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Ozone decontamination of medical and non-medical devices: an assessment of design and implementation considerations

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Abstract

The control of infectious diseases can be improved via carefully designed decontamination equipment and systems. Research interest in ozone (a powerful antimicrobial agent), has significantly increased over the past decade. The COVID-19 pandemic has also instigated the development of new ozone-based technologies for the decontamination of personal protective equipment, surfaces, materials and indoor environments. As this interest continues to grow, it is necessary to consider key factors affecting the applicability of lab-based findings to large-scale systems utilizing ozone. In this review, we present the recent developments pertaining to the key factors affecting the successful deployments of industrial ozone technologies. Some of these include the medium of application (air or water), material compatibility, efficient circulation and extraction, measurement and control, automation, scalability, and process economics. We also provide a comparative assessment of ozone relative to other decontamination methods/sterilisation technologies and further substantiate the necessity for increased developments in gaseous and aqueous ozonation. Modelling methodologies, which can be applied for the design and implementation of ozone contacting systems are also presented in this review. Key knowledge gaps and open research problems/opportunities are extensively covered, including our recommendations for the development of novel solutions with industrial importance.

Keywords

Ozone, Design, Safety, Material compatibility, Simulation, Decontamination

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1. Introduction

The critical role played by contaminated fomites in the spread of diseases over a myriad of environments (hospitals, offices, laboratories and schools) necessitates the application of surface disinfection methods for the prevention of infections [1,2]. The wide variation in the scale of the disinfection procedure (from small surfaces to huge environments) has also led to several engineering developments for the efficient application of numerous chemical disinfectants [3]. One of such disinfectants is ozone (a potent antimicrobial agent), which has received tremendous research interest for decontamination, since the advent of the COVID-19 pandemic [4–6]. Its excellent oxidation potential (2.07 V) and rapid decomposition into oxygen make it particularly attractive and versatile for not only decontamination but also bleaching and deodourisation in both air and water [7,8]. This multifunctional attribute also makes it widely applicable in different industries including, pulp and paper, textile processing, aquaculture, drinking water treatment, animal husbandry, wastewater treatment, food, healthcare (therapeutic applications) and medical equipment processing [9–18]. However, its highly unstable property implies it cannot be stored for subsequent applications and must be generated on-site for immediate use. A key area, requiring the application of ozone is that of medical device sterilization.

Currently, ethylene oxide is the most commonly used sterilization method in the US medical sector, accounting for ~50% of all devices requiring sterilization [19]. However, ethylene oxide is carcinogenic and there are concerns regarding the release of unsafe levels of ethylene oxide into the environment. These issues have led to several initiatives and innovation challenges by the US Food and Drug Administration (FDA), which are geared toward the development of novel sterilization methods, for the replacement of ethylene oxide [20]. Ozone has considerable potential to replace this disinfectant. Although the antimicrobial properties of ozone have been known for decades, it has mainly been utilised for the removal or degradation of pollutants in waste and drinking water. Its application for the decontamination of medical and non-medical devices is relatively new and has not been adequately explored. Furthermore, the cost of ozone production has dropped significantly over the last 2 decades, and this has paved the way for new lab-scale and industrial developments [21]. Nonetheless, a majority of these developments have been at a small scale, and there are several knowledge gaps surrounding the implementation of ozone decontamination systems at a large scale.

To the best of the authors' knowledge many reviews on the application of ozone are application specific – focusing on water/wastewater treatment [22,23], aquaculture [24], food decontamination [11,25–28], textile processing [29], medical device sterilization [30,31], pulp and paper processing

[10], biomass processing [32], ozone therapy [33] and SARS-CoV-2 (COVID-19) inactivation [8,34]. In this review, we present and discuss the current state of the art on the implementation of ozone decontamination systems, and draw applicable inferences from lab-scale studies that aid the industrial design and installation of ozonation systems. Firstly, the uniqueness of ozone, and why it is becoming the preferred decontamination method (for surface sterilization), in comparison to others is presented; thereafter, engineering considerations regarding material compatibility, ozone generation and decomposition, circulation and extraction, measurement and control, scalability and flexibility of operation (in gaseous and aqueous forms), automation, health and safety and the economics of ozonation processes are discussed. We also highlight key mathematical models that can aid the sizing of ozone contact equipment, and provide our recommendations on the way forward/future directions from both academic research and industrial application perspectives. In comparison to other reviews in the field of ozonation, this is the first review to simultaneously consider these key aspects of ozone systems design and implementation. It is worth-mentioning that in this review we pay particular attention to gaseous ozone application while providing some insights into its aqueous application. It is hoped that this review will appeal to a broad range of scientific communities for the continued development of ozone-related technologies.

2. How does ozone compare to other sterilisation methods

Besides ozone, several studies have demonstrated the effects of a variety of sterilisation methods, including cold plasma, gamma irradiation, ultraviolet irradiation (of type C), dry and moist heat, steam, hydrogen peroxide (gas and liquid) microwave, peracetic acid, ethanol, glutaraldehyde, orthophthalaldehyde (OPA), ethylene oxide, benzalkonium chloride and hypochlorite. The performance of these methods for diverse applications has mainly been assessed using factors such as decontamination efficacy, cycle time, penetration capability, substrate/material compatibility, operational safety, cost of implementation and environmental sustainability, with an overwhelming majority focusing on the decontamination efficacy. It should be highlighted that the cycle time in this paper represents the sum of the generation time (or time required to attain the desired concentration in the chamber), the contact time at the desired concentration and the decomposition time to concentrations below the safety limit.

Table 1, presents a summary of some of the reported merits and demerits of these sterilisation methods. Methods that rely on full immersion of the object/device/material to be disinfected (e.g. glutaraldehyde), tend to be less preferred due to the post-disinfection steps that are required before they can be reused. Thus, the applicability of a disinfectant in the gaseous form or via misting (e.g. ozone, hydrogen peroxide or peracetic acid [35–37]) is a favourable attribute that reduces the need

for long drying procedures after the main disinfection phase. In addition to ozone, supercritical CO_2 and non-thermal plasma are emerging methods that require further development. Other methods which are sparingly utilised in isolation, but not captured in Table 1, include high hydrostatic pressure, pulsed light, ultrasound and electrolysed water [38]; however, these have immense antimicrobial properties, particularly during hybrid application with other well-known/established methods. Fig. 1 demonstrates key differences in the inactivation mechanisms of selected disinfection methods, during contact with a bacterial cell.
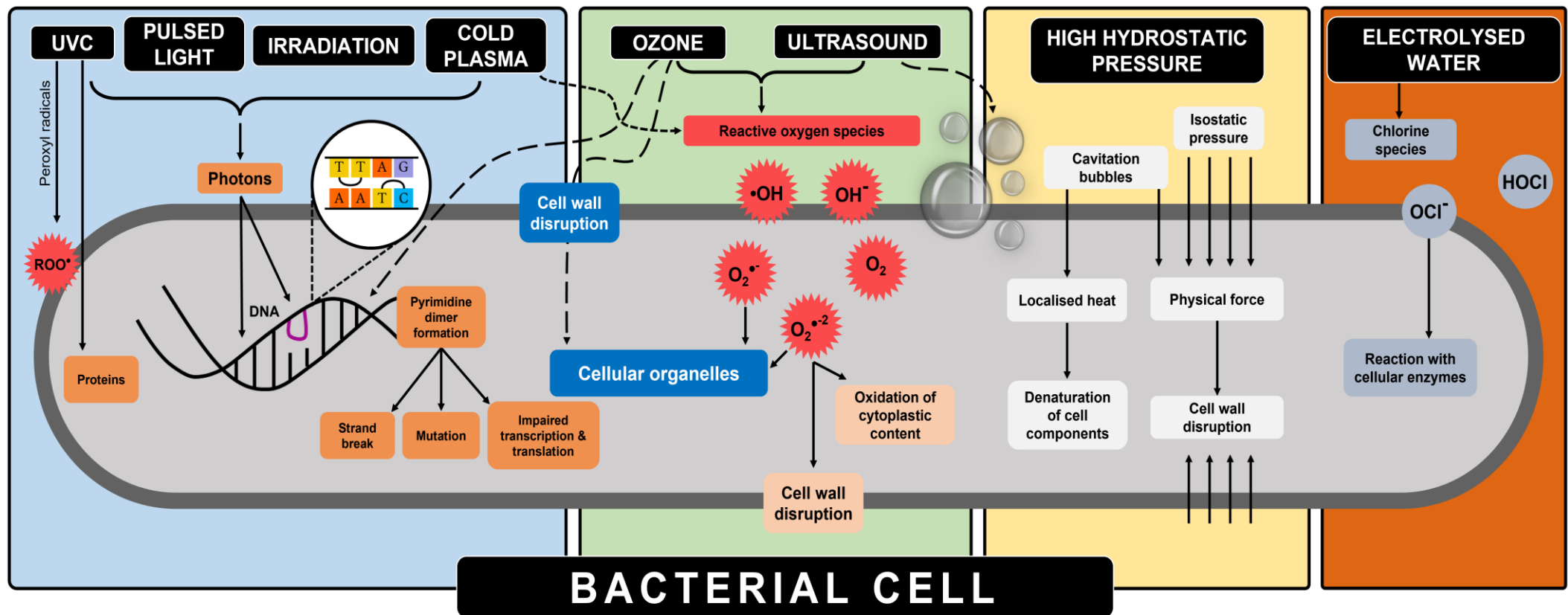
Figure 1: Comparative representation of the inactivation mechanisms of selected disinfection methods (adapted from [38,39]).

The direct attack on the DNA, cell wall disruption and subsequent oxidation, diffusion and reaction with key cell constituents and the creation of localised heat are key attributes of these methods. Ethylene oxide (EtO) is one of the most commonly applied methods for the terminal sterilization of medical devices, and this is due to their efficient microbial inactivation, excellent compatibility with a broad range of materials and the reasonable cost of implementation. Heat-based sterilization methods are usually not compatible with several polymeric components of medical devices that cannot withstand high temperatures. Unlike ozone which has to be generated on demand, EtO is usually pressurized and stored in liquid form for use in sterilization plants. Its inactivation mechanism is the alkylation of the amine groups of microbial DNA [40]. For efficient EtO application, a validated combination of humidity, gas concentration, temperature and exposure duration must be utilised [41]. Vacuum cycles are typically employed to increase the penetration of the gas into the substrate to be decontaminated. After treatment (which could last from 6 hrs to several days) [42], EtO concentrations can be brought below the permissible limits via vacuum purging and aeration. While there are several similarities between EtO's application and that of gaseous ozone (particularly the necessity of a humid environment for efficient inactivation), EtO poses a severe health and safety risk (a human carcinogen), as documented in Table 1. Its replacement with more environmentally friendly methods such as ozone and hydrogen peroxide has been a key subject of growing recent interest in the last 5 years. However, it should be noted that long-term chronic exposure to ozone can cause lung damage, and asthma and could be fatal [43]. Similarly, the inhalation of hydrogen peroxide causes irritation to the lungs and shortness of breath. Higher and long-term exposures may lead to the buildup of fluids in the lungs (pulmonary edema), bronchitis and even mortality (resulting from oxygen embolism) [44]. Thus, adequate control systems are required in facilities where these alternative decontamination techniques are applied to prevent uncontrolled exposure, and ensure safe working conditions. Further details of the exposure limits of different gaseous disinfectants are presented in Section 3.7.

Furthermore, the compatibility of key disinfection methods with commonly applied polymers as shown in Table 2 is a key differentiator affecting the applicability in different industries. Table 2 also illustrates why ethylene oxide has been predominantly applied for the sterilisation of medical equipment, despite its toxicity. Relative to other methods, it has the best compatibility with a wide range of polymers. The high reactivity of ozone coupled with its powerful oxidizing capabilities, make it a suitable candidate for the rapid sterilization of medical devices. However, a typical drawback of EtO is the long cycle times required for sterilization. Besides these benefits, ozone's (gas) ability to decontaminate items without leaving residues, coupled with its spontaneous

decomposition to oxygen makes it more environmentally friendly compared to ethylene oxide. Furthermore, since ozone gas must be generated on demand, storage and transportation requirements are minimal; thus reducing operational costs, compared to EtO. Although not directly applicable to gaseous sterilization of medical devices with ozone, it is important to highlight that the application of aqueous ozone may induce bromate formation (a toxic disinfection by-product) in waters with a high Bromine content, during water treatment. Furthermore, the degradation of ozone-incompatible materials may produce other harmful compounds. Thus, it is necessary to ensure that the devices to be sterilized are composed of ozone-compatible materials. While a direct comparison of the microbial inactivation efficiencies of EtO and ozone is scarce in the literature, the best-acting sterilant will depend on a myriad of factors and the specific application. Some of these factors include the applied dosage (concentration × exposure time), temperature, humidity, type of microorganisms present and the material properties of the substrate (porous or non-porous) to be disinfected. A detailed list of these factors is provided by Epelle et al. [45].

As indicated in Table 2, the compatibility of some polymers with ozone is still unknown – an indication of the recency of its application in diverse industries. The most stable materials with excellent resistance to the disinfection methods mentioned in Table 2 include PEEK, PTFE, PVC and potentially polyimides. Nonetheless, this review table has highlighted that ozone-based disinfection methods are relatively compatible with several popularly-applied polymers. Furthermore, hydrogen peroxide also appears to perform better than ozone (Table 2) in terms of material compatibility. Thus, future applications of ozone-based decontamination may involve a hybrid process (O_3 + H_2O_2 + UV + other environmentally-friendly gaseous disinfectants).

Table 1: Some advantages and disadvantages of popularly applied disinfection methods.

