+0.25,y = y,"10% FDR",adj = c(0,0.5))

```



```
par(p)
```

```

ENSG = c()
for (s in S) {
  load(file.path("result", "TranslationEfficiency", s, "res.rda"))
  ENSG = c(ENSG, row.names(res))
}
ENSG = sort(unique(ENSG))
X = matrix(factor("0", levels=c("repressed", "0", "enhanced")), nrow = length(ENSG), ncol=4)
row.names(X) = ENSG
colnames(X) = S
for (s in S) {
  load(file.path("result", "TranslationEfficiency", s, "res.rda"))
  a = res$significant
  a[a == "***] = "enhanced"
  a[a == **] = "enhanced"
  a[res$log2FoldChange < 0 & a == "enhanced"] = "repressed"
  a[a == ""] = "0"
  X[row.names(res),s] = a
}

t1 = table("NSC" = factor(X[,1],levels=c("repressed", "0", "enhanced")),
           "ENB"=factor(X[,2],levels=c("repressed", "0", "enhanced")))
t2 = table("ENB" = factor(X[,2],levels=c("repressed", "0", "enhanced")),
           "LNB"=factor(X[,3],levels=c("repressed", "0", "enhanced")))

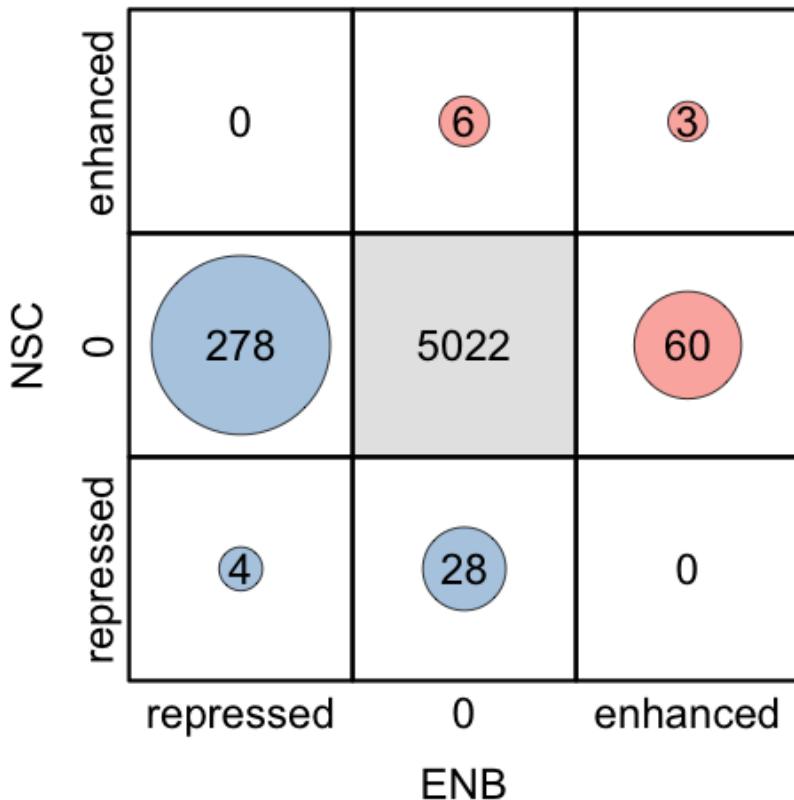
```

```

"LNB"=factor(X[,3],levels=c("repressed","0","enhanced")))
t3 = table("LNB" = factor(X[,3],levels=c("repressed","0","enhanced")),
           "NEURON"=factor(X[,4],levels=c("repressed","0","enhanced")))

col = matrix("gray90", nrow=3, ncol=3)
col[1,1] = col[2,1] = col[1,2] = brewer.pal(9, "Pastel1")[2]
col[3,3] = col[2,3] = col[3,2] = brewer.pal(9, "Pastel1")[1]
myballoonplot(t1, col = col, cex = 2)

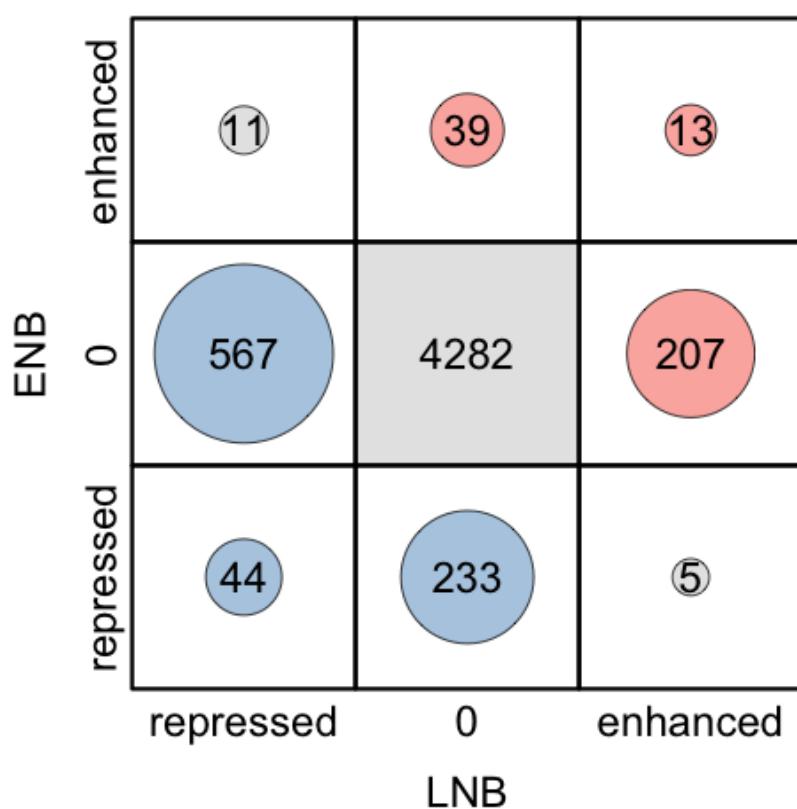
```



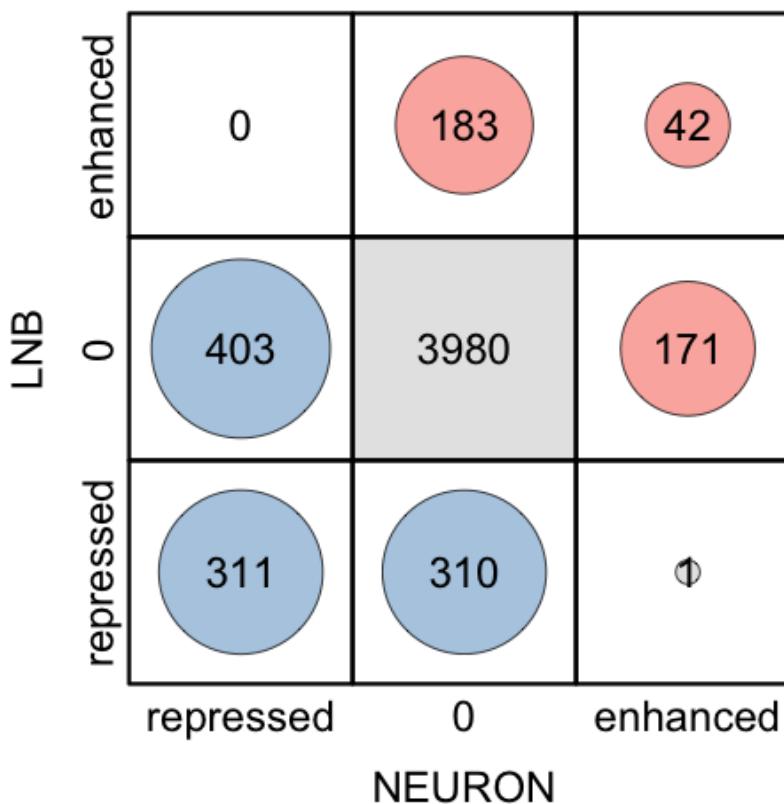
```

