# Title

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# Abstract

# Introduction

Why nutrient cycling in lakes is important globally (lakes as integrators of the landscape, lakes as hotspots of carbon cycling, small lakes disproportionately active in carbon cycle, lakes as sentinels of change)

Description of study sites

How can metagenomic/genome analysis shed light on ecosystems? Or how much did we not know or take for granted about how microbes cycle C? consistent with textbooks or surprises?

Acknowledge other papers that have use genomic data (Eiler fresh vs salt, bogs) much previous work done gene-centric vs genome centric

# Results/Discussion

## Overview of Dataset

Analyzing the genomes of uncultured microbes can provide insight into the potential metabolic functions of those organisms. 205 bacterial metagenome assembled genomes (MAGs) were recovered from a metagenomic time series in Trout Bog and Lake Mendota as described in Bendall, et al (Bendall et al. 2016). These MAGs range in completeness from 50 to 99% complete, and passed quality checks for contamination using CheckM (Parks et al. 2015) (supp table). Of the 205 MAGs, 102 were recovered from Lake Mendota, 31 were recovered from the epilimnion of Trout Bog, and 69 were recovered from the hypolimnion of Trout Bog. Several MAGs in the epilimnion and hypolimnion of Trout Bog appeared to be from the same population based on high average nucleotide identities (supp table). The phylogenetic distribution of MAGs was consistent with the classifications of 16S ribosomal rRNA gene amplicon sequencing results (Figure 1). These results are consistent with other 16S-based studies in these sites (Hall et al. 2017; Linz et al. 2017).

<Fig 1a Barchart of MAG phyla>

<Fig 1b Barchart of 16S phyla>

**Figure 1. How representative are the MAGs of the microbial community?** The taxonomic classifications of MAGs (A) reflect the community composition observed via 16S rRNA ribosomal amplicon sequencing (B).

## Photosynthesis and Carbon Fixation

Primary production is a critical component of the carbon cycle in lakes. Therefore, we looked at potential routes of primary production within the microbial community, expecting to find differences between our two ecosystems. In Lake Mendota, MAGs classified as Cyanobacteria comprised the majority of photoautotrophs in the dataset. These populations contained genes encoding enzymes in the Calvin-Benson-Bassham (CBB) pathway. In Trout Bog, genomes appearing to be from photoautotrophic organisms were classified as *Chlorobium clathratiforme*, a species of Chlorobiales widespread in humic lakes (Karhunen et al. 2013). The Chlorobiales MAGs in Trout Bog contained genes encoding citrate lyase and other key enzymes in the reductive TCA cycle, an alternative carbon fixation method to the CBB pathway commonly found in green sulfur bacteria (Kanao et al. 2002; Tang and Blankenship 2010). The primary producers Cyanobacteria and Chlorobiales seem to perform similar ecosystem functions, such as nitrogen and carbon fixation, in their respective lakes; however, oxygen availability drives both the type of microbe acting in this role and the pathways that it uses for primary production.

Marker genes for anoxygenic photosynthesis were identified in several other MAGs of Burkholderiales from both lakes (classified as groups such as *Polynucleobacter necessarius*, *Lautropia*, and *Albidoferax*) (Martinez-Garcia et al. 2012). However, genes potentially encoding carbon fixation were identified in only one of these MAGs, sequenced from Lake Mendota and classified as Burkholderiales.

Another form of harvesting sunlight for energy in freshwater is the use of light-activated proteins such as rhodopsins. Rhodopsins were observed in many phylogenetically diverse MAGs in both Trout Bog and Lake Mendota, and have been the subject of further study (cite Shaomei’s preprint when it comes out).

## Carbon Degradation

Carbon in lakes can either be produced in the water column (autochthonous) or received from the surrounding terrestrial landscape (allochthonous). To further understand bacterial carbon degradation in lakes, we identified and categorized putative carbon degradation pathways in our MAGs. We hypothesized that we would observe differences in carbon degradation pathways between lakes due to their contrasting landscapes and their unique primary producers.

Biopolymers in freshwater can be allochthonous or autochthonous in origin. Two common biopolymers, cellulose and chitin, are produce on land and in the water column, respectively. In both lakes, Bacteroidetes and Verrucomicrobia MAGs contained genes encoding cellulases, chitinases, and glucoside hydralases, without appearing to specialize in autochthonous or allochthonous carbon. Glucoside hydralase – encoding genes were also identified in Planctomycetes in Lake Mendota and in Burkholderiales, Actinobacteria, and Methylococcales in Trout Bog. Genes relating to the degradation of cellobiose and chitobiose, breakdown products of cellulose and chitin, were common in many taxa from both lakes. Degradation of phenol and salicylate, two aromatic compounds derived from terrestrial carbon sources, were potentially identified in MAGs of Burkholderiales in both lakes.

