Species Diversity of Thylakoid Proteins for Eudicots and Moncots with Photosynthesis and Chemical Carcinogenesis NADPH

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The Mathematical Learning Space Research Portfolio

1 Abstract

Photosynthesis uses light energy to synthesize organic compounds from carbon dioxide and water with a light dependent part (light reaction) and the light independent part (dark reaction, carbon fixation). The light reaction takes place in thylakoid, a membrane-bound compartment inside chloroplasts and cyanobacteria. The light energy is used by photosystems I and II to generate proton motive force and reducing power (NADPH or NADH). The proton motive force is used by ATP synthase to generate ATP, as in the mitochondrial respiratory chain where the supplies of ATP and NAD(P)H are then used to fix carbon dioxide. A major function of the thylakoid membrane and its integral photosystems is the establishment of chemiosmotic potential. The carriers in the electron transport chain use some of the electron's energy to actively transport protons from the stroma to the lumen. During photosynthesis, the lumen becomes acidic, pH 4, compared to pH 8 in the stroma where the change is a 10,000 fold concentration gradient for protons across the thylakoid membrane.

2 Introduction

In the Chemical carcinogenesis - reactive oxygen species, cancer is a multistep process which involves initiation, promotion and progression with chemical carcinogens, (1) directly or after xenobiotic metabolism, (2) induce DNA damage and act in a 'genotoxic' manner and carcinogens can induce cancer via nongenotoxic mechanisms such as oxidative stress. Trace metals and organic xenobiotics are typical classes of environmental pollutants with prooxidant effects that generate reactive oxygen species (ROS) depending on their ability to lose electrons and catalyze Haber Weiss and Fenton reactions. ROS is generated from the induction of various cytochrome P450 isoenzymes by detoxification of chemical carcinogens. Increased ROS generation creates DNA damage with mutations and have an important role in the initiation and progression of multistage carcinogenesis. Besides DNA damage, ROS further induce multiple

intracellular signaling pathways, notably (1) NF-kappa B, (2) JNK/SAPK/p38, as well as (3) Erk/MAPK. These signaling routes can lead to transcriptional induction of target genes to promote proliferation or confer apoptosis resistance to exposed cells. [401]

Figure 1 has the Chemical carcinogenesis - reactive oxygen species map

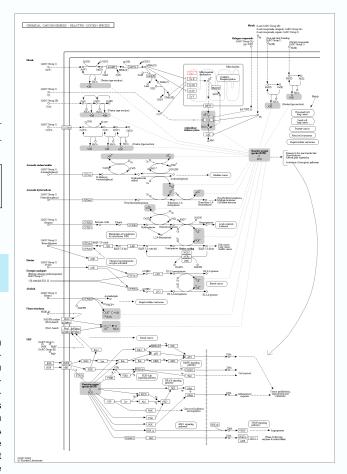


Figure 1: Chemical carcinogenesis - reactive oxygen species [401]

2.1. Water photolysis

The first step in photosynthesis is light dependent water separation to provide the electrons for the photosynthetic electron transport chains as well as protons for the establishment of a proton gradient. The water-seperation reactionis on the lumenal side of the thylakoid membranefrom the light energy captured by the photosystems. This oxidation of water conveniently produces the waste product O2 needed for cellular respiration with the molecular oxygen formed by the reaction released into the atmosphere. Thylakoids contain many integral and peripheral membrane proteins, as well as lumenal proteins.[1]

Figure 2 has the Photosynthesis map

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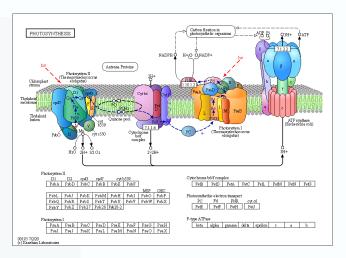


Figure 2: Photosynthesis with C00001 H2O, C00002 ATP, C00005 NADPH, C00006 NADP+, C00007 Oxygen, C00008 ADP, C00009 Orthophosphate, C00034 Manganese, C00080 H+, C02061 Plastoquinone and C02185 Plastoquinol-1 [401]

According to these studies, the thylakoid proteome consists of at least 335 different proteins. Out of these, 89 are in the lumen, 116 are integral membrane proteins, 62 are peripheral proteins on the stroma side, and 68 peripheral proteins on the lumenal side. Of the thylakoid proteins with known functions, 42 percent are involved in photosynthesis with the next largest functional groups include proteins involved in protein targeting, processing and folding with 11 percent, oxidative stress response (9 percent) and translation (8 percent). [1]

2.2. Integral membrane proteins

Thylakoid membranes contain integral membrane proteins with an important role in light-harvesting and the light-dependent reactions of photosynthesis. There are four major protein complexes in the thylakoid membrane: (a) Photosystems I and II (b) Cytochrome b6f complex and (c) ATP synthase. Photosystem II is in grana thylakoids and photosystem I and ATP synthase are in stroma thylakoids and the outer layers of grana. The cytochrome b6f complex is distributed evenly throughout thylakoid membranes. Because of separation the two photosystems in the thylakoid membrane system, mobile electron carriers (plastoquinone and plastocyanin) have to shuttle electrons between them. Plastoquinone shuttles electrons from photosystem II to the cytochrome b6f complex, whereas plastocyanin carries electrons from the cytochrome b6f complex to photosystem I. [1]

Together, these proteins use light energy to drive electron transport chains for a chemiosmotic potential across the thylakoid membrane and NADPH, a product of the terminal redox reaction. The ATP synthase uses the chemiosmotic potential to make ATP during photophosphorylation. [1]

2.3. NADPH

Figure 3 has the chemical structure of NADPH, TPNH, Reduced nicotinamide adenine dinucleotide phosphate with formula C21H30N7O17P3, EXACT MASS 745.0911 and MOL WEIGHT 745.4209 for KEGG compound C00005 and Pubchem 3307. [800]

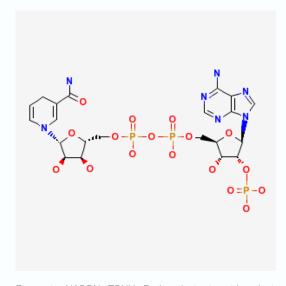


Figure 3: NADPH, TPNH, Reduced nicotinamide adenine dinucleotide phosphate C00005 [401]

There are N1=134 reactions and N2=588 Enzymes with 557 in class 1 Oxidoreductases and rest in 2,3 and 4 Transferases, Hydrolases and Lyases. Categories 1.1.1 Acting on the CH-OH group of donors With NAD+ or NADP+ as acceptor has 209 and 1.14 Acting on paired donors, with incorporation or reduction of molecular oxygen has 108. Fingerprint Tanimoto-based 2-dimensional similarity search generates N=845 structures in the PubChem database.

