

R Notebook

Code ▾

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```
#Loading the data file for Regression
attach(parkinsons_updrs)
```

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```
#Loading and cleaning the data file for Classification
attach(breast.cancer.wisconsin)

#Missing Value check and Solution
breast.cancer.wisconsin$V7 <- replace(breast.cancer.wisconsin$V7, breast.cancer.wisconsin$V7
== "?", NA)
breast.cancer.wisconsin$V7 <- as.integer(breast.cancer.wisconsin$V7)

breast_cancer_wisconsin<-na.omit(breast.cancer.wisconsin)
breast_cancer_wisconsin
```

	V1	V2	V3	V4	V5	V6	V7	V8	V9	
	<int>	<int>	<int>	<int>	<int>	<int>	<int>	<int>	<int>	
1	1000025	5	1	1	1	2	1	3	1	
2	1002945	5	4	4	5	7	10	3	2	
3	1015425	3	1	1	1	2	2	3	1	
4	1016277	6	8	8	1	3	4	3	7	
5	1017023	4	1	1	3	2	1	3	1	
6	1017122	8	10	10	8	7	10	9	7	
7	1018099	1	1	1	1	2	10	3	1	
8	1018561	2	1	2	1	2	1	3	1	
9	1033078	2	1	1	1	2	1	1	1	
10	1033078	4	2	1	1	2	1	2	1	
1-10 of 683 rows 1-10 of 11 columns				Previous	1	2	3	4	5	6 ... 69 Next

