

Epistatic Nested Effects Models

Inferring mixed epistasis from indirect measurements of knockout screens.

Madeline Diekmann & Martin Pirkl

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This package is an extension of the classic Nested Effects Models provided in package *nem*. Nested Effects Models is a pathway reconstruction method, which takes into account effects of downstream genes. Those effects are observed for every knockout of a pathway gene, and the nested structure of observed effects can then be used to reconstruct the pathway structure. However, classic Nested Effects Models do not account for double knockouts. In this package *epiNEM*, one additional layer of complexity is added. For every two genes, acting on one gene together, the relationship is evaluated and added to the model as a logic gate. Genetic relationships are represented by the logics OR (no relationship), AND (functional overlap), NOT (masking or inhibiting) and XOR (mutual prevention from acting on gene C).

Loading epiNEM

```
## install.packages("devtools", verbose = F, quiet = T)

library(devtools)

## install_github("cbg-ethz/epiNEM", quiet = T)

library(epiNEM)

## Loading required package: BoolNet
## Loading required package: e1071
## Loading required package: gtools
##
## Attaching package: 'gtools'
## The following object is masked from 'package:e1071':
##
##   permutations
## Loading required package: igraph
##
## Attaching package: 'igraph'
## The following object is masked from 'package:gtools':
##
##   permute
## The following objects are masked from 'package:stats':
##
##   decompose, spectrum
## The following object is masked from 'package:base':
##
##   union
```

Simulations

We compare epiNEM to several network inference methods.

```
library(bnem) # install_github("MartinFXP/B-NEM/package")

## Loading required package: CellNOptR
## Loading required package: RBGL
## Loading required package: graph
## Loading required package: BiocGenerics
## Loading required package: parallel
##
## Attaching package: 'BiocGenerics'
## The following objects are masked from 'package:parallel':
##
##   clusterApply, clusterApplyLB, clusterCall, clusterEvalQ, clusterExport,
##   clusterMap, parApply, parCapply, parLapply, parLapplyLB, parRapply,
##   parSapply, parSapplyLB
## The following objects are masked from 'package:igraph':
##
##   normalize, union
## The following objects are masked from 'package:stats':
##
##   IQR, mad, xtabs
## The following objects are masked from 'package:base':
##
##   Filter, Find, Map, Position, Reduce, anyDuplicated, append, as.data.frame,
##   cbind, colnames, do.call, duplicated, eval, evalq, get, grep, grepl,
##   intersect, is.unsorted, lapply, lengths, mapapply, match, mget, order, paste,
##   pmax, pmax.int, pmin, pmin.int, rank, rbind, rownames, sapply, setdiff,
##   sort, table, tapply, union, unique, unsplit, which, which.max, which.min
##
## Attaching package: 'graph'
## The following objects are masked from 'package:igraph':
##
##   degree, edges, intersection
##
## Attaching package: 'RBGL'
## The following objects are masked from 'package:igraph':
##
##   bfs, dfs, transitivity
## The following object is masked from 'package:e1071':
##
##   extractPath
## Loading required package: hash
## hash-2.2.6 provided by Decision Patterns
## Loading required package: ggplot2
## Warning: package 'ggplot2' was built under R version 3.3.2
## Loading required package: RCurl
## Loading required package: bitops
## Loading required package: Rgraphviz
## Loading required package: grid
## Loading required package: XML
## Warning: package 'XML' was built under R version 3.3.2
```

```
##
## Attaching package: 'XML'
## The following object is masked from 'package:graph':
##
##     addNode
## Loading required package: nem
##
## Attaching package: 'nem'
## The following object is masked from 'package:RBGL':
##
##     transitive.closure
## Loading required package: matrixStats
## matrixStats v0.51.0 (2016-10-08) successfully loaded. See ?matrixStats for help.
## Loading required package: snowfall
## Loading required package: snow
##
## Attaching package: 'snow'
## The following objects are masked from 'package:BiocGenerics':
##
##     clusterApply, clusterApplyLB, clusterCall, clusterEvalQ, clusterExport,
##     clusterMap, clusterSplit, parApply, parCapply, parLapply, parRapply,
##     parSapply
## The following objects are masked from 'package:parallel':
##
##     clusterApply, clusterApplyLB, clusterCall, clusterEvalQ, clusterExport,
##     clusterMap, clusterSplit, makeCluster, parApply, parCapply, parLapply,
##     parRapply, parSapply, splitIndices, stopCluster
## Loading required package: latticeExtra
## Loading required package: lattice
## Loading required package: RColorBrewer
##
## Attaching package: 'latticeExtra'
## The following object is masked from 'package:ggplot2':
##
##     layer
library(nem)

library(minet)

library(pcalg)
```

```
runs <- 100

noiselvls <- c(0.01, 0.025, 0.05, 0.1, 0.2, 0.3, 0.4, 0.5)

random <- list(FPrate = 0.1, FNrate = noiselvls, single = 4, double = 1, reporters = 100, replicates = 3)

spec <- sens <- logics <- array(0, dim = c(2, runs, length(noiselvls)))

sens2 <- spec2 <- time <- array(0, dim = c(5, runs, length(noiselvls)))

do <- c("n", "p", "a")
```

```

do <- c("e", "b", do)

popSize <- 100

maxTime <- F

forcelogic <- T

epinemsearch <- "greedy"

nIterations <- 3

bnemsearch <- "genetic"

parallel <- NULL

logicgate <- matrix("", runs, length(noiselvls))

edgenr <- matrix(0, runs, length(noiselvls))

## for (i in 1:runs) {

##     print(paste("run ", i, sep = ""))

##     for (j in 1:length(noiselvls)) {

##         print(paste("noiselvl ", j, sep = ""))

##         topology <- CreateTopology(random$single, random$double, force = forcelogic)

##         topology <- unlist(unique(topology), recursive = FALSE)

##         extTopology <- ExtendTopology(topology$model, random$reporters)

##         sortedData <- GenerateData(topology$model, extTopology, random$FPrate, random$FNrate[j], ran

##         logicgate[i, j] <- paste(topology$logics, collapse = "_")

##         edgenr[i, j] <- sum(topology$origModel == 1)

##         if ("e" %in% do) {
##             print("epiNEM")

##             start <- Sys.time()
##             TriplModel <- epiNEM(filename = sortedData, method = epinemsearch, nIterations = nIterat
##             time[1, i, j] <- difftime(Sys.time(), start, units = "secs")
##             print(time[1, i, j])

##             tp <- sum(topology$model == 1 & TriplModel$model == 1)
##             tn <- sum(topology$model == 0 & TriplModel$model == 0)
##             fp <- sum(topology$model == 0 & TriplModel$model == 1)
##             fn <- sum(topology$model == 1 & TriplModel$model == 0)
##             sens[1, i, j] <- tp/(tp+fn)

```

```

## spec[1, i, j] <- tn/(tn+fp)
## tp <- sum(topology$origModel == 1 & TriplModel$origModel == 1)
## tn <- sum(topology$origModel == 0 & TriplModel$origModel == 0)
## fp <- sum(topology$origModel == 0 & TriplModel$origModel == 1)
## fn <- sum(topology$origModel == 1 & TriplModel$origModel == 0)
## sens2[1, i, j] <- tp/(tp+fn)
## spec2[1, i, j] <- tn/(tn+fp)
## tp <- 0
## for (k in 1:length(topology$column)) {
##   for (l in 1:length(TriplModel$column)) {
##     if (topology$column[k] == TriplModel$column[l]) {
##       if (topology$logics[k] %in% TriplModel$logics[l]) {
##         tp <- tp + 1
##       }
##     }
##   }
## }
## logics[1, i, j] <- tp/(length(topology$logics) + length(TriplModel$logics) - tp)
## print(sens[1, i, j])
## print(spec[1, i, j])
## print(sens2[1, i, j])
## print(spec2[1, i, j])
## print(logics[1, i, j])

## }

## if ("b" %in% do) {
##   print("B-NEM")

##   gtn <- epi2bg(topology)

##   fc <- cbind(Ctrl_vs_S = -1, epi2bg(sortedData))*(-1)

##   bnemnoise <- sample(1:nrow(fc), floor(nrow(fc)*random$FNrate[j]))

##   fc[bnemnoise, 1] <- 0

##   ers <- t(topology$model)*(-1)
##   colnames(ers) <- paste("S_vs_S_", gsub("\\.", "_", colnames(ers)), sep = "")
##   ers <- cbind(Ctrl_vs_S = 1, ers)
##   ers <- ers[, order(colnames(ers))]

##   CNOList <- dummyCNOList(stimuli = "S", inhibitors = LETTERS[1:random$single], maxStim = 1)

##   parents <- unique(unlist(strsplit(colnames(sortedData)[grep("\\.", colnames(sortedData))], "\\."))

##   nodes <- unique(colnames(sortedData)[-grep("\\.", colnames(sortedData))])

##   child <- nodes[-which(nodes %in% parents)]

##   sijMatrix <- NULL
##   for (k in LETTERS[1:random$single]) {
##     sijMatrix <- rbind(sijMatrix, c("S", "1", k))#, c("S", "-1", k)) # bnem can set a priori

```

```

##           for (l in LETTERS[1:random$single]) {
##               if (k %in% l) { next() }
##               if (k %in% parents) {
##                   sifMatrix<- rbind(sifMatrix, c(k, "1", l), c(k, "-1", l))
##               } else {
##                   sifMatrix<- rbind(sifMatrix, c(k, "1", l))
##               }
##
## randfile <- paste("pkn_", as.numeric(Sys.time()), sep = "")
## write.table(sifMatrix, file = randfile, sep = "\t",
##             row.names = FALSE, col.names = FALSE, quote = FALSE)
## PKN <- readSIF(randfile)
## unlink(randfile)
##
## model <- preprocessing(CNOList, PKN)
##
## initBstring <- absorption(rep(1, length(model$reacID)), model)
##
## if (maxTime) { maxTime2 <- time[1, i, j] } else { maxTime2 <- Inf }
##
## start <- Sys.time()
## bga <- bnem(search = bnemsearch,
##             fc=fc,
##             CNOList=CNOList,
##             model=model,
##             initBstring=initBstring,
##             draw = F,
##             verbose = F,
##             popSize = popSize,
##             maxTime = maxTime2,
##             parallel = parallel
##             )
## time[2, i, j] <- difftime(Sys.time(), start, units = "secs")
## print(time[2, i, j])
##
## ers2 <- computeFc(CNOList, t(simulateStatesRecursive(CNOList, model, bga$bString)))
## ers2 <- ers2[, unique(colnames(fc))]
## ers2 <- ers2[, order(colnames(ers2))]
##
## tp <- sum(ers == -1 & ers2 == -1)
## tn <- sum(ers == 0 & ers2 == 0)
## fn <- sum(ers == -1 & ers2 == 0)
## fp <- sum(ers == 0 & ers2 == -1)
## sens[2, i, j] <- tp/(tp+fn)
## spec[2, i, j] <- tn/(tn+fp)
## gtn2 <- abs(dnf2adj(gtn))
## if (length(grep("S", rownames(gtn2))) > 0) {
##     gtn2 <- gtn2[-grep("S", rownames(gtn2)), -grep("S", colnames(gtn2))]
## }
## gtn2 <- gtn2[order(rownames(gtn2)), order(colnames(gtn2))]
## res <- abs(dnf2adj(bga$graph))
## if (length(grep("S", rownames(res))) > 0) {
##     res <- as.matrix(res[-grep("S", rownames(res)), -grep("S", colnames(res))])

```

```

##      }
##      if (dim(res)[1] == 1) {
##          colnames(res) <- rownames(res) <- gsub(".*=", "", bga$graph)
##      } else {
##          res <- res[order(rownames(res)), order(colnames(res))]
##      }
##      if (nrow(res) < nrow(gtn2)) {
##          res2 <- rbind(cbind(res, matrix(0, nrow(res), nrow(gtn2) - nrow(res))), matrix(0, nrow(res), nrow(gtn2) - nrow(res)))
##          colnames(res2)[(ncol(res)+1):ncol(res2)] <- colnames(gtn2)[which(!(colnames(gtn2) %in% colnames(res2)))]
##          rownames(res2)[(nrow(res)+1):nrow(res2)] <- rownames(gtn2)[which(!(rownames(gtn2) %in% rownames(res2)))]
##          res2 <- res2[order(rownames(res2)), order(colnames(res2))]
##          res <- res2
##      }
##      diag(gtn2) <- diag(res) <- 0
##      tp <- sum(gtn2 == 1 & res == 1)
##      tn <- sum(gtn2 == 0 & res == 0)
##      fn <- sum(gtn2 == 1 & res == 0)
##      fp <- sum(gtn2 == 0 & res == 1)
##      sens2[2, i, j] <- tp/(tp+fn)
##      spec2[2, i, j] <- tn/(tn+fp)
##      tp <- sum(bga$graph %in% gtn)
##      logics[2, i, j] <- tp/(length(gtn) + length(bga$graph) - tp) # (tp/(tp+fn) + tn/(tn+fp))
##      print(sens[2, i, j])
##      print(spec[2, i, j])
##      print(sens2[2, i, j])
##      print(spec2[2, i, j])
##      print(logics[2, i, j])

##      print(bga$graph)
##      print(gtn)

##  }

##  if (any(c("n", "p", "a") %in% do)) {

##      reddata <- sortedData[, -grep("\\.", colnames(sortedData))]
##      gtnadj <- topology$origModel
##      gtnadj <- gtnadj[order(apply(gtnadj, 1, sum), decreasing = T), order(apply(gtnadj, 2, sum))]
##      gtnadj[lower.tri(gtnadj)] <- gtnadj[upper.tri(gtnadj)]
##      gtnadj <- gtnadj[order(rownames(gtnadj)), order(colnames(gtnadj))]
##      eadj <- topology$origModel
##      eadj <- eadj[order(rownames(eadj)), order(colnames(eadj))]
##      reddata2 <- matrix(0, nrow(reddata)*random$replicates, length(unique(colnames(reddata))))
##      for (k in 1:length(unique(colnames(reddata)))) {
##          reddata2[, k] <- as.vector(reddata[, which(colnames(reddata) %in% unique(colnames(reddata)[k]))])
##      }
##      colnames(reddata2) <- unique(colnames(reddata))

##  }

##  if ("n" %in% do) {
##      print("NEM")

```

```

##      start <- Sys.time()
##      if (epinemsearch %in% "greedy") {
##          nemres <- nem(reddata, inference = "nem.greedy")
##      } else {
##          nemres <- nem(reddata, inference = "search")
##      }
##      nadj <- transitive.reduction(graph2adj(nemres$graph))
##      time[3, i, j] <- difftime(Sys.time(), start, units = "secs")
##      print(time[3, i, j])

##      tp <- sum(eadj == 1 & nadj == 1)
##      tn <- sum(eadj == 0 & nadj == 0)
##      fp <- sum(eadj == 0 & nadj == 1)
##      fn <- sum(eadj == 1 & nadj == 0)
##      sens2[3, i, j] <- tp/(tp+fn)
##      spec2[3, i, j] <- tn/(tn+fp)
##      print(sens2[3, i, j])
##      print(spec2[3, i, j])

##      }

##      if ("p" %in% do) {
##          print("PCalg")

##          start <- Sys.time()
##          pc.fit <- pc(suffStat = list(C = cor(reddata2), n = nrow(reddata2)),
##                      indepTest = gaussCItest, ## indep.test: partial correlations
##                      alpha=0.05, labels = colnames(reddata2), verbose = F)
##          pcadj <- graph2adj(pc.fit@graph)
##          time[4, i, j] <- difftime(Sys.time(), start, units = "secs")
##          print(time[4, i, j])

##          tp <- sum(gtnadj == 1 & pcadj == 1)
##          tn <- sum(gtnadj == 0 & pcadj == 0)
##          fp <- sum(gtnadj == 0 & pcadj == 1)
##          fn <- sum(gtnadj == 1 & pcadj == 0)
##          sens2[4, i, j] <- tp/(tp+fn)
##          spec2[4, i, j] <- tn/(tn+fp)
##          print(sens2[4, i, j])
##          print(spec2[4, i, j])

##          }

##      if ("a" %in% do) {
##          print("Aracne")

##          start <- Sys.time()
##          ares <- build.mim(reddata2)
##          ares <- aracne(ares)
##          ares <- disc(ares, 0)
##          ares <- ares[order(rownames(ares)), order(colnames(ares))]
##          nas <- which(is.na(ares) == T)
##          ares[nas] <- 0

```



```
##          diag(ares) <- 0
##          time[5, i, j] <- difftime(Sys.time(), start, units = "secs")
##          print(time[5, i, j])

##          tp <- sum(gtnadj == 1 & ares == 1)
##          tn <- sum(gtnadj == 0 & ares == 0)
##          fp <- sum(gtnadj == 0 & ares == 1)
##          fn <- sum(gtnadj == 1 & ares == 0)
##          sens2[5, i, j] <- tp/(tp+fn)
##          spec2[5, i, j] <- tn/(tn+fp)
##          print(sens2[5, i, j])
##          print(spec2[5, i, j])

##      }

##  }

## }
```

```
data(sim)

colvec <- c(rep("orange", length(noiselvls)), rep("blue", length(noiselvls)), rep("darkgreen", length(noiselvls)))

acc <- (sens + spec)/2

acc2 <- (sens2 + spec2)/2

m <- rbind(c(1,1), c(2,2), c(3,4))

layout(m)

timeframe <- as.data.frame(cbind(data.frame(epiNEM = time[1,,]), data.frame(BNEM = time[2,,]), data.frame(PC = time[3,,]), data.frame(NEM = time[4,,]), data.frame(ARACNE = time[5,,])))

colnames(timeframe) <- rep(noiselvls, 5) # c(paste(rep("epi", length(noiselvls)), noiselvls, sep = "-"), paste(rep("B", length(noiselvls)), noiselvls, sep = "-"), paste(rep("PC", length(noiselvls)), noiselvls, sep = "-"), paste(rep("NEM", length(noiselvls)), noiselvls, sep = "-"), paste(rep("ARACNE", length(noiselvls)), noiselvls, sep = "-"))

boxplot(timeframe, col = colvec, main = "running time", ylab = "seconds")

abline(v=(1:(length(do)-1)*length(noiselvls) + 0.5), col = "black", lty = 6)

axis(1, c(3, 11, 19, 28, 36)+1, c("epiNEM", "B-NEM", "NEM", "PC Algorithm", "ARACNE"), tick = F, pos = -1)

accframe2 <- as.data.frame(cbind(data.frame(epiNEM = acc2[1,,]), data.frame(BNEM = acc2[2,,]), data.frame(PC = acc2[3,,]), data.frame(NEM = acc2[4,,]), data.frame(ARACNE = acc2[5,,])))

colnames(accframe2) <- rep(noiselvls, 5) # c(paste(rep("E", length(noiselvls)), noiselvls, sep = "-"), paste(rep("B", length(noiselvls)), noiselvls, sep = "-"), paste(rep("PC", length(noiselvls)), noiselvls, sep = "-"), paste(rep("NEM", length(noiselvls)), noiselvls, sep = "-"), paste(rep("ARACNE", length(noiselvls)), noiselvls, sep = "-"))

boxplot(accframe2, col = colvec, main = "accuracy of the inferred edges", ylim = c(0,1))

abline(v=(1:(length(do)-1)*length(noiselvls) + 0.5), col = "black", lty = 6)

axis(1, c(3, 11, 19, 28, 36)+1, c("epiNEM", "B-NEM", "NEM", "PC Algorithm", "ARACNE"), tick = F, pos = -1)

## logical nems:
```

```

colvec2 <- c(rep("orange", length(noiselvls)), rep("blue", length(noiselvls)))

logicsframe <- as.data.frame(cbind(data.frame(epiNEM = logics[1,,]), data.frame(BNEM = logics[2,,])))

colnames(logicsframe) <- rep(noiselvls, 2) # c(paste(rep("E", length(noiselvls)), noiselvls, sep = "_"), p

boxplot(logicsframe, col = colvec2, main = "accuracy of the inferred logic gate", ylim = c(0,1))

abline(v=length(noiselvls)+0.5, col = "black", lty = 6)

axis(1, c(3, 11, 19, 28, 36)+1, c("epiNEM", "B-NEM", "NEM", "PC Algorithm", "ARACNE"), tick = F, pos = -

accframe <- as.data.frame(cbind(data.frame(epiNEM = acc[1,,]), data.frame(BNEM = acc[2,,])))

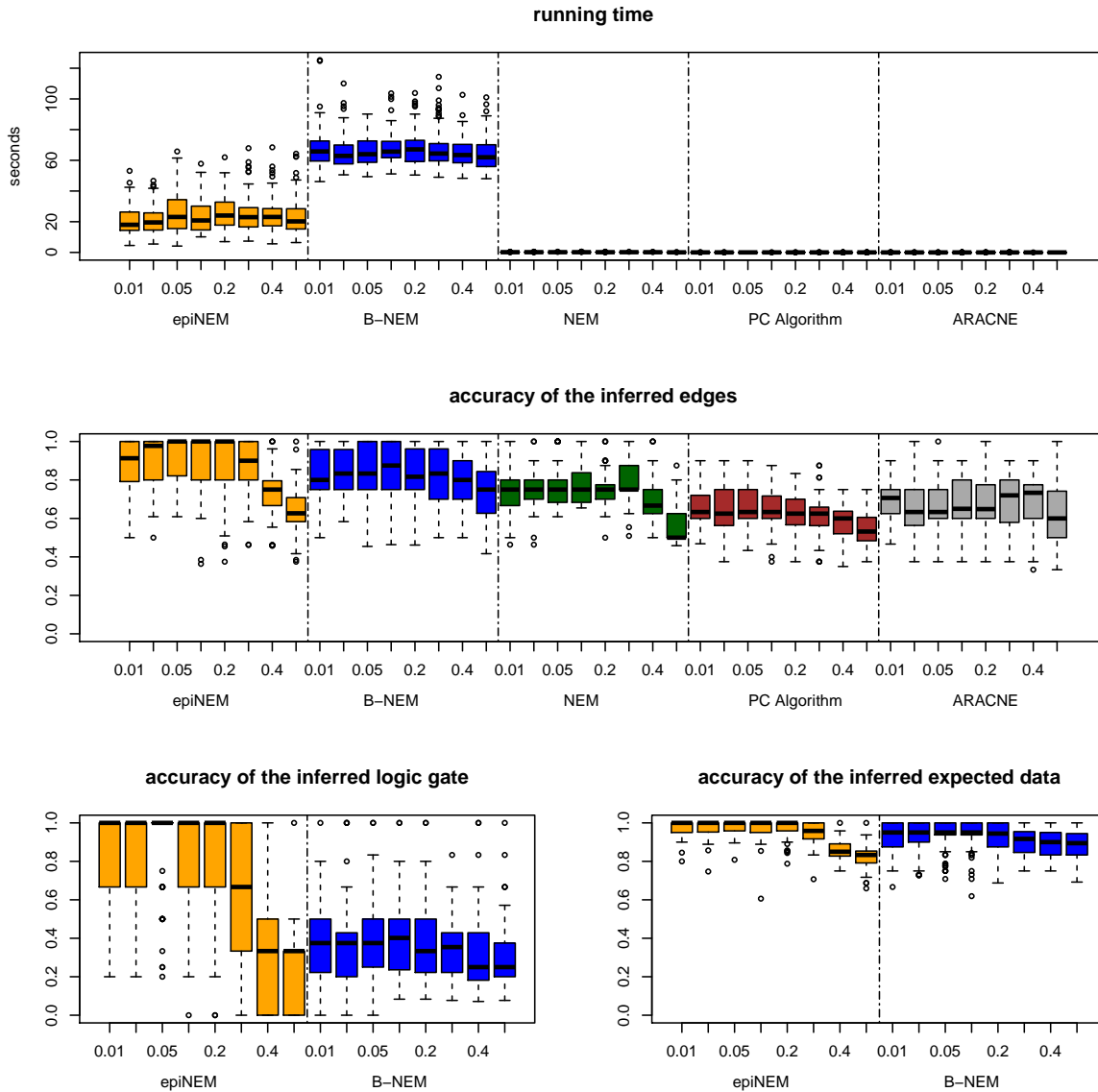
colnames(accframe) <- rep(noiselvls, 2) # c(paste(rep("E", length(noiselvls)), noiselvls, sep = "_"), p

boxplot(accframe, col = colvec2, main = "accuracy of the inferred expected data", ylim = c(0,1)) # what

abline(v=length(noiselvls)+0.5, col = "black", lty = 6)

axis(1, c(3, 11, 19, 28, 36)+1, c("epiNEM", "B-NEM", "NEM", "PC Algorithm", "ARACNE"), tick = F, pos = -