Table 1 has the frequency of the top 25 compounds from a subset of pH 7 liquid reactants or products. [401]

Frequency + 6654 C00001 2297 <=> 2296 2 674 C00007 608 C00080 531 C00014 230 C00006 220 C00005 219 C00003 201 C00004 197 C00003 201 C00004 197 C03024 129 C03161 129 C0011 116 C00027 101 3 86 C00028 74 C00030 74 C00030 59 C00031 59 C00031 56 C00022 47 C00031 56 C00022 47 C00018 47		
C00001 2297 <=> 2296 2 674 C00007 608 C00080 531 C00014 230 C00006 220 C00005 219 C00009 212 C00003 201 C00004 197 C03024 129 C00011 116 C00027 101 3 86 C00028 74 C00003 74 C0010 60 4 59 C00033 59 C00033 59 C00031 56 C000022 47		Frequency
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4 59 C00033 59 C00031 56 C00022 47	C00030	74
C00033 59 C00031 56 C00022 47	C00010	60
C00031 56 C00022 47	4	59
C00022 47		59
	C00031	56
C00138 47		47
	C00138	47

Table 2 has the KEGG Chemical Function (KCF) format. (a) KEGG Chemical Function (KCF) format of NADPH. [401]

	X
1	1 C1y C 45.6720 -22.9148
2	2 N4y N 46.0143 -20.6899
3	3 C1y C 45.2747 -24.1557
4	4 O2x O 44.5777 -22.1203
5	5 C8y C 44.7428 -20.2924
6	6 C8x C 46.7783 -19.6078
7	7 C1y C 43.9237 -24.1557
8	8 O2b O 46.0570 -25.2315
9	9 C1y C 43.5020 -22.8903
10	10 C8y C 44.7366 -18.9598
11	11 N5x N 43.5874 -20.9709
12	12 N5x N 45.9960 -18.5503
13	13 O1a O 43.1595 -25.2377
14	14 P1b P 46.0510 -27.0532
15	15 C1b C 42.2489 -22.4991
16	16 C8y C 43.5691 -18.2934
17	17 C8x C 42.4139 -20.3231
18	18 O1c O 46.0632 -28.6487
19	19 O1c O 47.4202 -27.0594
20	20 O1c O 44.7244 -27.0532
21	21 O2b O 41.1730 -22.4504
22	22 N5x N 42.4139 -18.9721
23	23 N1a N 43.5629 -16.9609
24	24 P1b P 39.7364 -22.4441
25	25 O2c O 37.9454 -22.4441
26	26 O1c O 39.7364 -23.9969
27	27 O1c O 39.7426 -20.9525
28	28 P1b P 36.2825 -22.4441
29	29 O2b O 34.8461 -22.4504
30	30 O1c O 36.2887 -23.9295
31	31 O1c O 36.2825 -20.9525
32	32 C1b C 33.4646 -22.4381
33	33 C1y C 32.2175 -22.8721
34	34 O2x O 31.1172 -22.1385
35	35 C1y C 31.8752 -24.2290
36	36 C1y C 30.0475 -22.9699
37	37 C1y C 30.4937 -24.2536
38	38 O1a O 32.6577 -25.2071
39	39 N1y N 28.7272 -21.6861
40	40 O1a O 29.7479 -25.2989
41	41 C2x C 29.7479 -20.9342
42	42 C2x C 27.4555 -20.9892
43	43 C2y C 29.7541 -19.5956
44	44 C2x C 27.4433 -19.7361
45	45 C1x C 28.5682 -18.9965
46	46 C5a C 30.8667 -18.8620
47	47 N1a N 30.8605 -17.6822
48	48 O5a O 32.0524 -19.4609

Table 1: KEGG Chemical Function (KCF) format. (a) KEGG Chemical Function (KCF) format of NADPH. KCF format has three sections; ENTRY, ATOM and BOND. ENTRY section describes the KEGG ID and the type of the entry. ATOM describes the numbering of the atoms, KEGG Atom Types for the labels on the atoms, atomic species (C for carbon, N for nitrogen, etc), and 2D coordinates of the atoms. [2]

Table 3 has the numbering of the bonds, the numbering of the two atoms in the bond, and the bond order, and steric configuration of the bond. The steric configuration of the bond are steric effects are nonbonding interactions that influence the shape (conformation) and reactivity of ions and molecules. ... Steric repulsive forces between overlapping electron clouds result in structured groupings of molecules stabilized by the way that opposites attract and like charges repel. [401]

Bond 1 52 2 11 2 1 #Up 3 2 131 4 4 31 4 1 5 4 2 5 1 6 5 2 6 1 7 63 7 1 8 73 8 1 #Down 9 8 4 9 1 10 9 5 10 2 11 10 5 11 1 12 11 6 12 2 13 12 7 13 1 #Down
2 1121#Up 3 2131 4 3141 5 4251 6 5261 7 6371 8 7381#Down 9 8491 10 95102 11 105111 12 116122
3 2131 4 3141 5 4251 6 5261 7 6371 8 7381 #Down 9 8491 10 95102 11 105111 12 116122
4 3141 5 4251 6 5261 7 6371 8 7381#Down 9 8491 10 95102 11 105111 12 116122
5 4251 6 5261 7 6371 8 7381#Down 9 8491 10 95102 11 105111 12 116122
6 5261 7 6371 8 7381 #Down 9 8491 10 95102 11 105111 12 116122
7 6 3 7 1 8 7 3 8 1 #Down 9 8 4 9 1 10 9 5 10 2 11 10 5 11 1 12 11 6 12 2
8 7 3 8 1 #Down 9 8 4 9 1 10 9 5 10 2 11 10 5 11 1 12 11 6 12 2
8 7 3 8 1 #Down 9 8 4 9 1 10 9 5 10 2 11 10 5 11 1 12 11 6 12 2
9 8491 10 95102 11 105111 12 116122
10 95 10 2 11 10 5 11 1 12 11 6 12 2
11 10 5 11 1 12 11 6 12 2
12 11 6 12 2
14 13 8 14 1
15 14 9 15 1 #Up
16 15 10 16 1
17 16 11 17 2
18 17 14 18 1
19 18 14 19 1
20 19 14 20 2
21 20 15 21 1
22 21 16 22 2
23 22 16 23 1
24 23 21 24 1
25 24 24 25 1
26 25 24 26 1
27 26 24 27 2
28 27 25 28 1
29 28 28 29 1
30 29 28 30 1
31 30 28 31 2
32 31 29 32 1
33 32 33 32 1 #Down
34 33 33 34 1
35 34 33 35 1
36 35 34 36 1
37 36 35 37 1
38 37 35 38 1 #Up
39 38 36 39 1 #Down
40 39 37 40 1 #Up
41 40 39 41 1
42 41 39 42 1
43 42 41 43 2
43 42 41 43 2
45 44 43 45 1
46 45 43 46 1
47 46 46 47 1
48 47 46 48 2
49 48 7 9 1
50 49 10 12 1
51 50 17 22 1
52 51 36 37 1
53 52 44 45 1

