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Physcraper: A Python package for continually updated phylogenetic trees using the Open Tree of Life

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Abstract

Background: Phylogenies are a key part of research in many areas of biology. Tools that automate some parts of the process of phylogenetic reconstruction, mainly molecular character matrix assembly, have been developed for the advantage of both specialists in the field of phylogenetics and non-specialists. However, interpretation of results, comparison with previously available phylogenetic hypotheses, and selection of one phylogeny for downstream analyses and discussion still impose difficulties to one that is not a specialist either on phylogenetic methods or on a particular group of study.

Results: Physcraper is a command-line Python program that automates the update of published phylogenies by adding public DNA sequences to underlying alignments of previously published phylogenies. It also provides a framework for straightforward comparison of published phylogenies with their updated versions, by leveraging upon tools from the Open Tree of Life project to link taxonomic information across databases. The program can be used by the nonspecialist, as a tool to generate phylogenetic hypotheses based on publicly available expert phylogenetic knowledge. Phylogeneticists and taxonomic group specialists will find it useful as a tool to facilitate molecular dataset gathering and comparison of alternative phylogenetic hypotheses (topologies).

Conclusions: The Physcraper workflow showcases the benefits of doing open science for phylogenetics, encouraging researchers to strive for better sharing practices. Physcraper can be used with any OS and is released under an open-source license. Detailed instructions for installation and usage are available at <https://physcraper.readthedocs.io>.

Keywords: gene tree; interoperability; open science; reproducibility; public database; DNA alignment

Background

Phylogenies capture the shared history of organisms and provide key evolutionary context for our biological observations [1]. Updating existing phylogenies with publicly available molecular sequence data ~~that has never been incorporated into any phylogenetic estimate~~ provides the opportunity to study the evolutionary history of many taxa in a reproducible and continuous manner. Here, we introduce Physcraper, a tool that establishes a data interoperability framework for biological databases to automate data connections across databases, ~~with the main goal of building on published alignments and extending~~. Physcraper's main goal is to build upon published alignments to extend existing phylogenetic inferences with more data and

taxa, [which improve phylogenetic reconstruction \[2, 3\]](#), [time of divergence estimates \[4, 5\]](#), [biogeographic analyses \[6\]](#), [as well as help in resolving phylogenetic conflict \[7, 8, 3\]](#). Physcraper updates a starting ~~tree and single locus alignments~~ [single locus alignment and corresponding tree](#) with public DNA data, and links the tips in ~~these~~ [the updated](#) trees to a unified, interoperable taxonomic resource [9].

Data such as geographical location, fossil ranges, and genetic and phenotypic information increasingly available in public databases constitute an amazing resource for biological discovery [10]. One of the main challenges for automatic integration of biological data across databases are varying taxonomic idiosyncrasies. To address this challenge, the Open Tree of Life project (OpenTree) created a unified taxonomy for name standardization, by integrating taxonomic data from several databases [9], including the USA National Center for Biodiversity Information (NCBI) taxonomy [11, 12], and the Global Biodiversity Information Facility (GBIF) [13] among others. By using the existing OpenTree taxonomy programmatic tools to map tip names, Physcraper has a framework for connecting updated phylogenies with data from any biological database.

Decades of single locus sequencing have generated massive amounts of homologous DNA datasets that have the potential to be used for phylogenetic reconstruction at many scales [14]. More than a decade ago, GenBank release 159 (April 15, 2007) already hosted 72 million DNA sequences that were gauged to have the potential to resolve phylogenetic relationships of 98.05% of the almost 241,000 distinct taxa in the NCBI taxonomy at the time [14]. However, even thirteen years later, phylogenetic estimates for ~~many-most~~ of these taxa are [still](#) not available [15]. ~~OpenTree's comprehensive~~ [The Open Tree of Life project assembles a comprehensive synthetic tree of life](#) ~~comprises-comprising~~ 2.3 million tips, of which around [only](#) 90,000 are supported by ~~phylogenies~~ [phylogenetic data](#) - the remaining 1.4 million taxa are placed in the tree based on taxonomy. There is a considerable amount of phylogenetically informative data in GenBank with the potential to fill these phylogenetic gaps in the tree of life, but this data either has not been analysed or the analyses have not been made publicly available [15].

Assembling a DNA alignment from such a massive database as GenBank can be done "by hand", but that is a time consuming approach which is not highly reproducible. A variety of computational pipelines that mine DNA databases fast, efficiently, and reproducibly have been developed and used to infer phylogenetic relationships in a variety of organisms (e.g., [16, 17, 18, 19]). While genomics has, and will continue to, revolutionize phylogenetic inference, the diversity of alternative genomic sequencing approaches implemented produce largely non-overlapping homology hypotheses across taxa [20], creating challenges for phylogenetic reconstruction. Phylogenomics addresses this problem by focusing on targeted capture of informative regions [21]. However, fine-grained curated markers and alignments can significantly improve phylogenetic reconstructions, even in phylogenomic analyses [22].

Physcraper improves on previous work in automating phylogenetic reconstruction by leveraging the power of existing homology hypotheses that taxon specialists have assessed and deemed appropriate for a specific phylogenetic scope. There are almost 8,200 publicly available, peer-reviewed curated alignments, covering around 100,000

distinct taxa in the TreeBASE database [23], which can be leveraged as seeds to mine molecular databases, and as “jump-start” alignments for phylogenetic reconstructions [24] to continually enrich, update and compare existing phylogenetic knowledge.