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```
summary(parkinsons_updrs)
```

subject.	age	sex	test_time
Min. : 1.00	Min. :36.0	Min. :0.0000	Min. : -4.263
1st Qu.:10.00	1st Qu.:58.0	1st Qu.:0.0000	1st Qu.: 46.847
Median :22.00	Median :65.0	Median :0.0000	Median : 91.523
Mean :21.49	Mean :64.8	Mean :0.3178	Mean : 92.864
3rd Qu.:33.00	3rd Qu.:72.0	3rd Qu.:1.0000	3rd Qu.:138.445
Max. :42.00	Max. :85.0	Max. :1.0000	Max. :215.490
motor_UPDRS	total_UPDRS	Jitter...	Jitter.Abs.
Min. : 5.038	Min. : 7.00	Min. :0.000830	Min. :2.250e-06
1st Qu.:15.000	1st Qu.:21.37	1st Qu.:0.003580	1st Qu.:2.244e-05
Median :20.871	Median :27.58	Median :0.004900	Median :3.453e-05
Mean :21.296	Mean :29.02	Mean :0.006154	Mean :4.403e-05
3rd Qu.:27.596	3rd Qu.:36.40	3rd Qu.:0.006800	3rd Qu.:5.333e-05
Max. :39.511	Max. :54.99	Max. :0.099990	Max. :4.456e-04
Jitter.RAP	Jitter.PPQ5	Jitter.DDP	
Min. :0.000330	Min. :0.000430	Min. :0.000980	
1st Qu.:0.001580	1st Qu.:0.001820	1st Qu.:0.004730	
Median :0.002250	Median :0.002490	Median :0.006750	
Mean :0.002987	Mean :0.003277	Mean :0.008962	
3rd Qu.:0.003290	3rd Qu.:0.003460	3rd Qu.:0.009870	
Max. :0.057540	Max. :0.069560	Max. :0.172630	
Shimmer	Shimmer.dB.	Shimmer.APQ3	Shimmer.APQ5
Min. :0.00306	Min. :0.026	Min. :0.00161	Min. :0.00194
1st Qu.:0.01912	1st Qu.:0.175	1st Qu.:0.00928	1st Qu.:0.01079
Median :0.02751	Median :0.253	Median :0.01370	Median :0.01594
Mean :0.03404	Mean :0.311	Mean :0.01716	Mean :0.02014
3rd Qu.:0.03975	3rd Qu.:0.365	3rd Qu.:0.02057	3rd Qu.:0.02375
Max. :0.26863	Max. :2.107	Max. :0.16267	Max. :0.16702
Shimmer.APQ11	Shimmer.DDA	NHR	HNR
Min. :0.00249	Min. :0.00484	Min. :0.000286	Min. : 1.659
1st Qu.:0.01566	1st Qu.:0.02783	1st Qu.:0.010955	1st Qu.:19.406
Median :0.02271	Median :0.04111	Median :0.018448	Median :21.920
Mean :0.02748	Mean :0.05147	Mean :0.032120	Mean :21.680
3rd Qu.:0.03272	3rd Qu.:0.06173	3rd Qu.:0.031463	3rd Qu.:24.444
Max. :0.27546	Max. :0.48802	Max. :0.748260	Max. :37.875
RPDE	DFA	PPE	
Min. :0.1510	Min. :0.5140	Min. :0.02198	
1st Qu.:0.4698	1st Qu.:0.5962	1st Qu.:0.15634	
Median :0.5423	Median :0.6436	Median :0.20550	
Mean :0.5415	Mean :0.6532	Mean :0.21959	
3rd Qu.:0.6140	3rd Qu.:0.7113	3rd Qu.:0.26449	
Max. :0.9661	Max. :0.8656	Max. :0.73173	

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```
#Descriptive Dataset's Info
```

```
#Class Label- 2 - Benign and 4- Malignant
```

```
names(breast_cancer_wisconsin)<-c('ID', 'C_Thickness', 'UCSize', 'UCShape', 'M_Adhesion', 'Single_ECSize', 'Bare_Nuclei', 'Bland_Chromatin', 'Normal_Nucleoli', 'Mitoses', 'Class_Label')
```

```
#Encondings of Class Label
```

```
breast_cancer_wisconsin$Class_Label<-ifelse(breast_cancer_wisconsin$Class_Label== 2, 0, 1)
breast_cancer_wisconsin$Class_Label
```

```
[1] 0 0 0 0 0 1 0 0 0 0 0 0 1 0 1 1 0 0 1 0 1 1 0 0 1 0 0 0 0 0 1 0 0 0
[36] 1 0 1 1 1 1 1 1 0 1 0 0 1 1 1 1 1 1 1 1 1 0 1 1 0 1 0 1 1 0 0 1
[71] 0 1 1 0 0 0 0 0 0 0 0 0 1 1 1 1 0 0 0 0 0 0 0 0 0 0 1 1 1 1 0 1 1 1
[106] 1 0 1 0 1 1 1 0 0 0 1 0 0 0 0 1 1 1 0 1 0 1 0 0 0 1 0 0 0 0 0 0 0 1
[141] 0 0 1 0 0 1 0 1 1 0 0 1 0 0 1 1 0 0 0 0 1 1 0 0 0 0 0 1 1 1 0 1 0 1 0
[176] 0 0 1 1 0 1 1 1 0 1 1 0 0 0 0 0 0 0 0 0 1 1 0 0 0 1 1 0 0 0 1 1 0 1 1 1
[211] 0 0 1 0 0 1 1 1 1 0 1 1 0 1 1 1 0 1 0 1 1 1 1 0 0 0 0 0 0 1 1 0 0 1 0
[246] 1 1 1 0 0 0 0 1 1 1 1 1 0 1 1 1 0 1 0 1 1 0 0 0 0 1 0 0 1 1 1 1 0 1
[281] 1 0 0 1 1 0 0 1 1 0 1 0 1 1 0 0 1 0 0 0 1 0 0 1 1 0 0 1 0 1 0 0 1 0 1
[316] 1 1 0 0 1 1 0 1 0 0 1 1 0 0 0 1 0 0 0 1 1 0 0 0 1 0 0 1 1 1 1 1 1 0 0
[351] 0 0 1 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 0 1 0 0 0 0 1 0 0 0 0 0 0
[386] 0 1 0 0 0 0 0 0 0 0 0 0 0 1 0 1 0 1 0 0 0 0 1 0 0 0 1 0 1 0 0 0 0 0 0
[421] 1 1 0 0 0 1 0 0 0 0 0 0 0 0 0 1 0 0 0 1 0 1 1 1 0 0 0 0 0 0 0 1 1 1 0 0
[456] 0 0 0 0 0 0 0 0 0 1 0 0 1 1 0 0 0 1 1 1 0 1 0 1 0 0 0 0 0 0 0 0 0 0 0
[491] 0 1 0 0 0 0 0 0 0 1 1 0 0 0 1 0 0 1 1 0 0 0 0 0 0 1 0 0 0 0 0 0 0 0 0
[526] 0 0 0 0 0 0 1 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 1 1 1 1 0 0 1
[561] 0 0 0 0 0 0 1 1 0 0 0 1 0 1 0 1 1 1 0 1 0 0 0 0 0 0 0 0 1 1 1 0 0 1 0
[596] 1 1 1 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 0 0 0 1 0 0 1 0 0 0 0 0 0 0 0 0
[631] 0 0 1 0 0 0 0 0 0 0 0 0 1 0 0 0 0 0 0 0 0 0 1 1 1 0 0 0 0 0 0 0 0 0 1
[666] 1 0 0 0 0 0 0 0 0 0 1 0 0 0 0 1 1 1
```