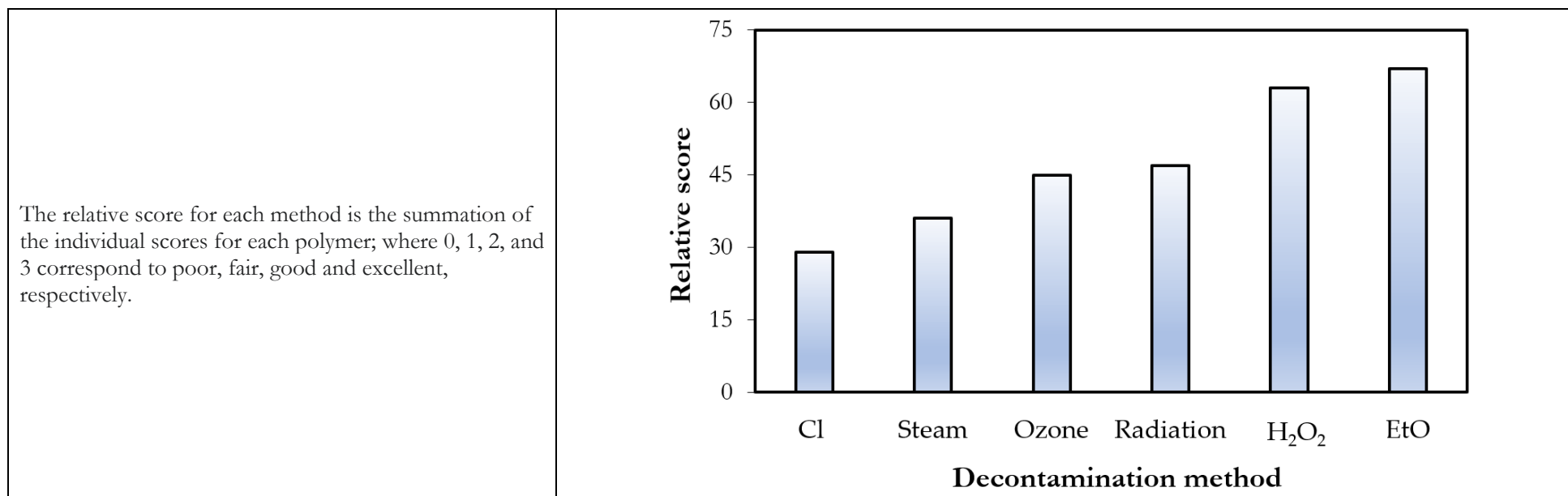| Disinfection method	Advantages	Disadvantages	References
Ozone	- Flexible application (low concentrations for longer durations, usually give the same effect as high concentrations for shorter times). - Environmentally friendly (decomposes to O_2). - Excellent for disinfecting heat-sensitive materials. - Can be readily applied as a gas, in aqueous form and via suspended mists. - Excellent penetration into hard-to-reach areas of an object. - Effective against a wide range of organisms (sporicidal & virucidal); no activation required.	- Relatively lower compatibility with some polymers, whereas the compatibility of some is still unknown (Table 2). - May produce bromates as a disinfection by-product, in the presence of Bromine (at high concentrations) during aqueous applications. - Large-scale generation costs can be high since it is difficult to store; may also pose a fire risk, particularly in oxygen-rich environments. - Inhalation causes shortness of breath, asthma, heaviness of the chest, dry throat, cough and headaches; long-term chronic exposure may be fatal.	[4,8,38,43,46,47]
UVC	- Particularly effective against different fungi. - Applicable in dry (air) and wet (water) conditions. - Effective against a wide range of organisms; sporicidal and virucidal.	- Poor penetration, particularly when materials of complex geometries are to be disinfected. - Effectiveness decreases as the distance from the UVC source increases. - High installation costs. - Induces colour change, and embrittlement of some plastics. - Exposure causes severe eye and skin irritation.	[5,48]
Chlorine-based methods	- Good microbiocidal activity - Inexpensive and commonly used in surface cleaning and bleaching agents.	- Non-sporicidal - Harmful disinfection by-products are formed. - Slower inactivation kinetics compared to ozone - Degradation of stainless steel and certain plastics	[6,49,50]
H_2O_2	- Better mater material compatibility compared to ozone. - Effective against a broad range of microorganisms; also sporicidal. - Readily applicable in gaseous and aqueous forms. - Excellent for heat-sensitive materials - No activation is required. - Non-staining	- Incompatible with some materials as shown in Table 2; metals such as silver, copper, brass and zinc may be affected. - An irritant to the eyes upon exposure. - Inhalation causes shortness of breath; long-term chronic exposure may be fatal. - Its instability implies an increased cost for large-scale continuous generation.	[36,46,51–53]

Ethylene oxide	• Excellent compatibility with a wide range of polymers (see Table 2). • Good penetration capabilities (particularly medical packaging). • Good antibacterial properties.	• Carcinogenic • Toxic, flammable and explosive • Longer cycle times are required. • High cost of operation • Higher pressures than atmospheric pressure are typically are required • Requires extensive aeration to remove excess EtO.	[48,54]
Steam	• Non-toxic • Good penetration into medical devices. • Easy to implement and control.	• Unsuitable for heat-sensitive materials. • Need for drying after treatment due to wetness of procedure. • May induce rust in equipment.	[55–57]
Surfactants – quaternary ammonium compounds	• Possesses good detergent properties • Does not cause irritation • Good surface disinfectant for non-critical items.	• Not sporicidal • May cause occupational asthma • May require full immersion for effective penetration.	[58]
Gamma radiation	• Excellent penetration • Effective for sterilising single-use medical materials • Rapid sterilizability	• May induce polymer weakening and further degradation. • Not effective for reusable materials. • High capital cost requirements	[57]
Alcohols	• Non-staining • Wide-ranged antibacterial activity • Inexpensive	• Non-sporicidal • Ineffective against some viruses. • Can be flammable.	[5,48,58]
Peracetic acid	• Effective against a wide range of organisms (also sporicidal). • Applicable for fogging purposes. • Activation not required • Good material compatibility	• Can cause severe irritation to the eyes and skin • Corrosive to metals • High operational costs • Medical instruments may require total immersion • Difficult sterile storage	[58–60]
Glutaraldehyde	• Relatively lower cost • Good material compatibility	• Activation required • Strong odour, which causes respiratory irritation • Slow bactericidal activity	[58,60]
Ortho-phthalaldehyde	• No activation required • Rapid disinfecting action • Insignificant odour compared to glutaraldehyde.	• Expensive • Slow sporicidal activity • Stains materials and surfaces	[59–61]

Supercritical CO_2	• Good penetration capability. • Effective for heat-sensitive materials. • Excellent antimicrobial properties.	• Expensive equipment for implementation. • Very sensitive to pressure and temperature conditions. • May require other chemical additives. • May compromise the functional properties of materials (e.g. the filtering and breathability properties of facemasks).	[62–64]
Cold atmospheric or non-thermal plasma	• Excellent penetration since it works at the atomic/molecular level. • Extensive application for a wide range of microorganisms (excellent fungicidal properties). • Fast-acting disinfection method. • Excellent therapeutic properties (for treatment of chronic wounds). • Environmentally friendly.	• High installation/investment costs. • Typical small volumes of generated plasma limit its application on a large scale (with many samples or samples with a large surface area). • Limited to batch-based application • There are concerns regarding its penetration efficiency. • Its adaptability to different scenarios is still an open research question. • Enumeration of the dosage may be difficult.	[65–74]
Metallic nanoparticles	• Can be used to functionalise or coat materials and surfaces, embedding them with antimicrobial properties. • Non-toxic • Effective at low concentrations.	• Intensive preparatory procedures. • Limited antimicrobial properties. • May require chemical additives to give desired antimicrobial properties.	[75–78]

A comparison of the exposure limits of the chemical disinfectants is presented in Table 5.

Table 2: Compatibility of different polymers with commonly applied disinfection methods [25,57,79–84].

Material	Ozone	H₂O₂	Ethylene oxide	Chlorine	Steam	UV & gamma radiation
Acetals	✓✓	✓✓✓	✓✓✓	✗	✓	✗
Acrylonitrile butadiene styrene	✓	✓✓✓	✓✓✓	✗	✗	✓✓
Butyl rubber	✗	✓✓	✓✓✓	✓	✓✓	✗
Cellulosics	✓	✗	✓✓✓	✓✓	✗	✓
Epoxies	✓✓	✓✓✓	✓✓	✓	✓✓	✓✓✓
Ethylene propylene diene monomer (EPDM)	✓	✓	✓✓✓	✓	✓✓	✓✓
Nitrile rubber	✗	✓	✓✓✓	✗	✓	✓✓
Phenolics	✓✓✓	✓✓	✓✓	✓✓	✓	✓✓✓
Polyacrylic rubber	✓✓	✓	✓	✗	✗	✓
Polyallomer	✓✓✓	✓✓✓	✓✓✓	✓✓✓	✓✓	✓
Polyamides (PA)	✓✓	✓✓	✓✓✓	✗	✓	✓
Polycarbonate (PC)	✓✓✓	✓✓✓	✓✓✓	✓	✓	✓✓
Polychloroprene	✗	✓✓✓	✓✓	✓	✓	✓✓
Polyesters (PES)	✓✓✓	✓✓✓	✓✓✓	✗	✓	✓
Polyethylene (PE)	✓✓✓	✓✓✓	✓✓✓	✓	✓	✓✓
Polyethylene terephthalate (PET)	??	??	✓✓✓	✗	✓✓✓	✓✓
Polyimides (PI)	??	✓✓✓	✓✓✓	✓✓✓	✓✓✓	✓✓✓
Polyketones, polyether ether ketone (PEEK)	✓✓✓	✓✓✓	✓✓✓	✓	✓✓✓	✓✓✓
Polymethyl methacrylate (PMMA)	✓✓	✓	✓✓	✓✓	✗	✓
Polypropylene (PP)	✓✓✓	✓✓✓	✓✓	✗	✓✓	✓
Polystyrene (PS)	✓	✓✓✓	✓	✓	✓	✓✓✓
Polysulfone (PSF)	✓✓	✓✓✓	✓✓✓	✗	✓✓✓	✓✓✓
Polytetrafluoroethylene (PTFE)	✓✓✓	✓✓✓	✓✓✓	✓✓✓	✓✓	✓
Polyurethane (PU)	✗	✓✓	✓	✗	✗	✓✓
Polyvinyl acetate (PVA)	??	✓✓✓	✗	✓✓✓	✗	✓✓
Polyvinyl chloride (PVC)	✓✓	✓✓✓	✓✓✓	✓✓✓	✓	✓✓
Silicone	✓✓✓	✓✓✓	✓✓✓	✗	✓✓	✓

The relative score for each method is the summation of the individual scores for each polymer; where 0, 1, 2, and 3 correspond to poor, fair, good and excellent, respectively.

Poor (✗); **Fair** (✔); **Good** (✔✔); **Excellent** (✔✔✔); **Unknown** (??). *Despite the superior compatibility of Ethylene oxide with most polymers compared to ozone, it is a carcinogen and thus poses a severe health and safety risk during its application. Gamma radiation is more cost-intensive compared to ozone; whereas UV radiation is not as effective when disinfecting objects with complex/narrow geometries due to poor penetration. Hydrogen peroxide also has a high compatibility index but is not as powerful as ozone (a lower oxidation potential – 1.80 V versus 2.07 V). Although UV and gamma radiation have been combined, the extent and type of damage caused by each radiation type will depend on the polymer type and dosage. The consulted studies collectively group their effects; however, newer studies that comparatively evaluate their effects on different polymers will be useful.*

Depending on its mechanical properties (toughness, ductility, hardness), the desired application, and cost, this material may be further utilised in diverse industries where frequent decontamination is paramount or required. Although mainly polymers are captured in Table 2, metals such as zinc and cast iron can be readily degraded by ozone; stainless steel (SAE 304 & 316) possess better stability against oxidation. The suitability of aluminium tends to depend on the application of the oxidising agent under wet or dry conditions; wet conditions can potentially cause degradation.

3. Engineering considerations for the deployment of ozone technologies

3.1 Generation and decomposition

The splitting of oxygen molecules in air to form ozone can be carried out via ultraviolet (UV) radiation (e.g. 185 nm low-pressure mercury lamps and 172 nm Xenon excimer vacuum UV lamps) or via a high voltage/energy electric field (at low or high frequencies), commonly referred to as corona discharge (CD). These procedures are energy-intensive and the ozone yield depends on the composition of the feed gas. Utilising high-purity oxygen (e.g. medical-grade oxygen) as the feed gas can produce 10-15 wt.% ozone concentration – which can be double to quadruple the ozone concentration, that can be produced by air [25,85]. The main disadvantages of mercury lamps are their low UV efficiency, the low absorption coefficient of oxygen and the simultaneous production of UVC (254 nm) which destroys ozone [86,87]. Although vacuum-ultraviolet (VUV) lamps principally emit the ozone-producing spectral line, VUV light sources are scarce and tend to have a pulsed operation profile; thus limiting the continuous production of ozone for disinfection [87]. These limitations, coupled with the higher electrical efficiency of corona discharge methods have made them more attractive. 1-16 wt.% of ozone can be produced by CD ozone generators, compared to 0.001-0.1 wt.% by UV methods; this corresponds to 10-1000 times lower ozone concentration than CD methods [88]. The specific energy consumption per gram of ozone produced from dry air is 0.515 kWh/g ozone for the UV method (185 nm); whereas it is 0.018 kWh/g ozone for the CD method [89,90]. Although CD is usually preferred in diverse industrial applications, it has to be fed with clean and dry air or pure oxygen, to prevent the formation of nitrogen oxides and corrosive compounds [91]. During the selection of ozone generators for disinfection applications, it is important to identify if the quoted production rate is based on an oxygen or air feed gas, as this could have massive impacts on the expected performance [4].

Dissolving ozone gas in water requires efficient mass transfer of the gas into the liquid phase, particularly because of the cost involvement of gaseous ozone production (which tends to be higher when high-purity oxygen is used). Venturi injection and bubble diffusion mechanisms have

been mainly applied for this purpose [25]. The former involves the use of a venturi equipped with multiple inlets maintained at a vacuum to facilitate the mixing of the gas and liquid phases via the created pressure difference upon liquid entry into the system. In the latter, pressurised ozone expands via nano/micro-sized pores on a porous stone into the liquid phase. Acoustic/ultrasound energy, high-intensity light photons in liquids (e.g. UV), and electrolysis are also other applied methods adopted to enhance ozone mass transfer [92]. It is worth mentioning that micro- and macro-bubbles may be generated during these procedures; however, it is the smaller-sized nanobubbles that are retained much longer in solution and facilitate ozone decontamination [93]. Larger-sized bubbles tend to be largely affected by buoyancy, leading to their collapse at the surface, and eventual ozone escape. Fig. 2a elucidates the increased stability attainable with ozone nanobubbles compared to ordinary bubbles of larger sizes. Furthermore, the increased dissolved oxygen (Fig. 2b) content facilitates the generation of radicals capable of oxidizing pollutants. Rice et al. [94] highlight some key methods of enhancing ozone mass transfer for laundry applications.
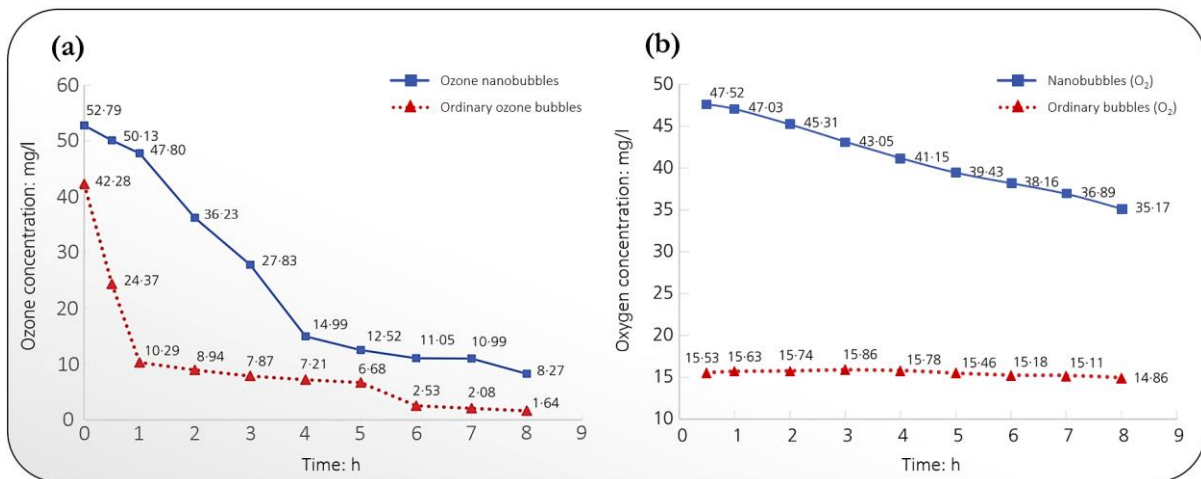

Figure 2: The effect of ozone nanobubble generation over time on (a) aqueous ozone stability and (b) oxygen stability (adapted with permission from [92]). Nanobubbles are generally more stable than ordinary bubbles in both scenarios.