Physcraper is implemented as a Python pipeline that uses OpenTree’s programmatic access protocols (API’s) to automatically link any phylogeny mapped to OpenTree’s standardized taxonomy [25], to alignments from TreeBASE [26], and data from GenBank [27]. Its utility and functionalities are presented with a case-study analysis of a group of flowering plants, the hollies.

Implementation

Physcraper is implemented with Python and can also be run on the command line. It currently consists of 12 modules. For testing and improving the quality of the Python code syntax in the Physcraper code, we used the Pylint [28, 29] software with a configuration file. We improved the code syntax on modules with low Pylint scores and fixed errors by following Pylint’s recommendations. Based on Physcraper’s software design choices, some of Pylint’s recommendations were overruled by using its check-disabling system, and are explained in the code. As of now, all modules have a Pylint score of 10/10.

The general Physcraper framework (Figure 1) consists of 4 steps: 1) identifying and processing a tree and its underlying alignment; 2) performing a BLAST search of DNA sequences from original alignment on GenBank, and filtering of new sequences; 3) profile-aligning new sequences to original alignment; 4) performing a phylogenetic analysis and comparing the updated tree to existing phylogenies.

The inputs: a tree and an alignment

Taxon names in the input tree must be standardized to OpenTree taxonomy [9] using OpenTree’s bulk Taxonomic Name Resolution Service (TNRS) tool [30]. Users can upload their own tree, or choose from among the 2, 950 standardized trees stored in OpenTree’s Phylsystem [31, 25] that also have alignments available on TreeBASE [23].

The input alignment is a single locus DNA dataset that was used in part or in whole to generate the input tree. Physcraper retrieves TreeBASE alignments automatically. Alternatively, users must provide the path to a local copy of the alignment. Only taxa that are both in the sequence alignment and in the tree are considered further for analysis; at least one taxon and its corresponding sequence are required.

DNA sequence search and filtering

The Basic Local Alignment Search Tool, BLAST [32] is used for DNA sequence search on a remote or local GenBank database. It is constrained to a “search taxon”, a taxonomic group in the NCBI taxonomy that is automatically identified using the OpenTree’s taxonomic Most Recent Common Ancestor (MRCA) API [33, 9], as the MRCA of all ingroup taxa that is also a named clade in the NCBI taxonomy (Figure 1).

BLAST is performed using the ‘blastn’ algorithm [34] implemented in BioPython’s [35] NCBIWWW module [36] modified to accept an alternative BLAST address.

Each sequence in the alignment is BLASTed once against all DNA sequences in GenBank. New sequences are excluded for analysis if they 1) are not in the search taxon; 2) have an e-value above the cutoff (default to 0.00001); 3) fall outside a min and max length threshold, defined as the proportion of the average length without gaps of all sequences in input alignment (default values of 80 respectively); 4) or if they are either identical to or shorter than an existing sequence in the input alignment and they represent the same taxon in OpenTree or NCBI taxonomy. An arbitrary maximum number of randomly chosen sequences per taxon are allowed (default to 5).

Reverse, complement, and reverse-complement sequences are identified and translated using BioPython internal functions [35]. Iterative cycles of BLAST searches can be performed, by blasting all new sequences until no new ones are found. By default only one BLAST cycle is performed.

New DNA sequence alignment

MUSCLE [37] is used to perform a profile alignment in which the original alignment is used as a template of homology criteria to align new sequences. The final alignment is not further automatically checked, and additional inspection and refinement are recommended.

Tree reconstruction and comparison

RAxML [38] is implemented to reconstruct a Maximum Likelihood (ML) gene tree for each input alignment with default settings (GTRCAT model and 100 bootstrap replicates with default algorithm), using input tree as starting tree for ML searches. Bootstrap results are summarized using DendroPy's SumTrees module [39].

Physcraper's main result is an updated phylogenetic hypothesis for the search taxon. Updated and original tree are compared with Robinson-Foulds weighted and unweighted metrics ~~estimated~~ calculated with Dendropy [39], and with a node by node comparison between the synthetic OpenTree and original and updated tree individually, using OpenTree's conflict API [40].

Results

Case Study: The hollies

A user is interested in phylogenetic relationships within the genus *Ilex*. Commonly known as "hollies", the genus encompasses between 400-700 living species, and is the only extant clade within the family Aquifoliaceae, order Aquifoliales of flowering plants.

An online literature review in June 2020 (Google scholar search for "ilex phylogeny") reveals that there are several published phylogenies showing relationships within the hollies [41, 42, 43, 44], but only two have data publicly available [45, 46]. [45] made original tree and alignment available in TreeBASE (study 1091 [47]). ~~The tree sampling~~ This tree (Gottlieb2005 tree from now on) samples 41 species ~~is also~~, is available from OpenTree's Phylesystem (study pg_2827 [48]), and has been integrated into OpenTree's synthetic tree [49]. The most recent *Ilex* tree [46] (Yao2020 tree from now on) is available in OpenTree's Phylesystem (study ot_1984 [50]), and in the DRYAD repository [51]. With 175 tips, the ~~[46] tree~~ Yao2020 tree [46] is

the best sampled phylogeny ~~yet~~-available for the ~~hollies~~-genus *Ilex*. In order to showcase Physcraper's performance, we updated the tree with the smallest number of tips (the Gottlieb2005 tree) with the expectation to use the tree with the largest number of tips (the Yao2020 tree) as a standard for comparison and verification of Physcraper's results.