```



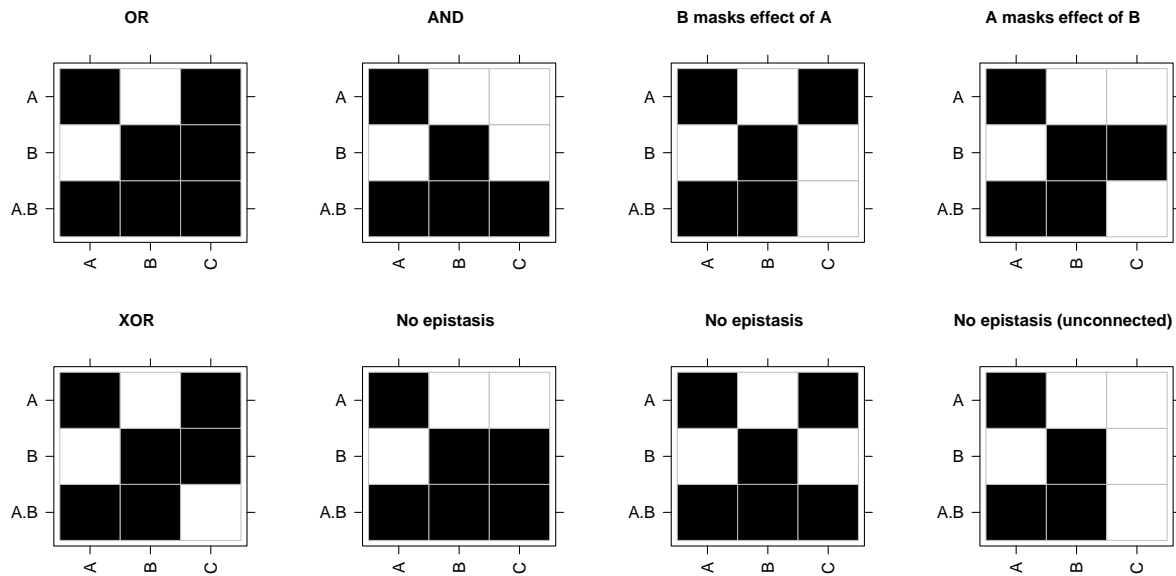
Yeast knockout screens

In this section we analyse previously published yeast knockout screens. The screens consist of gene expression data derived from double and single knockout mutants. We use epiNEM on each double mutant combined with each single mutant.

The results of the knockout screens have been annotated according to the following legend:

```
options(warn=-1)
heatmapOP(matrix(c(1,-1,1,-1,1,1, 1, 1, 1), 3, 3, dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), C
heatmapOP(matrix(c(1,-1,1,-1,1,1, -1, -1, 1), 3, 3, dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), C
heatmapOP(matrix(c(1,-1,1,-1,1,1, 1, -1, -1), 3, 3, dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), C
heatmapOP(matrix(c(1,-1,1,-1,1,1, -1, 1, -1), 3, 3, dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), C
heatmapOP(matrix(c(1,-1,1,-1,1,1, 1, 1, -1), 3, 3, dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), C
heatmapOP(matrix(c(1,-1,1,-1,1,1, -1, 1, 1), 3, 3, dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), C
```

```
heatmapOP(matrix(c(1,-1,1,-1,1,1, 1, -1, 1), 3, 3, dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), (
heatmapOP(matrix(c(1,-1,1,-1,1,1, -1, -1, -1), 3, 3, dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), (
options(warn=0)
```



Wageningen et al., 2010

```
data <- read.delim("http://www.holstegelab.nl/publications/sv/signaling_redundancy/downloads/DataS1.txt")
dataM <- data[-(1:2), (1+(1:(324/2))*2)]
dataP <- data[-(1:2), (2+(1:(324/2))*2)]
dataM <- dataM[-1, ]
dataP <- dataP[-1, ]
dataM <- apply(dataM, c(1,2), as.numeric)
dataP <- apply(dataP, c(1,2), as.numeric)
dataBin <- dataM
sig <- 0.05
cutoff <- log2(1.7)
dataBin[which(dataP < sig & dataP > 0 & abs(dataM) >= cutoff)] <- 1
dataBin[which(dataP >= sig | dataP == 0 | abs(dataM) < cutoff)] <- 0
dataBin <- dataBin[-which(apply(dataBin, 1, max) == 0), ]
genelist <- toupper(c('hs11', 'cla4', 'gin4', 'swe1', 'hs11.cla4'))
```

```

colnames(dataBin) <- gsub(".del.vs..wt", "", colnames(dataBin))

colnames(dataBin) <- gsub(".del", "", colnames(dataBin))

doubles <- colnames(dataBin)[grep("\\.", colnames(dataBin))]

doubles <- sort(doubles[-grep("vs", doubles)])

doubles.genes <- unique(unlist(strsplit(doubles, "\\.")))

singles <- colnames(dataBin)[-grep("\\.", colnames(dataBin))]

singles <- unique(sort(singles))

llmat <- logicmat <- matrix(0, length(singles), length(doubles))

rownames(llmat) <- rownames(logicmat) <- singles

colnames(llmat) <- colnames(logicmat) <- doubles

globalgenes <- which(apply(dataBin, 1, max) == 1)

## for (i in doubles[set]) {
##   if (which(doubles %in% i) == 8) { next() }
##   print(i)
##   doubles.singles <- unlist(strsplit(i, "\\."))
##   egenes <- which(apply(dataBin[, which(colnames(dataBin) %in% c(i, doubles.singles))], 1, max) == 1)
##   for (j in singles) {
##     print(j)
##     if (j %in% doubles.singles) { next() }

##     dataTmp <- dataBin[, grep(paste(paste("^", c(i, j, doubles.singles), "$", sep = ""), collapse = ""))]

##     if (path %in% "fixed_set") {
##       dataTmp <- dataTmp[egenes, ]
##     }
##     if (path %in% "global") {
##       dataTmp <- dataTmp[globalgenes, ]
##     }
##     if (path %in% "") {
##       dataTmp <- dataTmp[which(apply(dataTmp, 1, max) == 1), ]
##     }

##     i1 <- which(singles %in% j)
##     i2 <- which(doubles %in% i)

##     if (!(is.null(dim(dataTmp)))) {

##       if (any(dataTmp[, j] != 0)) {

##         epires <- epiNEM(dataTmp, method = "exhaustive")

##         tmp <- epires$logics

```

```

##           if ("OR" %in% tmp) {
##               if (sum(epires$origModel[, j]) != 2) {
##                   tmp <- "NOEPI"
##               } else {
##                   if (all(tmp %in% "OR")) {
##                       tmp <- "OR"
##                   } else {
##                       tmp <- tmp[which(!(tmp %in% "OR"))]
##                   }
##               }
##           }

##           logicmat[i1, i2] <- tmp
##           llmat[i1, i2] <- epires$score

##       } else {

##           logicmat[i1, i2] <- "UNCON"
##           llmat[i1, i2] <- -Inf

##       }

##   } else {

##       logicmat[i1, i2] <- "UNCON"
##       llmat[i1, i2] <- -Inf

##   }

## }

## }

```

Plot results.

```

palette(c("#4444cc", "#77aa77", "#009933", "#ff0000", "#dd8811", "#aa44bb", "#999900"))

data(wageningen_res)

llmat0 <- wageningen$ll

logicmat0 <- wageningen$logic

for (i in 1:length(doubles)) {

    #if (!(doubles[i] %in% c("ark1.prk1", "prk1.ark1", "ptp2.ptp3", "ptp3.ptp2", "bck1.ptp3", "ptp3.bck1"))) {

        if (i %in% 8) { next() }

        logicvec <- logicmat0[, i]

        llvec <- llmat0[, i]

        logicvec <- logicvec[order(llvec, decreasing = T)]
    }
}

```

```

llvec <- llvec[order(llvec, decreasing = T)]

parents <- unlist(strsplit(doubles[i], "\\\\"))

pchvec <- numeric(length(llvec))

pchvec[which(logicvec %in% "AND")] <- 1
pchvec[which(logicvec %in% "OR")] <- 2
pchvec[which(logicvec %in% "XOR")] <- 3
pchvec[grep(paste("^", parents[1], sep = ""), logicvec)] <- 4
pchvec[grep(paste("^", parents[2], sep = ""), logicvec)] <- 5
pchvec[which(logicvec %in% "NOEPI")] <- 6
pchvec[which(logicvec %in% c("NOINFO", "NOINF"))] <- 7

logicvec <- logicvec[-which(logicvec %in% "0")]
pchvec <- pchvec[-which(pchvec == 0)]
llvec <- llvec[-which(llvec == 0)]

colvec <- pchvec

if (all(is.infinite(llvec) == T)) {

  llvec[1:length(llvec)] <- -1000

  margin <- 100

  donames <- 30

} else {

  llvec[which(is.infinite(llvec) == T)] <- NA

  ## llvec[which(is.infinite(llvec) == T)] <- min(llvec) - 100

  margin <- abs(max(llvec[1:30], na.rm = T) - min(llvec[1:30], na.rm = T))

  offset <- 0.075

  if (margin == 0) { margin <- 10; offset <- 0.0375 }

  donames <- 30 - sum(is.na(llvec[1:30]) == T)

  if (any(is.na(llvec[1:30]) == T)) { margin2 <- margin*2 } else { margin2 <- margin }

  llvec[which(is.na(llvec) == T)] <- min(llvec, na.rm = T) - margin

  margin <- margin2

}

if (all(llvec[-(1:30)] - min(llvec[-(1:30)]) == 0)) {

  p2max <- max(llvec[-(1:30)]) + margin

```

```

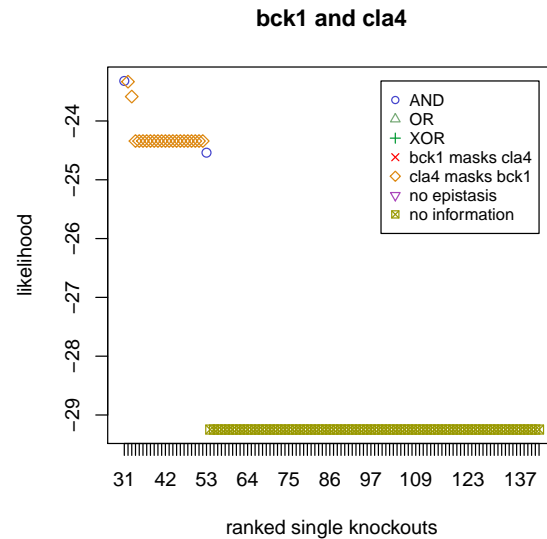
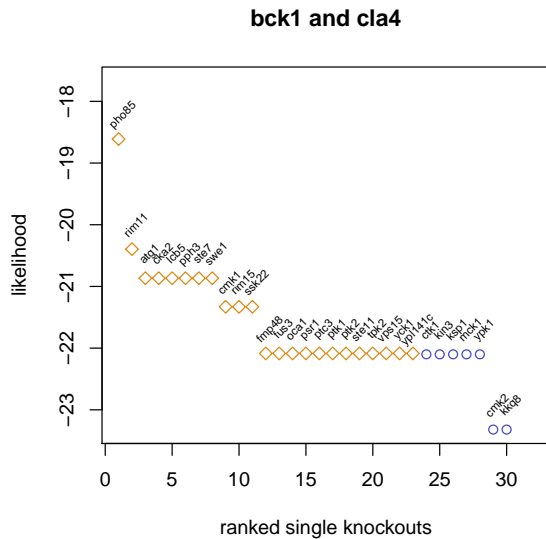
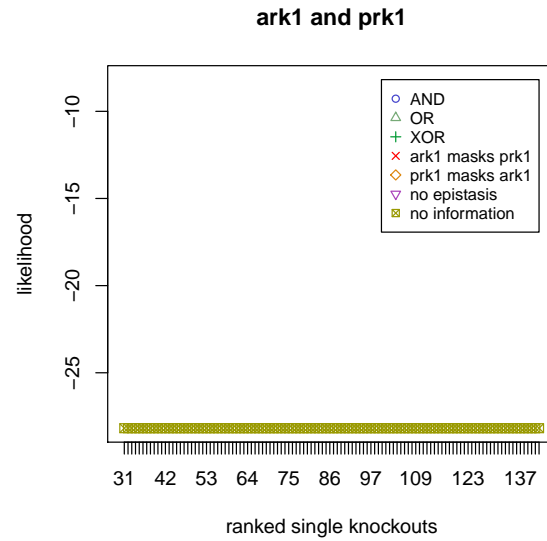
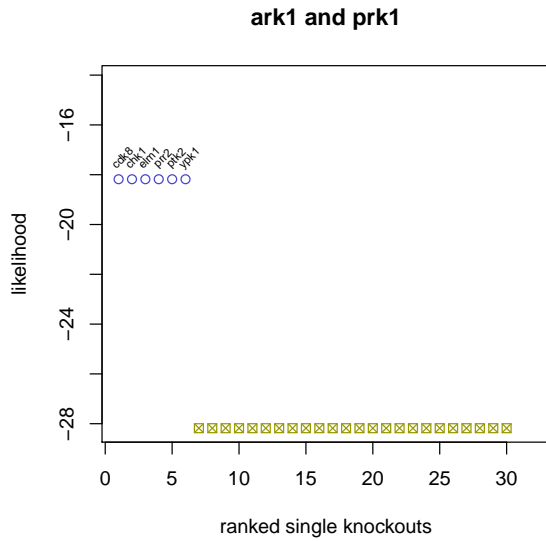
} else {

  p2max <- max(llvec[-(1:30)])

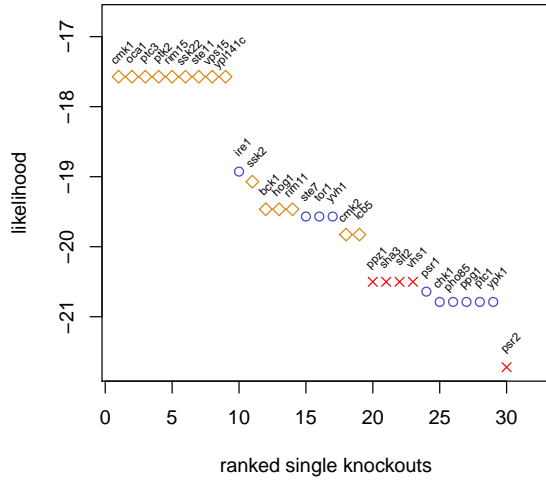
}

par = par(mfrow=c(1,2))
plot = plot(llvec[1:30], pch = pchvec[1:30], col = colvec[1:30], ylab = "likelihood", xlab = "ranked single knockouts")
text = text((1:30)+0.5, llvec[1:30]+(margin*offset), labels = c(names(llvec)[1:donames], rep("", 30 - donames)))
plot2 = plot(llvec[-(1:30)], pch = pchvec[-(1:30)], col = colvec[-(1:30)], ylab = "likelihood", xlab = "ranked single knockouts")
legend = legend(length(llvec[-(1:30)]), p2max,
               legend = c("AND", "OR", "XOR", paste(parents[1], " masks ", parents[2], sep = " "),
               no epistasis", no information"))
axis = axis(1, at = 1:length(llvec[-(1:30)]), labels = 31:length(llvec))
}

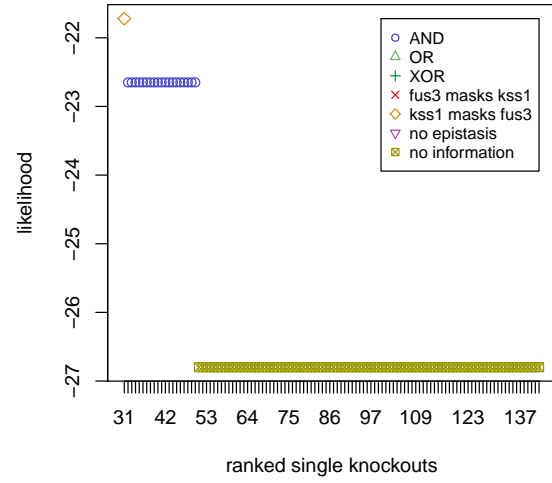
```



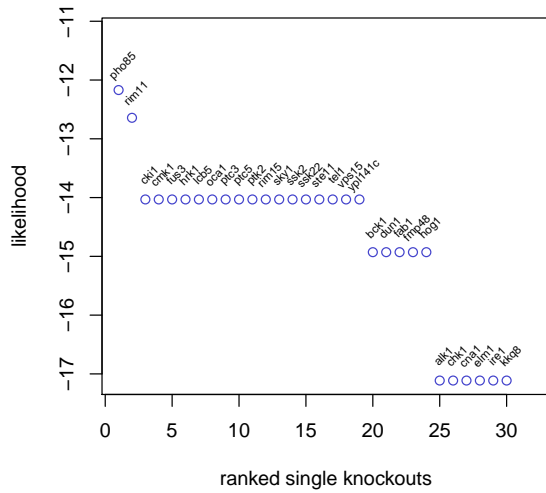
fus3 and kss1



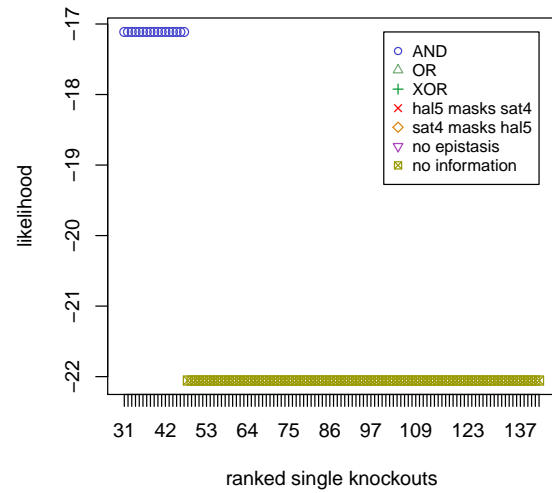
fus3 and kss1

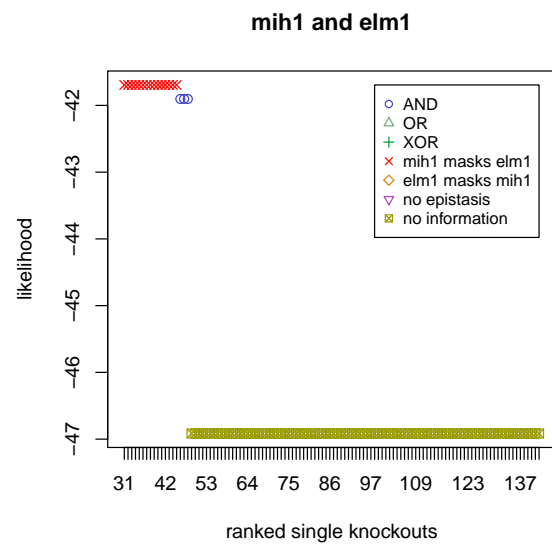
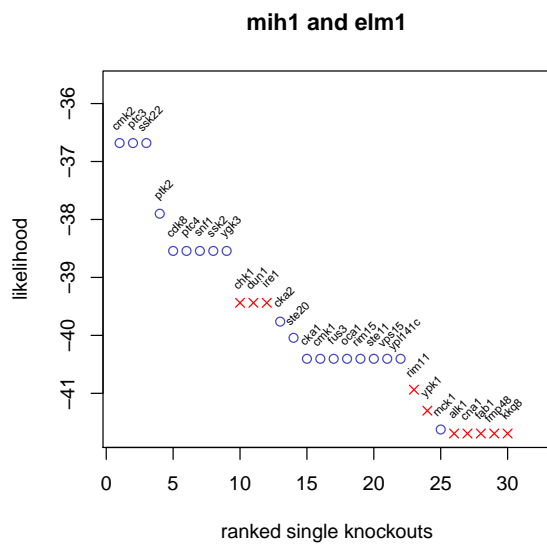
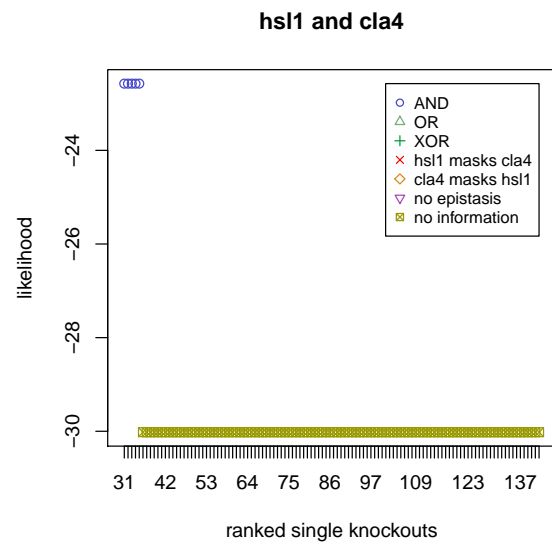
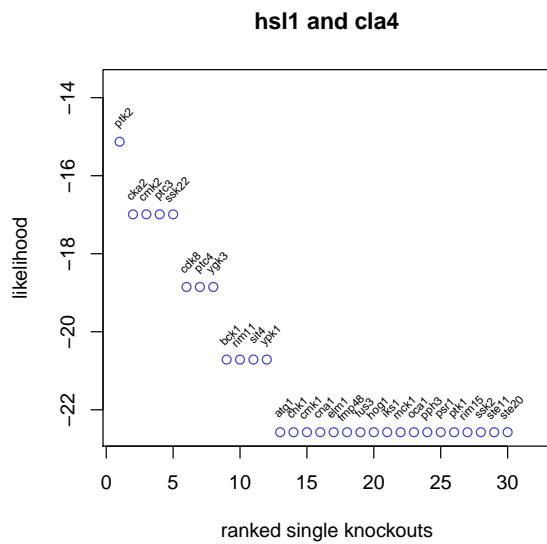