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```
summary(breast_cancer_wisconsin)
```

ID	C_Thickness	UCSize	UCShape
Min. : 63375	Min. : 1.000	Min. : 1.000	Min. : 1.000
1st Qu.: 877617	1st Qu.: 2.000	1st Qu.: 1.000	1st Qu.: 1.000
Median : 1171795	Median : 4.000	Median : 1.000	Median : 1.000
Mean : 1076720	Mean : 4.442	Mean : 3.151	Mean : 3.215
3rd Qu.: 1238705	3rd Qu.: 6.000	3rd Qu.: 5.000	3rd Qu.: 5.000
Max. : 13454352	Max. : 10.000	Max. : 10.000	Max. : 10.000

M_Adhesion	Single_ECSIZE	Bare_Nuclei	Bland_Chromatin
Min. : 1.00	Min. : 1.000	Min. : 1.000	Min. : 1.000
1st Qu.: 1.00	1st Qu.: 2.000	1st Qu.: 1.000	1st Qu.: 2.000
Median : 1.00	Median : 2.000	Median : 1.000	Median : 3.000
Mean : 2.83	Mean : 3.234	Mean : 3.545	Mean : 3.445
3rd Qu.: 4.00	3rd Qu.: 4.000	3rd Qu.: 6.000	3rd Qu.: 5.000
Max. : 10.00	Max. : 10.000	Max. : 10.000	Max. : 10.000

Normal_Nucleoli	Mitoses	Class_Label
Min. : 1.00	Min. : 1.000	Min. : 0.0000
1st Qu.: 1.00	1st Qu.: 1.000	1st Qu.: 0.0000
Median : 1.00	Median : 1.000	Median : 0.0000
Mean : 2.87	Mean : 1.603	Mean : 0.3499
3rd Qu.: 4.00	3rd Qu.: 1.000	3rd Qu.: 1.0000
Max. : 10.00	Max. : 10.000	Max. : 1.0000

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```
#Divide the data into training and testing
```

```
library(caret)
```

```
Loading required package: ggplot2
Loading required package: lattice
Registered S3 method overwritten by 'data.table':
  method      from
print.data.table
```

Hide

```
set.seed(1) # for reproducibility

# Create a vector of row indices
rows <- 1:nrow(parkinsons_updrs)

# Randomly sample 80% of the row indices for the training set
training_rows <- sample(rows, floor(0.8 * length(rows)))

# The remaining rows are for the testing set
testing_rows <- setdiff(rows, training_rows)

# Write the training and testing sets to separate files
write.table(parkinsons_updrs[training_rows, ], file = "Park_training_data.txt", row.names = F
ALSE, col.names = FALSE)
write.table(parkinsons_updrs[testing_rows, ], file = "Park_testing_data.txt", row.names = FAL
SE, col.names = FALSE)

training_data_old <- parkinsons_updrs[training_rows, ]
testing_data_old <- parkinsons_updrs[-training_rows, ]

# Remove the variable 'motor_UPDRS' (Training and Testing)
training_data <- subset(training_data_old, select = -motor_UPDRS)
testing_data <- subset(testing_data_old, select = -motor_UPDRS)

# Create X.train and X.test data frames that exclude the total_UPDRS
X.train <- training_data[, -which(names(training_data) == "total_UPDRS")]
X.test <- testing_data[, -which(names(testing_data) == "total_UPDRS")]
Y.train <- training_data$total_UPDRS
Y.test <- testing_data$total_UPDRS

#b) Division Verification in number of Examples
cat("Number of examples in training data:", nrow(training_data), "\n")
```

```
Number of examples in training data: 4700
```

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```
cat("Number of examples in testing data:", nrow(testing_data), "\n")
```

```
Number of examples in testing data: 1175
```

Hide

```
#Divide the data into training(80%) and testing(20%)

breast_cancer_wisconsin$Class_Label <- factor(breast_cancer_wisconsin$Class_Label)

#a) Divide the data into training and testing sets
library(caret)
set.seed(1) # for reproducibility

# Create a vector of row indices
rows1 <- 1:nrow(breast_cancer_wisconsin)

# Randomly sample 80% of the row indices for the training set
training_rows1 <- sample(rows1, floor(0.8 * length(rows1)))

# The remaining rows are for the testing set
testing_rows1 <- setdiff(rows1, training_rows1)

training_data1 <- breast_cancer_wisconsin[training_rows1, ]
testing_data1 <- breast_cancer_wisconsin[-training_rows1, ]

# Create X.train and X.test data frames that exclude the class label
X.train1 <- training_data1[, -which(names(training_data1) == "Class_Label")]
X.test1 <- testing_data1[, -which(names(testing_data1) == "Class_Label")]
Y.train1 <- training_data1$Class_Label
Y.test1 <- testing_data1$Class_Label

#b) Division Verification in number of Examples
cat("Number of examples in training data:", nrow(training_data1), "\n")
```

Number of examples in training data: 546

[Hide](#)

```
cat("Number of examples in testing data:", nrow(testing_data1), "\n")
```

Number of examples in testing data: 137

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```
#1.a) Bagging - Regression
library(randomForest)
```

```
randomForest 4.7-1.1
Type rfNews() to see new features/changes/bug fixes.
```

Attaching package: ‘randomForest’

The following object is masked from ‘package:ggplot2’:

margin

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```
# Bagging is just and RF with m=p
set.seed(1)
# mtry: number of predictors (m) to be taken into account in the model
bag.park=randomForest(total_UPDRS~.,training_data,mtry=20,importance=T)
bag.park
```

Call:

```
randomForest(formula = total_UPDRS ~ ., data = training_data,      mtry = 20, importance =
T)
```

Type of random forest: regression

Number of trees: 500

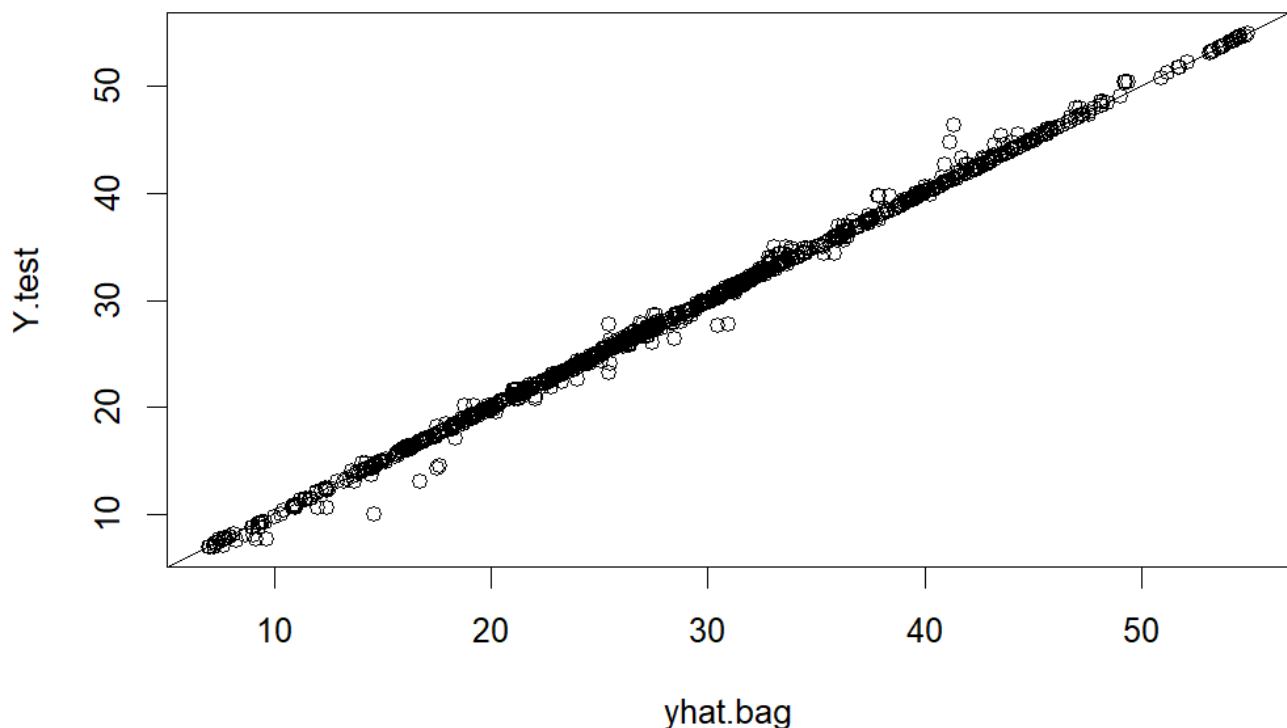
No. of variables tried at each split: 20

Mean of squared residuals: 0.2014956

% Var explained: 99.82

Hide

```
# Performance on test set
yhat.bag=predict(bag.park,testing_data)
plot(yhat.bag, Y.test)
abline(0,1)
```



Hide

```
#i)Performance Test MSE
mean((yhat.bag-Y.test)^2)
```

```
[1] 0.2111907
```

Hide

```
#####
# b) Random Forests - Regression

library(randomForest)
# By default randomForest() uses m=p/3 for regression and m=sqrt(p) for classification
set.seed(1)
rf.park=randomForest(total_UPDRS~.,training_data,mtry=20/3,importance =T)
yhat.rf = predict(rf.park ,testing_data)

