

Epistatic Nested Effects Models - Inferring mixed epistasis from indirect measurements of knockout screens.

Madeline Diekmann & Martin Pirkl

2017-01-12

Introduction

This package is an extension of the classic Nested Effects Models provided in package *nem*. Nested Effects Models is a pathway reconstruction method, which takes into account effects of downstream genes. Those effects are observed for every knockout of a pathway gene, and the nested structure of observed effects can then be used to reconstruct the pathway structure. However, classic Nested Effects Models do not account for double knockouts. In this package *epiNEM*, one additional layer of complexity is added. For every two genes, acting on one gene together, the relationship is evaluated and added to the model as a logic gate. Genetic relationships are represented by the logics OR (no relationship), AND (functional overlap), NOT (masking or inhibiting) and XOR (mutual prevention from acting on gene C).

Loading epiNEM

```
install.packages("devtools", verbose = F, quiet = T)

library(devtools)

install_github("cbg-ethz/epiNEM", quiet = T)
```

```
library(epiNEM)
```

Simulations

We compare epiNEM to several network inference methods.

```
library(bnem, quietly = T, verbose = F) # install_github("MartinFXP/B-NEM/package")

library(nem)

library(minet)

library(pcalg)

runs <- 100

noiselvls <- c(0.01, 0.025, 0.05, 0.1, 0.2, 0.3, 0.4, 0.5)

random <- list(FPrate = 0.1, FNrate = noiselvls,
               single = 4, double = 1, reporters = 100, replicates = 3)
```

```

spec <- sens <- logics <- array(0, dim = c(2, runs, length(noiselvls)))

sens2 <- spec2 <- time <- array(0, dim = c(5, runs, length(noiselvls)))

do <- c("n", "p", "a")

do <- c("e", "b", do)

popSize <- 100

maxTime <- F

forcelogic <- T

epinemsearch <- "greedy"

nIterations <- 3

bnemsearch <- "genetic"

parallel <- NULL

logicgate <- matrix("", runs, length(noiselvls))

edgenr <- matrix(0, runs, length(noiselvls))

for (i in 1:runs) {

  print(paste("run ", i, sep = ""))

  for (j in 1:length(noiselvls)) {

    print(paste("noiselvl ", j, sep = ""))

    topology <- CreateTopology(random$single, random$double, force = forcelogic)

    topology <- unlist(unique(topology), recursive = FALSE)

    extTopology <- ExtendTopology(topology$model, random$reporters)

    sortedData <- GenerateData(topology$model, extTopology,
                              random$FPrate, random$FNrate[j], random$replicates)

    logicgate[i, j] <- paste(topology$logics, collapse = "_")

    edgenr[i, j] <- sum(topology$origModel == 1)

    if ("e" %in% do) {
      print("epiNEM")

      start <- Sys.time()
      TriplModel <- epiNEM(filename = sortedData,
                          method = epinemsearch, nIterations = nIterations)
    }
  }
}

```

```

time[1, i, j] <- difftime(Sys.time(), start, units = "secs")
print(time[1, i, j])

tp <- sum(topology$model == 1 & TriplModel$model == 1)
tn <- sum(topology$model == 0 & TriplModel$model == 0)
fp <- sum(topology$model == 0 & TriplModel$model == 1)
fn <- sum(topology$model == 1 & TriplModel$model == 0)
sens[1, i, j] <- tp/(tp+fn)
spec[1, i, j] <- tn/(tn+fp)
tp <- sum(topology$origModel == 1 & TriplModel$origModel == 1)
tn <- sum(topology$origModel == 0 & TriplModel$origModel == 0)
fp <- sum(topology$origModel == 0 & TriplModel$origModel == 1)
fn <- sum(topology$origModel == 1 & TriplModel$origModel == 0)
sens2[1, i, j] <- tp/(tp+fn)
spec2[1, i, j] <- tn/(tn+fp)
tp <- 0
for (k in 1:length(topology$column)) {
  for (l in 1:length(TriplModel$column)) {
    if (topology$column[k] == TriplModel$column[l]) {
      if (topology$logics[k] %in% TriplModel$logics[l]) {
        tp <- tp + 1
      }
    }
  }
}
logics[1, i, j] <- tp/(length(topology$logics) +
                        length(TriplModel$logics) - tp)

print(sens[1, i, j])
print(spec[1, i, j])
print(sens2[1, i, j])
print(spec2[1, i, j])
print(logics[1, i, j])

}

if ("b" %in% do) {
  print("B-NEM")

  gtn <- epi2bg(topology)

  fc <- cbind(Ctrl_vs_S = -1, epi2bg(sortedData))*(-1)

  bmemnoise <- sample(1:nrow(fc), floor(nrow(fc)*random$FNrate[j]))

  fc[bmemnoise, 1] <- 0

  ers <- t(topology$model)*(-1)
  colnames(ers) <- paste("S_vs_S_",
                        gsub("\\.", "_", colnames(ers)), sep = "")
  ers <- cbind(Ctrl_vs_S = 1, ers)
  ers <- ers[, order(colnames(ers))]

  CNOList <- dummyCNOList(stimuli = "S",

```

```

        inhibitors = LETTERS[1:random$single],
        maxStim = 1, maxInhibit = 2,
        signals = LETTERS[1:random$single])

parents <- unique(unlist(strsplit(colnames(sortedData)[grep("\\\\.",
        colnames(sortedData))], "\\.")))

nodes <- unique(colnames(sortedData)[-grep("\\\\.", colnames(sortedData))])

child <- nodes[-which(nodes %in% parents)]

sifMatrix <- NULL
for (k in LETTERS[1:random$single]) {
  sifMatrix <- rbind(sifMatrix, c("S", "1", k))#, c("S", "-1", k))
  for (l in LETTERS[1:random$single]) {
    if (k %in% l) { next() }
    if (k %in% parents) {
      sifMatrix <- rbind(sifMatrix, c(k, "1", l), c(k, "-1", l))
    } else {
      sifMatrix <- rbind(sifMatrix, c(k, "1", l))
    }
  }

randfile <- paste("pkn_", as.numeric(Sys.time()), sep = "")
write.table(sifMatrix, file = randfile, sep = "\t",
  row.names = FALSE, col.names = FALSE, quote = FALSE)
PKN <- readSIF(randfile)
unlink(randfile)

model <- preprocessing(CNolist, PKN)

initBstring <- absorption(rep(1, length(model$reacID)), model)

if (maxTime) { maxTime2 <- time[1, i, j] } else { maxTime2 <- Inf }

start <- Sys.time()
bga <- bnem(search = bnemsearch,
  fc=fc,
  CNolist=CNolist,
  model=model,
  initBstring=initBstring,
  draw = F,
  verbose = F,
  popSize = popSize,
  maxTime = maxTime2,
  parallel = parallel
)
time[2, i, j] <- difftime(Sys.time(), start, units = "secs")
print(time[2, i, j])

ers2 <- computeFc(CNolist, t(simulateStatesRecursive(CNolist,
  model, bga$bString)))

ers2 <- ers2[, unique(colnames(fc))]
ers2 <- ers2[, order(colnames(ers2))]

```

```

tp <- sum(ers == -1 & ers2 == -1)
tn <- sum(ers == 0 & ers2 == 0)
fn <- sum(ers == -1 & ers2 == 0)
fp <- sum(ers == 0 & ers2 == -1)
sens[2, i, j] <- tp/(tp+fn)
spec[2, i, j] <- tn/(tn+fp)
gtn2 <- abs(dnf2adj(gtn))
if (length(grep("S", rownames(gtn2))) > 0) {
  gtn2 <- gtn2[-grep("S", rownames(gtn2)), -grep("S", colnames(gtn2))]
}
gtn2 <- gtn2[order(rownames(gtn2)), order(colnames(gtn2))]
res <- abs(dnf2adj(bga$graph))
if (length(grep("S", rownames(res))) > 0) {
  res <- as.matrix(res[-grep("S", rownames(res)),
    -grep("S", colnames(res))])
}
if (dim(res)[1] == 1) {
  colnames(res) <- rownames(res) <- gsub(".*=", "", bga$graph)
} else {
  res <- res[order(rownames(res)), order(colnames(res))]
}
if (nrow(res) < nrow(gtn2)) {
  res2 <- rbind(cbind(res, matrix(0, nrow(res), nrow(gtn2) - nrow(res))),
    matrix(0, nrow(gtn2) - nrow(res), ncol(gtn2)))
  colnames(res2)[(ncol(res)+1):ncol(res2)] <-
    colnames(gtn2)[which(!(colnames(gtn2)
      %in% colnames(res)))]
  rownames(res2)[(nrow(res)+1):nrow(res2)] <-
    rownames(gtn2)[which(!(rownames(gtn2)
      %in% rownames(res)))]
  res2 <- res2[order(rownames(res2)), order(colnames(res2))]
  res <- res2
}
diag(gtn2) <- diag(res) <- 0
tp <- sum(gtn2 == 1 & res == 1)
tn <- sum(gtn2 == 0 & res == 0)
fn <- sum(gtn2 == 1 & res == 0)
fp <- sum(gtn2 == 0 & res == 1)
sens2[2, i, j] <- tp/(tp+fn)
spec2[2, i, j] <- tn/(tn+fp)
tp <- sum(bga$graph %in% gtn)
logics[2, i, j] <- tp/(length(gtn) + length(bga$graph) - tp)
print(sens[2, i, j])
print(spec[2, i, j])
print(sens2[2, i, j])
print(spec2[2, i, j])
print(logics[2, i, j])

print(bga$graph)
print(gtn)
}

```

```

if (any(c("n", "p", "a") %in% do)) {

  reddata <- sortedData[, -grep("\\.", colnames(sortedData))]
  gtnadj <- topology$origModel
  gtnadj <- gtnadj[order(apply(gtnadj, 1, sum), decreasing = T),
                  order(apply(gtnadj, 2, sum), decreasing = F)]
  gtnadj[lower.tri(gtnadj)] <- gtnadj[upper.tri(gtnadj)]
  gtnadj <- gtnadj[order(rownames(gtnadj)), order(colnames(gtnadj))]
  eadj <- topology$origModel
  eadj <- eadj[order(rownames(eadj)), order(colnames(eadj))]
  reddata2 <- matrix(0, nrow(reddata)*random$replicates,
                    length(unique(colnames(reddata))))
  for (k in 1:length(unique(colnames(reddata)))) {
    reddata2[, k] <- as.vector(reddata[, which(colnames(reddata) %in%
                                              unique(colnames(reddata))[k])])
  }
  colnames(reddata2) <- unique(colnames(reddata))

}

if ("n" %in% do) {
  print("NEM")

  start <- Sys.time()
  if (epinemsearch %in% "greedy") {
    nemres <- nem(reddata, inference = "nem.greedy")
  } else {
    nemres <- nem(reddata, inference = "search")
  }
  nadj <- transitive.reduction(graph2adj(nemres$graph))
  time[3, i, j] <- difftime(Sys.time(), start, units = "secs")
  print(time[3, i, j])

  tp <- sum(eadj == 1 & nadj == 1)
  tn <- sum(eadj == 0 & nadj == 0)
  fp <- sum(eadj == 0 & nadj == 1)
  fn <- sum(eadj == 1 & nadj == 0)
  sens2[3, i, j] <- tp/(tp+fn)
  spec2[3, i, j] <- tn/(tn+fp)
  print(sens2[3, i, j])
  print(spec2[3, i, j])

}

if ("p" %in% do) {
  print("PCalg")

  start <- Sys.time()
  pc.fit <- pc(suffStat = list(C = cor(reddata2), n = nrow(reddata2)),
              indepTest = gaussCIttest, ## indep.test: partial correlations
              alpha=0.05, labels = colnames(reddata2), verbose = F)
  pcadj <- graph2adj(pc.fit@graph)
  time[4, i, j] <- difftime(Sys.time(), start, units = "secs")

```

```

    print(time[4, i, j])

    tp <- sum(gtnadj == 1 & pcadj == 1)
    tn <- sum(gtnadj == 0 & pcadj == 0)
    fp <- sum(gtnadj == 0 & pcadj == 1)
    fn <- sum(gtnadj == 1 & pcadj == 0)
    sens2[4, i, j] <- tp/(tp+fn)
    spec2[4, i, j] <- tn/(tn+fp)
    print(sens2[4, i, j])
    print(spec2[4, i, j])

  }

  if ("a" %in% do) {
    print("Aracne")

    start <- Sys.time()
    ares <- build.mim(reddata2)
    ares <- aracne(ares)
    ares <- disc(ares, 0)
    ares <- ares[order(rownames(ares)), order(colnames(ares))]
    nas <- which(is.na(ares) == T)
    ares[nas] <- 0
    diag(ares) <- 0
    time[5, i, j] <- difftime(Sys.time(), start, units = "secs")
    print(time[5, i, j])

    tp <- sum(gtnadj == 1 & ares == 1)
    tn <- sum(gtnadj == 0 & ares == 0)
    fp <- sum(gtnadj == 0 & ares == 1)
    fn <- sum(gtnadj == 1 & ares == 0)
    sens2[5, i, j] <- tp/(tp+fn)
    spec2[5, i, j] <- tn/(tn+fp)
    print(sens2[5, i, j])
    print(spec2[5, i, j])

  }

}

data(sim)

colvec <- c(rep("orange", length(noiselvls)), rep("blue", length(noiselvls)),
            rep("darkgreen", length(noiselvls)), rep("brown", length(noiselvls)),
            rep("darkgrey", length(noiselvls)))

acc <- (sens + spec)/2

acc2 <- (sens2 + spec2)/2

m <- rbind(c(1,1), c(2,2), c(3,4))

```

```

layout(m)

timeframe <- as.data.frame(
  cbind(data.frame(epiNEM = time[1,,]),
        data.frame(BNEM = time[2,,]), data.frame(NEM = time[3,,]),
        data.frame(Cor = time[4,,]), data.frame(MI = time[5,,])))

colnames(timeframe) <- rep(noiselvls, 5)

boxplot(timeframe, col = colvec, main = "running time", ylab = "seconds")

abline(v=(1:(length(do)-1)*length(noiselvls) + 0.5), col = "black", lty = 6)

axis(1, c(3, 11, 19, 28, 36)+1, c("epiNEM", "B-NEM", "NEM", "PC Algorithm", "ARACNE"),
     tick = F, pos = -25)

accframe2 <- as.data.frame(
  cbind(data.frame(epiNEM = acc2[1,,]),
        data.frame(BNEM = acc2[2,,]), data.frame(NEM = acc2[3,,]),
        data.frame(Cor = acc2[4,,]), data.frame(MI = acc2[5,,])))

colnames(accframe2) <- rep(noiselvls, 5)

boxplot(accframe2, col = colvec, main = "accuracy of the inferred edges", ylim = c(0,1))

abline(v=(1:(length(do)-1)*length(noiselvls) + 0.5), col = "black", lty = 6)

axis(1, c(3, 11, 19, 28, 36)+1, c("epiNEM", "B-NEM", "NEM", "PC Algorithm", "ARACNE"),
     tick = F, pos = -0.2)

## logical nems:

colvec2 <- c(rep("orange", length(noiselvls)), rep("blue", length(noiselvls)))

logicsframe <- as.data.frame(cbind(data.frame(epiNEM = logics[1,,]),
                                       data.frame(BNEM = logics[2,,])))

colnames(logicsframe) <- rep(noiselvls, 2)

boxplot(logicsframe, col = colvec2, main = "accuracy of the inferred logic gate",
        ylim = c(0,1))

abline(v=length(noiselvls)+0.5, col = "black", lty = 6)

axis(1, c(3, 11, 19, 28, 36)+1, c("epiNEM", "B-NEM", "NEM", "PC Algorithm", "ARACNE"),
     tick = F, pos = -0.2)

accframe <- as.data.frame(cbind(data.frame(epiNEM = acc[1,,]),
                                       data.frame(BNEM = acc[2,,])))

colnames(accframe) <- rep(noiselvls, 2)

boxplot(accframe, col = colvec2, main = "accuracy of the inferred expected data",

```



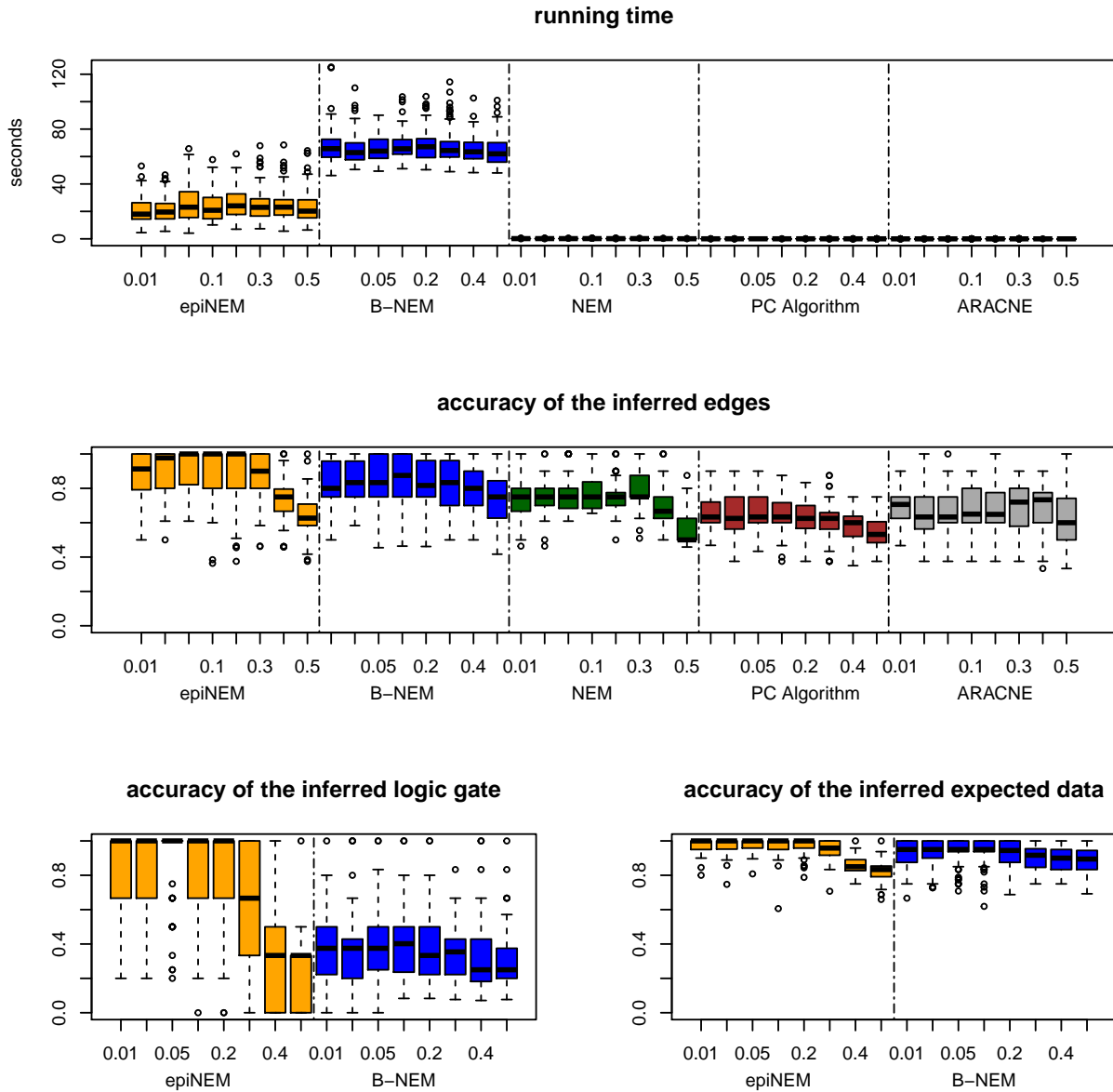
```

ylim = c(0,1))

abline(v=length(noiselvls)+0.5, col = "black", lty = 6)

axis(1, c(3, 11, 19, 28, 36)+1, c("epiNEM", "B-NEM", "NEM", "PC Algorithm", "ARACNE"),
     tick = F, pos = -0.2)

```

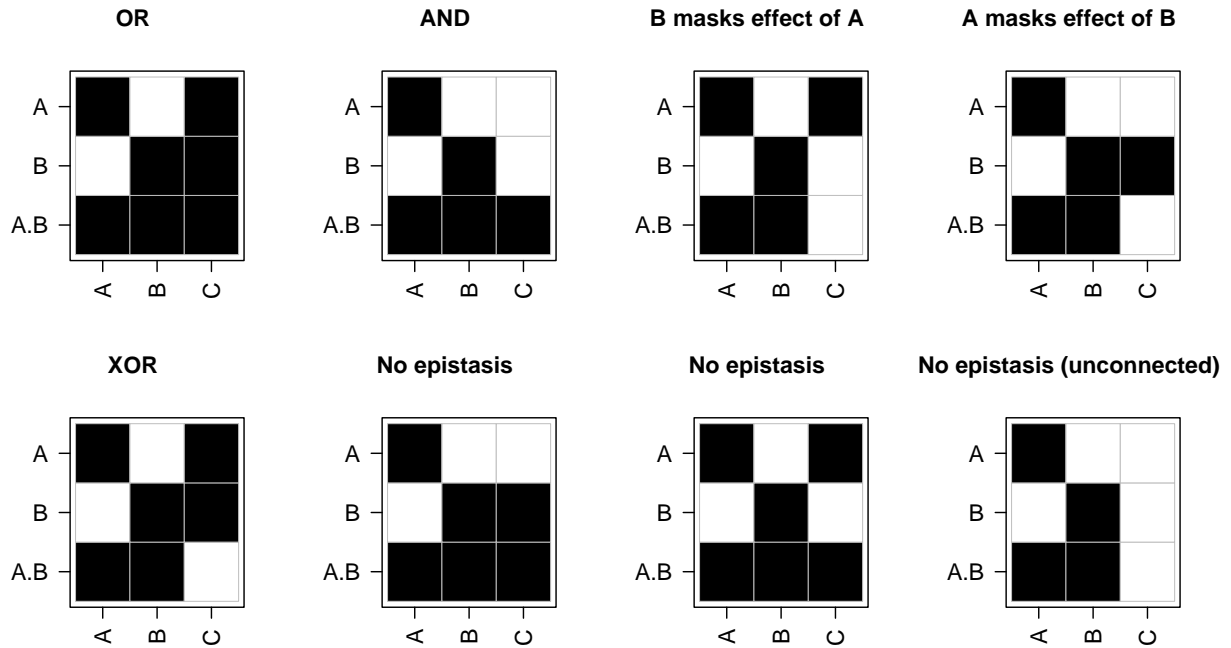


Yeast knockout screens

In this section we analyse previously published yeast knockout screens. The screens consist of gene expression data derived from double and single knockout mutants. We use epiNEM on each double mutant combined with each single mutant.\

The results of the knockout screens have been annotated according to the following legend:

```
a1 <- heatmapOP(matrix(c(1,-1,1,-1,1,1, 1, 1, 1), 3, 3,
  dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), Colv = F, Rowv = F,
  main = "OR", col = "Greys", sub = "", colorkey = NULL)
a2 <- heatmapOP(matrix(c(1,-1,1,-1,1,1, -1, -1, 1), 3, 3,
  dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), Colv = F, Rowv = F,
  main = "AND", col = "Greys", sub = "", colorkey = NULL)
a3 <- heatmapOP(matrix(c(1,-1,1,-1,1,1, 1, -1, -1), 3, 3,
  dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), Colv = F, Rowv = F,
  main = "B masks effect of A", col = "Greys", sub = "", colorkey = NULL)
a4 <- heatmapOP(matrix(c(1,-1,1,-1,1,1, -1, 1, -1), 3, 3,
  dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), Colv = F, Rowv = F,
  main = "A masks effect of B", col = "Greys", sub = "", colorkey = NULL)
a5 <- heatmapOP(matrix(c(1,-1,1,-1,1,1, 1, 1, -1), 3, 3,
  dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), Colv = F, Rowv = F,
  main = "XOR", col = "Greys", sub = "", colorkey = NULL)
a6 <- heatmapOP(matrix(c(1,-1,1,-1,1,1, -1, 1, 1), 3, 3,
  dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), Colv = F, Rowv = F,
  main = "No epistasis", col = "Greys", sub = "", colorkey = NULL)
a7 <- heatmapOP(matrix(c(1,-1,1,-1,1,1, 1, -1, 1), 3, 3,
  dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), Colv = F, Rowv = F,
  main = "No epistasis", col = "Greys", sub = "", colorkey = NULL)
a8 <- heatmapOP(matrix(c(1,-1,1,-1,1,1, -1, -1, -1), 3, 3,
  dimnames = list(c("A", "B", "A.B"), LETTERS[1:3])), Colv = F, Rowv = F,
  main = "No epistasis (unconnected)", col = "Greys", sub = "", colorkey = NULL)
print(a5, position = c(0,0, .25, .5), more = TRUE)
print(a6, position = c(.25,0, .5, .5), more = TRUE)
print(a7, position = c(.5,0, .75, .5), more = TRUE)
print(a8, position = c(.75,0, 1, .5), more = TRUE)
print(a1, position = c(0,.5, .25, 1), more = TRUE)
print(a2, position = c(.25,.5, .5, 1), more = TRUE)
print(a3, position = c(.5,.5, .75, 1), more = TRUE)
print(a4, position = c(.75,.5, 1, 1))
```



Wageningen et al., 2010

```
file <-
  "http://www.holstegelab.nl/publications/sv/signaling_redundancy/downloads/DataS1.txt"

data <- read.delim(file)

dataM <- data[-(1:2), (1+(1:(324/2))*2)]
dataP <- data[-(1:2), (2+(1:(324/2))*2)]

dataM <- dataM[-1, ]
dataP <- dataP[-1, ]

dataM <- apply(dataM, c(1,2), as.numeric)
dataP <- apply(dataP, c(1,2), as.numeric)

dataBin <- dataM

sig <- 0.05

cutoff <- log2(1.7)

dataBin[which(dataP < sig & dataP > 0 & abs(dataM) >= cutoff)] <- 1

dataBin[which(dataP >= sig | dataP == 0 | abs(dataM) < cutoff)] <- 0
```

```

dataBin <- dataBin[~which(apply(dataBin, 1, max) == 0), ]

dataBinWag <- dataBin

genelist <- toupper(c('hsl1', 'cla4', 'gin4', 'swe1', 'hsl1.cla4'))

colnames(dataBin) <- gsub(".del.vs..wt", "", colnames(dataBin))

colnames(dataBin) <- gsub(".del", "", colnames(dataBin))

doubles <- colnames(dataBin)[grep("\\.", colnames(dataBin))]

doubles <- sort(doubles[-grep("vs", doubles)])

doubles.genes <- unique(unlist(strsplit(doubles, "\\.")))

singles <- colnames(dataBin)[-grep("\\.", colnames(dataBin))]

singles <- unique(sort(singles))

llmat <- logicmat <- matrix(0, length(singles), length(doubles))

rownames(llmat) <- rownames(logicmat) <- singles

colnames(llmat) <- colnames(logicmat) <- doubles

globalgenes <- which(apply(dataBin, 1, max) == 1)

for (i in doubles) {
  if (which(doubles %in% i) == 8) { next() }
  print(i)
  doubles.singles <- unlist(strsplit(i, "\\."))
  egenes <- which(apply(dataBin[, which(colnames(dataBin) %in%
    c(i, doubles.singles))], 1, max) == 1)
  for (j in singles) {
    print(j)
    if (j %in% doubles.singles) { next() }

    dataTmp <- dataBin[, grep(paste(
      paste("^", c(i, j, doubles.singles), "$", sep = ""), collapse = "|"),
      colnames(dataBin))]

    if (path %in% "fixed_set") {
      dataTmp <- dataTmp[egenes, ]
    }
    if (path %in% "global") {
      dataTmp <- dataTmp[globalgenes, ]
    }
    if (path %in% "") {
      dataTmp <- dataTmp[which(apply(dataTmp, 1, max) == 1), ]
    }

    i1 <- which(singles %in% j)
  }
}