grid.newpage()
myballoonplot(t2, col = col, cex = 2)

```



```
grid.newpage()  
myballoonplot(t3, col = col, cex = 2)
```



```

fgdown = row.names(X)[which(X[,3] == "repressed" & X[,4] == "repressed")]
fgup = row.names(X)[which(X[,3] == "enhanced" & X[,4] == "enhanced")]
load(file.path("result", "TranslationEfficiency", "LNB", "res.rda"))
bg = row.names(res)
load(file.path("result", "TranslationEfficiency", "NEURON", "res.rda"))
bg = bg[bg %in% row.names(res)]
GSEAup = doGSEA(fgup, bg, ENSG2category, k=3)
GSEAdown = doGSEA(fgdown, bg, ENSG2category, k=3)

```

```

library(cluster)
colG = colH = G = H = list()
for (s in S) {
  load(file.path("result", "TranslationEfficiency", s, "res.rda"))
  K = rep(3, nrow(res))
  K[which(res$significant == "*" & res$log2FoldChange > 0)] = 4
  K[which(res$significant == "**" & res$log2FoldChange > 0)] = 5
  K[which(res$significant == "*" & res$log2FoldChange < 0)] = 2
  K[which(res$significant == "**" & res$log2FoldChange < 0)] = 1

  G[[s]] = row.names(res)[which(K == 5)]
  H[[s]] = row.names(res)[which(K == 1)]

  # X = cbind(log2FC, log2totalQ)
  # for (i in seq_len(ncol(X))) {
  #   X[, i] = X[, i] - median(X[, i])
  #   X[, i] = X[, i] / mad(X[, i])
  # }
  #
  # if (length(G[[s]]) >= 10) {
  #   CL<-1:5
  
```

```

# avgSil<-rep(NA, length(CL))
# for(cl in CL[2:length(CL)])
#   avgSil[cl]<-pam(X[G[[s]],], k=cl)$silinfo$avg.width
# barplot(avgSil, names.arg=CL, xlab="Number of clusters, K", ylab="Average silhouette", main=s)
# CL = which.max(avgSil)
# cl = pam(X[G[[s]],], k=CL)
# colG[[s]] = rainbow(CL)[cl$clustering]
# names(colG[[s]]) = G[[s]]
# } else {
#   colG[[s]] = rep("blue", length(G[[s]]))
#   names(colG[[s]]) = G[[s]]
# }
#
# if (length(H[[s]]) >= 10) {
#   CL<-1:5
#   avgSil<-rep(NA, length(CL))
#   for(cl in CL[2:length(CL)])
#     avgSil[cl]<-pam(X[H[[s]],], k=cl)$silinfo$avg.width
#   barplot(avgSil, names.arg=CL, xlab="Number of clusters, K", ylab="Average silhouette", main=s)
#   CL = which.max(avgSil)
#   cl = pam(X[H[[s]],], k=CL)
#   colH[[s]] = rainbow(CL)[cl$clustering]
#   names(colH[[s]]) = H[[s]]
# } else {
#   colH[[s]] = rep("blue", length(H[[s]]))
#   names(colH[[s]]) = H[[s]]
# }
}

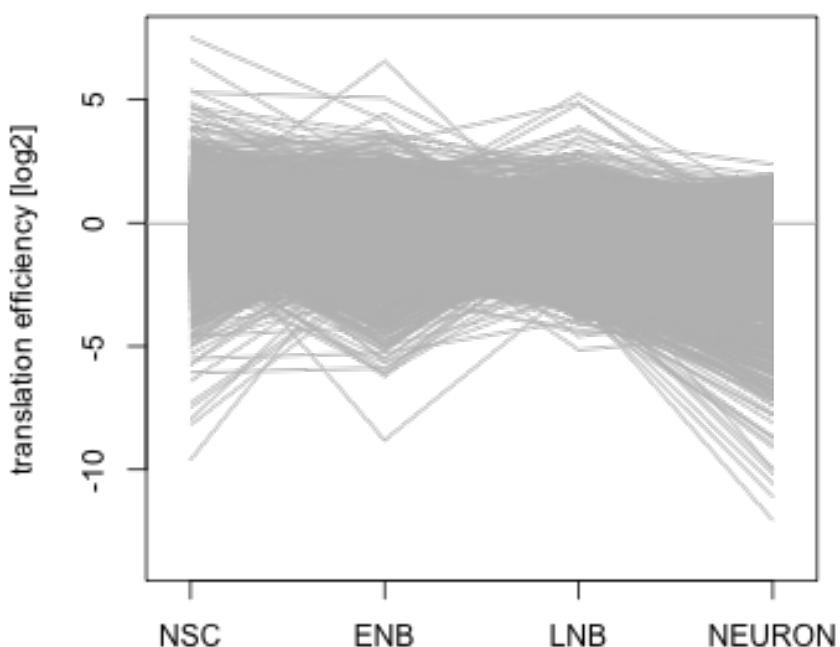
ylim = range(log2FC)
for (s in S) {
  load(file.path("result", "TranslationEfficiency", s, "res.rda"))