#i) Test MSE
mean((yhat.rf-Y.test)^2)
```

```
[1] 2.827568
```

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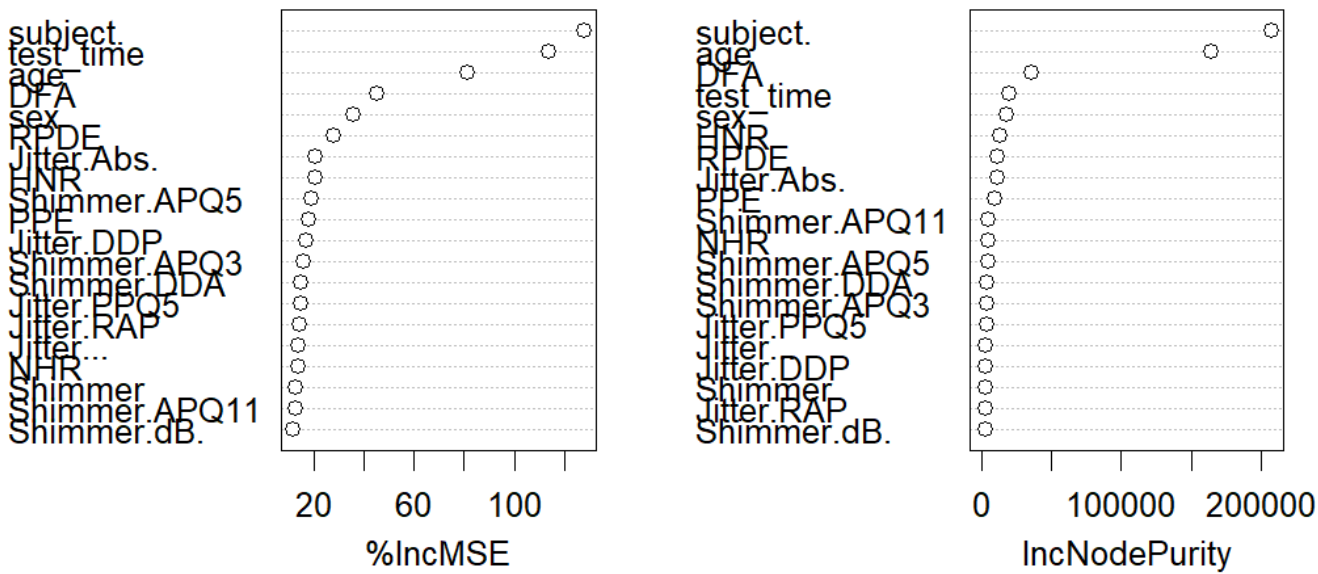
```
# importance(): view the importance of each variable
# %IncMSE: mean decrease of accuracy in predictions on the OOB samples when a
# given variable is excluded from the model
# IncNodeImpurity: total decrease in node impurity that results from splits over
# that variable, averaged over all trees (RSS in regr. vs. deviance in class)
importance(rf.park)
```

	%IncMSE	IncNodePurity
subject.	128.04524	207920.402
age	81.36791	164116.633
sex	35.83029	18302.275
test_time	114.15915	20017.791
Jitter...	13.80827	3562.756
Jitter.Abs.	20.67854	11222.622
Jitter.RAP	14.19765	3221.542
Jitter.PPQ5	14.66408	3938.261
Jitter.DDP	16.56443	3501.982
Shimmer	12.70978	3276.419
Shimmer.dB.	11.61624	3045.770
Shimmer.APQ3	15.51536	4003.207
Shimmer.APQ5	18.87825	4567.601
Shimmer.APQ11	12.56983	5045.740
Shimmer.DDA	14.82533	4172.176
NHR	13.67961	4998.148
HNR	20.19554	13443.800
RPDE	27.98000	11823.178
DFA	45.28773	36006.964
PPE	17.58112	9210.416

Hide

```
# varImpPlot(): Variance importance plot
varImpPlot(rf.park)
```

rf.park



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#c) Boosting - Regression

```
# gbm: library for boosting
library(gbm)
```

Warning: package ‘gbm’ was built under R version 4.2.3Loaded gbm 2.1.8.1

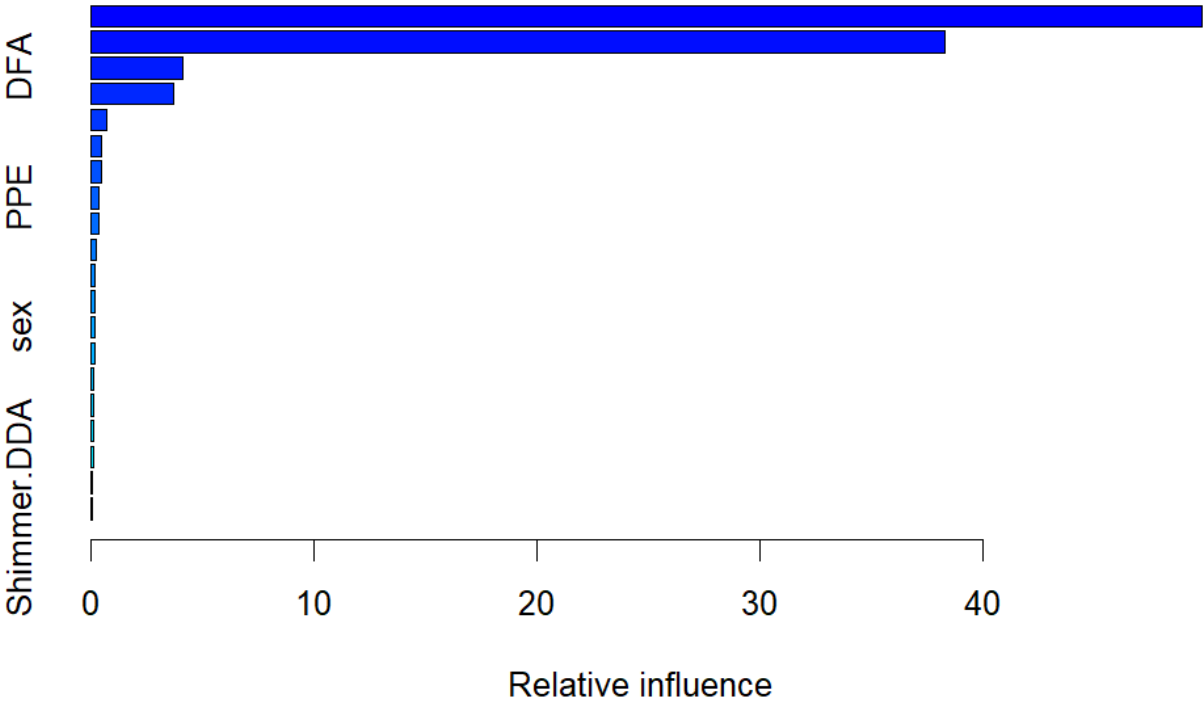
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```
set.seed(1)
# Since this is a regression problem, we set the distribution to "gaussian"
# For binary classification, we would use "bernoulli"
# n.trees: number of trees we want
# interaction.depth: limits the depth of each tree
boost.park=gbm(total_UPDRS~.,data=training_data,distribution="gaussian",n.trees=5000, interac
tion.depth=4)

# In this case, summary() produces the relative influence plot and outputs
# the relative influence statistics
summary(boost.park)
```