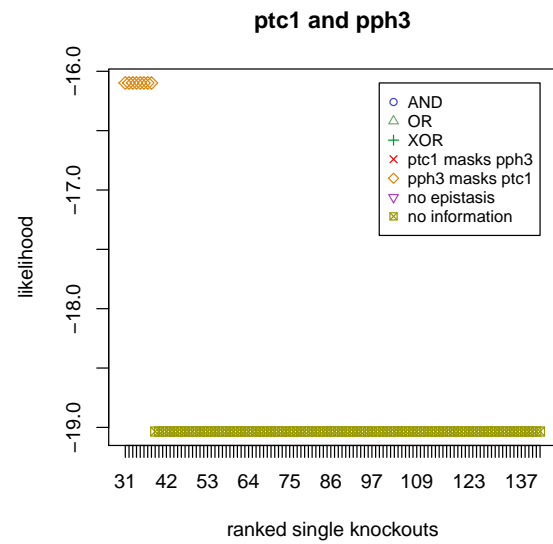
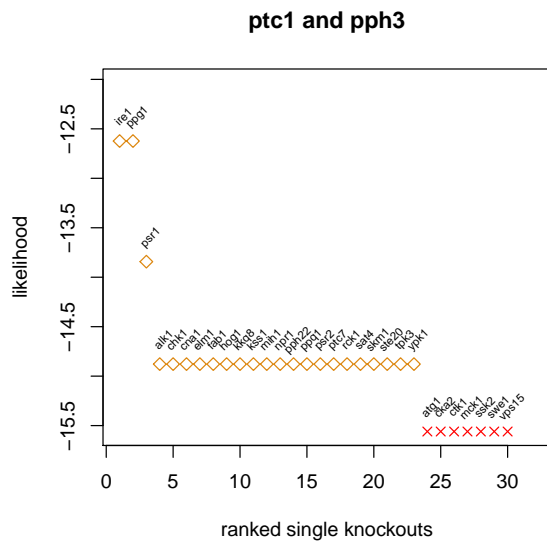
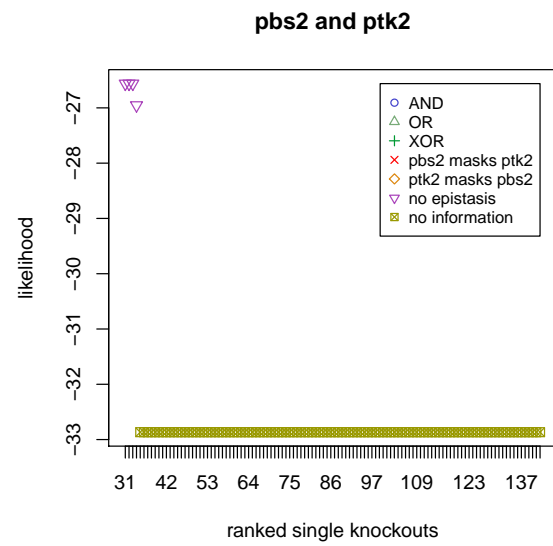
hal5 and sat4

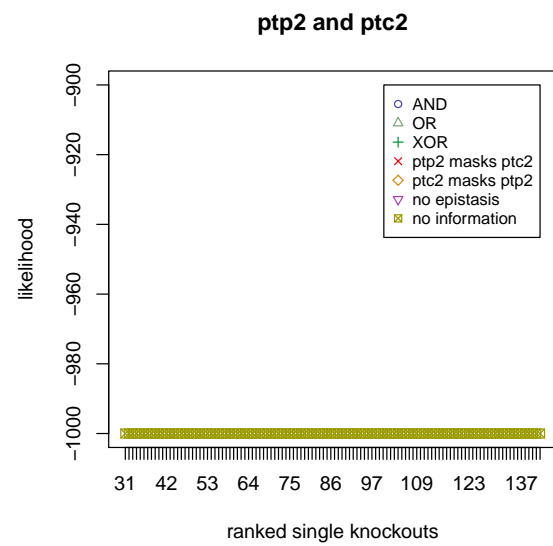
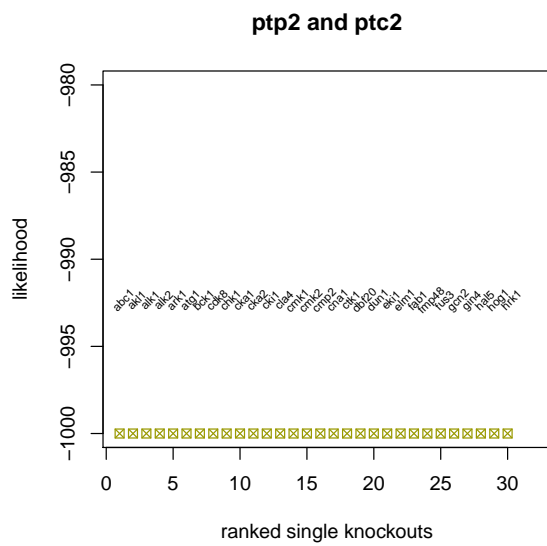
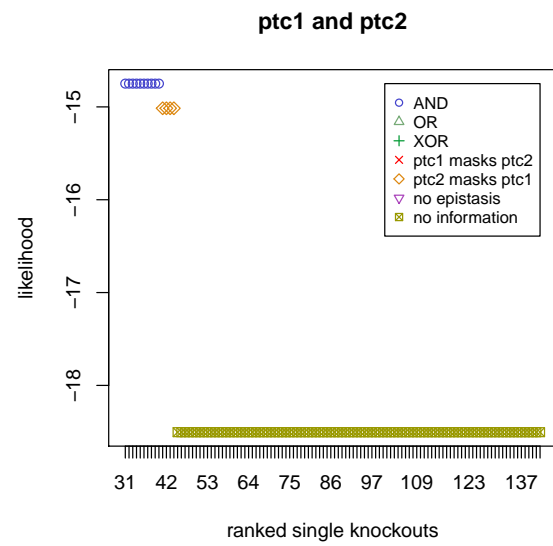
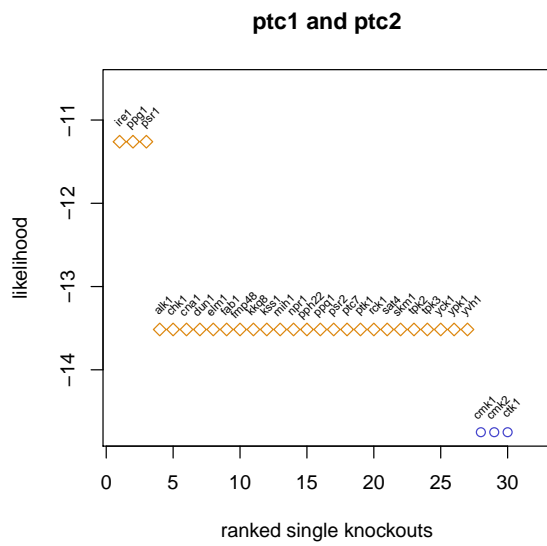


hal5 and sat4

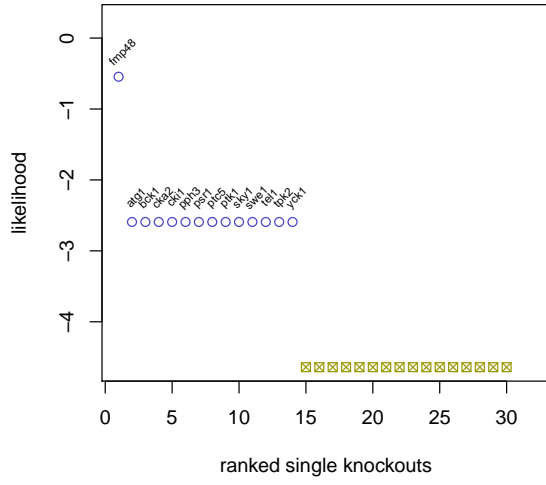




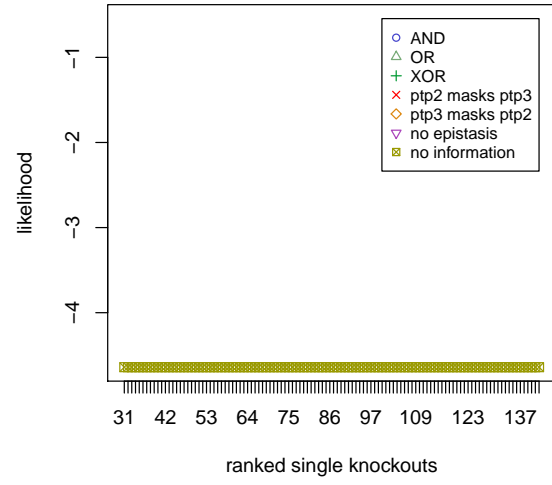




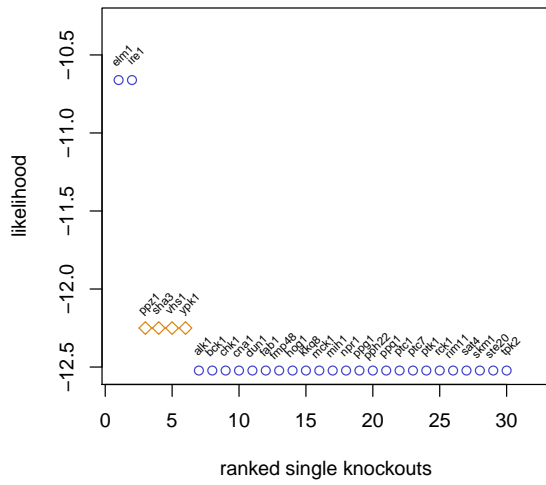
ptp2 and ptp3



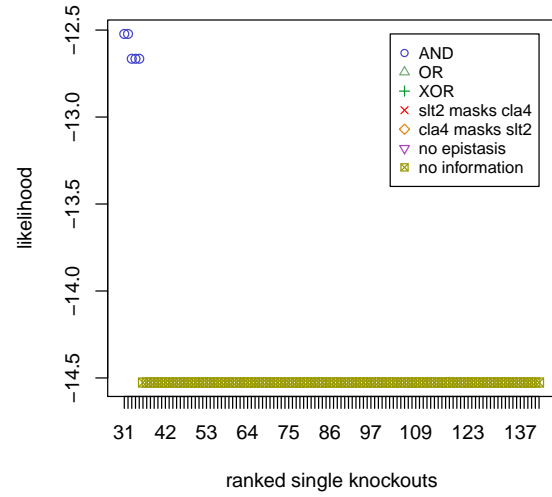
ptp2 and ptp3



slt2 and cla4



slt2 and cla4




```

    distmat[which(distmat[, i] %in% paste(genes[2], " masks the effect of ", genes[1], sep = "")), i] <- 0
  }

distmat <- apply(distmat, c(1,2), as.numeric)

for (i in 1:ncol(distmat)) {
  distmat[, i] <- rev(sort(distmat[, i]))
}

rownames(distmat) <- 1:nrow(distmat)

distmat <- distmat[-which(apply(distmat, 1, sum) == 0), ]

distmat <- distmat[, -which(apply(distmat, 2, max) == 0 | apply(distmat, 2, min) == 7)]

library(bnem)

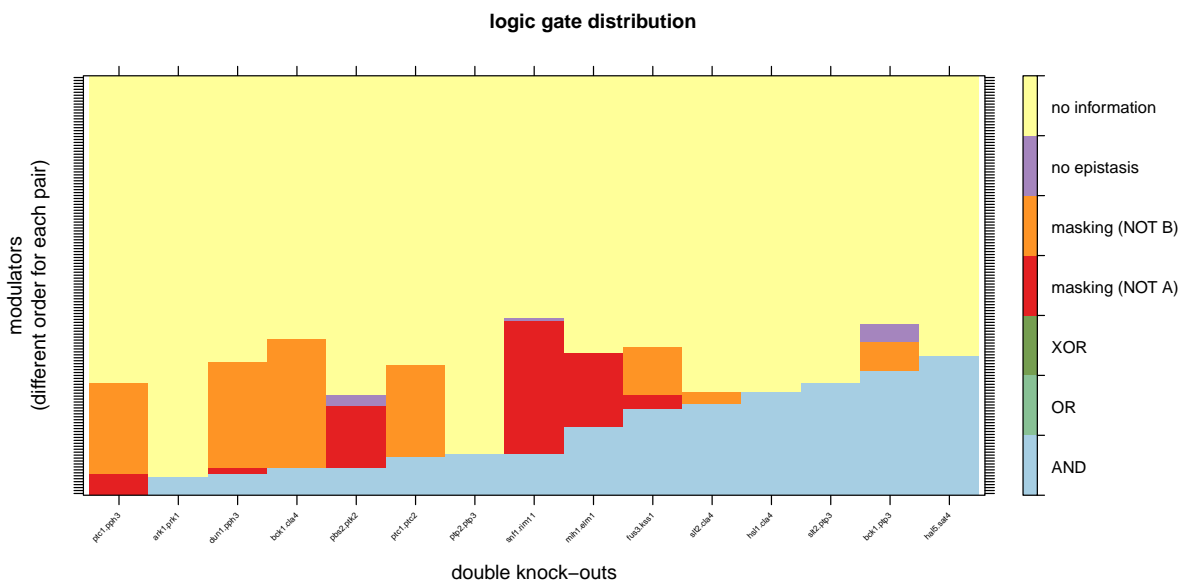
y <- distmat

distmat <- distmat[, order(apply(distmat, 2, function(x) { return(sum(x == 1)) }))]

y[which(y == 5)] <- 4

heatmapOP(distmat, Colv = F, Rowv = F, main = "logic gate distribution", sub = "", col = "Paired", break

```



Sameith et al., 2015

```

data <- read.delim("http://www.holstegelab.nl/publications/GSTF_geneticinteractions/downloads/del_mutant")

data <- apply(data, c(1,2), as.character)

dataM <- data[-1, which(data[1, ] %in% "M")]

```



```
dataM <- apply(dataM, c(1,2), as.numeric)

dataP <- data[-1, which(data[1, ] %in% "p.value")]

dataP <- apply(dataP, c(1,2), as.numeric)

dataBin <- dataM

sig <- 0.01

cutoff <- log2(1.5)

dataBin[which(dataP < sig & dataP > 0 & abs(dataM) >= cutoff)] <- 1

dataBin[which(dataP >= sig | dataP == 0 | abs(dataM) < cutoff)] <- 0

dataBin <- dataBin[-which(apply(dataBin, 1, max) == 0), ]

colnames(dataBin) <- gsub("\\.\\.\\.\\.\"", "\\.", colnames(dataBin))

## big screen:

doubles <- colnames(dataBin)[grep("\\.", colnames(dataBin))]

doubles.genes <- unique(unlist(strsplit(doubles, "\\.")))

singles <- colnames(dataBin)[-grep("\\.", colnames(dataBin))]

singles <- unique(sort(singles))

llmat <- logicmat <- matrix(0, length(singles), length(doubles))

rownames(llmat) <- rownames(logicmat) <- singles

colnames(llmat) <- colnames(logicmat) <- doubles

globalgenes <- which(apply(dataBin, 1, max) == 1)

## for (i in doubles[set]) {
##   print(i)
##   doubles.singles <- unlist(strsplit(i, "\\."))
##   egenes <- which(apply(dataBin[, which(colnames(dataBin) %in% c(i, doubles.singles))], 1, max) == 1)
##   for (j in singles) {
##     print(j)
##     if (j %in% doubles.singles) { next() }
##
##     dataTmp <- dataBin[, grep(paste(paste("^", c(i, j, doubles.singles), "$", sep = ""), collapse = ""))]
##
##     if (path %in% "fixed_set") {
##       dataTmp <- dataTmp[egenes, ]
##     }
##     if (path %in% "global") {
##       dataTmp <- dataTmp[globalgenes, ]
##     }
##   }
## }
```

```

##      }
##      if (path %in% "") {
##          dataTmp <- dataTmp[which(apply(dataTmp, 1, max) == 1), ]
##      }

##      i1 <- which(singles %in% j)
##      i2 <- which(doubles %in% i)

##      if (!(is.null(dim(dataTmp)))) {

##          if (any(dataTmp[, j] != 0)) {

##              epires <- epiNEM(dataTmp, method = "exhaustive")

##              tmp <- epires$logics
##              if ("OR" %in% tmp) {
##                  if (sum(epires$origModel[, j]) != 2) {
##                      tmp <- "NOEPI"
##                  } else {
##                      if (all(tmp %in% "OR")) {
##                          tmp <- "OR"
##                      } else {
##                          tmp <- tmp[which(!(tmp %in% "OR"))]
##                      }
##                  }
##              }

##              logicmat[i1, i2] <- tmp
##              llmat[i1, i2] <- epires$score

##          } else {

##              logicmat[i1, i2] <- "UNCON"
##              llmat[i1, i2] <- -Inf

##          }

##      } else {

##          logicmat[i1, i2] <- "UNCON"
##          llmat[i1, i2] <- -Inf

##      }

##  }

## }

```

```

data(sameith_res)

llmat0 <- sameith$ll

logicmat0 <- sameith$logic

```

```

paperdoubles <- c(4, 9, 17)

for (i in 1:length(doubles)) {

  ## if (!(doubles[i] %in% c("ECM22.UPC2", "GLN3.GZF3"))) { next() }

  logicvec <- logicmat0[, i]

  llvec <- llmat0[, i]

  logicvec <- logicvec[order(llvec, decreasing = T)]

  llvec <- llvec[order(llvec, decreasing = T)]

  parents <- unlist(strsplit(doubles[i], "\\\\"))

  pchvec <- numeric(length(llvec))

  pchvec[which(logicvec %in% "AND")] <- 1
  pchvec[which(logicvec %in% "OR")] <- 2
  pchvec[which(logicvec %in% "XOR")] <- 3
  pchvec[grep(paste("^", parents[1], sep = ""), logicvec)] <- 4
  pchvec[grep(paste("^", parents[2], sep = ""), logicvec)] <- 5
  pchvec[which(logicvec %in% "NOEPI")] <- 6
  pchvec[which(logicvec %in% c("NOINFO", "NOINF"))] <- 7

  logicvec <- logicvec[-which(logicvec %in% "0")]
  pchvec <- pchvec[-which(pchvec == 0)]
  llvec <- llvec[-which(llvec == 0)]

  colvec <- pchvec

  if (all(is.infinite(llvec) == T)) {

    llvec[1:length(llvec)] <- -1000

    margin <- 100

    donames <- 30

  } else {

    llvec[which(is.infinite(llvec) == T)] <- NA

    ## llvec[which(is.infinite(llvec) == T)] <- min(llvec) - 100

    margin <- abs(max(llvec[1:30], na.rm = T) - min(llvec[1:30], na.rm = T))

    if (margin == 0) { margin <- 10 }

    donames <- 30 - sum(is.na(llvec[1:30]) == T)

    if (any(is.na(llvec[1:30]) == T)) { margin2 <- margin*2 } else { margin2 <- margin }
  }
}

```

```

    llvec[which(is.na(llvec) == T)] <- min(llvec, na.rm = T) - margin

    margin <- margin2

  }

  if (all(llvec[-(1:30)] - min(llvec[-(1:30)])) == 0)) {

    p2max <- max(llvec[-(1:30)]) + margin

  } else {

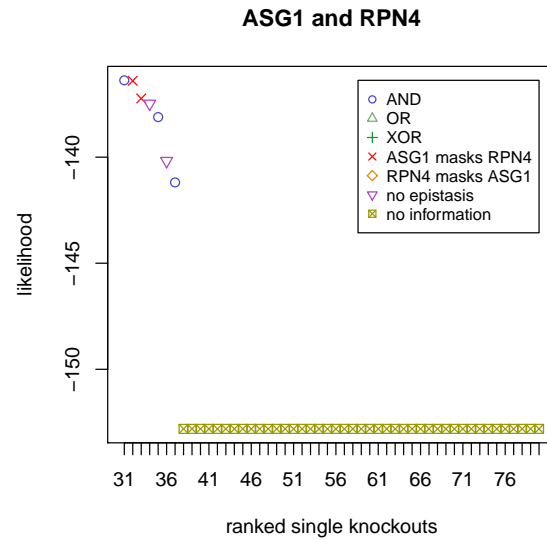
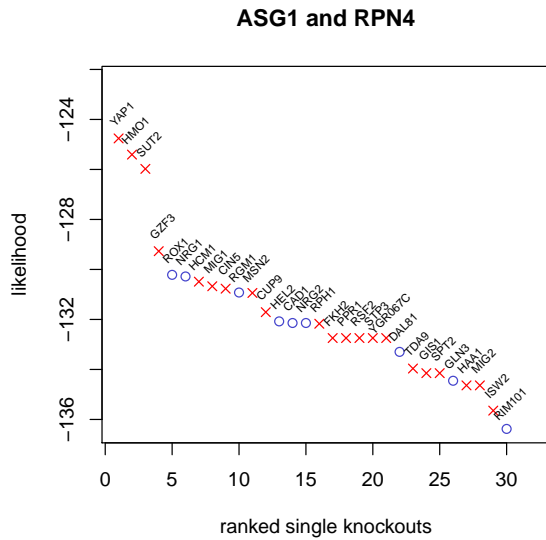
    p2max <- max(llvec[-(1:30)])

  }

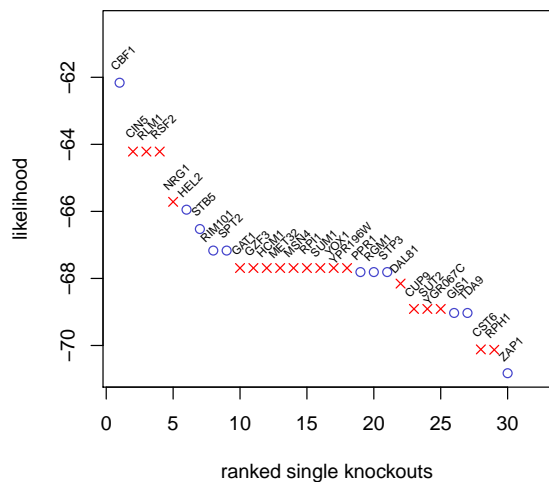
  par = par(mfrow=c(1,2))
  plot = plot(llvec[1:30], pch = pchvec[1:30], col = colvec[1:30], ylab = "likelihood", xlab = "ranked single knockouts",
    text = text((1:30)+0.5, llvec[1:30]+(margin*0.075), labels = c(names(llvec)[1:donames], rep("", 30 - donames)),
    plot2 = plot(llvec[-(1:30)], pch = pchvec[-(1:30)], col = colvec[-(1:30)], ylab = "likelihood", xlab = "ranked single knockouts",
    legend = legend(length(llvec[-(1:30)]), p2max,
      legend = c("AND", "OR", "XOR", paste(parents[1], " masks ", parents[2], sep = ")),
    axis = axis(1, at = 1:length(llvec[-(1:30)]), labels = 31:length(llvec))

}

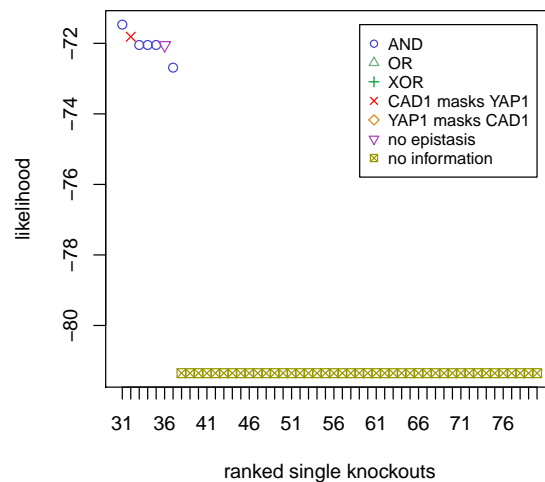
```