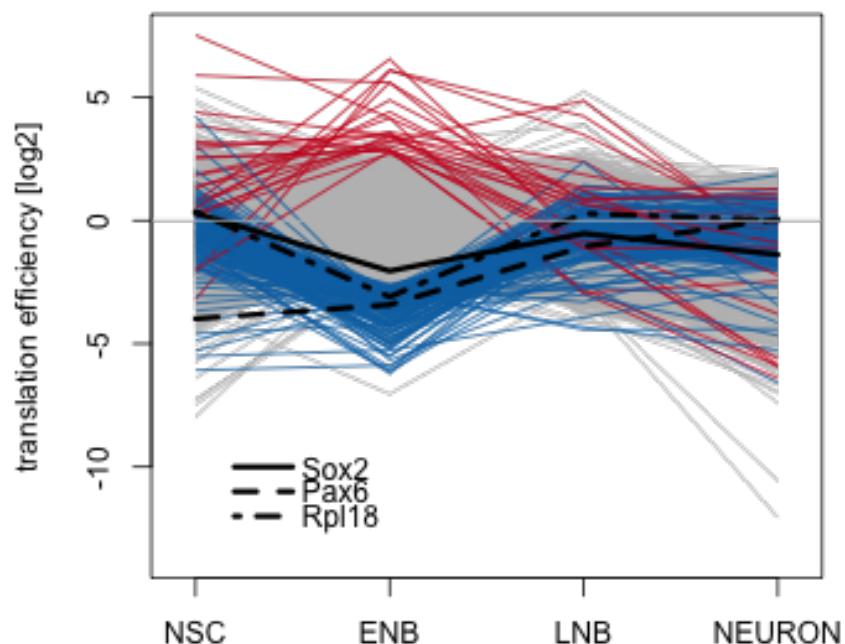
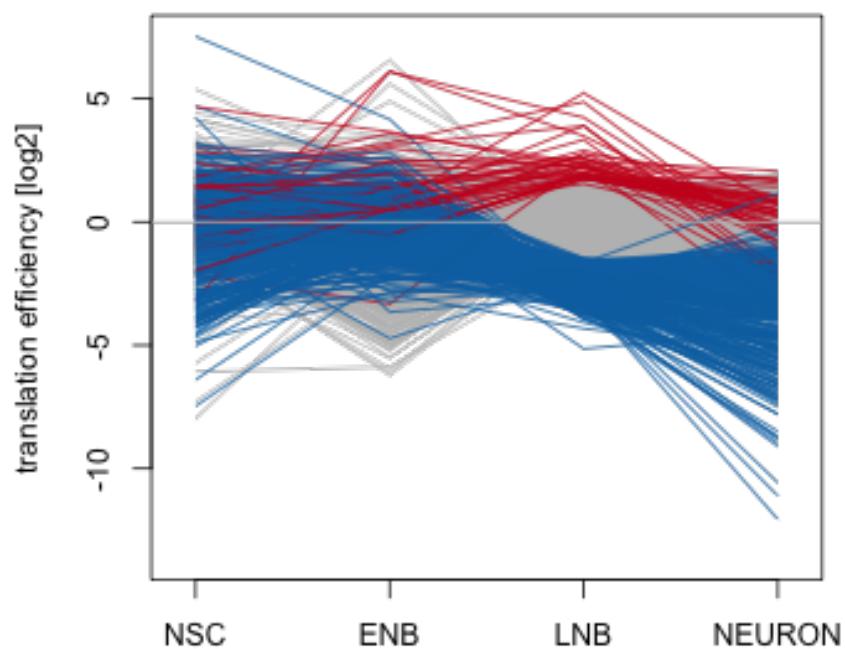
  plot(-100000, xlim=c(0.9, 4.1), ylim=ylim, xaxt="n",
       main=sprintf("enhanced or repressed translated in %s", s),
       xlab="", ylab="translation efficiency [log2]")
  axis(side=1, 1:4, colnames(log2total))
  col = rep("gray", nrow(log2FC))
  names(col) = row.names(log2FC)
  col[H[[s]]] = brewer.pal(4, "RdBu")[4]
  col[G[[s]]] = brewer.pal(4, "RdBu")[1]
  K = rep(1, nrow(log2FC))
  names(K) = row.names(log2FC)
  K[c(G[[s]], H[[s]])] = 2
  I = sample(row.names(res))
  I = I[order(K[I])]
  for (k in I) {
    lines(log2FC[k,], col=col[k])
  }
  if (s == "ENB") {
    if ("ENSMUSG00000074637" %in% rownames(log2FC)) {
      lines(log2FC["ENSMUSG00000074637",], col="black", lwd=3) # Sox2
    }
    if ("ENSMUSG00000027168" %in% rownames(log2FC)) {
      lines(log2FC["ENSMUSG00000027168",], col="black", lwd=3, lty="dashed") # Pax6
    }
    if ("ENSMUSG00000059070" %in% rownames(log2FC)) {
      lines(log2FC["ENSMUSG00000059070",], col="black", lwd=3, lty="dotdash") # Rpl18
    }
  }
}

```

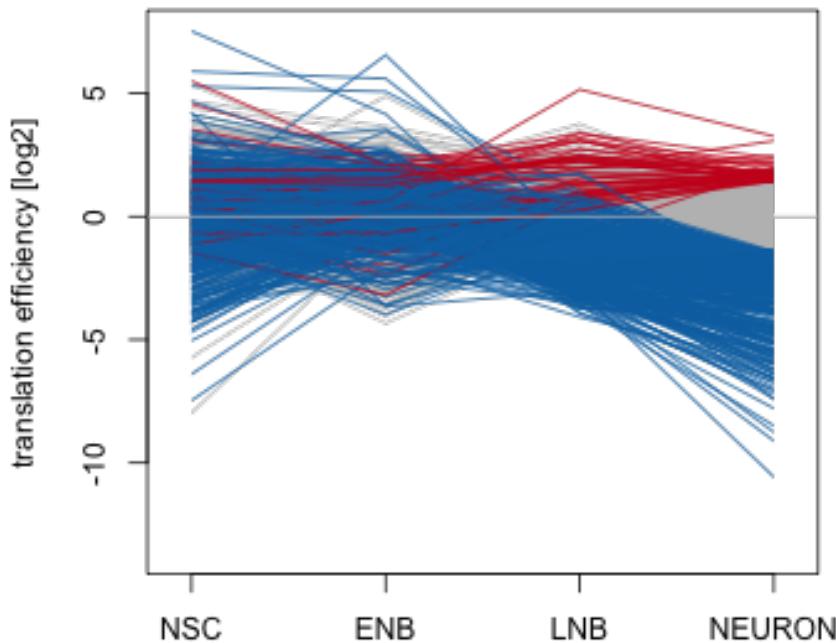
```
    }
  if (s == "ENB") {
    lines(c(1.2,1.5),c(-10,-10),col="black",lwd=3) # Sox2
    lines(c(1.2,1.5),c(-11,-11),col="black",lwd=3, lty="dashed") # Pax6
    lines(c(1.2,1.5),c(-12,-12),col="black",lwd=3, lty="dotdash") # Rpl18
    text(1.55,-10,"Sox2", adj = c(0,0.5))
    text(1.55,-11,"Pax6", adj = c(0,0.5))
    text(1.55,-12,"Rpl18", adj = c(0,0.5))
  }
}
abline(h=0,col="gray")
}
```

enhanced or repressed translated in NSC



enhanced or repressed translated in ENB**enhanced or repressed translated in LNB**

enhanced or repressed translated in NEURON



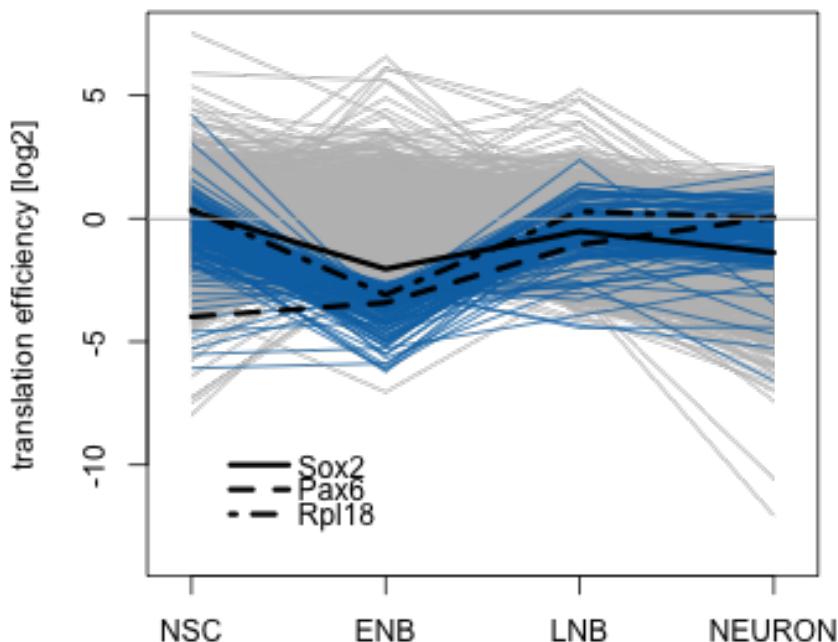
```