Various studies have reported different half-lives of ozone in air and water. The presence of ozone-consuming compounds in the type of water analysed is a key determinant of the half-life. While ordinary tap water gave a half-life of 10 mins in the study of Epelle et al. [6], the half of bottled mineral water was 39 min. Where stable nanobubbles are efficiently produced in ozone-demand-free water, the half-life of ozone may increase even more significantly. Some reports have mentioned stability for weeks of ozone nanobubbles in solution [95]. The degree of agitation also affects the stability; while this may aid dissolution, continuous stirring must be maintained at an optimal rate, or else increased bubble coalescence may be induced, thus causing a concentration

decline via outgassing or ozone decomposition via OH⁻ radical generation during collapse at the surface. Thus, the ozone diffusion mechanism plays a significant role in its stability and ultimate decomposition kinetics. In air, ozone tends to remain longer than in ordinary tap water with conventional bubbling. Half-lives between 20 and 50 min have been typically reported by different authors [4,96,97]. The application of catalytic ozone decomposition is commonly practised in several industries where spontaneous ozone decomposition is insufficient to meet the peculiar timelines of the process; activated carbon and manganese oxide are two popularly implemented catalysts for this purpose [98]. Epelle et al. demonstrated that gaseous ozone decomposition over an activated carbon catalyst yields a 24 times faster decomposition rate compared to spontaneous decomposition [4]. The factors affecting the stability and the corresponding efficiency of microbial inactivation by gaseous or aqueous ozone are numerous. They can be broadly classified into ambient conditions or properties of the disinfection environment, the material or substrate properties, and the operational conditions. The ambient factors involve parameters such as the pH, temperature, pressure, humidity, and dissolved organics concentration; the material or substrate properties include porosity, contact angle, and contamination level; whereas the operational aspects consider the type of organisms, the ozone concentration, exposure duration, ozone generation method, penetrability, and the homogeneity of the ozone distribution in the test chamber/facility. While the impact of the operational factors on the inactivation efficiency can be directly inferred (enhancing the listed factors increases the inactivation efficiency), the influence of the ambient conditions is not as straightforward. Table 3, provides further details on the impact of the ambient conditions (temperature, pressure, relative humidity, pH, conductivity, and the presence of certain additives). The stability of ozone and its inactivation kinetics are determined by these parameters. These parameters can be altered in the disinfection system/environment to facilitate the breakdown of ozone into oxygen. As indicated in Table 3, several studies have also utilised increased temperatures to catalyse ozone decomposition. Decomposition mechanisms of aqueous and gaseous ozone (Table 4) are already well established and can be found in the following studies [23,98].

Table 3: Influence of key parameters on ozone decontamination and stability characteristics.

Parameter	Influence on ozone stability	Influence on microbial inactivation
Gaseous ozone application		
Temperature	Increased temperature enhances gaseous ozone instability/depletion; at lower temperatures, ozone is more stable [11,98–101].	Increased temperature increases gaseous ozone's reactivity, and inactivation efficiency [11,98,100].

Relative Humidity	Increased RH enhances gaseous ozone decomposition [11,102,103].	Increased RH improves the inactivation efficiency of gaseous ozone [11,102,103].
Pressure	At pressures lower than atmospheric pressure, the effective lifetime of ozone increases with pressure; at pressures above atmospheric pressure, the effective lifetime of ozone gas decreases with pressure [101].	Increased pressure (above atmospheric pressure) enhances gaseous ozone inactivation efficiency [104]. Also, the creation of a vacuum before ozone entry enhances ozone penetration into a porous substrate, thus accelerating antimicrobial action [105].
Aqueous ozone application		
Temperature	Increased temperature enhances aqueous ozone instability/depletion; at lower temperatures, ozone is more stable [98,100,106].	Increased temperature increases aqueous ozone's reactivity, and inactivation efficiency [98,100,106].
Pressure	Increased pressure favours aqueous ozone solubility and stability [104].	Better stability implies prolonged antimicrobial activity in water [93].
pH	Aqueous ozone is less stable at high pH [11,94,107].	Aqueous ozone's efficacy increases at high pH due to the production of OH- radicals [11,94,107].
Conductivity	Increasing the conductivity has a dual effect depending on the additive (e.g. structure-breaking or forming salts); there tends to be a threshold beyond which the positive or negative effects may be reversed [6].	Increasing the conductivity may enhance the inactivation efficiency of aqueous ozone (however there tends to be a threshold beyond which the effect is reversed) [6].
Bubble size	Nanobubbles stay in solution longer and promote ozone stability, compared to macrobubbles, whose collapse aid ozone decomposition [92,93,108].	Reduced bubble size enhances mass transfer and induces better inactivation by aqueous ozone [92,93,108]
Viscosity	Increasing the viscosity of the solution enhances ozone stability, delaying its decomposition [109,110].	The microbial inactivation efficiency of the solution is retained for a long time at high viscosities [109,110].
Additives (surfactants)	Controlled surfactant concentration may favour ozone stability [6,107]. However, this may significantly consume ozone in solution beyond a certain threshold, which could be the critical micelle concentration for some surfactants.	Inactivation is enhanced by the joint action of the surfactant [6].

3.2 Disinfection by-products

The unintended formation of persistent transformation products and disinfection by-products (DBPs) particularly during aqueous ozone applications is a key environmental challenge, especially during immersive/aqueous treatment of medical devices or during wastewater treatment. They can be formed as a result of the reaction of ozone with dissolved organics and some inorganic compounds present in the utilised water and can be difficult to eliminate [111]. The type of DBP

produced and its toxicity depends on the original composition of the water. For example, bromide-containing waters will produce bromate during ozonation, and this is a human carcinogen. Additionally, the myriad of DBPs that could be formed, with unknown specific toxicities and health implications [22,112], is a prevalent source of concern. Although the most common toxicological behaviour of DBPs is carcinogenicity, some of them could be neurotoxic, mutagenic, cytotoxic, teratogenic and possibly genotoxic, with several adverse outcomes on human health [113]. Although harmful DBPs are mostly described as a challenge of aqueous ozone application, the radicals generated during gaseous ozone generation and decomposition may also induce the formation of partially-oxidised DBP particularly under humid environments and in the presence of UV radiation and volatile organic compounds (VOCs) [114]; however, these tend to be short-lived. The production of nitrogen oxides (NO_x) – another by-product, during the generation of ozone from air (instead of pure oxygen) [115] can be problematic at high concentrations. Exposure to high levels of NO_x can damage the respiratory airways; the Occupational Safety and Health Administration (OSHA), has set a permissible exposure limit (PEL) of 25 ppm for nitric oxide and a short-term exposure limit of 1 ppm for nitrogen dioxide. Gas-phase nitrous acid (HONO), which may be formed under these conditions has been identified as an emerging pollutant [116]. This species is known to be a major photolytic source of hydroxyl radicals in air [117].

The application of hybrid methods (advanced oxidation processes) is a popularly applied route towards mitigating the formation of DBPs [118]. Thus, proper characterisation of water composition pre-treatment and rigorous kinetic studies are required (as part of the design phase of treatment facilities) to predict the potential formation of DBPs or harmful transformation products, their concentrations and removal methods before and after the application of ozone for treatment [119]. Additionally, appropriate control of the ozone dosage (based on the microbiological and environmental requirements) should be carried out to avoid unnecessarily high production rates of ozone gas. Multiple passes through effective decomposition catalysts, and constant concentration monitoring of potentially-harmful DBPs in the ozone treatment environment should be carried out.

3.3 Scalability and flexibility

Before large-scale implementations of ozone disinfection systems are implemented, lab-scale experimentation on the required dosage is inevitable. Upscaling lab-scale conclusions to large systems should be done with care, particularly when differences in material type for chamber construction and ozone demand in the immediate environment are expected. However, the study by Zoutman et al. [120] demonstrated the scalability of ozone disinfection systems (obtaining

similar inactivation results in a small test chamber – 0.25 m³ and in a large room – 82 m³). Furthermore, for certain applications where there is the inherent flexibility of choosing the ozone disinfection medium (gaseous or aqueous), Table 4 provides a list of factors to consider before this choice and the corresponding investments are made.

Table 4: A comparison of the practical considerations required for gaseous and aqueous ozonation (adapted from Epelle et al. [7]).

Factor	Ozonation in air	Ozonation in water
Need for drying	Substrate is usually dry, eliminating the need for further drying after treatment.	Drying after treatment is required since the substrate becomes wet, particularly if porous (e.g., textiles).
Cleaning	Does not clean the substrate; only disinfects or sterilizes it. A separate cleaning step is required and is best to carry out before ozonation.	Cleaning and disinfection may occur simultaneously. Furthermore, the use of surfactants can promote aqueous ozone stability [6,107].
Limitations to ozone generation	Higher ozone concentrations (e.g., up to 50 ppm) can be attained rapidly; this depends on the capacity of the generator.	Attainable ozone concentration is limited by mass transfer factors, relative to gaseous ozonation, for the same volume and generator capacity.
Concentration homogenization	Efficient gas circulation systems are required for concentration homogenization.	Concentration homogenization strongly depends on efficient gas dispersion in water; this may cause high gas usage.
Penetration efficiency	Hard-to-reach areas of the substrate can be better disinfected due to increased penetration of gaseous ozone.	Liquid penetration efficiency may be adversely affected for certain substrates with difficult geometries (e.g., small-diameter endoscopes).
Parameters influencing ozone stability	Humidity and temperature are the key influencing factors on the stability of ozone.	The stability of ozone during a treatment cycle is a function of many variables (pH, conductivity, temperature, pressure, water composition & ozone demand constraints).
Safety	Ozone's detrimental impact on human health, implies air-tight ozone chambers are required if the ozone equipment is to be located in an inhabited area.	Ozone's impact on human health is significantly reduced when it is dissolved in water [35].

It has been recently reported that gaseous ozonation (T = 18 °C and RH = 50%) can be more effective than aqueous ozonation (T = 18 °C), particularly when disinfecting wet porous substrates

at the same ozone concentration and exposure duration [7]. In their study, the decontamination of *S. aureus* gave the reverse observation of all organisms tested. A similar observation was also made by Martinelli et al. [121]. However, Megahed et al. [122] reported the superiority of aqueous ozone treatment over gaseous treatment of non-porous substrates contaminated with cattle manure. Similar observations are also reported by Tizaoui et al. against the SARS-CoV-2 virus [103]. In these studies, the microbes were mainly dried onto the surfaces of the non-absorbent materials utilised. This indicates that the nature of the substrate (porous or non-porous and wet or dry) and the type of microorganism utilised, affect the performance of gaseous and aqueous ozonation. The respective contributions of direct and indirect oxidation during gaseous (humid or dry) and aqueous are also attributable to these observations. Thus, it is important to establish the required dosage threshold for the specific application involved before applying each ozonation method.

3.4 Automation of ozone systems

The advent of the COVID-19 pandemic led to several developments of automated gaseous ozone disinfection systems, for a variety of materials. Typical ozone disinfection cycles comprise the ozone generation duration, the stabilisation duration or dwell time for ozone to act at the desired concentration and the decomposition during which ozone is converted to oxygen spontaneously or via a catalyst [5]. The goal of automating this process is to reduce the total cycle time attributable to the load, disinfect and unload process; furthermore, automation reduces the need for manual handling, which could cause further contamination post treatment. Recently, Rodriguez [123] developed an automatic disinfection device that comprises an object conveyor that transports items to and from a cabin that houses an ozone generator. They proposed the use of sash doors to completely isolate the internal section of the cabin to prevent ozone exposure. The cabin consists of 3 compartments, where the 2 end chambers are fitted with an ozone decomposition catalyst. Another invention by Silla et al. [124] for the disinfection of apparel using ozone gas also featured a conveyor, with a system of automatic doors and an optional heating system to aid ozone decomposition after the main treatment cycle. The invention by Miller [105], particularly for mail articles involves a similar 3-chamber plus conveyor arrangement (with an optional single chamber scenario) and is equipped with a vacuum pump to enable efficient diffusion of ozone gas into the contaminated articles.

A recently developed automated ozone system (patent filed) by ACS Clothing Ltd. for garment decontamination utilises the slower decomposition rate of ozone in air (relative to aqueous ozone) for continuous decontamination of clothing items and PPE. The items are fed by a conveyor into

a chamber with a pre-set ozone concentration. This concentration is maintained by a control system that regulates the ozone generators' outputs depending on the concentration set point (as determined by extensive experimentation). Thus, the cycle time is cut down significantly by eliminating the need for continuous ozone decomposition and regeneration from scratch. An interconnected system of conveyors, multiple chambers and air curtains is applied to prevent the escape of ozone from the system. By applying an ozone dose consisting of a high exposure concentration and shorter exposure duration, thousands of garments can be treated within an hour. From the experience of the authors, a logistics problem that has arisen from embedding automation is matching the high throughput of the disinfecting chamber with other workstations during garment processing. To resolve this, the cycle time in the disinfection chamber was increased, and the ozone concentration was lowered while maintaining the required dosage. This way, the potential for a bottleneck at the next station (usually the bagging machine) is effectively managed. As suggested by Farooq and Tizaoui [8], lower ozone concentrations for longer durations, are likely to give the same inactivation efficiency as high concentrations over shorter durations (so far the same dose is administered). This excellent flexibility provided by ozone, makes it very attractive, particularly for ensuring worker safety and solving logistical problems during industrial implementation.

There have also been several developments in the application of ozonated water sprays for the disinfection of different articles, employing varying degrees of automation. One of these developments is that by Maurya et al. [125], where an autonomous disinfection spray tunnel was applied to disinfect external surfaces at the height of the pandemic in India. Mascarenhas et al. [3] provide a detailed overview of the subject. However, conveyor systems, automatic doors and automatic ozone level control systems are all common features of these inventions. These advances and several others [35,126] will better position relevant authorities, and organisations to prevent or effectively combat the next pandemic.