Although eukaryotic genomes were not included in this analysis, eukaryotic algae are known photoautotrophs in both lakes (Descy et al. 2000; Hurley and Armstrong 1990) Algae produce amino acids, carbohydrates, and carboxylic acids that fuel growth of the heterotrophic community (Salcher, Posch, and Pernthaler 2013). We observed many MAGs in both Lake Mendota and Trout Bog containing putative pathways for the degradation of carbohydrates such as glucose, galactose, maltose, rhamnose, mannose, and xylose. These compounds are all documented algal exudates in freshwater (Giroldo, Augusto, and Vieira 2005; Juttner and Matuschek 1977). They can also be derived from the breakdown of biopolymers; leaky extracellular degradation of biopolymers may result in these sugars being made available to community members without the ability to break down biopolymers. Degradation of additional sugars involved in galactose metabolism (sucrose, stachyose, raffinose, trehalose, lactose, and melibiose) were identified in MAGs classified as Bacteroidetes, Verrucomicrobia, and Actinobacteria from Trout Bog, but not from Lake Mendota.

One type of carbon degradation is methylotrophy, the ability to grow solely on one carbon compounds such as methane, methanol, formaldehyde, or methylamines. Multiple MAGs classified as well-studied methylotrophs Methylococcales and Methylophilaceae contained genes for methylotrophic pathways in both lakes (Kalyuzhnaya et al. 2012; Salcher et al. 2015) (Figure 3). Lake Mendota additionally had MAGs containing potential methylotrophs belonging to Planctomyces and Rhodocyclaceae, while additional potential methylotrophs in Trout Bog included Burkholderiales, Rhizobiales, Nitrosomonadales, Geobacteraceae, and Solirubrobacterales. Given the rapid rate at which the known diversity of methylotrophs is increasing, this finding is not surprising (Chistoserdova, Kalyuzhnaya, and Lidstrom 2009). The methylotrophs Methylobacter and Methylotenera have been observed to exchange carbon cooperatively, perhaps outcompeting other community members via denitrification (Beck et al. 2013). This, along with evidence from our MAGs suggesting involvement in sulfur cycling, points to an ecological role for methylotrophs such as these at the intersection of multiple nutrient cycles.

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Description generated with high confidence

**Fig 2. Carbon cycling in Lake Mendota vs Trout Bog.** Carbon cycling between the two lakes was largely similar, with a few key differences. Carbon fixation is carried out by different taxa with different pathways. MAGs from Trout Bog tend to over more genes encoding enzymes in galactose metabolism than those from Lake Mendota.

**A close up of a map

Description generated with high confidenceFigure 3. Methylotrophy in freshwater.** *Methylococcaceae* and *Methylotenera* are taxa capable of methylotrophy in both Lake Mendota and Trout Bog. While both are likely capable of subsisting only on one carbon compounds, both have the potential to utilize other carbon sources. This, along with their putative pathways for assimilatory sulfate reduction and Methylococcaceae’s potential for denitrification and nitrogen fixation, place these populations at the intersection of multiple nutrient cycles.

## Nitrogen Cycling

While carbon cycling was relatively similar between lakes, the drastically different concentrations of nitrogen in Trout Bog versus Mendota led us to hypothesize that steps in the water column nitrogen cycle may be altered between these two systems. One key difference we found in the MAGs was that in Mendota, very few MAGs had genes encoding nitrogen fixation, and they belong mainly to Cyanobacteria. Conversely, more MAGs in Trout Bog contained these genes, and they were in phylogenetically diverse populations. Genes annotated as nitrate and nitrite reductases, key enzymes in denitrification, were found in MAGs from both lakes. However, nitrate reductases were far less common than nitrite reductases in Lake Mendota (19 vs 53, respectively), and found primarily in Cyanobacteria. This difference was not as pronounced in Trout Bog. Genes annotated as ammonia monooxygenase were not found in MAGs from either lake, aside from the ammonia/methane monooxygenases found in MAGs classified as putative methanotroph Methylococcales, which are likely not involved in nitrogen cycling. No genes potentially encoding the anammox pathway were identified any of the MAGs. Genes encoding steps in the urea cycle or ammonia assimilation were found in nearly every MAG.

Genes potentially encoding the biosynthesis, degradation, and transport of the polyamines and non-proteinogenic amino acids putrescine, spermidine, and canavanine were widespread in both lakes. While the importance of polyamines as nitrogen sources for bacteria has been established in coastal marine systems (Mou et al. 2011), these compounds have been less studied in freshwater. Our results lend support to the hypothesis that these compounds are important parts of the dissolved organic nitrogen pool in freshwater. Polyamines play a critical but poorly understood role in bacterial metabolism (Igarashi and Kashiwagi 1999), and the exchange of these nitrogen compounds may be a factor structuring freshwater bacterial communities.

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**Fig 3A. Nitrogen cycling in Trout Bog vs Lake Mendota.** The numbers of genes relating to nitrogen cycling in the MAGs were analyzed by lake. Genes annotated as nitrogenase subunits were more common in Trout Bog than Lake Mendota. Nitrite reductases were observed more frequently than nitrate reductases in both lakes, but this difference was more pronounced in Lake **A close up of a map

Description generated with high confidence**Mendota. Polyamine transport was widespread in both lakes.