```

```

i2 <- which(doubles %in% i)

if (!(is.null(dim(dataTmp)))) {

  if (any(dataTmp[, j] != 0)) {

    epires <- epiNEM(dataTmp, method = "exhaustive")

    tmp <- epires$logics
    if ("OR" %in% tmp) {
      if (sum(epires$origModel[, j]) != 2) {
        tmp <- "NOEPI"
      } else {
        if (all(tmp %in% "OR")) {
          tmp <- "OR"
        } else {
          tmp <- tmp[which(!(tmp %in% "OR"))]
        }
      }
    }

    logicmat[i1, i2] <- tmp
    llmat[i1, i2] <- epires$score

  } else {

    logicmat[i1, i2] <- "UNCON"
    llmat[i1, i2] <- -Inf

  }

} else {

  logicmat[i1, i2] <- "UNCON"
  llmat[i1, i2] <- -Inf

}

}

}

```

```

palette(c("#4444cc", "#77aa77", "#009933", "#ff0000", "#dd8811", "#aa44bb", "#999900"))

```

```

data(wageningen_res)

```

```

llmat0 <- wageningen$ll

```

```

logicmat0 <- wageningen$logic

```

```

for (i in 1:length(doubles)) {

```

```

  if (i %in% 8) { next() }

```

```

logicvec <- logicmat0[, i]

llvec <- llmat0[, i]

logicvec <- logicvec[order(llvec, decreasing = T)]

llvec <- llvec[order(llvec, decreasing = T)]

parents <- unlist(strsplit(doubles[i], "\\\\"))

pchvec <- numeric(length(llvec))

pchvec[which(logicvec %in% "AND")] <- 1
pchvec[which(logicvec %in% "OR")] <- 2
pchvec[which(logicvec %in% "XOR")] <- 3
pchvec[grep(paste("^", parents[1], sep = ""), logicvec)] <- 4
pchvec[grep(paste("^", parents[2], sep = ""), logicvec)] <- 5
pchvec[which(logicvec %in% "NOEPI")] <- 6
pchvec[which(logicvec %in% c("NOINFO", "NOINF"))] <- 7

logicvec <- logicvec[-which(logicvec %in% "0")]
pchvec <- pchvec[-which(pchvec == 0)]
llvec <- llvec[-which(llvec == 0)]

colvec <- pchvec

if (all(is.infinite(llvec) == T)) {

  llvec[1:length(llvec)] <- -1000

  margin <- 100

  donames <- 30

} else {

  llvec[which(is.infinite(llvec) == T)] <- NA

  ## llvec[which(is.infinite(llvec) == T)] <- min(llvec) - 100

  margin <- abs(max(llvec[1:30], na.rm = T) - min(llvec[1:30], na.rm = T))

  offset <- 0.075

  if (margin == 0) { margin <- 10; offset <- 0.0375 }

  donames <- 30 - sum(is.na(llvec[1:30]) == T)

  if (any(is.na(llvec[1:30]) == T)) { margin2 <- margin*2
  } else { margin2 <- margin }

  llvec[which(is.na(llvec) == T)] <- min(llvec, na.rm = T) - margin

```

```

margin <- margin2

}

if (all(llvec[-(1:30)] - min(llvec[-(1:30)]) == 0)) {

  p2max <- max(llvec[-(1:30)]) + margin

} else {

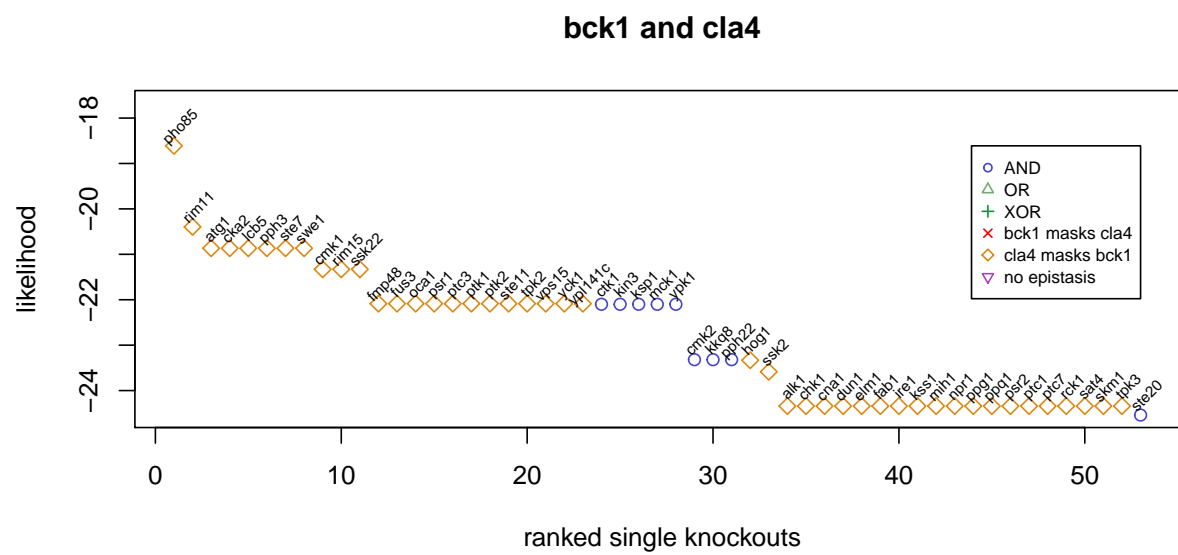
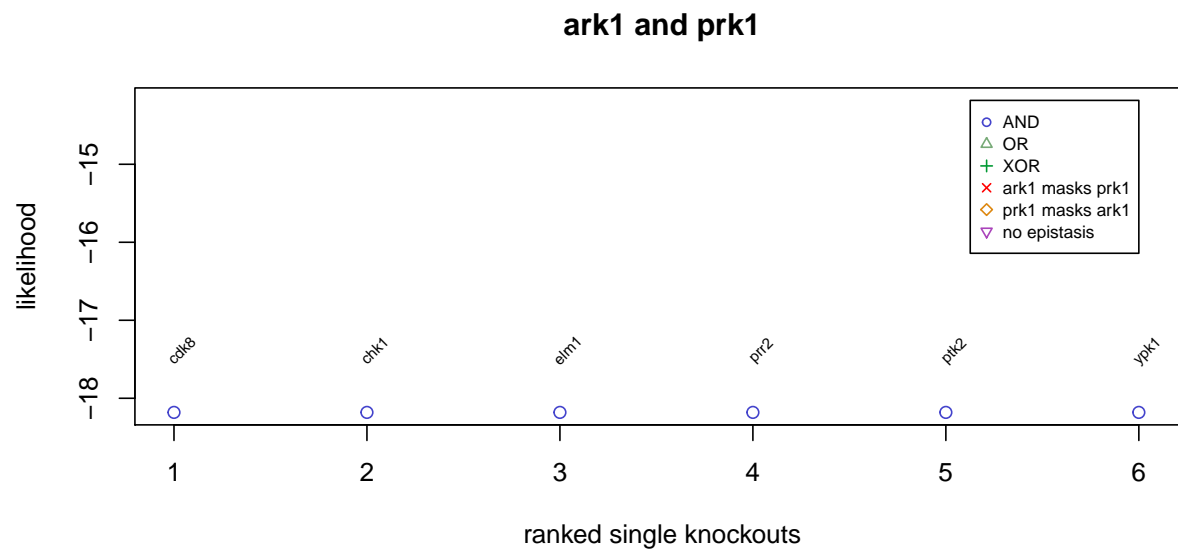
  p2max <- max(llvec[-(1:30)])

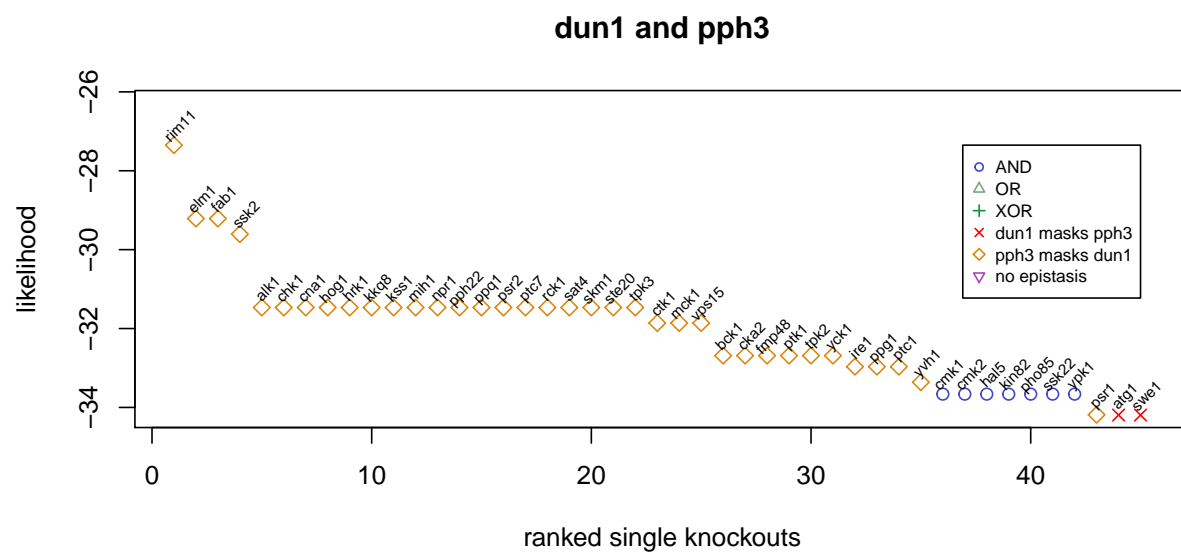
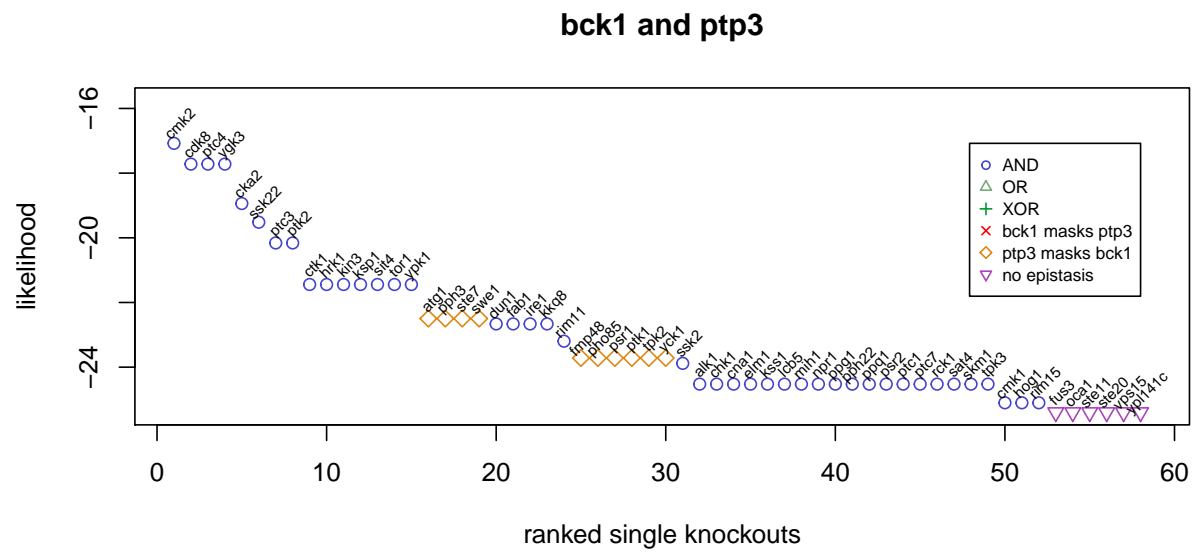
}

mark <- ""
thetop <- sum(!(logicvec %in% c("NOINFO", "NOINF")))
legendx <- length(llvec[1:thetop])
p2max <- max(llvec[1:thetop])
if (p2max == min(llvec[1:thetop])) {
  p2max <- p2max+margin*0.2
}
legendtext <- c("AND", "OR", "XOR", paste(parents[1]," masks ", parents[2], sep = ""),
               paste(parents[2], " masks ", parents[1], sep = ""), "no epistasis")
if (thetop == 0) { next() }
plot = plot(llvec[1:thetop], pch = pchvec[1:thetop], col = colvec[1:thetop],
            ylab = "likelihood", xlab = "ranked single knockouts",
            ylim = c(min(llvec[1:thetop]), max(llvec[1:thetop])+margin*0.2),
            xlim = c(1, thetop+(thetop/100)),
            main = paste(unlist(strsplit(doubles[i], "\\.")), collapse = " and "))
text = text((1:thetop)+(thetop/100), llvec[1:thetop]+(margin*offset),
            labels = names(llvec)[1:thetop], cex = 0.6, srt = 45, pos = 3,
            offset = 0)
mtext = mtext(mark, side = 3, line = 1, outer = F, cex = 4, adj = 0)
legend = legend(legendx, p2max,
               legend = legendtext,
               col = 1:6, pch = 1:6, xjust = 1, yjust = 1, cex = 0.7)

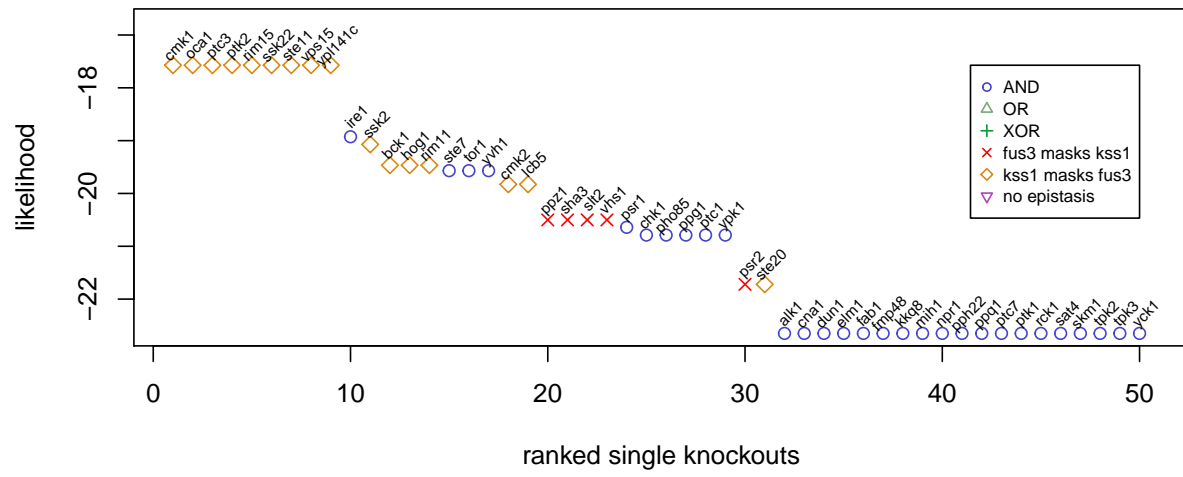
}

```

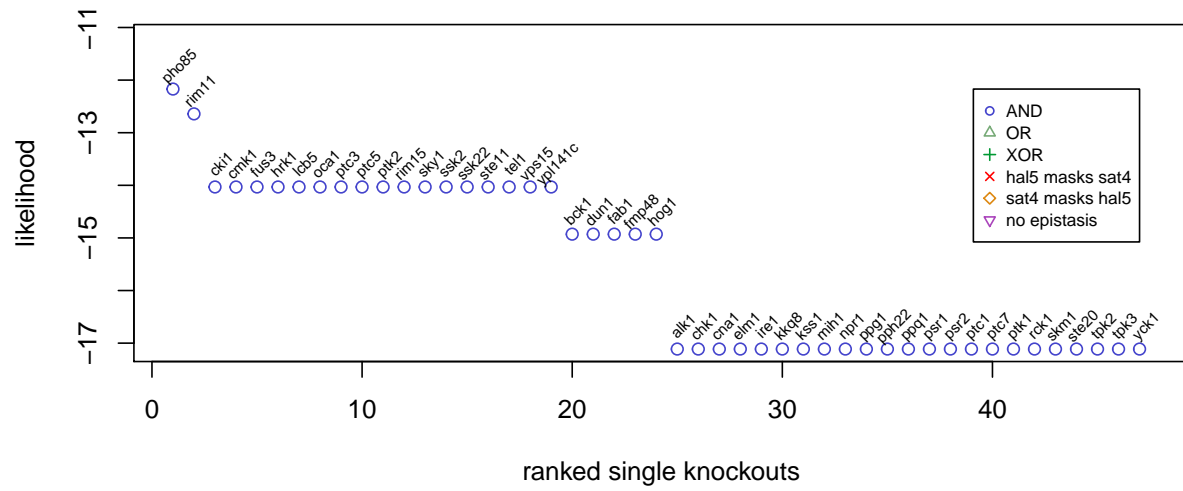




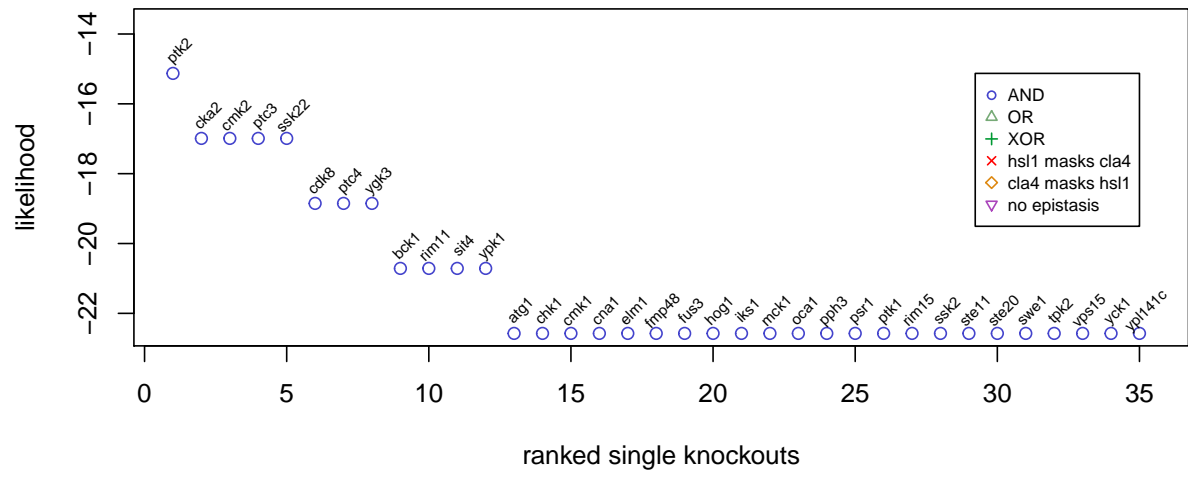
fus3 and kss1



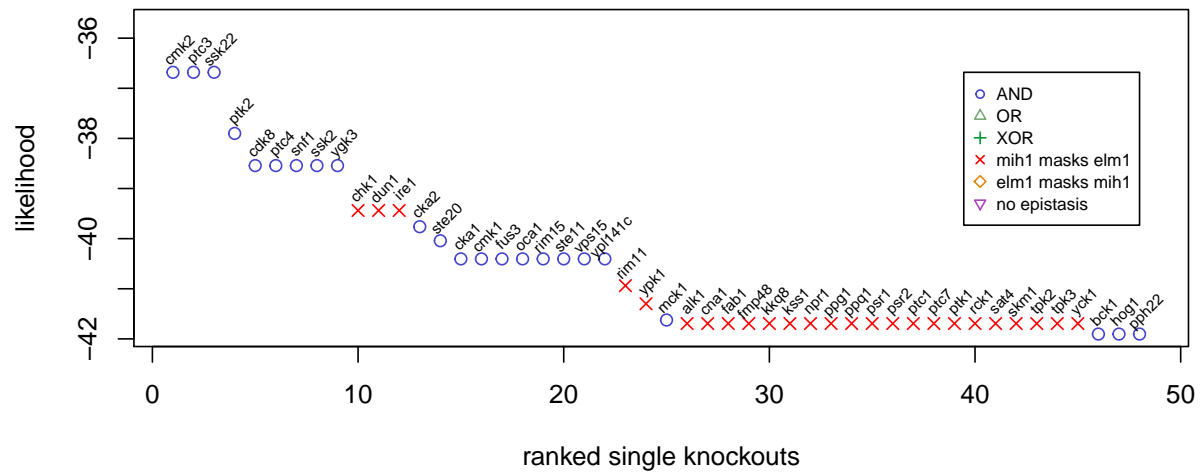
hal5 and sat4

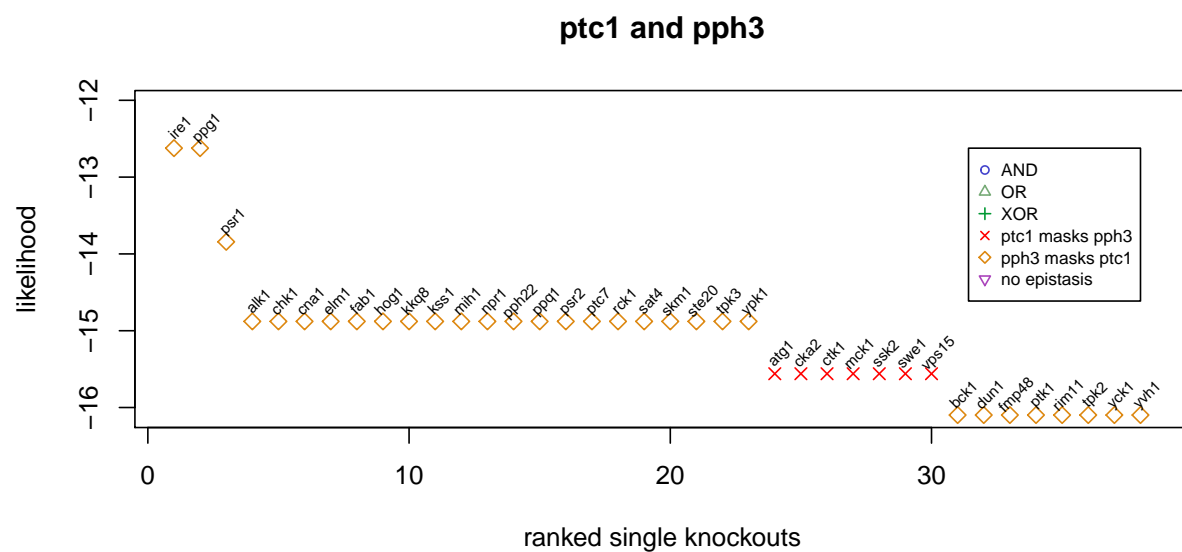
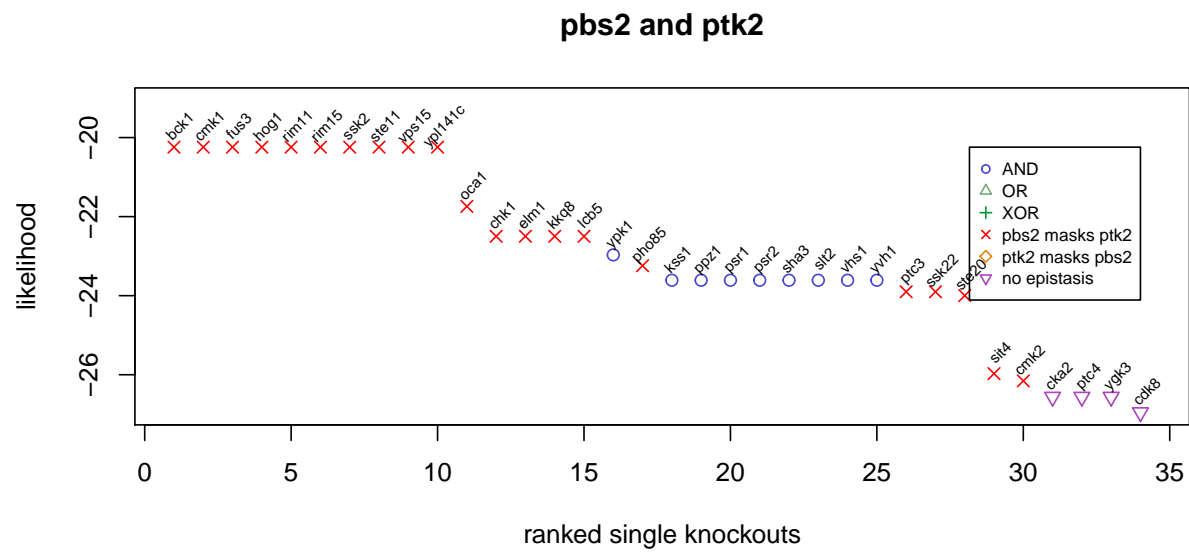


hsl1 and cla4

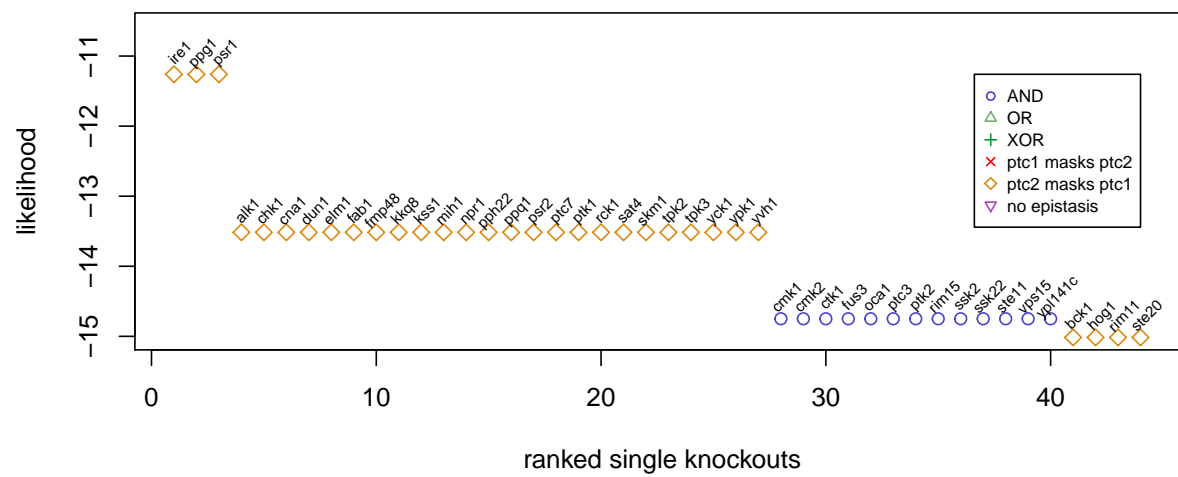


mih1 and elm1

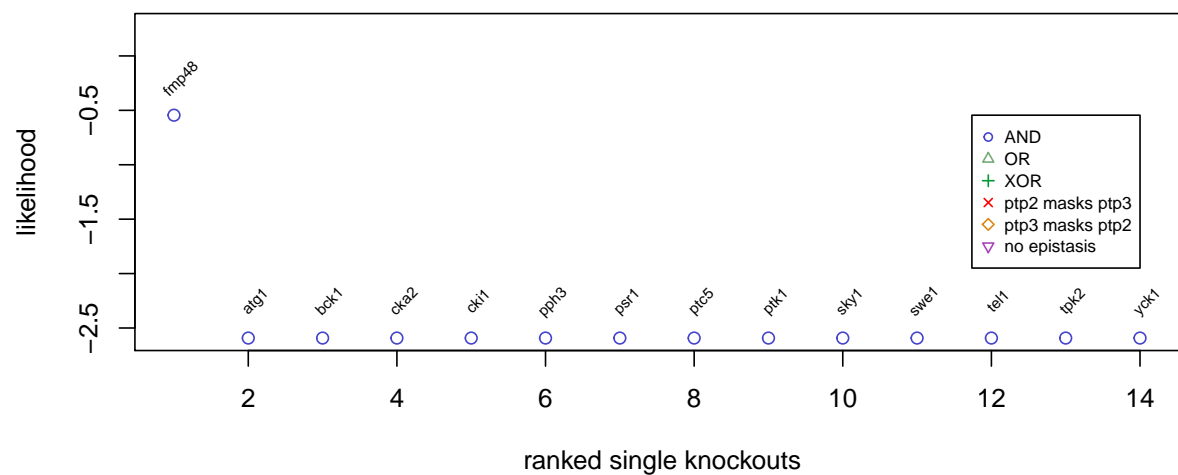


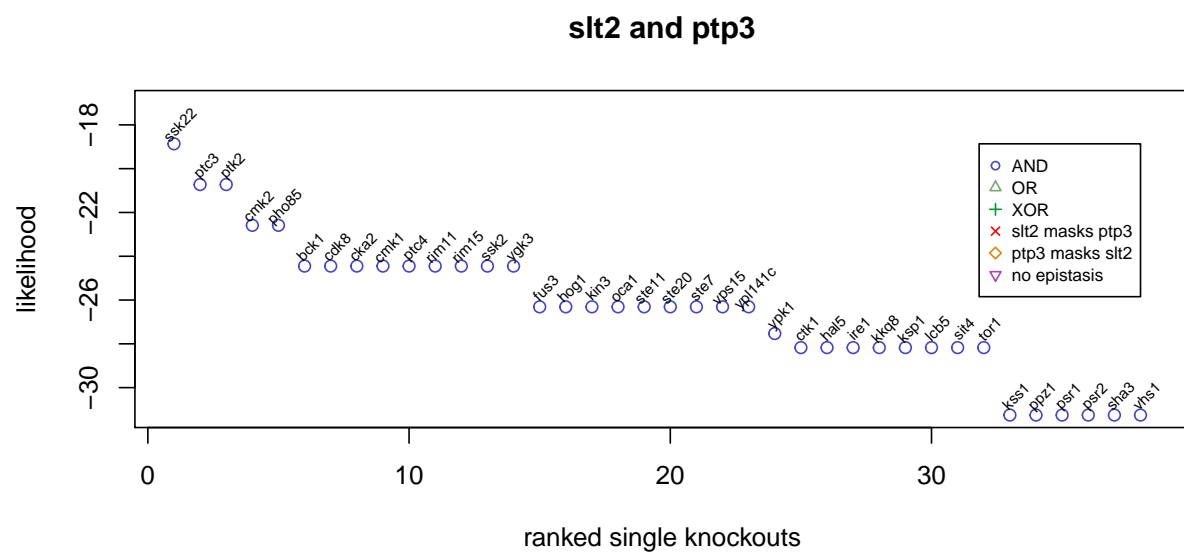
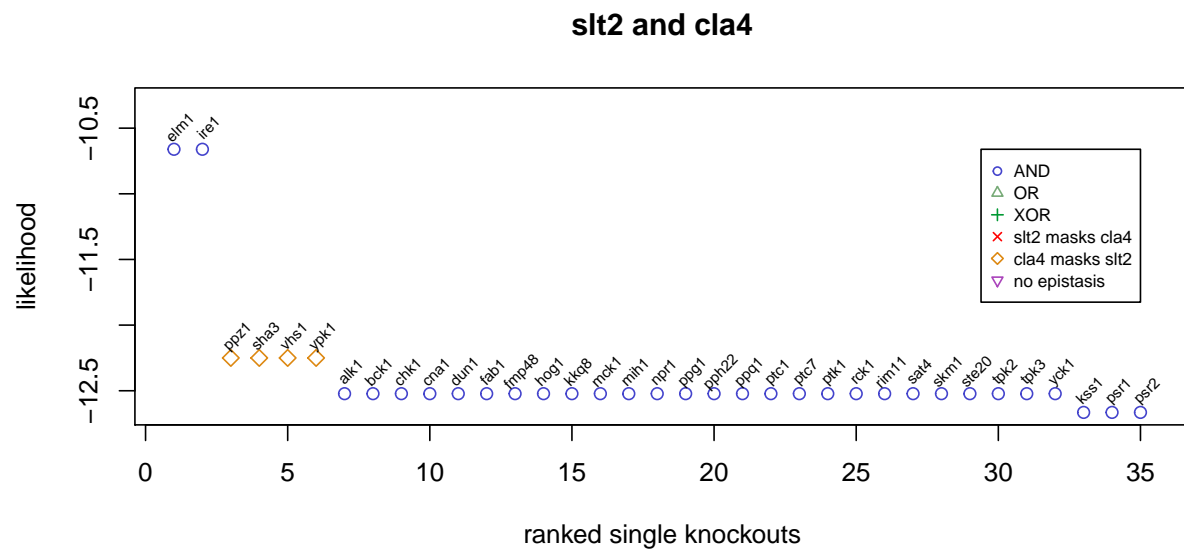