	var <chr>	rel.inf <dbl>
	subject.	49.81842382
	age	38.28416168
	DFA	4.14339575

	var<chr>	rel.inf<dbl>
test_time	test_time	3.69974374
HNR	HNR	0.73524494
Shimmer.APQ5	Shimmer.APQ5	0.49513030
RPDE	RPDE	0.45997826
PPE	PPE	0.36989422
Jitter.Abs.	Jitter.Abs.	0.33782891
Shimmer.APQ11	Shimmer.APQ11	0.26981946
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```
#MSE Test
# Performance on the test set
yhat.boost=predict(boost.park,testing_data,n.trees=5000)
mean((yhat.boost -Y.test)^2)
```

```
[1] 0.8775685
```

Hide

```
# We can set the shrinkage parameter (i.e., lambda). Default value: 0.001
boost.park1=gbm(total_UPDRS~.,training_data,distribution= "gaussian",
                n.trees=5000,interaction.depth=4,shrinkage=0.2,verbose=F)
yhat.boost1=predict(boost.park1,testing_data,n.trees=5000)

#MSE Test
# Performance on the test set
mean((yhat.boost1-Y.test)^2)
```

```
[1] 1.094136
```

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#d) Compare and comment on the error obtained with each approach. Which model seems to perform the best?

Overall bagging performed better compared to boosting and random forests as it resulted in a average value for MSE of 0.21 vs random forests 2.92 and boosting 0.88 which performed more closely to bagging.

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```
# 1. a) Decision Tree Classification
library(tree)
```

```
# Performance on the test set
# tree(): Fit a classification tree
# Similar syntax to lm()
```

```
tree.breast=tree(Class_Label~.,training_data1)
```

```
# summary() lists the variables that are used as internal nodes in the tree,
# the number of terminal nodes, and the (training) error rate
# A small deviance indicates a tree that provides a good fit to the (training) data
summary(tree.breast)
```

Classification tree:

```
tree(formula = Class_Label ~ ., data = training_data1)
```

Variables actually used in tree construction:

```
[1] "UCSize"          "Bare_Nuclei"      "Bland_Chromatin" "C_Thickness"
[5] "Normal_Nucleoli" "ID"               "M_Adhesion"
```

Number of terminal nodes: 11

Residual mean deviance: 0.1006 = 53.8 / 535

Misclassification error rate: 0.01832 = 10 / 546

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```
# The object name prints split criterion, the number of observations
# in that branch, the deviance, the overall prediction for the branch, and the
# fraction of observations in that branch that take on values of Yes and No.
# Branches that lead to terminal nodes are indicated using asterisks
tree.breast
```