We ran Physcraper on a laptop Linux computer to update an internal transcribed spacer DNA region (ITS) alignment ~~that was used to construct the tree~~ from [45], using a local GenBank database. BLAST and RAxML analyses ran for 19hrs 45min, with bootstrap analyses taking an additional 13hrs. The ~~updated [45] tree~~ ~~Gottlieb2005 tree~~ [45] updated using Physcraper (Figure 2; ~~Physcraper updated tree from now on~~) displays all 41 distinct taxa from the original study plus 231 new tips, contributing phylogenetic data to 84 additional *Ilex* taxa. The best ~~RaxML tree~~ ~~ML tree from the RAxML analysis~~ is 99% resolved, with 25% of nodes with bootstrap support < 0.1 and 48% nodes with bootstrap support > 0.75. A large portion of internal branches are negligibly small, with 30 branches < 0.00001 substitution rate units, from which only 9 have a bootstrap support > 0.75 (Figure 2). ~~For comparison,~~ ~~As comparison with the Physcraper updated tree,~~ ~~the Yao2020 tree~~ [46] also contains all 41 distinct taxa ~~from the original [45] study,~~ ~~and contributes~~ ~~sampled in the Gottlieb2005 tree~~ [45], while contributing phylogenetic data to 134 additional *Ilex* taxa, ~~from which,~~ ~~From these,~~ 67 taxa are also in ~~updated [45].~~ ~~While [46] also used the Physcraper updated tree.~~ ~~While the Yao2020 tree [46] was also constructed using~~ ITS as a marker, their GenBank data is not released yet, ~~so,~~ ~~Hence,~~ Physcraper was unable to incorporate 68 ~~additional taxa~~ ~~into the analysis.~~ ~~However,~~ ~~taxa that are only on the Yao2020 tree because the DNA data is unavailable.~~ We also note that Physcraper incorporates 18 *Ilex* taxa that are not in the Yao2020 tree [46]. These taxa appear nested among other *Ilex* species and visual inspection of the DNA sequences suggests they are correctly assigned as *Ilex*. The ITS alignment that underlies the Yao2020 tree was constructed without any tool to scrape GenBank [46], which could explain why Physcraper was able to incorporate ~~these 18 taxa that were not in [46].~~ additional *Ilex* taxa in the updated tree (Figure 2).

Verification test

We pruned 9 out of the 41 original tips of the Gottlieb et al. tree, corresponding to a 20% trim, excluding the outgroups. We then performed a Physcraper run to test if we would recover the dropped tips. We successfully recovered 6 out of 9 pruned tips in the updated tree. Closer examination of results revealed that the 3 missing tips were correctly retrieved with BLAST along with the other 6 tips that were included in the updated tree. Examining the ITS sequences belonging to the tips missed by the test, by following the GenBank accession numbers reported in the original publication, showed that these sequences contain a 100 bp long gap of unidentified nucleotides (Ns). These Ns were absent in the 3 sequences published in the original alignment.

Discussion

Databases preserving and democratizing access to biological data have become essential resources for science. New molecular data keep accumulating and tools

facilitating its integration into existent evolutionary knowledge contribute to the acceleration of scientific discovery.

Physcraper is a tool that builds upon previous knowledge stored in published alignments, with the main goal to extend our knowledge of phylogenetic relationships across the tree of life.

We believe this is a key step to successfully establish an open, reproducible workflow for phylogenetics, facilitating phylogenetic knowledge for ecologists and other non-specialists, effectively democratizing phylogenetic studies.

As a tool for automatizing phylogenetic reconstruction from molecular databases, Physcraper presents several advantages over existing phylogenetic pipelines designed to make evolutionary sense of the vast amount of public molecular data available.

Several analysis tools create full phylogenies *de novo* by mining of molecular databases [17, 14, 52, 53, 19]. In particular, Phylota [14], and PHLAWD [16], have been cited and used abundantly.

Physcraper builds on this automated database mining concept by incorporating prior phylogenetic work and existing taxonomic domain knowledge on appropriate markers and alignment construction. This decreases error (requiring less manual downstream processing) and eases comparison with previous phylogenetic knowledge.

Results from the verification test highlight the importance of incorporating existing expertly curated homology statements to automatically update phylogenetic relationships, instead of ignoring the information they contain and building homology statements fully *de novo*. The three sequences that were in the original tree but were not included in the updated tree by Physcraper in the verification test were excluded because they contain a 100 bp long gap of unidentified nucleotides (Ns) in GenBank that was manually curated and removed from in the original alignment.

These sequences do appear in the Physcraper results in the list of matches from GenBank which did not fit the sequence length cutoffs set in the configuration. This "seqlen_mismatch.txt" file includes the accession number, taxon, and sequence length of all sequences filtered based on sequence length.

We encourage users to look at this file, and potentially modify their sequence length cutoff parameters based on the excluded sequences. The default relative sequence length thresholds are arbitrary, but we hope that by making the process of locating homologous sequences online reproducible, and tracking what cutoffs are used, we make it easier for researchers to delve into the effect that different choices have on their inferences. This is in contrast to 'manual' searches for taxa, where similarly arbitrary cutoffs are applied, but are difficult to trace. As many studies have shown [54] the effect of missing data can be enigmatic, and interact with the true phylogenetic relationships for the data set at hand. There is not currently strong support in the literature for any cutoffs, and rather than prescribe specific approaches, we encourage users to explore the effects of different choices on inference.