ptc1 and ptc2

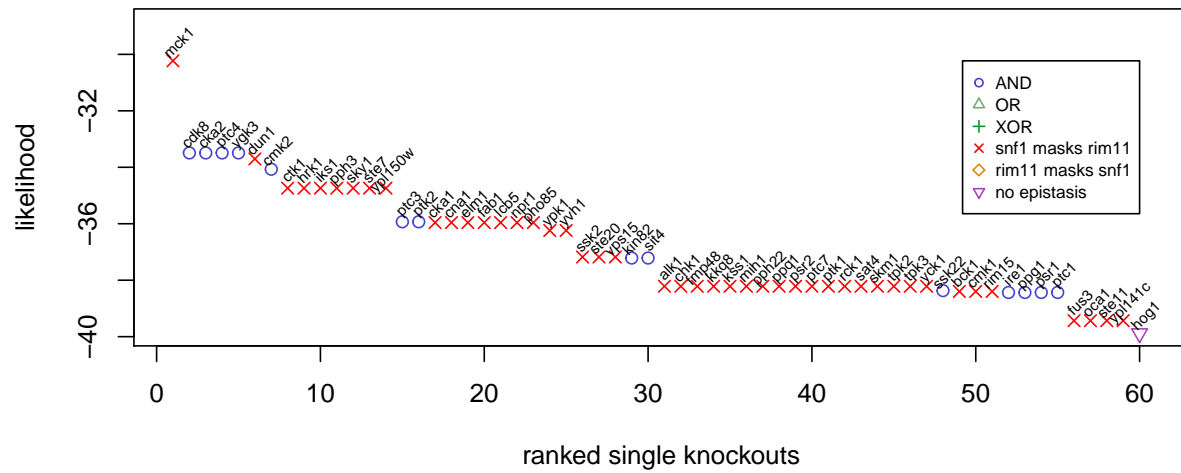


ptp2 and ptp3





snf1 and rim11



```

distmat <- wageningen$logic

distmat[which(distmat %in% "AND")] <- 1
distmat[which(distmat %in% "OR")] <- 2
distmat[which(distmat %in% "XOR")] <- 3
distmat[which(distmat %in% "NOEPI")] <- 6
distmat[which(distmat %in% c("NOINFO", "NOINF"))] <- 7

for (i in 1:ncol(distmat)) {

  genes <- unlist(strsplit(colnames(distmat)[i], "\\."))

  distmat[which(distmat[, i] %in%
    paste(genes[1], " masks the effect of ", genes[2], sep = "")), i] <- 4

  distmat[which(distmat[, i] %in%
    paste(genes[2], " masks the effect of ", genes[1], sep = "")), i] <- 5

}

distmat <- apply(distmat, c(1,2), as.numeric)

for (i in 1:ncol(distmat)) {
  distmat[, i] <- rev(sort(distmat[, i]))
}

distmat <- distmat[-which(apply(distmat, 1, sum) == 0), ]

distmat <- distmat[, -which(apply(distmat, 2, max) == 0 | apply(distmat, 2, min) == 7)]

y <- distmat

```

```

dismat <- dismat[, order(apply(dismat, 2, function(x) { return(sum(x == 1)) }))]

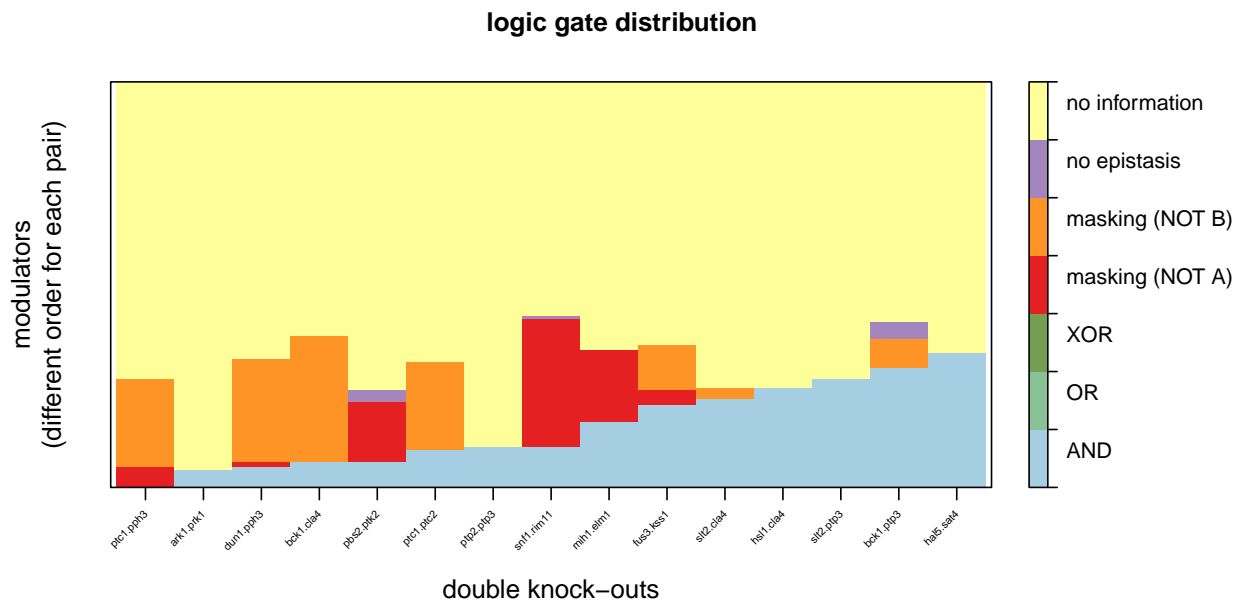
y[which(y == 5)] <- 4

rownames(dismat) <- NULL

labeltext <- c("", "no information\n\n\n", "no epistasis\n\n\n", "masking (NOT B)\n\n\n",
               "masking (NOT A)\n\n\n", "XOR\n\n\n", "OR\n\n\n", "AND\n\n\n")

heatmapOP(dismat, Colv = F, Rowv = F, main = "logic gate distribution",
          sub = "", col = "Paired", breaks = seq(0.5, 7.5, length.out = 8),
          cexRow = 0, cexCol = 0.4, aspect = "fill",
          colorkey = list(space = "right",
                          labels = rev(labeltext), width = 1,
                          at = seq(1.5, 7.5, length.out = 8)),
          xlab = "double knock-outs",
          ylab = "modulators\n(different order for each pair)",
          xrot = 45, bordercol = "transparent")

```



Sameith et al., 2015

```

file <- paste("http://www.holstegelab.nl/publications/GSTF_geneticinteractions/",
              "downloads/del_mutants_limma.txt", sep = "")

data <- read.delim(file)

data <- apply(data, c(1,2), as.character)

dataM <- data[-1, which(data[1, ] %in% "M")]

dataM <- apply(dataM, c(1,2), as.numeric)

```



```
dataP <- data[-1, which(data[1, ] %in% "p.value")]

dataP <- apply(dataP, c(1,2), as.numeric)

dataBin <- dataM

sig <- 0.01

cutoff <- log2(1.5)

dataBin[which(dataP < sig & dataP > 0 & abs(dataM) >= cutoff)] <- 1

dataBin[which(dataP >= sig | dataP == 0 | abs(dataM) < cutoff)] <- 0

dataBin <- dataBin[-which(apply(dataBin, 1, max) == 0), ]

colnames(dataBin) <- gsub("\\.\\.\\.\\.\\.\"", "\\\".", colnames(dataBin))

## big screen:

doubles <- colnames(dataBin)[grep("\\\".", colnames(dataBin))]

doubles.genes <- unique(unlist(strsplit(doubles, "\\\".")))

singles <- colnames(dataBin)[-grep("\\\".", colnames(dataBin))]

singles <- unique(sort(singles))

llmat <- logicmat <- matrix(0, length(singles), length(doubles))

rownames(llmat) <- rownames(logicmat) <- singles

colnames(llmat) <- colnames(logicmat) <- doubles

globalgenes <- which(apply(dataBin, 1, max) == 1)

for (i in doubles[set]) {
  print(i)
  doubles.singles <- unlist(strsplit(i, "\\\"."))
  egenes <- which(apply(dataBin[,
    which(colnames(dataBin) %in% c(i, doubles.singles))], 1, max) == 1)
  for (j in singles) {
    print(j)
    if (j %in% doubles.singles) { next() }

    dataTmp <- dataBin[, grep(paste(paste("^", c(i, j, doubles.singles), "$", sep
      = ""), collapse = "|"), colnames(dataBin))]

    if (path %in% "fixed_set") {
      dataTmp <- dataTmp[egenes, ]
    }
    if (path %in% "global") {
      dataTmp <- dataTmp[globalgenes, ]
    }
  }
}
```

```

}
if (path %in% "") {
  dataTmp <- dataTmp[which(apply(dataTmp, 1, max) == 1), ]
}

i1 <- which(singles %in% j)
i2 <- which(doubles %in% i)

if (!(is.null(dim(dataTmp)))) {

  if (any(dataTmp[, j] != 0)) {

    epires <- epiNEM(dataTmp, method = "exhaustive")

    tmp <- epires$logics
    if ("OR" %in% tmp) {
      if (sum(epires$origModel[, j]) != 2) {
        tmp <- "NOEPI"
      } else {
        if (all(tmp %in% "OR")) {
          tmp <- "OR"
        } else {
          tmp <- tmp[which(!(tmp %in% "OR"))]
        }
      }
    }

    logicmat[i1, i2] <- tmp
    llmat[i1, i2] <- epires$score

  } else {

    logicmat[i1, i2] <- "UNCON"
    llmat[i1, i2] <- -Inf

  }

} else {

  logicmat[i1, i2] <- "UNCON"
  llmat[i1, i2] <- -Inf

}

}
}

```

```

data(sameith_res)

llmat0 <- sameith$ll

logicmat0 <- sameith$logic

```

```

for (i in 1:length(doubles)) {

  logicvec <- logicmat0[, i]

  llvec <- llmat0[, i]

  logicvec <- logicvec[order(llvec, decreasing = T)]

  llvec <- llvec[order(llvec, decreasing = T)]

  parents <- unlist(strsplit(doubles[i], "\\\\"))

  pchvec <- numeric(length(llvec))

  pchvec[which(logicvec %in% "AND")] <- 1
  pchvec[which(logicvec %in% "OR")] <- 2
  pchvec[which(logicvec %in% "XOR")] <- 3
  pchvec[grepl(paste("^", parents[1], sep = ""), logicvec)] <- 4
  pchvec[grepl(paste("^", parents[2], sep = ""), logicvec)] <- 5
  pchvec[which(logicvec %in% "NOEPI")] <- 6
  pchvec[which(logicvec %in% c("NOINFO", "NOINF"))] <- 7

  logicvec <- logicvec[-which(logicvec %in% "0")]
  pchvec <- pchvec[-which(pchvec == 0)]
  llvec <- llvec[-which(llvec == 0)]

  colvec <- pchvec

  if (all(is.infinite(llvec) == T)) {

    llvec[1:length(llvec)] <- -1000

    margin <- 100

    donames <- 30

  } else {

    llvec[which(is.infinite(llvec) == T)] <- NA

    margin <- abs(max(llvec[1:30], na.rm = T) - min(llvec[1:30], na.rm = T))

    if (margin == 0) { margin <- 10 }

    donames <- 30 - sum(is.na(llvec[1:30]) == T)

    if (any(is.na(llvec[1:30]) == T)) { margin2 <- margin*2 }
    else { margin2 <- margin }

    llvec[which(is.na(llvec) == T)] <- min(llvec, na.rm = T) - margin

    margin <- margin2
  }
}

```

```

}

if (all(llvec[-(1:30)] - min(llvec[-(1:30)]) == 0)) {

  p2max <- max(llvec[-(1:30)]) + margin

} else {

  p2max <- max(llvec[-(1:30)])

}

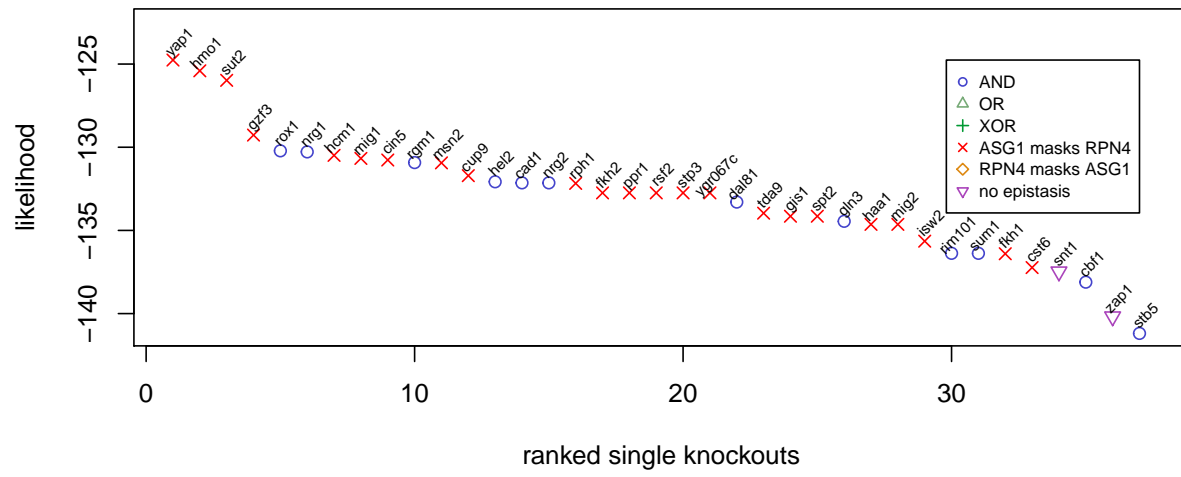
labeltext <- c("AND", "OR", "XOR", paste(parents[1], " masks ", parents[2], sep = ""),
              paste(parents[2], " masks ", parents[1], sep = ""), "no epistasis")

mark <- ""
pointx <- 10000
thetop <- sum(!(logicvec %in% c("NOINFO", "NOINF")))
legendx <- length(llvec[1:thetop])
p2max <- max(llvec[1:thetop])
if (p2max == min(llvec[1:thetop])) {
  p2max <- p2max+margin*0.2
}
if (thetop == 0) { next() }
plot = plot(llvec[1:thetop], pch = pchvec[1:thetop], col = colvec[1:thetop],
            ylab = "likelihood", xlab = "ranked single knockouts",
            ylim = c(min(llvec[1:thetop]), max(llvec[1:thetop])+margin*0.2),
            xlim = c(1, thetop+(thetop/100)),
            main = paste(tolower(unlist(strsplit(doubles[i], "\\."))),
                        collapse = " and "))
text = text((1:thetop)+(thetop/100), llvec[1:thetop]+(margin*offset),
            labels = tolower(names(llvec)[1:thetop]), cex = 0.6, srt = 45, pos = 3,
            offset = 0)
mtext = mtext(mark, side = 3, line = 1, outer = F, cex = 4, adj = 0)
legend = legend(legendx, p2max,
               legend = labeltext, col = 1:6, pch = 1:6, xjust = 1, yjust = 1,
               cex = 0.7)

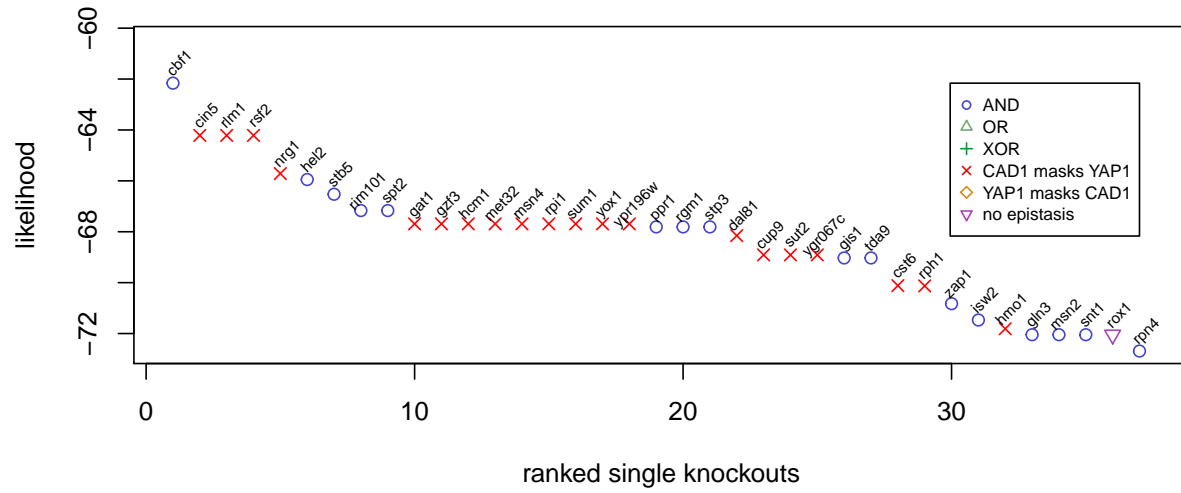
}

```

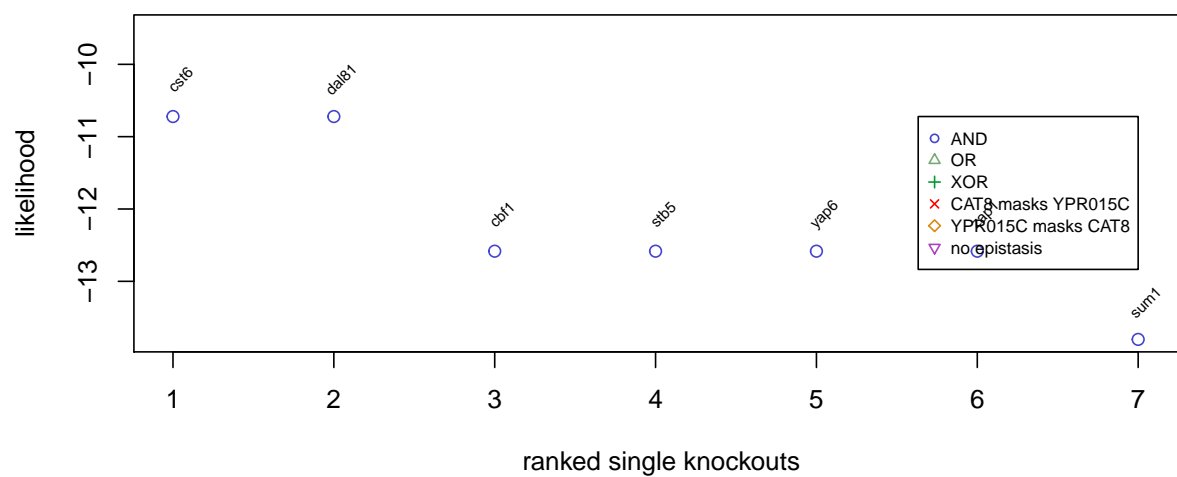
asg1 and rpn4



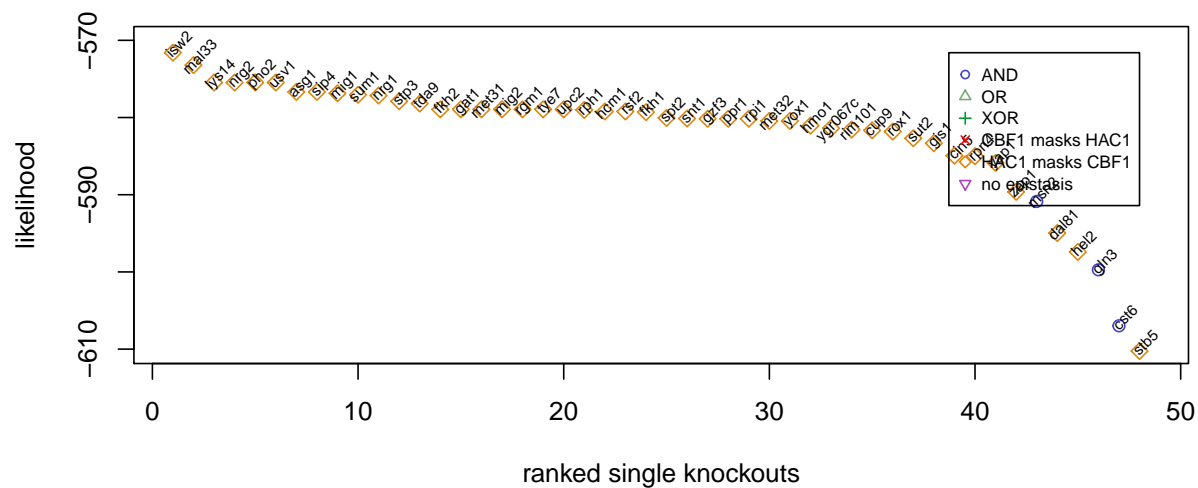
cad1 and yap1

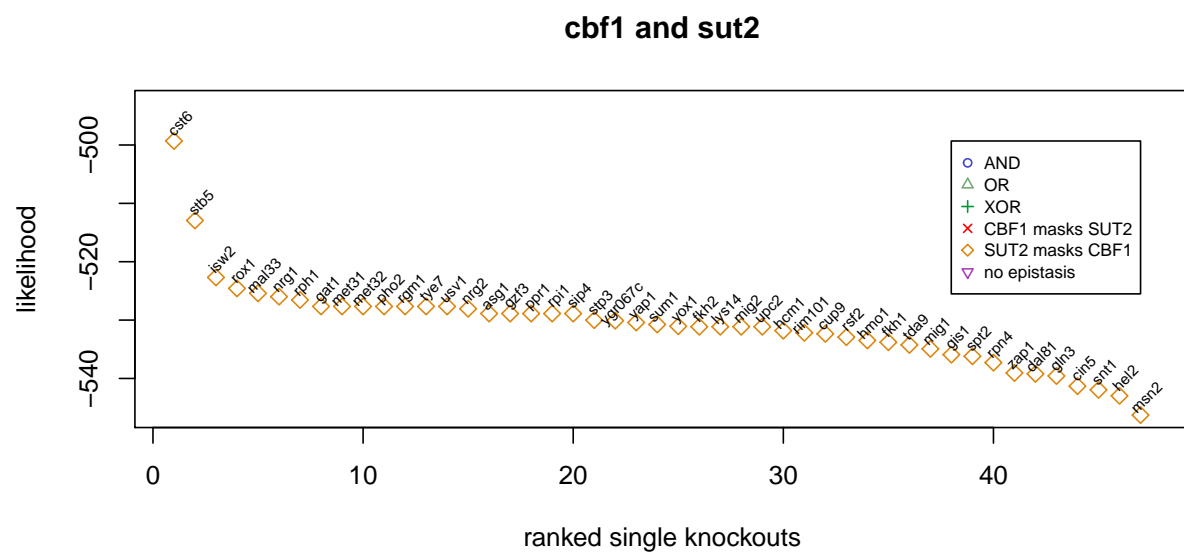
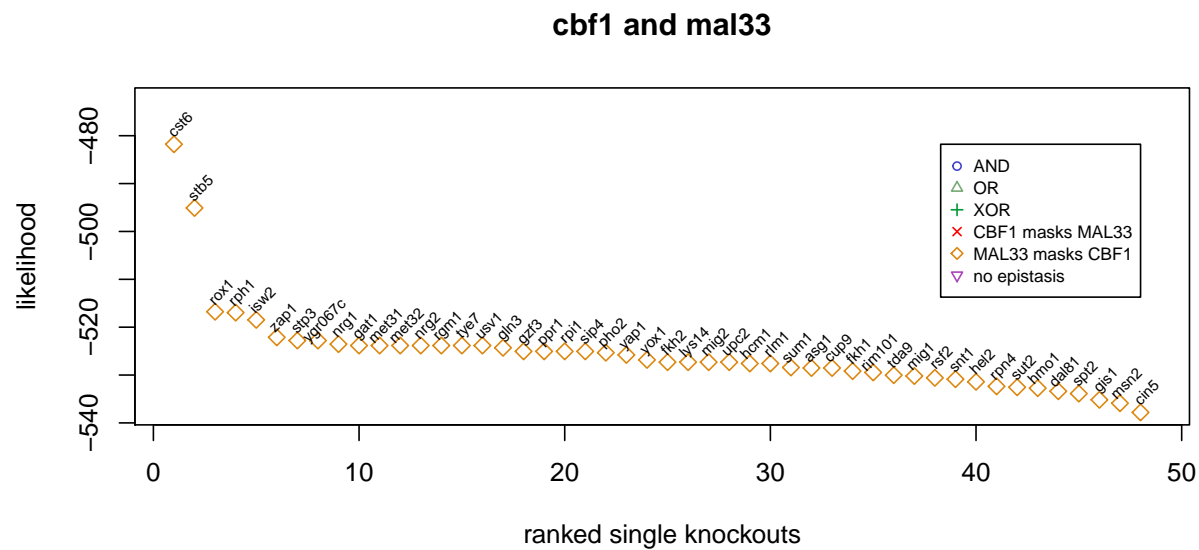