```
node), split, n, deviance, yval, (yprob)
  * denotes terminal node
```

```
1) root 546 716.300 0 ( 0.635531 0.364469 )
 2) USize < 2.5 328 96.320 0 ( 0.966463 0.033537 )
   4) Bare_Nuclei < 3.5 308 13.460 0 ( 0.996753 0.003247 )
     8) Bland_Chromatin < 3.5 303 0.000 0 ( 1.000000 0.000000 ) *
     9) Bland_Chromatin > 3.5 5 5.004 0 ( 0.800000 0.200000 ) *
   5) Bare_Nuclei > 3.5 20 27.730 0 ( 0.500000 0.500000 )
     10) C_Thickness < 3.5 9 0.000 0 ( 1.000000 0.000000 ) *
     11) C_Thickness > 3.5 11 6.702 1 ( 0.090909 0.909091 ) *
 3) USize > 2.5 218 174.700 1 ( 0.137615 0.862385 )
   6) Bare_Nuclei < 3.5 54 74.560 1 ( 0.462963 0.537037 )
     12) USize < 4.5 33 36.550 0 ( 0.757576 0.242424 )
       24) Normal_Nucleoli < 1.5 15 0.000 0 ( 1.000000 0.000000 ) *
       25) Normal_Nucleoli > 1.5 18 24.730 0 ( 0.555556 0.444444 )
         50) ID < 1.17042e+06 9 9.535 1 ( 0.222222 0.777778 ) *
         51) ID > 1.17042e+06 9 6.279 0 ( 0.888889 0.111111 ) *
     13) USize > 4.5 21 0.000 1 ( 0.000000 1.000000 ) *
 7) Bare_Nuclei > 3.5 164 44.750 1 ( 0.030488 0.969512 )
   14) C_Thickness < 6.5 65 35.250 1 ( 0.076923 0.923077 )
     28) M_Adhesion < 5.5 28 26.280 1 ( 0.178571 0.821429 ) *
     29) M_Adhesion > 5.5 37 0.000 1 ( 0.000000 1.000000 ) *
   15) C_Thickness > 6.5 99 0.000 1 ( 0.000000 1.000000 ) *
```

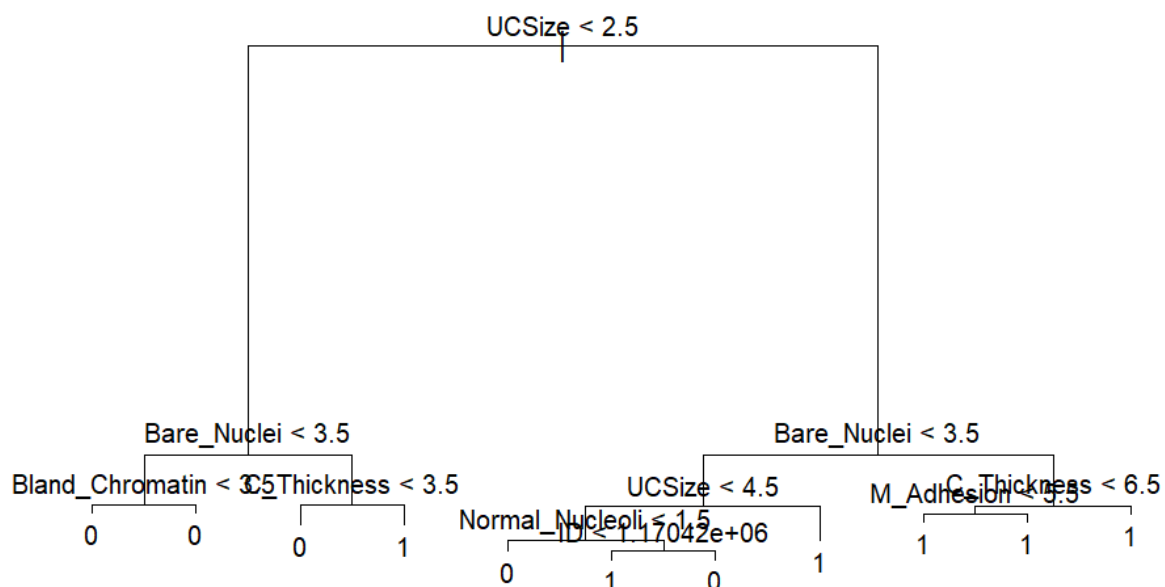
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```
# Test on test set using predict()
# type="class" to return the class prediction
tree.pred=predict(tree.breast,testing_data1,type="class")
```

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```
# plot() to display the tree structure
plot(tree.breast)

# text() to display the node labels
# pretty=0: include the category names for any qualitative predictors,
# rather than simply displaying a letter for each category
# cex for font size
text(tree.breast, pretty = 0, cex=0.75)
```



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```
# Confusion matrix
conf.matrix <- table(tree.pred,Y.test1)
conf.matrix
```

```
      Y.test1
tree.pred 0  1
      0 91  1
      1  6 39
```

Hide

```
#i) Accuracy on test set
(conf.matrix[1,1] + conf.matrix[2,2])/200
```

```
[1] 0.65
```

Hide

```
##b) Tree pruning

# cv.tree(): CV to determine the optimal level of tree complexity
# FUN=prune.misclass: classification error rate to guide the CV and pruning process
# default: deviance
set.seed(3)

cv.breast=cv.tree(tree.breast,FUN=prune.misclass)

# Number of terminal nodes of each tree considered (size)
cv.breast$size
```

```
[1] 11  8  6  4  2  1
```

[Hide](#)

```
# Corresponding CV error rate
cv.breast$dev
```

```
[1] 19 19 28 29 35 199
```