Organelle genome sequences, such as chloroplasts and mitochondria will also generally be excluded from automatic addition based on length cutoffs. Multiple

sequence alignment of loci of drastically different lengths is unfeasible, and we have found in testing that it often returns incorrect results, splitting shorter sequences with many long gaps to align with exact matches across the entire longer locus. While it would be possible to directly extract the blast match from genomes, this match would exclude potentially homologous flanking regions which are not matched by blasts local search algorithm, but may be important for phylogenetic inference. Instead we list the accession numbers for these matches in the "seqlen_mismatch.txt" file, for users to assess appropriate homologous regions for their alignment of interest.

Single gene phylogenies with very high numbers of taxa may lack sufficient signal for accurate resolution [55]. The Physcraper workflow avoids some of these challenges, by focussing on ingroup taxa of an existing phylogeny, using markers that have been assessed and proven appropriate for that phylogenetic scope in past publications. In addition, Physcraper thins alignments by removing identical sequences, and by setting a maximum number of sequences per taxon. Nonetheless, it is incumbent on users to assess their final inference with respect to statistical support and biological plausibility.

Most of the default thresholds parameter values and software choices in the Physcraper pipeline are arbitrary. We hope that by making the process of locating homologous sequences online reproducible, and tracking what cutoffs are used, we make it easier for researchers to delve into the effect that different choices have on their inferences. In addition, by providing the output files at each step of the analysis, it is straightforward to assess how changing parameter and software choices do or do not drive differences in phylogenetic inference. By gathering the sequences, and making the unaligned files easily available to users, researchers can compare if applying any alternate alignment tool of their choice affects inferences. Once sequences are aligned, they can apply and compare inferences from any Phylogenetic software.

Unlike phylogenetic placement approaches [56, 57], which add new taxa without modifying the input tree, Physcraper estimates all the relationships anew in the context of the new data. PUMPER [18] shares these conceptual strengths, but is no longer under active development, is challenging to install and run, and has resulted in very few phylogenetic analyses since its publication.

Physcraper generates gene trees, which individually do not capture the full complexity of species' evolutionary history [58]. However, Physcraper facilitates gathering alignments and gene trees for multiple loci from a group of interest, that together can be used to reconstruct species trees taking into account coalescent processes with ASTRAL [59], BEAST2 [60], or SVD Quartets [61]. Rigorous analyses of multiple loci allows for more complex evolutionary models than analyses of large genomic data sets, and can provide better evolutionary estimates.

For example [62] show that when applying coalescent models, there is more information in two genes of 300 bp each than in 600 independent sites. We have included a script ("multi_locus.py") and examples of how to automatically merge the outputs of Physcraper runs at different loci into input files for ASTRAL, SVD Quartets, or as concatenated alignments for concatenated analyses.

Physcraper has the added advantage of facilitating the linkage of taxonomic information about tips in the output phylogenies to data available in a variety of

biological databases [9], such as geographical locations for taxa from the GBIF [13]. Taxonomic links, and comparisons to existing published phylogenies in the OpenTree data store can also help flag paralogous sequences. Accidentally including paralogs as homologs is a risk in phylogenetic analyses, and can be more prevalent in automated analyses than in manually curated analyses. We provide users with several tools to try to assess homology of their aligned sequences. The estimated gene tree itself is an evolutionarily explicit way to visualize gene evolution, which in concert with taxonomic labelling can reveal paralogy. Our conflict analysis tool informs the users of whether their tree contains major conflicts with established taxonomy. This conflict tool also returns information on if these taxonomic conflicts existed in the original input tree. These conflicts may be a sign that taxonomy needs to be updated, or may be a sign that non-homologous sequences have been included in the analysis. These taxonomic and phylogenetic conflicts flag regions for the tree for the researcher to more closely examine and assess homology.

The updated holly tree is based on a single marker, so we expected for it to be not as well resolved as trees using multiple markers. Nevertheless, all samples corresponding to the ingroup cluster together in a monophyletic group. We also note that most samples belonging to the same species also cluster together forming monophyletic groups. Notable exceptions are *Ilex theezans*, which also appears as non-monophyletic in the original Gottlieb et al. 2005 tree. A visual comparison of the Yao et al. 2020 tree and the original Gottlieb et al. 2005 tree suggests that the relationships within the genus *Ilex* are still being determined, and that increased taxon sampling is helping with it.

The Physcraper workflow can be used to rapidly (in a matter of hours) create phylogenies which can address challenges overarching both fields of ecology and evolution, such as phylogenetically placing newly discovered species [63], curating taxonomic assignments [64], and generating custom trees for ecological [65] and evolutionary downstream analyses [66].

Conclusions

Data repositories hold more information than meets the eye. Beyond the main data, they are rich sources of metadata that can be leveraged for the advantage of all areas of biology as well as the advancement of scientific policy and applications. Initial ideas about the data are constantly changed by results from new analyses. Physcraper provides a framework for reproducible ~~phylogenetics~~ Phylogenetics that has the potential to consistently provide context for these ideas, highlighting the importance of data sharing and open science in the field, biology and science.