cat8 and ypr015c

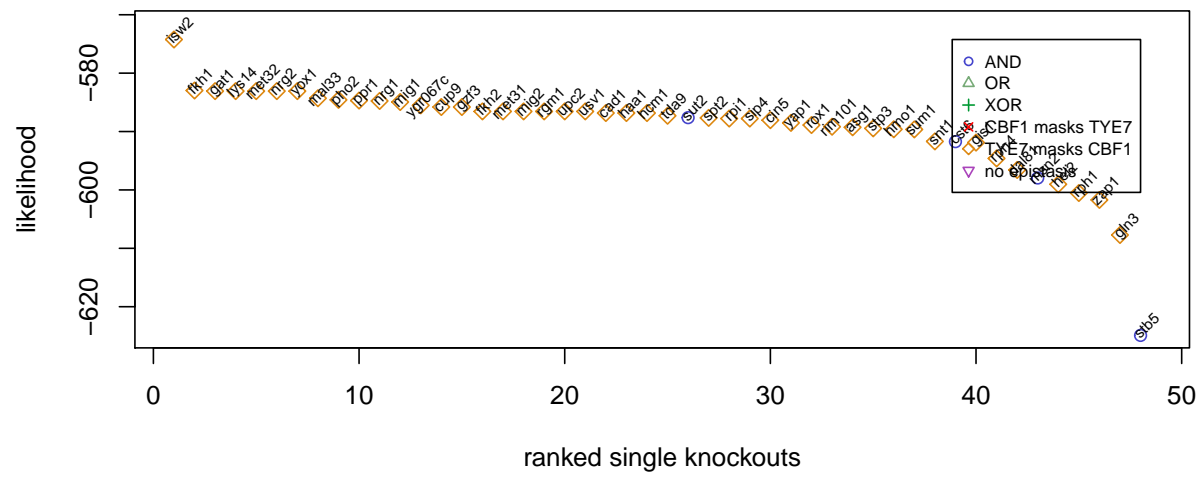


cbf1 and hac1

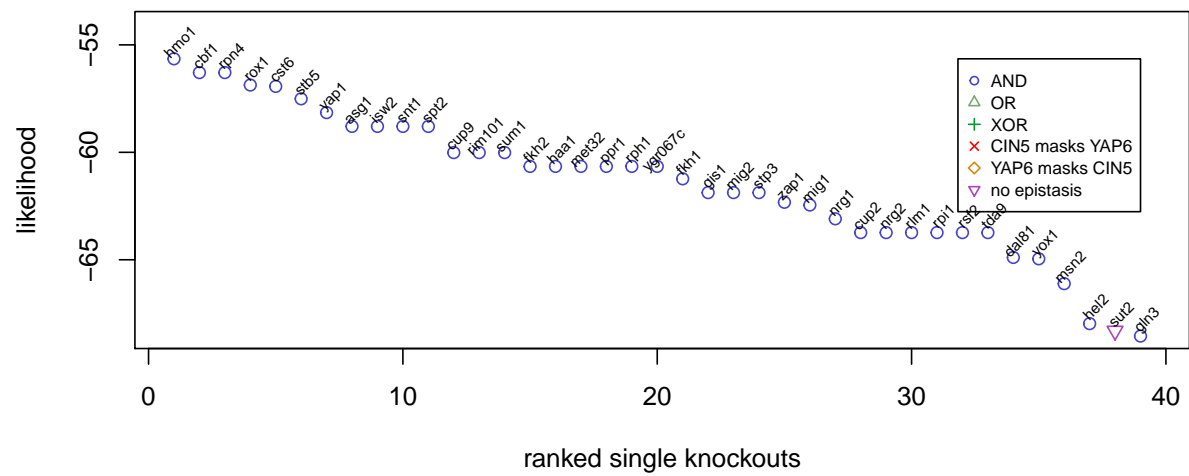


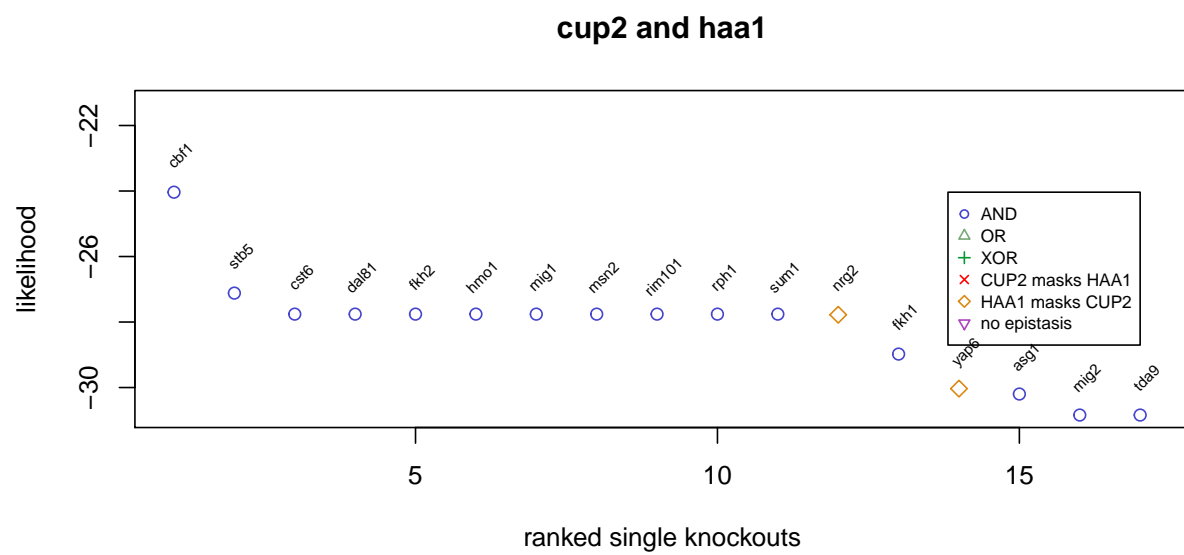
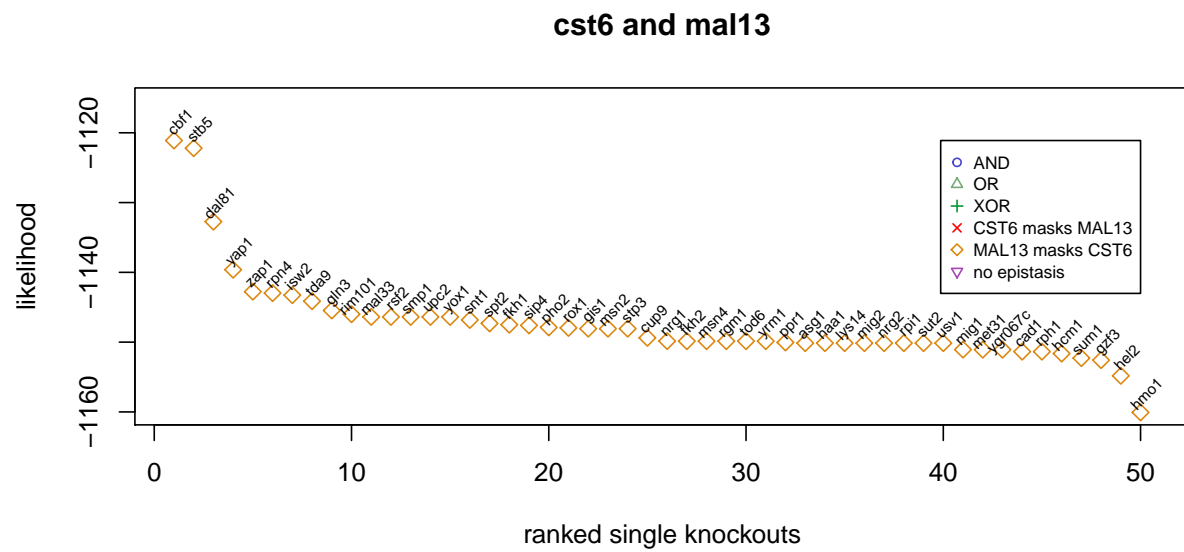


cbf1 and tye7

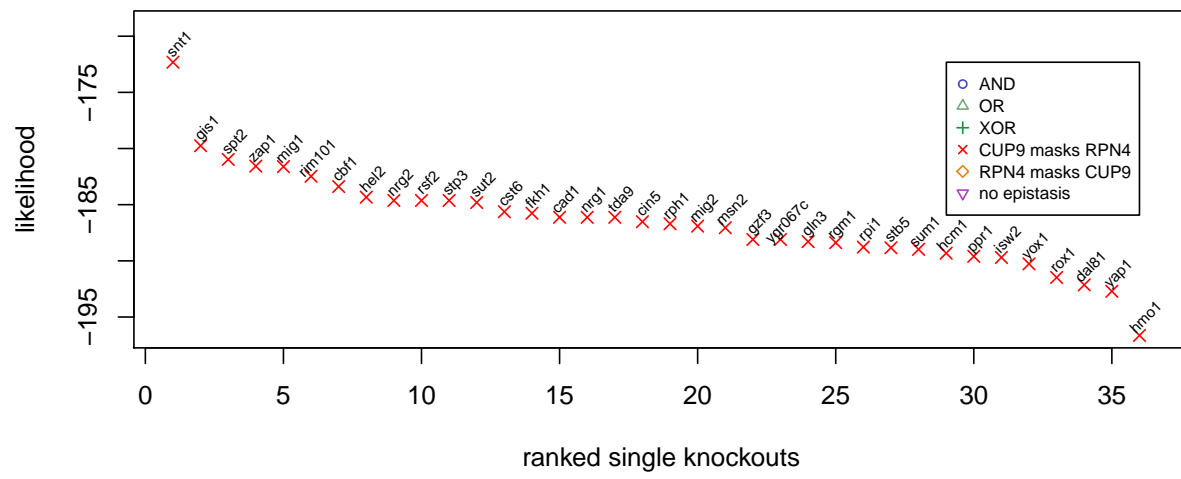


cin5 and yap6

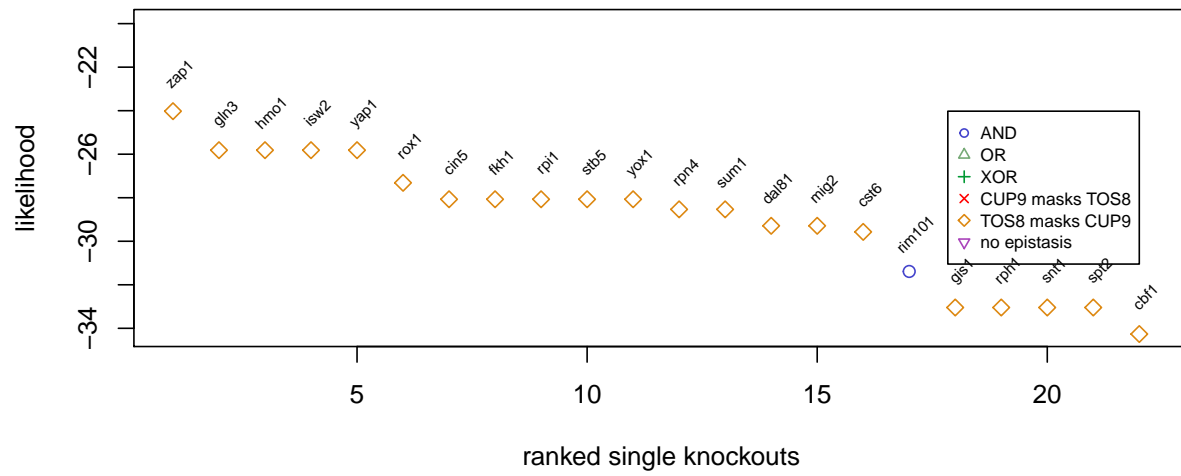




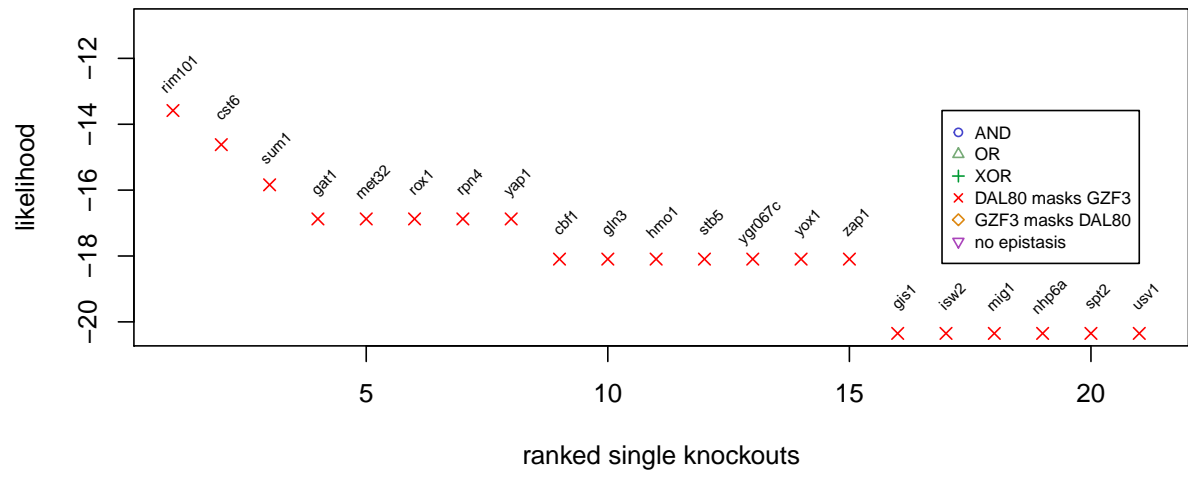
cup9 and rpn4



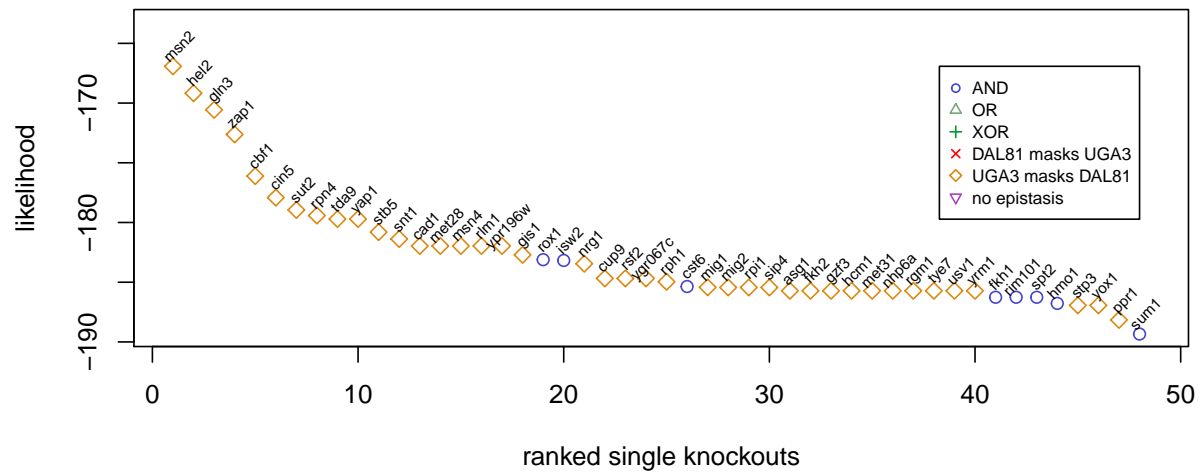
cup9 and tos8

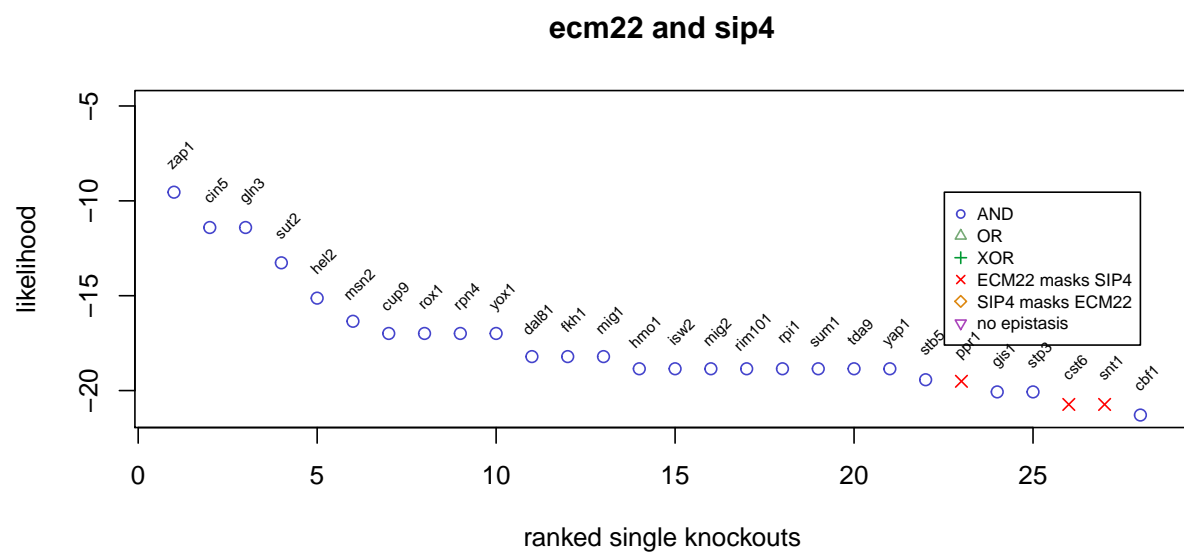
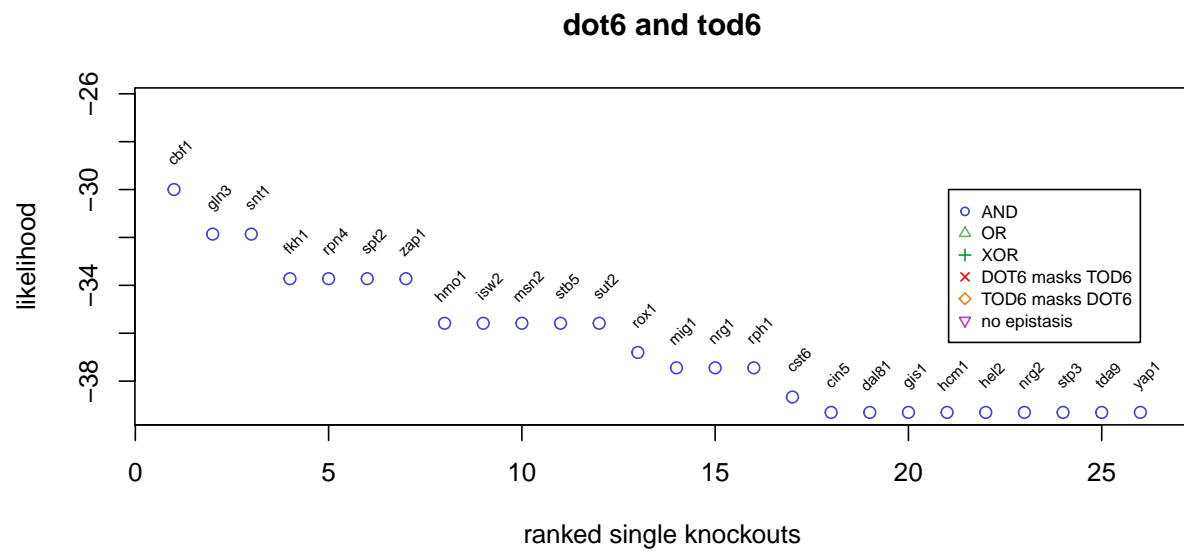


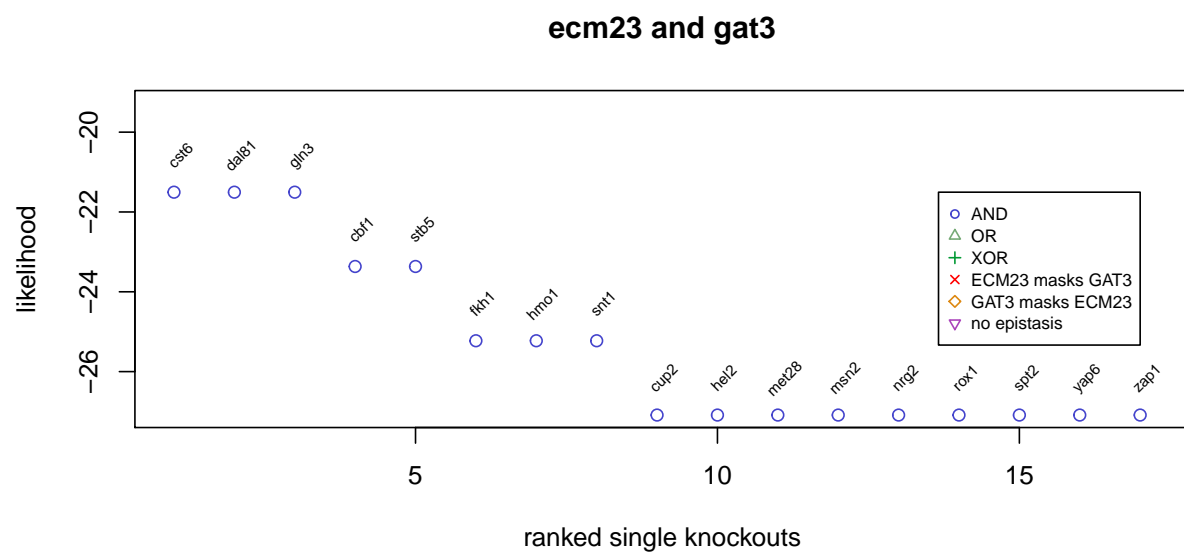
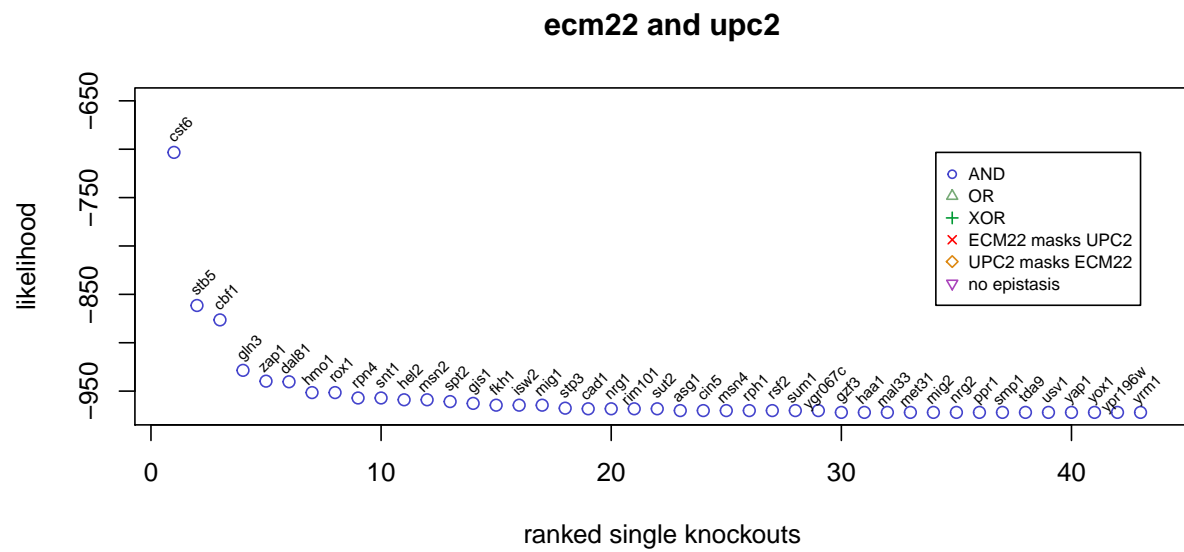
dal80 and gzf3



