

## RESEARCH

# Physcraper: A Python package for continually updated phylogenetic trees using the Open Tree of Life

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## Abstract

**Background:** Phylogenies are a key part of research in many areas of biology. Tools that automate some parts of the process of phylogenetic reconstruction, mainly molecular character matrix assembly, have been developed for the advantage of both specialists in the field of phylogenetics and non-specialists. However, interpretation of results, comparison with previously available phylogenetic hypotheses, and selection of one phylogeny for downstream analyses and discussion still impose difficulties to one that is not a specialist either on phylogenetic methods or on a particular group of study.

**Results:** Physcraper is a command-line Python program that automates the update of published phylogenies by adding public DNA sequences to underlying alignments of previously published phylogenies. It also provides a framework for straightforward comparison of published phylogenies with their updated versions, by leveraging upon tools from the Open Tree of Life project to link taxonomic information across databases. The program can be used by the nonspecialist, as a tool to generate phylogenetic hypotheses based on publicly available expert phylogenetic knowledge. Phylogeneticists and taxonomic group specialists will find it useful as a tool to facilitate molecular dataset gathering and comparison of alternative phylogenetic hypotheses (topologies).

**Conclusions:** The Physcraper workflow showcases the benefits of doing open science for phylogenetics, encouraging researchers to strive for better sharing practices. Physcraper can be used with any OS and is released under an open-source license. Detailed instructions for installation and usage are available at <https://physcraper.readthedocs.io>.

**Keywords:** gene tree; interoperability; open science; reproducibility; public database; DNA alignment

## Background

Phylogenies capture the shared history of organisms and provide key evolutionary context for our biological observations [1]. Public biological databases constitute an amazing resource for evolutionary studies. Connecting existing phylogenies to molecular data that has never been incorporated into any phylogenetic estimate, geographical location, fossils, and other data in a reproducible and continuous manner is possible by establishing a data interoperability framework for biological databases. Here, we introduce Physcraper, a tool that automates biological database connections with the main goal of building upon homology hypotheses that taxon specialists have assessed and deemed appropriate for a specific phylogenetic scope to

update a starting tree and single locus alignments with public DNA data, with the added capability to connect the lineages represented in the phylogeny to biological data in public databases.

One of the main challenges for automatic integration of biological data into phylogenies are varying taxonomic idiosyncrasies across databases. To address this challenge, the Open Tree of Life project (OpenTree) unifies taxonomic data from various databases [2], including the USA National Center for Biodiversity Information (NCBI) molecular database GenBank [3], among others. The OpenTree taxonomy represents a key resource that remains to be leveraged for connecting data from any biological database that has been integrated to it.

The Open Tree of Life project (OpenTree) also constructs a comprehensive tree of life by synthesizing published phylogenies and taxonomy. OpenTree's "synthetic" tree comprises 2.3 million tips, of which around 90,000 (87,740) are supported by phylogenies - the remaining 1.4 million taxa are placed in the tree based on taxonomy. Meaning that there is still a lot of work to do to complete a full phylogenetic tree of all life.

Decades of single locus sequencing have generated massive amounts of homologous DNA datasets that have the potential to be used for phylogenetic reconstruction at many scales [4]. More than a decade ago, GenBank release 159 (April 15, 2007) already hosted 72 million DNA sequences that were gauged to have the potential to resolve phylogenetic relationships of 98.05% of the almost 241,000 distinct taxa in the NCBI taxonomy at the time [4].

With more than XXXX million single locus sequences in GenBank today, there is a considerable amount of phylogenetically informative data in GenBank that has not been analysed and/or made publicly available with the potential to give phylogenetic light to the tree of life.

Phylogenies constructed from single locus super matrix datasets are still widely generated and relevant to the field. Notable examples include the

Notable examples of phylogenies constructed using multi-locus, database mined, super matrix approach: Magallon, Soltis, examples with fish, bats, [5] mining by hand - flowering plant families - has been cited 561 times [6] 38k clades of plants with seeds - cited 176 times [7] 6k bird species - cited [8] mining with phlawd - cited 202 times

These studies are widely cited and immensely relevant for the biological community.

