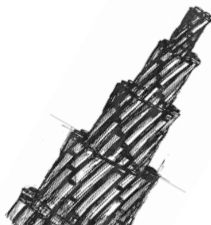


Modelling Cross-Linking in Collagen Fibrils

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Collagen Fibrils

- Collagen fibrils are a principal structural component of many animal tissues such as tendons, bones, cartilage, and corneas.
- A fibril is a nematic liquid crystal composed of tropocollagen molecules oriented along a director field \mathbf{n} .

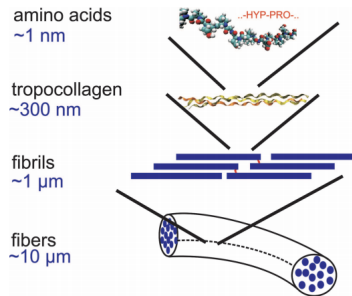


Figure: Buehler *et al.* *PNAS*, (2006).

Cross-linking

Collagen fibrils are held together by enzymatic cross-links (Lysyl Oxidase or LOX), spring-like molecules that connect two tropocollagen filaments at specific sites along their lengths.

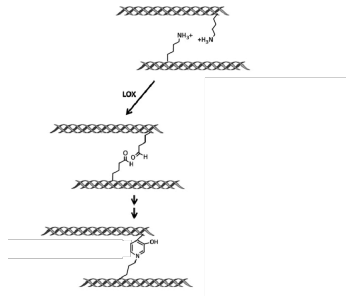


Figure: Perla-Kajan *et al.* *FASEB Journal*, (2016)

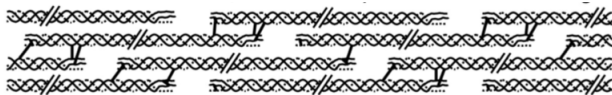


Figure: Kaku *et al.* *Journal of Prosthodontic Research*, (2014).

Question:

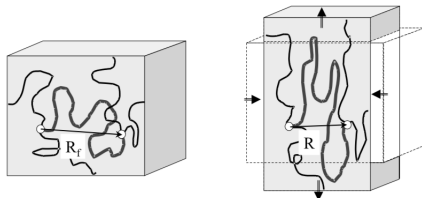
- How does the structure of a fibril determine its elastic properties?

Cross-links Modelled as Gaussian Springs

We model cross-links as entropic springs with end-to-end distance \mathbf{R} following the distribution:

$$P(\mathbf{R}) = \left[\left(\frac{3}{2\pi L} \right)^3 \frac{1}{\text{Det}[\underline{\ell}_0]} \right]^{1/2} \exp \left(-\frac{3}{2L} \mathbf{R}^\top \underline{\ell}_0^{-1} \mathbf{R} \right). \quad (1)$$

Here $\underline{\ell}_0$ is a tensor describing the structure of the fibril and its cross-links. We define $\zeta = \ell_{\parallel}/\ell_{\perp}$ to be the anisotropy ratio.



[1] Warner, M., and Terentjev, E. M. "Nematic elastomers – a new state of matter?" *Progress in Polymer Science* (1996).

The free energy density for a cross-linked liquid crystal elastomer subjected to a strain field $\underline{\underline{\lambda}}$ is:

$$\boxed{f_{\text{Cross-Link}} = \frac{1}{2} \rho k_B T \text{Tr}(\underline{\underline{\ell}}_0 \underline{\underline{\lambda}}^\top \underline{\underline{\ell}}^{-1} \underline{\underline{\lambda}}).} \quad (2)$$

Here ρ is the (assumed to be constant) cross-linking density, and $\underline{\underline{\ell}}$ is a tensor describing the post-strain fibril structure.

$$\text{Minimize } F = \frac{2}{R^2} \int_0^R r f dr \quad \xRightarrow{\text{Numerically}} \quad \begin{cases} \text{Post-strain fibril structure } \underline{\underline{\ell}} \\ \text{Stress field } \underline{\underline{\sigma}} \left(\sigma_{ij} = \frac{\partial F}{\partial \lambda_{ij}} \right) \end{cases}$$

$\underline{\underline{\ell}}_0$: Double Twist Structure

Our ansatz:

$$\mathbf{n}_0 = -\sin \psi(r) \hat{\phi} + \cos \psi(r) \hat{z}. \quad (3)$$

Then we have

$$\underline{\underline{\ell}}_0 = \underline{\underline{\delta}} + [\zeta - 1] \mathbf{n}_0 \otimes \mathbf{n}_0. \quad (4)$$

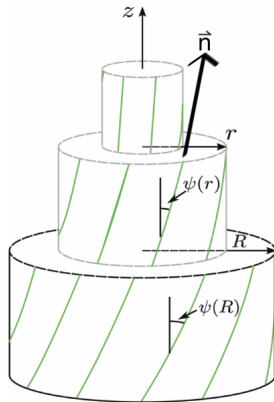


Figure: Cameron, Kreplak, and Rutenberg. *Soft Matter*, (2018).