For aqueous ozone application, it is also important to highlight the necessity of robust control systems for the ozone concentration, as this avoids ozone overdose/oversaturation, which is could cause loss of ozone gas to the environment above the air-water interface. Although the oxidation-reduction potential (ORP) has been frequently utilised [127] as the control parameter for this purpose, it is an indirect measure (which considers multiple oxidants, including ozone); thus making it difficult to ascertain the actual amount of ozone required to treat the immersed object [128]. Nonetheless, the use of the ORP may be attributed to the fact that the presence of other oxidizing agents may affect the microbial inactivation efficacy. Nonetheless, direct control of ozone concentration is mostly desirable.

3.5 Circulation and extraction

Given the high relative density of ozone gas (1.7), adequate circulation systems are required to ensure proper contact with the objects and surfaces of interest to be disinfected. This is particularly important, for objects of complex geometries, and for large-scale applications, where several items are to be disinfected. Computational fluid dynamics (CFD) becomes an important tool to apply
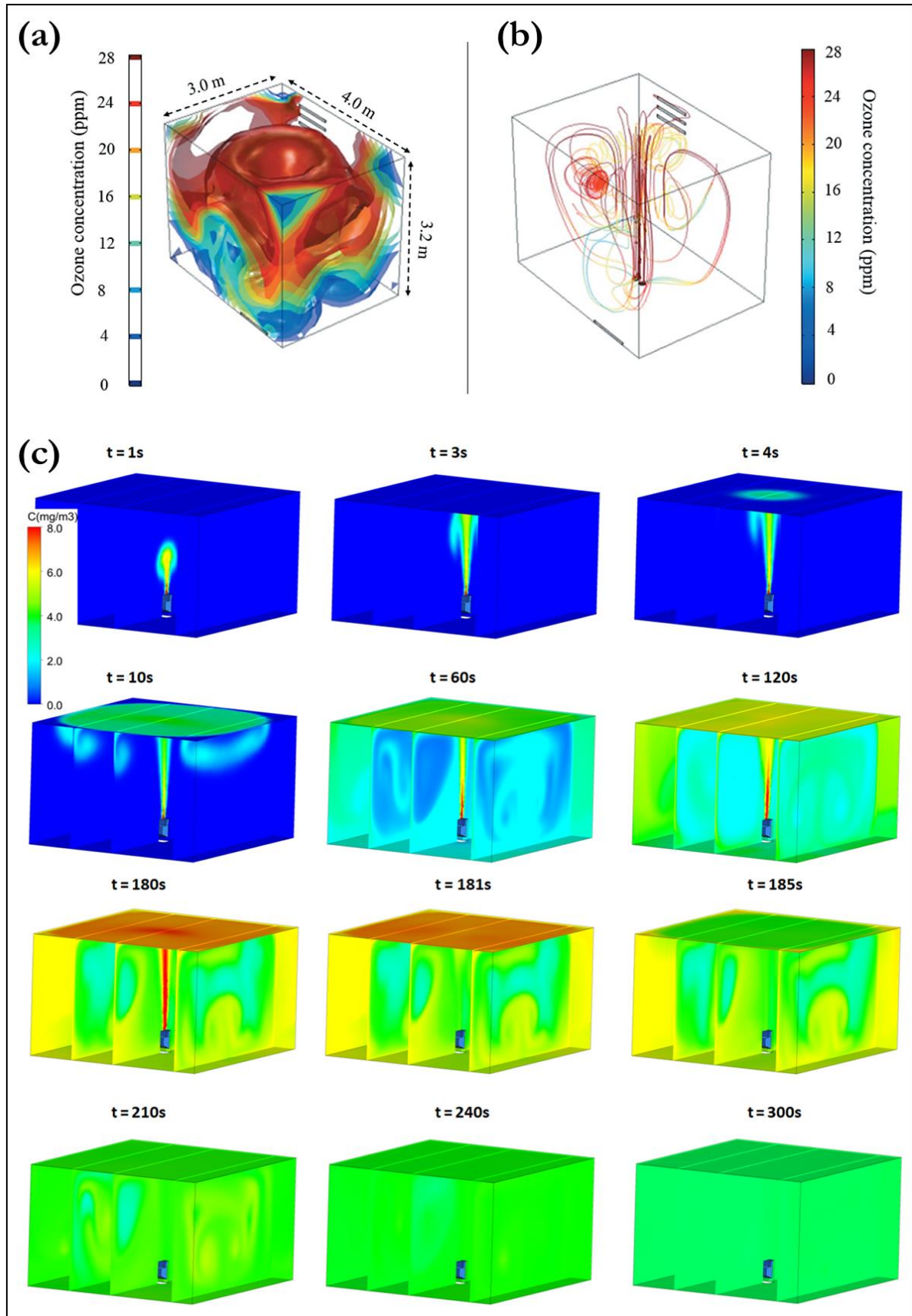
Figure 3: Application of computational fluid dynamics (CFD) to analyse the circulation of ozone gas in a room for the decontamination of surfaces and objects (a, b) show the contours and

streamlines of ozone concentration using COMSOL Multiphysics in the work of de Souza et al. [129]; (c) shows the time variation of ozone spread in a room via contour plots on different planes using ANSYS Fluent [130].

for this purpose. As demonstrated in Figs. 3a and 3b, de Souza et al. applied finite element CFD simulations using COMSOL Multiphysics® to examine ozone's spread in an office via an ozone generator [129]. The validated model enabled the authors to identify regions of the room with low concentrations, due to poor mixing. A similar study by Jarohmi et al. [130] shown in Fig. 3c, utilises the scalar transport model in Ansys Fluent (a finite volume solver) to model the dispersion of ozone gas from a generator in a room. Excellent agreements with experiments were observed in their work. The conditions shown in Fig. 3c represent a generation time of 3 min (at 7 g/h) followed by a 2 min dwell time. CFD studies of this kind provide insights into the number of generators required based on the desired concentration level desired, and the optimal locations of the generators and circulation systems to ensure adequate ozone interaction with the items to be decontaminated. CFD methods can also be applied to study ozone fogging or ozone misting operations, using Eulerian-Eulerian or Lagrangian-Eulerian multiphase flow models.
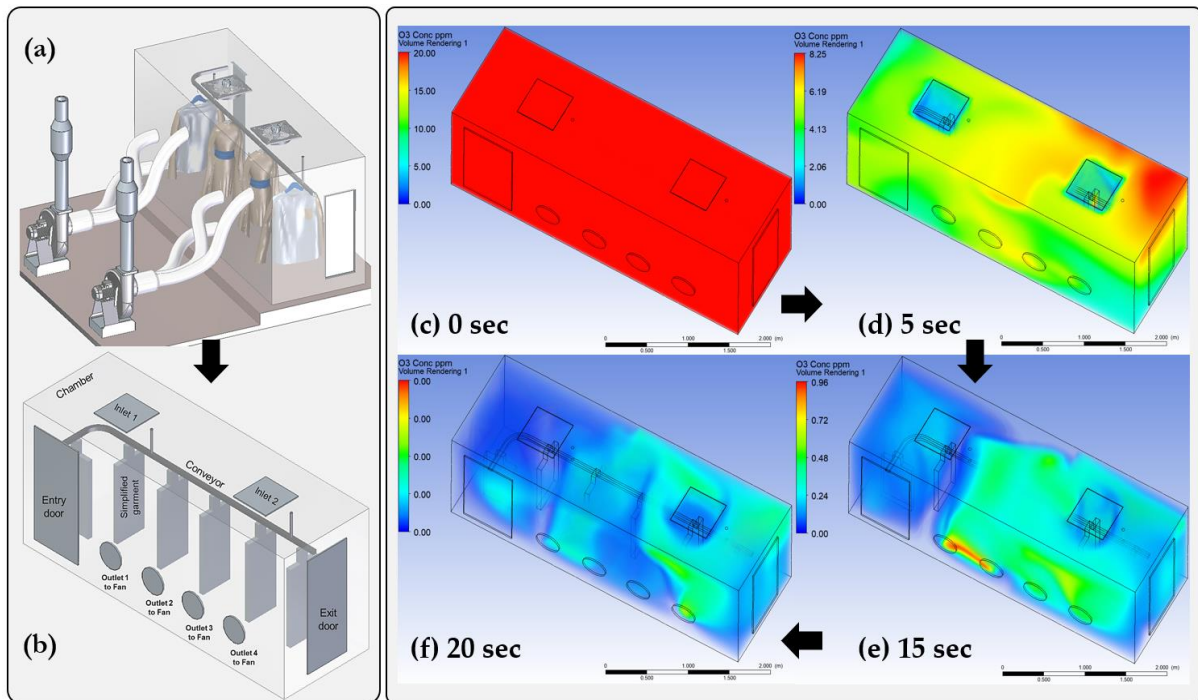

Figure 4: Application of computational fluid dynamics (CFD) to analyse the removal rate of ozone gas from a clothing disinfection chamber using extraction fans; (a) shows the chamber's configuration (b) a simplified 3D model (to reduce the number of meshing elements) used for the CFD simulation in ANSYS Fluent; (c-f) time variation of ozone removal from the system.

During emergency procedures, it may be required to rapidly extract ozone gas from the chamber, to ensure worker safety. As an example, Fig. 4 shows a downscaled simulation of an ozone chamber, which the authors have designed for industrial disinfection applications. The pressure drop through the catalytic destruct units at the fan outlets (Fig. 4a), the fan static pressure drop, the attainable air flow rate, duct size and the pressure at sealable inlets, are key factors affecting the efficiency of ozone removal rates from the system. Numerical simulations of an ozone misting process for the sanitisation of hospital facilities have been performed by Schroer et al. [131]. Their validated CFD model enabled an accurate distribution of the ozone mist concentration to be obtained. Besides the application of numerical CFD computations, other important analytical models, to bear in mind when designing ozone disinfection systems are shown in Table 6.

3.6 Measurement of ozone concentration

The measurement methods of ozone in gas and water are already well-established in the literature [25,29,91]. Fourier transform infrared spectroscopy (FTIR) is often used for gaseous ozone measurement; this method is however not suitable for aqueous ozone measurements [72]. UV absorption has also been extensively used and is adaptable to both gaseous and aqueous ozone measurements. Nonetheless, organic compounds in culture media may limit their applicability [72,132]. Pang et al. reported that most UV-based methods for gas-phase measurements require high amounts of power, and sampling gas flow rates; they also tend to be expensive [133]. Sensors based on gas-sensitive semiconducting oxide technologies are also commonly used [134]. Electrochemical ozone sensors can produce a voltage signature between an anode and cathode that correspond to the amount of ozone present [25]. They usually involve the use of a porous membrane that allows ozone to pass through into a cell containing electrodes and an electrolyte. The contact of ozone and the electrolyte causes a change in the electrochemical potential between the electrodes causing a flow of electrons. This method is robust and can be used for both gas and liquid phase measurements, but may be expensive and require frequent calibration [6].

Aqueous ozone can be readily measured via colourimetric methods such as the N,N-diethyl-p-phenylenediamine (Palin DPD) method, and the indigo method. The oxidation of the iodine ion in a DPD + KI buffered solution causes a pink colouration whose relative absorbance can be measured spectrophotometrically [29]. Similarly, ozone interacts with the carbon-double bonds of sulfonated indigo dye to decolourise it, and the change in absorbance gives an indication of the ozone concentration [91,135]. However, these methods tend to be affected by other oxidizing agents present in the solution (e.g. Cl^-, Br^- Mn^{+2}, OH^-) [29,72]. Palin DPD method implements a correction for the presence of Cl^-, Br^- and in solution using glycine tablets [136]. However, this

increases the difficulty of continuous data collection, which is often required for decomposition kinetics studies. Although these colourimetric methods tend to be cheaper than electrochemical methods, they are not as robust. More recently, Wright et al. [72] highlighted Pittsburgh Green fluorescence probes as being sensitive and specific to ozone. Although not commercially available, it has considerable potential to be widely applied if further developed. The interested reader is referred to the work of Korlu et al. which provides a good overview of aqueous measurement techniques not covered herein [29]. In summary, electrochemical methods, although costly, provide accurate and robust ozone concentration measurements. Whichever ozone-sensing method is adopted, extensive and reasonably-frequent calibration should be performed.

3.7 Material selection, health and safety

Given ozone's degradative properties, construction materials must be carefully selected during the design and installation of gaseous or aqueous ozone equipment. Cost-effective metals like aluminium may be affected by moist conditions during ozone treatment [137,138]. It is also important to pay close attention to the grading of steel materials to be used (stainless steel 304 and 316 are the most resistant to ozone degradation). As shown in Table 2, several popularly applied polymers have limited compatibility with ozone; thus critical consideration is required for the material selection phase of any design endeavour involving ozone. Routine checks are also important to ensure that installed safety systems (e.g. rubber seals) are not compromised due to the degradation of the materials as a result of ozone application. While we concentrate on the resistance of several polymers (which are often degraded via chain scission and the breakage of crosslinks) in Table 2, the ozone resistance of commonly utilised metals can be found in [138].

The extent of ozone degradation of highly unsaturated polymers tends to be higher than that of saturated polymers, according to a study by Giurginca et al. [139]. Using IR spectra and kinetic data they established that ethylene-propylene-diene elastomer is more susceptible to ozone attack as a result of the presence of double bonds in the macromolecules. A study [140] on the degradation of a high-temperature epoxy, showed an ozone oxidation depth of up to 120 μm. Exposure of the neat resin to 1% of ozone for 3 months at room temperature showed that crosslinking dominated in the first week, resulting in a slight stiffening of the polymer. However, as ageing continued, a chain scission mechanism became dominant, resulting in a reduction of the load to failure. This demonstrates the intricate relationship between ozone degradation and the chemistry of the polymer, and thus the increased necessity need for regular inspections since ozone's impact on certain polymers is not always a progressive deterioration.

With ozone disinfection systems (particularly gaseous ozonation), there is a need for continuous monitoring. It is quite often the case that the detection limit increases with the maximum measurable concentration of the sensor. Thus, large-scale ozone applications that utilise high concentrations of ozone, require sensors capable of detecting ozone concentrations within the occupational exposure limit values. In the UK, the short-term (usually 15 min) worker exposure level for ozone is 0.2 ppm (Table 5). Grignani et al. [141] present a comprehensive list of the exposure levels in different countries. Ozone causes severe irritation of the respiratory tract as well as lung damage; coughing and chest tightness are characteristics of uncontrolled exposure to ozone [100]. There has been some discussion throughout the COVID-19 pandemic regarding the use of low ozone concentrations in occupied spaces to reduce the risk of disease transmission. However, the intentional generation of ozone in occupied spaces for this purpose is not encouraged. Table 5 presents the exposure limits of ozone relative to other gaseous disinfectants.