Some of these multilocus phylogenetic studies have implemented pipelines such as PHLAWD, other studies have curated their matrices by hand, which is largely non-reproducible.

Make emphasis on these really great studies (and pipelines) that do not rely on genomic data, and that there is a lot of single locus data that can be analysed.

Assembling a DNA alignment from such a massive database can be done "by hand", but it is a largely time consuming and mostly non-reproducible approach. Computational pipelines that mine DNA databases fast, efficiently, and reproducibly, have been applied to infer phylogenetic relationships in a variety of organisms (e.g., [9, 10, 11]). However, fine-grained curated markers and alignments can improve phylogenetic reconstructions, even in phylogenomic analyses [12].

There are almost 8,200 publicly available, peer-reviewed alignments, covering around 100,000 distinct taxa in the TreeBASE database [13], which can be used as seeds to mine molecular databases, and as "jump-start" alignments for phylogenetic reconstructions [14] to continually enrich, update and compare existing phylogenetic knowledge.

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Physcraper is a Python pipeline using OpenTree's taxonomy and programmatic access protocols (API's) to implement a database interoperability framework that automatically links phylogenies that have been standardized to OpenTree taxonomy, to alignments from TreeBASE, data from GenBank, and phylogenies from OpenTree's Phylesystem. Physcraper aims to demonstrate the benefits of reproducible workflows and open science in phylogenetics, and encourage better data sharing practices in the community.

## Implementation

The general Physcraper framework (Figure 1) consists of 4 steps: 1) identifying and processing a tree and its underlying alignment; 2) performing a BLAST search of DNA sequences from original alignment on GenBank, and filtering of new sequences; 3) profile-aligning new sequences to original alignment; 4) performing a phylogenetic analysis and comparing the updated tree to existing phylogenies.

### The inputs: a tree and an alignment

Taxon names in the input tree must be standardized to OpenTree taxonomy [2] using OpenTree's bulk Taxonomic Name Resolution Service (TNRS) tool [15]. Users can upload their own tree, or choose from among the 2, 950 standardized trees stored in OpenTree's Phylesystem [16, 17] that also have alignments available on TreeBASE [13].

The input alignment is a single locus DNA dataset that was used in part or in whole to generate the input tree. Physcraper retrieves TreeBASE alignments automatically. Alternatively, users must provide the path to a local copy of the alignment. Only taxa that are both in the sequence alignment and in the tree are considered further for analysis; at least one taxon and its corresponding sequence are required.

### DNA sequence search and filtering

The Basic Local Alignment Search Tool, BLAST [18] is used for DNA sequence search on a remote or local GenBank database. It is constrained to a "search taxon", a taxonomic group in the NCBI taxonomy that is automatically identified using the OpenTree's taxonomic Most Recent Common Ancestor (MRCA) API [19, 2], as the MRCA of all ingroup taxa that is also a named clade in the NCBI taxonomy (Figure 1).

BLAST is performed using the 'blastn' algorithm [20] implemented in BioPython's [21] NCBIWWW module [22] modified to accept an alternative BLAST address. Each sequence in the alignment is BLASTed once against all DNA sequences in GenBank. New sequences are excluded for analysis if they 1) are not in the search taxon; 2) have an e-value above the cutoff (default to 0.00001); 3) fall outside a min

and max length threshold, defined as the proportion of the average length without gaps of all sequences in input alignment (default values of 80 respectively); 4) or if they are either identical to or shorter than an existing sequence in the input alignment and they represent the same taxon in OpenTree or NCBI taxonomy. An arbitrary maximum number of randomly chosen sequences per taxon are allowed (default to 5).