ylim = range(log2FC)
s = "ENB"
load(file.path("result", "TranslationEfficiency", s, "res.rda"))

plot(-100000,xlim=c(0.9,4.1), ylim=ylim,xaxt="n",
      main=sprintf("enhanced or repressed translated in %s",s),
      xlab="",ylab="translation efficiency [log2]")
axis(side=1,1:4,colnames(log2total))
col = rep("gray", nrow(log2FC))
names(col) = row.names(log2FC)
col[H[[s]]] = brewer.pal(4,"RdBu")[4]
#col[G[[s]]] = brewer.pal(4,"RdBu")[1]
K = rep(1, nrow(log2FC))
names(K) = row.names(log2FC)
K[c(H[[s]])] = 2
I = sample(row.names(res))
I = I[order(K[I])]
for (k in I) {
  lines(log2FC[k,],col=col[k])
}
lines(log2FC["ENSMUSG00000074637",],col="black",lwd=3) # Sox2
lines(log2FC["ENSMUSG00000027168",],col="black",lwd=3, lty="dashed") # Pax6
lines(log2FC["ENSMUSG00000059070",],col="black",lwd=3, lty="dotdash") # Rpl18
lines(c(1.2,1.5),c(-10,-10),col="black",lwd=3) # Sox2
lines(c(1.2,1.5),c(-11,-11),col="black",lwd=3, lty="dashed") # Pax6
lines(c(1.2,1.5),c(-12,-12),col="black",lwd=3, lty="dotdash") # Rpl18
text(1.55,-10,"Sox2", adj = c(0,0.5))
text(1.55,-11,"Pax6", adj = c(0,0.5))
text(1.55,-12,"Rpl18", adj = c(0,0.5))
abline(h=0,col="gray")

```

enhanced or repressed translated in ENB



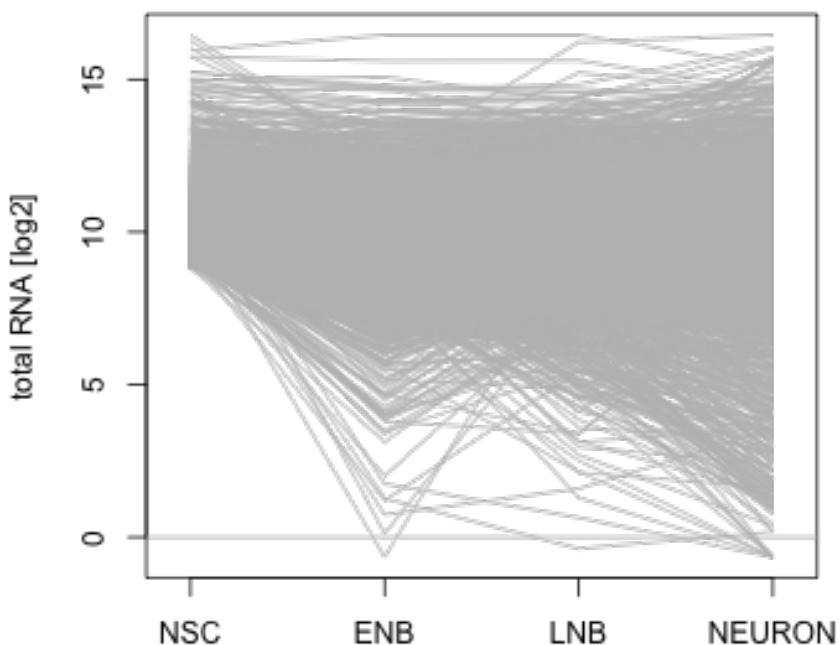
```