Table 5: Exposure limits of some gaseous disinfectants.

Disinfectant	Exposure limits	Reference
Ozone	0.1 ppm (OSHA-PEL-TWA) 0.3 ppm (Cal/OSHA-PEL-STEL) 0.1 ppm (NIOSH-REL-C)	[142]
Glutaraldehyde (GTA)	0.05 ppm (ACGIH-TLV-TWA) 0.2 ppm (NIOSH-REL-C)	[143]
Peracetic acid	0.4 ppm (ACGIH-TLV-STEL)	[144]
Hydrogen peroxide	1 ppm (OSHA-PEL-TWA)	[145]
Ethylene oxide	1 ppm (Cal/OSHA-PEL-STEL)	[146]
Chlorine	1 ppm (OSHA-PEL-TWA) 1 ppm (OSHA-PEL-C) 0.5 ppm (NIOSH-REL-C)	[146,147]
Ortho-phthalaldehyde (OPA)	—	—

PEL: Permissible exposure limit; STEL: Short-term exposure limit (usually 15 min); TWA: Time-weighted average over an 8-hr shift; TLV: Threshold limit value; REL: Recommended exposure limit; C: Ceiling; OSHA: Occupational Safety and Health Administration; ACGIH: American Conference of Governmental Industrial Hygienists; NIOSH: National Institute for Occupational Safety & Health; OPA is generally considered safer that GTA; however, exposure limit data for this disinfectant is scarce; — : No data.

3.8 Economics of large-scale disinfection systems

The cost of a large-scale ozone disinfection system is dependent on the intended application (water treatment, textiles disinfection medical equipment sterilisation etc.), and the capacity of the disinfection facility. Furthermore, the desired ozone dose affects the runtime and capacity of the ozone generators, which in turn affects electricity consumption and operating costs; as previously highlighted, ozone generation is an energy-intensive process. Data provided by Champion Technology in 1998 suggests a capital expenditure – CAPEX > $250,000 for the treatment of 1 million gallons of wastewater per day (which had undergone pre-treatment), and an annual

operating expenditure (OPEX) > $18,000 [148]. The presented analysis concluded that the costs are site-specific and depend on the plant's effluent limitations. Rice et al. [149] provided a comparative assessment of capital and operating ozonation costs for drinking water treatment plants in Belgium, Switzerland, France, and the US. For a 687 ML/day plant in Belgium, a capital cost of $4,024,000 was stated; whereas the electrical cost was ¢3.01/kWh. Similar electrical costs were quoted for different regions. Remondino and Valdenassi [18] presented a case study of ozone's therapeutic application in animal husbandry (specifically, a pig farm). They reported a €90,000 cost for purchasing the ozone plant, whereas, between €5,000 – 6,500 is incurred per year for the plant's maintenance (€4,000 of which are related to electrical costs). ACS Clothing Ltd, a clothing rental and fulfilment company in Scotland UK, is on the verge of completing the installation of an automated (semi-continuous) gaseous ozone disinfection system for garments. The system, which is capable of disinfecting 20,000 garments within an 8-hr shift, involves an approximate investment of £270,000. Ozone generators, ozone sensors, pin and clip conveyors, disinfecting and housing chambers, air curtains, extraction fans, circulation fans, ducting, catalytic destruct beds, and the control system/software are the main cost components - the automatic conveyor systems constituting approximately 32% of the CAPEX. A thorough economic analysis of this system will be presented in a separate study.

Table 6: Relevant models and correlations to consider when designing aqueous and gaseous ozone decontamination systems.

No.	Description (LHS)	Model	Definition of terms (RHS) & Reference
1	O_3 dose ($\boldsymbol{D_{O_3}}$, ppm.min)	$D_{O_3} = C \times t$	$\boldsymbol{C:}$ O_3 concentration (ppm); $\boldsymbol{t:}$ exposure time (min) [8]
2	Microbial log reduction ($\boldsymbol{LR}$)	$LR = \log_{10}\left(\dfrac{N_0}{N}\right)$	$\boldsymbol{N_0:}$ number of viable organisms before treatment; $\boldsymbol{N:}$ number of viable organisms after treatment [103]
3	O_3 concentration conversion (ppm to mg/m³); $\boldsymbol{C_{ppm}}$ is O_3 concentration in ppm	$C_{ppm} = \dfrac{24.45\, C_{mg/m^3}}{M_{O_3}}$	$\boldsymbol{C_{mg/m^3}:}$ O_3 concentration (mg/m³); $\boldsymbol{M_{O_3}:}$ the molecular weight; $\boldsymbol{24.45:}$ volume of a mole of O_3 gas at 1 atm and 25 °C [150]
4	Time to run an O_3 generator to give a target ozone concentration ($\boldsymbol{t_{min}}$, min)	$t_{min} = 117.9 \times F \times \dfrac{C_{ppm} \times V_{m^3}}{R_{mg/hr}}$	$\boldsymbol{111.79:}$ conversion factor from mg/m³ to ppm and hours to minutes. $\boldsymbol{C_{ppm}:}$ O_3 concentration (ppm); $\boldsymbol{V_m{}^3:}$ chamber volume (m³); $\boldsymbol{R_{mg/hr}:}$ O_3 production rate from generator (mg/hr); $\boldsymbol{F:}$ multiplication factor that depends on set-up; for an airtight chamber, with 0 start-up time and nonreactive walls, $\boldsymbol{F} = 1$ [150]
5	Equilibrium rate of O_3 formation ($\boldsymbol{\dot{n}_{O_3}^{eq}}$, ppm/s), when using a low-pressure mercury lamp	$\dot{n}_{O_3}^{eq} = 2\left[1 - e^{\left(-\kappa_{185}^{air} L\right)}\right]\dot{n}_{185} - \left[1 - e^{\left(-\kappa_{254}^{air} \frac{\dot{n}_{O_3}^{eq}}{\dot{n}_{air}} L\right)}\right]\dot{n}_{254}$	$\boldsymbol{\kappa_{185}^{air}:}$ absorption coefficient of air at the wavelength of 185 nm; $\boldsymbol{\kappa_{254}^{air}:}$ absorption coefficient of air at the wavelength of 254 nm; $\boldsymbol{L:}$ distance of the lamp from the reactor wall (m); $\boldsymbol{\dot{n}_{air}:}$ air flow (m³/s); $\boldsymbol{\dot{n}_{185}:}$ lamp efficiency at 185 nm; $\boldsymbol{\dot{n}_{254}:}$ lamp efficiency at 254 nm [151]
6	Current efficiency ($\boldsymbol{\Phi_{O_3}^e}$, %) during O_3 production by electrolysis	$\Phi_{O_3}^e = \dfrac{5 \times F \times Q}{3 \times M_{O_3} \times I} \times 100\%$	$\boldsymbol{Q:}$ O_3 production rate (kg/h); $\boldsymbol{I:}$ current applied to the cell (A); $\boldsymbol{F:}$ Faraday's constant (9.6485×10^4 C/mol); $\boldsymbol{M_{O_3}:}$ Molecular weight of O_3 [152]
7	First-order decomposition kinetics of O_3 in air; $\boldsymbol{C_t}$ is O_3 concentration at any given time	$\ln C_t = \ln C_0 - kt$ $t_{1/2} = \dfrac{\ln 2}{k}$	$\boldsymbol{C_0:}$ initial O_3 concentration; $\boldsymbol{k:}$ decay rate; $\boldsymbol{t:}$ time; $\boldsymbol{t_{1/2}:}$ O_3 half-life in air [25]
8	Decomposition kinetics of O_3 in water	$C_t = (C_0 - D)e^{(-kt)}$	$\boldsymbol{C_0:}$ initial O_3 concentration; $\boldsymbol{k:}$ O_3 decay rate; $\boldsymbol{t:}$ time; $\boldsymbol{D:}$ instantaneous O_3 demand of water [25]
9	Modified Chick-Watson disinfection kinetics of dissolved O_3; $\boldsymbol{S_t}$ is the survival ratio	$\log(S_t) = -p\dfrac{(C_0 - D)^q}{qk}[1 - \exp(-qkt)]$	$\boldsymbol{C_0:}$ initial O_3 concentration; $\boldsymbol{D:}$ instantaneous O_3 demand; $\boldsymbol{t:}$ time; $\boldsymbol{k:}$ O_3 decay rate; $\boldsymbol{k, p, q:}$ rate parameters [25,153,154]
10	Pseudo-first-order virus inactivation rate constant by ozone ($\boldsymbol{k_V'}$)	$k_V' = \dfrac{2.303 \times LR}{D_{O_{3,gas}}}$	$\boldsymbol{LR:}$ $\log_{10}$ reduction, $\boldsymbol{D_{O_{3,gas}}:}$ O_3 gas dosage [8]

11	>90% virus inactivation kinetics via O_3 gas; where t_* is the inactivation time (min)	$t_* = -24.18 \ln(C) + 77.10$ (inactivation data of several viruses were used to develop this correlation)	**C:** O_3 concentration (ppm) [141]
12	SARS-CoV-2 virus inactivation (VI)	$VI = 1 - \frac{N}{N_0} = 1 - 2.09 \exp(-5.68 \, RH)$ --- SARS-CoV-2 valid for **CT** ~ 5.8 g.min/m³ and 13% < **RH** < 70%	**N_0:** control virus count per unit volume; **N:** count of the surviving virus per unit volume after ozone treatment; **CT:** ozone doze (g.min/m³); **RH:** relative humidity (%) [103]
13	Inactivation kinetics of *B. cereus* spores and *E. coli* by aqueous O_3; LR: log reduction	$LR = 0.3482t + 1.5365$ --- *B. cereus* $LR = 0.2413t + 4.7716$ --- *E. coli*	**t:** exposure time (min) [155]
14	Electrical energy per order (**EE/O**, KW/m³/order) – the energy required by O_3 to degrade a contaminant by 1 order of magnitude.	$EE/O = \dfrac{P \times t \times 1000}{V \times \log\left(\frac{C_i}{C_f}\right)}$	**P:** electric power (kW); **t:** treatment time (h); **V:** volume of treated wastewater (L); **Ci:** initial concentration of contaminant (mol/L); **Cf:** final concentration of contaminant (mol/L) [23,156]
15	Diffusion of ozone in water (**r**, m²/s)	$r = 2S(C' - C_0)\sqrt{\dfrac{D \times t}{\pi}}$	**S:** bubble surface area; **C':** O_3 concentration in the gas-liquid interface; **C_0:** initial concentration assumed to be 0; **D:** diffusivity (1.76×10^{-9} m²/s); **t:** time of liquid exposure [92]
16	Amount of dissolved O_3 in water (**$C_{O_3,liq}$**, mg/L; **$C^*_{O_3,liq}$**, mg/mL)	$C_{O_3,liq} = \dfrac{C_{O_3,gas} \times Q_{O_3,gas}}{Q_{WW}}$ OR $C^*_{O_3,liq} = \dfrac{C^*_{O_3,gas} \times t \times D}{60 \times V}$	**$C_{O_3,gas}$:** O_3 concentration in the feed gas; **$Q_{O_3,gas}$:** feed O_3 flow rate (L/min); **Q_{WW}:** wastewater flow rate (L/min); **$C^*_{O_3,gas}$:** O_3 concentration in the inlet gas (g/m³); **t:** duration of O_3 application (min); **D:** O_3 flow rate (L/hr); **V:** volume of solution (mL) [157,158]
17	2ⁿᵈ kinetics of pharmaceuticals removal; where **S_t** (mg/m³) is the concentration of the compound of interest	$S_t = \dfrac{S_0}{(1 + S_0 k_2 t)}$	**S_0:** Initial concentration of compound of interest (mg/m³); **k_2:** rate constant for second-order kinetics (mg/m³/h); **t:** time [159]

3.9 Mathematical models

Table 6 highlights some key equations and models to consider when determining the optimal generation capacity for a particular system. One key question that transcends several industrial applications involving the utilisation of gaseous ozone generation is the length of time to operate an ozone generator of a certain capacity to achieve a desired concentration level. Eq. 4 (Table 6) provides a guide towards developing a reasonable estimate of the operational time. Since ozone autodecomposes to oxygen, the knowledge of the ozone generator performance coupled with the half-life (Eq. 7 & 8) of ozone in the specific environment of interest, can facilitate the design of an optimal control strategy that maintains the ambient concentration at a set value. Furthermore, the table includes the inactivation kinetic correlations of different organisms, which allows a design team to make recommendations on the length of the treatment cycle for effective inactivation. The electrical energy requirements as well as the diffusion rate and attainable concentration during aqueous ozone application are also presented. It is important to mention that the models in Table 6 do not fully consider important design concepts such as complex gas-liquid mass transfer effects and ozone gas dispersion and penetration; the impacts of which may be adequately studied using computational fluid dynamics (CFD), as presented in Figs. 3 and 4.

4. Open research problems, opportunities & recommendations

In this section, we highlight some of the key gaps in the literature that warrant further development. Furthermore, we also draw inferences from our industrial experience of implementing industrial-scale aqueous and gaseous ozone disinfection systems in providing the following recommendations for future exploration.

Life-cycle assessments (LCA) and environmental impact assessments are required for large-scale ozone systems to quantify their benefits relative to other popularly applied decontamination methods. More research efforts are also required to demonstrate the scalability of different disinfection methods; this can be achieved by comparing the disinfection efficiency attainable within the lab-scale test chamber with that in a large room under the same dosage conditions.

Automated and rapid ozone disinfection systems require further development for throughput and safety enhancement in industrial operations. New advancements in the design of robust control systems for the accurate regulation of ozone concentration levels in air and water are required. Some studies rely on the ORP (mV), which is not a direct implication of the disinfection ability of ozone. Comparative studies that examine the additional compounds generated by industrial and domestic ozone generators will help enhance personnel safety. Furthermore, a systematic

quantification of the ozone generation efficiencies when using medicinal oxygen and ambient air should be elucidated. A direct comparison of the efficacies of EtO, hydrogen peroxide and ozone (at the same set of conditions) is also required to evaluate the inactivation kinetics and performance characteristics of these methods.

As highlighted in Table 2, the compatibility of some materials (e.g. PET and PVA) with ozone is yet to be determined. This demonstrates the relatively recent application of ozone for the sterilisation of devices made from these materials. A detailed ozone-compatibility assessment of these materials is necessary. The ozone-adsorbent capability of some polymers (e.g. polystyrene [160]), and their subsequent efficient release, enhances the biocidal properties of surfaces made from this material. Further exploration into other polymers or classes of materials (e.g. zeolites) with a similar potential is required as they hold great potential for the development of self-disinfecting surfaces. In addition, there appears to be some conflicting information regarding the resistance of aluminium to degradation over prolonged ozone exposure. More clarity is required in this regard, considering the prevalence and cost-effectiveness of this metal for constructing large-scale chambers.