Reverse, complement, and reverse-complement sequences are identified and translated using BioPython internal functions [21]. Iterative cycles of BLAST searches can be performed, by blasting all new sequences until no new ones are found. By default only one BLAST cycle is performed.

#### New DNA sequence alignment

MUSCLE [23] is used to perform a profile alignment in which the original alignment is used as a template of homology criteria to align new sequences. The final alignment is not further automatically checked, and additional inspection and refinement are recommended.

#### Tree reconstruction and comparison

RAxML [24] is implemented to reconstruct a Maximum Likelihood (ML) gene tree for each input alignment with default settings (GTRCAT model and 100 bootstrap replicates with default algorithm), using input tree as starting tree for ML searches. Bootstrap results are summarized using DendroPy's SumTrees module [25].

Physcraper's main result is an updated phylogenetic hypothesis for the search taxon. Updated and original tree are compared with Robinson-Foulds weighted and unweighted metrics estimated with Dendropy [25], and with a node by node comparison between the synthetic OpenTree and original and updated tree individually, using OpenTree's conflict API [26].

## Results

### Case Study: The hollies

A user is interested in phylogenetic relationships within the genus *Ilex*. Commonly known as "hollies", the genus encompasses between 400-700 living species, and is the only extant clade within the family Aquifoliaceae, order Aquifoliales of flowering plants.

An online literature review in June 2020 (Google scholar search for "ilex phylogeny") reveals that there are several published phylogenies showing relationships within the hollies [27, 28, 29, 30], but only two have data publicly available [31, 32]. [31] made original tree and alignment available in TreeBASE (study 1091 [33]).

The tree sampling 41 species is also available from OpenTree's Phylesystem (study pg-2827 [34]), and has been integrated into OpenTree's synthetic tree [35].

The most recent *Ilex* tree [32] is available in OpenTree's Phylesystem (study ot\_1984 [36]), and in the DRYAD repository [37]. With 175 tips, the [32] tree is the best sampled phylogeny yet available for the hollies.

We ran Physcraper on a laptop Linux computer to update an internal transcribed spacer DNA region (ITS) alignment from [31], using a local GenBank database. BLAST and RAxML analyses ran for 19hrs 45min, with bootstrap analyses taking

an additional 13hrs. The updated [31] tree (Figure 2) displays all 41 distinct taxa from the original study plus 231 new tips, contributing phylogenetic data to 84 additional *Ilex* taxa. The best RaxML tree is 99% resolved, with 25% of nodes with bootstrap support < 0.1 and 48% nodes with bootstrap support > 0.75. A large portion of internal branches are negligibly small, with 30 branches < 0.00001 substitution rate units, from which only 9 have a bootstrap support > 0.75 (Figure 2). For comparison, [32] also contains all 41 distinct taxa from the original [31] study, and contributes phylogenetic data to 134 additional *Ilex* taxa, from which ‘r 135-68’ are also in updated [31]. While [32] also used ITS as a marker, their GenBank data is not released yet, so Physcraper was unable to incorporate 68 additional taxa into the analysis. However, Physcraper was able to incorporate 18 taxa that were not in [32].

## Discussion

Databases preserving and democratizing access to biological data have become essential resources for science. New molecular data keep accumulating and tools facilitating its integration into existent evolutionary knowledge are needed.

Phylogenetic pipelines designed to make evolutionary sense of the vast amount of public molecular data (e.g., Phylota [4], PHLAWD [9], SUPERSMART [10]) focus on generating full phylogenies *de novo*, i.e., inferring phylogenetic relationships from a newly generated homology hypothesis, as opposed to e.g., supertrees, that are generated by summarizing previous phylogenetic estimates. While Physcraper does not generate phylogenies *de novo* in a traditional sense, it successfully generates new phylogenetic knowledge, revealing the importance of open science in facilitating phylogenetic placement of public molecular data and accelerating enrichment and updating of phylogenetic relationships in any region of the tree of life. The PUMPER pipeline [11] also uses the concept of updating pre-existing alignments to incorporate public molecular data into phylogenies. Unfortunately, installation was unsuccessful following instructions from the author, and a comparison analysis between Physcraper and PUMPER is unfeasible at present.