Availability and requirements

Project name: Physcraper

Project home page: <https://physcraper.readthedocs.io/en/latest/index.html>

Operating System: Linux, Mac, Windows

Programming Language: Python

Other requirements: Dependencies

License: GNU

Any restrictions to use by non-academics: As specified by the License

Abbreviations

OpenTree: The Open Tree of Life project

TNRS: Taxonomic Name Resolution Service

MRCA: Most Recent Common Ancestor

BLAST: Basic Local Alignment Search Tool

NCBI: USA National Center for Biodiversity Information

GBIF: Global Biodiversity Information Facility

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Availability of data and materials

The datasets generated and analysed during the current study are available in the repositories "physcraper" containing the source code, <https://github.com/McTavishLab/physcraper>; "physcraperex" containing the examples, <https://github.com/McTavishLab/physcraperex>; and, "physcraper.ms" containing this reproducible manuscript, <https://github.com/McTavishLab/physcraper.ms>.

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

LLSR wrote manuscript, alignment code, documentation, performed analyses and developed examples; MK wrote code for ncbitdataparser module, filtering of sequences per OTU and using offline blast searches, wrote documentation and tests; EJM conceived study, wrote most of the code, documentation and tests. All authors contributed to the manuscript and gave final approval for publication. . . .

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References

1. Dobzhansky, T.: Nothing in biology makes sense except in the light of evolution. *The American Biology Teacher* **35**(3), 125–129 (1973)
2. Hillis, D.M.: Inferring complex phylogenies. *Nature* **383**(6596), 130–131 (1996)
3. Natsidis, P., Tsakogiannis, A., Pavlidis, P., Tsigenopoulos, C.S., Manousaki, T.: Phylogenomics investigation of sparids (teleostei: Spariformes) using high-quality proteomes highlights the importance of taxon sampling. *Communications Biology* **2**(1), 1–10 (2019)
4. Schulte, J.A.: Undersampling taxa will underestimate molecular divergence dates: an example from the south american lizard clade liolaemini. *International Journal of Evolutionary Biology* **2013** (2013)
5. Soares, A.E., Schrago, C.G.: The influence of taxon sampling on bayesian divergence time inference under scenarios of rate heterogeneity among lineages. *Journal of Theoretical Biology* **364**, 31–39 (2015)
6. Kayaalp, P., Stevens, M.I., Schwarz, M.P.: Back to africa: increased taxon sampling confirms a problematic australia-to-africa bee dispersal event in the eocene. *Systematic Entomology* **42**(4), 724–733 (2017)
7. Hedtke, S.M., Townsend, T.M., Hillis, D.M.: Resolution of phylogenetic conflict in large data sets by increased taxon sampling. *Systematic Biology* **55**(3), 522–529 (2006)
8. Townsend, J.P., Lopez-Giraldez, F.: Optimal selection of gene and ingroup taxon sampling for resolving phylogenetic relationships. *Systematic Biology* **59**(4), 446–457 (2010)
9. Rees, J.A., Cranston, K.: Automated assembly of a reference taxonomy for phylogenetic data synthesis. *Biodiversity Data Journal* (5) (2017). doi:[10.3897/BDJ.5.e12581](https://doi.org/10.3897/BDJ.5.e12581)
10. Baxevanis, A.D., Bateman, A.: The importance of biological databases in biological discovery. *Current Protocols in Bioinformatics* **50**(1), 1–1 (2015)
11. Federhen, S.: The NCBI Taxonomy database. *Nucleic Acids Research* **40**(D1), 136–143 (2012). doi:[10.1093/nar/gkr1178](https://doi.org/10.1093/nar/gkr1178). Publisher: Oxford Academic. Accessed 2020-12-03