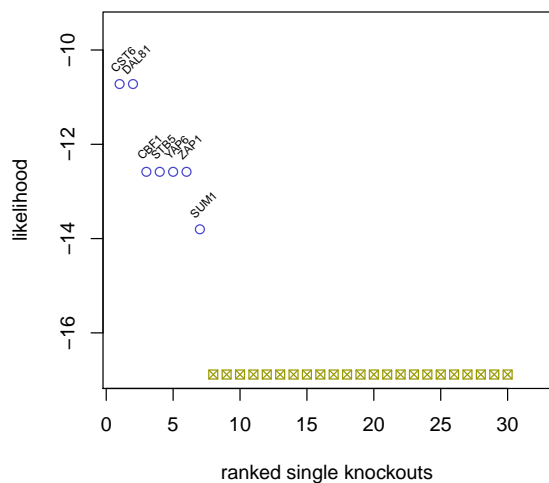
CAD1 and YAP1



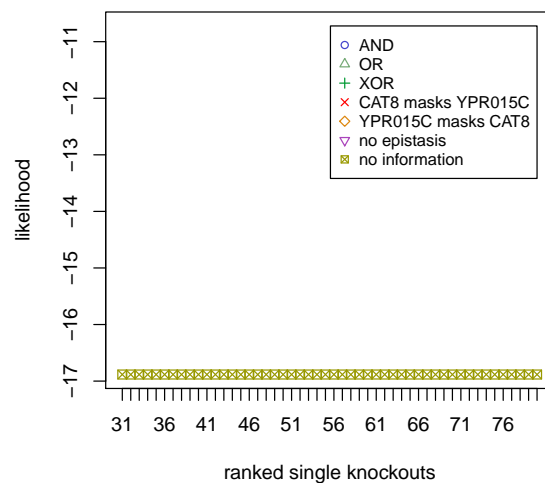
CAD1 and YAP1

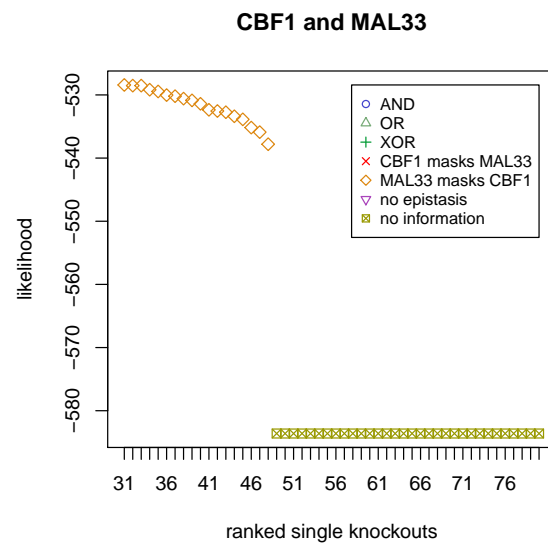
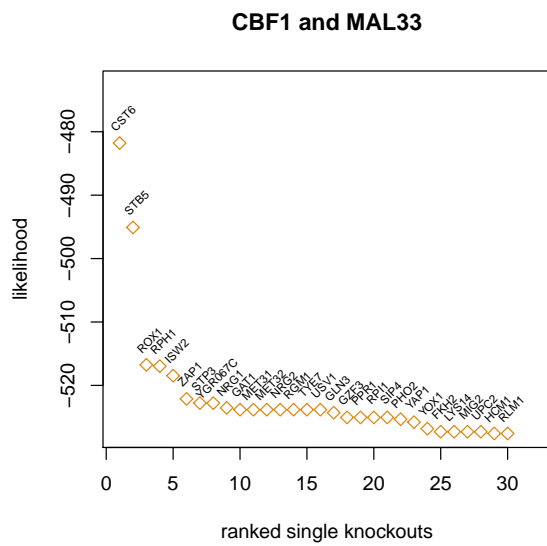
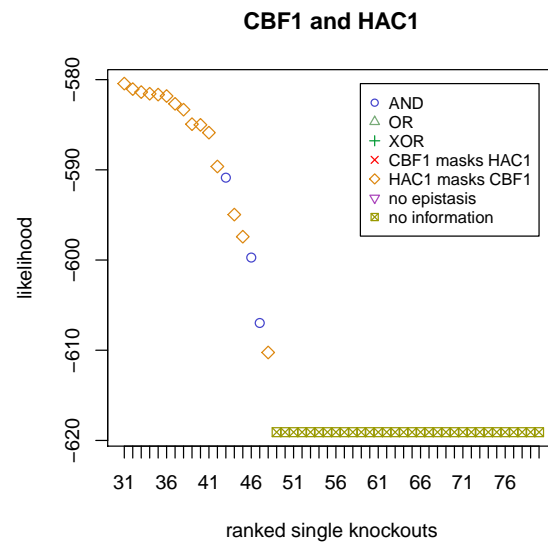
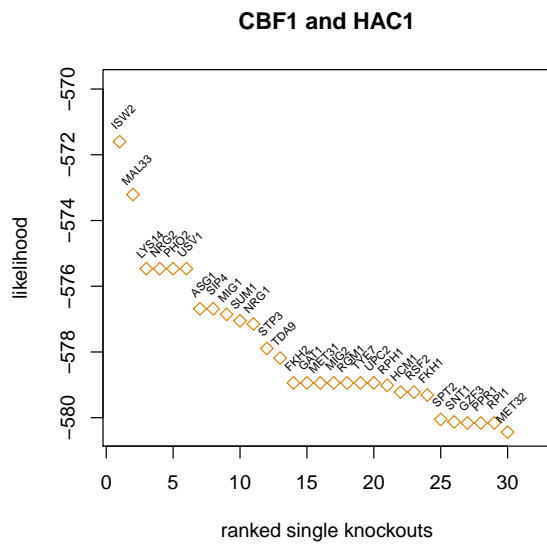


CAT8 and YPR015C

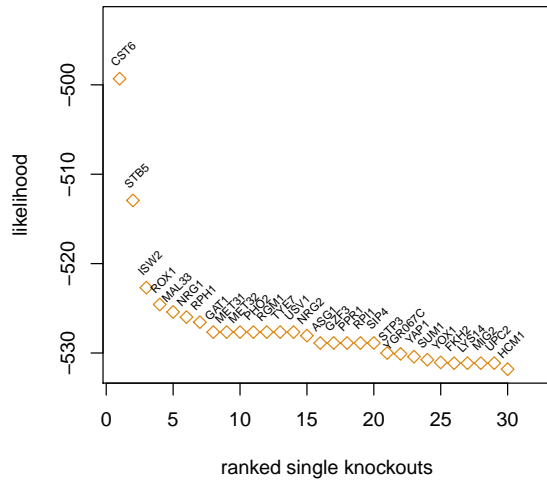


CAT8 and YPR015C

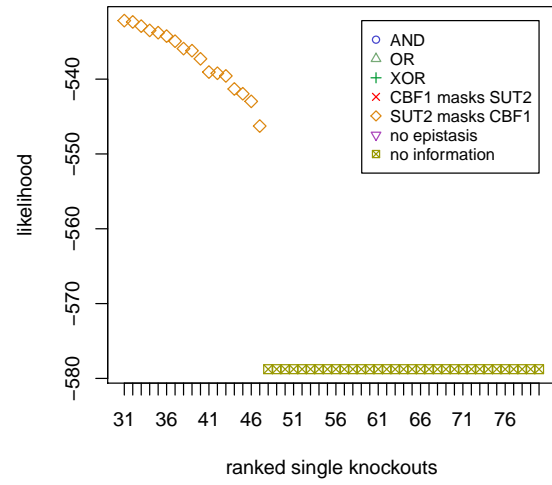




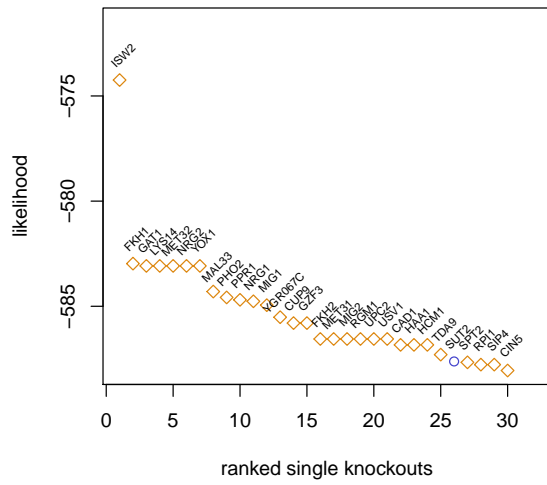
CBF1 and SUT2



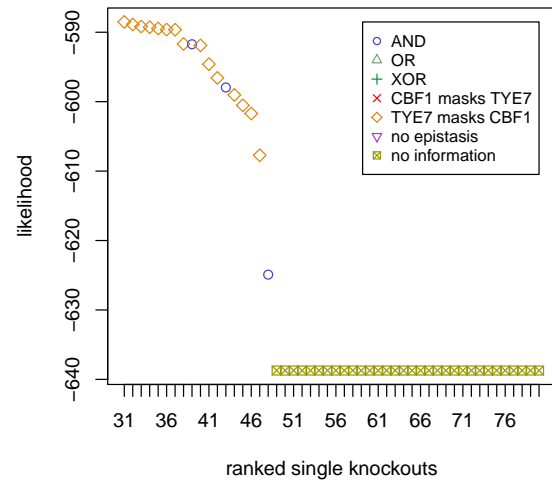
CBF1 and SUT2

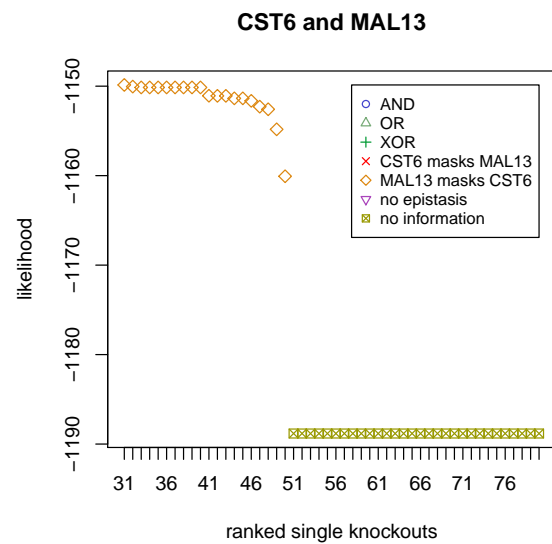
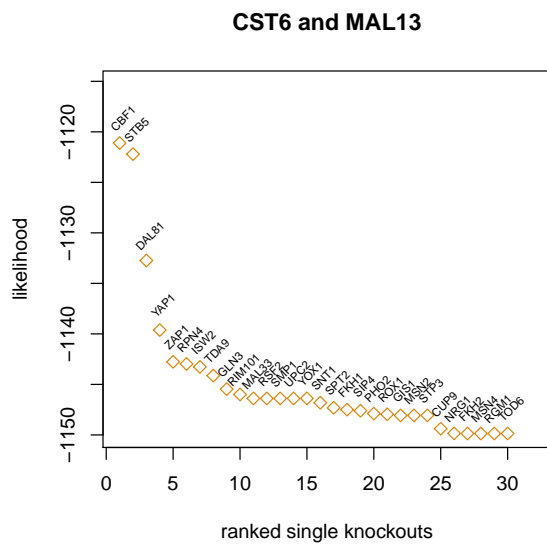
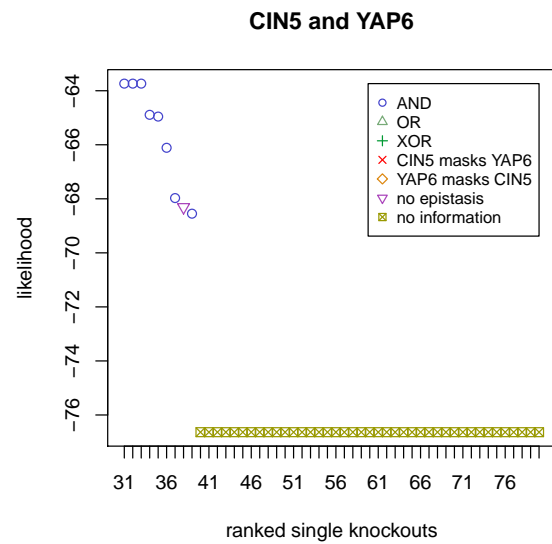
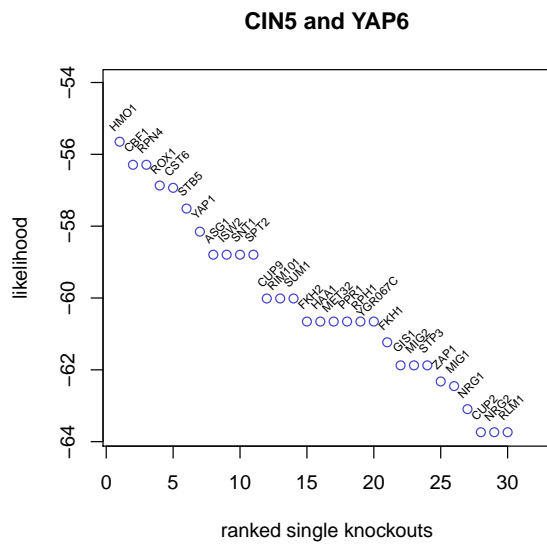


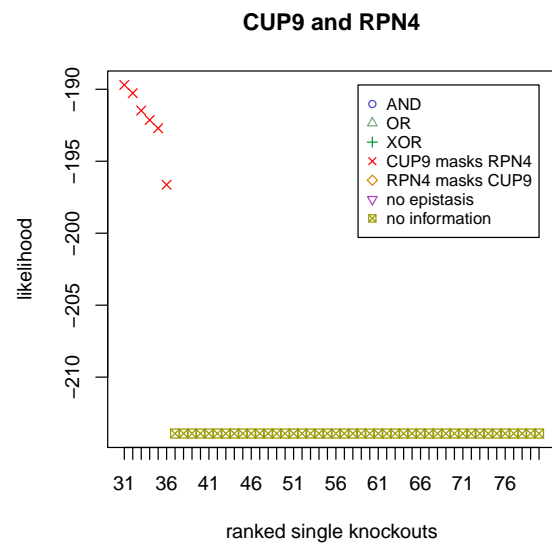
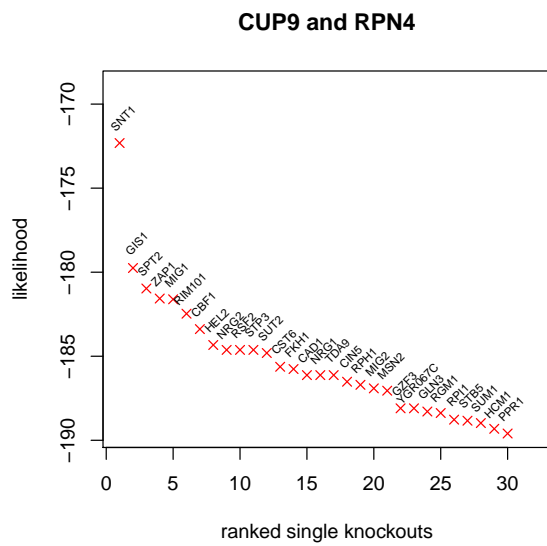
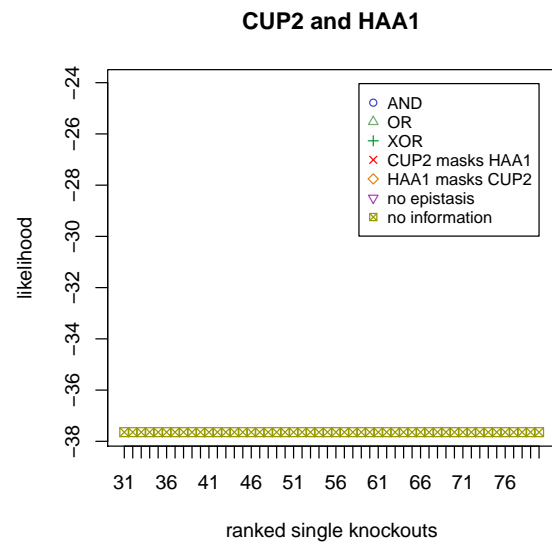
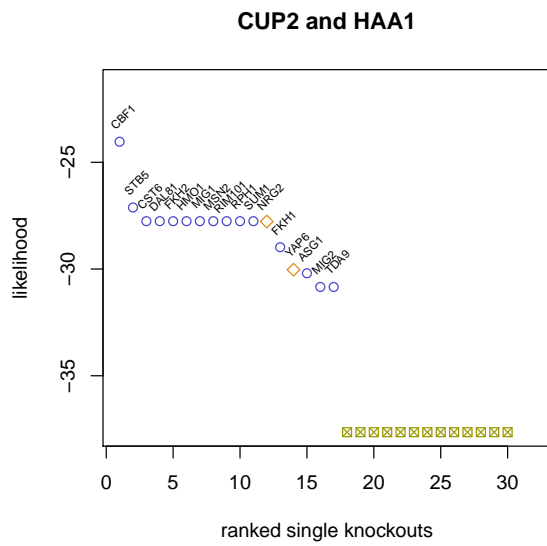
CBF1 and TYE7

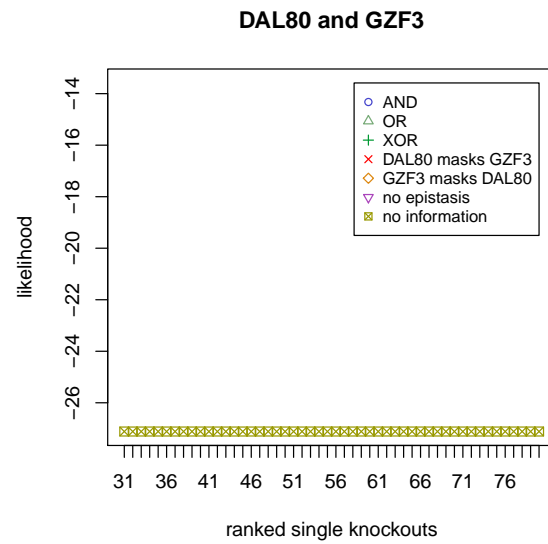
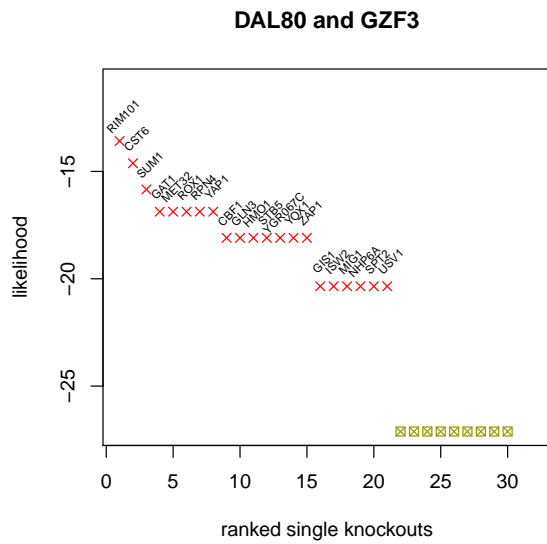
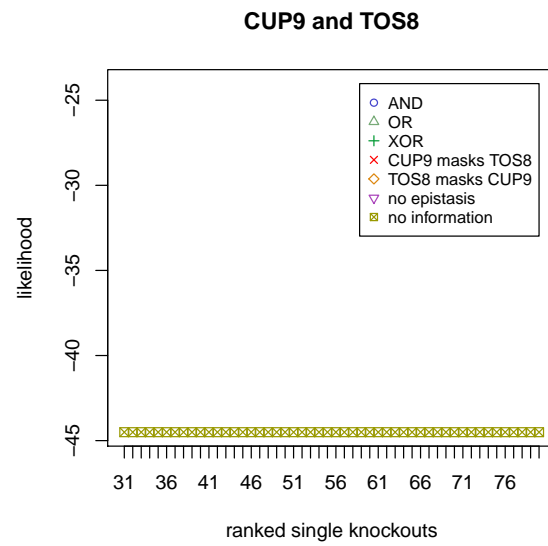
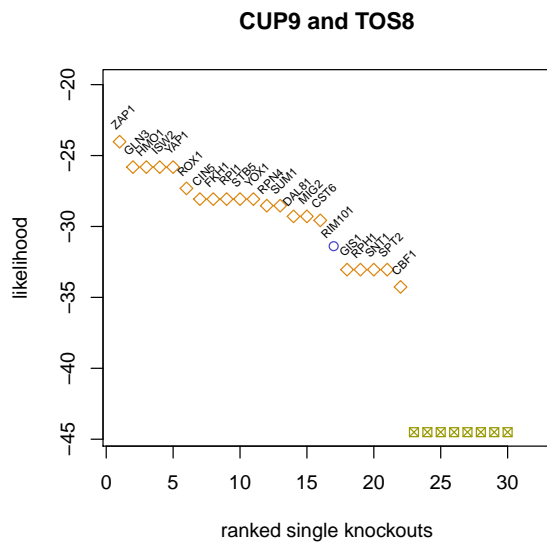


