

# Class 10: Structural Bioinformatics (pt1)

Melanda Aboueid (PID: A17473102)

## PDB Statistics

The Protein Data Bank (PDB) is the main repository of biomolecular structures. Let's see what it contains:

```
stats <- read.csv("Data Export Summary.csv")
stats
```

	Molecular.Type	X.ray	EM	NMR	Integrative	Multiple.methods
1	Protein (only)	178,795	21,825	12,773	343	226
2	Protein/Oligosaccharide	10,363	3,564	34	8	11
3	Protein/NA	9,106	6,335	287	24	7
4	Nucleic acid (only)	3,132	221	1,566	3	15
5	Other	175	25	33	4	0
6	Oligosaccharide (only)	11	0	6	0	1
	Neutron	Other	Total			
1	84	32	214,078			
2	1	0	13,981			
3	0	0	15,759			
4	3	1	4,941			
5	0	0	237			
6	0	4	22			

```
stats$X.ray
```

```
[1] "178,795" "10,363" "9,106" "3,132" "175" "11"
```

```
sum(stats$Neutron)
```

```
[1] 88
```

The comma in these numbers leads to the numbers here being read as character.

```
c(100, 10, "barry")
```

```
[1] "100"    "10"     "barry"
```

```
library(readr)
stats <- read_csv("Data Export Summary.csv")
```

```
Rows: 6 Columns: 9
-- Column specification -----
Delimiter: ","
chr (1): Molecular Type
dbl (4): Integrative, Multiple methods, Neutron, Other
num (4): X-ray, EM, NMR, Total

i Use `spec()` to retrieve the full column specification for this data.
i Specify the column types or set `show_col_types = FALSE` to quiet this message.
```

```
stats
```

```
# A tibble: 6 x 9
`Molecular Type` `X-ray`   EM    NMR Integrative `Multiple methods` Neutron
<chr>           <dbl> <dbl> <dbl>      <dbl>          <dbl> <dbl>
1 Protein (only) 178795 21825 12773      343        226     84
2 Protein/Oligosacch~ 10363  3564   34        8         11      1
3 Protein/NA       9106   6335   287       24         7      0
4 Nucleic acid (only) 3132    221  1566       3        15      3
5 Other            175     25    33        4         0      0
6 Oligosaccharide (o~ 11      0     6        0         1      0
# i 2 more variables: Other <dbl>, Total <dbl>
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
n.xray <- sum(stats$`X-ray`)
#n.em <-
n.total <- sum(stats$Total)
n.xray/n.total
```

```
[1] 0.8095077
```

80%

```
n.em <- sum(stats$EM)
n.em/n.total
```

```
[1] 0.1283843
```

Q2: What proportion of structures in the PDB are protein?

```
n.protein <- stats[1, "Total"]
p_protein <- n.protein/n.total
p_protein
```

```
      Total
1 0.8596889
```

85% > Q3: SKIP

### **Visualizing the HIV-1 protease structure**

We can use the molstar viewer online: <https://molstar.org/viewer/>.