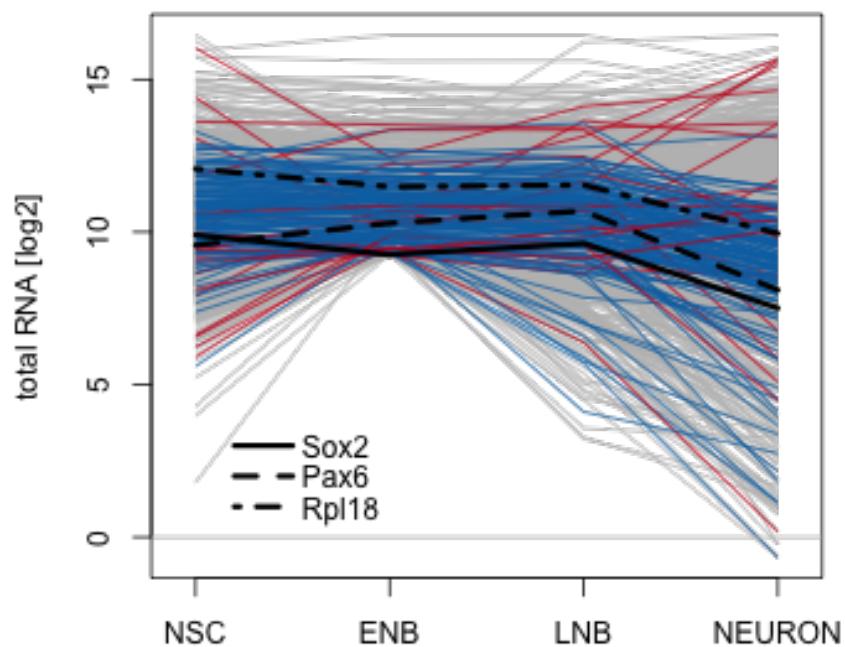
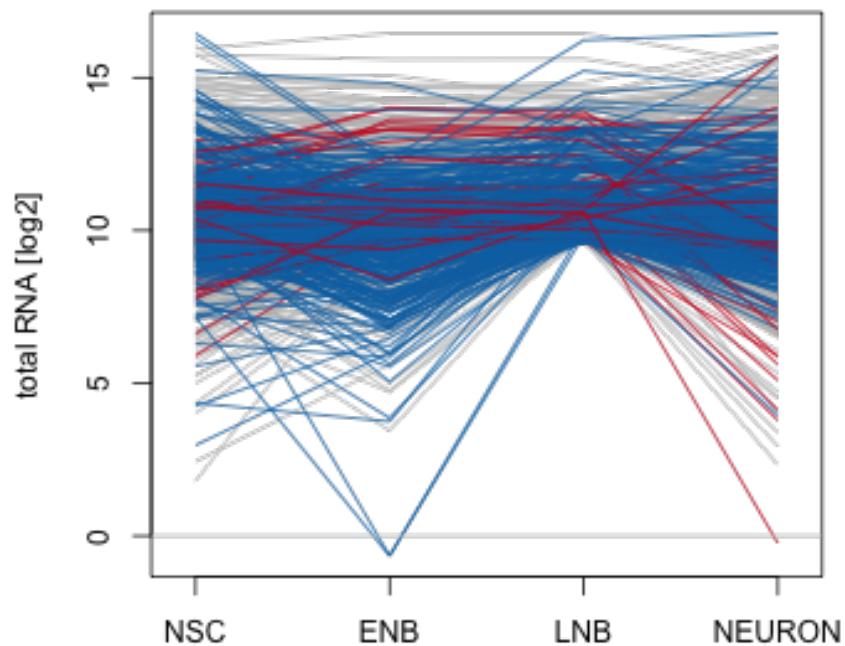
ylim = range(log2totalQ)
for (s in S) {
  load(file.path("result", "TranslationEfficiency", s, "res.rda"))

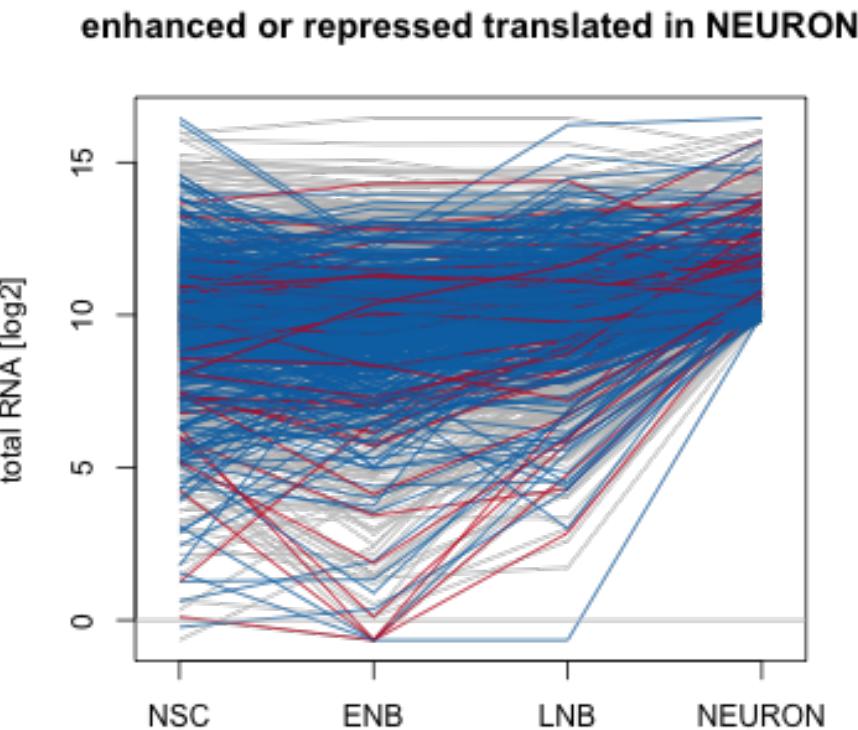
  plot(-100000, xlim=c(0.9,4.1), ylim=ylim, xaxt="n",
    main=sprintf("enhanced or repressed translated in %s",s),
    xlab="", ylab="total RNA [log2]")
  axis(side=1,1:4,colnames(log2total))
  col = rep("gray", nrow(log2FC))
  names(col) = row.names(log2FC)
  col[H[[s]]] = brewer.pal(4,"RdBu")[4]
  col[G[[s]]] = brewer.pal(4,"RdBu")[1]
  K = rep(1, nrow(log2FC))
  names(K) = row.names(log2FC)
  K[c(G[[s]],H[[s]])] = 2
  I = sample(row.names(res))
  I = I[order(K[I])]
  for (k in I) {
    lines(log2totalQ[k,],col=col[k])
  }
  if (s == "ENB") {
    if ("ENSMUSG00000074637" %in% rownames(log2totalQ)) {
      lines(log2totalQ["ENSMUSG00000074637",],col="black",lwd=3) # Sox2
    }
    if ("ENSMUSG00000027168" %in% rownames(log2totalQ)) {
      lines(log2totalQ["ENSMUSG00000027168",],col="black",lwd=3, lty="dashed") # Pax6
    }
    if ("ENSMUSG00000059070" %in% rownames(log2totalQ)) {
  
```