Table 2: BOND describes the numbering of the bonds, the numbering of the two atoms in the bond, and the bond order, and steric configuration of the bond. [401] [3]

Figure 3 has the Tanimoto Structural Similarities N=845 with CID5884 with (a) oxygen (b) carbon (c) hydrogen (d) Nitrogen with a location scale [0,1] with 0=start and 1=end for the SMILE sequence.

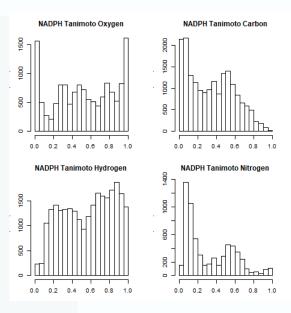


Figure 4: Tanimoto Structural Similarities N=845 with CID5884 with (a) oxygen (b) carbon (c) hydrogen (d) Nitrogen with a location scale [0,1] with 0=start and 1=end for the SMILE sequence. [601] The Pearson distribution with moments are (a) type 2 a 0.9360405 location -0.0176163 scale 1.017616, (b) type 3 shape 1.31062 location 0.003256247 scale 0.2609505, (c) type 2 a 0.9463522 location 0.002016147 scale 0.9966398 and (d) type 3 shape 1.365883 location 0.0120604 scale 0.2251447.

3 Thylakoid Protein Networks

Cyanobacteria are photosynthetic prokaryotes with highly differentiated membrane systems with an internal system of thylakoid membranes of fully functional electron transfer chains of photosynthesis for respiration. The presence of different membrane systems lends these cells a unique complexity among bacteria and reorganization the membranes, synthesize new membrane lipids, and properly target proteins to the correct membrane system is important. The outer membrane, plasma membrane, and thylakoid membranes each have specialized roles in the cyanobacterial cell. [1]

In contrast to the thylakoid network of higher plants, thylakoids in cyanobacteria are organized into multiple concentric shells that split and fuse to parallel layers forming a highly connected network that encloses a single lumen (as in higher-plant chloroplasts) and allows water-soluble and lipid-soluble molecules to diffuse through the entire membrane network. Perforations within the parallel thylakoid sheets allow for the traffic of particles of different sizes throughout the cell: ribosomes, glycogen granules, and lipid bodies. The relatively large distance between the thylakoids provides space for the external light-harvesting antennae, the phycobilisomes. This macrostructure, as in the case of higher plants has flexibility during changes in the physicochemical environment. [1]

In Eudicots where N=93, the distribution is given by Mustard family (9), Caper family (1), Papaya family (1), Rue family (2), Sumac family (2), Mallow family (5), Myrtle family (1), Pea family (15), Rose family (8), Buckthorn family (1), Mulberry family (1), Cucumber family (7), Spurge family (4), Willow family (3), Walnut family (1), Beech family (2), Bittersweet family (1), Grape family (2), Nightshade family (8), Morning-glory family (2), Sesame family (1), Olive family (1), Lopseed family (1), Mint family (1), Daisy family (4), Parsley family (1), Tea family (1), Amaranth family (3), Lotus family (1), Protea family (1), Poppy family (1) and Water-lily family (1). In Monocots, N=18 and the distribution is Grass family (12), Palm family (2), Banana family (1), Orchid family (2) and Asparagus family (1).

3.1. Oryza sativa ssp. japonica cultivar Nipponbare (Japanese rice)

Monocots with the Grass family includes: (a) osa Oryza sativa japonica (Japanese rice) (RefSeq) (b) dosa Oryza sativa japonica (Japanese rice) (RAPDB), (c) obr Oryza brachyantha (malo sina) ,(d) bdi Brachypodium distachyon ,(e) ats Aegilops tauschii (wheat D) ,(f) tdc Triticum dicoccoides (wild emmer wheat) ,(g) taes Triticum aestivum (bread wheat) ,(h) sbi Sorghum bicolor (sorghum) ,(i) zma Zea mays (maize) ,(j) sita Setaria italica (foxtail millet) ,(k) pvir Panicum virgatum (switchgrass) and (l) phai Panicum hallii

Oryza sativa ssp. japonica cultivar Nipponbare (Japanese rice) with T01015 complete Genome Org code osa with the number of protein genes: 28073 and the number of RNA genes: 71.

3.2. Rosa chinensis (China rose)

In the Rose family, the following varieties: (a) fve Fragaria vesca (woodland strawberry), (b) rcn Rosa chinensis (China rose) (c) pper Prunus persica (peach), (d) pmum Prunus mume (Japanese apricot) (e) pavi Prunus avium (sweet cherry), (f) pdul Prunus dulcis (almond), (g) mdm Malus domestica (apple) and (h) pxb Pyrus x bretschneideri (Chinese white pear).

Rosa chinensis (China rose) is from Eukaryota; Viridiplantae; Streptophyta; Embryophyta; Tracheophyta; Spermatophyta; Magnoliopsida; eudicotyledons; Gunneridae; Pentapetalae; rosids; fabids; Rosales; Rosaceae; Rosoideae; Rosoideae incertae sedis; Rosa. The Number of protein genes is 30924 and the Number of RNA genes: 866.

3.3. Species Eudicots and Monocots Families and Thylakoid Proteins

Consider N1=149 unique protein names and N2=165 unique species in both Table 1 and 2 obtained from the KEGG database. The combination generated N3=1597 sequences to examine.

	1	2	3	4
1	Nothobranchius furzeri (turquoise killifish)	Morus notabilis	Spinacia oleracea (spinach)	Synechocystis sp. PCC 6803 GT-S
2	Arabidopsis thaliana (thale cress)	Cucumis sativus (cucumber)	Chenopodium quinoa (quinoa)	Synechocystis sp. PCC 6803 GT-3
3	Arabidopsis Iyrata (Iyrate rockcress)	Cucumis melo (muskmelon)	Nelumbo nucifera (sacred lotus)	Synechocystis sp. PCC 6803 PCC-N
4	Capsella rubella	Benincasa hispida (wax gourd)	Macadamia integrifolia (macadamia nut)	Synechocystis sp. PCC 6803 PCC-N
5	Camelina sativa (false flax)	Momordica charantia (bitter melon)	Papaver somniferum (opium poppy)	Synechocystis sp. PCC 6714
6	Brassica rapa (field mustard)	Cucurbita maxima (winter squash)	Nymphaea colorata	Synechocystis sp. IPPAS B-1465
7	Brassica rapa (fleid mustard) Brassica napus (rape)	Cucurbita maxima (winter squash) Cucurbita moschata (crookneck pumpkin)	Oryza sativa japonica (Japanese rice) (RefSeg)	Synechococcus elongatus PCC7942
8	Brassica oleracea (wild cabbage)	Cucurbita pepo subsp. pepo (vegetable marrow)	Oryza sativa japonica (Japanese rice) (RAPDB)	Synechococcus sp. CC9902
9	Raphanus sativus (radish)	Ricinus communis (castor bean)	Oryza brachyantha (malo sina)	Synechococcus sp. WH 8103
10	Tarenaya hassleriana (spider flower)	Jatropha curcas	Brachypodium distachyon	Synechococcus sp. PCC 73109
11	Carica papaya (papaya)	Hevea brasiliensis (rubber tree)	Aegilops tauschii (wheat D)	Synechococcus sp. PCC 7003
12	Citrus sinensis (Valencia orange)	Manihot esculenta (cassava)	Triticum dicoccoides (wild emmer wheat)	Prochlorococcus sp. MIT 0604
13	Pistacia vera (pistachio)	Populus trichocarpa (black cottonwood)	Triticum aestivum (bread wheat)	Prochlorococcus sp. MIT 0801
14	Mangifera indica (mango)	Populus euphratica (Euphrates poplar)	Sorghum bicolor (sorghum)	Synechococcus lividus
15	Theobroma cacao (cacao)	Populus alba (white poplar)	Zea mays (maize)	Thermosynechococcus sp. NK55
16	Gossypium raimondii	Juglans regia (English walnut)	Setaria italica (foxtail millet)	Kovacikia sp. CCNU0001
17	Gossypium hirsutum (upland cotton)	Quercus suber (cork oak)	Panicum virgatum (switchgrass)	Halomicronema hongdechloris
18	Gossypium arboreum	Quercus lobata (valley oak)	Panicum hallii	Pseudanabaena sp. ABRG5-3
19	Durio zibethinus (durian)	Tripterygium wilfordii	Phoenix dactylifera (date palm)	Geminocystis sp. NIES-3709
20	Eucalyptus grandis (rose gum)	Vitis vinifera (wine grape)	Elaeis guineensis (African oil palm)	Geminocystis sp. NIES-3708
21	Glycine max (soybean)	Vitis riparia (riverbank grape)	Musa acuminata (wild Malaysian banana)	Microcystis panniformis
22	Glycine soja (wild soybean)	Solanum lycopersicum (tomato)	Dendrobium catenatum	Microcystis sp. MC19
23	Vigna radiata (mung bean)	Solanum pennellii	Phalaenopsis equestris	Cyanobacterium sp. HL-69
24	Vigna angularis (adzuki bean)	Solanum tuberosum (potato)	Asparagus officinalis (garden asparagus)	Trichodesmium erythraeum
25	Vigna unguiculata (cowpea)	Capsicum annuum	Amborella trichopoda	Oxynema aestuarii
26	Cajanus cajan (pigeon pea)	Nicotiana tabacum (common tobacco)	Physcomitrium patens	Limnospira fusiformis
27	Abrus precatorius (Indian licorice)	Nicotiana sylvestris	Chlamydomonas reinhardtii	Moorea producens
28	Medicago truncatula (barrel medic)	Nicotiana tomentosiformis	Volvox carteri f. nagariensis	Phormidium sp. PBR-2020
29	Cicer arietinum (chickpea)	Nicotiana attenuata	Monoraphidium neglectum	Nostoc sp. PCC 7120
30	Arachis duranensis	Ipomoea nil (Japanese morning glory)	Coccomyxa subellipsoidea	Nostoc sp. NIES-3756
31	Arachis ipaensis	Ipomoea triloba (trilobed morning glory)	Ostreococcus lucimarinus	Nostoc sp. CENA543
32	Arachis hypogaea (peanut)	Sesamum indicum (sesame)	Micromonas commoda	Nostoc sphaeroides
33	Lupinus angustifolius (narrow-leaved blue lupine)	Olea europaea var. sylvestris (wild olive)	Cyanidioschyzon merolae	Nostoc edaphicum
34	Fragaria vesca (woodland strawberry)	Erythranthe guttata (spotted monkey flower)	Galdieria sulphuraria	Anabaena sp. WA102
35	Rosa chinensis (China rose)	Salvia splendens (scarlet sage)	Thalassiosira pseudonana	Calothrix sp. 336/3
36	Prunus persica (peach)	Helianthus annuus (common sunflower)	Acetobacter oryzifermentans	Nodularia spumigena
37	Prunus mume (Japanese apricot)	Erigeron canadensis (horseweed)	Acetobacter pomorum	Cylindrospermopsis curvispora
38	Prunus avium (sweet cherry)	Lactuca sativa (garden lettuce)	Acetobacter sp. JWB	Nostocales cyanobacterium HT-58-2
39	Prunus dulcis (almond)	Cynara cardunculus var. scolymus (artichoke)	Acetobacter oryzoeni	Viruses (Paramecium bursaria Chlorella virus NYs1)
40	Malus domestica (apple)	Daucus carota (carrot)	Clostridium pasteurianum DSM 525 = ATCC 6013	Nothobranchius furzeri (turquoise killifish)
41	Pyrus x bretschneideri (Chinese white pear)	Camellia sinensis	Thermoclostridium stercorarium subsp. stercorarium DSM 8532	Arabidopsis thaliana (thale cress)
42	Ziziphus jujuba (Chinese jujube)	Beta vulgaris (sugar beet)	Syntrophomonas wolfei	Arabidopsis lyrata (lyrate rockcress)
42	Zizipnus jujuba (Gninese jujubė)	Beta vulgaris (sugar beet)	Syntropnomonas woilei	Arabidopsis iyrata (iyrate rockcress)

Table 2 has the list of the thylakoid proteins.

	1	2	3	4
1	(RefSeq) thylakoid ADP,ATP carrier protein, chloroplastic-like	(RefSeq) thylakoid membrane protein TERC, chloro- plastic isoform X4	(GenBank) thylakoid lumenal 17.4 kDa protein	(RefSeq) thylakoid lumenal 30 kDa protein, probable
2	(RefSeq) cold regulated 314 thylakoid membrane 2	(RefSeq) thylakoid membrane protein slr0575	(GenBank) thylakoid lumenal 17.9 kDa protein	(RefSeq) chloroplast thylakoid protein kinase STN7, probable
3	(RefSeq) thylakoid lumen 18.3 kDa protein	(RefSeq) thylakoid membrane protein TERC, chloro- plastic isoform X2	(GenBank) thylakoid membrane phosphoprotein 14 kDa	(RefSeq) thylakoid lumenal 20 kDa protein, putative
4	(RefSeq) thylakold lumenal protein (Mog1/PsbP/DUF1795-like photosystem II reaction center PsbP family protein)	(RefSeq) LOW QUALITY PROTEIN: thylakoidal pro- cessing peptidase 1, chloroplastic-like	(GenBank) thylakoid lumenal protein	(RefSeq) thylakoid lumenal protein, chloroplast precursor
5	(RefSeq) thylakoidal ascorbate peroxidase	(RefSeq) thylakoid lumenal 15.0 kDa protein 2, chloroplastic-like	(GenBank) thylakoid lumenal 29 kDa protein	(RefSeq) thylakoid lumenal 25.6kDa protein like, chloro- plast precursor
6	(RefSeq) thylakoid processing peptide	(RefSeq) thylakoid membrane protein TERC, chloroplastic-like isoform X1	(GenBank) thylakoid membrane slr0575-like protein	(RefSeq) similar to thylakoid lumenal 17.4 kD protein, chloroplast precursor
7	(RefSeq) thylakoid lumenal P17.1 protein	(RefSeq) thylakoid membrane protein TERC, chloroplastic-like isoform X2	(GenBank) oxygen-evolving complex/thylakoid lumenal 25.6 kDa protein	(RefSeq) probable thylakoid lumen rotamase
8	(RefSeq) thylakoid soluble phosphoprotein	(RefSeq) thylakoid lumenal 15 kDa protein 1, chloroplastic-like isoform X1	(GenBank) thylakoid rhodanese-like protein	(RefSeq) similar to thylakoid lumen rotamase
9	(RefSeq) thylakoid rhodanese-like protein	(RefSeq) thylakoid lumenal protein TL20.3, chloroplastic-like	(GenBank) thylakoid lumenal 16.5 kDa protein	(RefSeq) similar to thylakoidal sec-independent protein transporter Tha4
10	(RefSeq) chloroplast thylakoid lumen protein	(RefSeq) thylakoid membrane protein TERC, chloro- plastic isoform X1	(RefSeq) thylakoidal processing peptidase 1, chloro- plastic isoform X1	(RefSeq) thylakoid protein
11	(RefSeq) thylakoid lumenal 17.9 kDa protein, chloro- plast	(RefSeq) thylakoid lumenal 29 kDa protein, chloroplas- tic isoform X1	(RefSeq) thylakoid lumenal protein TL20.3, chloroplastic isoform X2	(RefSeq) thylakoid lumenal 17.4 kDa protein, chloro- plast precursor
12	(RefSeq) thylakoid-associated phosphatase 38	(RefSeq) thylakoid lumenal 16.5 kDa protein, chloroplastic-like	(RefSeq) thylakoid lumenal 15.0 kDa protein 2, chloroplastic-like isoform X1	(RefSeq) thylakoidal processing peptidase
13	(RefSeq) thylakoid ATP/ADP carrier	(RefSeq) thylakoid membrane protein slr0575-like	(RefSeq) LOW QUALITY PROTEIN: thylakoid lumenal 17.4 kDa protein, chloroplastic	(RefSeq) thylakoid lumen 15.0-kDa protein
14	(RefSeq) thylakoid lumen 15.0 kDa protein	(RefSeq) thylakoid lumenal protein At1g12250, chloro- plastic	(RefSeq) thylakoid lumenal 15 kDa protein 1, chloro- plastic isoform X2	(RefSeq) ATP synthase alpha chain, thylakoid ATPase, plastid protein
15	(RefSeq) thylakoid lumenal 17.9 kDa protein, chloro- plastic	(RefSeq) thylakoid lumenal 15.0 kDa protein 2, chloro- plastic isoform X1	(RefSeq) thylakoid lumenal 15.0 kDa protein 2, chloro- plastic isoform X2	(RefSeq) plastid thylakoid ATPase delta chain, plastid protein
16	(RefSeq) thylakoid lumenal 16.5 kDa protein, chloro- plastic	(RefSeq) thylakoidal processing peptidase 1, chloroplastic-like isoform X1	(RefSeq) probable thylakoidal processing peptidase 2, chloroplastic-like	(RefSeq) plastid thylakoid ATPase subunit, plastid pro- tein
17	(RefSeq) thylakoid ADP,ATP carrier protein, chloroplas- tic	(RefSeq) LOW QUALITY PROTEIN: thylakoid lumenal 16.5 kDa protein, chloroplastic	(RefSeq) LOW QUALITY PROTEIN: thylakoid mem- brane protein slr0575	(GenBank) thylakoid lumen protein
18	(RefSeq) thylakoid lumenal 19 kDa protein, chloroplas- tic	(RefSeq) thylakoid lumenal 29 kDa protein, chloroplastic-like isoform X1	(RefSeq) thylakoid-bound ascorbate peroxidase 6	(GenBank) thylakoid-associated protein
19	(RefSeq) thylakoidal processing peptidase 1, chloro- plastic	(RefSeq) thylakoid membrane protein ssl2009-like	(RefSeq) probable thylakoid lumen protein sll1769	(GenBank) thylakoid-like protein
20	(RefSeq) probable thylakoidal processing peptidase 2, chloroplastic	(RefSeq) thylakoid lumenal 17.9 kDa protein, chloro- plastic isoform X1	(RefSeq) LOW QUALITY PROTEIN: thylakoid lumenal protein TL20.3, chloroplastic	(GenBank) thylakoidal processing peptidase. Serine peptidase. MEROPS family S26A
21	(RefSeq) thylakoid lumenal protein TL20.3, chloroplas- tic isoform X1	(RefSeq) thylakoid membrane protein slr0575 isoform X1	(RefSeq) thylakoid ADP,ATP carrier protein, chloroplastic-like isoform X1	(GenBank) plasma membrane protein essential for thy-
22	(RefSeq) thylakoid lumenal 15.0 kDa protein 2, chloro- plastic	(RefSeq) thylakoid lumenal 17.9 kDa protein, chloro- plastic isoform X2	(RAP-DB) Similar to thylakoid lumenal 17.9 kDa protein, chloroplast.	(GenBank) Putative thylakoid membrane protein, con- tains 8 pentapeptide repeats
23	(RefSeq) thylakoid lumenal 17.4 kDa protein, chloro- plastic	(RefSeq) thylakoid lumenal 17.4 kDa protein, chloroplastic-like isoform X3	(RAP-DB) Similar to thylakoid-bound ascorbate peroxi- dase APx8.	(GenBank) thylakoid membrane protein
24	(RefSeq) thylakoid membrane protein TERC, chloro- plastic	(RefSeq) thylakoid lumenal 15 kDa protein 1, chloroplastic-like	(RefSeq) LOW QUALITY PROTEIN: thylakoid lumenal 17.9 kDa protein, chloroplastic	(GenBank) thylakoid-associated single-stranded DNA- binding protein
25	(RefSeq) LOW QUALITY PROTEIN: thylakoid lumenal 29 kDa protein, chloroplastic	(RefSeq) thylakoid lumenal 17.4 kDa protein, chloroplastic-like isoform X1	(RefSeq) thylakoid lumenal 29 kDa protein, chloroplas- tic isoform X4	(GenBank) thylakoid lumenal 15 kDa protein 1, chloro- plastic
26	(RefSeq) thylakoid lumenal 15 kDa protein 1, chloro- plastic isoform X1	(RefSeq) thylakoid membrane protein slr0575-like iso- form X1	(RefSeq) thylakoid lumenal 17.9 kDa protein, chloroplastic-like isoform X1	(GenBank) Putative thylakoid membrane protein
27	(RefSeq) thylakoid lumenal 17.4 kDa protein, chloro- plastic isoform X1	(RefSeq) thylakoid lumenal 16.5 kDa protein, chloro- plastic isoform X1	(RefSeq) thylakoid assembly 8	(GenBank) ring forming protein essential for thylakoid formation Vino1
28	(RefSeq) thylakoid lumenal 29 kDa protein, chloroplastic	(RefSeq) thylakoid lumenal 16.5 kDa protein, chloroplastic-like isoform X1	(RefSeq) chloroplast oxygen-evolving complex/thy- lakoid lumenal 25.6kDa protein	(GenBank) putative thylakoid membrane protein
29	(RefSeq) thylakoid lumenal 15 kDa protein 1, chloro- plastic	(RefSeq) thylakoid ADP,ATP carrier protein, chloroplas- tic isoform X1	(RefSeq) thylakoid lumenal 19 kDa protein	(GenBank) thylakoid signal peptidase I LepB1
30	(RefSeq) thylakoidal processing peptidase 1, chloroplastic-like	(RefSeq) LOW QUALITY PROTEIN: thylakoid lumenal 19 kDa protein, chloroplastic	(RefSeq) LOW QUALITY PROTEIN: probable thy- lakoidal processing peptidase 2, chloroplastic	(GenBank) high-affinity Na+(Li+):H+ thylakoid mem- brane antiporter NhaS3
31	(RefSeq) thylakoid lumenal 17.9 kDa protein, chloroplastic-like	(RefSeq) thylakoid lumenal 19 kDa protein, chloroplas- tic isoform X1	(RefSeq) LOW QUALITY PROTEIN: thylakoidal pro- cessing peptidase 1, chloroplastic	(GenBank) thylakoid membrane photosystem I accumu- lation factor
32	(RefSeq) thylakoid lumenal 29 kDa protein, chloroplastic-like	(GenBank) thylakoid lumenal 15.0 kDa protein	(RefSeq) photosystem II thylakoid lumenal 29.8 kDa protein PsbP	(GenBank) putative thylakoid lumen protein
33	(RefSeq) thylakoid lumenal protein TL20.3, chloroplastic-like isoform X1	(GenBank) thylakoid lumen 18.3 kDa protein	(RefSeq) thylakoid membrane protein	(GenBank) biogenesis of thylakoid protein
34	(RefSeq) thylakoid lumenal 17.4 kDa protein, chloroplastic-like	(GenBank) thylakoid lumenal 19 kDa protein	(RefSeq) thylakoid lumenal protein	(GenBank) biogenesis of thylakoid protein A
35	(RefSeq) probable thylakoidal processing peptidase 2, chloroplastic isoform X1	(GenBank) thylakoid soluble phosphoprotein TSP9 pro- tein	(RefSeq) thylakoid lumen protein	(RefSeq) Paramecium bursaria Chlorella virus NYs1; thylakoid formation protein
36	(RefSeq) thylakoid membrane protein TERC, chloroplastic-like	(GenBank) thylakoid lumenal 15 kDa protein	(RefSeq) thylakoid formation protein 1	(RefSeq) thylakoid ADP,ATP carrier protein, chloroplastic-like
37	(RefSeq) thylakoid lumenal 19 kDa protein, chloroplastic-like	(GenBank) thylakoid-bound ascorbate peroxidase	(RefSeq) putative thylakoid lumenal 29.8 kDa protein	(RefSeq) cold regulated 314 thylakoid membrane 2
	(RefSeq) thylakoid lumenal protein TL20.3, chloroplas-	(GenBank) thylakoid lumenal 29.8 kDa protein	(RefSeq) Tat family transporter: pH-dependent thy-	(RefSeq) thylakoid lumen 18.3 kDa protein

Molecular properties are abundant in dimensional reduction for collections of sequences. Examples such as stability, binding potential aliphatic and hydrophobicity along with the charge at different pH is a few of available molecular properties to examine based on the gene ontology ids in Table 2. The net charge of a protein sequence based on the Henderson-Hasselbalch equation based on pH 5, 7 and 9. The aliphatic index is the relative volume occupied by aliphatic side chains (Alanine, Valine, Isoleucine, and Leucine) and is a positive factor for the increase of thermostability of globular proteins. The potential protein interaction index proposed by Boman (2003) based in the amino acid sequence of a protein and provides an overall estimate of the potential of a peptide to bind to membranes or other proteins as receptors. A protein have high binding potential if the index value is higher than 2.48. This index predicts the stability of a protein based on its amino acid composition, a protein whose instability index is smaller than 40 is predicted as stable, a value above 40 predicts that the protein may be unstable. Hydrophobicity is an important stabilization force in protein folding; this force changes depending on the solvent in which the protein is found. [1001]

Table 3 has the molecular properties for Rosa chinensis (China rose) for each of the thylakoid proteins.

	Species	Protein	Stability Index	Binding Potential	ALiphatic	f.1	CpH5	CpH7	CpH9
498	Rosa chinensis (China rose)	(RefSeq) probable thylakoidal processing peptidase 2, chloroplastic	50.229	1.2327	94.0	-0.012772	8.44e+00	1.81158	-4.55458
499	Rosa chinensis (China rose)	(RefSeq) thylakoid lumenal 15 kDa protein 1, chloroplastic	19.088	1.3862	84.4	-0.128169	8.62e-01	-2.65045	-7.21622
500	Rosa chinensis (China rose)	(RefSeq) thylakoid lumenal 29 kDa protein, chloroplastic	35.773	1.7112	84.1	-0.344092	3.05e+00	-1.08483	-5.24162
501	Rosa chinensis (China rose)	(RefSeq) thylakoid lumenal 29 kDa protein, chloroplastic isoform X1	35.506	1.7827	81.4	-0.359574	3.63e+00	0.90610	-2.61588
502	Rosa chinensis (China rose)	(RefSeq) thylakoid lumenal protein TL20.3, chloroplastic isoform X1	48.416	1.6834	76.1	-0.234743	1.04e+01	3.19342	-5.46992
503	Rosa chinensis (China rose)	(RefSeq) thylakoid lumenal 17.9 kDa protein, chloroplastic	49.541	1.3600	81.1	-0.307627	4.99e+00	0.62118	-3.57338
504	Rosa chinensis (China rose)	(RefSeq) thylakoid lumenal 17.4 kDa protein, chloroplastic	34.418	1.4846	86.8	-0.083691	6.19e+00	3.05174	-2.71363
505	Rosa chinensis (China rose)	(RefSeq) thylakoid lumenal 16.5 kDa protein, chloroplastic	41.827	1.5464	92.1	-0.100935	5.13e+00	0.65678	-2.49249
506	Rosa chinensis (China rose)	(RefSeq) thylakoid membrane protein slr0575	32.448	1.3082	87.4	-0.128058	1.42e+01	5.82286	0.10253
507	Rosa chinensis (China rose)	(RefSeq) thylakoid membrane protein TERC, chloroplastic	20.010	0.1643	122.7	0.732061	9.69e-01	-0.84109	-3.84147
508	Rosa chinensis (China rose)	(RefSeq) thylakoid lumenal 15.0 kDa protein 2, chloroplastic	39.004	1.1985	87.4	-0.082063	9.77e-01	-2.61777	-6.39816
509	Rosa chinensis (China rose)	(RefSeq) LOW QUALITY PROTEIN: thylakoid lumenal 19 kDa protein, chloroplastic	29.919	1.5465	68.3	-0.512917	4.18e+00	-1.84206	-5.77130
510	Rosa chinensis (China rose)	(RefSeq) thylakoid membrane protein TERC, chloroplastic isoform X1	32.997	0.7072	104.4	0.390160	5.96e+00	-1.68130	-7.71025
511	Rosa chinensis (China rose)	(RefSeq) LOW QUALITY PROTEIN: thylakoid lumenal 29 kDa protein, chloroplastic	31.883	1.6064	80.8	-0.402190	-2.59e+00	-6.09156	-10.20765
512	Rosa chinensis (China rose)	(RefSeg) probable thylakoidal processing peptidase 2, chloroplastic	30.410	0.9423	91.5	-0.017391	-2.64e+00	-7.14469	-11.51717

Table 4 has the molecular properties for Rosa chinensis (China rose) for each of the thylakoid proteins.

	Species	Protein	Stability Index	Binding Potential	ALiphatic	f.1	CpH5	СрН7	СрН9
1220	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid ADP,ATP carrier protein, chloroplastic	39.808	1.0101	90.9	0.000262	2.19e+01	15.07598	9.08661
1221	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoidal processing peptidase 1, chloroplastic	65.680	1.3368	75.5	-0.143552	1.39e+01	5.52554	-1.70164
1222	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid lumenal 15 kDa protein 1, chloroplastic	15.641	1.2523	84.4	-0.075253	3.42e+00	0.40663	-2.62287
1223	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid lumenal 17.4 kDa protein, chloroplastic	39.262	1.3700	79.9	0.010169	3.15e+00	0.77958	-5.50364
1224	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid lumenal protein TL20.3, chloroplastic isoform X1	38.770	1.6883	71.4	-0.209386	1.01e+01	6.25959	-0.54193
1225	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid lumenal 17.9 kDa protein, chloroplastic isoform X1	58.846	1.5643	73.2	-0.208494	9.69e+00	5.49351	-1.75768
1226	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid lumenal 29 kDa protein, chloroplastic	44.285	1.6712	84.1	-0.293768	1.08e+01	4.57527	-1.53218
1227	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid membrane protein TERC, chloroplastic	49.331	0.9880	101.1	0.249344	4.69e+00	-0.57573	-4.35237
1228	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid lumenal 16.5 kDa protein, chloroplastic	45.977	1.3544	89.6	-0.027615	7.13e+00	3.38410	-0.38791
1229	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid lumenal 15.0 kDa protein 2, chloroplastic	36.908	1.0797	86.5	0.099593	4.12e+00	1.02092	-5.41130
1230	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid membrane protein slr0575	54.038	1.1617	87.2	0.003690	9.50e+00	5.38260	1.08299
1231	Oryza sativa japonica (Japanese rice) (RefSeq)	(RefSeq) thylakoid lumenal 19 kDa protein, chloroplastic	40.832	1.2830	80.9	-0.140329	5.29e+00	0.68164	-2.43518
1232	Oryza sativa japonica (Japanese rice) (RAPDB)	(RAP-DB) Similar to thylakoid lumenal 17.9 kDa protein, chloroplast.	58.477	1.5790	72.7	-0.223346	9.69e+00	5.49351	-1.75768
1233	Oryza sativa japonica (Japanese rice) (RAPDB)	(RAP-DB) Similar to thylakoid-bound ascorbate peroxidase APx8.	47.748	1.5927	68.7	-0.565635	-1.00e+01	-17.29263	-20.83042