12. Schoch, C.L., Ciufu, S., Domrachev, M., Hottton, C.L., Kannan, S., Khovanskaya, R., Leipe, D., Mcveigh, R., O'Neill, K., Robbertse, B., Sharma, S., Soussov, V., Sullivan, J.P., Sun, L., Turner, S., Karsch-Mizrachi, I.: NCBI Taxonomy: a comprehensive update on curation, resources and tools. *Database* **2020** (2020)
13. Secretariat, G.: GBIF Backbone Taxonomy (2019). doi:[10.15468/39omei](https://doi.org/10.15468/39omei). Accessed 2020-12-03
14. Sanderson, M.J., Boss, D., Chen, D., Cranston, K.A., Wehe, A.: The PhyLoTA Browser: Processing GenBank for Molecular Phylogenetics Research. *Systematic Biology* **57**(3), 335–346 (2008). doi:[10.1080/10635150802158688](https://doi.org/10.1080/10635150802158688). <https://academic.oup.com/sysbio/article-pdf/57/3/335/24203605/57-3-335.pdf>
15. McTavish, E.J., Drew, B.T., Redelings, B., Cranston, K.A.: How and Why to Build a Unified Tree of Life. *BioEssays* **39**(11) (2017). doi:[10.1002/bies.201700114](https://doi.org/10.1002/bies.201700114). Number: 11. Accessed 2018-04-10
16. Smith, S.A., Beaulieu, J.M., Donoghue, M.J.: Mega-phylogeny approach for comparative biology: an alternative to supertree and supermatrix approaches. *BMC Evolutionary Biology* **9**(1), 37 (2009). doi:[10.1186/1471-2148-9-37](https://doi.org/10.1186/1471-2148-9-37)
17. Antonelli, A., Hettling, H., Condamine, F.L., Vos, K., Nilsson, R.H., Sanderson, M.J., Sauquet, H., Scharn, R., Silvestro, D., Töpel, M., et al.: Toward a self-updating platform for estimating rates of speciation and migration, ages, and relationships of taxa. *Systematic Biology* **66**(2), 152–166 (2017). doi:[10.1093/sysbio/syw066](https://doi.org/10.1093/sysbio/syw066)
18. Izquierdo-Carrasco, F., Cazes, J., Smith, S.A., Stamatakis, A.: Pumper: phylogenies updated perpetually. *Bioinformatics* **30**(10), 1476–1477 (2014). doi:[10.1093/bioinformatics/btu053](https://doi.org/10.1093/bioinformatics/btu053)
19. Pearse, W.D., Purvis, A.: phylogenerator: an automated phylogeny generation tool for ecologists. *Methods in Ecology and Evolution* **4**(7), 692–698 (2013)
20. Jones, M.R., Good, J.M.: Targeted capture in evolutionary and ecological genomics. *Molecular Ecology* **25**(1), 185–202 (2016). doi:[10.1111/mec.13304](https://doi.org/10.1111/mec.13304)
21. Andermann, T., Torres Jiménez, M.F., Matos-Maraví, P., Batista, R., Blanco-Pastor, J.L., Gustafsson, A.L.S., Kistler, L., Liberal, I.M., Oxelman, B., Bacon, C.D., Antonelli, A.: A guide to carrying out a phylogenomic target sequence capture project. *Frontiers in Genetics* **10**(1407), 1–20 (2020). doi:[10.3389/fgene.2019.01407](https://doi.org/10.3389/fgene.2019.01407)
22. Fragoso-Martínez, I., Salazar, G.A., Martínez-Gordillo, M., Magallón, S., Sánchez-Reyes, L., Lemmon, E.M., Lemmon, A.R., Sazatornil, F., Mendoza, C.G.: A pilot study applying the plant Anchored Hybrid Enrichment method to New World sages (*Salvia* subgenus *Calospatha*; Lamiaceae). *Molecular Phylogenetics and Evolution* **117**, 124–134 (2017). doi:[10.1016/j.ympev.2017.02.006](https://doi.org/10.1016/j.ympev.2017.02.006)
23. Piel, W., Chan, L., Dominus, M., Ruan, J., Vos, R., Tannen, V.: Treebase v. 2: A database of phylogenetic knowledge. e-Biosphere. London (2009)
24. Morrison, D.A.: Multiple sequence alignment for phylogenetic purposes. *Australian Systematic Botany* **19**(6), 479–539 (2006). doi:[10.1071/SB06020](https://doi.org/10.1071/SB06020)
25. McTavish, E.J., Hinchliff, C.E., Allman, J.F., Brown, J.W., Cranston, K.A., Holder, M.T., Rees, J.A., Smith, S.A.: Phylsystem: A git-based data store for community-curated phylogenetic estimates. *Bioinformatics* **31**(17), 2794–2800 (2015). doi:[10.1093/bioinformatics/btv276](https://doi.org/10.1093/bioinformatics/btv276)