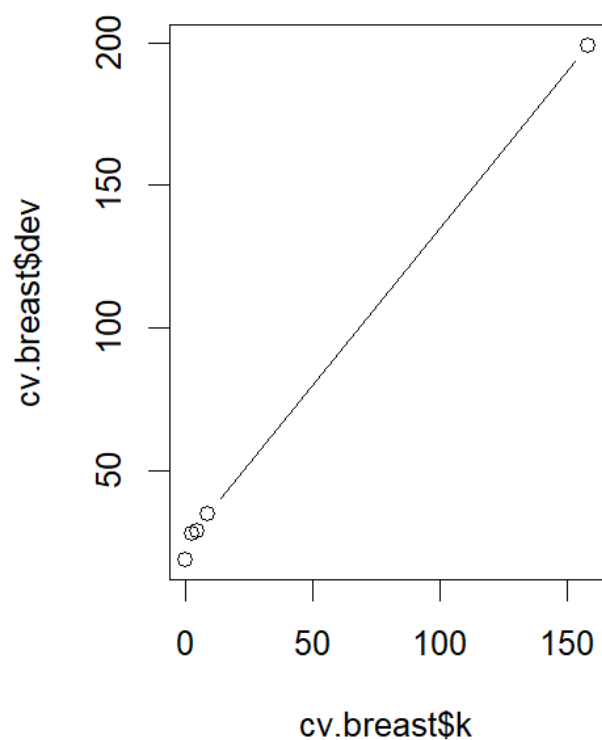
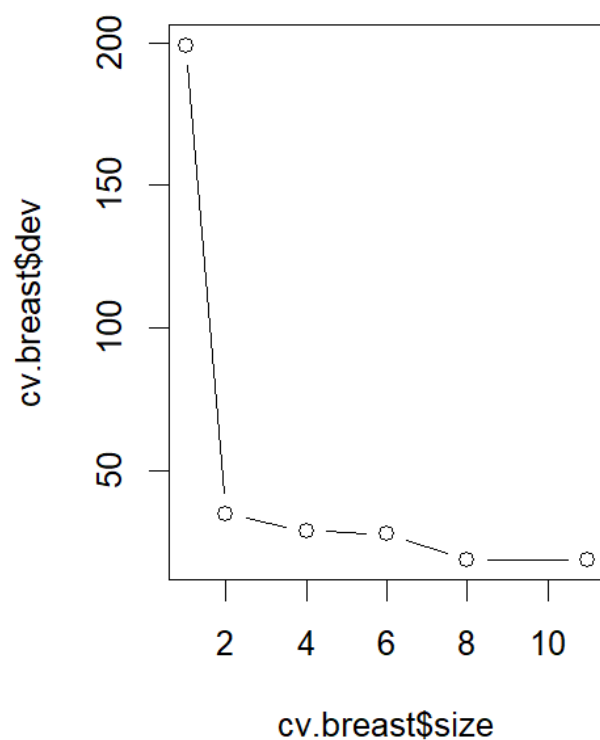
[Hide](#)

```
# Value of the cost-complexity parameter used (k here, alpha in formula - see 8.4)
cv.breast$k
```

```
[1] -Inf  0.0  2.5  4.5  8.5 158.0
```

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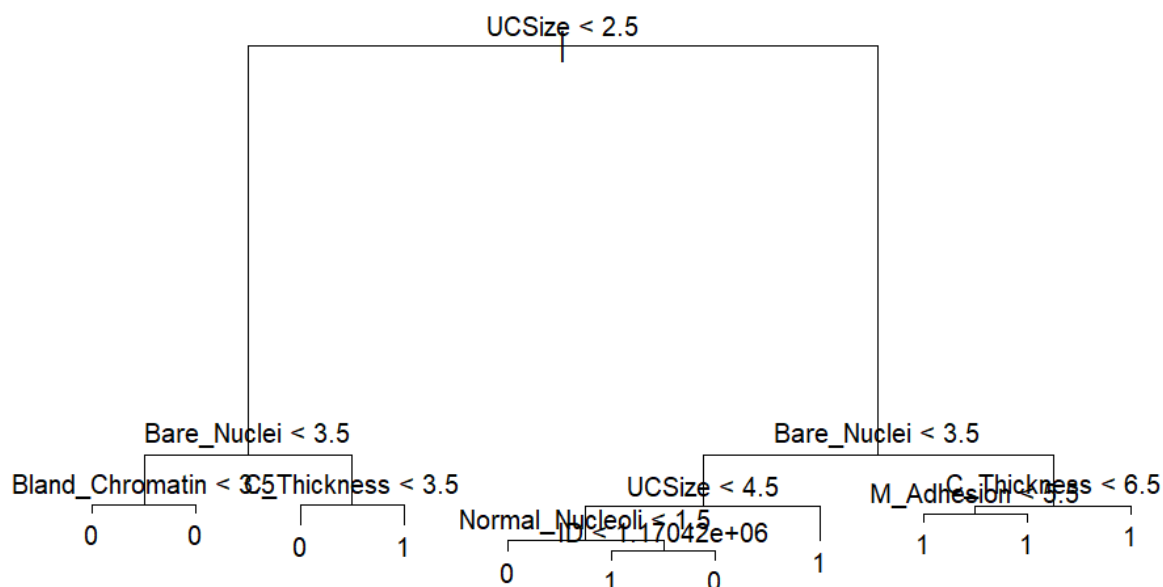
```
# Plot the error rate as a function of both size and k
par(mfrow=c(1,2))
plot(cv.breast$size, cv.breast$dev, type="b")
plot(cv.breast$k, cv.breast$dev, type="b")
```

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```
# prune.misclass(): prune the tree based on the CV error
prune.breast=prune.misclass(tree.breast,best=11)
```

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```
# Plot resulting decision tree
plot(prune.breast)
text(prune.breast, pretty=0, cex=0.75)
```



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```
# Let's check how well the pruned tree performs on the test set
tree.pred=predict(prune.breast,testing_data1,type="class")
conf.matrix1 <- table(tree.pred,Y.test1)
conf.matrix1
```

```
      Y.test1
tree.pred 0  1
0      91  1
1       6 39
```

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```
#ii)Prune Accuracy
(conf.matrix1[1,1] + conf.matrix1[2,2])/200
```

```
[1] 0.65
```