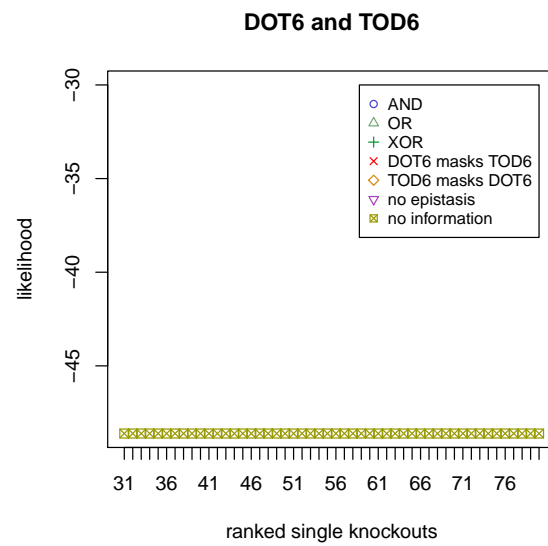
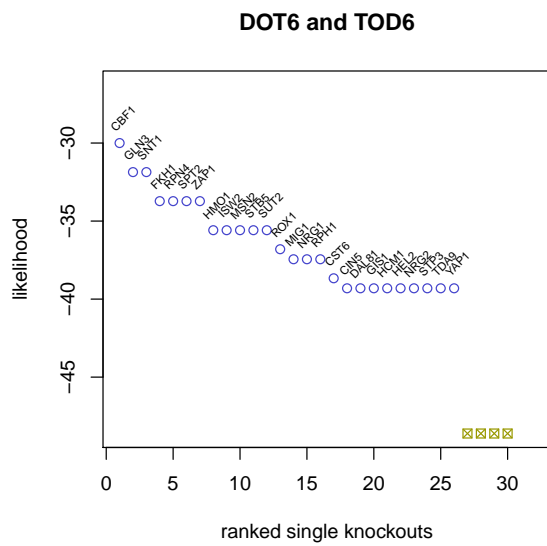
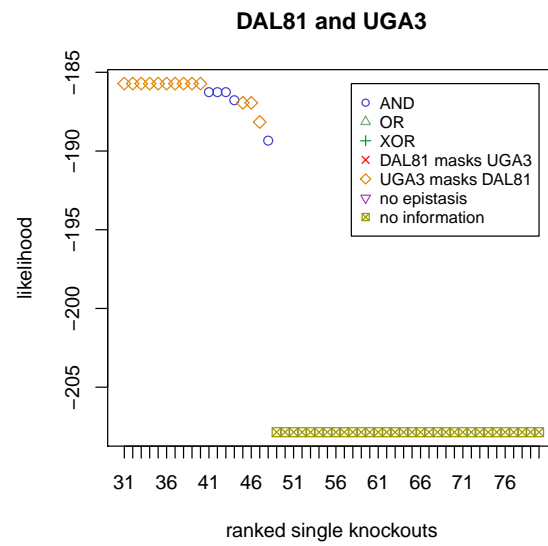
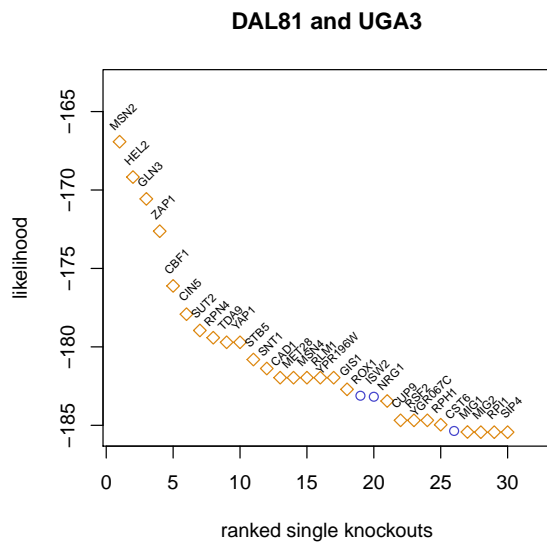
CBF1 and TYE7



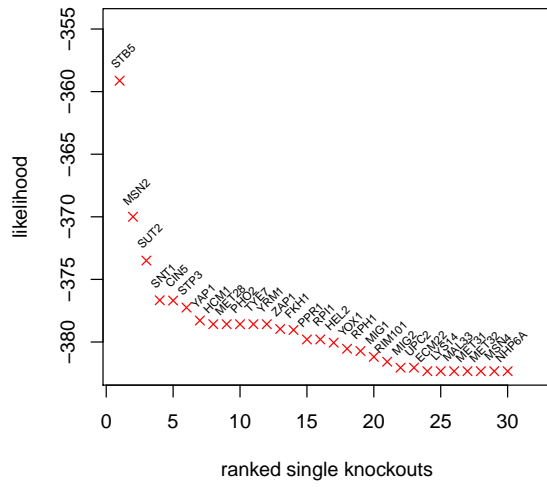




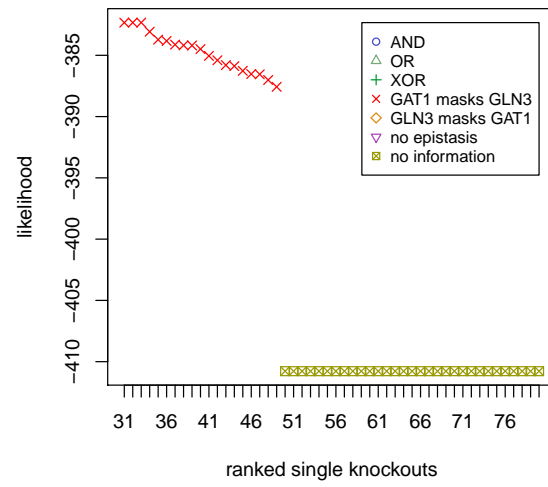




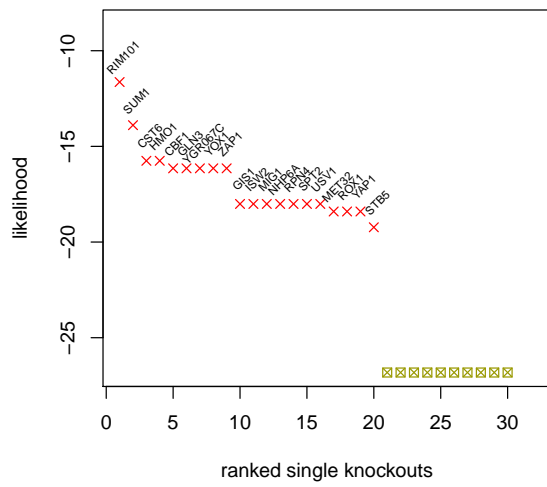
GAT1 and GLN3



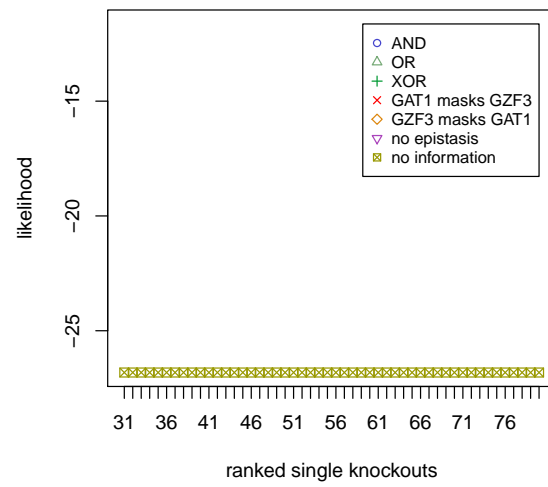
GAT1 and GLN3

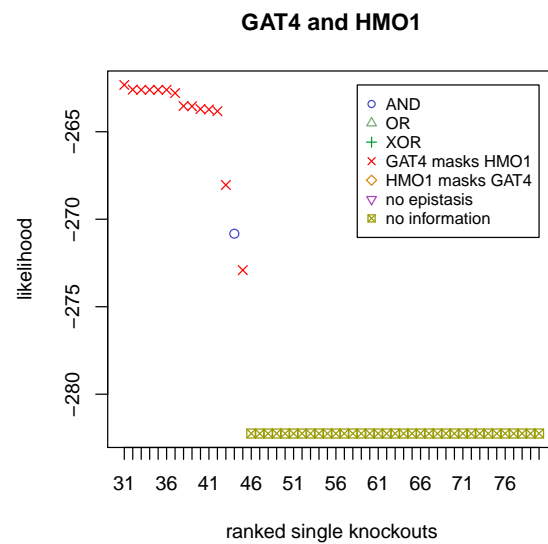
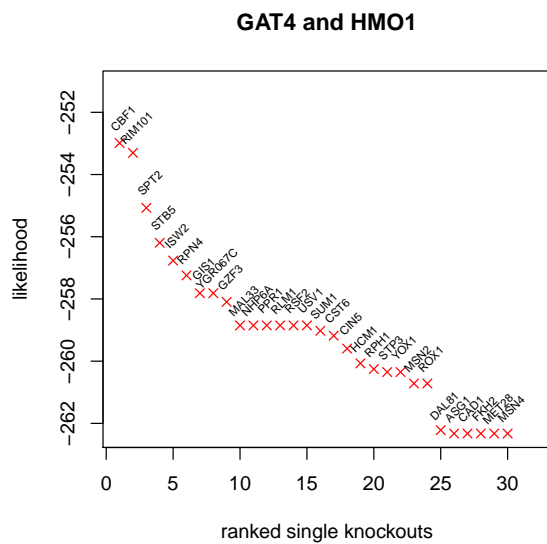
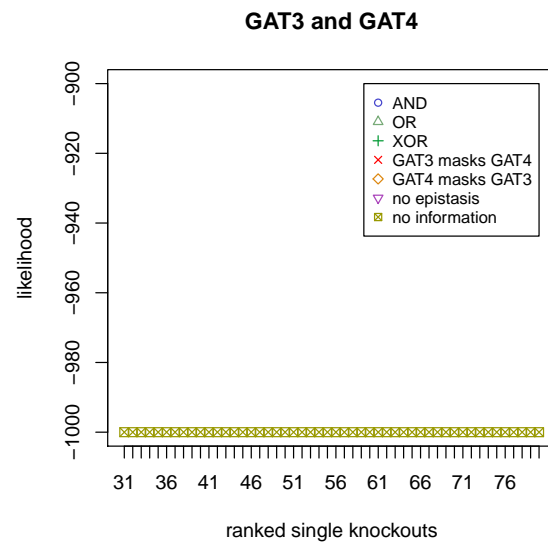
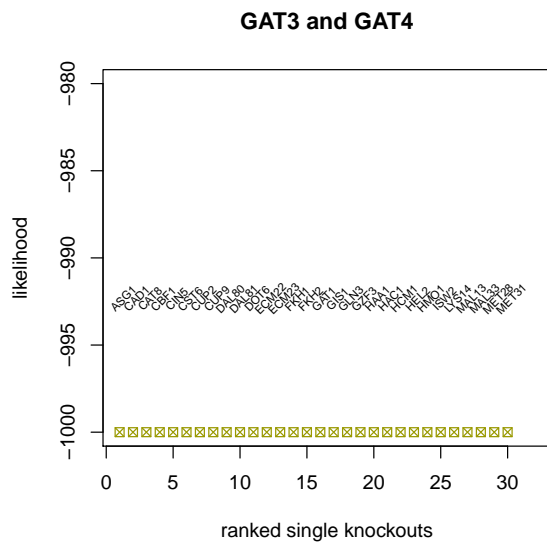


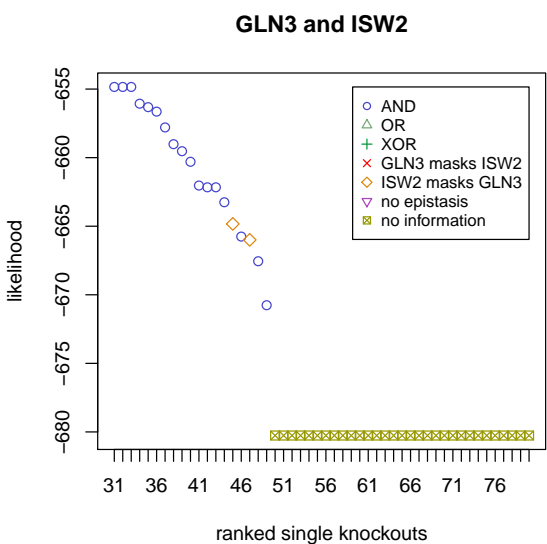
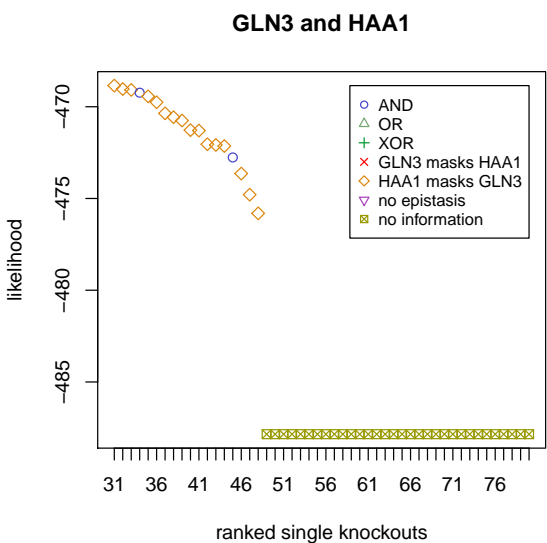
GAT1 and GZF3



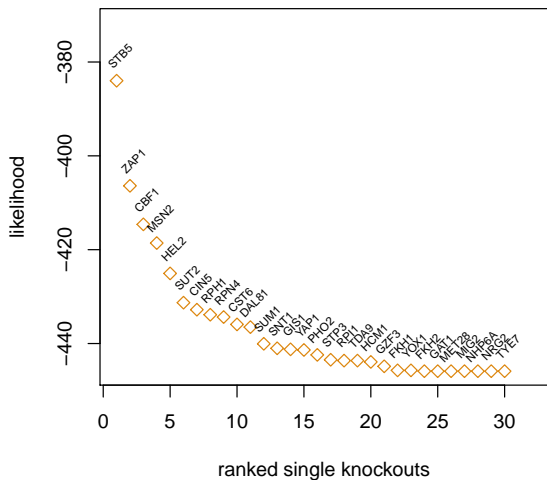
GAT1 and GZF3



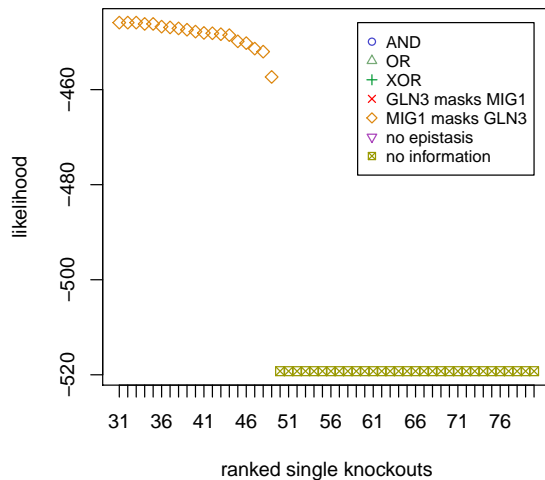




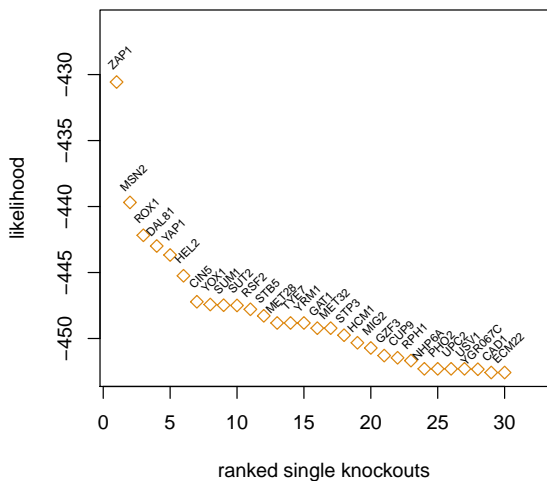
GLN3 and MIG1



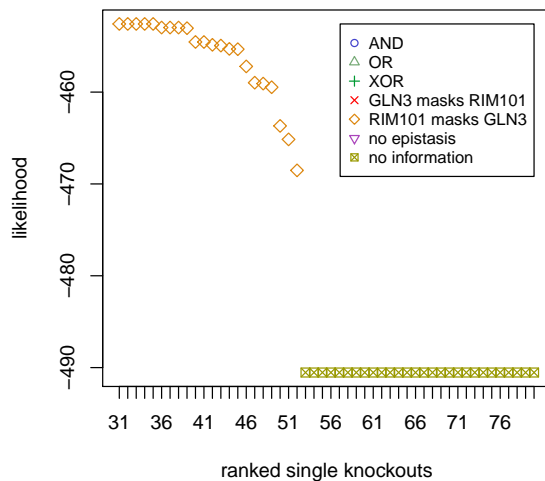
GLN3 and MIG1

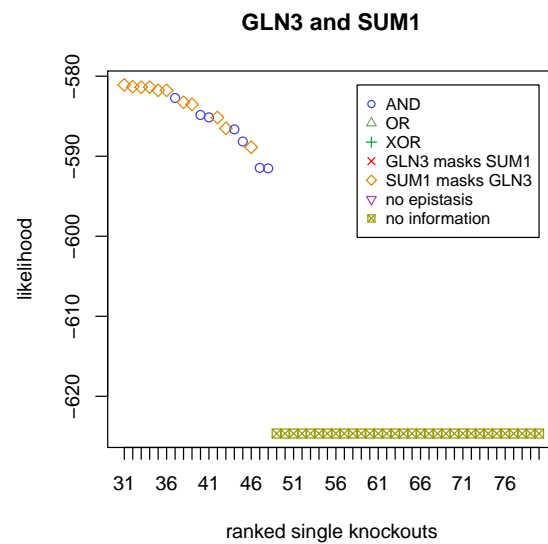
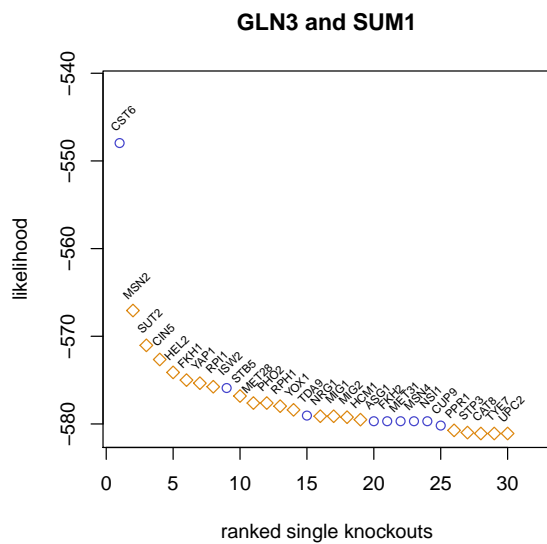
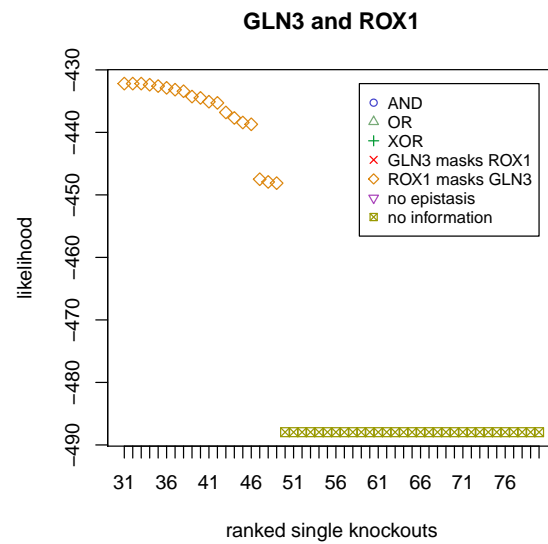
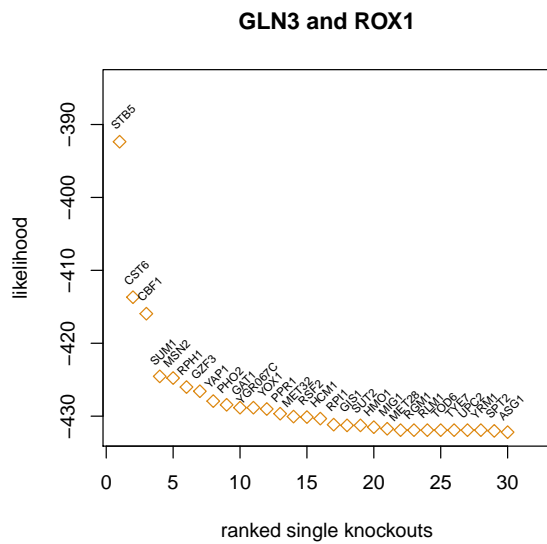