26. Sanderson, M., Donoghue, M., Piel, W., Eriksson, T.: Treebase: a prototype database of phylogenetic analyses and an interactive tool for browsing the phylogeny of life. *American Journal of Botany* **81**(6), 183 (1994). doi:[publication does not exist](https://doi.org/10.1093/ajbot/81.6.183)
27. Benson, D.A., Karsch-Mizrachi, I., Lipman, D.J., Ostell, J., Rapp, B.A., Wheeler, D.L.: Genbank. *Nucleic Acids Research* **28**(1), 15–18 (2000). doi:[10.1093/nar/28.1.15](https://doi.org/10.1093/nar/28.1.15)
28. S.A.), S.T.L.: Pylint. <https://www.pylint.org/>
29. S.A.), S.T.L., PyCQA, contributors: Pylint User Manual. <http://pylint.pycqa.org/en/latest/>
30. OpenTreeOfLife: Name Resolution (TNRS) bulk mapping tool. <https://tree.opentreeoflife.org/curator/tncs/>
31. OpenTreeOfLife, E.J. McTavish, Hinchliff, C.E., Allman, J.F., Brown, J.W., Cranston, K.A., Holder, M.T., Rees, J.A., Smith, S.A.: Phylsystem's top-level repository in the Open Tree of Life phylogenetic study document store. <https://github.com/opentreeoflife/phylsystem>
32. Altschul, S.F., Gish, W., Miller, W., Myers, E.W., Lipman, D.J.: Basic local alignment search tool. *Journal of Molecular Biology* **215**(3), 403–410 (1990). doi:[10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2)
33. OpenTreeOfLife, Rees, J.A., Cranston, K.: OpenTree's taxonomic MRCA API. <https://github.com/OpenTreeOfLife/germinator/wiki/Taxonomy-API-v3#mrca>
34. Camacho, C., George, C., Vahram, A., Ning, M., Jason, P., Kevin, B., Thomas, L.: BLAST+: Architecture and applications. *BMC Bioinformatics* **10**(1), 421 (2009). doi:[10.1186/1471-2105-10-421](https://doi.org/10.1186/1471-2105-10-421)
35. Cock, P.J., Antao, T., Chang, J.T., Chapman, B.A., Cox, C.J., Dalke, A., Friedberg, I., Hamelryck, T., Kauff, F., Wilczynski, B., et al.: Biopython: freely available Python tools for computational molecular biology and bioinformatics. *Bioinformatics* **25**(11), 1422–1423 (2009). doi:[10.1093/bioinformatics/btp163](https://doi.org/10.1093/bioinformatics/btp163)
36. The BioPython Contributors, -: BioPython's Bio.Blast.NCBIWWW module. <https://biopython.org/DIST/docs/api/Bio.Blast.NCBIWWW-module.html>
37. Edgar, R.C.: Muscle: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* **32**(5), 1792–1797 (2004). doi:[10.1093/nar/gkh340](https://doi.org/10.1093/nar/gkh340)
38. Stamatakis, A.: Raxml version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**(9), 1312–1313 (2014). doi:[10.1093/bioinformatics/btu033](https://doi.org/10.1093/bioinformatics/btu033)
39. Sukumaran, J., Holder, M.T.: DendroPy: a Python library for phylogenetic computing. *Bioinformatics* **26**(12), 1569–1571 (2010). doi:[10.1093/bioinformatics/btq228](https://doi.org/10.1093/bioinformatics/btq228)
40. Redelings, B.D., Holder, M.T.: A supertree pipeline for summarizing phylogenetic and taxonomic information for millions of species. *PeerJ* **5**, 3058 (2017). doi:[10.7717/peerj.3058](https://doi.org/10.7717/peerj.3058)
41. Cuénoud, P., Martinez, M.A.d.P., Loizeau, P.-A., Spichiger, R., Andrews, S., Manen, J.-F.: Molecular phylogeny and biogeography of the genus *Ilex* L.(aquifoliaceae). *Annals of Botany* **85**(1), 111–122 (2000). doi:[10.1006/anbo.1999.1003](https://doi.org/10.1006/anbo.1999.1003)
42. Manen, J.-F., Barriera, G., Loizeau, P.-A., Naciri, Y.: The history of extant *Ilex* species (Aquifoliaceae): evidence of hybridization within a Miocene radiation. *Molecular Phylogenetics and Evolution* **57**(3), 961–977