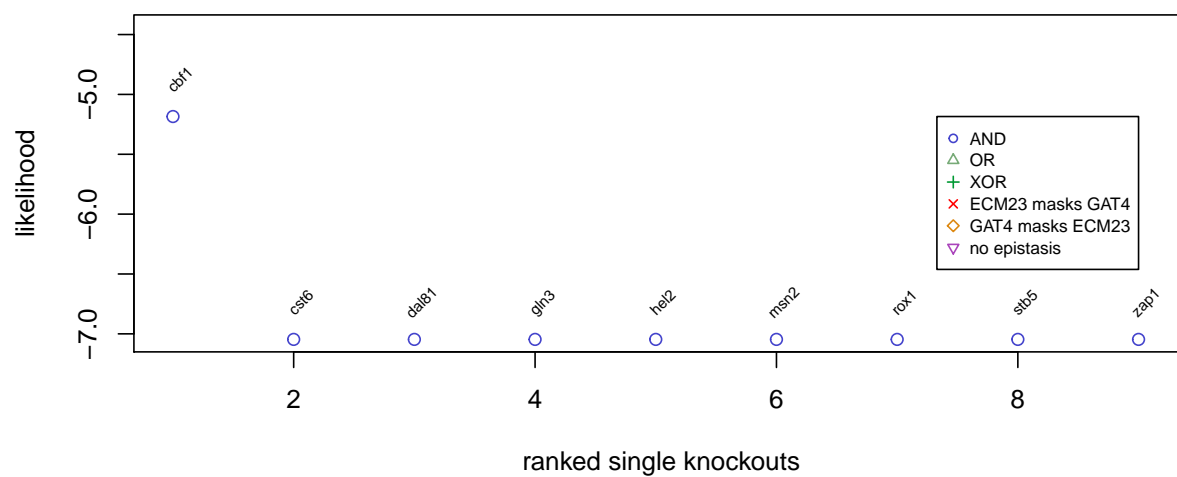
dal81 and uga3



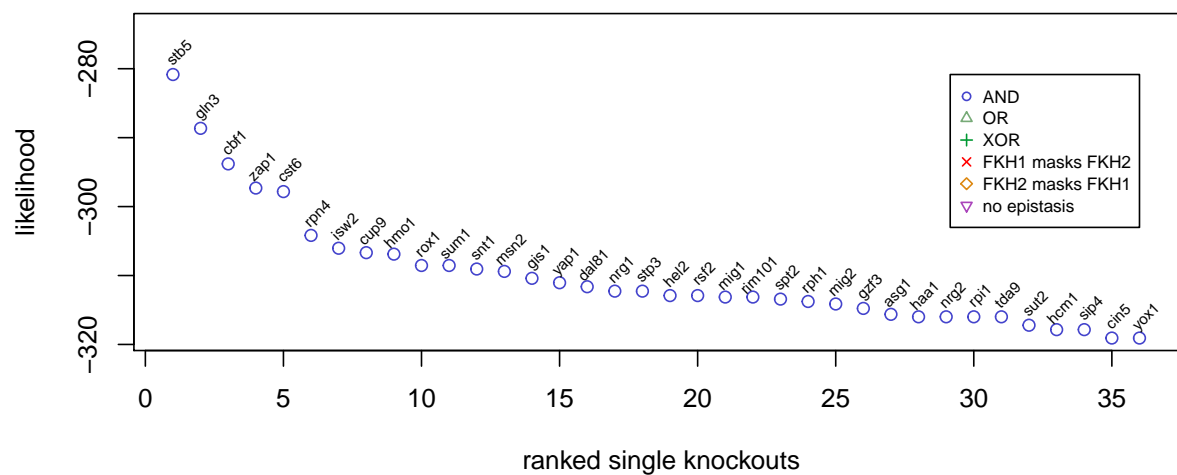




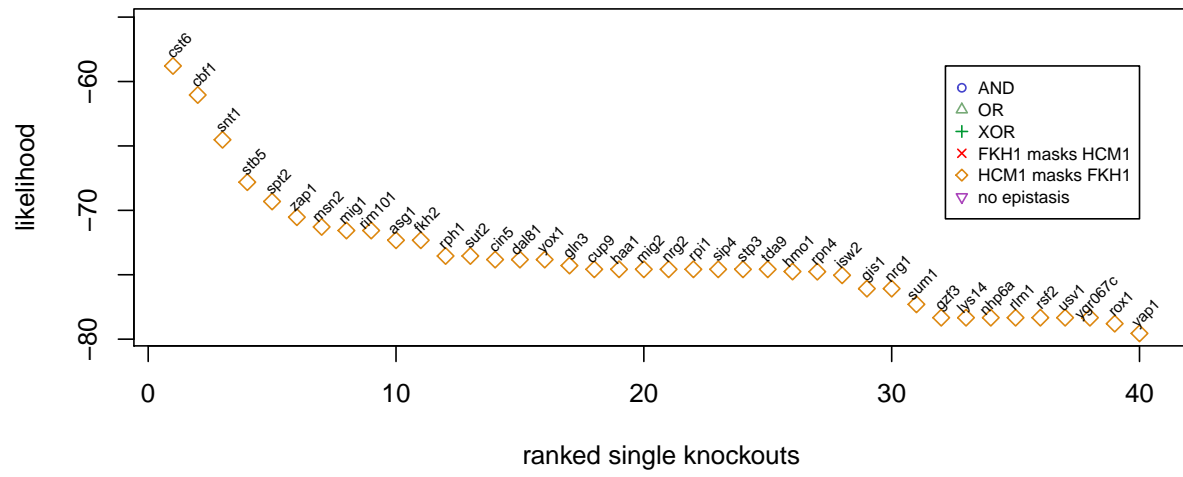
ecm23 and gat4



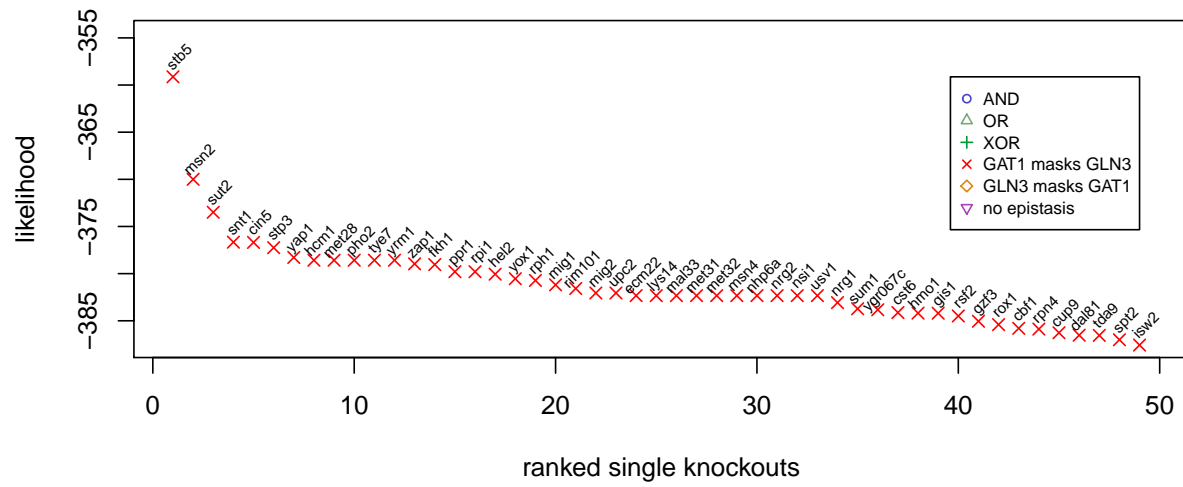
fkf1 and fkh2

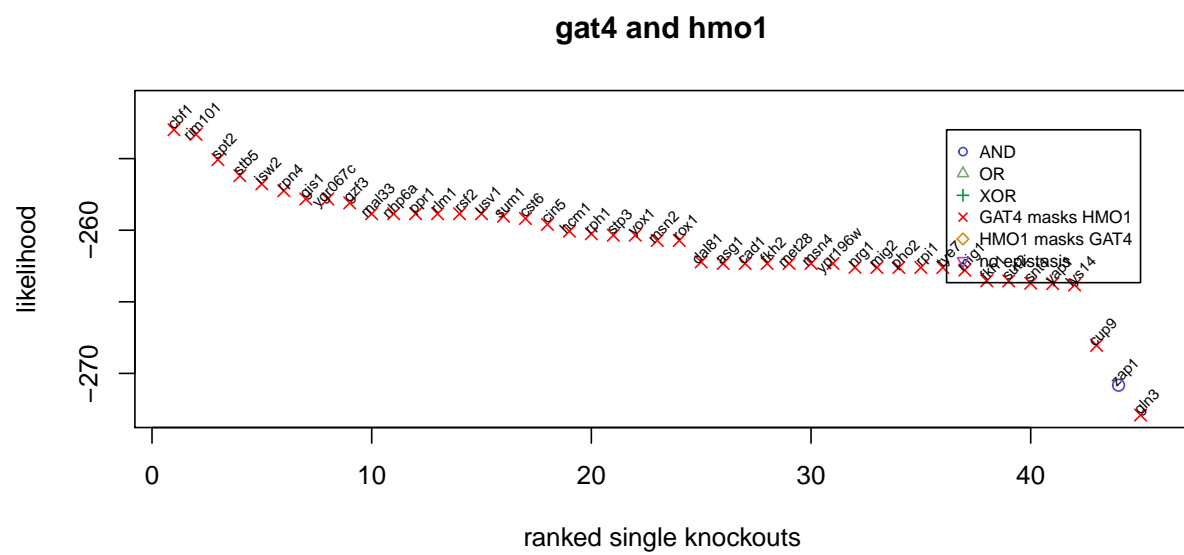
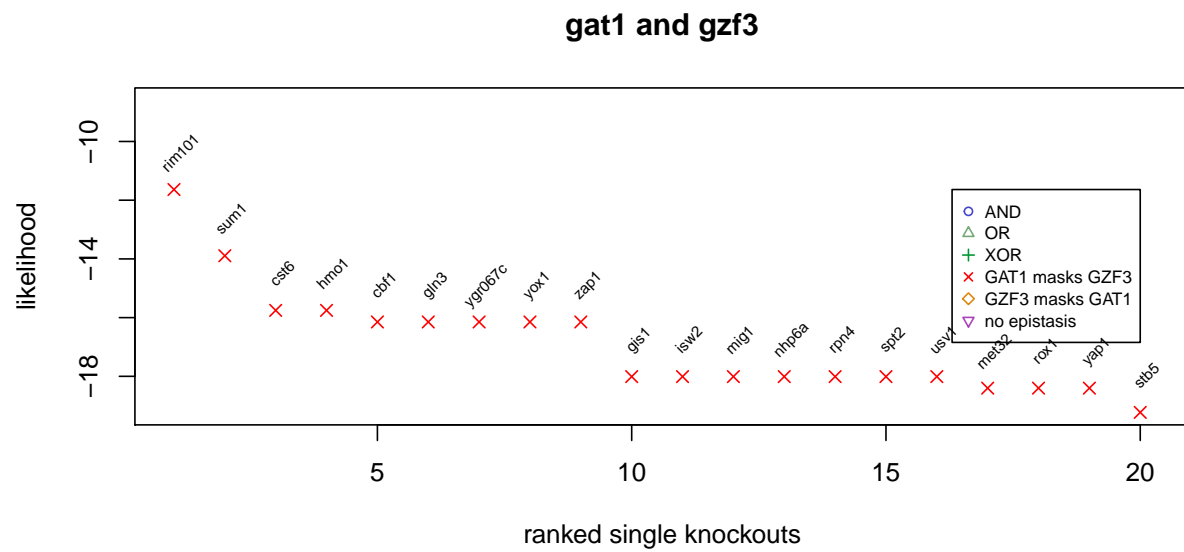


fkh1 and hcm1

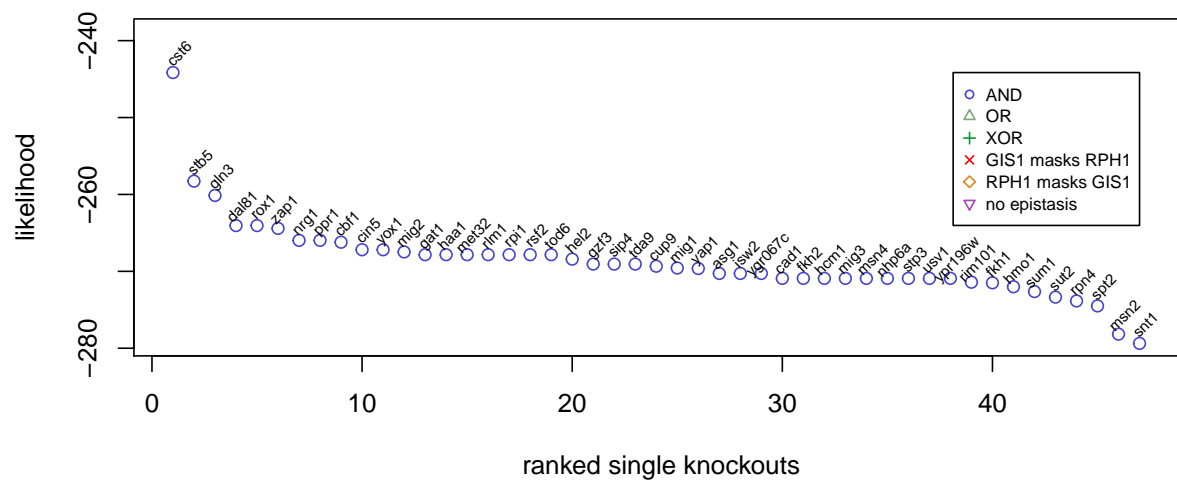


gat1 and gln3

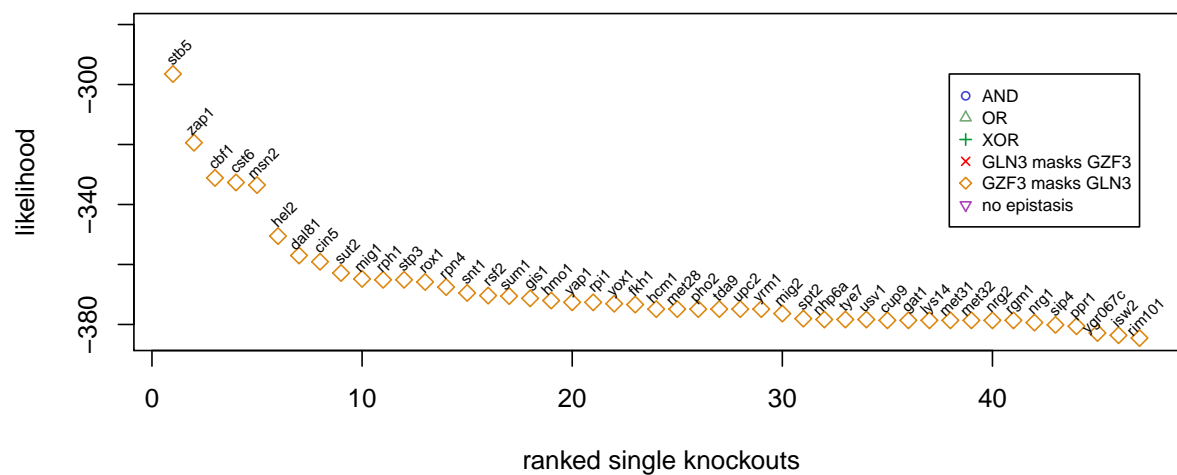




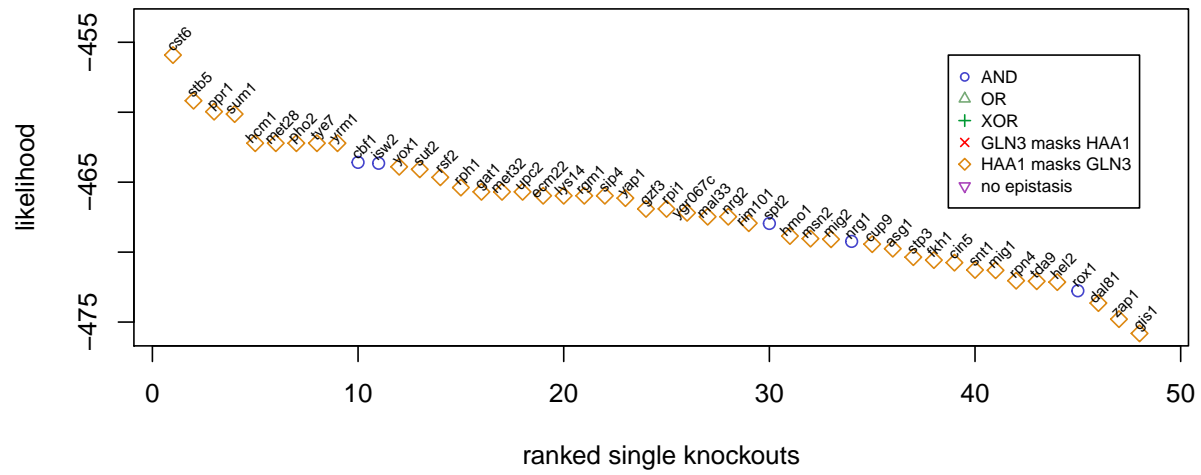
gis1 and rph1



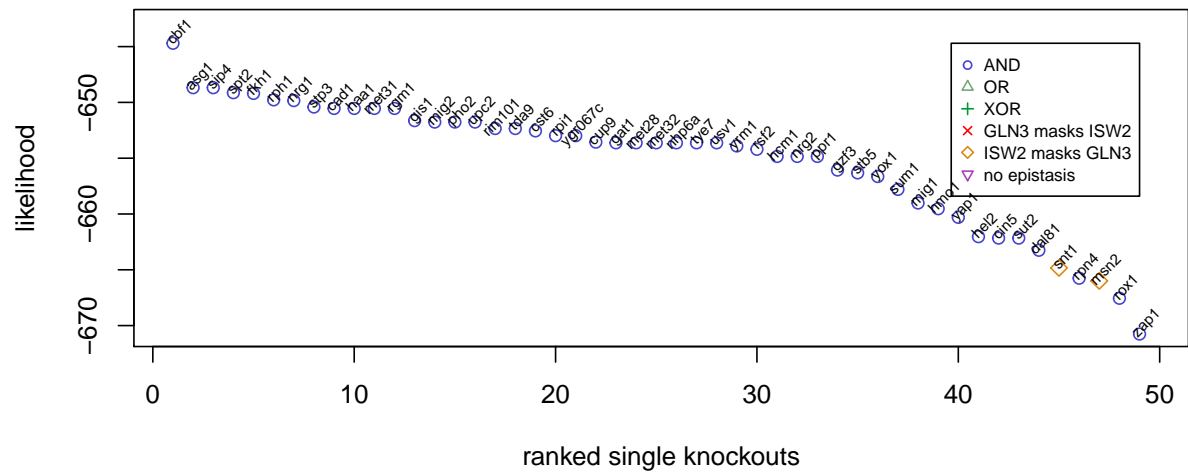
gln3 and gzf3



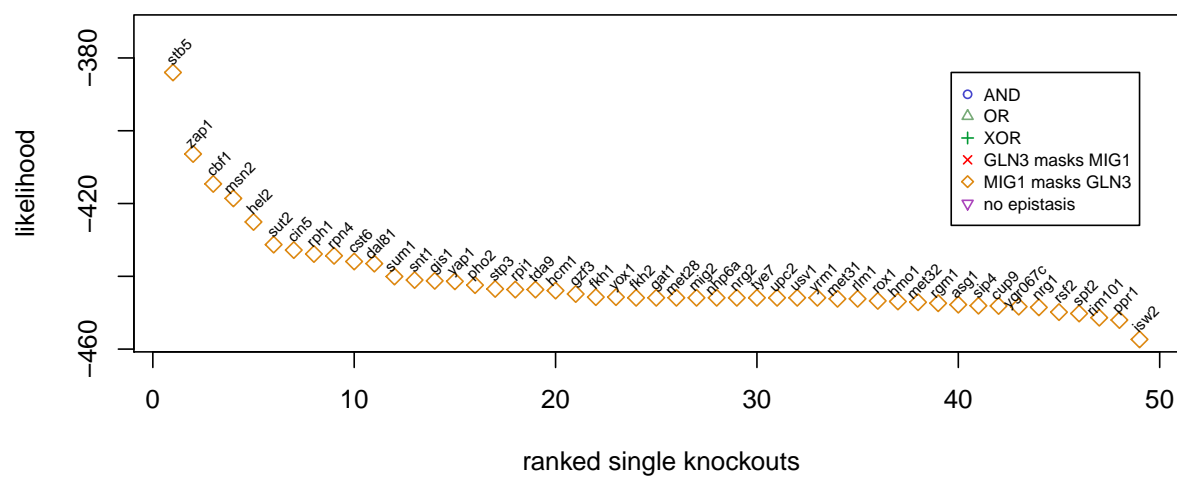
gln3 and haa1



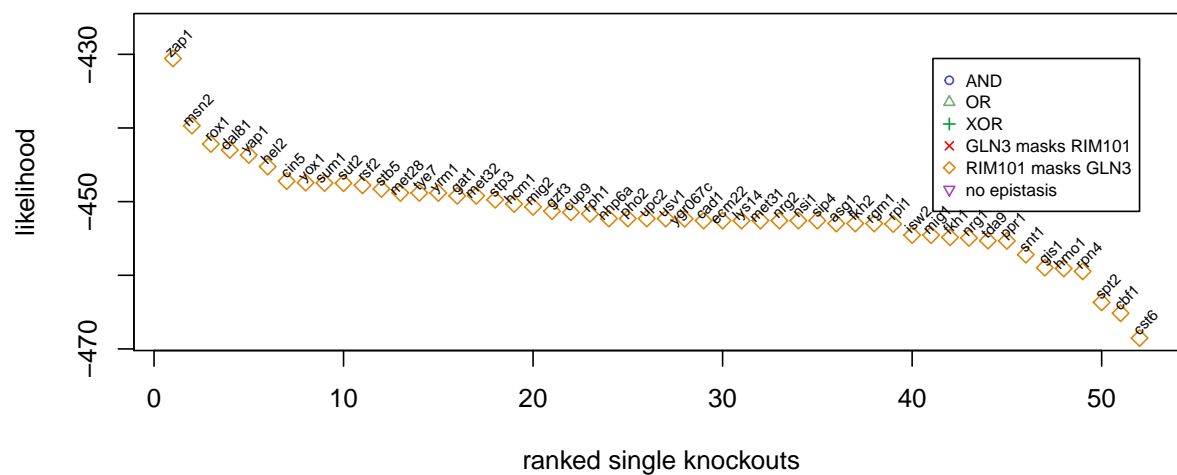
gln3 and isw2



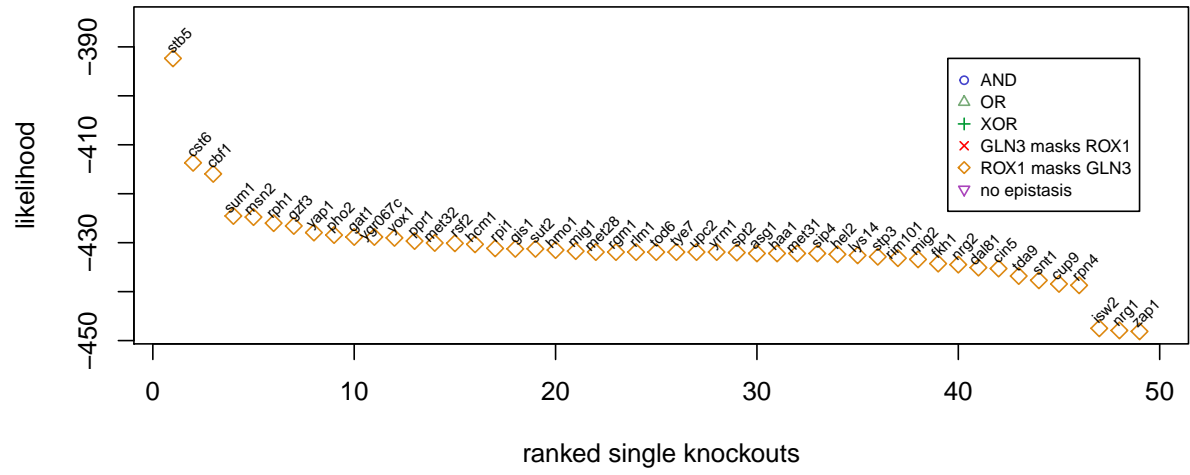
gln3 and mig1



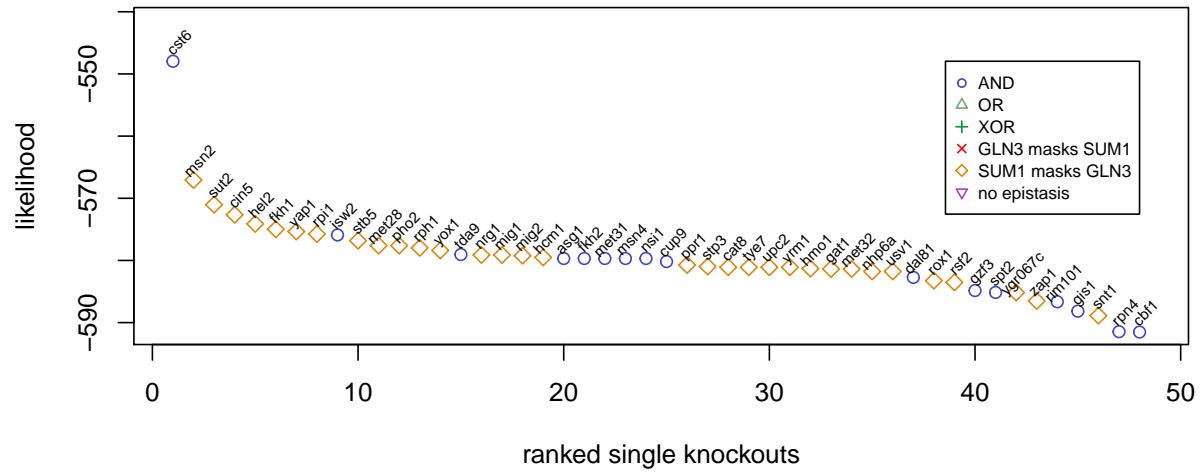
gln3 and rim101

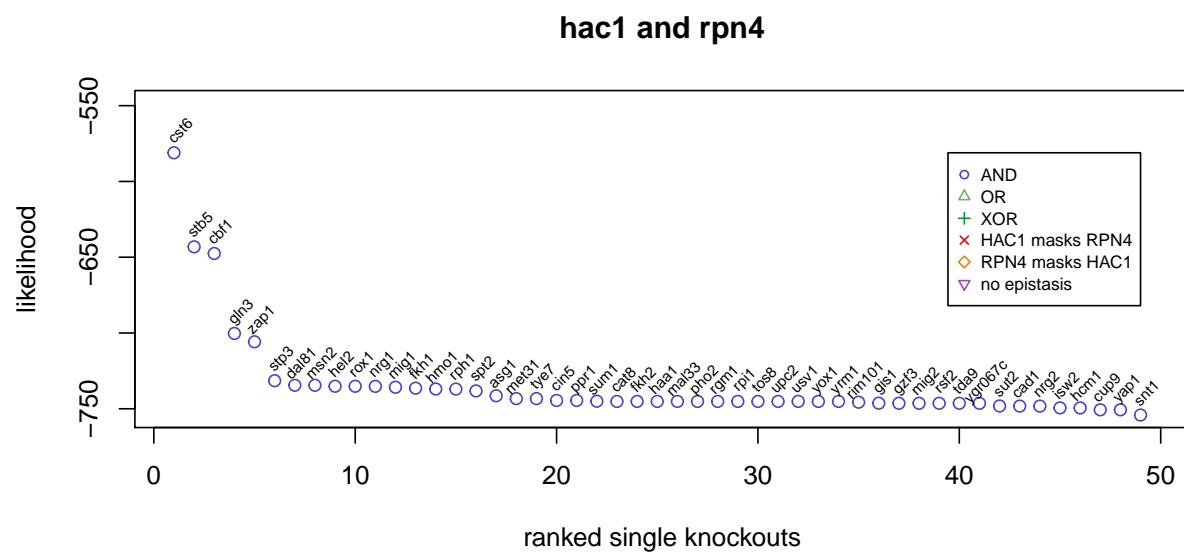
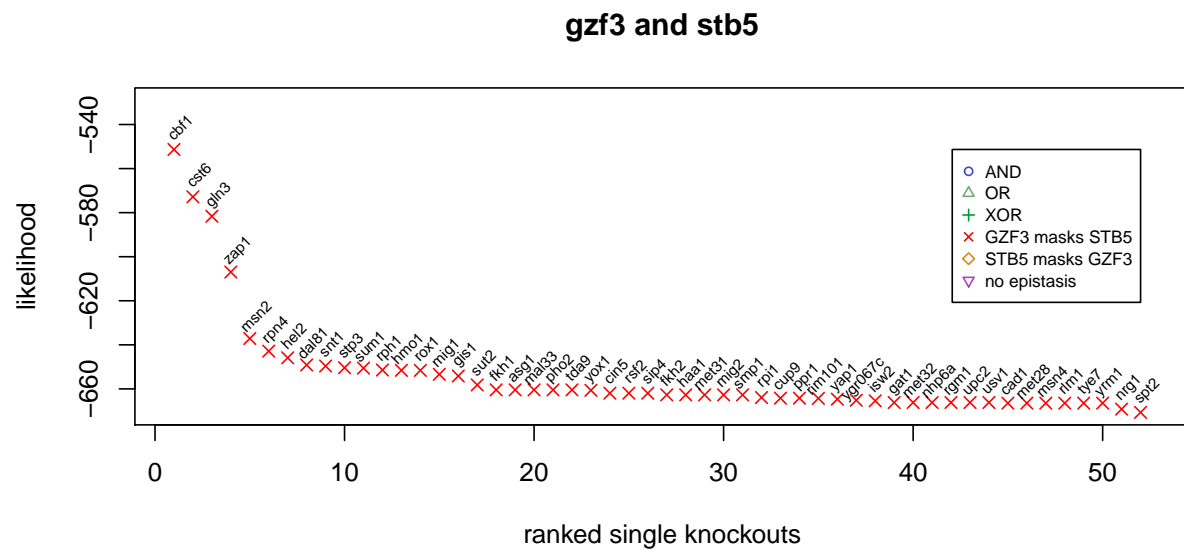


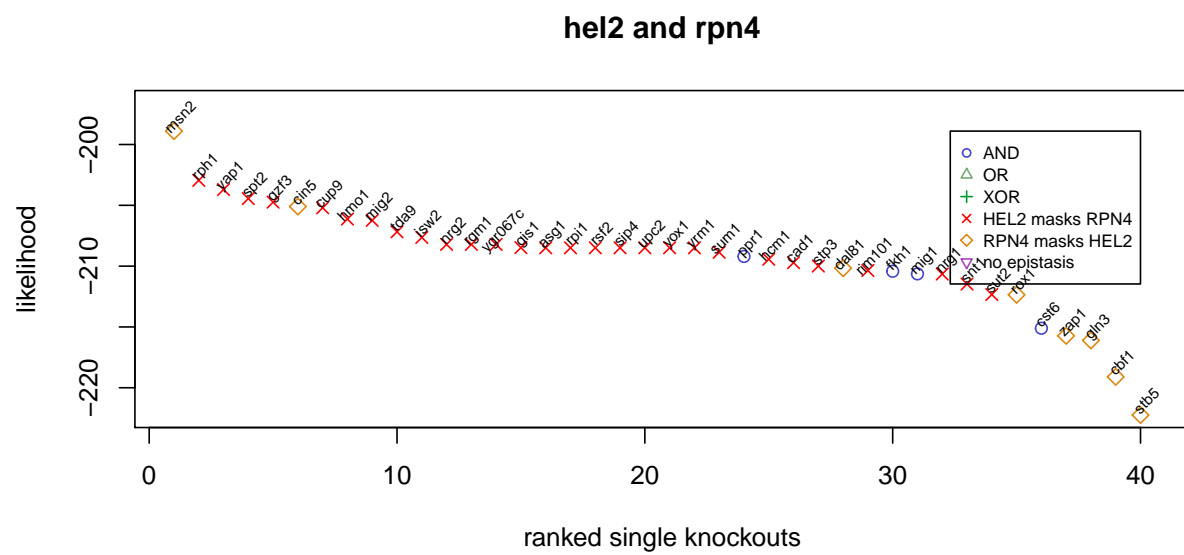
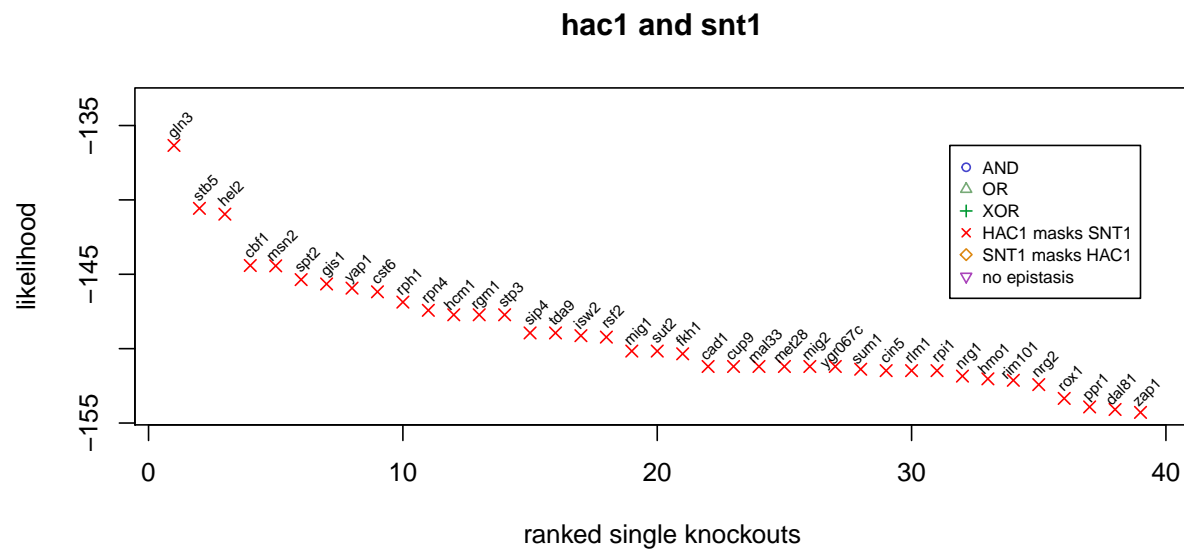
gln3 and rox1

