# Package 'SureTypeSCR'

# November 4, 2019

Title Interface to python based package SureTypeSC via reticulate

**Description** SureTypeSCR is the implementation of algorithm for regenotyping of single cell data coming from Illumina BeadArrays without genome studio.

**Version** 0.99.0

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Suggests testthat

**Depends** R (>= 3.5.0), reticulate, knitr

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License Artistic-2.0

biocViews Software, GenotypingArray,SingleCell

VignetteBuilder knitr

**SystemRequirements** python (>= 2.7), sklearn, numpy, pandas,

SureTypeSC, IlluminaBeadArrayFiles

NeedsCompilation no

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2 apply\_thresh

allele_freq	The frequency function is to calculate the allele frequency over all the samples
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# Description

The frequency function is to calculate the allele frequency over all the samples

#### Usage

```
allele_freq(df,th=0)
```

# Arguments

df the pandas dataframe from GenomeStudio or scbasic function

th the threshold

#### Value

The frequency values of all samples

# **Examples**

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)

# The Random Forest classifier
call <- allele_freq(df,th=0.2)</pre>
```

apply\_thresh

To apply threshold over all samples on GenCall score

# Description

To apply threshold over all samples on GenCall score

```
apply_thresh(df,th)
```

calculate\_ma 3

# **Arguments**

df the pandas dataframe from GenomeStudio or schasic function

th the threshold

#### Value

Data object only with applied threshold

# **Examples**

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)

df <- create_from_frame(df)

df <- apply_thresh(df,0.01)</pre>
```

calculate\_ma

To calculate m and a features

# Description

To calculate m and a features

# Usage

```
calculate_ma(df)
```

# **Arguments**

df

the pandas dataframe from GenomeStudio or scbasic function

# Value

Data object with adding m and a features

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### **Examples**

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)

df <- create_from_frame(df)

dfs <- calculate_ma(df)</pre>
```

callrate

The callrate function is to calculate the allele frequency over all the samples

# **Description**

The callrate function is to calculate the allele frequency over all the samples

# Usage

```
callrate(df,th=0)
```

# Arguments

df the pandas dataframe from GenomeStudio or scbasic function

th the threshold

#### Value

The callrate values of all samples

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)
# The Random Forest classifier</pre>
```

callrate\_chr 5

```
call <- callrate(df,th=0.2)</pre>
```

callrate\_chr The callrate function is to calculate the allele frequency over all the samples of one specific chromosome

# **Description**

The callrate function is to calculate the allele frequency over all the samples of one specific chromosome

# Usage

```
callrate_chr(df,chr_name,th=0)
```

# **Arguments**

df the pandas dataframe from GenomeStudio or scbasic function

chr\_name the name of the selected chromsome

th the threshold

# Value

The callrate values of all samples of one specific chromosome

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)

# The Random Forest classifier
call <- callrate_chr(df,'1',th=0.2)</pre>
```

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create\_from\_frame

convert pandas dataframe to Data object

# **Description**

Convert pandas dataframe to Data object and rearrange the index level

#### Usage

```
create_from_frame(df)
```

#### **Arguments**

df

genotyping pandas dataframe from scbasic function

#### Value

Data object with index rearrangement (multi-leve index)

#### **Examples**

```
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

# get genotyping data from gtc files and meta file
df <- scbasic(manifest_path,cluster_path,samplesheet)

# create Data object and rearrange the index
df <- create_from_frame(df)</pre>
```

locus\_cluster

To do intensity aggregation at a specific locus

# **Description**

To do intensity aggregation at a specific locus

# Usage

```
locus_cluster(df,locus)
```

# **Arguments**

df the pandas dataframe from GenomeStudio or scbasic function

locus the name of the locus

locus\_ma 7

#### Value

the intensity of one locus

#### **Examples**

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)

# The Random Forest classifier
call <- locus_cluster(df,'rs3128117')</pre>
```

locus\_ma

To do m and a aggregation at a specific locus

# Description

To do m and a aggregation at a specific locus

#### Usage

```
locus_ma(df,locus)
```

#### **Arguments**

df the pandas dataframe from GenomeStudio or scbasic function

locus the name of the locus

### Value

the m and a of one locus

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')
df <- scbasic(manifest_path,cluster_path,samplesheet)</pre>
```

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```
# The Random Forest classifier
call <- locus_ma(df,'rs3128117')</pre>
```

pca\_chr

To apply principle component annalysis on frequency dataframe of samples of one chromosome

# Description

To apply principle component annalysis on frequency dataframe of samples

# Usage

```
pca_chr(df,chr_name,th=0,n=2)
```

### **Arguments**

df the pandas dataframe from GenomeStudio or scbasic function

chr\_name the name of the specific chromosome

th the threshold

n is the number of components

# Value

Component values

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)

# The Random Forest classifier
call <- pca_chr(df,'X')</pre>
```

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pca_samples	To apply principle component annalysis on frequency dataframe of samples
-------------	--

# Description

To apply principle component annalysis on frequency dataframe of samples

#### Usage