GLN3 and RIM101

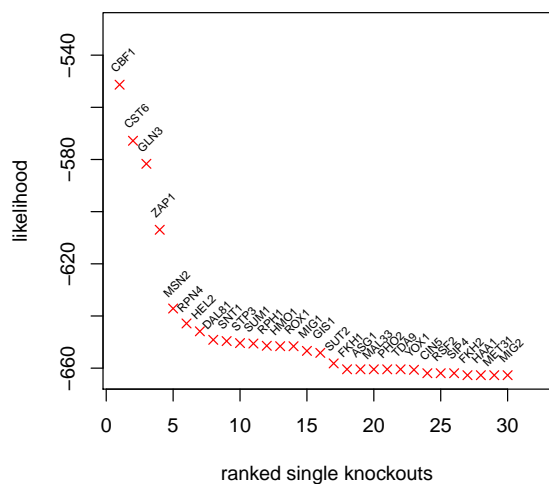


GLN3 and RIM101

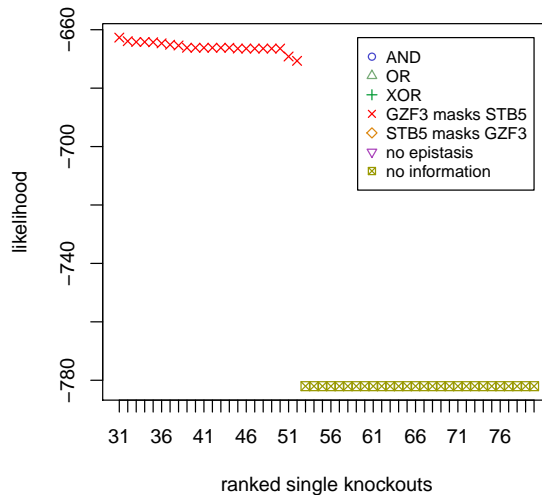




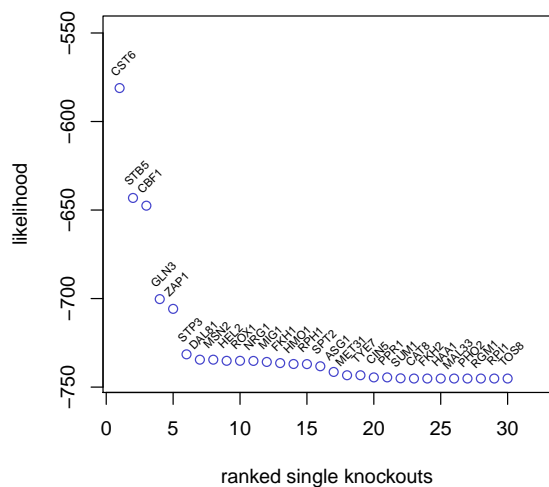
GZF3 and STB5



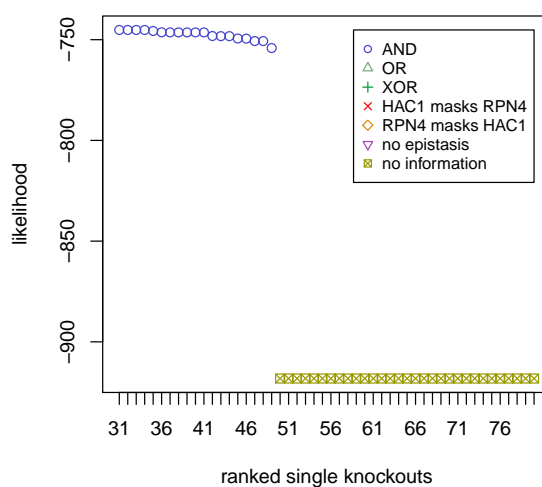
GZF3 and STB5

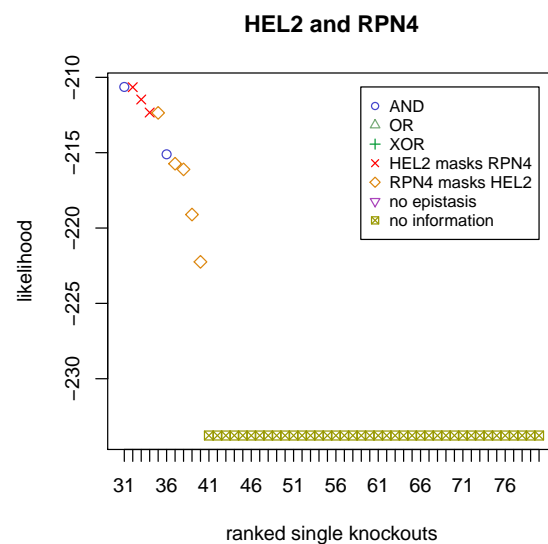
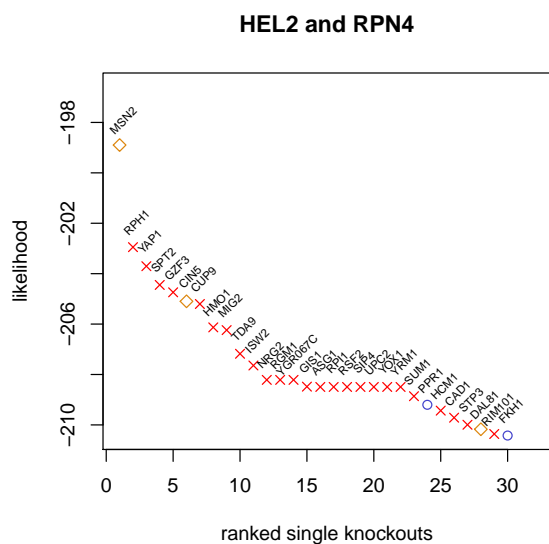
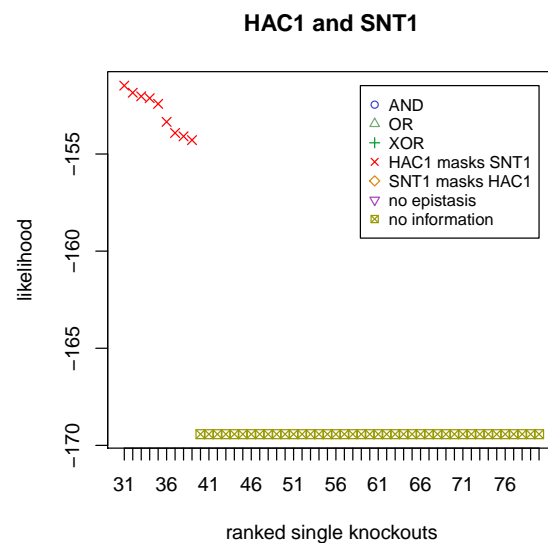
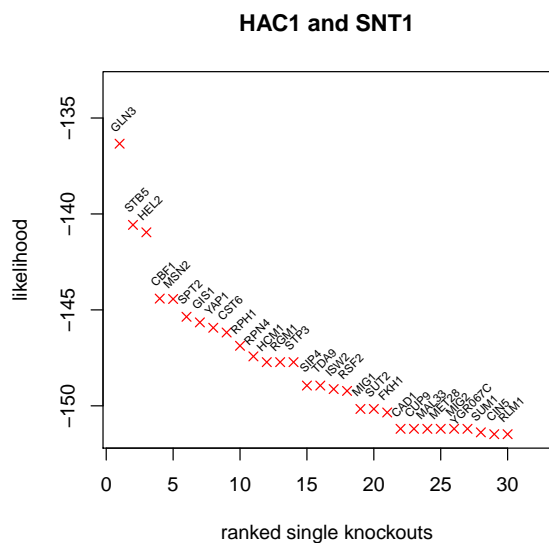


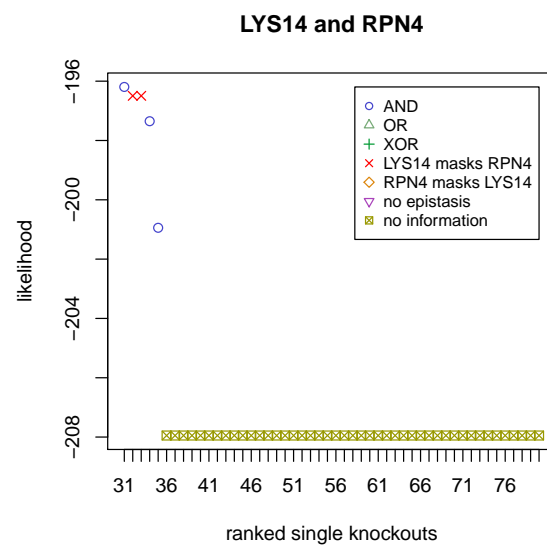
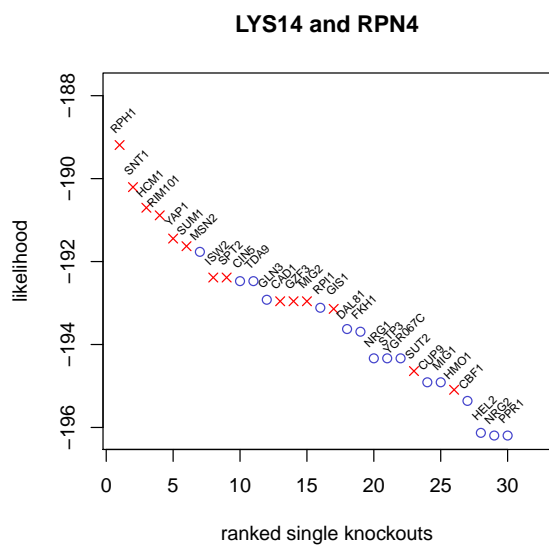
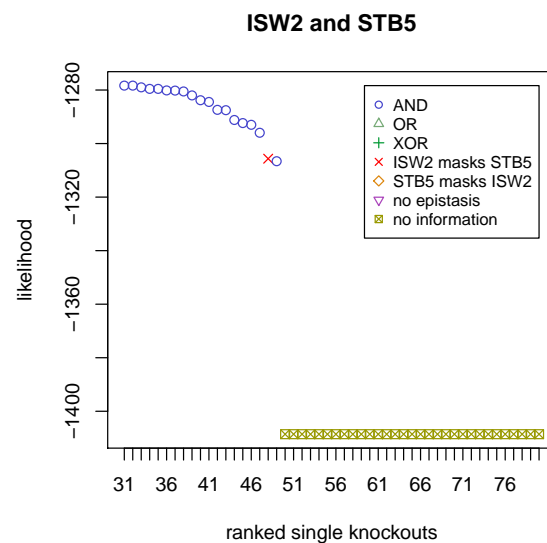
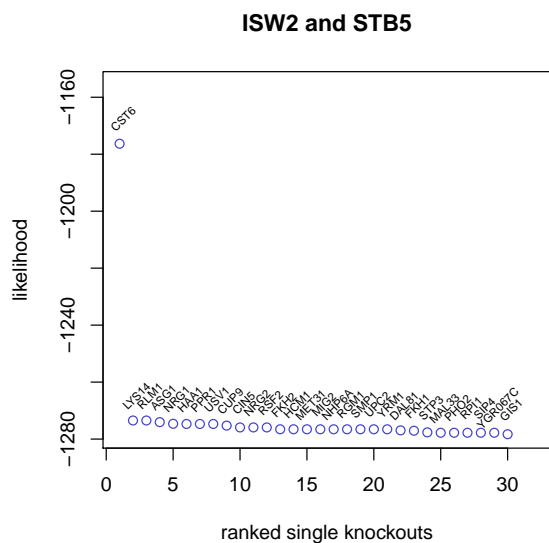
HAC1 and RPN4

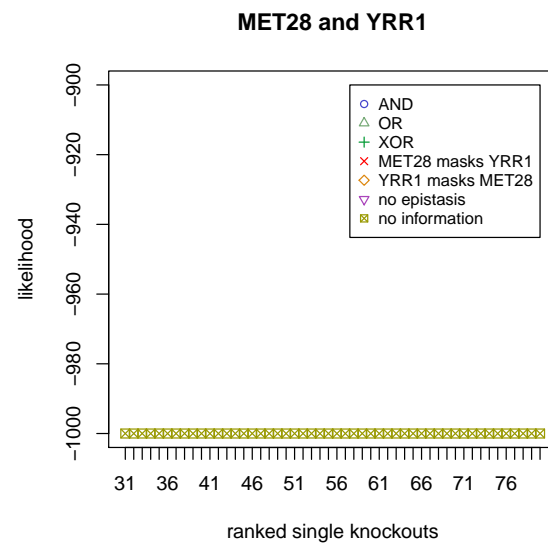
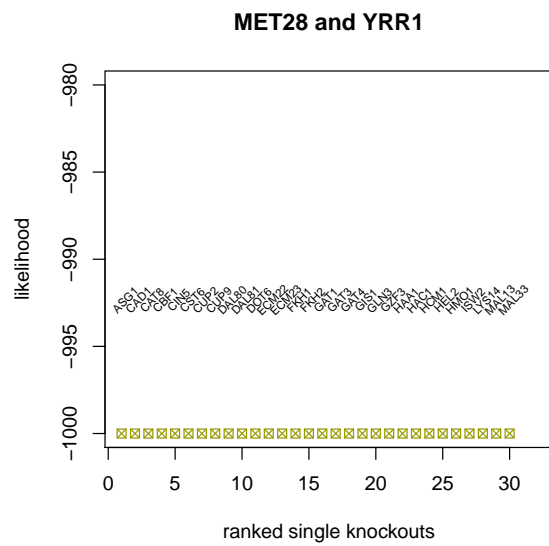
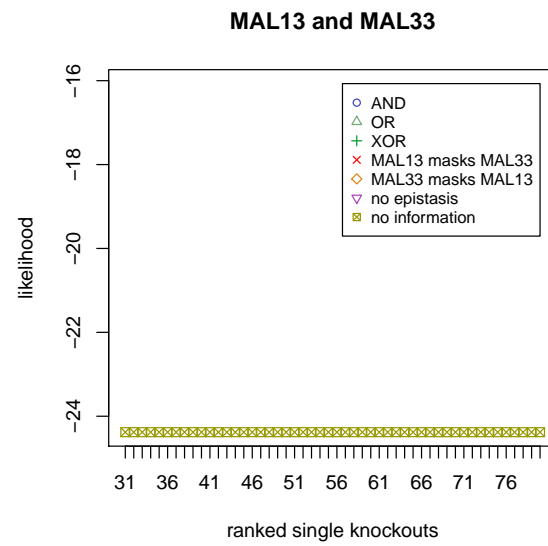
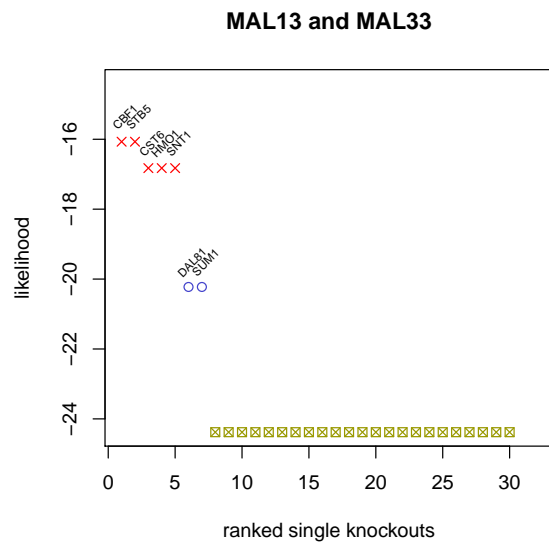


HAC1 and RPN4

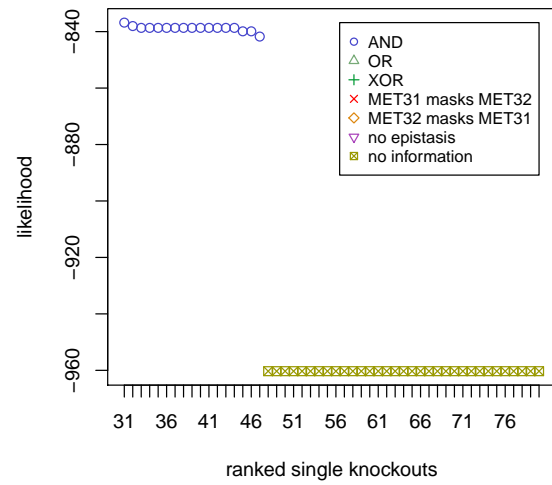




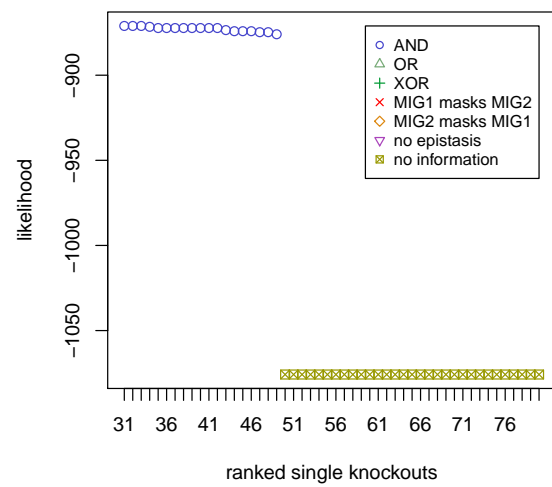


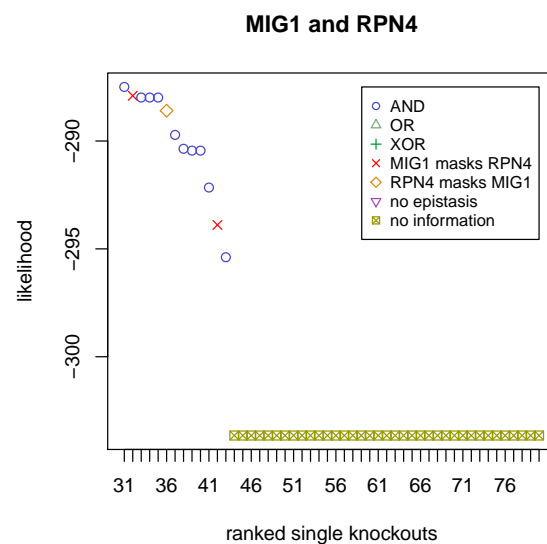
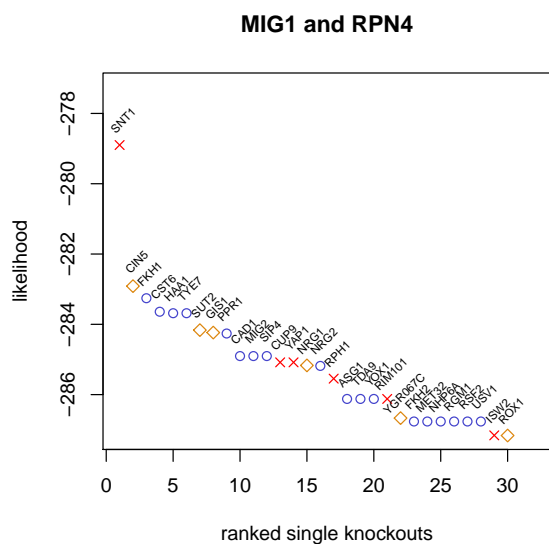
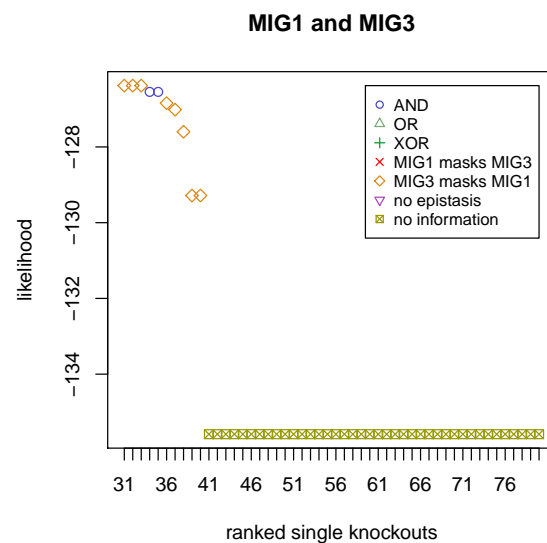
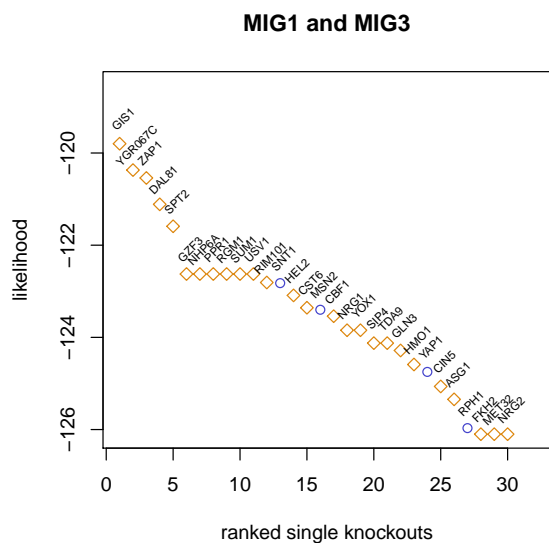


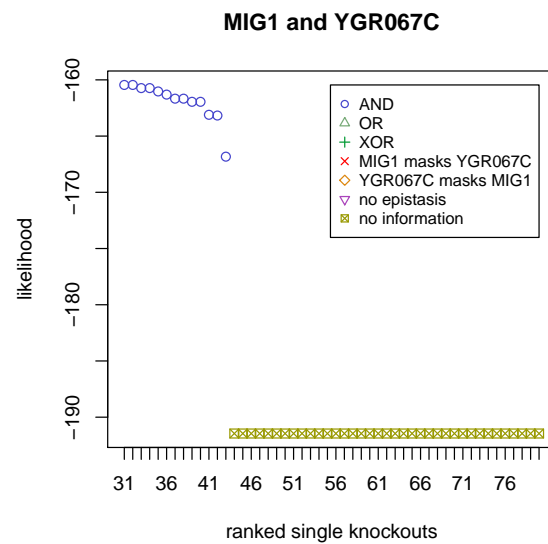
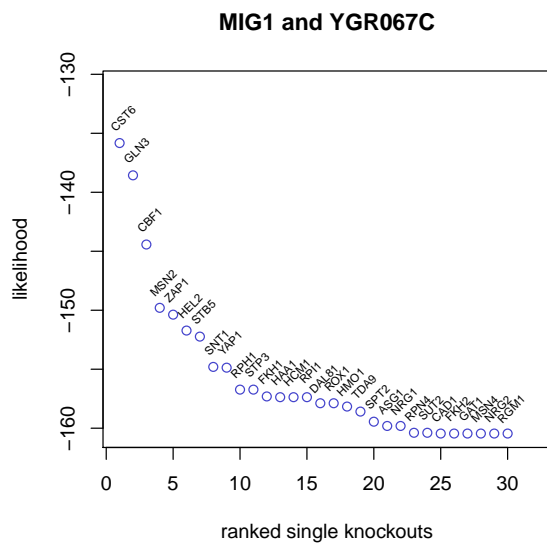
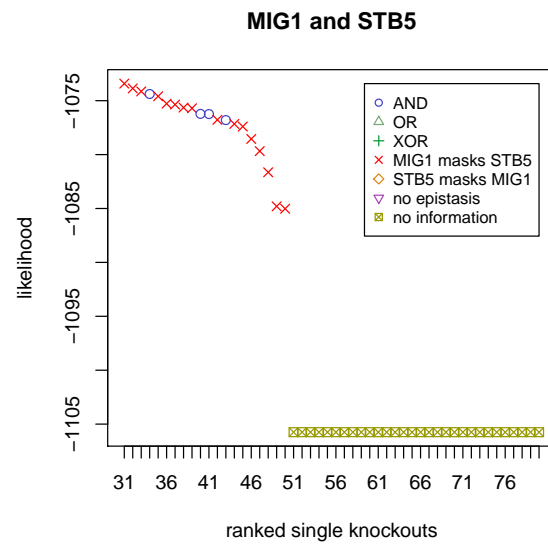
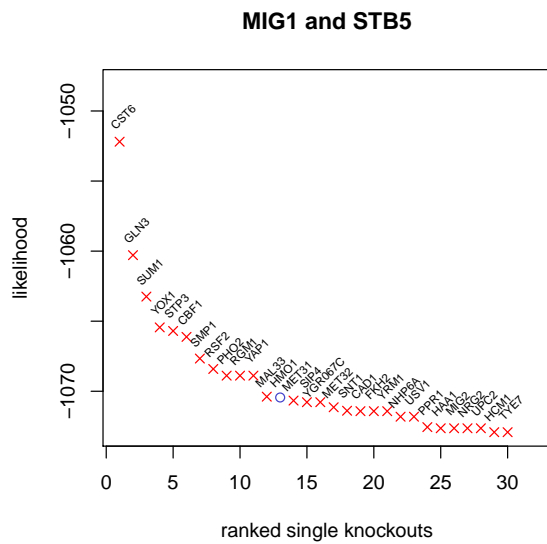
MET31 and MET32



