

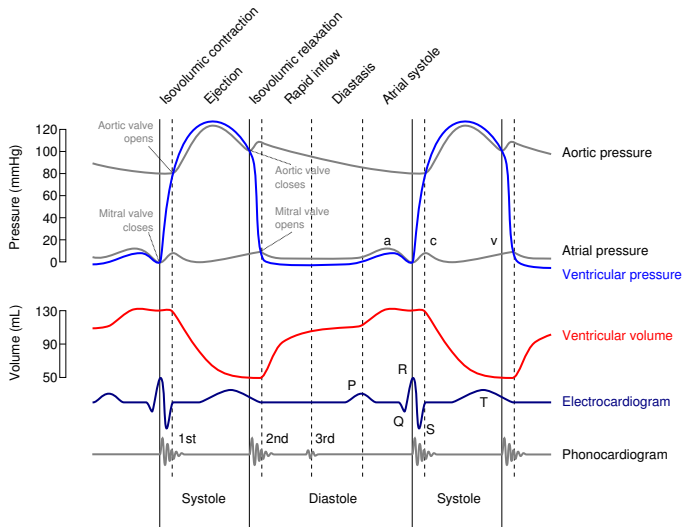
Fundamental cardiac mechanics  
Part I: Ode models of circulation and cell  
contraction

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Jun 17, 2016

- The heart as a pump
  - The pressure volume loop
  - Phases of the cardiac cycle
  - Elastance models of the heart and circulation
- Introduction to sliding filament theory
  - Micro-structure of force development
  - Cross-bridge cycling
  - Regulation of force
  - Models of cell contraction

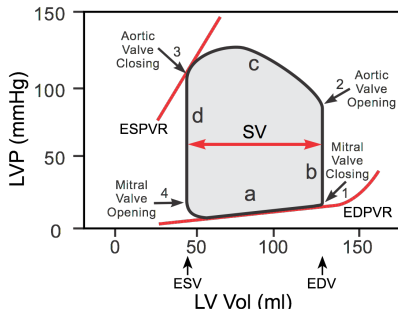
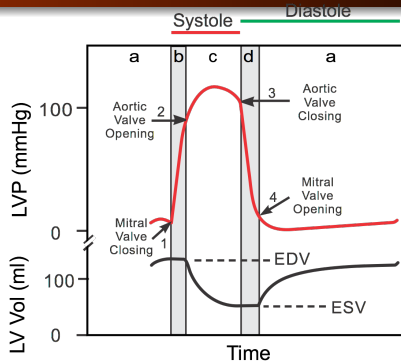
# Overview of cardiac function (1)



# The four phases of the heart cycle

- Passive filling; the muscle is relaxed and is filled with blood from the venous system (and the atria). Increase of pressure (small) and volume (large)
- Isovolumic contraction; the heart muscle contracts while all valves are closed. The cavity pressure increases while the volume stays constant
- Ejection; the valves open to allow blood to be ejected into the arteries. Pressure increases at first, then drops. Volume decreases
- Isovolumic relaxation; the muscle is relaxing while all valves are closed. The volume remains constant while the pressure drops

# The pressure-volume loop



# Why model all this?

- Understand fundamental mechanisms
- Investigate pathologies and interventions:
  - What happens if ...?
- Linking cell level processes to overall heart function (advanced)

# How can it be modeled?

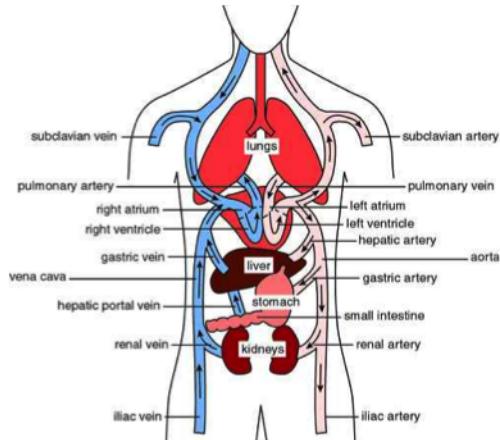
- Top-down (Today (June 17), morning lecture):
  - Derive relations for macroscopic function of the heart and other parts of the circulatory system
- Bottom-up/multiscale (Late morning/afternoon):
  - Model the biophysics of cell contraction
  - Integrate into tissue/organ models
  - Couple to models of circulation