$\ell_{=0}$: Imposed Twist Phases

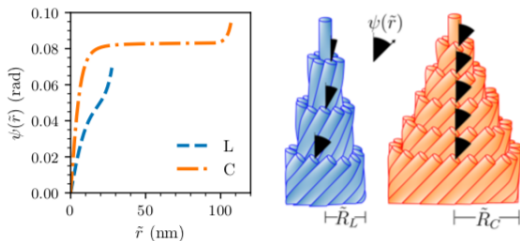


Figure: Cameron, Kreplak, and Rutenberg. *Unpublished*, (2019).

We consider two functional forms (phases) for $\psi(r)$:

- $\psi(r) = \alpha r$ (linear twist phase) (e.g. Tendon Fibrils),
- $\psi(r) = \beta \cdot \mathbb{1}_{[\frac{1}{10}R, R]}$ (constant twist phase) (e.g. Corneal Fibrils).

λ : The Strain Field

We take as our strain tensor ($\mathbf{R} = \underline{\underline{\lambda}}\mathbf{R}_0$) a volume-preserving longitudinal extension with a shear-twist around the fibril axis:

$$\underline{\underline{\lambda}} = \begin{bmatrix} \frac{1}{\sqrt{\epsilon}} & 0 & 0 \\ 0 & \frac{1}{\sqrt{\epsilon}} & \kappa \\ 0 & 0 & \epsilon \end{bmatrix}. \quad (5)$$

Equilibrium Twist Angle

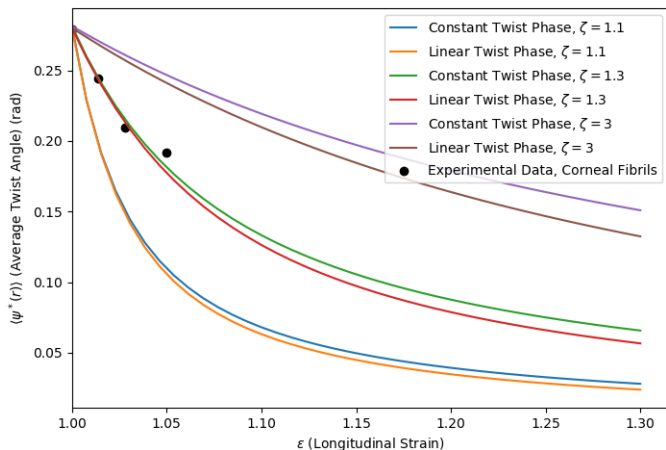


Figure: Data from Bell *et al. Acta Biomaterialia*, (2017).

Stress-Strain Curves

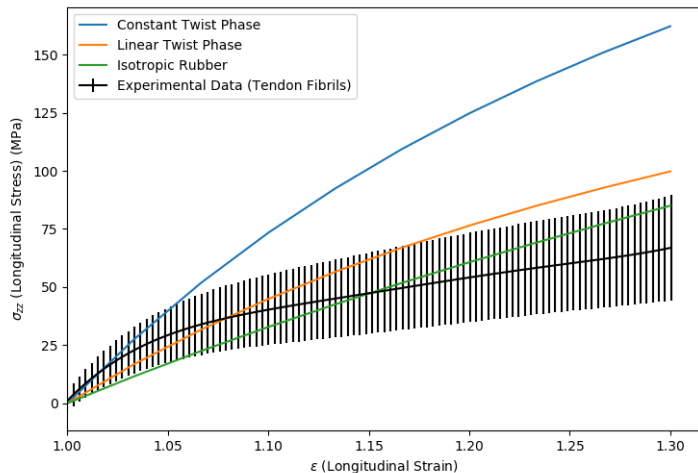


Figure: Data from Quigley, ..., and Kreplak. *Scientific Data*, (2018).

Thanks!