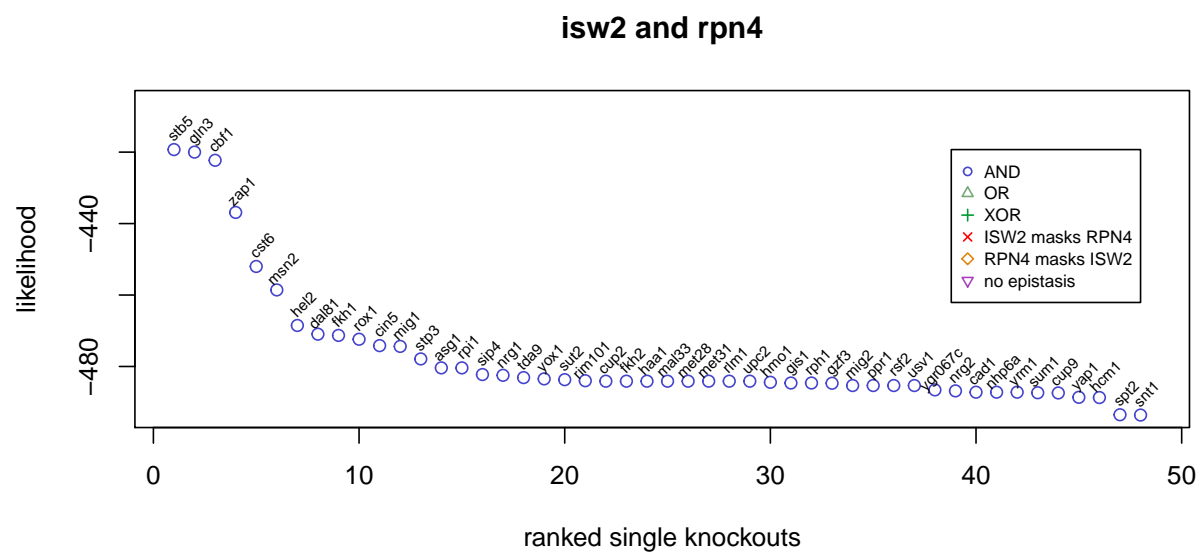
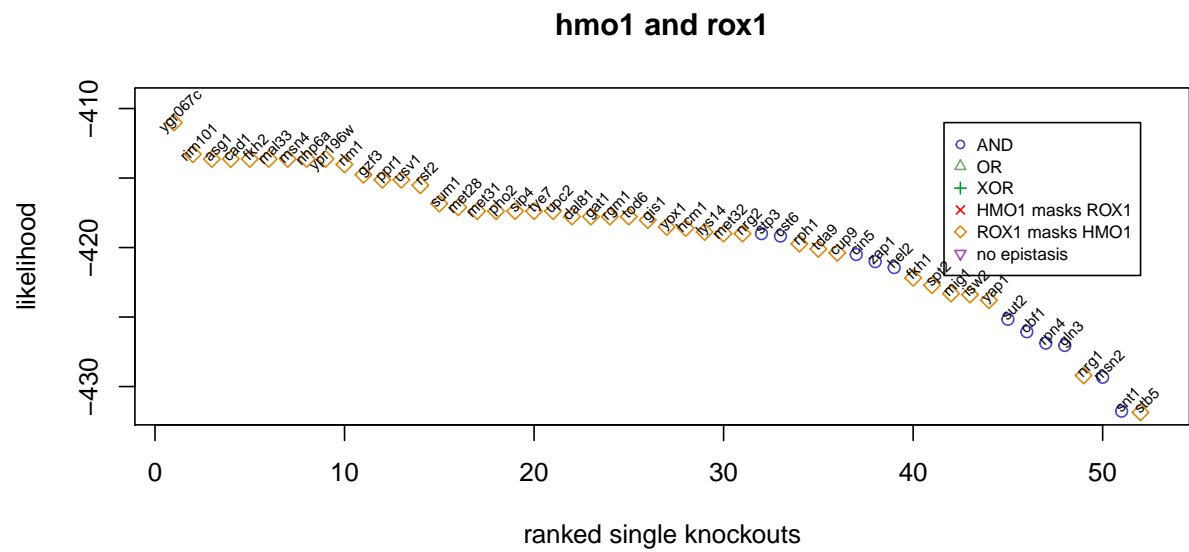


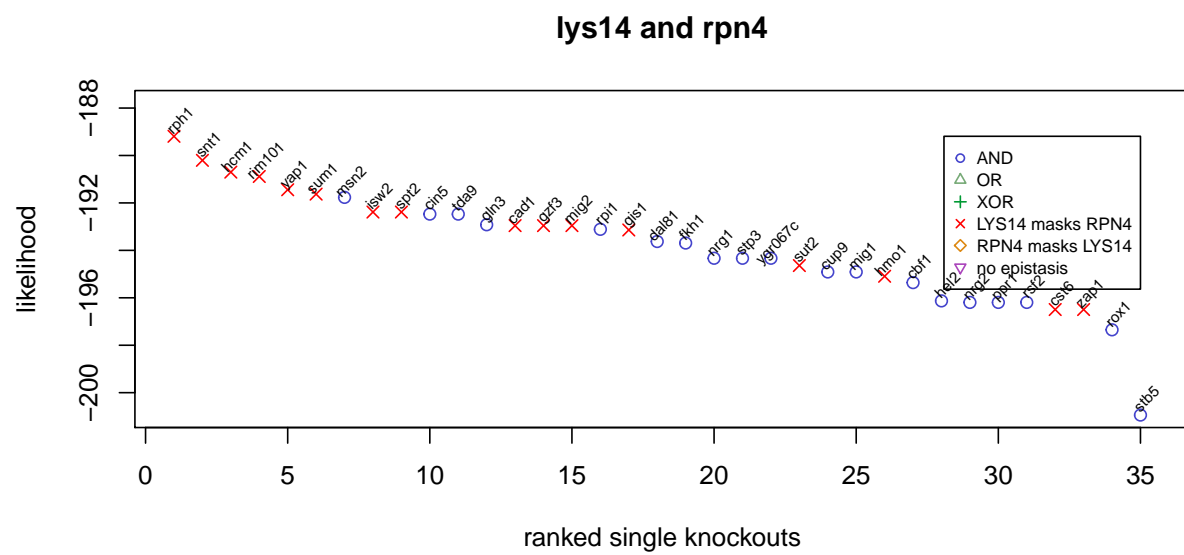
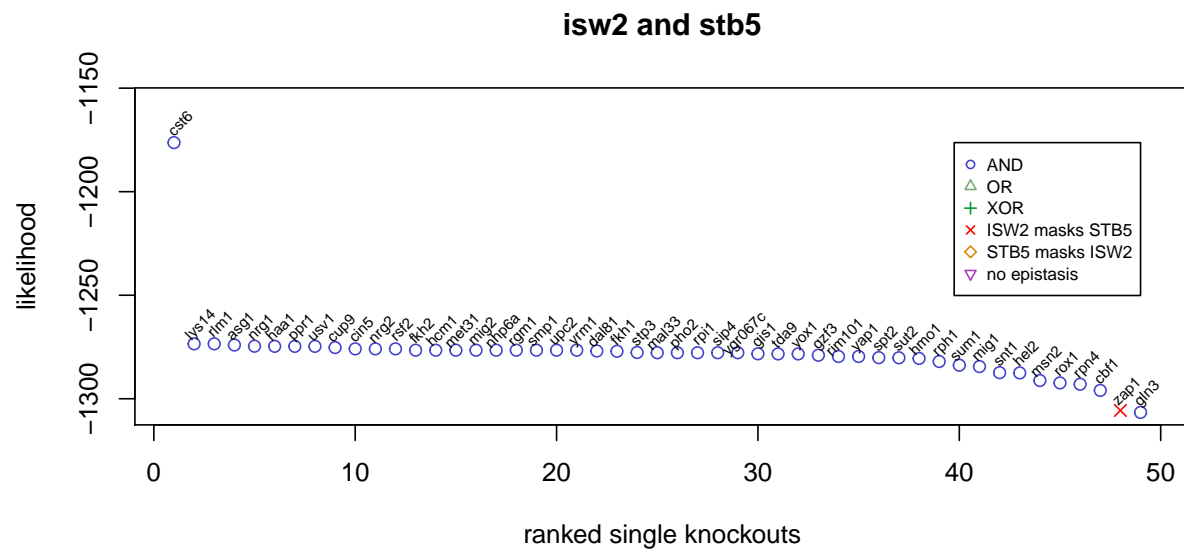
gln3 and sum1

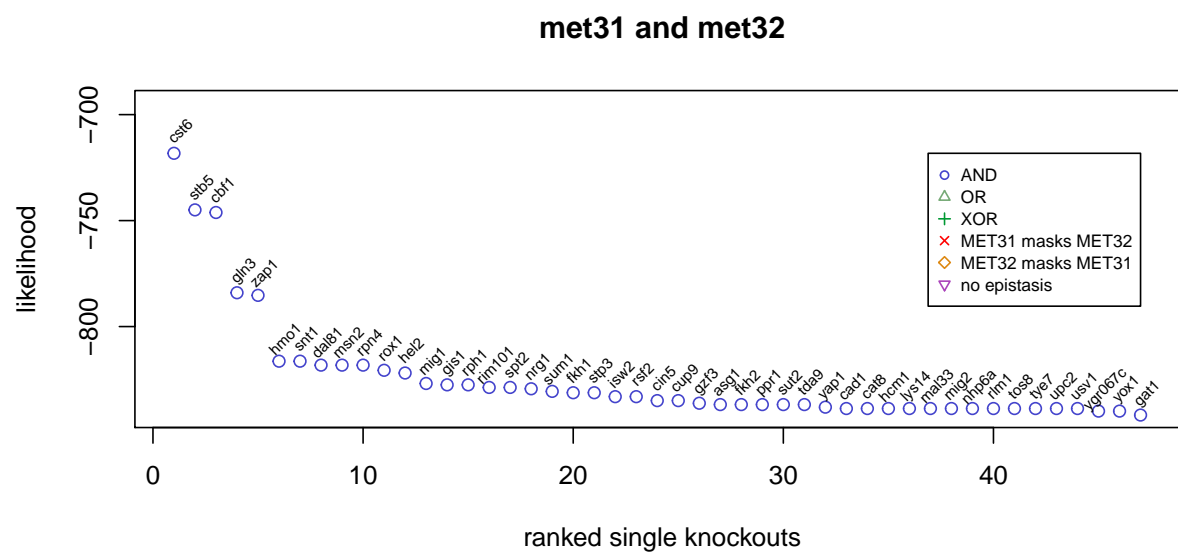
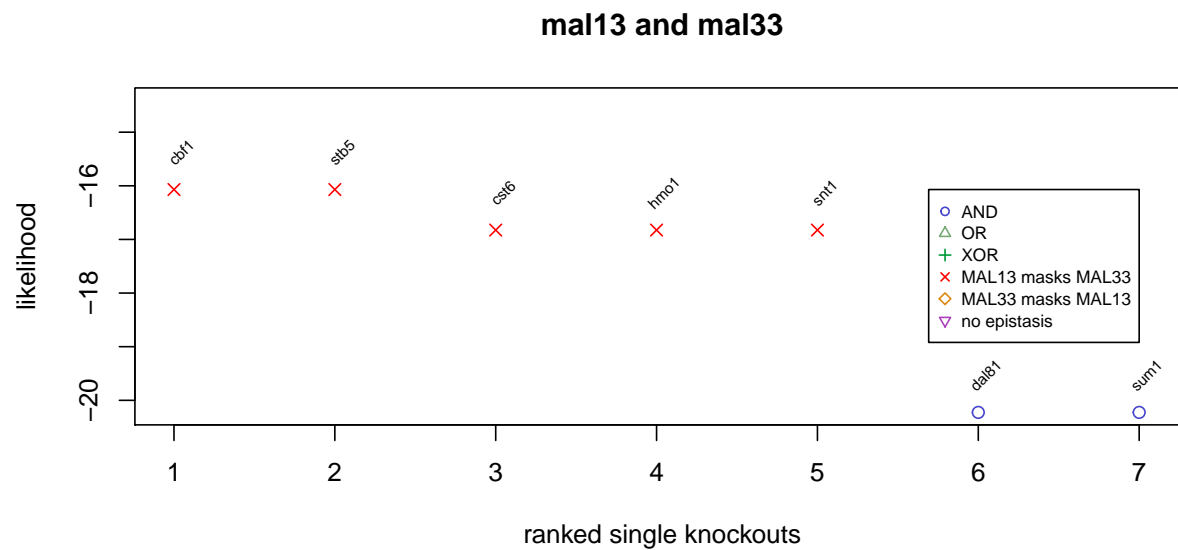




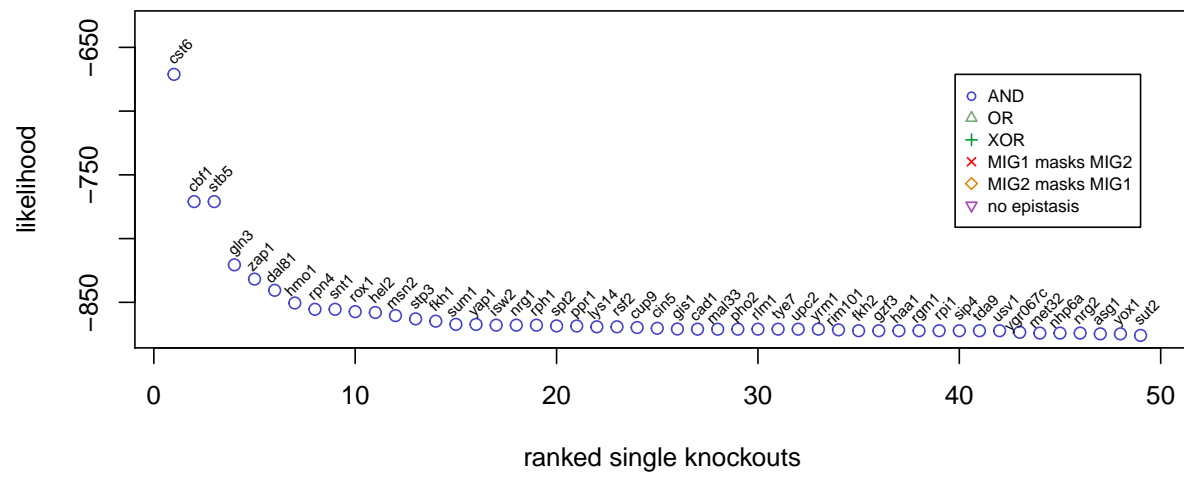




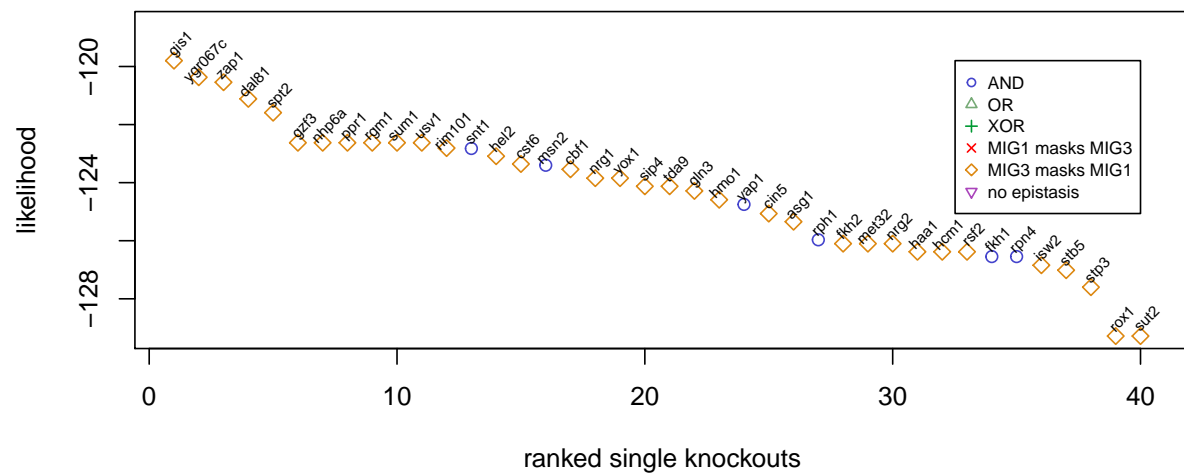




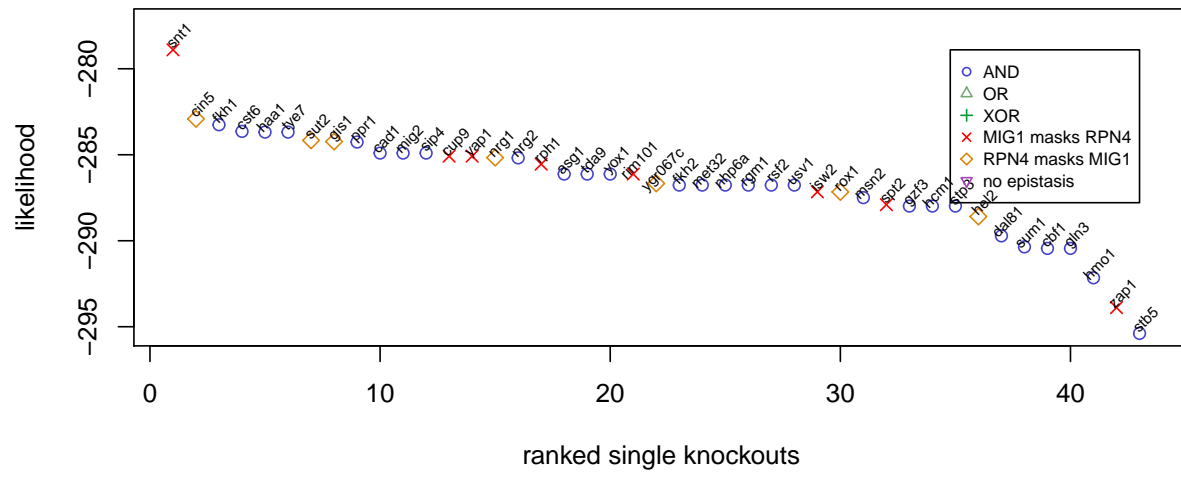
mig1 and mig2



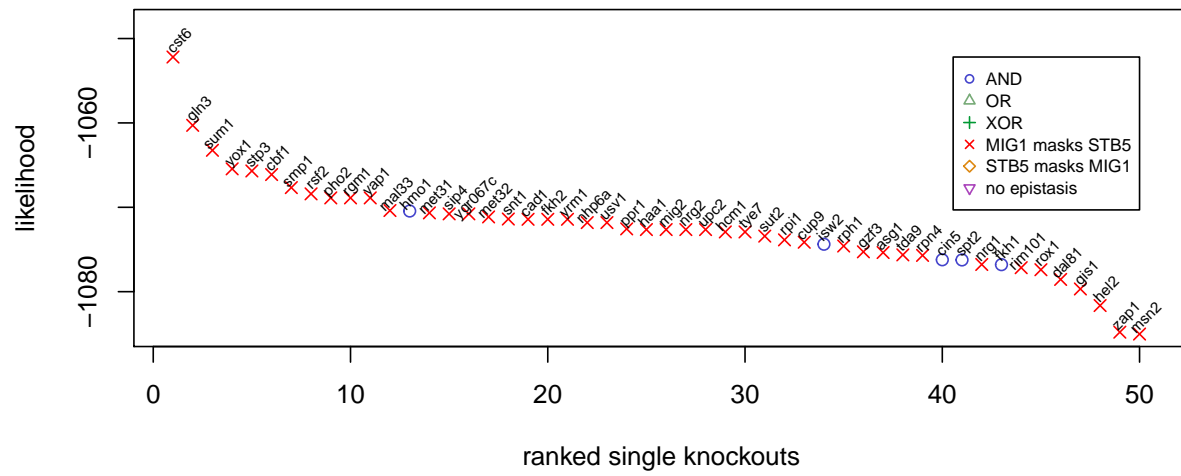
mig1 and mig3



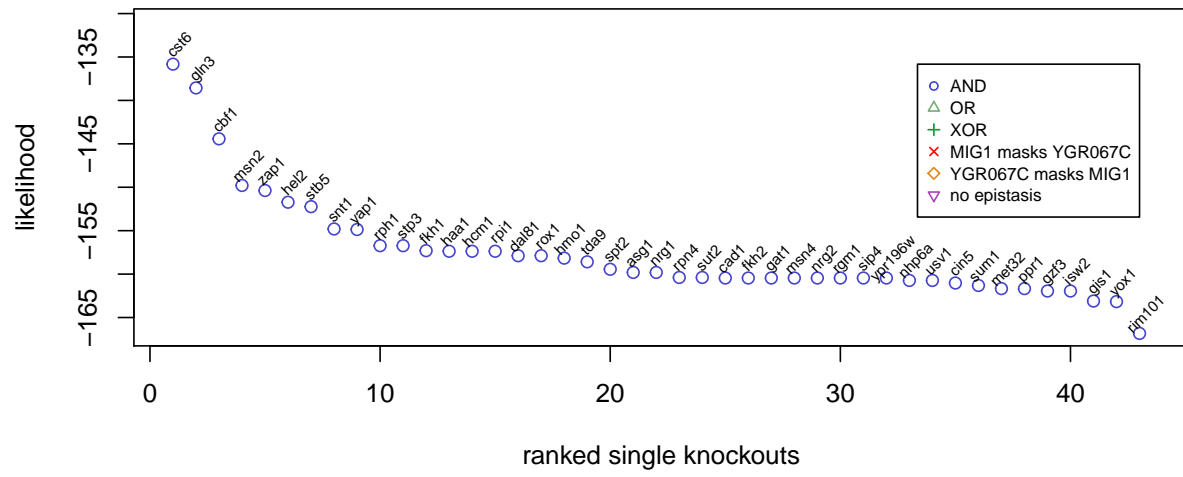
mig1 and rpn4



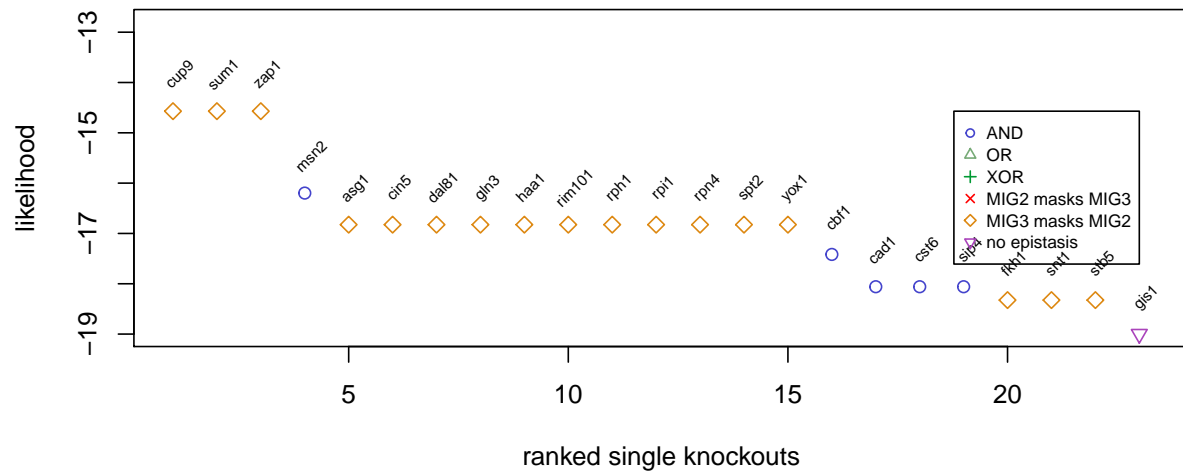
mig1 and stb5

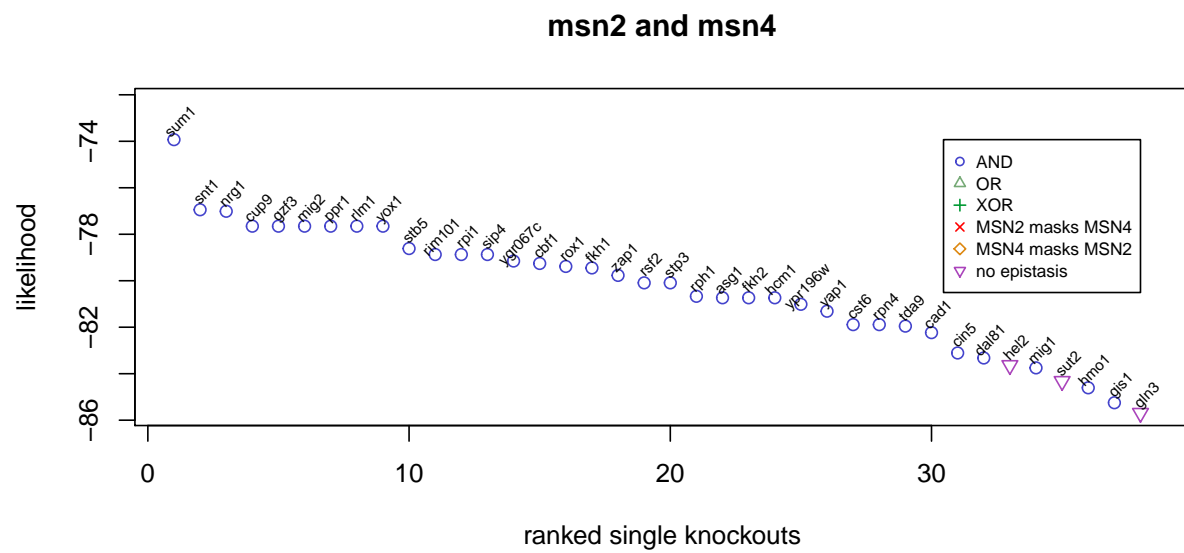
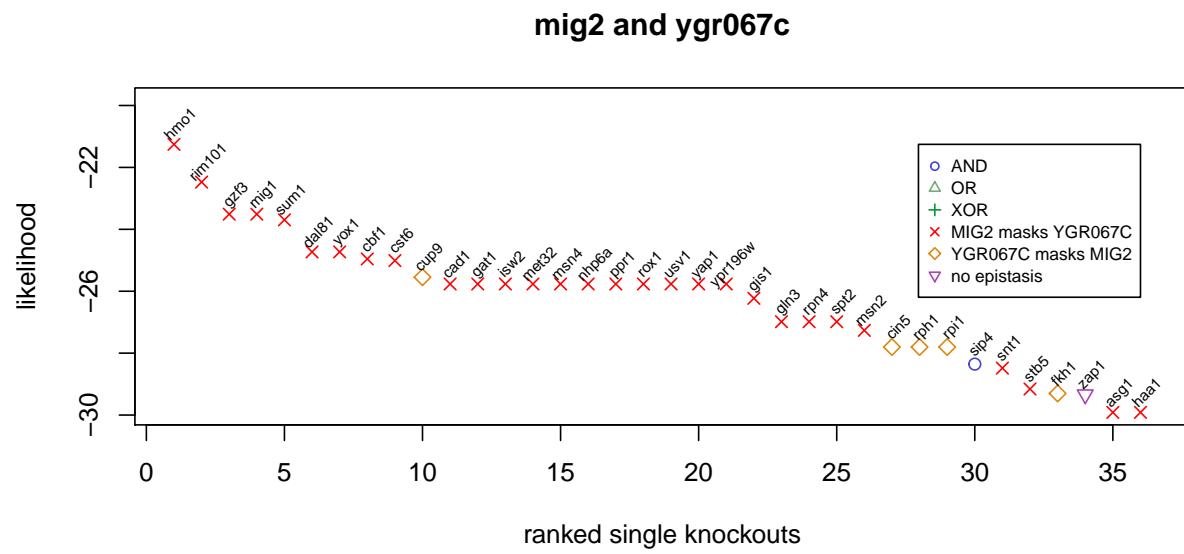