# Modeling the heart and circulation

- Overview of the circulatory system
- Important quantities
- Resistance and compliance vessels
- Models for the circulatory system
- Examples and extensions



# The circulatory system



## Important quantities (1)

- Heart rate, measured in beats per minute.
- Cardiac output: The rate of blood flow through the circulatory system, measured in liters/minute.
- Stroke volume: the difference between the end-diastolic volume and the end-systolic volume, i.e. the volume of blood ejected from the heart during a heart beat, measured in liters.

## Important quantities (2)

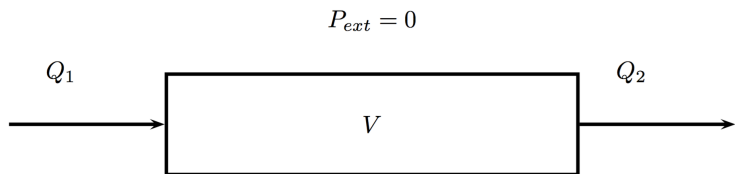
The cardiac output  $Q$  is given by

$$Q = FV_{stroke}$$

Typical values:

- $F = 80$  beats/minute.
- $V_{stroke} = 70\text{cm}^3/\text{beat} = 0.070$  liters/beat.
- $Q = 5.6$  liters/minute.

# Resistance and compliance vessels



- $V$  = vessel volume,
- $P_{ext}$  = external pressure,
- $P_1$  = upstream pressure,
- $P_2$  = downstream pressure,
- $Q_1$  = inflow,
- $Q_2$  = outflow.

Assume that the vessel is rigid, so that  $V$  is constant. Then we have

$$Q_1 = Q_2 = Q_*.$$

The flow through the vessel will depend on the pressure drop through the vessel. The simplest assumption is that  $Q_*$  is a linear function of the pressure difference  $P_1 - P_2$ :

$$Q_* = \frac{P_1 - P_2}{R},$$

where  $R$  is the resistance of the vessel.

Assume that the resistance over the vessel is negligible. This gives

$$P_1 = P_2 = P_*$$

Assume further that the volume depends on the pressure  $P_*$ . We assume the simple linear relation

$$V = V_d + CP_*,$$

where  $C$  is the compliance of the vessel and  $V_d$  is the “dead volume”, the volume at  $P_* = 0$ .

# Vessels in the circulation

- All blood vessels can be viewed as either resistance vessels or compliance vessels. (This is a reasonable assumption, although all vessels have both compliance and resistance.)
- Large arteries and veins; negligible resistance, significant compliance.
- Arterioles and capillaries; negligible compliance, significant resistance.

# The heart as a compliance vessel (1)

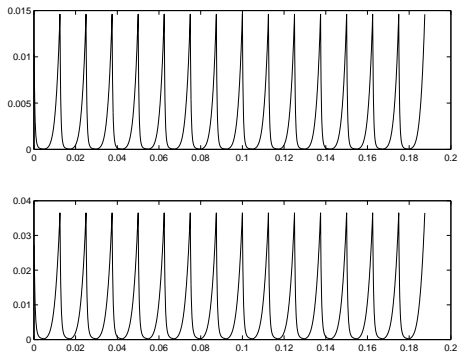
The heart may be viewed as a pair of compliance vessels, where the compliance changes with time,

$$V(t) = V_d + C(t)P.$$

The function  $V(t)$  should be specified so that it takes on a large value  $C_{diastole}$  when the heart is relaxed, and a small value  $C_{systole}$  when the heart contracts.



# The heart as a compliance vessel (2)



Characteristic properties of a heart valve:

- Low resistance for flow in the “forward” direction.
- High resistance for flow in the “backward” direction.