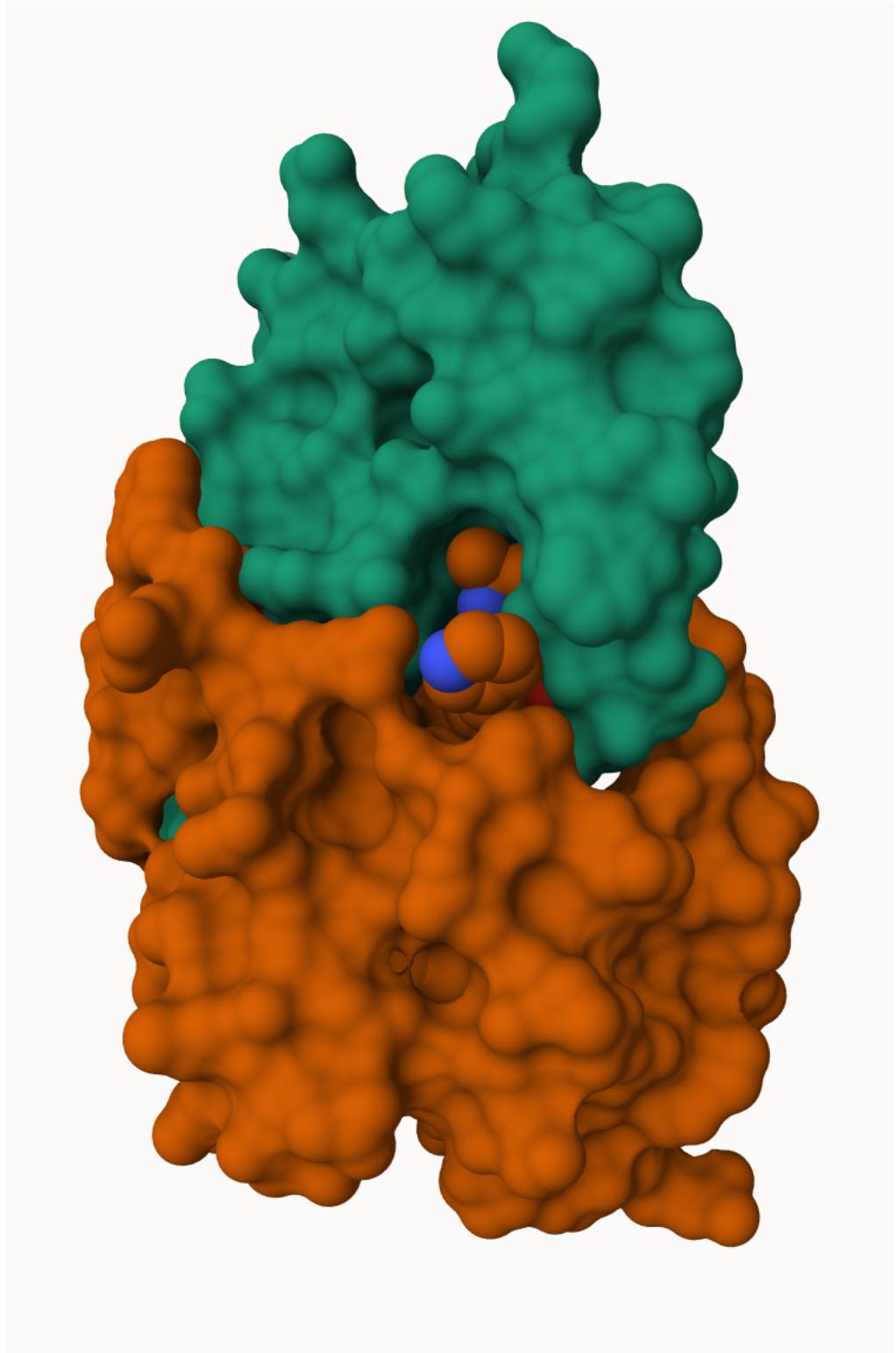
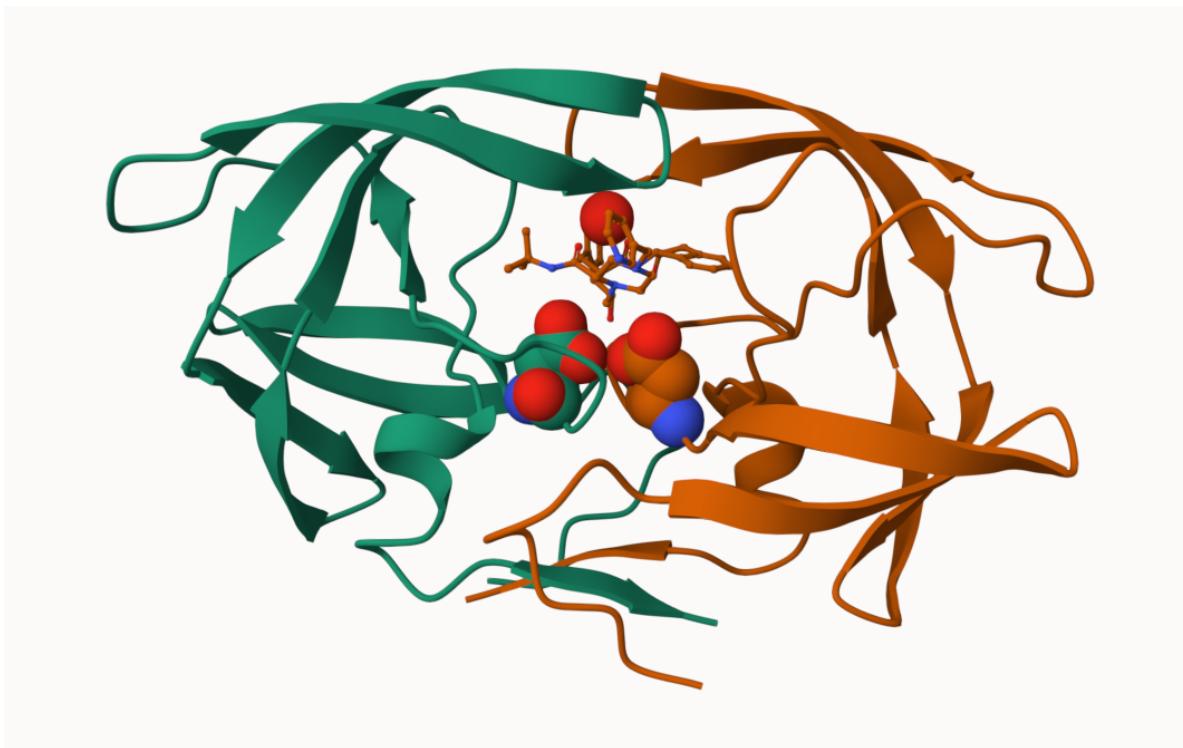