Physcraper generates individual gene trees, failing to capture the complexity of species’ evolutionary history [38]. Yet, Physcraper facilitates gathering alignments and gene trees for multiple loci from a group of interest, that can be used to reconstruct species trees with ASTRAL [39], BEAST2 [40], or SVD Quartets [41]).

Physcraper can potentially link phylogenies to data available in any of the taxonomies integrated in the OpenTree taxonomy [2], such as geographical locations from the Global Biodiversity Information Facility, or fossils from the Paleobiology Database. The Physcraper workflow can be used to rapidly (in a matter of hours) address challenges overarching both fields of ecology and evolution, such as phylogenetically placing newly discovered species [42], systematizing molecular (and other) databases, i.e., curating taxonomic assignments [43], and generating custom trees for ecological [44] and evolutionary downstream analyses [45].

Data repositories hold more information than meets the eye. Beyond the main data, they are rich sources of metadata that can be leveraged for the advantage of all areas of biology as well as the advancement of scientific policy and applications. Initial ideas about the data are constantly changed by results from new analyses.

Physcraper provides a framework for reproducible phylogenetics that has the potential to consistently provide context for these ideas, highlighting the importance of data sharing and open science in the field, biology and science.

## Conclusions

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## Availability and requirements

**Project name:** Physcraper

**Project home page:** <https://physcraper.readthedocs.io/en/latest/index.html>

**Operating System:** Linux, Mac, Windows

**Programming Language:** Python

**Other requirements:** Dependencies

**License:** GNU

**Any restrictions to use by non-academics:** As specified by the License

## Abbreviations

**OpenTree:** The Open Tree of Life project

**TNRS:** Taxonomic Name Resolution Service

**MRCA:** Most Recent Common Ancestor

**BLAST:** Basic Local Alignment Search Tool

**NCBI:** USA National Center for Biodiversity Information

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Availability of data and materials

The datasets generated and analysed during the current study are available in the repositories "physcraper" containing the source code, <https://github.com/McTavishLab/physcraper>; "physcraperex" containing the examples, <https://github.com/McTavishLab/physcraperex>; and, "physcraper.ms" containing this reproducible manuscript, <https://github.com/McTavishLab/physcraper.ms>.

### Competing interests

The authors declare that they have no competing interests.

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### Authors' contributions

LLSR wrote manuscript, alignment code, documentation, performed analyses and developed examples; MK wrote code for ncbitaparser module, filtering of sequences per OTU and using offline blast searches, wrote documentation and tests; EJM conceived study, wrote most of the code, documentation and tests. All authors contributed to the manuscript and gave final approval for publication. ...

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## Figures

**Figure 1** The Physcraper framework consists of 4 general steps (see text). The software is fully described on its documentation website at <https://physcraper.readthedocs.io>, along with installation instructions, function usage descriptions, examples and tutorials.

**Figure 2** A) Phylogeny updated with Physcraper from original [31] tree in B. Tips in original alignment and new tips added with Physcraper are depicted in black and red, respectively. Physcraper obtained sequences from the GenBank database via local BLAST of all sequences in the original alignment that generated tree in B), filtered them following criteria from section "DNA sequence search and filtering", aligned them to original alignment using MUSCLE and performed a phylogenetic reconstruction using RAxML with 100 bootstraps. B-D conflict analyses performed with OpenTree tools.

## Tables

### Additional Files

Additional file 1 — Sample additional file title

Additional file descriptions text (including details of how to view the file, if it is in a non-standard format or the file extension). This might refer to a multi-page table or a figure.



**Table 1** Sample table title. This is where the description of the table should go

	B1	B2	B3
A1	0.1	0.2	0.3
A2	...	..	.
A3	..	.	.

Additional file 2 — Sample additional file title  
Additional file descriptions text.