```
    lines(log2totalQ["ENSMUSG00000059070"], col="black", lwd=3, lty="dotdash") # Rpl18
  }
  if (s == "ENB") {
    lines(c(1.2,1.5),c(3,3),col="black",lwd=3) # Sox2
    lines(c(1.2,1.5),c(2,2),col="black",lwd=3, lty="dashed") # Pax6
    lines(c(1.2,1.5),c(1,1),col="black",lwd=3, lty="dotdash") # Rpl18
    text(1.55,3,"Sox2", adj = c(0,0.5))
    text(1.55,2,"Pax6", adj = c(0,0.5))
    text(1.55,1,"Rpl18", adj = c(0,0.5))
  }
}
abline(h=0,col="gray")
}
```

enhanced or repressed translated in NSC



enhanced or repressed translated in ENB**enhanced or repressed translated in LNB**

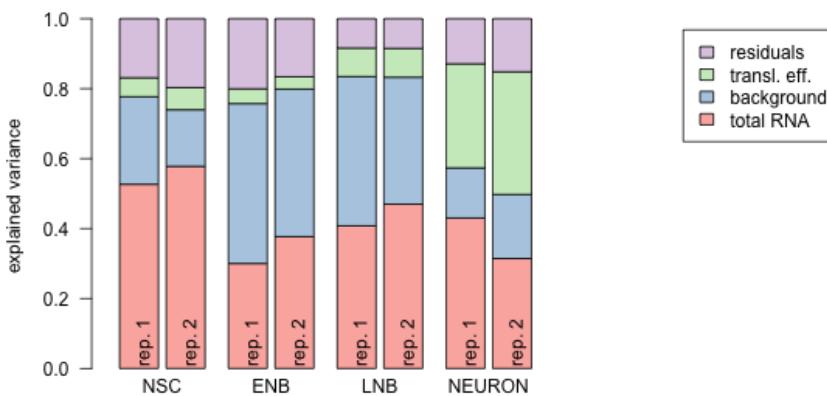


The explained variance is compared along the four differential stages.

```
AllExpVar = NULL
for (s in S) {
  load(file.path("result", "TranslationEfficiency", s, "ExpVar.rda"))
#  row.names(ExpVar) = paste0(s, "rep. ", 1:2)
  AllExpVar = rbind(AllExpVar, ExpVar)
}

row.names(AllExpVar) = NULL

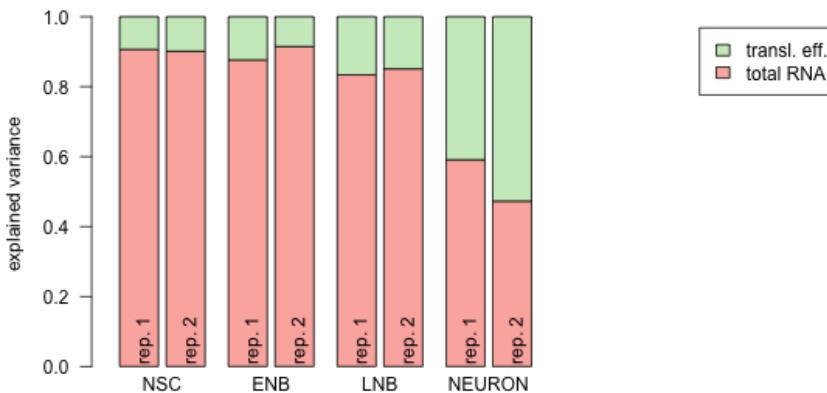
p = par(xpd = NA)
bp = barplot(t(AllExpVar), col=brewer.pal(4, "Pastel1"), las=2, space = c(0.0, 0.2, 0.6, 0.2, 0.6, 0.2,
text((bp[seq(1,8,by=2)]+bp[seq(2,8,by=2)])/2, y = -0.05, S)
text(bp[seq(1,8,by=2)], y = 0.01, "rep. 1", adj=c(0,0.8), srt=90)
text(bp[seq(2,8,by=2)], y = 0.01, "rep. 2", adj=c(0,0.8), srt=90)
```



```
par(p)
```

```
AllExpVar = AllExpVar[,c(1,3)]
s = rowSums(AllExpVar)
AllExpVar[,1] = AllExpVar[,1] / s
AllExpVar[,2] = AllExpVar[,2] / s

p = par(xpd = NA)
bp = barplot(t(AllExpVar), col=brewer.pal(4, "Pastel1")[c(1,3)], las=2, space = c(0.0, 0.2, 0.6, 0.2, 0
text((bp[seq(1,8,by=2)]+bp[seq(2,8,by=2)])/2, y = -0.05, S)
text(bp[seq(1,8,by=2)], y = 0.01, "rep. 1", adj=c(0,0.8), srt=90)
text(bp[seq(2,8,by=2)], y = 0.01, "rep. 2", adj=c(0,0.8), srt=90)
```



```
par(p)
```

3.6 Compare RNAseq and RiboTag data

Create an output directory.

```
d = file.path("result", "CompareRNaseqRiboTag", "")
dir.create(d, recursive = TRUE, showWarnings = FALSE)
```

```

mainmenu = file.path("../","..","mainmenu.html")

library(grid)
source(file.path("R","myballoonplot.R"))
load(file.path("data","ENSG2category.rda"))
source(file.path("R","doGSEA.R"))

RNAseq = RiboTag = list()
S = names(SamplesRNaseq)
for (s in S) {
  load(file.path("result","RNAseq",s,"res.rda"))
  RNAseq[[s]] = res
  load(file.path("result","RIBOseq",s,"res.rda"))
  RiboTag[[s]] = res
}

DFA = data.frame(ENSG = row.names(RNAseq[[1]]),symbol=RNAseq[[1]][,"symbol"],
                 stringsAsFactors = FALSE)
DF1 = data.frame(RNAseq[[1]][,c("baseMean","log2FoldChange","pvalue","padj","significant")],
                 stringsAsFactors = FALSE)
colnames(DF1) = paste(S[1],colnames(DF1),sep=".")
DF2 = data.frame(RNAseq[[2]][,c("baseMean","log2FoldChange","pvalue","padj","significant")],
                 stringsAsFactors = FALSE)
colnames(DF2) = paste(S[2],colnames(DF2),sep=".")
DF3 = data.frame(RNAseq[[3]][,c("baseMean","log2FoldChange","pvalue","padj","significant")],
                 stringsAsFactors = FALSE)
colnames(DF3) = paste(S[3],colnames(DF3),sep=".")
DF = cbind(DFA,DF1,DF2,DF3)
DF = DF[order(DF$ENSG),]
write.table(DF, file = file.path(d, "TableS2-RNaseq.txt"), sep="\t", row.names=FALSE, quote=FALSE)

DFA = data.frame(ENSG = row.names(RiboTag[[1]]),symbol=RiboTag[[1]][,"symbol"],
                 stringsAsFactors = FALSE)
DF1 = data.frame(RiboTag[[1]][,c("baseMean","log2FoldChange","pvalue","padj","significant")],
                 stringsAsFactors = FALSE)
colnames(DF1) = paste(S[1],colnames(DF1),sep=".")
DF2 = data.frame(RiboTag[[2]][,c("baseMean","log2FoldChange","pvalue","padj","significant")],
                 stringsAsFactors = FALSE)
colnames(DF2) = paste(S[2],colnames(DF2),sep=".")
DF3 = data.frame(RiboTag[[3]][,c("baseMean","log2FoldChange","pvalue","padj","significant")],
                 stringsAsFactors = FALSE)
colnames(DF3) = paste(S[3],colnames(DF3),sep=".")
DF = cbind(DFA,DF1,DF2,DF3)
DF = DF[order(DF$ENSG),]
write.table(DF, file = file.path(d, "TableS3-RiboTag.txt"), sep="\t", row.names=FALSE, quote=FALSE)