## Modeling the heart valves (2)

The operation of the valve can be seen as a switching function that depends on the pressure difference across the valve. The switching function can be expressed as

$$S = \begin{cases} 1 & \text{if } P_1 > P_2 \\ 0 & \text{if } P_1 < P_2 \end{cases}$$

## Modeling the heart valves (3)

The flow through the valve can be modeled as flow through a resistance vessel multiplied by the switching function. We have

$$Q_* = \frac{(P_1 - P_2)S}{R},$$

where  $R$  will typically be very low for a healthy valve.

For a compliance vessel that is not in steady state, we have

$$\frac{dV}{dt} = Q_1 - Q_2.$$

If we assume the vessel is connected to resistance vessels, the flows are given by

$$Q_j = \frac{P^{in} - P^{out}}{R_j}.$$

The flows  $Q_j$  depend on the pressures, which can be computed from the linear pressure-volume relation:

$$P_i = (V_i - V_{d,i})/C_i,$$

where  $V_{d,i}$ ,  $C_i$  are the unloaded volume and the compliance for vessel  $i$ . These are assumed constant for arteries and veins, and time dependent for the heart.

Another common formulation is to use the elastance instead of the compliance,  $E_i = 1/C_i$ , which gives

$$P_i = E_i(V_i - V_{d,i}),$$

## Circulation dynamics (3)

The circulatory system can now be viewed as a set of compliance vessels connected by valves and resistance vessels. For each compliance vessel we have

$$\frac{V_i}{dt} = Q_i^{in} - Q_i^{out},$$

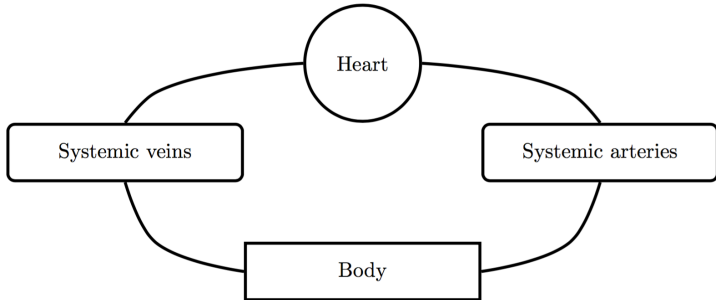
with flows given by

$$Q_i^{in} = \frac{P_{i-1} - P_i}{R_{i-1,i}} = \frac{E_{i-1}(V_{i-1} - V_{d,i-1}) - E_i(V_i - V_{d,i})}{R_{i-1,i}}$$
$$Q_i^{out} = \frac{P_i - P_{i+1}}{R_{i,i+1}} = \frac{E_i(V_i - V_{d,i}) - E_{i+1}(V_{i+1} - V_{d,i+1})}{R_{i,i+1}}$$

where we have introduced  $R_{i,i+1}$  to denote the resistance between vessel  $i$  and vessel  $i + 1$ .

# A simple model (1)

Consider first a simple model consisting of three compliance vessels; the left ventricle, the systemic arteries, and the systemic veins. These are connected by two valves, and a resistance vessel describing the flow through the systemic tissues.





## A simple model (2)

For the left ventricle we have

$$\frac{dV_{lv}}{dt} = Q^{in} - Q^{out},$$

with  $Q^{in}$  and  $Q^{out}$  given by

$$Q_{in} = \frac{S_{mi}(P_{sv} - P_{lv})}{R_{mi}},$$
$$Q_{out} = \frac{S_{ao}(P_{lv} - P_{sa})}{R_{ao}}.$$

We get

$$\frac{dV_{lv}}{dt} = \frac{P_{sv} - P_{lv}}{S_{mi}R_{mi}} - \frac{P_{lv} - P_{sa}}{S_{ao}R_{ao}},$$