mig1 and ygr067c

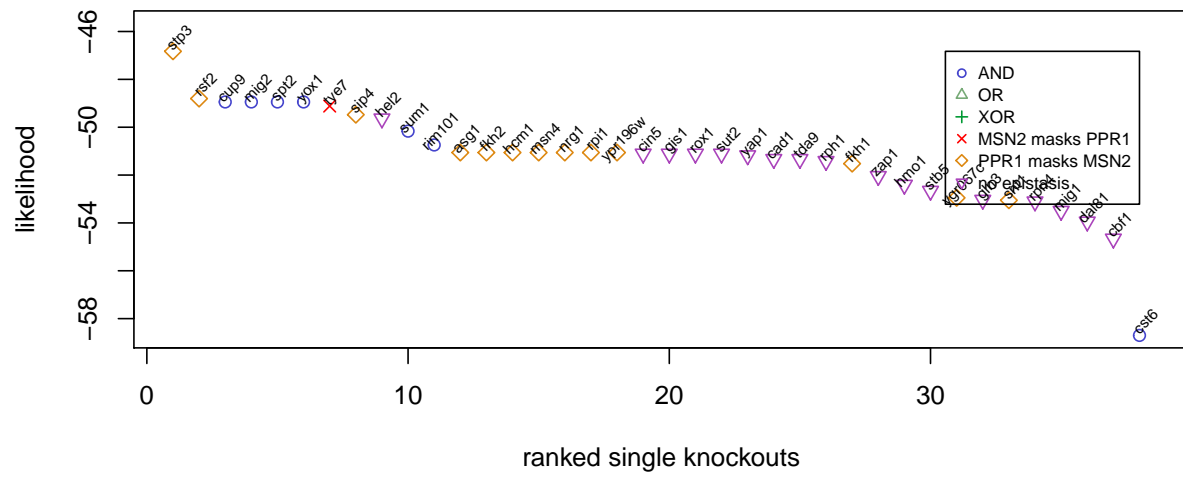


mig2 and mig3

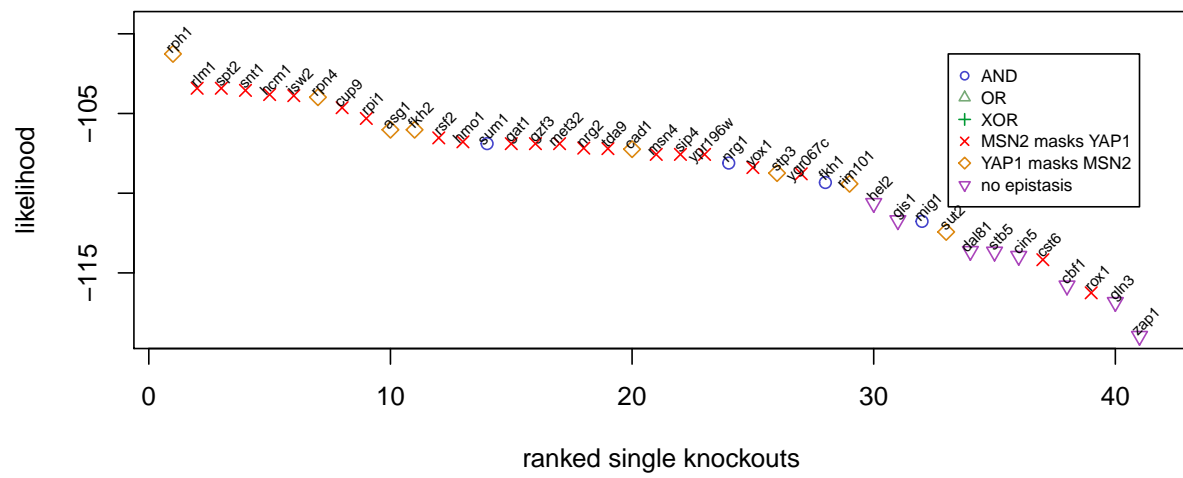




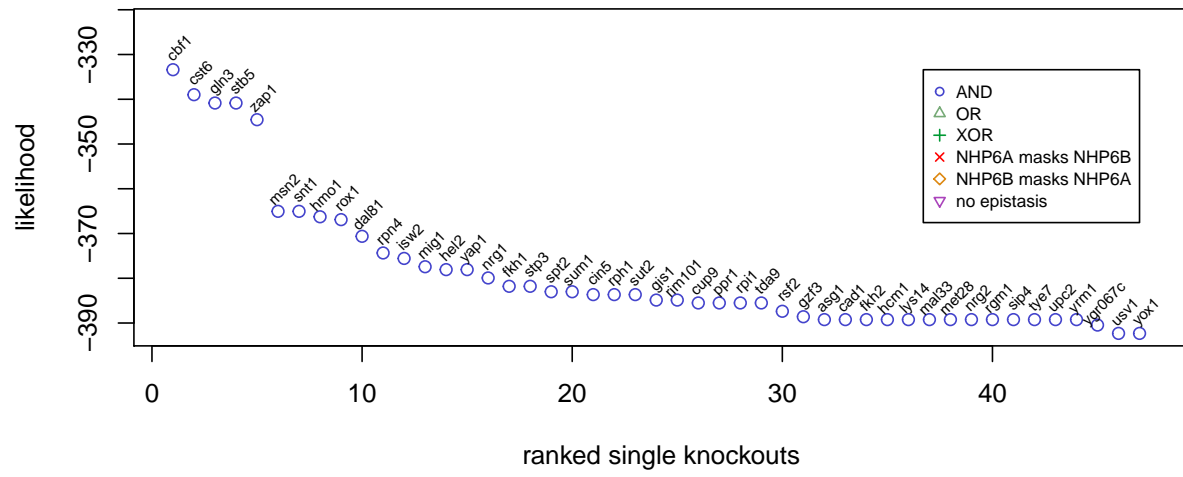
msn2 and ppr1



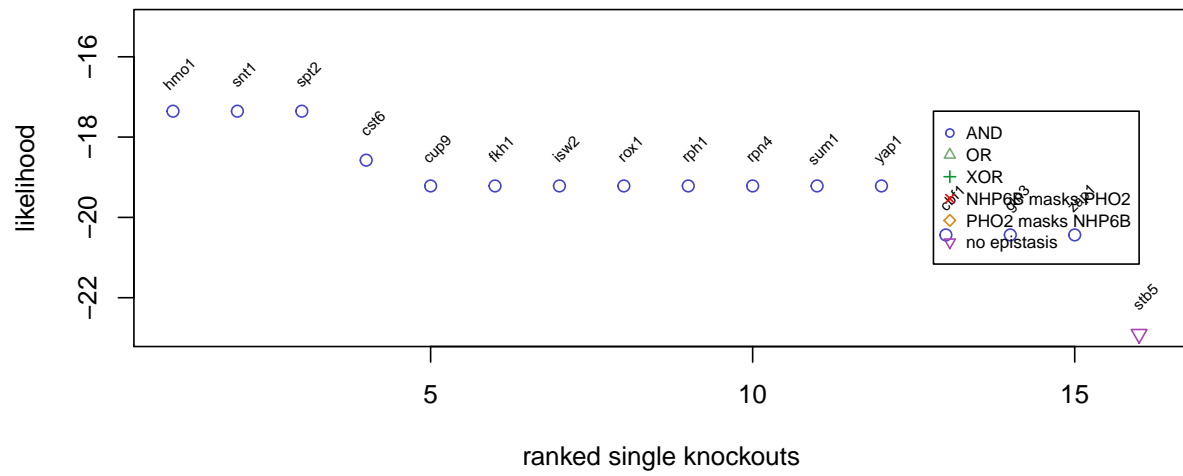
msn2 and yap1



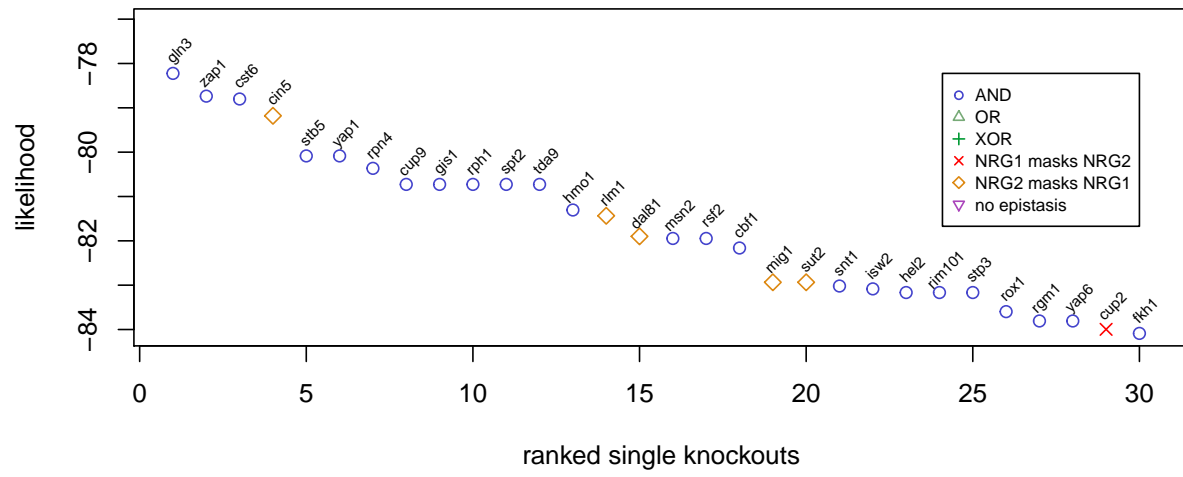
nhp6a and nhp6b



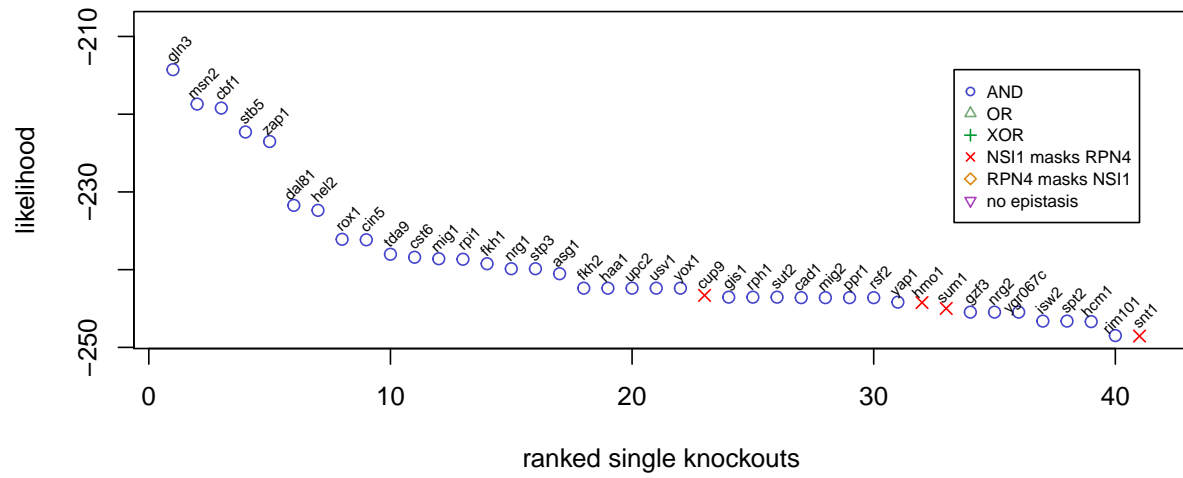
nhp6b and pho2



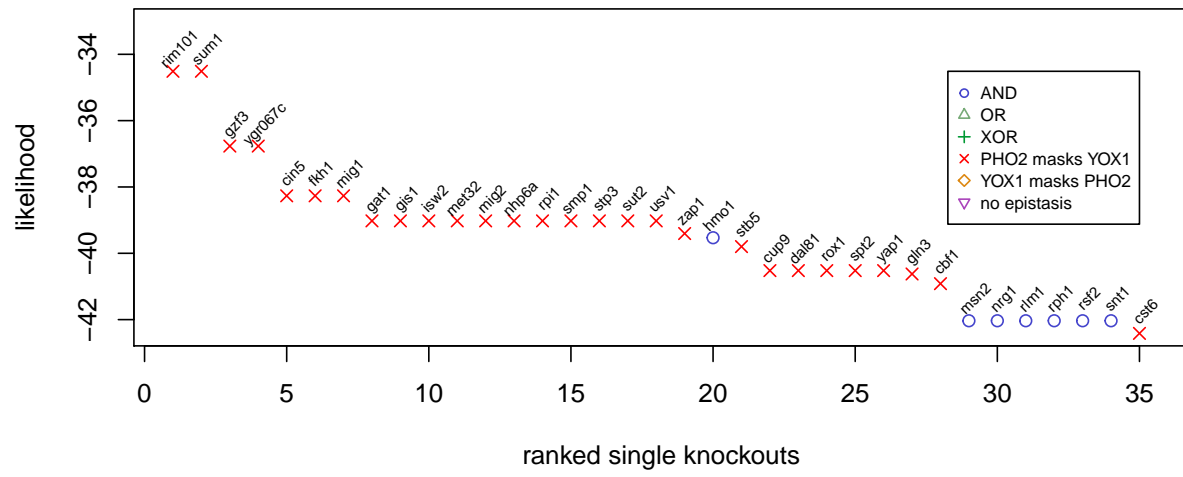
nrg1 and nrg2



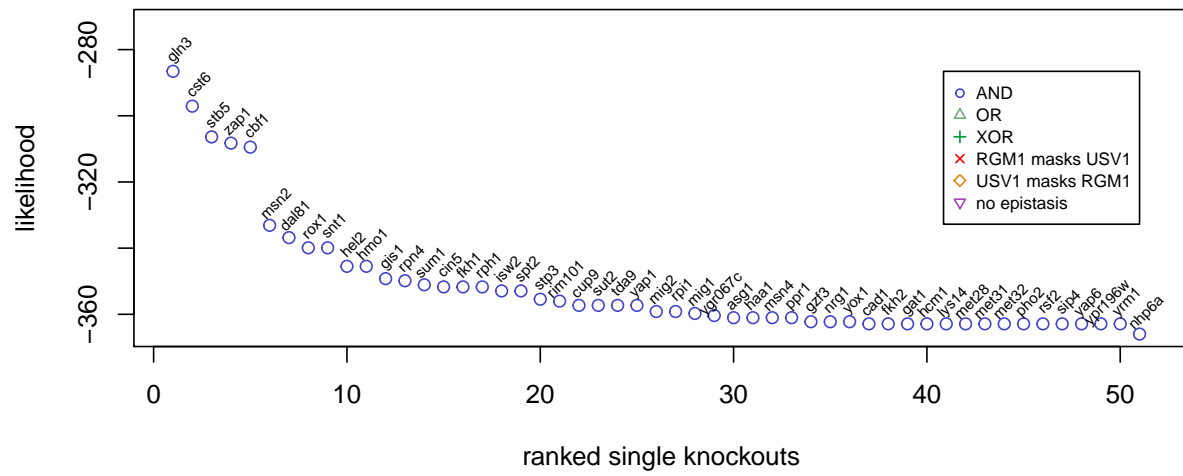
nsi1 and rpn4

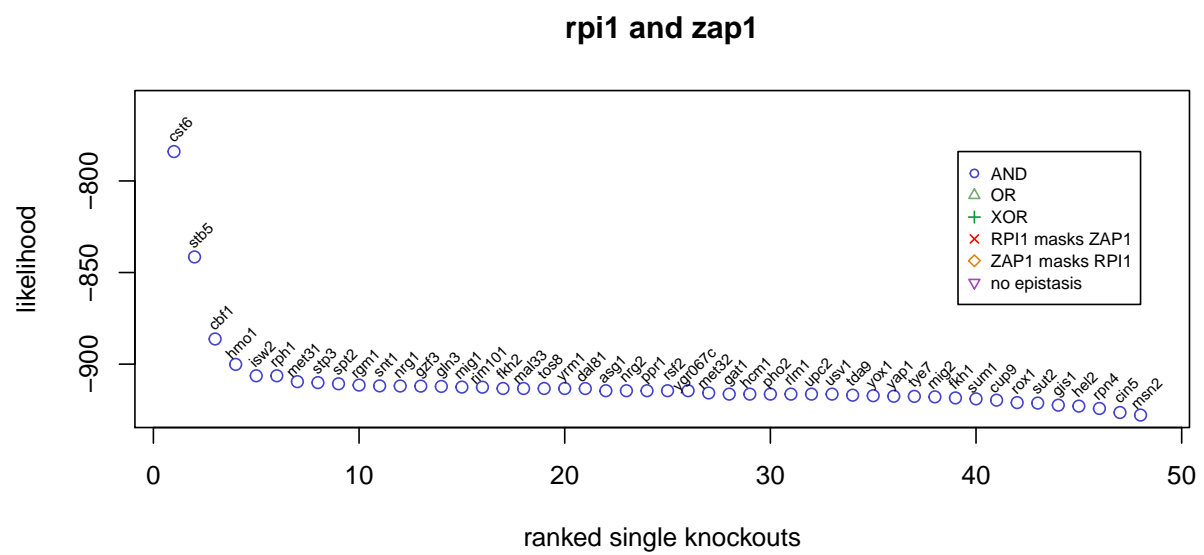
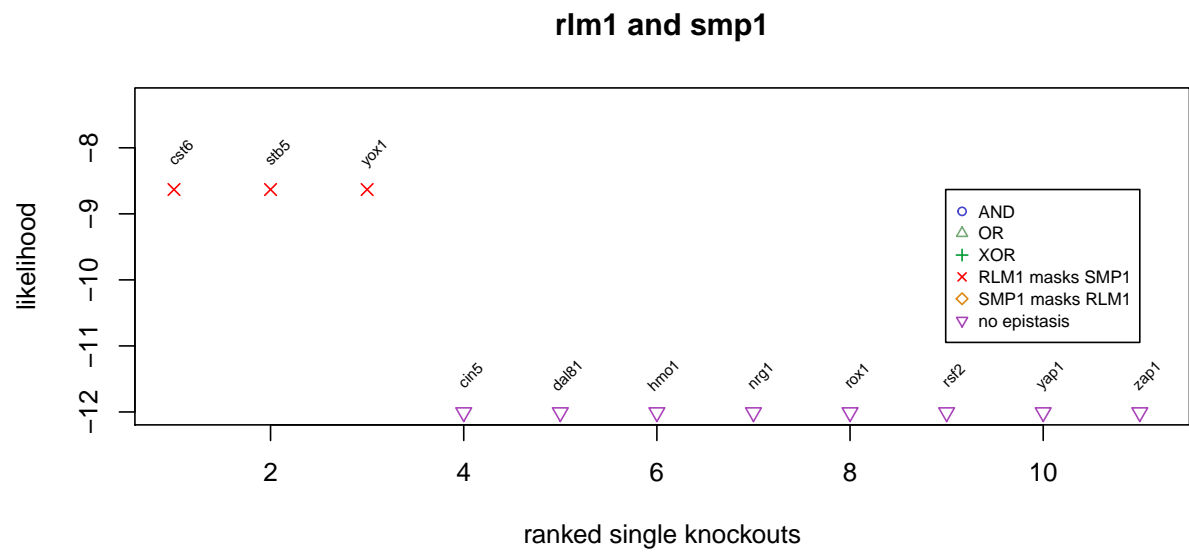


pho2 and yox1

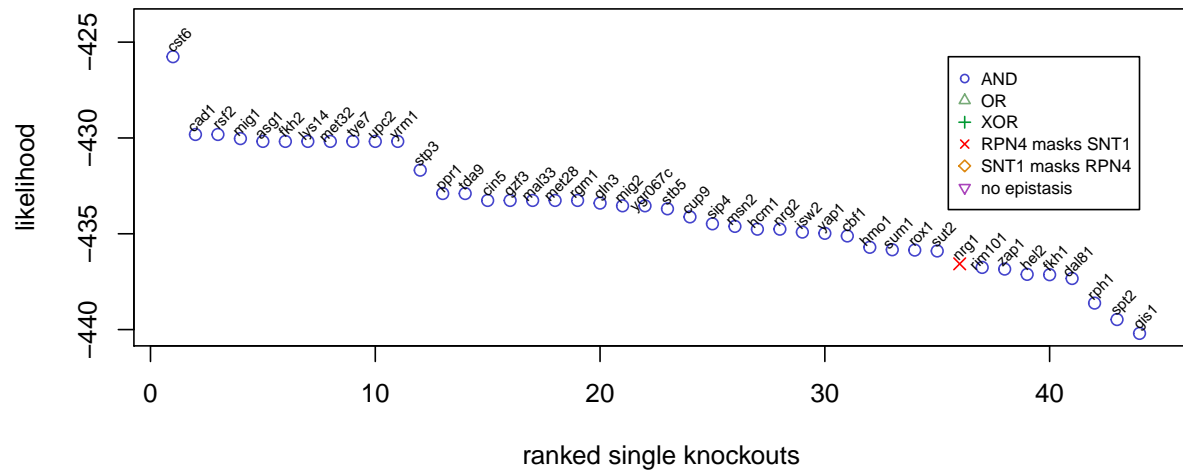


rgm1 and usv1

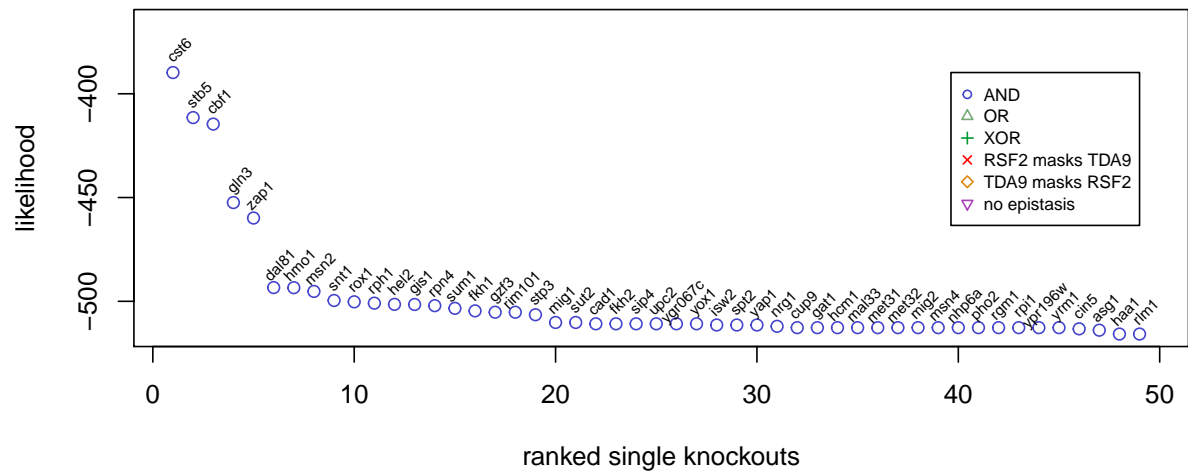


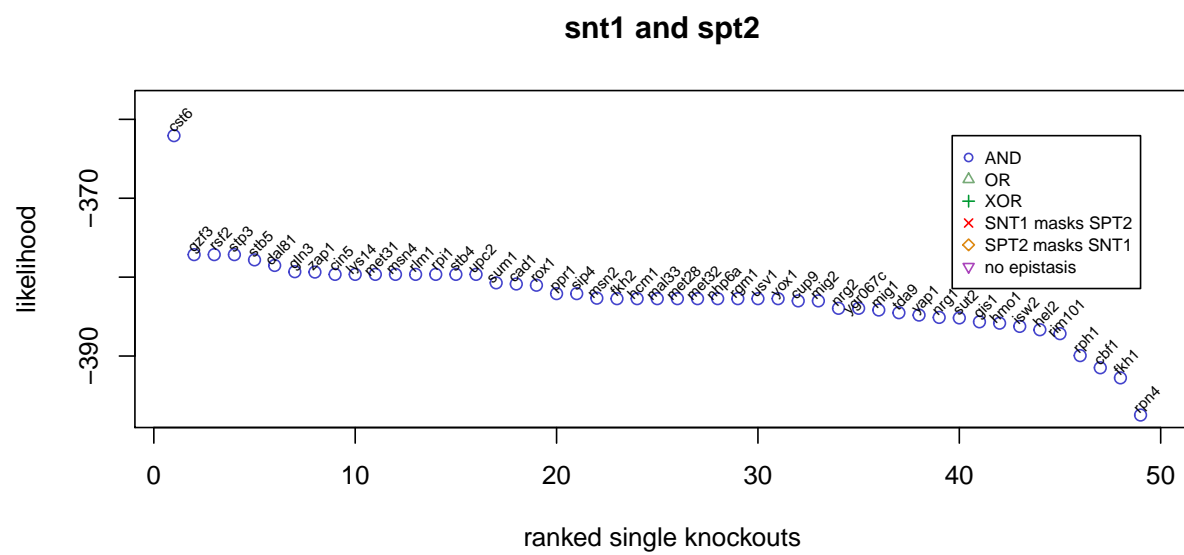
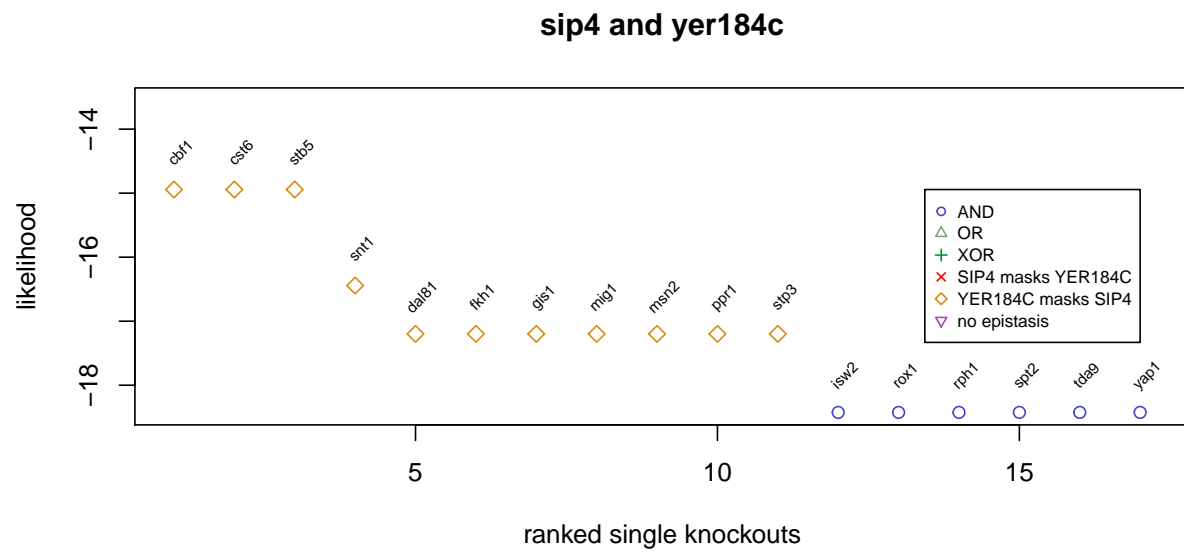