```
pca_samples(df,th=0)
```

# Arguments

df the pandas dataframe from GenomeStudio or scbasic function

th the threshold

#### Value

Component values

# **Examples**

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)

# The Random Forest classifier
call <- pca_samples(df,th=0.2)</pre>
```

restrict\_chrom

To choose certain chromosomes with Data object

# Description

To choose certain chromosomes with Data object

```
restrict_chrom(df,chrom)
```

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### **Arguments**

df the pandas dataframe from GenomeStudio or scbasic function

chrom the list of selected chromosomes

#### Value

Data object only with certain chromosomes

#### **Examples**

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)

df <- create_from_frame(df)

df <- restrict_chrom(df,c('1','2'))</pre>
```

sample\_ma

To do m and a aggregation at a specific chromosome of a specific sample

# **Description**

To do m and a at a specific chromosome of a specific sample

### Usage

```
sample_ma(df,sample_name,chr_name)
```

# Arguments

df the pandas dataframe from GenomeStudio or scbasic function

sample\_name the name of the sample chr\_name the name of the chromosome

# Value

the m and a a of specific chromosome of a specific sample

scbasic 11

#### **Examples**

```
# parsing file from gtc raw files
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

df <- scbasic(manifest_path,cluster_path,samplesheet)

# The Random Forest classifier
call <- sample_ma(df,'Kit4_4mos_SC21','1')</pre>
```

scbasic

Function to process raw gtc data and meta data without genomestudio

# **Description**

Function to process raw gtc data and meta data without genomestudio

# Usage

```
scbasic(bpm,egt,samplesheet)
```

#### **Arguments**

bpm a pathname to manifest file
egt a pathname to cluster file
samplesheet a pathname to samplesheet file

### Value

pandas data frame of genotyping data

```
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')

# get genotyping data from gtc files and meta file
df <- scbasic(manifest_path,cluster_path,samplesheet)

#/Users/apple/anaconda3/envs/gtc2/lib/python2.7/site-packages/sklearn/ensemble/weight_boosting.py:29:
#DeprecationWarning: numpy.core.umath_tests is an internal NumPy module and should not be imported.
#It will be removed in a future NumPy release.
    #from numpy.core.umath_tests import inner1d
#Reading cluster file</pre>
```

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```
#Reading sample file
#Number of samples: 2
#Reading manifest file
#Initializing genotype data
#Generating
#9968648019_R06C01
#9968648019_R06C02
#Finish parsing
```

scEls

mediate access to python modules

# Description

mediate access to python modules

# Usage

```
scEls()
```

# Value

list of (S3) "python.builtin.module"

# Note

Returns a list with elements sc (SureTypeSC), pd (pandas)each referring to python modules.

```
els = scEls()
els
##$sc
##Module(SureTypeSC)
##$pd
##Module(pandas)
```

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scload

Load Random Forest classifier or Gaussian Discrinimate Analysis

# **Description**

Load Random Forest classifier or Gaussian Discrinimate Analysis

# Usage

```
scload(filename)
```

# Arguments

filename

a pathname to an classifier

#### Value

instance of a classifier

# **Examples**

```
clf_rf_path = system.file('files/clf_30trees_7228_ratio1_lightweight.clf',package='SureTypeSCR')
clf_gda_path = system.file('files/clf_30trees_7228_ratio1_lightweight.clf',package='SureTypeSCR')
# The Random Forest classifier
clf_rf <- scload(clf_rf_path)
# The Gaussian Discriminate Analysis classifier
clf_gda <- scload(clf_gda_path)</pre>
```

scpredict

Predictions from Random Forest classifier or Gaussian Discriminant Analysis

# Description

Predictions from Random Forest classifier or Gaussian Discriminant Analysis

```
scpredict(clf_rf,test,clftype='rf')
```

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#### **Arguments**

clf\_rf classifier load by using scload

test Data object including m and a feature

clftype The type of classifier (rf: Random Forest;

gda: Gaussian Discriminant Analysis;

rf-gda: the cascade of Random Forest and Gaussian Discriminant Analysis)

#### Value

The prediction Data object.

The predicted items might include:

rf\_ratio:1\_pred: Random Forest prediction (binary)

rf\_ratio:1\_prob: Random Forest Score for the positive class

gda\_ratio:1\_prob: Gaussian Discriminant Analysis score for the positive class

gda\_ratio:1\_pred: Gaussian Disciminant Analysis prediction (binary)

rf-gda\_ratio:1\_prob: combined 2-layer RF and GDA - probability score for the positive class

rf-gda\_ratio:1\_pred: binary prediction of RF-GDA

```
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')
clf_rf_path = system.file('files/clf_30trees_7228_ratio1_lightweight.clf',package='SureTypeSCR')
clf_gda_path = system.file('files/clf_30trees_7228_ratio1_lightweight.clf',package='SureTypeSCR')
# The Random Forest classifier
clf_rf <- scload(clf_rf_path)</pre>
# The Gaussian Disciminant Analysis
clf_gda <- scload(clf_gda_path)</pre>
# get genotyping data from gtc files and meta file
df <- scbasic(manifest_path,cluster_path,samplesheet)</pre>
# create Data object and rearrange the index
dfs <- create_from_frame(df)</pre>
\# extract the chromosomes 1 and 2
dfs <- restrict_chrom(dfs,c('1','2'))</pre>
\# mask the Gencall score lower than 0.01
dfs <- apply_thresh(dfs,0.01)</pre>
# calculate the m and a feature
dfs <- calculate_ma(dfs)</pre>
# prediction by Random Forest
```