## A simple model (3)

Similar calculations for the two other compliance vessels gives the system

$$\begin{aligned}\frac{dV_{lv}}{dt} &= \frac{S_{mi}(P_{sv} - P_{lv})}{R_{mi}} - \frac{S_{ao}(P_{lv} - P_{sa})}{R_{ao}}, \\ \frac{dV_{sa}}{dt} &= \frac{S_{ao}(P_{lv} - P_{sa})}{R_{ao}} - \frac{P_{sa} - P_{sv}}{R_{sys}}, \\ \frac{dV_{sv}}{dt} &= \frac{P_{sa} - P_{sv}}{R_{sys}} - \frac{S_{mi}(P_{sv} - P_{lv})}{R_{mi}}.\end{aligned}$$

With the pressures given by relations  $P_i = E_i(V_i - Vd, i)$  To have a solvable system we need to specify parameters  $R_{mi}, R_{ao}, R_{sys}, C_{sa}, C_{sv}$  and the function  $C_{lv}(t)$ .

## A more realistic model (1)

The model can easily be improved to a more realistic model describing six compliance vessels:

- The left ventricle,  $V_{lv}, E_{lv}(t)$ ,
- the right ventricle,  $V_{rv}, E_{rv}(t)$ ,
- the systemic arteries,  $V_{sa}, E_{sa}$ ,
- the systemic veins,  $V_{sv}, E_{sv}$ ,
- the pulmonary arteries, and  $V_{pa}, E_{pa}$ , and
- the pulmonary veins,  $V_{pv}, E_{pv}$ .

## A more realistic model (2)

The flows are governed by two resistance vessels and four valves:

- Systemic circulation,  $R_{sys}$ ,
- pulmonary circulation,  $R_{pu}$ ,
- aortic valve (left ventricle to systemic arteries),  $R_{ao}, S_{ao}$ ,
- tricuspid valve (systemic veins to right ventricle),  $R_{tri}, S_{tri}$ ,
- pulmonary valve (right ventricle to pulmonary arteries),  
 $R_{puv}, S_{puv}$ ,
- mitral valve (pulmonary veins to left ventricle) , $R_{mi}, S_{mi}$ .

## A more realistic model (3)

This gives the ODE system

$$\begin{aligned}\frac{dV_{lv}}{dt} &= \frac{S_{mi}(P_{sv} - P_{lv})}{R_{mi}} - \frac{S_{ao}(P_{lv} - P_{sa})}{R_{ao}}, \\ \frac{dV_{sa}}{dt} &= \frac{S_{ao}(P_{lv} - P_{sa})}{R_{ao}} - \frac{P_{sa} - P_{sv}}{R_{sys}}, \\ \frac{dV_{sv}}{dt} &= \frac{P_{sa} - P_{sv}}{R_{sys}} - \frac{S_{tri}(P_{sv} - P_{rv})}{R_{tri}}, \\ \frac{dV_{rv}}{dt} &= \frac{S_{tri}(P_{sv} - P_{rv})}{R_{tri}} - \frac{S_{puv}(P_{rv} - P_{pa})}{R_{puv}}, \\ \frac{dV_{pa}}{dt} &= \frac{S_{puv}(P_{rv} - P_{pa})}{R_{puv}} - \frac{P_{pa} - P_{pv}}{R_{pu}}, \\ \frac{dV_{pv}}{dt} &= \frac{P_{pa} - P_{pv}}{R_{pu}} - \frac{S_{mi}(P_{pv} - P_{lv})}{R_{mi}}.\end{aligned}$$

with pressures given by the formulas above.

- Python/matlab code in repo:  
*SUURPh-summer-school/L14/circ\_models*
- Three models:
  - Systemic (simple three-compartment model)
  - `circ_no_atria` (full circulation, six compartments)
  - `circ_full` (full circulation with atria, eight compartments)
- Run two test cases:
  - Aortic stenosis: Increase aortic resistance from 0.5 to 50
  - Physical exercise: Reduce systemic resistance from 246.9382 to 50
- What happens in the two cases?
- Is the model response realistic?