**Fig 3B. Nitrogen cycling in Trout Bog vs Lake Mendota.** The potential to fix nitrogen was confirmed in MAGs by searching for operons containing multiple nitrogen fixation – related genes. Trout Bog had greater numbers of MAGs that met this condition than Lake Mendota. The putative nitrogen fixers in Lake Mendota were all Cyanobacteria, while putative nitrogen fixers in Trout Bog were more diverse.

## Sulfur Cycling

Sulfur is another important element in freshwater. Because measurements of sulfur species measurements were not available for either lake, we did not predict any differences in sulfur cycling between lakes. We found that, as with carbon cycling, the identity and numbers of genes associated with steps in the sulfur cycle were similar between Lake Mendota and Trout Bog. Sulfate reduction genes dominated over sulfide oxidation genes in both systems, presumably reflecting the ecological importance of oxidation vs reduction. Genes potentially encoding assimilatory sulfate reduction were far more common than those potentially encoding dissimilatory sulfate reduction. This indicates that sulfate is likely more often used as a building block in biosynthesis rather than as a terminal electron acceptor. Genes for sulfur cycling were identified in diverse MAGs in both lakes.

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**Fig 4. Number of MAGs containing sulfur metabolism genes by lake.** Sulfate reduction dominates over sulfide oxidation in both ecosystems. By the number of MAGs encoding putative enzymes for each pathway, assimilatory sulfate reduction is favored over dissimilatory sulfate reduction. These results likely reflect the availability of sulfur species in Trout Bog and Lake Mendota.

## Unusual microbes

Although our primary goal was to use genome content to investigate differences in nutrient cycling between lakes, we recovered the genomes of unusual microorganisms in this dataset and report their genome content here. One MAG from Elusimicrobiales was recovered from Trout Bog. While this genome is only 44% complete, we can propose that it uses sugars such as maltooligosaccharides, maltose, and arabinogalactan as a carbon source. This population of Elusimicrobiales likely reduces sulfate via the assimilatory pathway. It also contains one nitrogenase subunit, suggesting that it may be capable of fixing nitrogen.

Thirteen MAGs classified as Planctomycetes were recovered from Lake Mendota. While some Planctomycetes isolates have been known to perform anammox, no genes encoding the enzymes necessary for this pathway were found in the Lake Mendota MAGs. The Mendota Plantcomycetes MAGs contained genes encoding glucoside hydrolases and sulfatase enzymes. These pathways could potentially be used to break down complex sulfur-containing polysaccharides, such as those produced by Cyanobacteria or eukaryotic algae. Similar genes were found in Planctomycetes genomes in a freshwater algal biofilm (cite when published). This suggests a new potential ecological role for Planctomycetes in freshwater as a motile specialist in polysaccharide degradation.

A picture containing text, map

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**Figure 5. Proposed metabolisms of Planctomycetes in Lake Mendota.** Several MAGs classified as Planctomycetes were recovered from Lake Mendota. The observation of genes annotated as glycoside hydrolases and sulfatases suggest a role for Planctomycetes as algal polysaccharide degraders. These populations also appear to consume saccharides such as galactose, fructose, and mannose, and synthesize both spermidine and putrescine. The presence of genes encoding flagellar and chemotaxis proteins suggests that they are motile.

## Conclusions

Analysis of gene content and predicted pathways in our MAGs reveals potential similarities and differences in the ecology of bacteria in two freshwater lakes. Surprisingly, predicted pathways involved in carbon cycling were often similar between Lake Mendota and Trout Bog. However, exceptions included differences in the identity and carbon fixation pathways of primary producers, and the greater numbers of genes encoding enzymes involved in galactose metabolism in Trout Bog. Sulfur cycling was also highly similar between these two lakes, with both indicating a bias towards reduction over oxidation, and furthermore towards assimilatory over dissimilatory reduction. Nitrogen cycling did appear altered between the lakes, with the greater levels of nitrogen fixation (and diversity of nitrogen fixers) in Trout Bog compared to Lake Mendota in concordance with their known nitrogen concentrations. Polyamines and nonproteinogenic amino acids appear to be important sources of nitrogen in both lakes. Finally, comprehensive analysis of individual MAGs can suggest ecological roles for uncultured organisms, such as polysaccharide degradation for Lake Mendota’s Planctomycetes, saccharide degradation for Trout Bog’s Elusimicrobiales, and a key position at the intersection of carbon, nitrogen, and sulfur cycling for methylotrophs in both lakes.

The insights gained from this study provide hypotheses for further testing of bacterial nutrient cycling in freshwater through metatranscriptomics and chemical assays. Additionally, genomes from this study can be compared to those sequenced from other lakes and environments. We hope that this will serve as a resource to other researchers with similar genomes and datasets.

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