scsave 15

```
result_rf <- scpredict(clf_rf,dfs,clftype='rf')
# prediction by Guassian Discriminate Analysis
result_gda <- scpredict(clf_gda,dfs,clftype='gda')</pre>
```

scsave

Save the predictions from different classifiers

# **Description**

Save the predictions from different classifiers

# Usage

```
# save different mdoes based on the full prediction table
scsave(result,filename,header=TRUE,clftype='rf',threshold=0.15,all=FALSE)
```

#### **Arguments**

result The predicted result

filename The path where the result will be saved

header the index clftype classifier type

threshold the threshold of gencall score

all if the users want to save the full table or not

### Value

txt file of the results

```
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')
clf_rf_path = system.file('files/clf_30trees_7228_ratio1_lightweight.clf',package='SureTypeSCR')
clf_gda_path = system.file('files/clf_30trees_7228_ratio1_lightweight.clf',package='SureTypeSCR')

# The Random Forest classifier
clf_rf = scload(clf_rf_path)

# The Gaussian Disciminant Analysis
clf_gda = scload(clf_gda_path)

# get genotyping data from gtc files and meta file
```

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```
df <- scbasic(manifest_path,cluster_path,samplesheet)</pre>
# create Data object and rearrange the index
dfs <- create_from_frame(df)</pre>
#original shape (294602, 15)
#shape after operation (294602, 12)
\# extract the chromosomes 1 and 2
dfs <- restrict_chrom(dfs,c('1','2'))</pre>
\# mask the Gencall score lower than 0.01
dfs <- apply_thresh(dfs,0.01)</pre>
# calculate the m and a feature
dfs <- calculate_ma(dfs)</pre>
# prediction by Random Forest
result_rf <- scpredict(clf_rf,dfs,clftype='rf')</pre>
# prediction by Guassian Discriminate Analysis
result_gda <- scpredict(clf_gda,dfs,clftype='gda')</pre>
# Train the rf-gda classifier
trainer <- scTrain(result_gda,clfname='gda')</pre>
# The prediction from the cascade of Random Forest and Gaussian Discriminate Analysis
result_end <- scpredict(trainer,result_gda,clftype='rf-gda')</pre>
# Save the complete prediction table
scsave(result_end, 'fulltable.txt',clftype='rf',header=TRUE,threshold=0.15,all=TRUE)
# recall mode
scsave(result_end,'fulltable.txt',clftype='rf',threshold=0.15,header=TRUE,all=FALSE)
```

scTrain

Train Gaussian Discriminate Analysis by using the output of predicitons of Random Forest

# **Description**

Train Gaussian Discriminate Analysis by using the output of predicitons of Random Forest

```
scTrain(trainingdata, clfname='gda')
```

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#### **Arguments**

trainingdata Data object, the training data of scTrain

clfname The classifier type

#### Value

instance of a classifier

```
gtc_path = system.file("files/GTCs",package='SureTypeSCR')
cluster_path = system.file('files/HumanKaryomap-12v1_A.egt',package='SureTypeSCR')
manifest_path = system.file('files/HumanKaryomap-12v1_A.bpm',package='SureTypeSCR')
samplesheet = system.file('files/Samplesheetr.csv',package='SureTypeSCR')
clf_rf_path = system.file('files/clf_30trees_7228_ratio1_lightweight.clf',package='SureTypeSCR')
# The Random Forest classifier
clf_rf <- scload(clf_rf_path)</pre>
# The Gaussian Disciminant Analysis
clf_gda <- scload(clf_gda_path)</pre>
# get genotyping data from gtc files and meta file
df <- scbasic(manifest_path,cluster_path,samplesheet)</pre>
# create Data object and rearrange the index
dfs <- create_from_frame(df)</pre>
\# extract the chromosomes 1 and 2
dfs <- restrict_chrom(dfs,c('1','2'))</pre>
# mask the Gencall score lower than 0.01
dfs <- apply_thresh(dfs,0.01)</pre>
# calculate the m and a feature
dfs <- calculate_ma(dfs)</pre>
# prediction by Random Forest
result_rf <- scpredict(clf_rf,dfs,clftype='rf')</pre>
# prediction by Guassian Discriminate Analysis
result_gda <- scpredict(clf_gda,dfs,clftype='gda')</pre>
# Train the rf-gda classifier
trainer <- scTrain(result_rf,clfname='gda')</pre>
# The prediction from the cascade of Random Forest and Gaussian Discriminate Analysis
result_end <- scpredict(trainer,result_gda,clftype='rf-gda')</pre>
```

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