# Summary

- The entire circulation can be described using simple the building blocks of compliance and resistance vessels
- Simple models fail to describe correct physiological response, but are easily extended with baroreflex and more compartments, for remarkably realistic pressure- and flow profiles (See for instance <http://www.physiome.org/Models/> )
- Even with 50+ compartments, the heart is described as a time varying elastance:
  - Hard to relate changes on cell level to overall contractility and stiffness (elastance)
  - What about regional injuries such as an infarct?
- Multiscale models offer another dimension of modeling, integrating from the cell level to tissue and the complete organ

# Multiscale mechanical models

- Describe how the electrical signal triggers contraction of a single cell
- Connect cells, embed in tissue
- Couple with circulation models for flows and pressures (boundary conditions)



# Modeling cardiac cell contraction

- Crossbridge theory. How do muscles contract?
- A model for muscle contraction.
- Coupling to electrophysiology (excitation-contraction coupling)

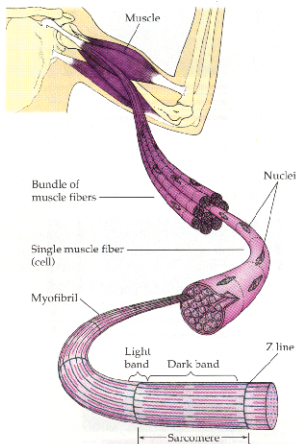
# Three main groups of muscle cells

- Smooth muscle
- Cardiac muscle
- Skeletal muscle

Cardiac muscle cells are so-called striated muscles, and have very similar contractile mechanisms. Skeletal muscle has been most extensively studied and modeled.

# Striated muscle cells

- A muscle cell (cardiac or skeletal) contains smaller units called myofibrils, which in turn are made up of sarcomeres.
- The sarcomere contains overlapping thin and thick filaments, which are responsible for the force development in the muscle cells.



# Overlapping thin and thick filaments

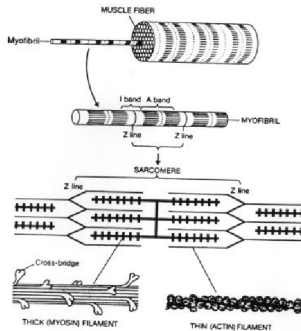
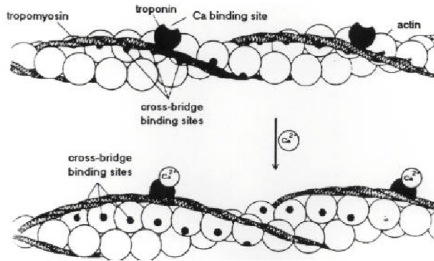


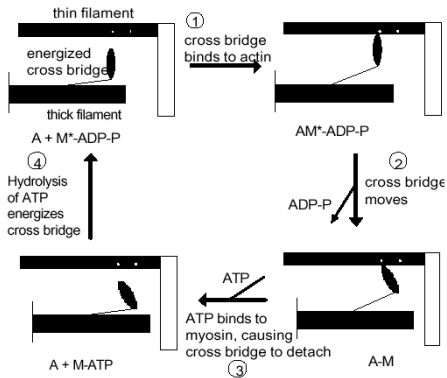
Figure. molecules have heads which form cross-bridges that interact with the thin filaments to generate force. \* Thick filaments are made up of the protein myosin. The myosin \* Thin filaments contain the three proteins actin, tropomyosin and troponin. \* The actin forms a double helix around a backbone formed by tropomyosin.

# Force regulation



- In the base configuration, tropomyosin blocks the cross-bridge binding sites on the actin.
- Troponin contains binding sites for calcium, and binding of calcium causes the tropomyosin to move, exposing the actin binding sites for the cross-bridges to attach.

# The cross-bridge cycle (1)



## The cross-bridge cycle (2)

After calcium has bound to the troponin to expose the binding sites, the force development in the muscle happens in four stages:

- An energized cross-bridge binds to actin.
- The cross-bridge moves to its energetically preferred position, pulling on the thin filament.
- ATP binds to the myosin, causing the cross-bridge to detach.
- Hydrolysis of ATP energizes the cross-bridge.