rpn4 and snt1

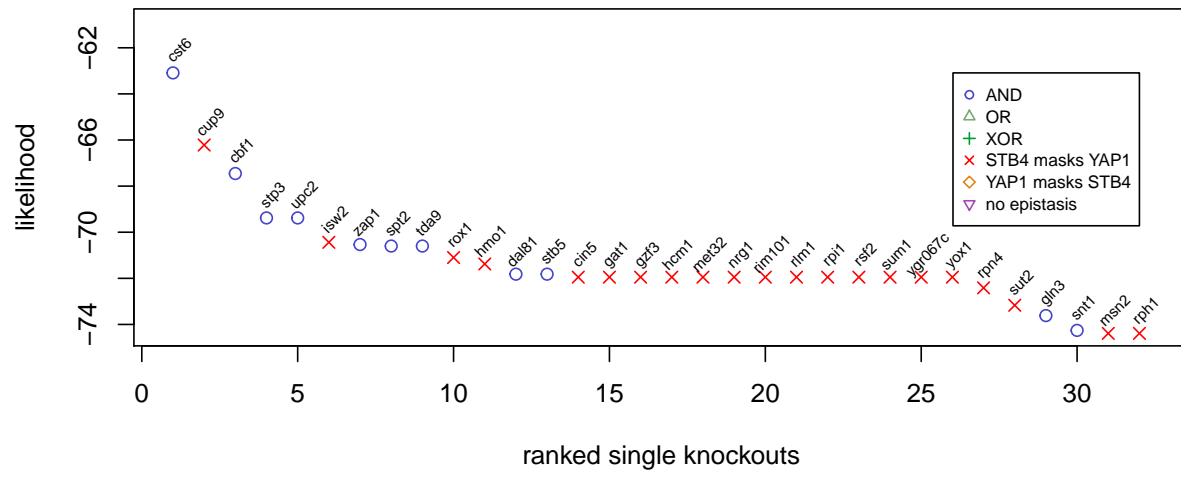


rsf2 and tda9

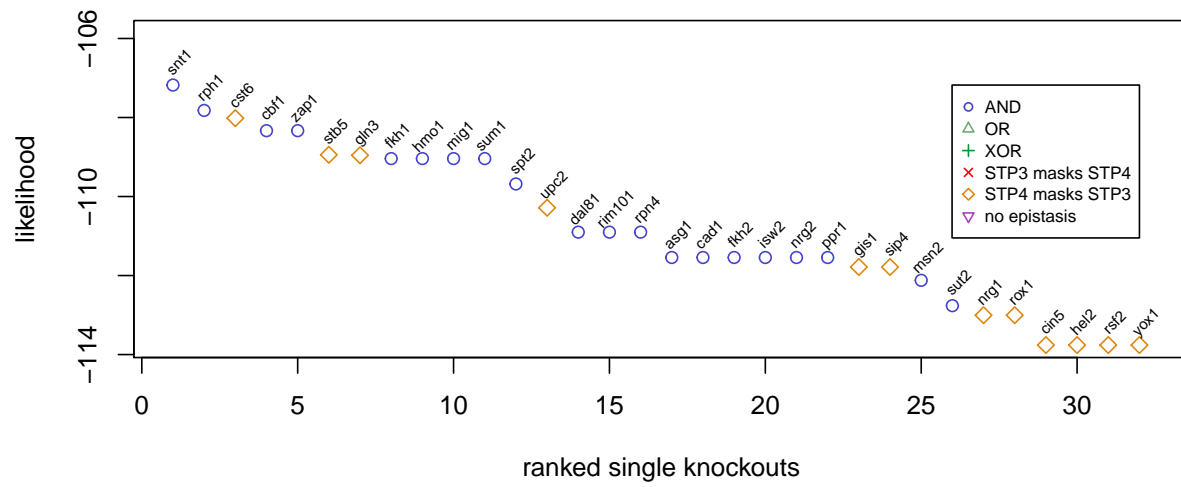


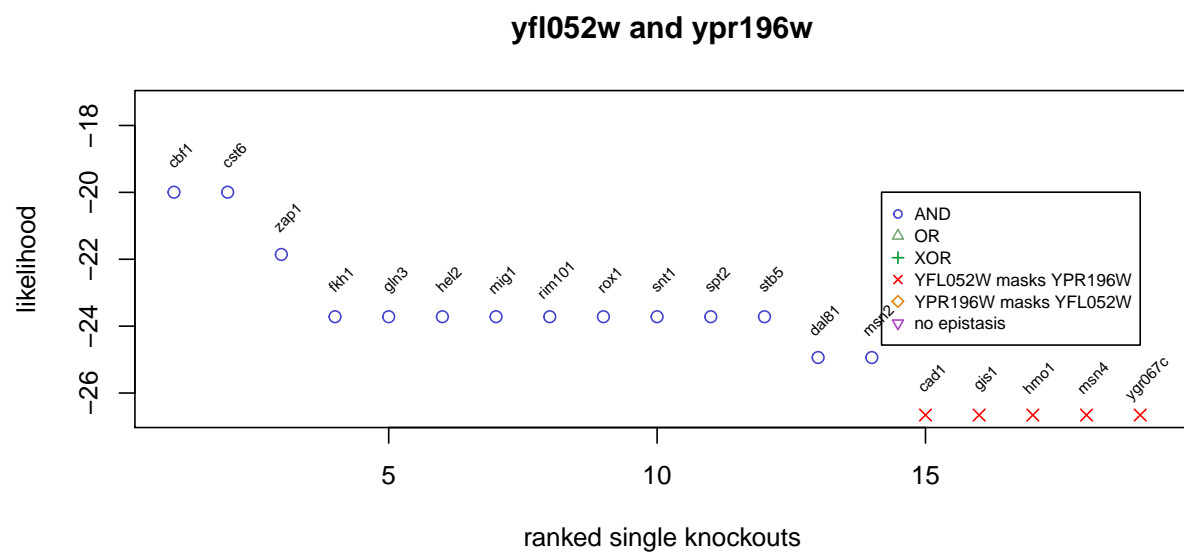
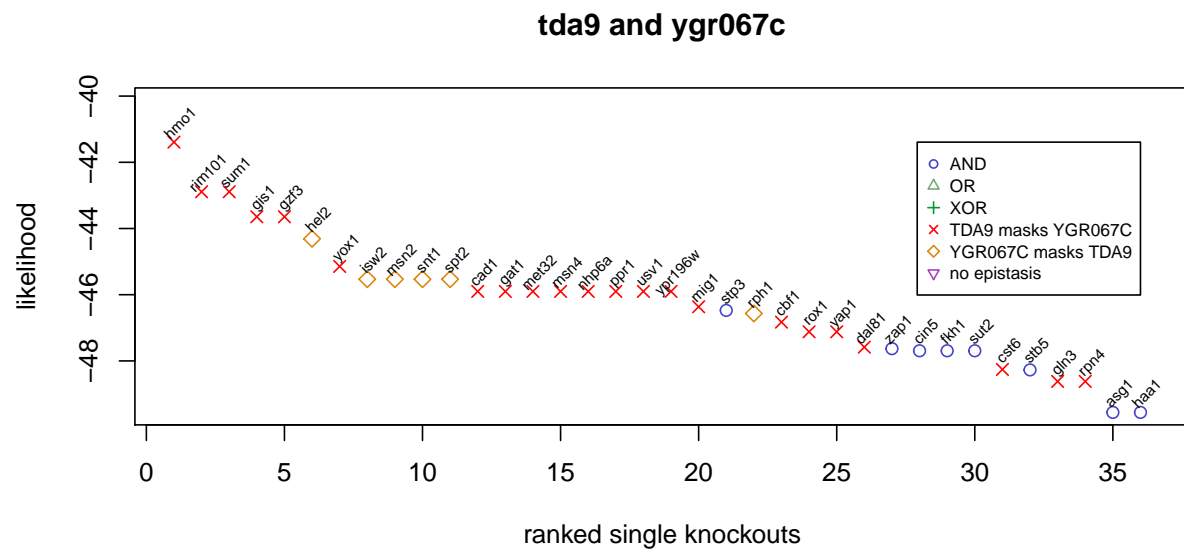


stb4 and yap1

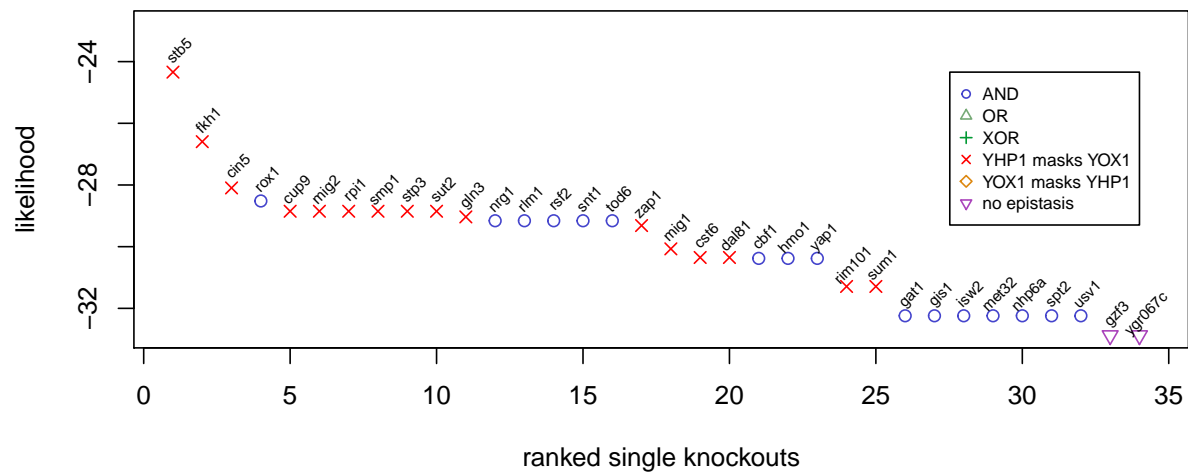


stp3 and stp4

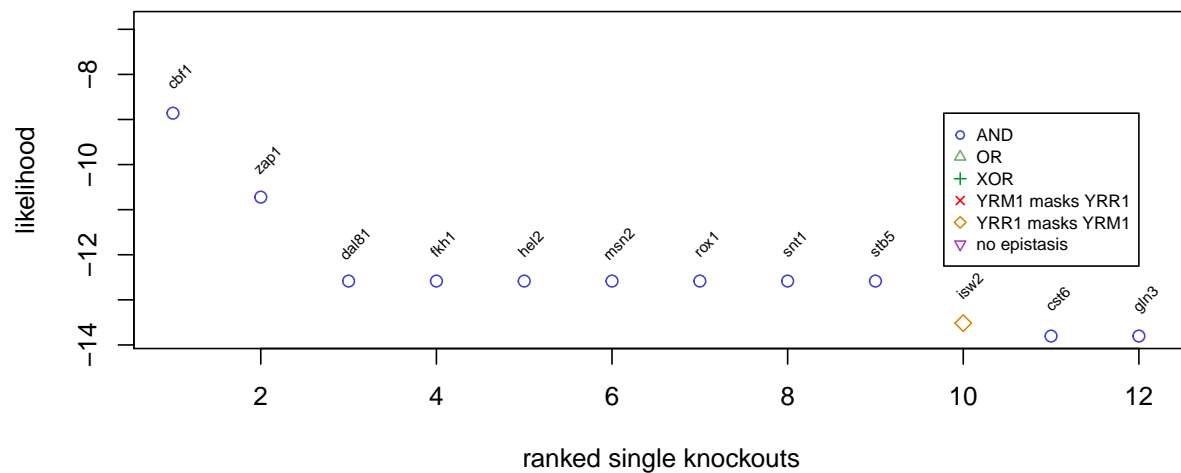




yhp1 and yox1



yrm1 and yrr1



```
distmat <- sameith$logic
```

```
distmat[which(distmat %in% "AND")] <- 1
distmat[which(distmat %in% "OR")] <- 2
distmat[which(distmat %in% "XOR")] <- 3
distmat[which(distmat %in% "NOEPI")] <- 6
distmat[which(distmat %in% c("NOINFO", "NOINF"))] <- 7
```

```
for (i in 1:ncol(distmat)) {
```

```
  genes <- unlist(strsplit(colnames(distmat)[i], "\\."))
```

```
  distmat[which(distmat[, i] %in% paste(genes[1], " masks the effect of ",
```

```

genes[2], sep = "")), i] <- 4

distmat[which(distmat[, i] %in% paste(genes[2], " masks the effect of ",
genes[1], sep = "")), i] <- 5
}

distmat <- apply(distmat, c(1,2), as.numeric)

for (i in 1:ncol(distmat)) {
  distmat[, i] <- rev(sort(distmat[, i]))
}

distmat <- distmat[-which(apply(distmat, 1, sum) == 0), ]

library(bnem)

y <- distmat

distmat <- distmat[, order(apply(distmat, 2, function(x) { return(sum(x == 1)) }))]

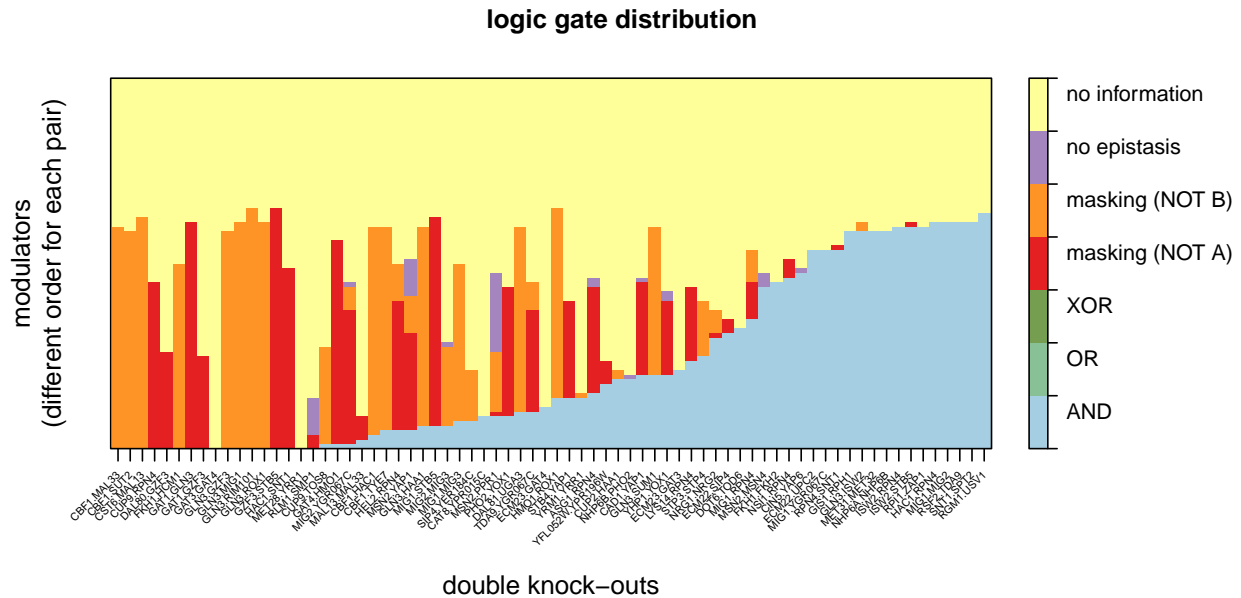
y[which(y == 5)] <- 4

rownames(distmat) <- NULL

labeltext <- c("", "no information\n\n\n", "no epistasis\n\n\n",
  "masking (NOT B)\n\n\n", "masking (NOT A)\n\n\n",
  "XOR\n\n\n", "OR\n\n\n", "AND\n\n\n")

heatmapOP(distmat, Colv = F, Rowv = F, main = "logic gate distribution", sub = "",
  col = "Paired", breaks = seq(0.5,7.5, length.out = 8), cexRow = 0,
  cexCol = 0.4, aspect = "fill",
  colorkey = list(space = "right", labels = rev(labeltext), width = 1,
    at = seq(1.5,7.5, length.out = 8)),
  xlab = "double knock-outs",
  ylab = "modulators\n(different order for each pair)",
  xrot = 45, bordercol = "transparent")

```

Now we plot the densities of the string-db interaction scores of our identified modulators and a random draw.

```
par(mfrow=c(1,2))

library(Stringdb)

get_STRING_species(version="10", species_name=NULL)[26, ] # 4932

##      species_id      official_name      compact_name  kingdom
## 26      4932 Saccharomyces cerevisiae Saccharomyces cerevisiae eukaryota
##      type
## 26 core

string_db <- STRINGdb$new( version="10", species=4932, score_threshold=0,
                           input_directory=~/" )

llmat <- wageningen$ll

logicmat <- wageningen$logic

string.scores <- list()

string.names <- character()

for (i in 1:ncol(llmat)) {

  if (sum(!(llmat[, i] %in% c(0,-Inf))) > 0) {
    top30 <- llmat[, i]
    top30[which(top30 == 0)] <- -Inf
    top30 <- top30[which(!(llmat[, i] %in% c(0,-Inf)))]
    top30 <- top30[order(top30,decreasing = T)[1:min(30, sum(!(llmat[, i]
      %in% c(0,-Inf)))]]
```

```

doubles <- unlist(strsplit(colnames(llmat)[i], "\\."))

for (j in names(top30)) {
  tmp <- string_db$get_interactions(string_db$mp(c(doubles[1], j)))
  string.scores <- c(string.scores, tmp$combined_score)
  string.names <- c(string.names, paste(sort(c(doubles[1], j)), collapse = "_"))
  tmp <- string_db$get_interactions(string_db$mp(c(doubles[2], j)))
  string.scores <- c(string.scores, tmp$combined_score)
  string.names <- c(string.names, paste(sort(c(doubles[2], j)), collapse = "_"))
}

} else {
  next()
}
}

```

```

data(wageningen_string)

tmp <- string_db$get_interactions(string_db$mp(unique(unlist(strsplit(colnames(dataBinWag), "\\.")))))

stsc <- unlist(string.scores)

denspval <- wilcox.test(stsc, unlist(tmp$combined_score), alternative = "greater")$p.value

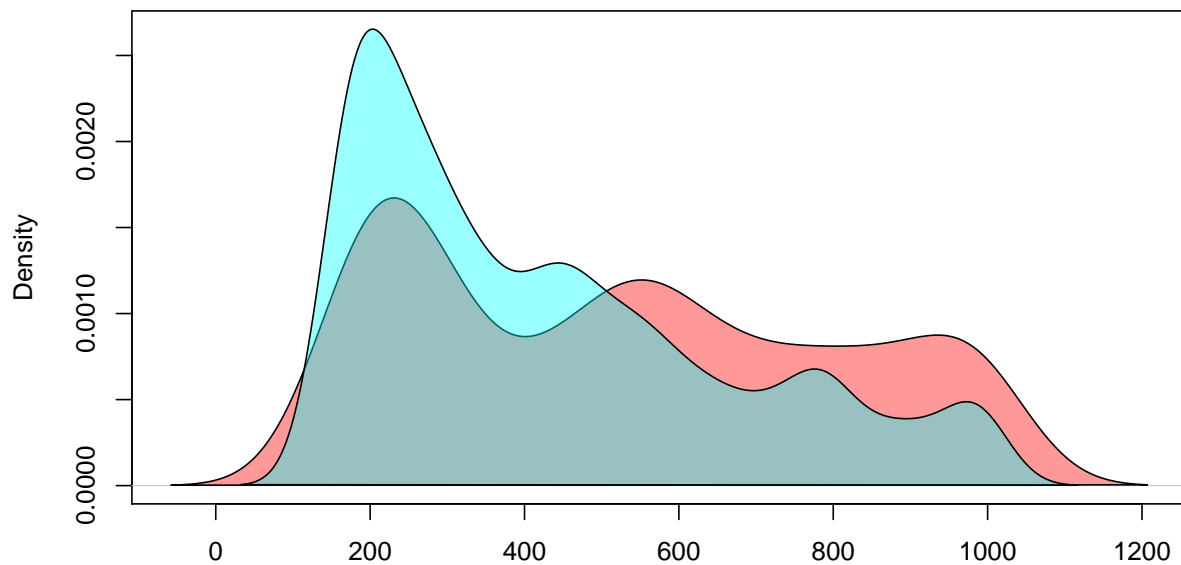
for (i in 100:1) {
  if (denspval < 10^(-i)) {
    denspval <- paste("< ", 10^(-i), sep = "")
  }
}

plot(density(stsc), col = "#00000000",
     ylim = c(0, max(c(max(density(stsc)$y), max(density(unlist(tmp$combined_score))$y)))),
     main = paste("Mann-Whitney test p-value ", denspval, sep = ""), xlab = "",
     cex.main = 1.5)
polygon(density(stsc), col = "#ff000066")

lines(density(unlist(tmp$combined_score)), col = "#00000000")
polygon(density(unlist(tmp$combined_score)), col = "#00ffff66")

```

Mann-Whitney test p-value < 1e-15



```
llmat <- sameith$ll
logicmat <- sameith$logic

string.scores2 <- list()
string.names2 <- character()

for (i in 1:ncol(llmat)) {

  if (sum(!(llmat[, i] %in% c(0,-Inf))) > 0) {
    top30 <- llmat[, i]
    top30[which(top30 == 0)] <- -Inf
    top30 <- top30[which(!(llmat[, i] %in% c(0,-Inf)))]
    top30 <- top30[order(top30, decreasing = T)[1:min(30, sum(!(llmat[, i]
      %in% c(0,-Inf))))]]

    doubles <- unlist(strsplit(colnames(llmat)[i], "\\."))

    for (j in names(top30)) {
      tmp <- string_db$get_interactions(string_db$mp(c(doubles[1], j)))
      string.scores2 <- c(string.scores2, tmp$combined_score)
      string.names2 <- c(string.names2, paste(sort(c(doubles[1], j)), collapse = "_"))
      tmp <- string_db$get_interactions(string_db$mp(c(doubles[2], j)))
      string.scores2 <- c(string.scores2, tmp$combined_score)
      string.names2 <- c(string.names2, paste(sort(c(doubles[2], j)), collapse = "_"))
    }
  }
}
```

```

    } else {
      next()
    }
  }

data(sameith_string)

tmp <- string_db$get_interactions(string_db$tmp(unique(unlist(strsplit(colnames(dataBin)
, "\\."))))))

stsc <- unlist(string.scores2)

denspval <- wilcox.test(stsc, unlist(tmp$combined_score), alternative = "greater")$p.value

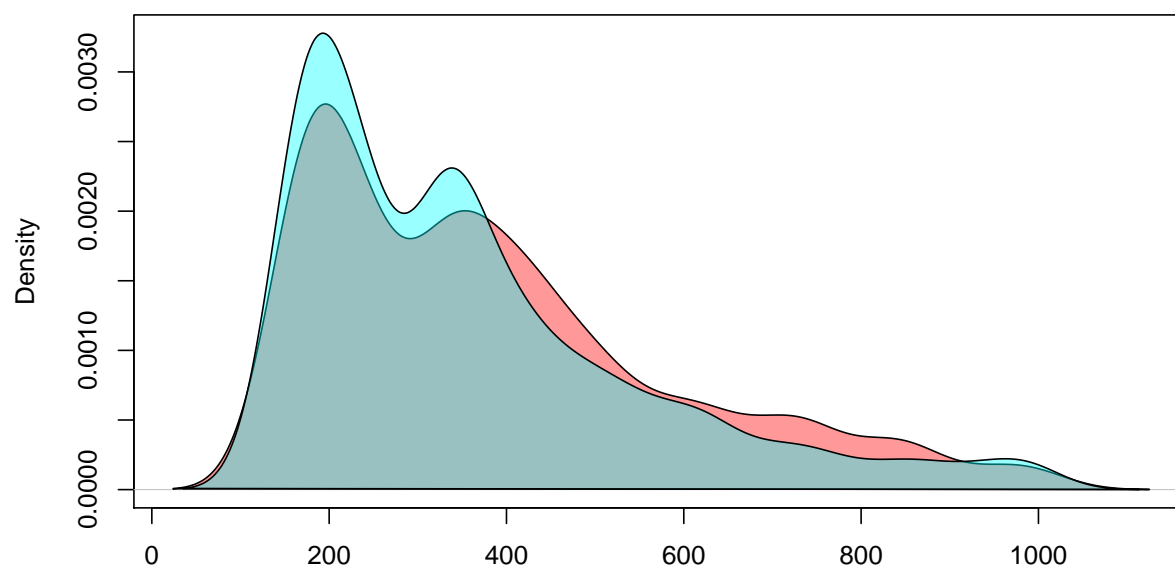
for (i in 100:1) {
  if (denspval < 10^(-i)) {
    denspval <- paste("< ", 10^(-i), sep = "")
  }
}

plot(density(stsc), col = "#00000000",
      ylim = c(0, max(c(max(density(stsc)$y), max(density(unlist(tmp$combined_score))$y)))),
      main = paste("Mann-Whitney test p-value ", denspval, sep = ""), xlab = "",
      cex.main = 1.5)
polygon(density(stsc), col = "#ff000066")

lines(density(unlist(tmp$combined_score)), col = "#00000000")
polygon(density(unlist(tmp$combined_score)), col = "#00ffff66")

```

Mann-Whitney test p-value < 1e-04



```
sessionInfo()
```

```
## R version 3.3.2 (2016-10-31)
## Platform: x86_64-apple-darwin13.4.0 (64-bit)
## Running under: OS X El Capitan 10.11.5
##
## locale:
## [1] C/UTF-8/C/C/C/C
##
## attached base packages:
## [1] grid      parallel  stats      graphics  grDevices  utils      datasets
## [8] methods   base
##
## other attached packages:
## [1] bnem_0.99.0      latticeExtra_0.6-28 RColorBrewer_1.1-2
## [4] lattice_0.20-34  snowfall_1.84-6.1  snow_0.4-2
## [7] matrixStats_0.51.0 nem_2.48.0          CellNOptR_1.20.0
## [10] XML_3.98-1.5      Rgraphviz_2.18.0    RCurl_1.95-4.8
## [13] bitops_1.0-6      ggplot2_2.2.0        hash_2.2.6
## [16] RBGL_1.50.0        graph_1.52.0         BiocGenerics_0.20.0
## [19] devtools_1.12.0   STRINGdb_1.14.0     minet_3.32.0
## [22] pcalg_2.4-3        roxygen2_5.0.1       epiNEM_0.99.0
## [25] knitr_1.15.1       igraph_1.0.1         gtools_3.5.0
## [28] e1071_1.6-7        BoolNet_2.1.3
##
## loaded via a namespace (and not attached):
## [1] Rcpp_0.12.8        bdsmatrix_1.3-2      corpcor_1.6.8
## [4] png_0.1-7          class_7.3-14         assertthat_0.1
```

## [7] rprojroot_1.1	digest_0.6.10	gmp_0.5-12
## [10] plyr_1.8.4	chron_2.3-47	backports_1.0.4
## [13] stats4_3.3.2	RSQLite_1.0.0	evaluate_0.10
## [16] sqldf_0.4-10	BiocInstaller_1.24.0	gplots_3.0.1
## [19] lazyeval_0.2.0	gdata_2.17.0	rmarkdown_1.3
## [22] gsubfn_0.6-6	proto_1.0.0	statmod_1.4.26
## [25] stringr_1.1.0	munsell_0.4.3	htmltools_0.3.5
## [28] tcltk_3.3.2	tibble_1.2	withr_1.0.2
## [31] ggm_2.3	gtable_0.2.0	DBI_0.5-1
## [34] magrittr_1.5	scales_0.4.1	KernSmooth_2.23-15
## [37] stringi_1.1.2	limma_3.30.4	robustbase_0.92-6
## [40] boot_1.3-18	fastICA_1.2-0	tools_3.3.2
## [43] DEoptimR_1.0-6	sfsmisc_1.1-0	abind_1.4-5
## [46] plotrix_3.6-3	yaml_2.1.14	clue_0.3-51
## [49] colorspace_1.3-0	cluster_2.0.5	caTools_1.17.1
## [52] memoise_1.0.0		

Reference:

Martin Pirkel, Madeline Diekmann, Marlies van der Wees, Niko Beerenwinkel, Holger Fröhlich, Florian Markowetz. Inferring Modulators of Genetic Interactions with Epistatic Nested Effects Models. submitted.