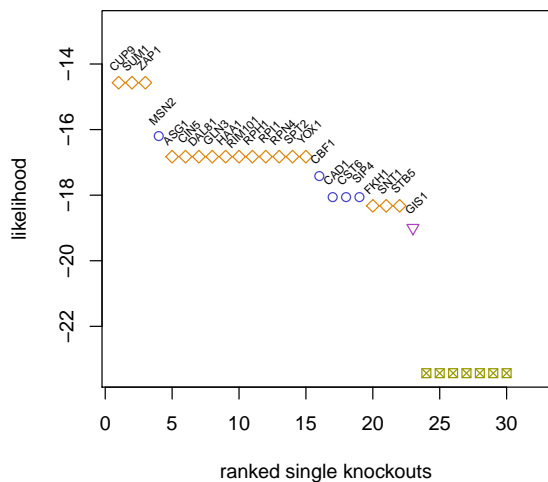
MIG1 and MIG2



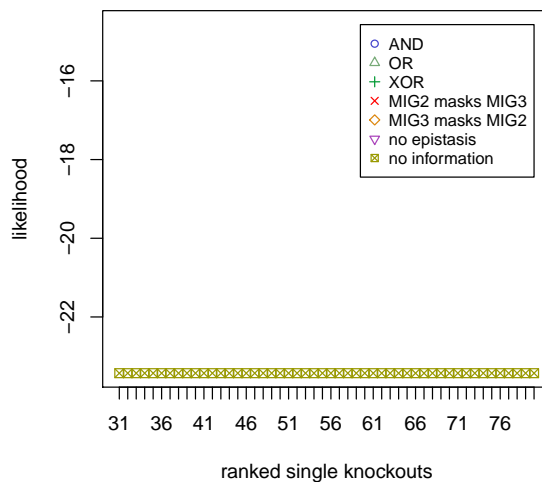




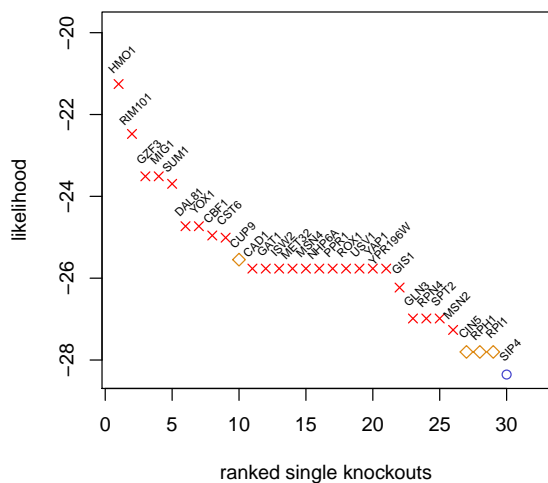
MIG2 and MIG3



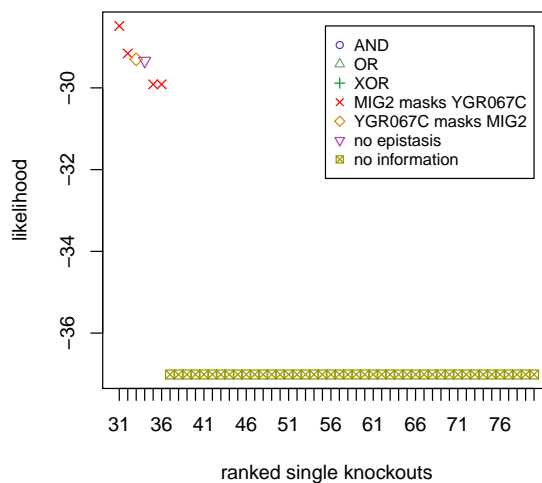
MIG2 and MIG3

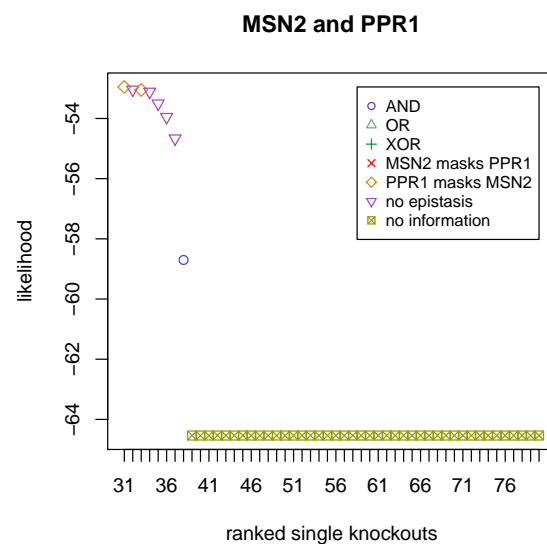
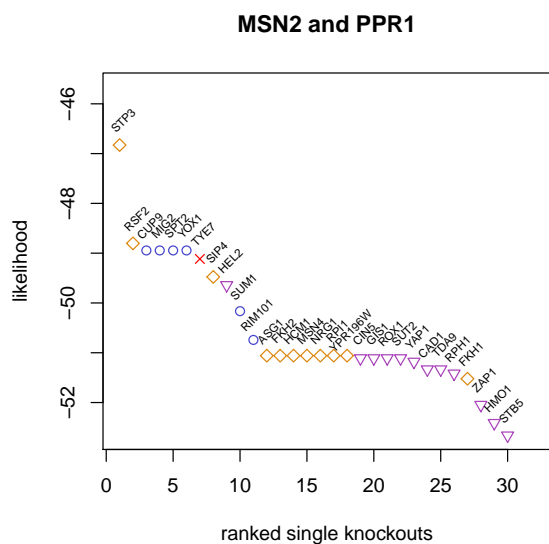
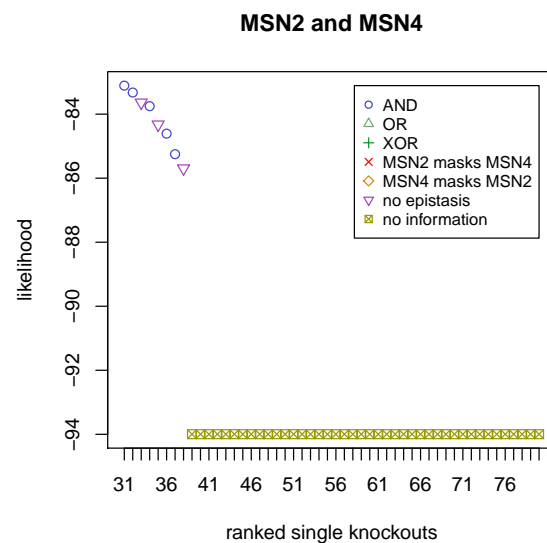
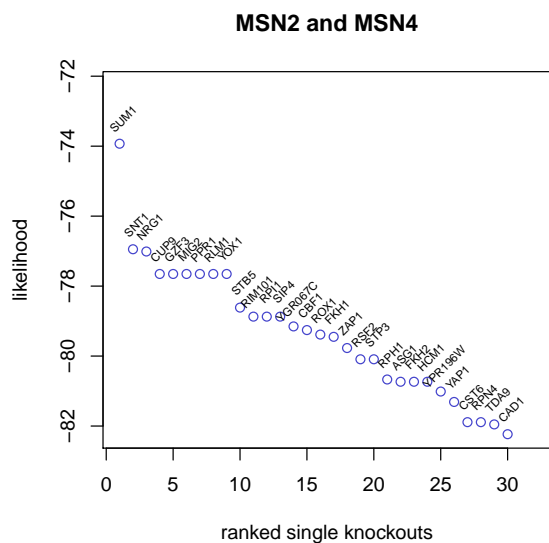


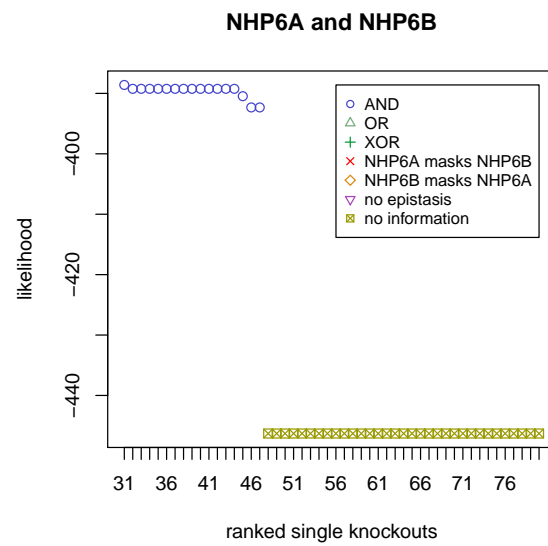
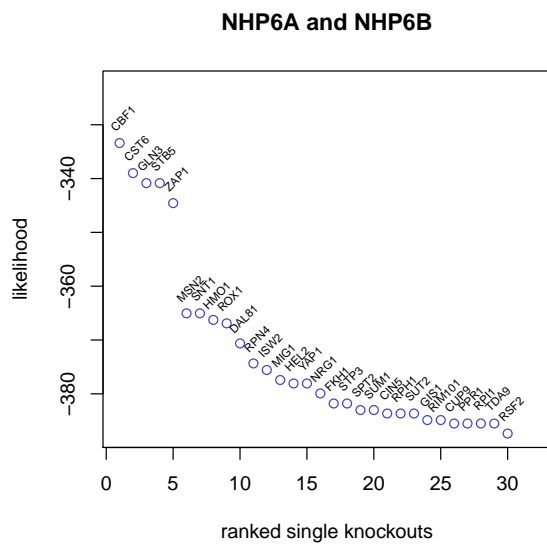
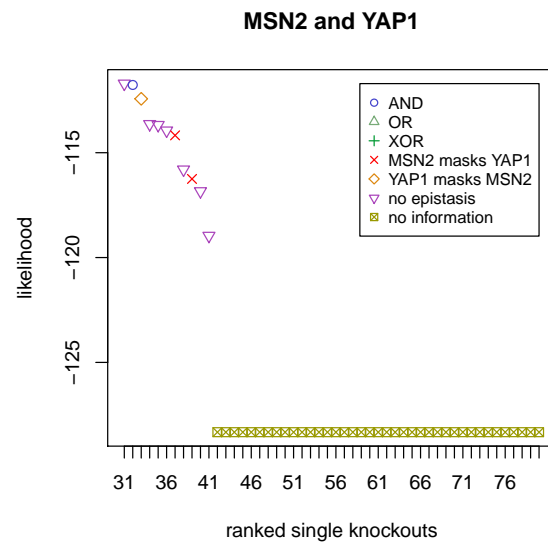
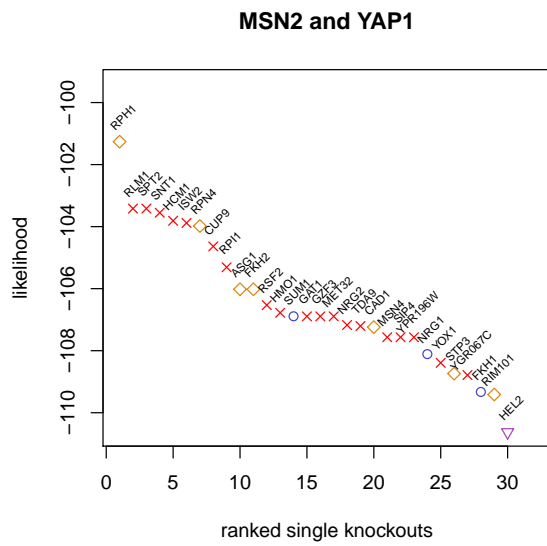
MIG2 and YGR067C

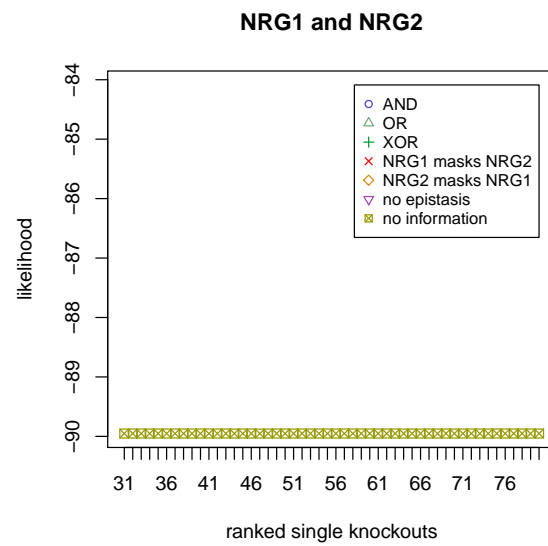
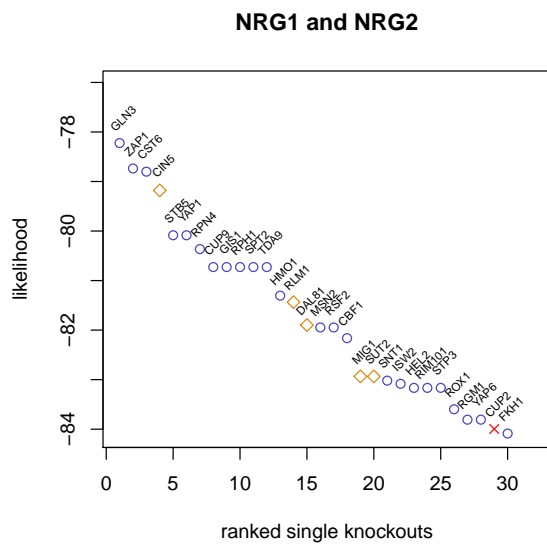
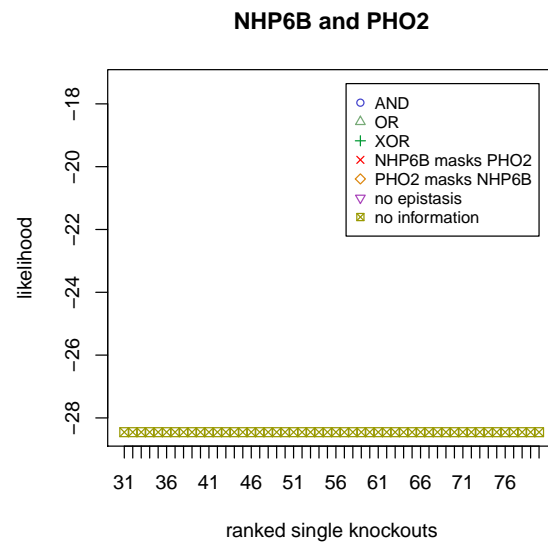
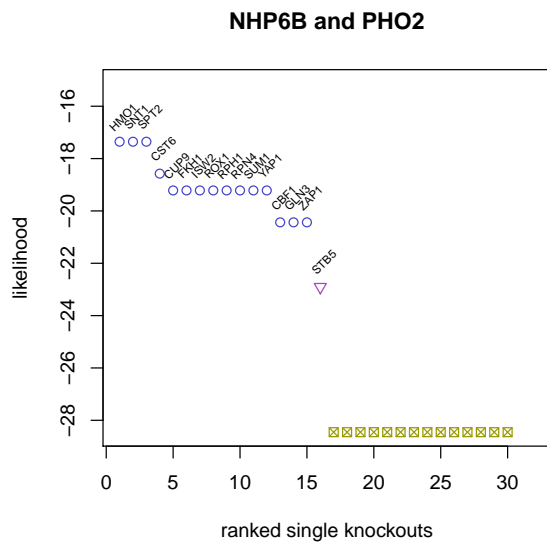


MIG2 and YGR067C

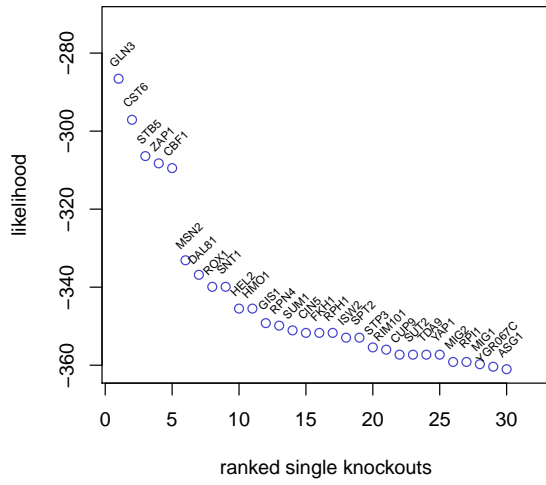




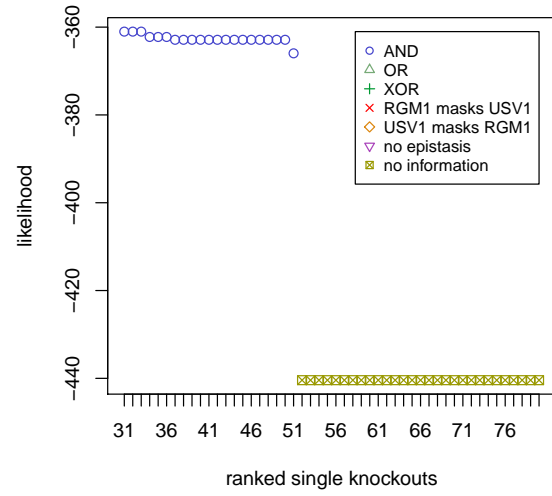




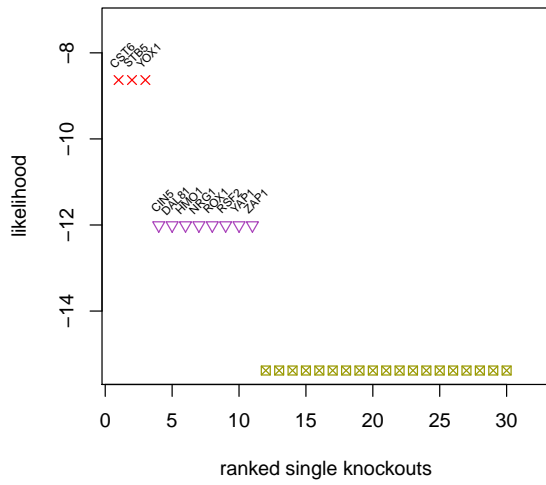
RGM1 and USV1



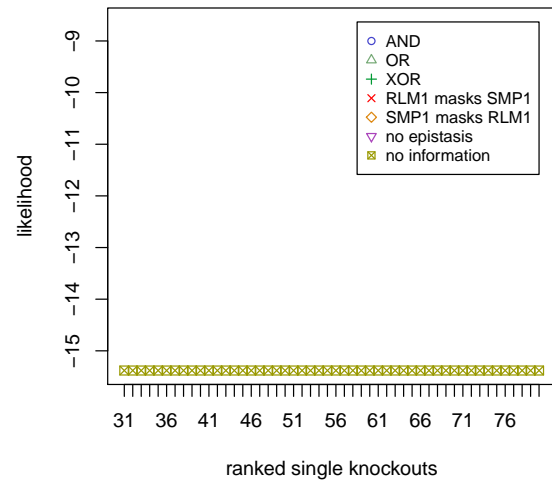
RGM1 and USV1

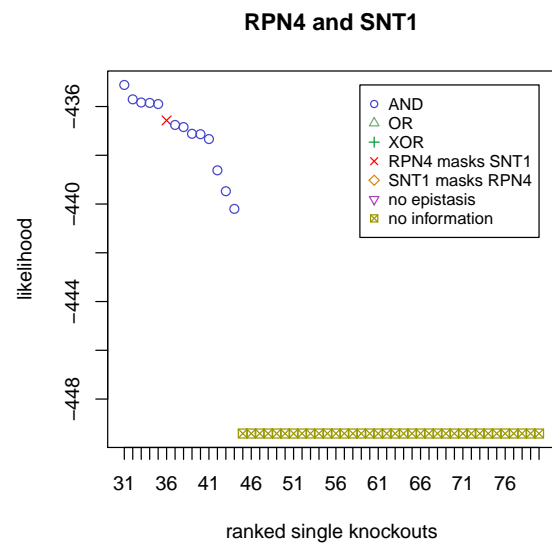
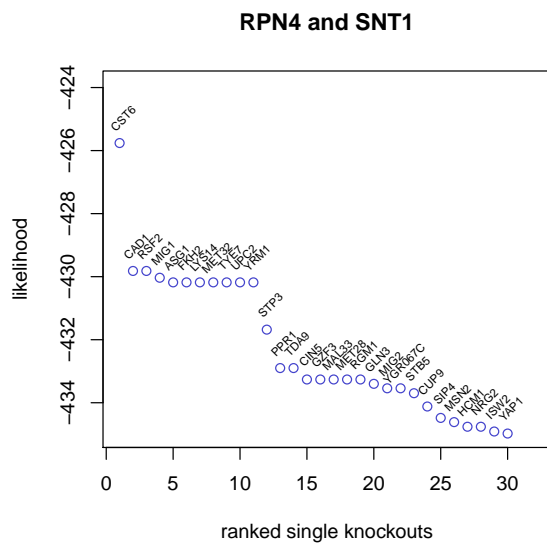
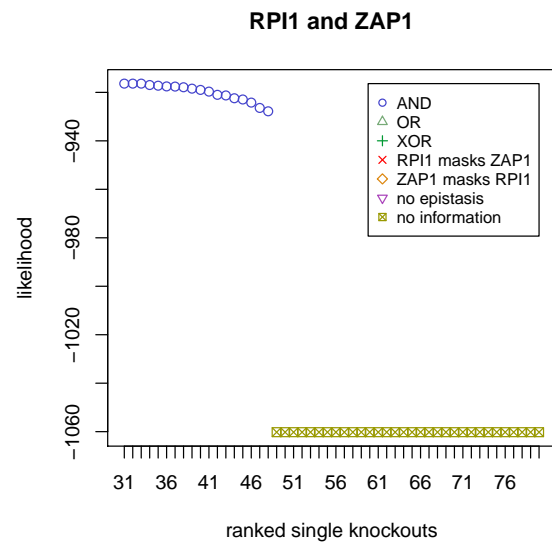
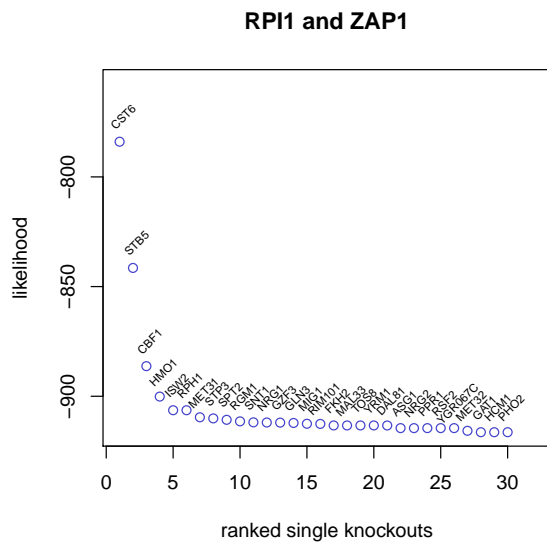