Form of the Anisotropy Tensor

$$\begin{aligned}\underline{\underline{\ell}}_0 &= \ell_{\perp} \underline{\underline{\delta}} + [\ell_{\parallel} - \ell_{\perp}] \mathbf{n}_0 \otimes \mathbf{n}_0 \\ &= \ell_{\perp} \begin{bmatrix} 1 & 0 & 0 \\ 0 & 1 + (\zeta - 1) \sin^2 \psi & (1 - \zeta) \sin \psi \cos \psi \\ 0 & (1 - \zeta) \sin \psi \cos \psi & 1 + (\zeta - 1) \cos^2 \psi \end{bmatrix},\end{aligned}$$

where $\zeta = \ell_{\parallel} / \ell_{\perp}$.

(6)

$$\begin{aligned}\underline{\underline{\ell}}^{-1} &= 1/\ell_{\perp} \underline{\underline{\delta}} + [1/\ell_{\parallel} - 1/\ell_{\perp}] \mathbf{n} \otimes \mathbf{n} \\ &= \ell_{\perp}^{-1} \begin{bmatrix} 1 & 0 & 0 \\ 0 & 1 + (\zeta^{-1} - 1) \sin^2 \psi^* & (1 - \zeta^{-1}) \sin \psi^* \cos \psi^* \\ 0 & (1 - \zeta^{-1}) \sin \psi^* \cos \psi^* & 1 + (\zeta^{-1} - 1) \cos^2 \psi^* \end{bmatrix},\end{aligned}$$

where ψ^* is the post-strain twist angle.

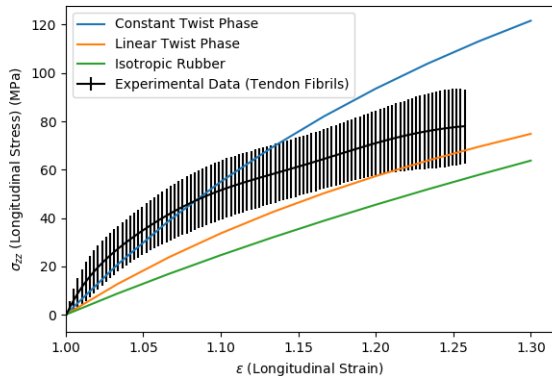
Frank Free Energy

The Frank free energy describes the energy contributions from the spatial arrangement of the tropocollagen filaments inside the fibril. For our double-twist director field $\mathbf{n}(\psi)$, the Frank free energy density is:

$$\begin{aligned} f_{\text{Frank}} = & \frac{1}{2} K_{22} \left(\frac{K_2}{K_{22}} - \psi' - \frac{\sin(2\psi)}{2r} \right)^2 \\ & + \frac{1}{2} K_{33} \frac{\sin^4(\psi)}{r^2} \\ & - \frac{1}{2} (K_{22} + K_{24}) \frac{1}{r} \frac{d}{dr} (\sin^2(\psi)) . \end{aligned} \tag{7}$$

Here $K_2, K_{22}, K_{24}, K_{33}$ are the standard Frank elastic constants.

More Stress Strain Curves



Experimental Data

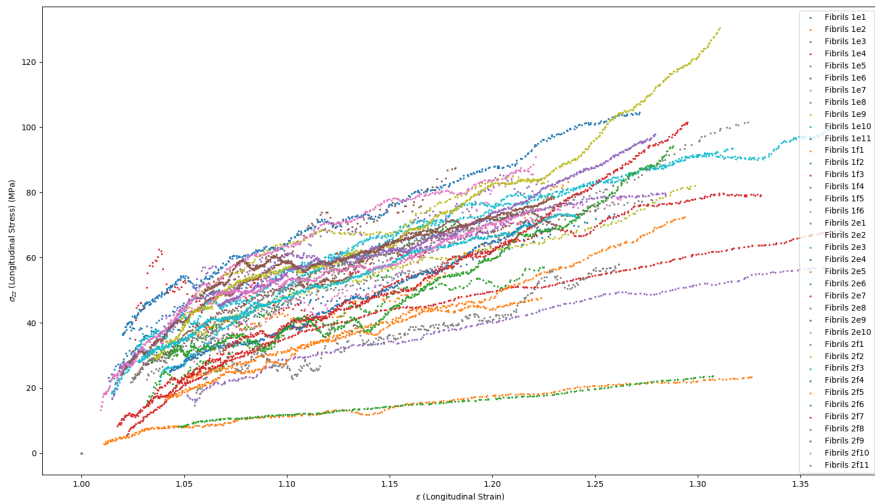


Figure: Data from Quigley, ..., and Kreplak. *Scientific Data*, (2018).