As demonstrated by Epelle et al. [4], the ozone decomposition rate via activated carbon catalyst is approximately 24 times that of ozone's natural decomposition. Further improvements in catalytic ozone decomposition will help ensure cycle time reduction and operational safety in automated ozonation systems. More developments are also required on the stabilisation of ozone in water via viscosity enhancement of the solution. Although glycerol has been successfully applied, further studies utilising safe and environmentally friendly polymers are needed. These may be useful as hand sanitisers – a friendlier alternative to ethanol-based sanitisers. Alkyl polyglycosides for example can be investigated in this regard; the OH groups present in their structure may also enhance the biocidal action of ozone. For certain applications as in ozone therapy, the release rate of ozone is key to achieving the desired efficacy. The development of materials capable of controlled ozone release in aqueous and dry environments is essential.

Further analyses of ozone decontamination of textile materials are required, to demonstrate its effectiveness in accordance with established standards such as the European Standard BS EN16616 [161]. The benchmarking of ozone's performance against these standards (applying the recommended microorganisms, and required contamination levels), will further facilitate its large-scale adoption. The textile industry will also benefit from the application of ozone in the mist form (dry fogging), particularly where the textile materials are required to be dry, post-treatment, and

where material compatibility or additional cost concerns limit the thorough application of gaseous ozone.

5. Conclusions

This review summarises key engineering factors to consider in the design and implementation of ozone decontamination systems. While several successful lab-scale demonstrations of ozone's effectiveness against a myriad of microorganisms exist in the literature, details of large-scale deployments of ozone technology are lacking. Factors such as the medium of application (air or water) material compatibility, efficient circulation and extraction, measurement and control, automation, scalability, and process economics must be carefully considered in the design and implementation phases of industrial ozone decontamination systems. Nonetheless, we present some progress made by the authors on the application of automation technologies to ozone systems for the disinfection of clothing and PPE items. The compatibility of ozone with several polymeric materials as shown in Table 2 also appears to be a key determinant of its applicability, despite its widely acknowledged antimicrobial efficiency. The evaluation of concentration thresholds over a repeated number of cycles for a variety of popularly applied materials with limited ozone compatibility will go a long way towards mitigating potential degradation. The application of hybrid ozonation methods, particularly green methods, with no toxic residues (e.g. $UV + O_3$, $H_2O_2 + O_3$, Peracetic-acid $+ O_3$) also holds great potential for addressing these compatibility constraints, without compromising the disinfection efficacy and material compatibility. It is hoped that the insights presented in this study will support the transition from carcinogenic ethylene oxide to ozone-based methods, for the sterilisation of medical devices and other materials – a current and global innovation challenge.

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7. Declaration of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

8. Credit author statement

Emmanuel I. Epelle: Conceptualization, Methodology, Software, Data Curation, Writing Original Draft, Writing Review Draft

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9. Acronyms

ACGIH	American Conference of Governmental Industrial Hygienists
CAPEX	Capital expenditure
CD	Corona discharge
CFD	Computational fluid dynamics
CFU	Colony-forming unit
EtO	Ethylene oxide
LCA	Life cycle assessment
NIOSH	National Institute for Occupational Safety & Health
OPEX	Operating Expenditure
OSHA	Occupational Safety and Health Administration
PET	Polyethylene terephthalate
PPE	Personal protective equipment
STEL	Short-term exposure limit
TLV	Threshold limit value
TWA	Time-weighted average

10. References

1. Castaño, N.; Cordts, S.C.; Kurosu Jalil, M.; Zhang, K.S.; Koppaka, S.; Bick, A.D.; Paul, R.; Tang, S.K.Y. Fomite Transmission, Physicochemical Origin of Virus-Surface Interactions, and Disinfection Strategies for Enveloped Viruses with Applications to SARS-CoV-2. *ACS Omega* 2021, *6*.

2. Goodwin, L.; Hayward, T.; Krishan, P.; Nolan, G.; Nundy, M.; Ostrishko, K.; Attili, A.; Cárceles, S.B.; Epelle, E.I.; Gabl, R.; et al. Which factors influence the extent of indoor transmission of SARS-CoV-2? A rapid evidence review. *J. Glob. Health* 2021.

3. Mascarenhas, L.A.B.; Oliveira, F.O.; Da Silva, E.S.; Dos Santos, L.M.C.; Rodrigues, L.D.A.P.; Neves, P.R.F.; Santos, A.Á.B.; Moreira, G.A.F.; Lobato, G.M.; Nascimento, C.; et al. Technological advances in ozone and ozonized water spray disinfection devices. *Appl. Sci.* **2021**, *11*, doi:10.3390/app11073081.

4. Epelle, E.I.; Macfarlane, A.; Cusack, M.; Burns, A.; Amaeze, N.; Mackay, W.; Yaseen, M. The Impact of Gaseous Ozone Penetration on the Disinfection Efficiency of Textile Materials. *Ozone Sci. Eng.* **2022**.

5. Epelle, E.I.; Macfarlane, A.; Cusack, M.; Burns, A.; Thissera, B.; Mackay, W.; Rateb, M.E.; Yaseen, M. Bacterial and fungal disinfection via ozonation in air. *J. Microbiol. Methods* **2022**.

6. Epelle, E.I.; Macfarlane, A.; Cusack, M.; Burns, A.; Amaeze, N.; Richardson, K.; Mackay, W.; Rateb, M.E.; Yaseen, M. Stabilisation of Ozone in Water for Microbial Disinfection. *Environments* **2022**, 1–19.

7. Epelle, E.I.; Emmerson, A.; Nekrasova, M.; Macfarlane, A.; Cusack, M.; Burns, A.; Mackay, W.; Yaseen, M. Microbial Inactivation: Gaseous or Aqueous Ozonation? *Ind. Eng. Chem. Res.* **2022**, 1–11.

8. Farooq, S.; Tizaoui, C. A critical review on the inactivation of surface and airborne SARS-CoV-2 virus by ozone gas. *Crit. Rev. Environ. Sci. Technol.* 2022.

9. de Wilt, A.; van Gijn, K.; Verhoek, T.; Vergnes, A.; Hoek, M.; Rijnaarts, H.; Langenhoff, A. Enhanced pharmaceutical removal from water in a three step bio-ozone-bio process. *Water Res.* **2018**, *138*, doi:10.1016/j.watres.2018.03.028.

10. Hostachy, J.-C.; van Wyk, B.; Metais, A. The Use of Ozone in the Pulp and Paper Industry. *paperASia* **2014**, *30*, 22–28.

11. Varga, L.; Szigeti, J. Use of ozone in the dairy industry: A review. *Int. J. Dairy Technol.* 2016, *69*.

12. Ben Fraj, A.; Jaouachi, B. Effects of ozone treatment on denim garment properties. *Color. Technol.* **2021**, *137*, doi:10.1111/cote.12568.

13. Powar, A.S.; Perwuelz, A.; Behary, N.; Hoang, L.; Aussenac, T. Application of ozone treatment for the decolorization of the reactive-dyed fabrics in a pilot-scale process-optimization through response surface methodology. *Sustain.* **2020**, *12*, doi:10.3390/su12020471.

14. Manasfi, T. Ozonation in drinking water treatment: an overview of general and practical aspects, mechanisms, kinetics, and byproduct formation. In *Comprehensive Analytical Chemistry*; 2021; Vol. 92.

15. Pumkaew, M.; Taweephitakthai, T.; Satanwat, P.; Yocawibun, P.; Chumtong, P.; Pungrasmi, W.; Powtongsook, S. Use of ozone for Vibrio parahaemolyticus inactivation alongside nitrification biofilter treatment in shrimp-rearing recirculating aquaculture system. *J. Water Process Eng.* **2021**, *44*, doi:10.1016/j.jwpe.2021.102396.

16. Araimo, F.; Imperiale, C.; Tordiglione, P.; Ceccarelli, G.; Borrazzo, C.; Alessandri, F.; Santinelli, L.; Innocenti, G. Pietro; Pinacchio, C.; Mauro, V.; et al. Ozone as adjuvant support in the treatment of COVID-19: A preliminary report of probiozovid trial. *J. Med. Virol.* **2021**, *93*, doi:10.1002/jmv.26636.

17. Lopes, M.S.; Ferreira, J.R.F.; da Silva, K.B.; de Oliveira Bacelar Simplício, I.; de Lima, C.J.; Fernandes, A.B. Disinfection of corrugated tubing by ozone and ultrasound in

mechanically ventilated tracheostomized patients. *J. Hosp. Infect.* **2015**, *90*, doi:10.1016/j.jhin.2015.03.004.

18. Remondino, M.; Valdenassi, L. Different uses of ozone: Environmental and corporate sustainability. Literature review and case study. *Sustain.* **2018**, *10*, doi:10.3390/su10124783.

19. FDA FDA Continues Efforts to Support Innovation in Medical Device Sterilization Available online: https://www.fda.gov/news-events/press-announcements/fda-continues-efforts-support-innovation-medical-device-sterilization (accessed on Aug 12, 2022).

20. FDA FDA Innovation Challenge 1: Identify New Sterilization Methods and Technologies Available online: https://www.fda.gov/medical-devices/general-hospital-devices-and-supplies/fda-innovation-challenge-1-identify-new-sterilization-methods-and-technologies (accessed on May 21, 2022).

21. Da Silva, L.M.; Jardim, W.F. Trends and strategies of ozone application in environmental problems. *Quim. Nova* **2006**, *29*, doi:10.1590/s0100-40422006000200023.

22. Von Gunten, U. Oxidation Processes in Water Treatment: Are We on Track? *Environ. Sci. Technol.* 2018, *52*.

23. Joseph, C.G.; Farm, Y.Y.; Taufiq-Yap, Y.H.; Pang, C.K.; Nga, J.L.H.; Li Puma, G. Ozonation treatment processes for the remediation of detergent wastewater: A comprehensive review. *J. Environ. Chem. Eng.* 2021, *9*.

24. Powell, A.; Scolding, J.W.S. Direct application of ozone in aquaculture systems. *Rev. Aquac.* 2018, *10*.

25. Aslam, R.; Alam, M.S.; Saeed, P.A. Sanitization Potential of Ozone and Its Role in Postharvest Quality Management of Fruits and Vegetables. *Food Eng. Rev.* 2020, *12*.

26. Botondi, R.; Barone, M.; Grasso, C. A review into the effectiveness of ozone technology for improving the safety and preserving the quality of fresh-cut fruits and vegetables. *Foods* **2021**, *10*, 748.

27. Pandiselvam, R.; Kothakota, A. Recent Applications of Ozone in Agri-Food Industry. *Ozone Sci. Eng.* 2022, *44*, 1–2.

28. Premjit, Y.; Sruthi, N.U.; Pandiselvam, R.; Kothakota, A. Aqueous ozone: Chemistry, physiochemical properties, microbial inactivation, factors influencing antimicrobial effectiveness, and application in food. *Compr. Rev. Food Sci. Food Saf.* **2022**, *21*, doi:10.1111/1541-4337.12886.

29. Körlü, A. Use of Ozone in the Textile Industry. In *Textile Industry and Environment*; 2019.

30. Murphy, L. Ozone--the latest advance in sterilization of medical devices. *Can. Oper. room Nurs. journal.* 2006, *24*.

31. Sousa, C.S.; Torres, L.M.; Azevedo, M.P.F.; de Camargo, T.C.; Graziano, K.U.; Lacerda, R.A.; Turrini, R.N.T. Sterilization with ozone in health care: An integrative literature review. *Rev. da Esc. Enferm.* **2011**, *45*.

32. Travaini, R.; Martín-Juárez, J.; Lorenzo-Hernando, A.; Bolado-Rodríguez, S. Ozonolysis: An advantageous pretreatment for lignocellulosic biomass revisited. *Bioresour. Technol.* 2016, *199*.

33. Cattel, F.; Giordano, S.; Bertiond, C.; Lupia, T.; Corcione, S.; Scaldaferri, M.; Angelone,

L.; De Rosa, F.G. Ozone therapy in COVID-19: A narrative review. *Virus Res.* 2021, *291*.

34. Tizaoui, C. Ozone: A Potential Oxidant for COVID-19 Virus (SARS-CoV-2). *Ozone Sci. Eng.* **2020**, *42*, doi:10.1080/01919512.2020.1795614.

35. Oliveira, F.; Dos Santos, L.M.C.; da Silva, E.S.; de Alencar Pereira Rodrigues, L.; Freitas Neves, P.R.; Fernandes Moreira, G.A.; Lobato, G.M.; Nascimento, C.; Gerhardt, M.; Bandeira Santos, A.A. Disinfecting Efficacy of an Ozonated Water Spray Chamber: Scientific Evidence of the Total and Partial Biocidal Effect on Personal Protective Equipment and in Vitro Analysis of a Viral Experimental Model. *Ozone Sci. Eng.* **2022**, 1–19.

36. McEvoy, B.; Rowan, N.J. Terminal sterilization of medical devices using vaporized hydrogen peroxide: a review of current methods and emerging opportunities. *J. Appl. Microbiol.* 2019, *127*.

37. Kruszewska, E.; Grześ, H.; Czupryna, P.; Pancewicz, S.; Groth, M.; Wondim, M.; Moniuszko-Malinowska, A. Fogging With Peracetic Acid in Schools and Kindergartens. *Front. Public Heal.* **2021**, *9*, doi:10.3389/fpubh.2021.697917.

38. Bhilwadikar, T.; Pounraj, S.; Manivannan, S.; Rastogi, N.K.; Negi, P.S. Decontamination of Microorganisms and Pesticides from Fresh Fruits and Vegetables: A Comprehensive Review from Common Household Processes to Modern Techniques. *Compr. Rev. Food Sci. Food Saf.* 2019.

39. Epelle, E.I.; Macfarlane, A.; Cusack, M.; Burns, A.; Mackay, W.G.; Rateb, M.E.; Yaseen, M. Application of Ultraviolet-C Radiation and Gaseous Ozone for Microbial Inactivation on Different Materials. *ACS Omega* **2022**, 1–16, doi:https://doi.org/10.1021/acsomega.2c05264.

40. Ratner, B.D.; Hoffman, A.S.; Schoen, F.J.; Lemons, J.E. Biomaterials science: an introduction to materials in medicine. *San Diego, Calif.* **2004**, 162–164.

41. Instrumentation, A. for the A. of M. *Sterilization of health care products-Ethylene oxide-Part 1: Requirments for development, validation, and routine control of a sterilization process for medical devices*; ANSI/AAMI/ISO 11135-1: 2007, 1-47 2007 Arlington, VA Association for the …, 2007;

42. Lambert, B.J.; Mendelson, T.A.; Craven, M.D. Radiation and ethylene oxide terminal sterilization experiences with drug eluting stent products. *AAPS PharmSciTech* 2011, *12*.