RLM1 and SMP1

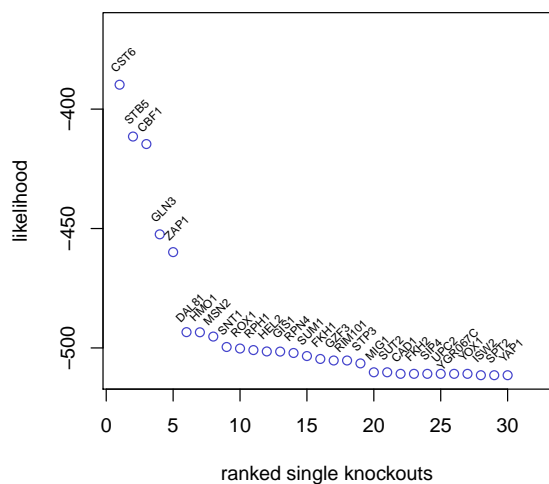


RLM1 and SMP1

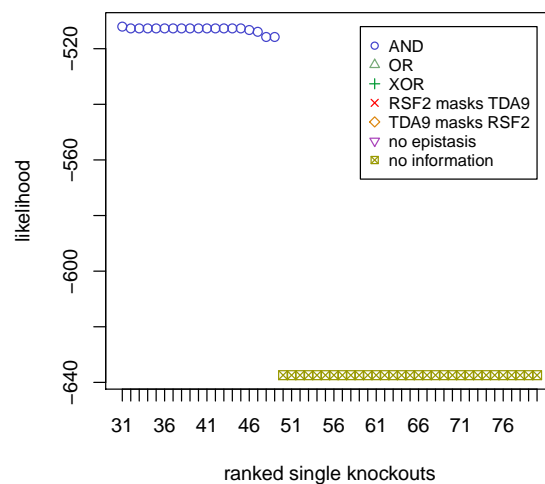




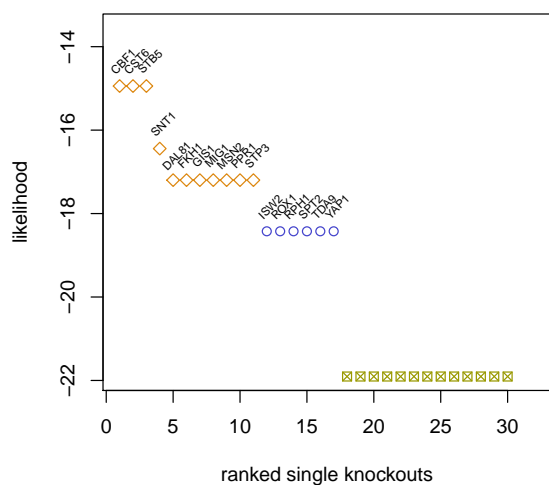
RSF2 and TDA9



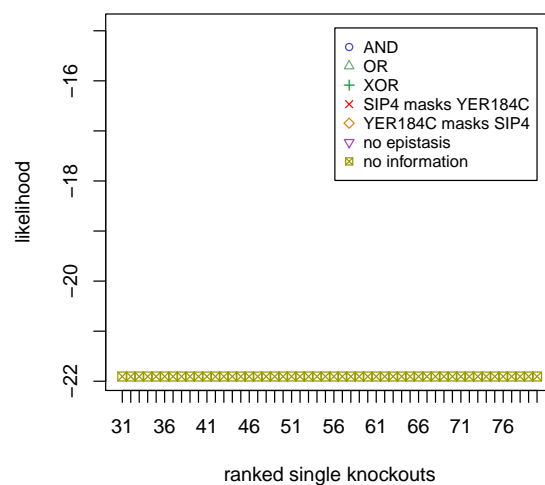
RSF2 and TDA9



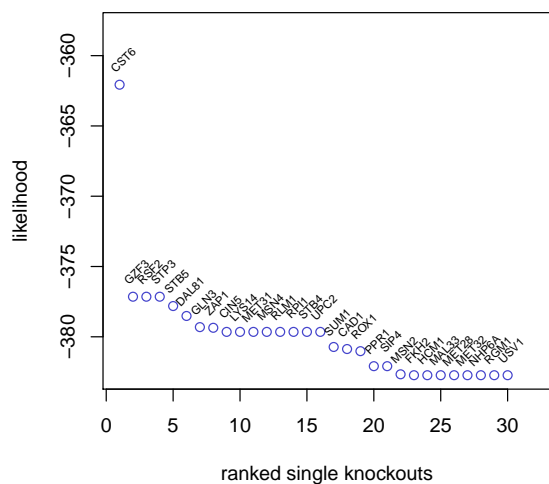
SIP4 and YER184C



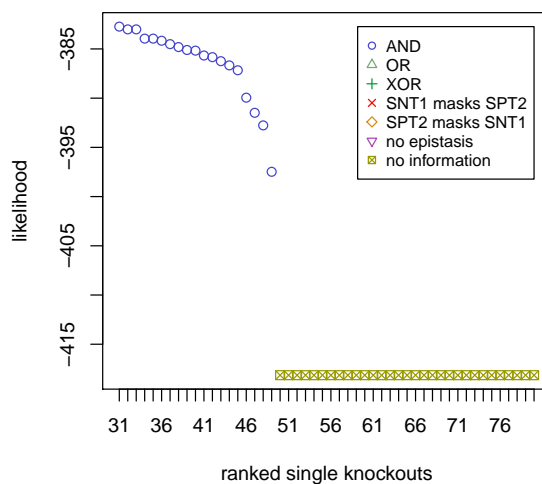
SIP4 and YER184C



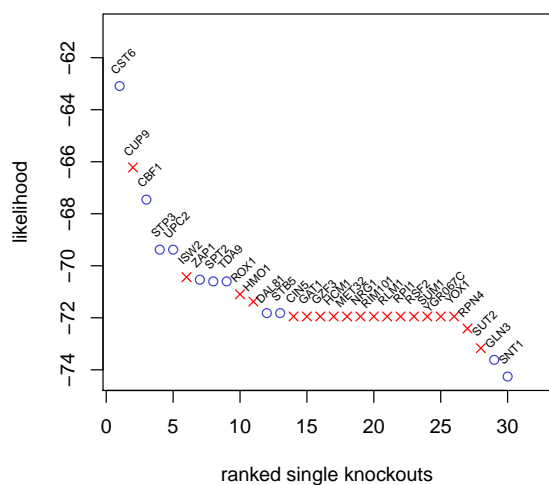
SNT1 and SPT2



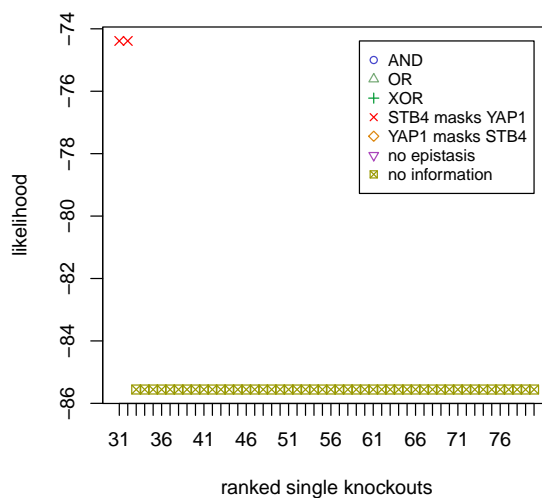
SNT1 and SPT2

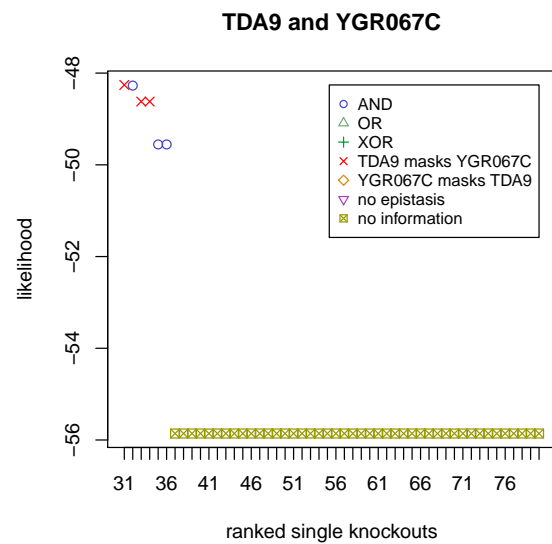
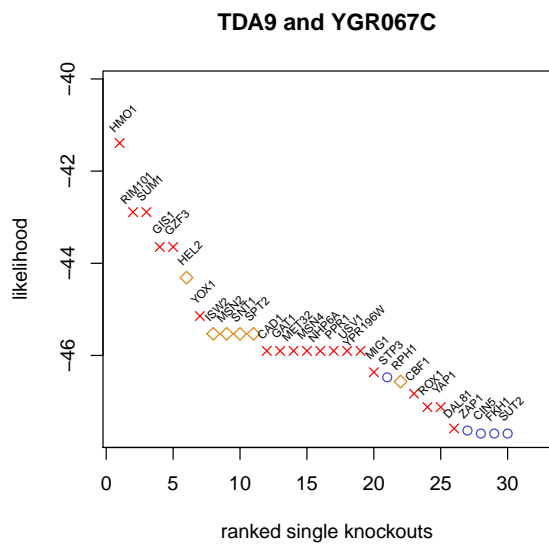
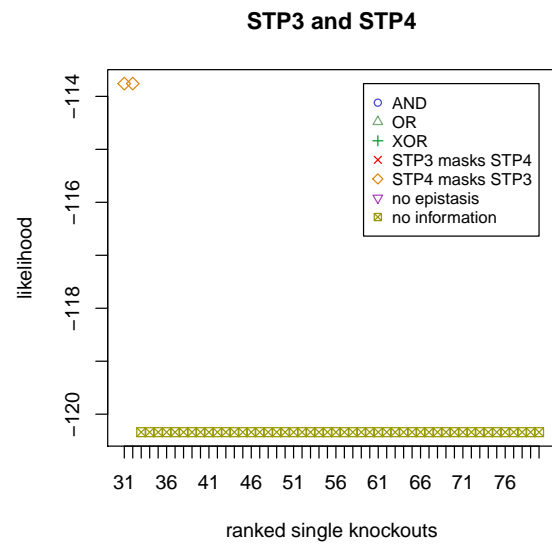
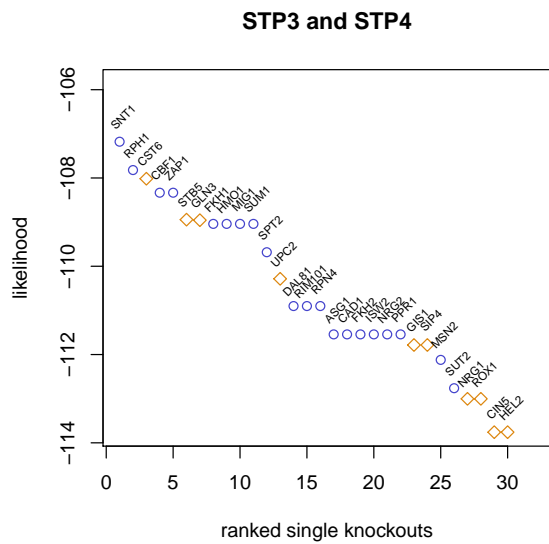


STB4 and YAP1

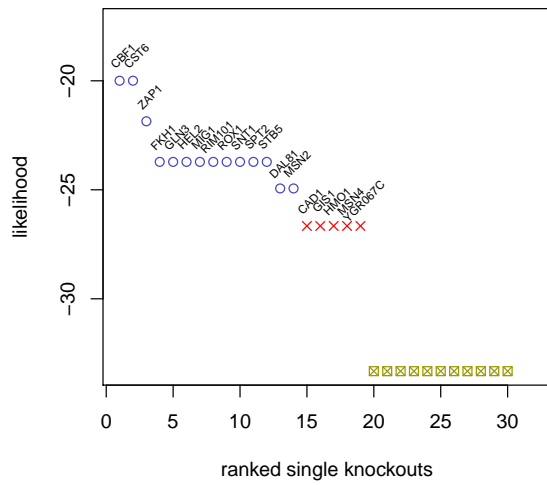


STB4 and YAP1

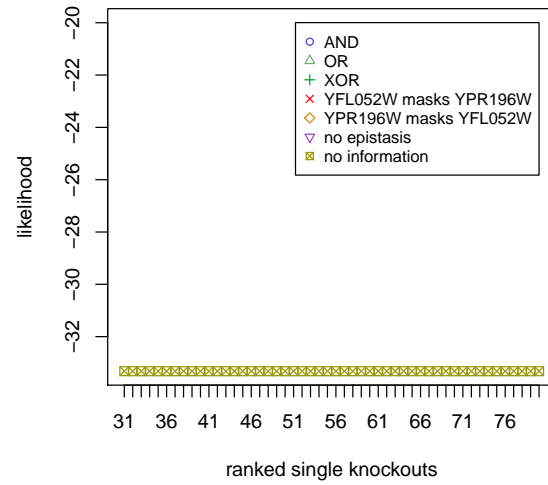




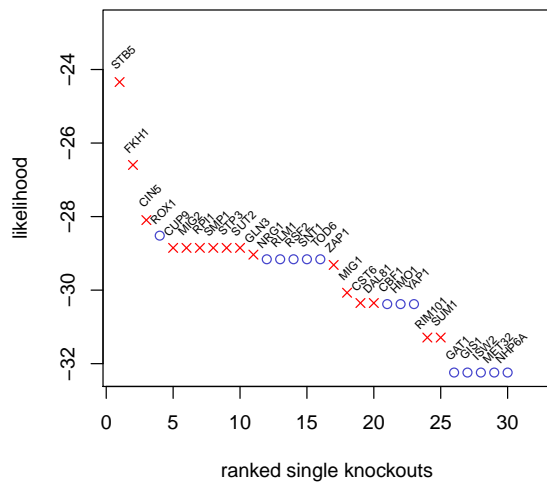
YFL052W and YPR196W



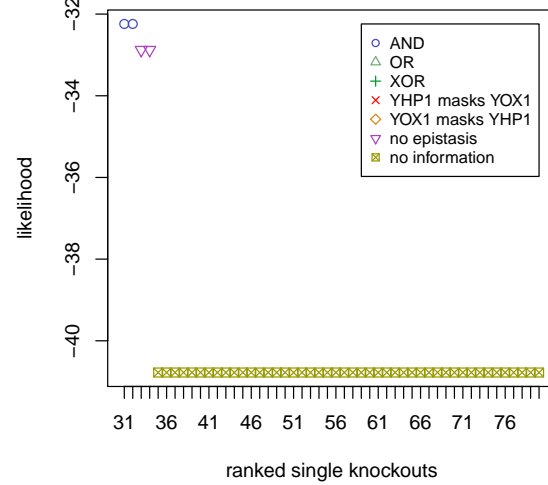
YFL052W and YPR196W

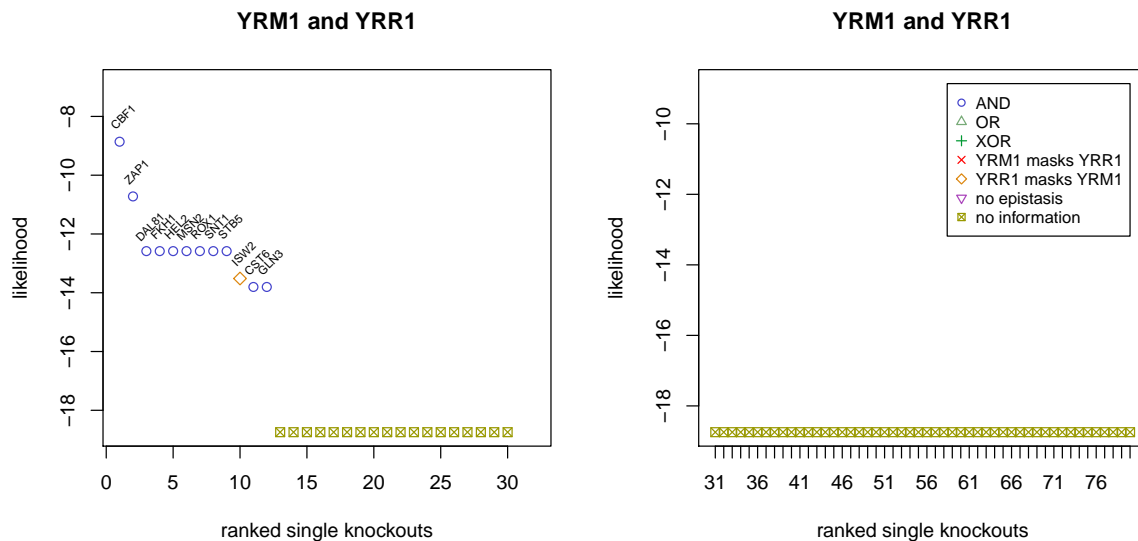


YHP1 and YOX1



YHP1 and YOX1





```

distmat <- sameith$logic

distmat[which(distmat %in% "AND")] <- 1
distmat[which(distmat %in% "OR")] <- 2
distmat[which(distmat %in% "XOR")] <- 3
distmat[which(distmat %in% "NOEPI")] <- 6
distmat[which(distmat %in% c("NOINFO", "NOINF"))] <- 7

for (i in 1:ncol(distmat)) {

  genes <- unlist(strsplit(colnames(distmat)[i], "\\."))

  distmat[which(distmat[, i] %in% paste(genes[1], " masks the effect of ", genes[2], sep = "")), i] <- 7

  distmat[which(distmat[, i] %in% paste(genes[2], " masks the effect of ", genes[1], sep = "")), i] <- 7

}

distmat <- apply(distmat, c(1,2), as.numeric)

for (i in 1:ncol(distmat)) {
  distmat[, i] <- rev(sort(distmat[, i]))
}

rownames(distmat) <- 1:nrow(distmat)

distmat <- distmat[-which(apply(distmat, 1, sum) == 0), ]

library(bnem)

y <- distmat

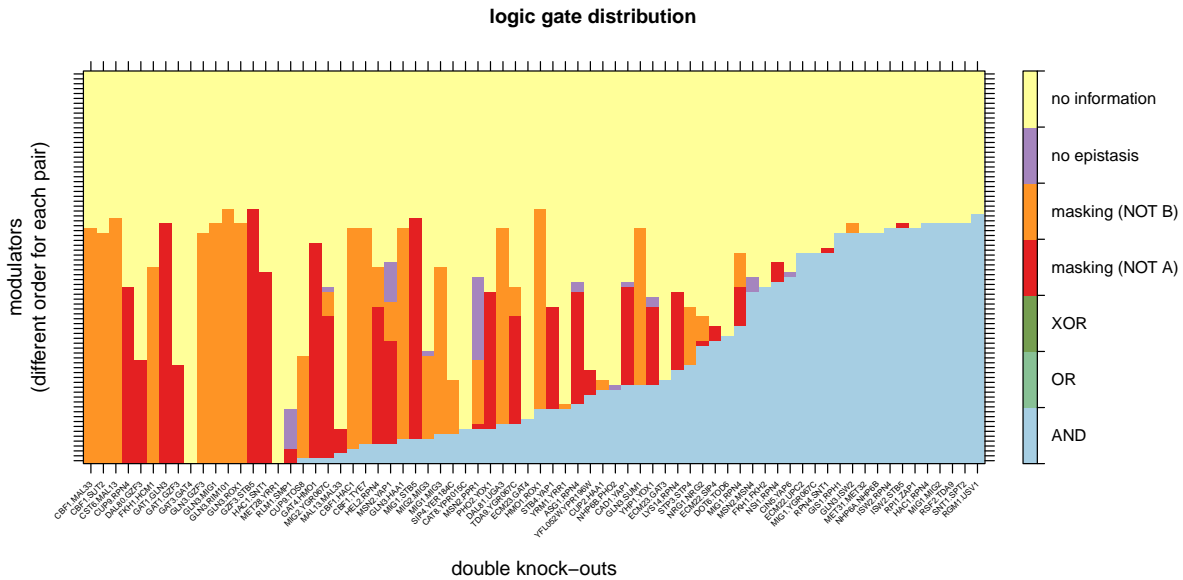
distmat <- distmat[, order(apply(distmat, 2, function(x) { return(sum(x == 1)) }))]

```



```
y[which(y == 5)] <- 4
```

```
heatmapOP(distmat, Colv = F, Rowv = F, main = "logic gate distribution", sub = "", col = "Paired", break
```



```
sessionInfo()
```

```
## R version 3.3.1 (2016-06-21)
## Platform: x86_64-apple-darwin13.4.0 (64-bit)
## Running under: OS X 10.11.5 (El Capitan)
##
## locale:
## [1] C/UTF-8/C/C/C/C
##
## attached base packages:
## [1] grid      parallel  stats      graphics  grDevices  utils      datasets  methods
## [9] base
##
## other attached packages:
## [1] pcalg_2.4-3          minet_3.32.0          bnem_0.99.0           latticeExtra_0.6-28
## [5] RColorBrewer_1.1-2   lattice_0.20-34       snowfall_1.84-6.1     snow_0.4-2
## [9] matrixStats_0.51.0  nem_2.48.0            CellNOptR_1.20.0      XML_3.98-1.5
## [13] Rgraphviz_2.18.0     RCurl_1.95-4.8        bitops_1.0-6          ggplot2_2.2.0
## [17] hash_2.2.6           RBGL_1.50.0           graph_1.52.0          BiocGenerics_0.20.0
## [21] epiNEM_0.99.0        igraph_1.0.1          gtools_3.5.0          e1071_1.6-7
## [25] BoolNet_2.1.1        knitr_1.15            devtools_1.12.0
##
## loaded via a namespace (and not attached):
## [1] statmod_1.4.26       colorspace_1.3-0      stats4_3.3.1          fastICA_1.2-0
## [5] gmp_0.5-12           withr_1.0.2           plyr_1.8.4           robustbase_0.92-6
## [9] stringr_1.1.0        munsell_0.4.3         gtable_0.2.0         bdsmatrix_1.3-2
## [13] memoise_1.0.0        evaluate_0.10         ggm_2.3              BiocInstaller_1.24.0
## [17] class_7.3-14         highr_0.6             DEoptimR_1.0-6       Rcpp_0.12.8
## [21] corpcor_1.6.8        scales_0.4.1          limma_3.30.4         plotrix_3.6-3
## [25] abind_1.4-5          digest_0.6.10         stringi_1.1.2        clue_0.3-51
```

```
## [29] tools_3.3.1      magrittr_1.5      lazyeval_0.2.0    tibble_1.2
## [33] cluster_2.0.5    assertthat_0.1    boot_1.3-18       sfsmisc_1.1-0
```

References: