

# Mass spectrometric analysis of proteins using Electron Transfer Dissociation

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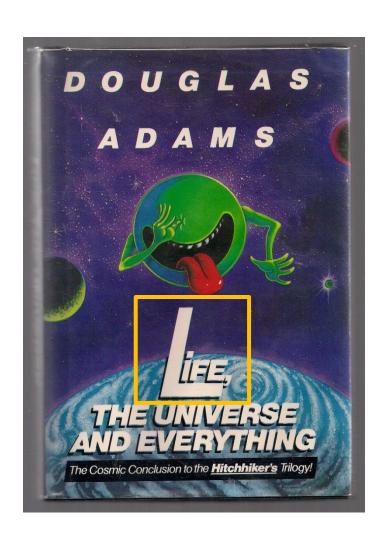
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### Introduction



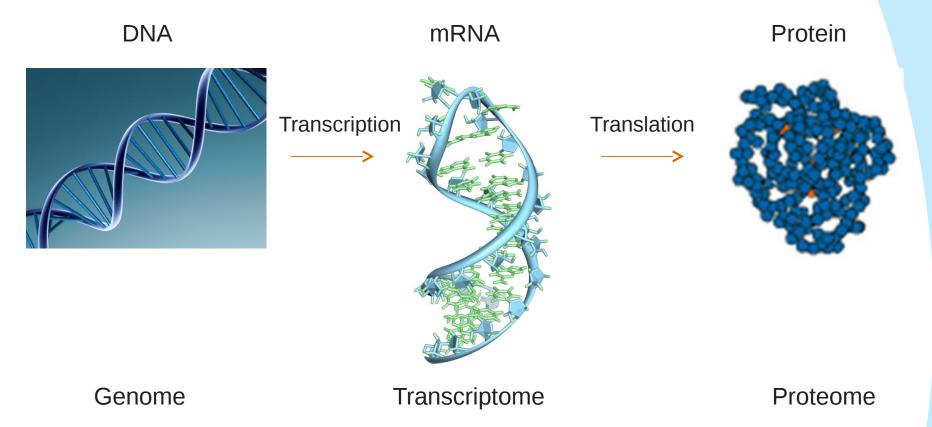


### Overview

- » Biochemistry recap
  - » Central dogma
  - » Protein structure
- » Mass spectrometry
- » Bottom-up vs. top-down
  - » Native vs. denatured
- » Electron transfer dissociation (ETD)
  - » Case study: alcohol dehydrogenase
  - » Effect of ETD on isotopic distribution (bioinformatics)



### Basic biochemistry



Fraser, C.M. et al.: The minimal gene complement of Mycoplasma genitalium. Science 270(5235), 397-404 (1995) Adams, M.D. et al.: The genome sequence of Drosophila melanogaster. Science 287(5461), 2185-2195 (2000) Lander, E. S. et al.: Initial sequencing and analysis of the human genome. Nature, 409(6822), 860-921 (2001) Venter, J. C. et al.: The sequence of the human genome. Science, 291(5507), 1304-1351 (2001) Venter, J.C.: A Part of the Human Genome Sequence. Science, 299(5610), 1183-1184 (2003)

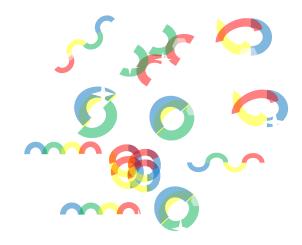


### Basic biochemistry – information content

Genome 2,3x104 Transcripto me >106

Proteom e

>108





### Protein structure

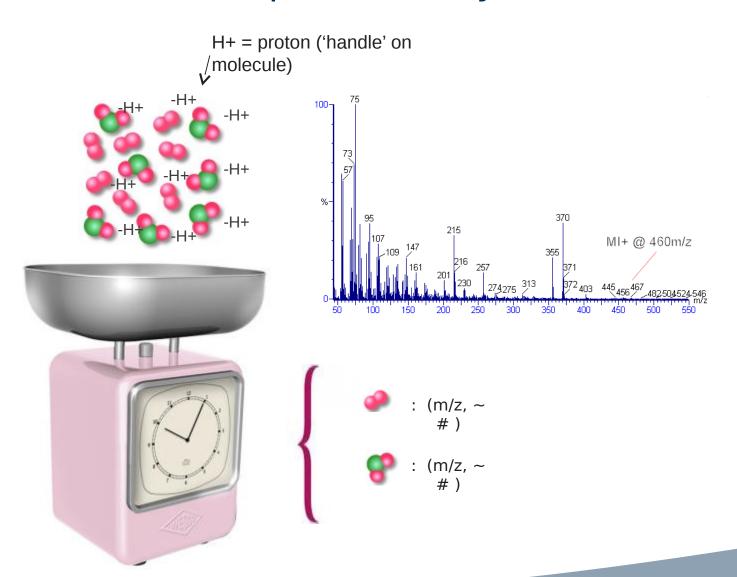
» (Large) linear polymer (chain) of amino acids

$$H_2N \stackrel{\alpha}{=} C \stackrel{O}{=} C \stackrel{O}{=} O$$
R group peptide bond

- » Protein = amino acid sequence
  - + post-translational modifications (PTMs)
  - + 3D structure
  - + protein-protein/protein-ligand complexes
- Now do we analyze this? → Mass spectrometry!



### Mass spectrometry

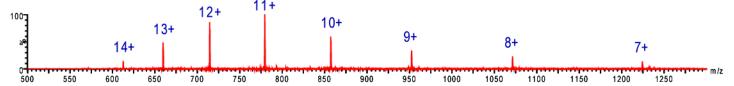




### Charge state distributions

» Number of protons (charges) placed on protein is not a single value, but a distribution!

e.g. ubiquitin (76 amino acids, 8.6 kDa):





### Tandem mass spectrometry

$$m/z = 46.07 \text{ Th}$$

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C2H6O

C2H6O

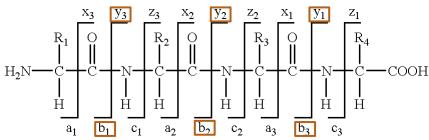
- H2O

СНЗОН

- » Mass analysis: vacuum
- » Collision-induced dissociation (CID): let ions collide with

inert gas molecules

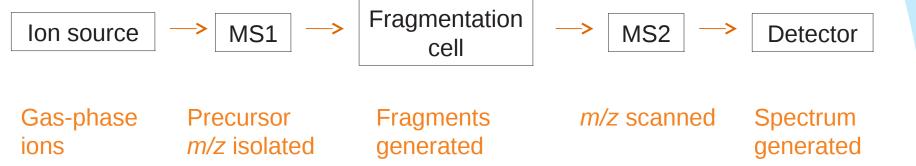
- » Internal energy increases
- » Weakest bond(s) break



» Mass of precursor and fragments reveal molecular structure

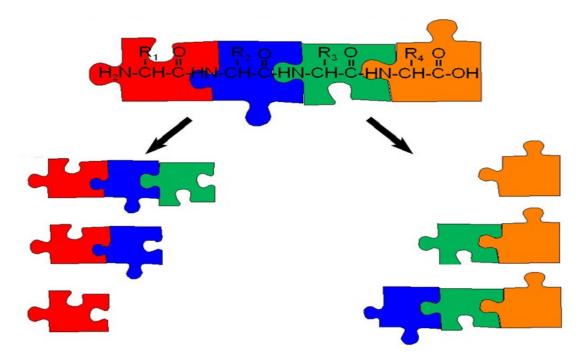


### Tandem mass spectrometry



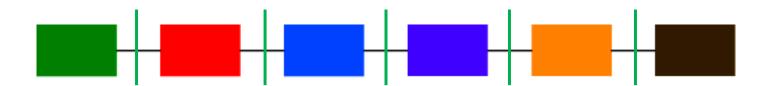


### Tandem mass spectrometry

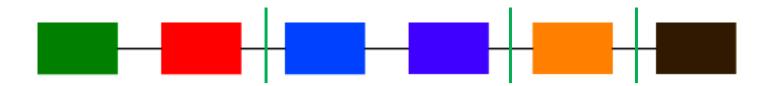


- » Theory:  $\Delta M \rightarrow amino acid sequence$
- » Practice: internal energy buildup  $\rightarrow$  side reactions
  - » e.g. loss of PTMs, protein complexes, ligands, etc.

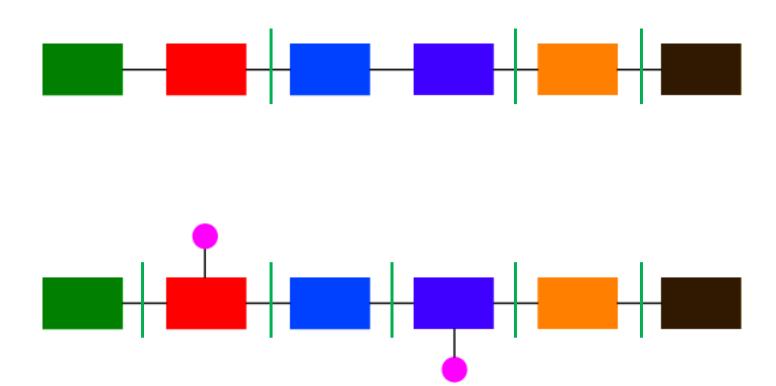




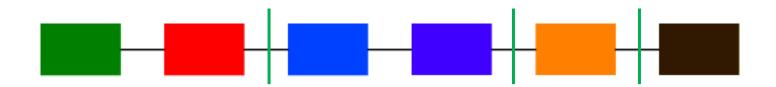


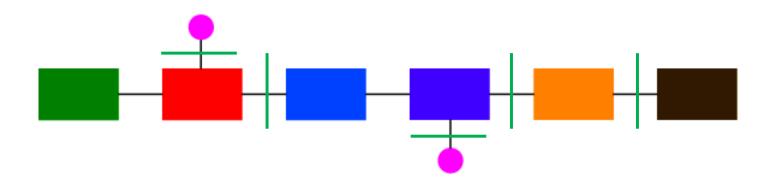






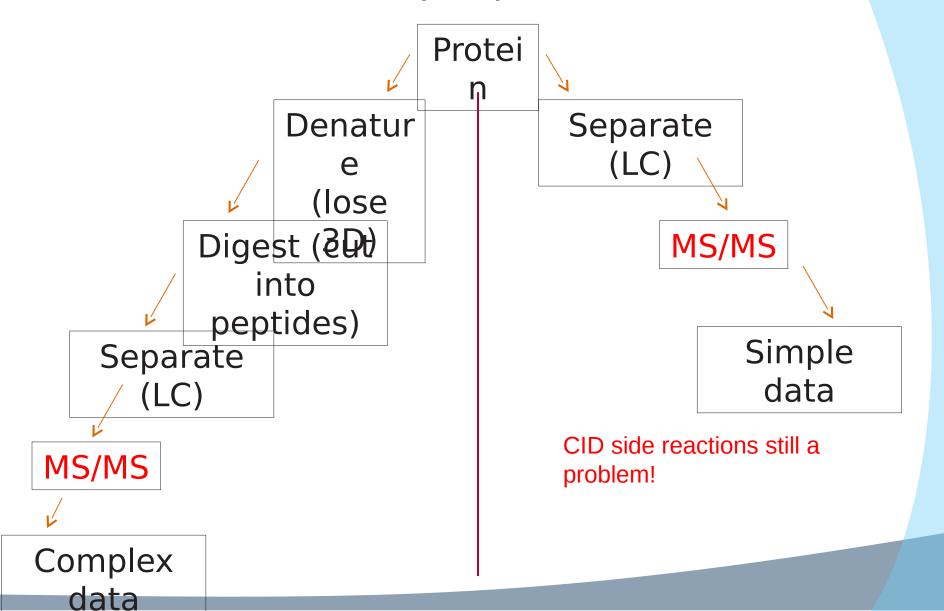








### Bottom-up/top-down





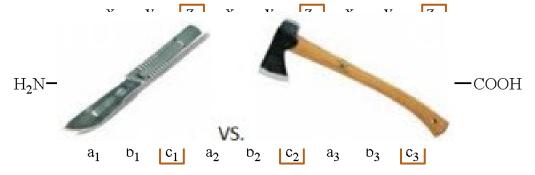
### Native/denatured proteins

- » Native: keep 3D structure (including complexes) intact
- » Why?
  - » Study protein conformations
  - » Study protein/protein or protein/ligand interactions directly
- » How?
  - » Solution phase: no acids/organic solvents
  - » Ionization: 'soft' methods (electrospray ionization/ESI)
  - » Careful control of voltages/pressures in gas phase (minimal internal energy buildup)
- » Can we still perform fragmentation?



### Electron capture/transfer dissociation

- » CID: gradually increase internal energy by 103 collisions
- $^{\circ}$  ECD/ETD: H+ + e-  $\rightarrow$  H $_{\bullet}$  (radical causes fragmentation)



- » Advantage: very specific reaction → minimal side reactions
- » Also: limited preference for certain residues (exception: proline)



### Electron capture/transfer dissociation

» Proline:

» Difference ECD/ETD: source of e-

» ECD: electron beam

» ETD: radical anion

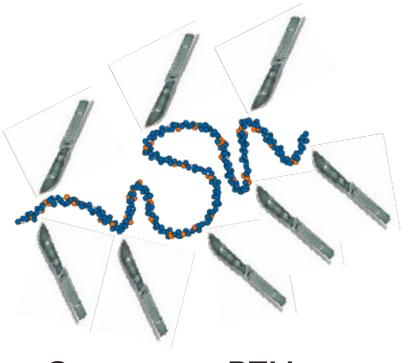
 $^{*}$  ECD: Mz+ + e- → [M<sub>•</sub>(z-1)+]\* → fragments

ETD:  $Mz+ + A_{\bullet}- \rightarrow [M_{\bullet}(z-1)+]^* + A \rightarrow fragments$ 

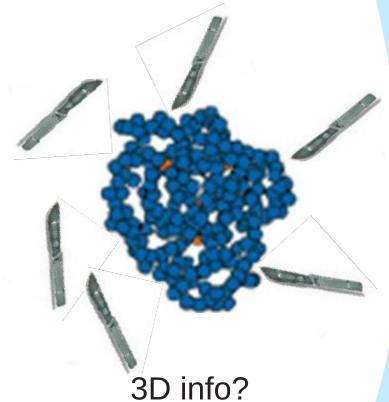


### Electron transfer dissociation

» Is ETD gentle enough to perform on native proteins?

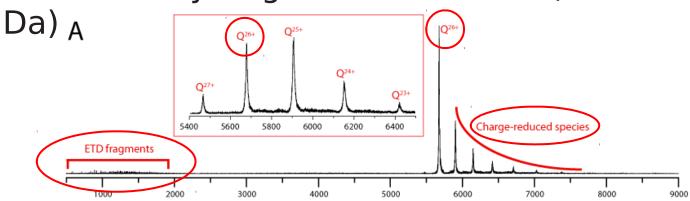


Sequence + PTMs



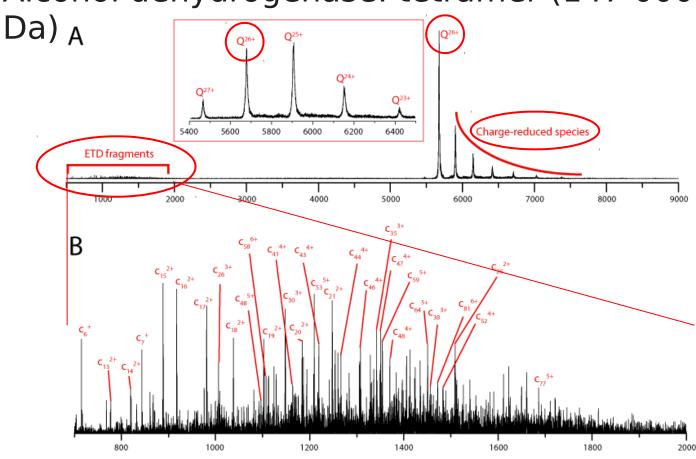


» Alcohol dehydrogenase: tetramer (147 000





Alcohol dehydrogenase: tetramer (147 000)





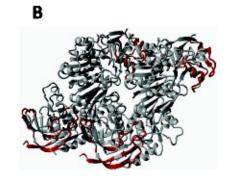
#### A

```
ac-SIPETOKGVI FYESHGKLEY KDIPVPKPKA NELLINVKYS GVCHTDLHAW HGDWPLPVKL
PLVGGHEGAG VVVGMGENVK GWKIGDYAGI KWLNGSCMAC EYCELGNESN CPHADLSGYT
HDGSFQEYAT ADAVQAAHIP QGTDLAEVAP VLCAGITVYK ALKSANLMAG HWVAISGAAG
GLGSLAVQYA KAMGYRVLGI DGGEGKEELF RSIGGEVFID FTKEKDIVGA VLKATDGGAH
GVINVSVSEA AIEASTRYVR ANGTTVLVGM PAGAKCCSDV FNQVVKSISI VGSYVGNRAD
TREALDFFAR GLIKSPIKVV GLSTLPEIYE KMEKGQIVGR YVVDTSK
```



A

AC-SIPETOKGVI FYESHGKLEY KDIPVPKPKA NELLINVKYS GVCHTDLHAW HGDWPLPVKL
PLVGGHEGAG VVVGMGENVK GWKIGDYAGI KWLNGSCMAC EYCELGNESN CPHADLSGYT
HDGSFQEYAT ADAVQAAHIP QGTDLAEVAP VLCAGITVYK ALKSANLMAG HWVAISGAAG
GLGSLAVQYA KAMGYRVLGI DGGEGKEELF RSIGGEVFID FTKEKDIVGA VLKATDGGAH
GVINVSVSEA AIEASTRYVR ANGTTVLVGM PAGAKCCSDV FNQVVKSISI VGSYVGNRAD
TREALDFFAR GLIKSPIKVV GLSTLPEIYE KMEKGQIVGR YVVDTSK



- Fragmentation occurs at surface of native tetramer (confirmed through calculations)
- » Current research: can we extend this to other proteins/complexes?



## Isotopic distributions in ETD-MS







### Isotopic distributions in MS

» Elements naturally occur as different isotopes (same # of protons, different # of neutrons)

» C: 98.93% 12C, 1.07% 13C

» H: 99.99% 1H, 0.01% 2H

» O: 99.76% 16O, 0.04% 17O, 0.20% 18O

» N: 99.63% 14N, 0.37% 15N

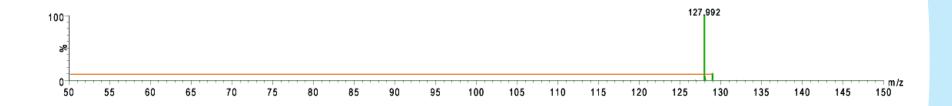
» S: 94.93% 32S, 0.76% 33S, 4.29% 34S, 0.02% 36S

Monoisotopic (lightest) peak, M+1, M+2, etc. determined by statistics



### Isotopic distributions in MS

» 1,4-dicyanobenzene (C8H4N2): chance of M+1  $\sim$  1 - 0.98938 = 8.2% IM+1/IM  $\sim$  8.2/98.93 = 8.3%

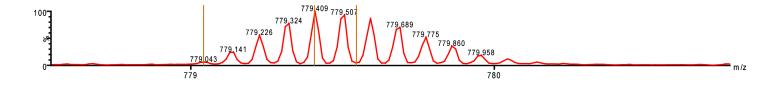


- » Monoisotopic peak = most abundant peak
- » Monoisotopic mass ~ average mass



### Isotopic distributions in MS

» Small protein (ubiquitin, 76 amino acids, 8.6 kDa): hundreds of  $C \rightarrow$  monoisotopic no longer most likely

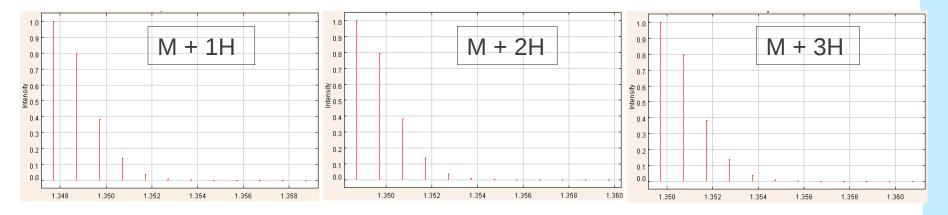


- » Monoisotopic peak != most abundant peak
- Monoisotopic mass !~ average mass



### Isotopic distributions in ETD-MS

- » In ESI-MS: measured mass = Mprotein + (n x MH)
  - » H: 99.99% 1H  $\rightarrow$  distribution same shape, shifted to the right
  - » Example: substance P (11 amino acids, 1.3 kDa)



- » Different charge states  $\rightarrow$  appear in different m/z regions
- » No problem as long as 'extra' H are protons H+



### Isotopic distributions in ETD-MS

- $^{*}$  ECD/ETD: H+ + e-  $\rightarrow$  H∎ (radical causes fragmentation)
  - » Potential problem!

$$^{\circ}$$
 [M + nH]n+ + A<sub>-</sub>-  $\rightarrow$  [c + xH]x+ [z + (n-x)H]<sub>-</sub>(n-x-1)+ + A ETD

$$| [M + nH]n + + A_{\bullet} - \rightarrow [M + (n-1)H] (n-1) + + HA$$
PTR
$$| [M + nH]n + + A_{\bullet} \rightarrow [M + nH]_{\bullet} (n-1) + + A$$
ETnoD

- » Contribution (probability) of each reaction not known
- » Mix of these reactions distorts isotopic distribution
  - » And it gets worse...



### ETD mechanism in detail

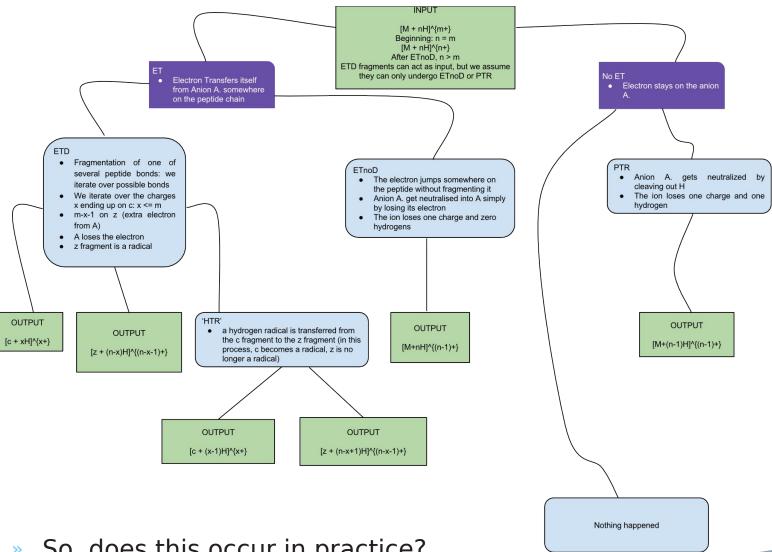


### ETD mechanism in detail

- Possible step 5: H
   migrates from c to z fragment
- [c + xH]x + [z + (n-x)H] (n-x-1) + + [c + (x-1)H]x + [z + (n-x+1)H] (n-x-1) +

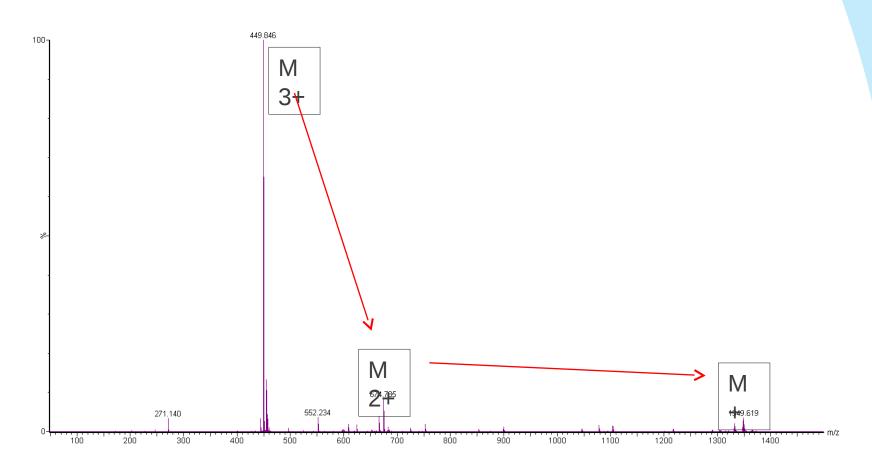


### ETD mechanism in detail



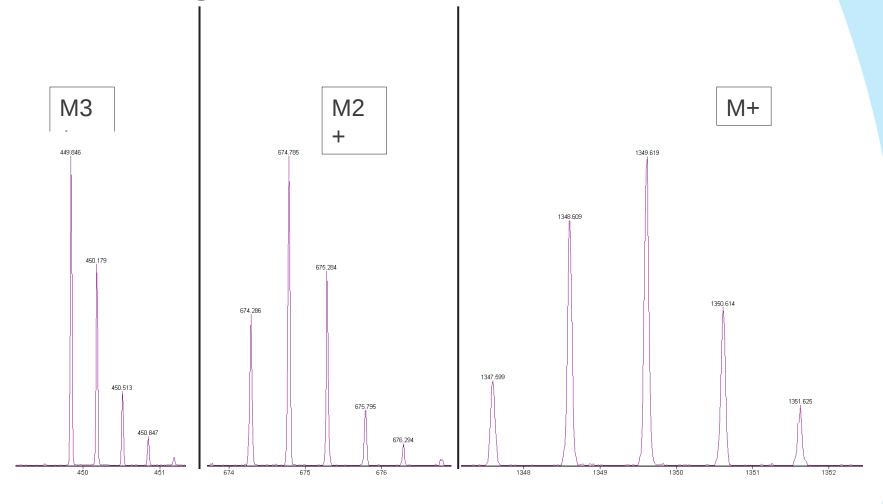
So, does this occur in practice?



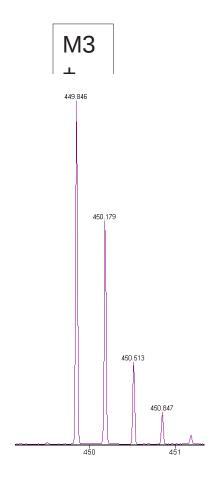


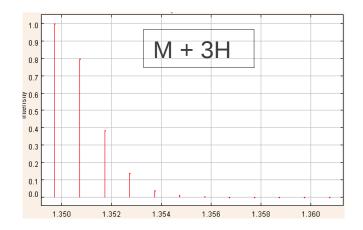
PTR or ETnoD? → Look at isotopic distribution



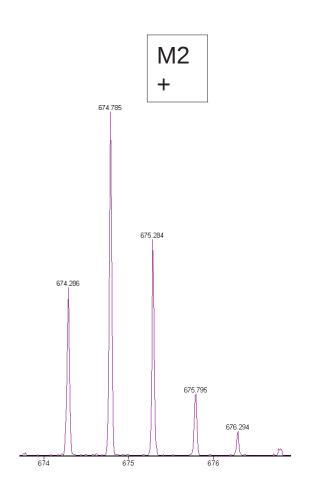


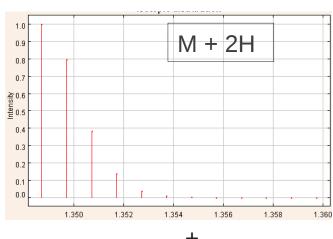


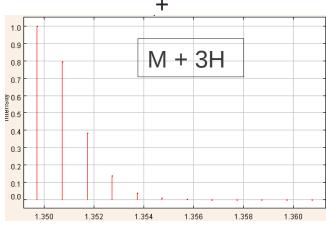




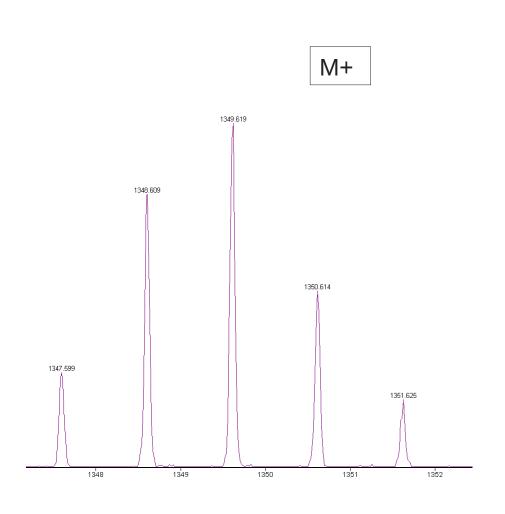




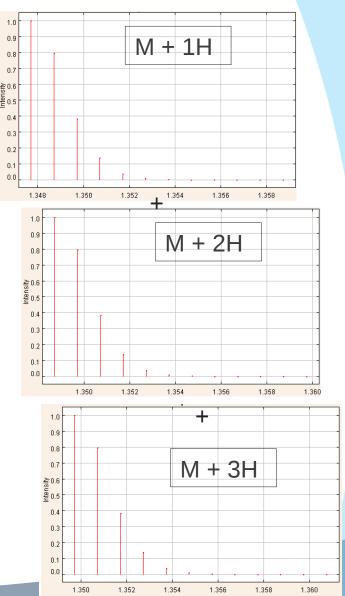






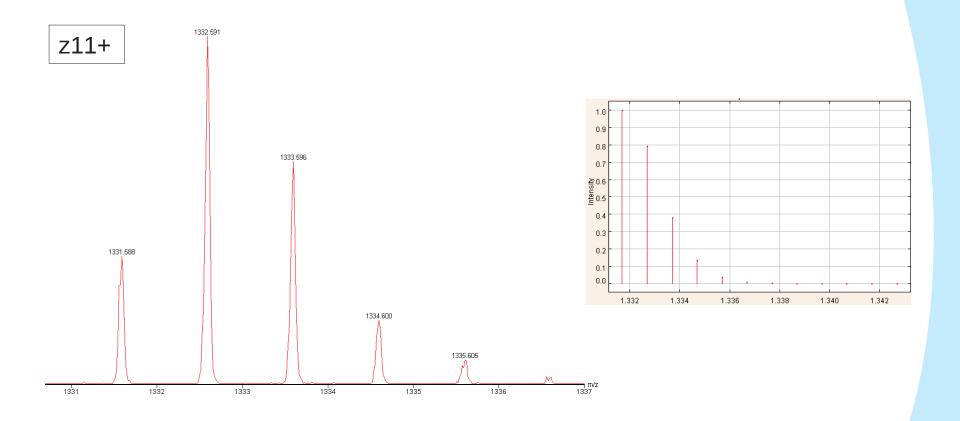


» Both PTR and ETnoD occur!



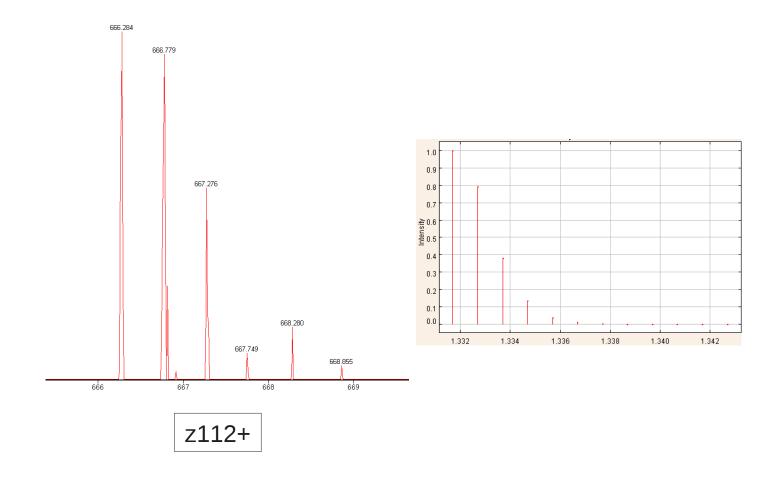


### ETD fragmentation of substance P





### ETD fragmentation of substance P





### Isotopic distributions in ETD-MS

- » Substance P (1.3 kDa): 3+ precursor used
  - » Maximum of 2 ET steps
- » Ubiquitin (8.6 kDa): 10 12+ precursors often used
- » ADH tetramer: (147 kDa): 26+ precursor used
  - » Most observed c fragments heavier than expected
- **»**
- » Can get very complex → bioinformatics required



### Goals of the project

- Deconvolute observed isotopic distributions and determine contribution of each component
- Learn how important each of the possible reactions is and gain insight into the (multistep) ETD process