43. Jerrett, M.; Burnett, R.T.; Pope III, C.A.; Ito, K.; Thurston, G.; Krewski, D.; Shi, Y.; Calle, E.; Thun, M. Long-term ozone exposure and mortality. *N. Engl. J. Med.* **2009**, *360*, 1085–1095.

44. Abdollahi, M.; Hosseini, A. Hydrogen peroxide. *Encycl. Toxicol.* **2014**, *3*, 967–970.

45. Epelle, E.I.; Yaseen, M.; Macfarlane, A.; Cusack, M.; Burns, A.; Rolland, L. Automation of Large-Scale Gaseous Ozonation: A Case Study of Textile and PPE Decontamination. *Sustainability* **2023**, *15*, 2216.

46. Centers for Disease Control and Prevention (CDC) *Guideline for disinfection and sterilization in healthcare facilities*; 2008;

47. Epelle, E.I.; Okoye, P.U.; Roddy, S.; Gunes, B.; Okolie, J.A. Advances in the Applications of Nanomaterials for Wastewater Treatment. *Environments* **2022**, *9*, 1–27.

48. Chidambaranathan, A.S.; Balasubramanium, M. Comprehensive Review and Comparison

of the Disinfection Techniques Currently Available in the Literature. *J. Prosthodont.* 2019, *28*.

49. Dosti, B.; Guzel-Seydim, Z.; Greene, A.K. Effectiveness of ozone, heat and chlorine for destroying common food spoilage bacteria in synthetic media and biofilms. *Int. J. Dairy Technol.* **2005**, *58*, doi:10.1111/j.1471-0307.2005.00176.x.

50. Rosenblum, J.; Ge, C.; Bohrerova, Z.; Yousef, A.; Lee, J. Ozonation as a clean technology for fresh produce industry and environment: Sanitizer efficiency and wastewater quality. *J. Appl. Microbiol.* **2012**, *113*, doi:10.1111/j.1365-2672.2012.05393.x.

51. Johnston, M.D.; Lawson, S.; Otter, J.A. Evaluation of hydrogen peroxide vapour as a method for the decontamination of surfaces contaminated with Clostridium botulinum spores. *J. Microbiol. Methods* **2005**, *60*, doi:10.1016/j.mimet.2004.10.021.

52. McDonnell, G. The Use of Hydrogen Peroxide for Disinfection and Sterilization Applications. In *PATAI'S Chemistry of Functional Groups*; 2014.

53. Fichet, G.; Antloga, K.; Comoy, E.; Deslys, J.P.; McDonnell, G. Prion inactivation using a new gaseous hydrogen peroxide sterilisation process. *J. Hosp. Infect.* **2007**, *67*, doi:10.1016/j.jhin.2007.08.020.

54. Wallace, C.A. New developments in disinfection and sterilization. *Am. J. Infect. Control* **2016**, *44*, doi:10.1016/j.ajic.2016.02.022.

55. van Straten, B.; Robertson, P.D.; Oussoren, H.; Pereira Espindola, S.; Ghanbari, E.; Dankelman, J.; Picken, S.; Horeman, T. Can sterilization of disposable face masks be an alternative for imported face masks? A nationwide field study including 19 sterilization departments and 471 imported brand types during COVID-19 shortages. *PLoS One* **2021**, *16*, doi:10.1371/journal.pone.0257468.

56. de Man, P.; van Straten, B.; van den Dobbelsteen, J.; van der Eijk, A.; Horeman, T.; Koeleman, H. Sterilization of disposable face masks by means of standardized dry and steam sterilization processes; an alternative in the fight against mask shortages due to COVID-19. *J. Hosp. Infect.* 2020, *105*.

57. Rogers, W.J. Sterilisation techniques for polymers. In *Sterilisation of Biomaterials and Medical Devices*; 2012.

58. Juwarkar, C.S. Cleaning and sterilisation of anaesthetic equipment. *Indian J. Anaesth.* 2013, *57*.

59. Rutala, W.A.; Weber, D.J. New disinfection and sterilization methods. In Proceedings of the Emerging Infectious Diseases; 2001; Vol. 7.

60. Rutala, W.A.; Weber, D.J. Disinfection, sterilization, and antisepsis: An overview. *Am. J. Infect. Control* 2019, *47*.

61. Rutala, W.A.; Weber, D.J. Guideline for disinfection and sterilization in healthcare facilities, 2008. **2008**.

62. Santos-Rosales, V.; López-Iglesias, C.; Sampedro-Viana, A.; Alvarez-Lorenzo, C.; Ghazanfari, S.; Magariños, B.; García-González, C.A. Supercritical CO2 sterilization: An effective treatment to reprocess FFP3 face masks and to reduce waste during COVID-19 pandemic. *Sci. Total Environ.* **2022**, *826*, doi:10.1016/j.scitotenv.2022.154089.

63. Soares, G.C.; Learmonth, D.A.; Vallejo, M.C.; Davila, S.P.; González, P.; Sousa, R.A.;

Oliveira, A.L. Supercritical CO2 technology: The next standard sterilization technique? *Mater. Sci. Eng. C* **2019**, *99*, 520–540.

64. Meyer, M.; Prade, I.; Leppchen-Fröhlich, K.; Felix, A.; Herdegen, V.; Haseneder, R.; Repke, J.U. Sterilisation of collagen materials using hydrogen peroxide doted supercritical carbon dioxide and its effects on the materials properties. *J. Supercrit. Fluids* **2015**, *102*, doi:10.1016/j.supflu.2015.04.006.

65. Li, Y.F.; Shimizu, T.; Zimmermann, J.L.; Morfill, G.E. Cold atmospheric plasma for surface disinfection. *Plasma Process. Polym.* **2012**, *9*, doi:10.1002/ppap.201100090.

66. Selcuk, M.; Oksuz, L.; Basaran, P. Decontamination of grains and legumes infected with Aspergillus spp. and Penicillum spp. by cold plasma treatment. *Bioresour. Technol.* **2008**, *99*, doi:10.1016/j.biortech.2007.09.076.

67. Cotter, J.J.; Maguire, P.; Soberon, F.; Daniels, S.; O'Gara, J.P.; Casey, E. Disinfection of meticillin-resistant Staphylococcus aureus and Staphylococcus epidermidis biofilms using a remote non-thermal gas plasma. *J. Hosp. Infect.* **2011**, *78*, doi:10.1016/j.jhin.2011.03.019.

68. Scholtz, V.; Pazlarova, J.; Souskova, H.; Khun, J.; Julak, J. Nonthermal plasma - A tool for decontamination and disinfection. *Biotechnol. Adv.* 2015, *33*.

69. Nosenko, T.; Shimizu, T.; Morfill, G.E. Designing plasmas for chronic wound disinfection. *New J. Phys.* **2009**, *11*, doi:10.1088/1367-2630/11/11/115013.

70. Satoh, K.; MacGregor, S.J.; Anderson, J.G.; Woolsey, G.A.; Fouracre, R.A. Pulsed-plasma disinfection of water containing Escherichia coli. *Japanese J. Appl. Physics, Part 1 Regul. Pap. Short Notes Rev. Pap.* **2007**, *46*, doi:10.1143/JJAP.46.1137.

71. Kitazaki, S.; Tanaka, A.; Hayashi, N. Sterilization of narrow tube inner surface using discharge plasma, ozone, and UV light irradiation. *Vacuum* **2014**, *110*, doi:10.1016/j.vacuum.2014.06.014.

72. Wright, A.; Fuster, J.; Shaw, A.; Bandulasena, H.; Buckley, B.R.; Iza, F. Quantification of the Ozone Dose Delivered into a Liquid by Indirect Plasma Treatments: Method and Calibration of the Pittsburgh Green Fluorescence Probe. *Plasma Chem. Plasma Process.* **2018**, *38*, doi:10.1007/s11090-018-9923-1.

73. Sakudo, A.; Yagyu, Y.; Onodera, T. Disinfection and sterilization using plasma technology: Fundamentals and future perspectives for biological applications. *Int. J. Mol. Sci.* 2019, *20*.

74. Šimončicová, J.; Kryštofová, S.; Medvecká, V.; Ďurišová, K.; Kaliňáková, B. Technical applications of plasma treatments: current state and perspectives. *Appl. Microbiol. Biotechnol.* 2019, *103*.

75. Karim, N.; Afroj, S.; Lloyd, K.; Oaten, L.C.; Andreeva, D. V.; Carr, C.; Farmery, A.D.; Kim, I.D.; Novoselov, K.S. Sustainable personal protective clothing for healthcare applications: A review. *ACS Nano* 2020, *14*.

76. Bansod, S.; Bawskar, M.; Rai, M. In vitro effect of biogenic silver nanoparticles on sterilisation of tobacco leaf explants and for higher yield of protoplasts. *IET Nanobiotechnology* **2015**, *9*, doi:10.1049/iet-nbt.2014.0031.

77. Sreeja, S.; Vidya Shetty, K. Microbial disinfection of water with endotoxin degradation by photocatalysis using Ag@TiO2 core shell nanoparticles. *Environ. Sci. Pollut. Res.* **2016**, *23*, doi:10.1007/s11356-016-6841-8.

78. Caratto, V.; Aliakbarian, B.; Casazza, A.A.; Setti, L.; Bernini, C.; Perego, P.; Ferretti, M. Inactivation of Escherichia coli on anatase and rutile nanoparticles using UV and fluorescent light. *Mater. Res. Bull.* **2013**, *48*, doi:10.1016/j.materresbull.2013.02.024.

79. IS MED Specialties Hydrogen Peroxide Material Compatibility Chart Available online: https://www.industrialspec.com/images/files/hydrogen-peroxide-material-compatibility-chart-from-ism.pdf.

80. Nordion, M.D.S. Gamma Compatible Materials. *Datasheet, Aug* **2007**.

81. Hong, S.; Kim, I.-C.; Tak, T.; Kwon, Y.-N. Interfacially synthesized chlorine-resistant polyimide thin film composite (TFC) reverse osmosis (RO) membranes. *Desalination* **2013**, *309*, 18–26.

82. Stolov, M.; Freger, V. Degradation of polyamide membranes exposed to chlorine: an impedance spectroscopy study. *Environ. Sci. Technol.* **2019**, *53*, 2618–2625.

83. Novotema Elastomer Types and Chemical Compatibility Available online: https://en.novotema.com/assets/uploads/brochures/N1021A_Novotema_Elastomer_Chemical_Compatibility_Brochure_EN.pdf (accessed on Jun 19, 2022).

84. Kendron Chemical resistance of plastics Available online: https://www.kendrion.com/fileadmin/user_upload/Downloads/Datasheets_Operating_instructions/Valves_Fluid_Control/Chemical-resistance-valve-technology-Kendrion-EN.pdf (accessed on Jun 19, 2022).

85. Summerfelt, S.T. Ozonation and UV irradiation - An introduction and examples of current applications. In Proceedings of the Aquacultural Engineering; 2003; Vol. 28.

86. Eliasson, B.; Kogelschatz, U. Ozone Generation with Narrow-Band UV Radiation. *Ozone Sci. Eng.* **1991**, *13*, doi:10.1080/01919519108552472.

87. Szeto, W.; Yam, W.C.; Huang, H.; Leung, D.Y.C. The efficacy of vacuum-ultraviolet light disinfection of some common environmental pathogens. *BMC Infect. Dis.* **2020**, *20*, doi:10.1186/s12879-020-4847-9.

88. Oxidation Technologies LLC Corona vs. UV Ozone Available online: https://www.oxidationtech.com/blog/corona-vs-uv-ozone/ (accessed on May 31, 2022).

89. Blazek, P.; Harrison, J. *Ozone for Point-of-Use, Point-of-Entry and Small Water System Water Treatment Application - A Reference Manual*; Water Quality Association, 2004;

90. Lykins, B.W.; Clark, R.M.; Goodrich, J.A. *Point-of-use/point-of-entry for drinking water treatment*; CRC Press, 2018; ISBN 1351075829.

91. Kim, J.G.; Yousef, A.E.; Dave, S. Application of ozone for enhancing the microbiological safety and quality of foods: A review. *J. Food Prot.* 1999, *62*.

92. Batagoda, J.H.; Hewage, S.D.A.; Meegoda, J.N. Nano-ozone bubbles for drinking water treatment. *J. Environ. Eng. Sci.* **2018**, *14*, doi:10.1680/jenes.18.00015.

93. Seridou, P.; Kalogerakis, N. Disinfection applications of ozone micro- And nanobubbles. *Environ. Sci. Nano* 2021, *8*.

94. Rice, R.G.; DeBrum, M.; Cardis, D.; Tapp, C. The ozone laundry handbook: A comprehensive guide for the proper application of ozone in the commercial laundry industry. *Ozone Sci. Eng.* **2009**, *31*, doi:10.1080/01919510903091318.

95.   Li, M.; Ma, X.; Eisener, J.; Pfeiffer, P.; Ohl, C.D.; Sun, C. How bulk nanobubbles are stable over a wide range of temperatures. *J. Colloid Interface Sci.* **2021**, *596*, doi:10.1016/j.jcis.2021.03.064.

96.   Hudson, J.B.; Sharma, M.; Vimalanathan, S. Development of a practical method for using ozone gas as a virus decontaminating agent. *Ozone Sci. Eng.* **2009**, *31*, 216–223.

97.   Irie, M.S.; Dietrich, L.; de Souza, G.L.; Soares, P.B.F.; Moura, C.C.G.; da Silva, G.R.; Paranhos, L.R. Ozone disinfection for viruses with applications in healthcare environments: a scoping review. *Braz. Oral Res.* **2022**, *36*, doi:10.1590/1807-3107BOR-2022.VOL36.0006.

98.   Batakliev, T.; Georgiev, V.; Anachkov, M.; Rakovsky, S.; Zaikov, G.E. Ozone decomposition. *Interdiscip. Toxicol.* 2014, *7*.

99.   Kitayama, J.; Kuzumoto, M. Theoretical and experimental study on ozone generation characteristics of an oxygen-fed ozone generator in silent discharge. *J. Phys. D. Appl. Phys.* **1997**, *30*, doi:10.1088/0022-3727/30/17/011.

100.   UK HSE Ozone: Health hazards and control measures: Guidance Note EH38 Available online: https://www.hse.gov.uk/pubns/eh38.pdf (accessed on May 30, 2022).

101.   Suzuki, S.; Rusinov, I.M.; Teranishi, K.; Shimomura, N.; Itoh, H. Re-evaluation of rate coefficients for ozone decomposition by oxygen in wide range of gas pressures (20–1000 Torr) and temperatures (293–423 K). *J. Phys. D. Appl. Phys.* **2018**, *51*, 305201.

102.   Mahfoudh, A.; Moisan, M.; Séguin, J.; Barbeau, J.; Kabouzi, Y.; Ḱroack, D. Inactivation of vegetative and sporulated bacteria by dry gaseous ozone. *Ozone Sci. Eng.* **2010**, *32*, doi:10.1080/01919511003791971.