GSEA = list(up = list(
  both = list(),
  RNAspecific = list(),
  RiboTagSpecific = list()
))
GSEA$down = GSEA$up
Nhits = list()
for (s in S) {
  Nhits[[s]] = matrix(NA_integer_, nrow=2, ncol = 3,
                     dimnames=list(c("up","down"),
                                   c("RNAspecific","both","RiboTagSpecific")))
}

```

```

I = which(log2(RNAseq[[s]]$baseMean) > 2 | log2(RiboTag[[s]]$baseMean) > 2)
R = RNAseq[[s]][I,]
RT = RiboTag[[s]][I,]
bg = row.names(R)

J = which(R$padj <= 0.1 & R$log2FoldChange > 0 &
           RT$padj <= 0.1 & RT$log2FoldChange > 0)
fgUp = bg[J]
K = which(R$padj <= 0.1 & R$log2FoldChange < 0 &
           RT$padj <= 0.1 & RT$log2FoldChange < 0)
fgDown = bg[K]
GSEA$up$both[[s]] = doGSEA(fgUp,bg,ENSG2category, k=3)
GSEA$down$both[[s]] = doGSEA(fgDown,bg,ENSG2category, k=3)
Nhits[[s]]["up","both"] = length(J)
Nhits[[s]]["down","both"] = length(K)

J2 = which(R$padj <= 0.1 & R$log2FoldChange > 0)
J2 = setdiff(J2, J)
fgUp = bg[J2]
K2 = which(R$padj <= 0.1 & R$log2FoldChange < 0)
K2 = setdiff(K2, K)
fgDown = bg[K2]
GSEA$up$RNAspecific[[s]] = doGSEA(fgUp,bg,ENSG2category, k=3)
GSEA$down$RNAspecific[[s]] = doGSEA(fgDown,bg,ENSG2category, k=3)
Nhits[[s]]["up","RNAspecific"] = length(J2)
Nhits[[s]]["down","RNAspecific"] = length(K2)

J3 = which(RT$padj <= 0.1 & RT$log2FoldChange > 0)
J3 = setdiff(J3, J)
fgUp = bg[J3]
K3 = which(RT$padj <= 0.1 & RT$log2FoldChange < 0)
K3 = setdiff(K3, K)
fgDown = bg[K3]
GSEA$up$RiboTagSpecific[[s]] = doGSEA(fgUp,bg,ENSG2category, k=3)
GSEA$down$RiboTagSpecific[[s]] = doGSEA(fgDown,bg,ENSG2category, k=3)
Nhits[[s]]["up","RiboTagSpecific"] = length(J3)
Nhits[[s]]["down","RiboTagSpecific"] = length(K3)
}

```

3.7 Sequence motifs

Create an output directory.

```

d = file.path("result","Motifs","")
dir.create(d, recursive = TRUE, showWarnings = FALSE)
mainmenu = file.path("../..","mainmenu.html")

S = names(SamplesTranslationEfficiency)
TransEff = list()
for (s in S) {
  load(file.path("result","TranslationEfficiency",s,"res.rda"))
  TransEff[[s]] = res
}

alphabet = function() {
  list(A = c("A", "R", "N", "D", "C", "E", "Q", "G", "H", "I",
            "L", "K", "M", "F", "P", "S", "T", "W", "Y", "V"),

```

```

        N = c("T", "C", "G", "A"))
}

getKMers <- function(seq, K, toMatrix, n) {
  SP = strsplit(as.character(seq), split="")
  tmp = sapply(SP, function(x) {
    res = rep("", length(x)-K+1)
    for (i in seq_len(K)) {
      res = paste(res,x[seq_len(length(x)-K+1)+i-1],sep="")
    }
    res
  })
  if (toMatrix) {
    a = alphabet() [[n]]
    A = a
    if (K > 1) {
      for (k in 2:K) {
        A = paste0(rep(a, each=length(A)),rep(A, times=length(a)))
      }
    }
    tmp = t(sapply(tmp, function(x) { table(factor(x, levels=A)) }))
  }
  tmp
}

load("data/Seq.rda")

Seq = Seq[1:3]
Seq = lapply(Seq, function(x) {
  I = which(x$gene_biotype == "protein_coding" & x$transcript_biotype == "protein_coding")
  x = x[I,]
  x
})

Seq = lapply(Seq, function(x) {
  n = nchar(x[[1]])
  I = order(n, decreasing = TRUE)
  I = I[!duplicated(x$ensembl_gene_id[I])]
  x = x[I,]
  x
})

```

The samples are annotated.

```

K = 6
tests = list()
for (s in names(Seq)) {
  tests[[s]] = list()
  for (ct in S) {
#    load(file.path("result", "controledTranslation", ct, "res.rda"))
    Seq2 = lapply(Seq, function(x) {
      x = x[which(x[, "ensembl_gene_id"] %in% row.names(TransEff[[ct]])),]
      x
    })
    Seq2 = Seq[[s]][which(Seq[[s]][, "ensembl_gene_id"] %in%
                           row.names(TransEff[[ct]])),]
    seq = Seq2[,1]
    seq[seq == "Sequence unavailable"] = ""
    I = which(nchar(seq) < K)
  }
}

```

```

seq[I] = ""
I = which(nchar(seq) >= K)
X = getKMers(seq[I], K = K, toMatrix = TRUE, n = "N")