For the 1597 proteins for all species the descriptive statistics are in Table 5.

	vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis	se
Stability Index	1	1597.00	39.32	10.17	39.81	39.74	8.98	0.03	73.37	73.34	-0.39	0.49	0.25
Binding Potential	2	1597.00	1.38	0.33	1.41	1.40	0.28	-0.57	2.45	3.02	-0.62	2.04	0.01
ALiphatic	3	1597.00	86.09	10.34	84.56	85.19	8.03	45.00	143.27	98.27	0.92	2.50	0.26
f.1	4	1597.00	-0.12	0.21	-0.13	-0.14	0.16	-0.98	1.11	2.09	0.90	2.60	0.01
CpH5	5	1597.00	6.54	5.32	5.68	6.07	3.61	-13.85	39.44	53.29	1.21	4.11	0.13
CpH7	6	1597.00	1.59	4.69	1.09	1.26	3.22	-22.61	28.43	51.04	0.94	4.20	0.12
CpH9	7	1597.00	-3.63	4.63	-3.79	-3.88	3.28	-27.43	16.63	44.07	0.69	3.75	0.12

Consider Figure 1 with Drug Activity Z scores with NSC:284751 adenosine, 8-chloro-, cyclic 3',5'-(hydrogen phosphate) Pubchem 144036 and SMILES Nc1ncnc2c1nc(Cl)n2C3OC4COP(=O)(O)OC4C3O for the NCI-60 Cell lines.

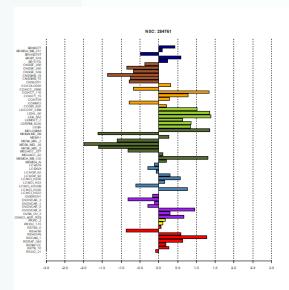


Figure 5: Drug Activity Z scores with NSC:284751 adenosine, 8-chloro-, cyclic 3',5'-(hydrogen phosphate) Pubchem 144036 and SMILES Nc1ncnc2c1nc(Cl)n2C3OC4COP(=O)(O)OC4C3O

4 Conclusions

In this brief mathematical note, the chemical structure of NADPH was examined with respect to the pathways of map00195 Photosynthesis, map00480 Glutathione metabolism, map01100 Metabolic pathways, map04918 Thyroid hormone synthesis, map04925 Aldosterone synthesis and secretion, map04936 Alcoholic liver disease map05208 Chemical carcinogenesis - reactive oxygen species map05415 Diabetic cardiomyopathy with a focus on Photosythesis, Chemical carcinogenesis - reactive oxygen species and Thylakoid Protein Networks.

Here molecular properties for Monocot and Eudicot Families for Oryza sativa japonica (Japanese rice) and Rosa chinensis (China rose) were examined as well as the descriptive statistics for all species in the data set.

5 Bibliography

- [1] Wikipedia contributors. "Thylakoid." Wikipedia, The Free Encyclopedia. Wikipedia, The Free Encyclopedia, 19 Nov. 2021. Web. 6 Jan. 2022.
- [2] Kotera, M., Tabei, Y., Yamanishi, Y. et al. KCF-S: KEGG Chemical Function and Substructure for improved interpretability and prediction in chemical bioinformatics. BMC Syst Biol 7, S2 (2013). https://doi.org/10.1186/1752-0509-7-S6-S2
- [3] Wikipedia contributors. "Steric effects." Wikipedia, The Free Encyclopedia. Wikipedia, The Free Encyclopedia, 27 Oct. 2021. Web. 6 Jan. 2022.
- [400] Kanehisa, Furumichi, M., Tanabe, M., Sato, Y., and Morishima, K.; KEGG: new perspectives on genomes, pathways, diseases and drugs. Nucleic Acids Res. 45, D353-D361 (2017).

- [401] Kanehisa, M., Sato, Y., Kawashima, M., Furumichi, M., and Tanabe, M.; KEGG as a reference resource for gene and protein annotation. Nucleic Acids Res. 44, D457-D462 (2016).
- [402] Kanehisa, M. and Goto, S.; KEGG: Kyoto Encyclopedia of Genes and Genomes. Nucleic Acids Res. 28, 27-30 (2000).
- [403] Petri, V., Jayaraman, P., Tutaj, M., Hayman, G. T., Smith, J. R., De Pons, J., ... Jacob, H. J. (2014). The pathway ontology – updates and applications. Journal of Biomedical Semantics, 5, 7. http://doi.org/10.1186/2041-1480-5-7
- [601] Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, Simonovic M, Doncheva NT, Morris JH, Bork P, Jensen LJ, von Mering C. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. Nucleic Acids Res. 2019 Jan; 47:D607-613.PubMed
- [800] Kim, Sunghwan et al. "PubChem in 2021: new data content and improved web interfaces." Nucleic Acids Res. vol. 49,D1 (2021): D1388-D1395. doi:10.1093/nar/gkaa971
- [1000] R Core Team. "R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria." (2015). URL https://www.R-project.org/.
- [1001] Osorio, D., Rondon-Villarreal, P. and Torres, R. Peptides: A package for data mining of antimicrobial peptides. The R Journal. 7(1), 4-14 (2015).
- [1002] Grant, B.J. et al. (2006) Bioinformatics 22, 2695-2696.