103.   Tizaoui, C.; Stanton, R.; Statkute, E.; Rubina, A.; Lester-Card, E.; Lewis, A.; Holliman, P.; Worsley, D. Ozone for SARS-CoV-2 inactivation on surfaces and in liquid cell culture media. *J. Hazard. Mater.* **2022**, *428*, doi:10.1016/j.jhazmat.2022.128251.

104.   Bialka, K.L.; Demirci, A. Utilization of gaseous ozone for the decontamination of Escherichia coli O157:H7 and Salmonella on raspberries and strawberries. *J. Food Prot.* **2007**, *70*, doi:10.4315/0362-028X-70.5.1093.

105.   Miller, J.D. Apparatus for sterilizing mail and textile articles 2011, US20110283661A1.

106.   Epelle, E.I.; Macfarlane, A.; Cusack, M.; Burns, A.; Mackay, W.; Mostafa, R.; Yaseen, M. Application of Ultraviolet Radiation and Gaseous Ozone for Microbial Inactivation on Different Materials. *J. Environ. Chem. Eng.* **2022**, 1–15.

107.   Eriksson, M. Ozone chemistry in aqueous solution: ozone decomposition and stabilisation 2005.

108.   Meegoda, J.N.; Aluthgun Hewage, S.; Batagoda, J.H. Stability of nanobubbles. *Environ. Eng. Sci.* **2018**, *35*, 1216–1227.

109.   Fukui, T.; Masuno, K.; Makita, Y.; Fujiwara, S.; Shiota, G.; Imamura, Y.; Shiba, A.; Wang, P.-L. Antimicrobial effects of ozone gel against periodontal bacteria. *J. Hard Tissue Biol.* **2014**, *23*, 445–448.

110.   Takeda, Y.; Jamsransuren, D.; Makita, Y.; Kaneko, A.; Matsuda, S.; Ogawa, H.; Oh, H. Inactivation of SARS-CoV-2 by Ozonated Glycerol. *Food Environ. Virol.* **2021**, *13*, doi:10.1007/s12560-021-09485-x.

111. Alharbi, S.K.; Price, W.E.; Kang, J.; Fujioka, T.; Nghiem, L.D. Ozonation of carbamazepine, diclofenac, sulfamethoxazole and trimethoprim and formation of major oxidation products. *Desalin. Water Treat.* **2016**, *57*, 29340–29351.

112. Lim, S.; Shi, J.L.; von Gunten, U.; McCurry, D.L. Ozonation of organic compounds in water and wastewater: A critical review. *Water Res.* 2022, *213*.

113. Vizioli, B.C.; Hantao, L.W.; Montagner, C.C. Disinfection byproducts in emerging countries. In *Emerging Freshwater Pollutants*; Elsevier, 2022; pp. 241–266.

114. National Academies of Sciences and Medicine, E. Why Indoor Chemistry Matters. **2022**.

115. Touati, H.; Guerin, A.; Swesi, Y.; Dupeyrat, C.B.; Philippe, R.; Meille, V.; Clacens, J.-M. Unexpected role of NOx during catalytic ozone abatement at low temperature. *Catal. Commun.* **2021**, *148*, 106163.

116. Gligorovski, S. Nitrous acid (HONO): An emerging indoor pollutant. *J. Photochem. Photobiol. A Chem.* **2016**, *314*, 1–5.

117. Wang, W.; Ezell, M.J.; Lakey, P.S.J.; Aregahegn, K.Z.; Shiraiwa, M.; Finlayson-Pitts, B.J. Unexpected formation of oxygen-free products and nitrous acid from the ozonolysis of the neonicotinoid nitenpyram. *Proc. Natl. Acad. Sci.* **2020**, *117*, 11321–11327.

118. Rosenfeldt, E.J.; Linden, K.G.; Canonica, S.; Von Gunten, U. Comparison of the efficiency of OH radical formation during ozonation and the advanced oxidation processes O3/H2O2 and UV/H2O2. *Water Res.* **2006**, *40*, 3695–3704.

119. Epelle, E.I.; Macfarlane, A.; Cusack, M.; Burns, A.; Okolie, J.A.; Mackay, W.; Rateb, M.; Yaseen, M. Ozone application in different industries: a review of recent developments. *Chem. Eng. J.* **2023**, 140188.

120. Zoutman, D.; Shannon, M.; Mandel, A. Effectiveness of a novel ozone-based system for the rapid high-level disinfection of health care spaces and surfaces. *Am. J. Infect. Control* **2011**, *39*, doi:10.1016/j.ajic.2011.01.012.

121. Martinelli, M.; Giovannangeli, F.; Rotunno, S.; Trombetta, C.M.; Montomoli, E. Water and air ozone treatment as an alternative sanitizing technology. *J. Prev. Med. Hyg.* **2017**, *58*, doi:10.15167/2421-4248/jpmh2017.58.1.757.

122. Megahed, A.; Aldridge, B.; Lowe, J. The microbial killing capacity of aqueous and gaseous ozone on different surfaces contaminated with dairy cattle manure. *PLoS One* **2018**, *13*, doi:10.1371/journal.pone.0196555.

123. Rodriguez Lopez, F. Automatic disinfection device 2020.

124. Silla Vidal, E.; Albert Revert, V.; Fuster Fuster, V.; Jesus, G.C. Apparatus and method for sanitizing items with ozone gas (EP3900747A1) 2021, 1–65.

125. Maurya, D.; Gohil, M.K.; Sonawane, U.; Kumar, D.; Awasthi, A.; Prajapati, A.K.; Kishnani, K.; Srivastava, J.; Age, A.; Pol, R.; et al. Development of Autonomous Advanced Disinfection Tunnel to Tackle External Surface Disinfection of COVID-19 Virus in Public Places. *Trans. Indian Natl. Acad. Eng.* **2020**, *5*, doi:10.1007/s41403-020-00141-7.

126. Greene, C.; Schindler, B. Efficacy of Ultraviolet Light Coupled with Ozone as a Germicidal Against Eight Healthcare Associated Environmental Pathogens. *Infect. Prev. Strateg.* **2020**.

127. Summerfelt, S.T.; Sharrer, M.J.; Tsukuda, S.M.; Gearheart, M. Process requirements for achieving full-flow disinfection of recirculating water using ozonation and UV irradiation. *Aquac. Eng.* **2009**, *40*, doi:10.1016/j.aquaeng.2008.10.002.

128. Gonçalves, A.A.; Gagnon, G.A. Ozone application in recirculating aquaculture system: An overview. *Ozone Sci. Eng.* **2011**, *33*, doi:10.1080/01919512.2011.604595.

129. de Souza, H.M.; Savi, G.D.; Gomes, T.; Cardoso, W.A.; Cargnin, M.; Angioletto, E. Ozone Application in COVID-19 Triage Areas and Its Efficiency of Microbial Decontamination. *Ozone Sci. Eng.* **2021**, *43*, doi:10.1080/01919512.2021.1908880.

130. Tamaddon Jahromi, H.R.; Rolland, S.; Jones, J.; Coccarelli, A.; Sazonov, I.; Kershaw, C.; Tizaoui, C.; Holliman, P.; Worsley, D.; Thomas, H.; et al. Modelling ozone disinfection process for creating COVID-19 secure spaces. *Int. J. Numer. Methods Heat Fluid Flow* **2022**, *32*, doi:10.1108/HFF-12-2020-0797.

131. Anton Schroer, I.; da Silva, J.; Brochier, B.; da Silva, P.R.S.; da Silva, S.B.; Hansen, É. Study of ozone misting for sanitization of hospital facilities: A CFD approach. *Ozone Sci. Eng.* **2022**, 1–15.

132. Hofman-Caris, R.C.H.M.; Harmsen, D.J.H.; Beerendonk, E.F.; Knol, T.H.; Houtman, C.J.; Metz, D.H.; Wols, B.A. Prediction of advanced oxidation performance in various pilot UV/H2O2 reactor systems with MP- and LP- and DBD-UV lamps. *Chem. Eng. J.* **2012**, *210*, doi:10.1016/j.cej.2012.09.041.

133. Pang, X.; Shaw, M.D.; Lewis, A.C.; Carpenter, L.J.; Batchellier, T. Electrochemical ozone sensors: A miniaturised alternative for ozone measurements in laboratory experiments and air-quality monitoring. *Sensors Actuators, B Chem.* **2017**, *240*, doi:10.1016/j.snb.2016.09.020.

134. Bart, M.; Williams, D.E.; Ainslie, B.; McKendry, I.; Salmond, J.; Grange, S.K.; Alavi-Shoshtari, M.; Steyn, D.; Henshaw, G.S. High density ozone monitoring using gas sensitive semi-conductor sensors in the lower Fraser valley, British Columbia. *Environ. Sci. Technol.* **2014**, *48*, doi:10.1021/es404610t.

135. Bader, H.; Hoigné, J. Determination of ozone in water by the indigo method. *Water Res.* **1981**, *15*, doi:10.1016/0043-1354(81)90054-3.

136. Palintest Ozone Meter User Manual Available online: https://www.palintest.com/wp-content/uploads/2019/04/Ozone-Meter_User-Manual_Web.pdf (accessed on Jan 10, 2022).

137. Technologies, S.E. Ozone Material Compatibility Available online: https://spartanwatertreatment.com/ozone-material-compatibility-chart/.

138. Solutions, O. Ozone Compatible Materials Available online: https://ozonesolutions.com/blog/ozone-compatible-materials/ (accessed on May 31, 2022).

139. Giurginca, M.; Zaharescu, T.; Meghea, A. Degradation of ethylene-propylene elastomers in the presence of ozone. *Polym. Degrad. Stab.* **1995**, *50*, doi:10.1016/0141-3910(95)00108-X.

140. Middleton, J.; Burks, B.; Wells, T.; Setters, A.M.; Jasiuk, I.; Predecki, P.; Hoffman, J.; Kumosa, M. The effect of ozone on polymer degradation in Polymer Core Composite Conductors. *Polym. Degrad. Stab.* **2013**, *98*, doi:10.1016/j.polymdegradstab.2012.08.018.

141. Grignani, E.; Mansi, A.; Cabella, R.; Castellano, P.; Tirabasso, A.; Sisto, R.; Spagnoli, M.; Fabrizi, G.; Frigerio, F.; Tranfo, G. Safe and Effective Use of Ozone as Air and Surface Disinfectant in the Conjuncture of Covid-19. *Gases* **2020**, *1*, doi:10.3390/gases1010002.

142. OSHA Occupational Safety and Health Administration: Ozone Available online: https://www.osha.gov/chemicaldata/9 (accessed on Jun 17, 2022).

143. OSHA Occupational Safety and Health Administration: Glutaraldehyde Available online: https://www.osha.gov/etools/hospitals/hospital-wide-hazards/glutaraldehyde#:~:text=OSHA does not currently have,ppm (parts per million). (accessed on Jun 17, 2022).

144. OSHA Occupational Safety and Health Administration: Peracetic Acid Available online: https://www.osha.gov/chemicaldata/866 (accessed on Jun 17, 2022).

145. OSHA Occupational Safety and Health Administration: Hydrogen Peroxide Available online: https://www.osha.gov/chemicaldata/630 (accessed on Jun 17, 2022).

146. OSHA Permissible Exposure Limits – Annotated Tables Available online: https://www.osha.gov/annotated-pels/table-z-1#notes (accessed on Feb 5, 2023).

147. OSHA Occupational Safety and Health Administration: Chlorine Available online: https://www.osha.gov/chemicaldata/650 (accessed on Jun 17, 2022).

148. Environmental Protection Agency, U.S. Wastewater Technology Fact Sheet Ozone Disinfection. *United States Environ. Prot. Agnecy* **1999**.

149. Rice, R.G.; Robson, C.M.; Miller, G.W.; Hill, A.G. Uses of ozone in drinking water treatment. *J. / Am. Water Work. Assoc.* **1981**, *73*, doi:10.1002/j.1551-8833.1981.tb04637.x.

150. Dennis, R.; Cashion, A.; Emanuel, S.; Hubbard, D. Ozone Gas: Scientific Justification and Practical Guidelines for Improvised Disinfection using Consumer-Grade Ozone Generators and Plastic Storage Boxes. *J. Sci. Med.* **2020**, *2*, doi:10.37714/josam.v2i1.35.

151. Voronov, A. New generation of low pressure mercury lamps for producing ozone. *Ozone Sci. Eng.* **2008**, *30*, doi:10.1080/01919510802341012.

152. Okada, F.; Nay, K. Electrolysis for Ozone Water Production. In *Electrolysis*; 2012.

153. Cho, M.; Chung, H.; Yoon, J. Disinfection of water containing natural organic matter by using ozone-initiated radical reactions. *Appl. Environ. Microbiol.* **2003**, *69*, doi:10.1128/AEM.69.4.2284-2291.2003.

154. Kaymak, B. *Effects of initial microbial density on disinfection efficiency and explanatory mechanisms*; Drexel University, 2003; ISBN 0496428497.

155. Ding, W.; Jin, W.; Cao, S.; Zhou, X.; Wang, C.; Jiang, Q.; Huang, H.; Tu, R.; Han, S.F.; Wang, Q. Ozone disinfection of chlorine-resistant bacteria in drinking water. *Water Res.* **2019**, *160*, doi:10.1016/j.watres.2019.05.014.

156. Bilińska, L.; Blus, K.; Gmurek, M.; Ledakowicz, S. Coupling of electrocoagulation and ozone treatment for textile wastewater reuse. *Chem. Eng. J.* **2019**, *358*, doi:10.1016/j.cej.2018.10.093.

157. Pagès, M.; Kleiber, D.; Violleau, F. Ozonation of three different fungal conidia associated with apple disease: Importance of spore surface and membrane phospholipid oxidation. *Food Sci. Nutr.* **2020**, *8*, doi:10.1002/fsn3.1618.

158. Paucar, N.E.; Kim, I.; Tanaka, H.; Sato, C. Ozone treatment process for the removal of pharmaceuticals and personal care products in wastewater. *Ozone Sci. Eng.* **2019**, *41*, doi:10.1080/01919512.2018.1482456.

159. Szabová, P.; Hencelová, K.; Sameliaková, Z.; Marcová, T.; Staňová, A.V.; Grabicová, K.; Bodík, I. Ozonation: effective way for removal of pharmaceuticals from wastewater. *Monatshefte fur Chemie* **2020**, *151*, doi:10.1007/s00706-020-02600-x.

160. Levif, P.; Larocque, S.; Séguin, J.; Moisan, M.; Barbeau, J. Inactivation of Bacterial Spores on Polystyrene Substrates Pre-exposed to Dry Gaseous Ozone: Mechanisms and Limitations of the Process. *Ozone Sci. Eng.* 2021, *43*.

161. BS EN 16616:2015-10 *Chemical disinfectants and antiseptics - Chemical-thermal textile disinfection - Test method and requirements*; 2015;

Graphical Abstract