- (2010). doi:[10.1016/j.jympev.2010.09.006](https://doi.org/10.1016/j.jympev.2010.09.006)
43. Setoguchi, H., Watanabe, I.: Intersectional gene flow between insular endemics of *Ilex* (Aquifoliaceae) on the Bonin Islands and the Ryukyu Islands. *American Journal of Botany* **87**(6), 793–810 (2000). doi:[10.2307/2656887](https://doi.org/10.2307/2656887)
 44. Selbach-Schnadelbach, A., Cavalli, S.S., Manen, J.-F., Coelho, G.C., De Souza-Chies, T.T.: New information for *Ilex* phylogenetics based on the plastid psbA-trnH intergenic spacer (Aquifoliaceae). *Botanical Journal of the Linnean Society* **159**(1), 182–193 (2009). doi:[10.1111/j.1095-8339.2008.00898.x](https://doi.org/10.1111/j.1095-8339.2008.00898.x)
 45. Gottlieb, A.M., Giberti, G.C., Poggio, L.: Molecular analyses of the genus *Ilex* (aquifoliaceae) in southern south america, evidence from aifp and its sequence data. *American Journal of Botany* **92**(2), 352–369 (2005). doi:[10.3732/ajb.92.2.352](https://doi.org/10.3732/ajb.92.2.352)
 46. Yao, X., Song, Y., Yang, J.-B., Tan, Y.-H., Corlett, R.T.: Phylogeny and biogeography of the hollies (*Ilex* L., aquifoliaceae). *Journal of Systematics and Evolution* **58**(5), 1–10 (2020). doi:[10.1111/jse.12567](https://doi.org/10.1111/jse.12567)
 47. Gottlieb, A.M., Giberti, G.C., Poggio, L.: TreeBASE study 1091. <https://treebase.org/treebase-web/search/study/summary.html?id=1091>
 48. Gottlieb, A.M., Giberti, G.C., Poggio, L.: Phylsystem study pg.2827. https://tree.opentreeoflife.org/curator/study/edit/pg_2827/?tab=home
 49. OpenTreeOfLife, Redelings, B., Reyes, L.L.S., Cranston, K.A., Allman, J., Holder, M.T., McTavish, E.J.: Open Tree of Life Synthetic subtree, node id mrcaott68451ott89474. <https://tree.opentreeoflife.org/opentree/opentree12.3@mrcaott68451ott89474/Ilex-theizans--Ilex-dumosa>
 50. Yao, X., Song, Y., Yang, J.-B., Tan, Y.-H., Corlett, R.T.: Phylsystem study ot.1984. https://tree.opentreeoflife.org/curator/study/view/ot_1984
 51. Yao, X., Song, Y., Yang, J.-B., Tan, Y.-H., Corlett, R.T.: Phylogeny and biogeography of the hollies (*Ilex* L., Aquifoliaceae), Dryad, Dataset. <https://datadryad.org/stash/dataset/doi:10.5061/dryad.k0p2nngf4x>. Accessed April 2020
 52. Smith, S.A., Walker, J.F.: Pyphlawd: A python tool for phylogenetic dataset construction. *Methods in Ecology and Evolution* **10**(1), 104–108 (2019). doi:[10.1111/2041-210X.13096](https://doi.org/10.1111/2041-210X.13096)
 53. Bennett, D.J., Hettling, H., Silvestro, D., Zizka, A., Bacon, C.D., Faurby, S., Vos, R.A., Antonelli, A.: phylotar: An automated pipeline for retrieving orthologous dna sequences from genbank in r. *Life* **8**(2), 20 (2018). doi:[10.3390/life8020020](https://doi.org/10.3390/life8020020)
 54. Huang, H., Knowles, L.L.: What Is the Danger of the Anomaly Zone for Empirical Phylogenetics? *Systematic Biology*, 047 (2009). doi:[10.1093/sysbio/syp047](https://doi.org/10.1093/sysbio/syp047). Accessed 2015-07-24
 55. Morel, B., Barbera, P., Czech, L., Bettisworth, B., Hübner, L., Lutteropp, S., Serdari, D., Kostaki, E.-G., Mamais, I., Kozlov, A.M., Pavlidis, P., Paraskevis, D., Stamatakis, A.: Phylogenetic Analysis of SARS-CoV-2 Data Is Difficult. *Molecular Biology and Evolution* (msaa314) (2020). doi:[10.1093/molbev/msaa314](https://doi.org/10.1093/molbev/msaa314). Accessed 2021-03-30
 56. Berger, S.A., Krompass, D., Stamatakis, A.: Performance, Accuracy, and Web Server for Evolutionary Placement of Short Sequence Reads under Maximum Likelihood. *Systematic Biology*, 010 (2011). doi:[10.1093/sysbio/syr010](https://doi.org/10.1093/sysbio/syr010). Accessed 2015-01-12
 57. Matsen, F., Kodner, R., Armbrust, E.V.: pplacer: linear time maximum-likelihood and Bayesian phylogenetic placement of sequences onto a fixed reference tree. *BMC bioinformatics* **11**(1), 538 (2010). Number: 1. Accessed 2013-12-17
 58. Song, S., Liu, L., Edwards, S.V., Wu, S.: Resolving conflict in eutherian mammal phylogeny using phylogenomics and the multispecies coalescent model. *Proceedings of the National Academy of Sciences* **109**(37), 14942–14947 (2012). doi:[10.1073/pnas.1211733109](https://doi.org/10.1073/pnas.1211733109)
 59. Mirarab, S., Reaz, R., Bayzid, M.S., Zimmermann, T., Swenson, M.S., Warnow, T.: ASTRAL: genome-scale coalescent-based species tree estimation. *Bioinformatics* **30**(17), 541–548 (2014). doi:[10.1093/bioinformatics/btu462](https://doi.org/10.1093/bioinformatics/btu462)
 60. Bouckaert, R., Vaughan, T.G., Barido-Sottani, J., Duchêne, S., Fourment, M., Gavryushkina, A., Heled, J., Jones, G., Kühnert, D., Maio, N.D., Matschiner, M., Mendes, F.K., Müller, N.F., Ogilvie, H.A., Plessis, L.d., Poppinga, A., Rambaut, A., Rasmussen, D., Siveroni, I., Suchard, M.A., Wu, C.-H., Xie, D., Zhang, C., Stadler, T., Drummond, A.J.: BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis. *PLOS Computational Biology* **15**(4), 1006650 (2019). doi:[10.1371/journal.pcbi.1006650](https://doi.org/10.1371/journal.pcbi.1006650). Publisher: Public Library of Science
 61. Chifman, J., Kubatko, L.: Quartet inference from SNP data under the coalescent model. *Bioinformatics* **30**(23), 3317–3324 (2014). doi:[10.1093/bioinformatics/btu530](https://doi.org/10.1093/bioinformatics/btu530). Publisher: Oxford Academic
 62. Zhu, T., Yang, Z.: Complexity of the simplest species tree problem. *Molecular Biology and Evolution* (msab009) (2021). doi:[10.1093/molbev/msab009](https://doi.org/10.1093/molbev/msab009). Accessed 2021-03-30
 63. Webb, C.O., Slik, J.F., Triono, T.: Biodiversity inventory and informatics in Southeast Asia. *Biodiversity and Conservation* **19**(4), 955–972 (2010). doi:[10.1007/s10531-010-9817-x](https://doi.org/10.1007/s10531-010-9817-x)
 64. San Mauro, D., Agorreta, A.: Molecular systematics: a synthesis of the common methods and the state of knowledge. *Cellular & Molecular Biology Letters* **15**(2), 311 (2010). doi:[10.2478/s11658-010-0010-8](https://doi.org/10.2478/s11658-010-0010-8)
 65. Helmus, M.R., Ives, A.R.: Phylogenetic diversity–area curves. *Ecology* **93**(sp8), 31–43 (2012). doi:[10.1890/11-0435.1](https://doi.org/10.1890/11-0435.1)
 66. Stoltzfus, A., Lapp, H., Matasci, N., Deus, H., Sidlauskas, B., Zmasek, C.M., Vaidya, G., Pontelli, E., Cranston, K., Vos, R., et al.: Phylotastic! making tree-of-life knowledge accessible, reusable and convenient. *BMC Bioinformatics* **14**(1), 158 (2013). doi:[10.1186/1471-2105-14-158](https://doi.org/10.1186/1471-2105-14-158)

Figures

Figure 1 The Physcraper framework consists of four general steps. The software is fully described on its documentation website at <https://physcraper.readthedocs.io>, along with installation instructions, function usage descriptions, examples and tutorials.

Figure 2 A) Phylogeny updated with Physcraper from original [45] tree in B. Tips in original alignment and new tips added with Physcraper are depicted in black and red, respectively. Physcraper obtained sequences from the GenBank database via local BLAST of all sequences in the original alignment that generated tree in B), filtered them following criteria from section "DNA sequence search and filtering", aligned them to original alignment using MUSCLE and performed a phylogenetic reconstruction using RAxML with 100 bootstraps. B-D conflict analyses performed with OpenTree tools.