GN = factor(Seq2[I, "ensembl_gene_id"], levels=row.names(TransEff[[ct]]))
KM = matrix(nrow=nrow(TransEff[[ct]]), ncol=ncol(X))
row.names(KM) = levels(GN)
colnames(KM) = colnames(X)
for (i in seq_len(ncol(KM))) {
  KM[,i] = tapply(X[,i], GN, sum, na.rm=TRUE)
}
KM[KM>1] = 1
for (upDown in c("up", "down")) {
  c = paste0(ct, "_", upDown)
  tests[[s]][[c]] = data.frame(pval = rep(NA, ncol(X)),
                                padj = NA, est = NA, p=NA)
  row.names(tests[[s]][[c]]) = colnames(X)

  if (upDown == "up") {
    Sel = row.names(TransEff[[ct]])[which(TransEff[[ct]]$padj <= 0.1 & TransEff[[ct]]$log2FoldChange
                                          !is.na(KM[,1]))]
  } else {
    Sel = row.names(TransEff[[ct]])[which(TransEff[[ct]]$padj <= 0.1 & TransEff[[ct]]$log2FoldChange
                                          !is.na(KM[,1]))]
  }
  BG = row.names(TransEff[[ct]])[which(is.finite(TransEff[[ct]]$padj) & !is.na(KM[,1]))]
  p = median(apply(KM[Sel,], 1, sum)) / median(apply(KM[setdiff(BG, Sel),], 1, sum))
  M = apply(KM[setdiff(BG, Sel),], 2, sum)
  m = apply(KM[Sel,], 2, sum)
  n = length(Sel)
  N = length(BG)
  Tab = matrix(0, nrow=2, ncol=2)
  for (i in seq_len(ncol(KM))) {
    Tab[2,2] = m[i]
    Tab[1,2] = M[i]
    Tab[2,1] = n-m[i]
    Tab[1,1] = N-M[i]-n
    test = fisher.test(Tab, or = p)
    # test = binom.test(x[i], n, p=p[i])
    tests[[s]][[c]]$pval[i] = test$p.value
    tests[[s]][[c]]$est[i] = test$estimate
  }
  tests[[s]][[c]]$padj[] = p.adjust(tests[[s]][[c]]$pval[], method = "BH")
  tests[[s]][[c]]$p = p
  cat("s=", s, " ct=", ct, " d=", upDown, " FDR20%=", sum(tests[[s]][[c]]$padj < 0.2), " FDR5%=", sum(tests[[s]][[c]]$padj < 0.05), "\n")
}
}

save(tests, file=file.path(d, "tests.rda"))

for (s in names(Seq)) {
  print(sapply(tests[[s]], function(x) { sum(x$padj <= 0.1) }))
}

##      NSC_up     NSC_down      ENB_up     ENB_down      LNB_up     LNB_down     NEURON_up
##          0           1           0           1           0           3           0
## NEURON_down
##          4

```

```

##      NSC_up    NSC_down     ENB_up    ENB_down     LNB_up    LNB_down  NEURON_up
##      0          0          0          9          0          2          0
## NEURON_down
##      6
##      NSC_up    NSC_down     ENB_up    ENB_down     LNB_up    LNB_down  NEURON_up
##      0          0          1          34          0          0          0
## NEURON_down
##      1064

M = matrix("", ncol = length(Seq), nrow=length(tests[[1]]))
row.names(M) = names(tests[[1]])
colnames(M) = names(Seq)
or = data.frame(pval=c(), padj=c(), est=c(), mRNA=c(), celltype=c())
for (s in names(Seq)) {
  for (ct in names(tests[[1]])) {
    M[ct,s] = sprintf("volcano_%s_%s.png", s, ct)
    png(file.path(d, M[ct,s]))
    x = log2(tests[[s]][[ct]]$est/tests[[s]][[ct]]$p)
    y = -log2(tests[[s]][[ct]]$padj)
    x[x > 2.5] = 2.5
    x[x < -2.5] = -2.5
    y[y > 10] = 10
    y[y < -10] = -10
    plot(x,y,col=ifelse(tests[[s]][[ct]]$padj < 0.1,"red","gray"), xlim=c(-2.5,2.5), ylim=c(0,10),
          main = sprintf("%s %s",s, ct))
    dev.off()
    I = which(tests[[s]][[ct]]$padj < 0.1)
    if (length(I) > 0) {
      x = tests[[s]][[ct]][I,]
      x$est = x$est / x$p
      x$p = NULL
      x$motif = row.names(x)
      x$mRNA = s
      x$celltype = ct
      or = rbind(or, x)
    }
  }
}
colnames(or)[3] = "oddsRatio"
row.names(or) = NULL
file.copy(system.file("images", "hwriter.css", package="hwriter"),
          file.path(d, "hwriter.css"))

## [1] TRUE

page = openPage(file.path(d, "index_motifs.html"), link.css="hwriter.css")
hwrite("Overrepresented motifs", heading=1, page=page)
write.table(or, file=file.path(d, "motifs.txt"), sep="\t", quote=FALSE, row.names=FALSE)
hwrite("[Download txt-file]", link="motifs.txt", page=page, br=TRUE)
hwrite(or, page=page)
closePage(page, splash = FALSE)
page = openPage(file.path(d, "index_volcano.html"), link.css="hwriter.css")
hwrite("Volcano plots", heading=1, page=page)
hwrite("[Download txt-file motifs]", link="motifs.txt", page=page, br=TRUE)
hwrite("[Table of motifs]", link="index_motifs.html", page=page, br=TRUE)
hwriteImage(M, page=page)
closePage(page, splash = FALSE)

p = par(xpd = NA)

```

```

TOPmRNAs = readLines(file.path("data/knownTOPmRNAs.txt"))
for (ct in S) {
  load(file.path("result","TranslationEfficiency",ct,"res.rda"))

  sig = res[, "significant"]
  reg = ifelse(res[, "log2FoldChange"] > 0, "up", "down")
  reg = paste0(reg, " (", sig, ")")
  reg[sig == ""] = "notRegulated"
  reg = factor(reg, levels=c("down (**)", "down (*)", "notRegulated", "up (*)", "up (**)"))
  names(reg) = res$symbol
  png(file.path(d, sprintf("barplot_TOP_%s.png",ct)))
  t = table(reg[res$symbol %in% TOPmRNAs])
  bp = barplot(t,main=sprintf("known TOP mRNAs in %s",ct),ylim=c(0,max(t)*1.05))
  text(bp[,1],t+max(t)*0.03,t)
  dev.off()
  png(file.path(d, sprintf("barplot_all_%s.png",ct)))
  t = table(reg)
  bp = barplot(t,main=sprintf("all mRNAs in %s",ct),ylim=c(0,max(t)*1.05))
  text(bp[,1],t+max(t)*0.03,t)
  dev.off()
  r = reg[res$symbol %in% TOPmRNAs]
  r = r[order(r, names(r))]
  write.table(r, file=file.path(d, sprintf("table_TOP_%s.txt",ct)),sep="\t", col.names=FALSE)
}
par(p)

page = openPage(file.path(d, "index_TOP.html"), link.css="hwriter.css")
hwrite("TOP mRNAs", heading=1, page=page)

M = matrix(S, nr=length(S), ncol=1)
L = matrix(sprintf("table_TOP_%s.txt",S), nr=length(S), ncol=1)
hwrite(M, link = L, page=page, br=TRUE)

IMG = cbind(sprintf("barplot_TOP_%s.png",S),sprintf("barplot_all_%s.png",S))
M = hwriteImage(IMG, table=FALSE)
row.names(M) = S
colnames(M) = c("TOP mRNAs", "all mRNAs")
hwrite(M, page=page)
closePage(page, splash = FALSE)

```