Figure 1: My first image of HIV-Pr with surface display showing ligand binding

A new clean image showing the catalytic ASP25 amino acids in both chains of the HIV-PR dimer along with the inhibitor and the all important active site water



### Bio3D package for structural bioinformatics

```
library(bio3d)
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
 pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPCKMIGGIGGFVKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWPCKMIGGIGGFVKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

```
head( pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

```
#library(bio3dview)
```

```
#view.pdb(pdb)
```

## Predicting functional motions of a single structure

Read an ADK structure from the PDB database

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file  
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: read.pdb(file = "6s36")

Total Models#: 1

Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]

Protein sequence:

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMLRAAVKSGSELGKQAKDIDMAGKLVT  
DELVIALVKERIAQEDCRNGFLLDGFPRТИPQADAMKEAGINVDYVLEFDVPDELVDKI  
VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

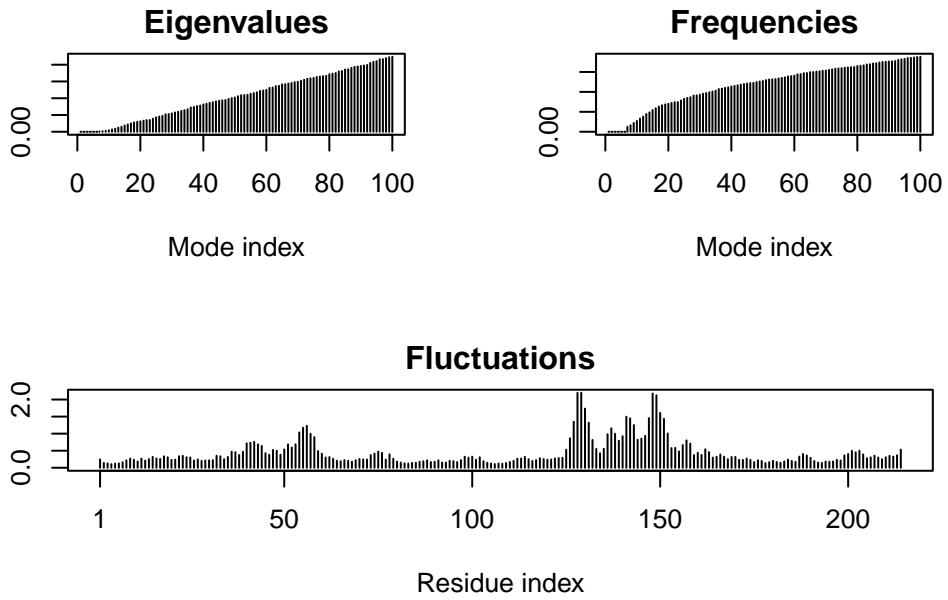
+ attr: atom, xyz, seqres, helix, sheet,  
calpha, remark, call

```
m <- nma(adk)
```

Building Hessian... Done in 0.011 seconds.

Diagonalizing Hessian... Done in 0.045 seconds.

```
plot(m)
```



Write out results as a wee trajectory/movie of predicted motions:

```
mktrj(m, file="adk_m7.pdb")
```