During muscle contraction, each cross-bridge goes through this cycle repeatedly.

# Many factors affect force development

- Effective overlap of thick and thin filaments; overlap and force increase when cell is stretched, up to a limit
- Velocity of contraction; cross-bridge cycle cannot "keep up" with high contraction rates, reducing developed tension
- Cooperativity; a formed cross bridge increases the likelihood of nearby cross-bridges to form



## Important quantities

- Isometric tension ( $T_0$ ): the tension generated by a muscle contracting at a fixed length. Depends on the activation level (i.e. Ca concentration) and length (stretch) of the muscle cell
- Tension ( $T$ ): Tension developed during active shortening. In phenomenological models, this is often obtained by a scaling of the isometric tension:  $T = T_0 f(V)$ , where  $V$  is the rate of shortening and  $f(V) < 1$  is some *force-velocity relation*.

# Force-velocity relations (1)

- The classical equation of Hill (1938) describes the relation between velocity and tension in a muscle that contracts against a constant load (*isotonic* contraction):  $(T + a)V = b(T_0 - T)$
- $T_0$  is the isometric tension and  $V$  is the velocity.  $a$  and  $b$  are parameters which are fitted to experimental data.
- Recall that  $T_0$  is constant for skeletal muscle cells, dependent on length in cardiac cells

## Force-velocity relations (2)

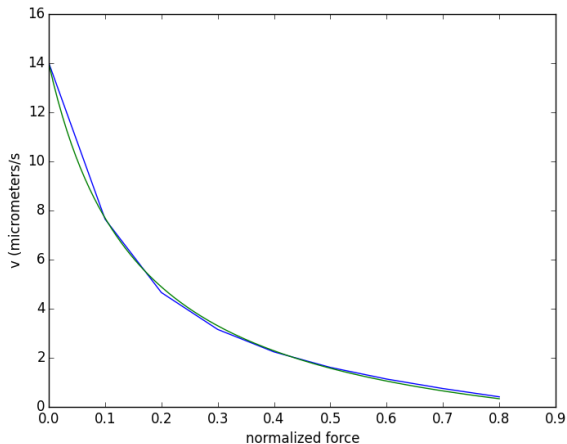
Velocity as function of force:

$$V = b \frac{T_0 - T}{T + a}$$

Force as function of velocity:

$$T = \frac{bT_0 - aV}{b + V}$$

# A typical Hill-curve



Force-velocity relation for the model by Rice et al (2008).

# Summary of model requirements

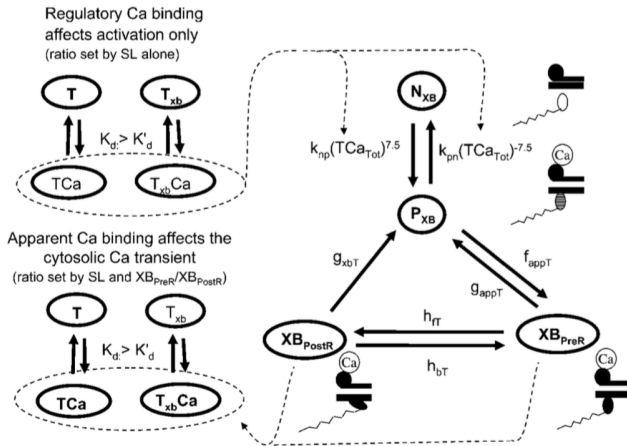
A model of cell contraction must include the following:

- Calcium binding to Troponin C
- Exposure of cross bridge binding sites on the actin
- Cycling of cross-bridges to develop force
- Force-dependence on filament overlap and contraction velocity
  - Often phenomenological (Hunter et al (1998), Niederer et al (2006))
  - Recent models are more mechanistic (Rice et al (2008), Campbell et al (2009))

## An example model; Rice et al (2008)

- Rice, Wang, Bers, de Tombe: *Approximate Model of Cooperative Activation and Crossbridge Cycling in Cardiac Muscle Using Ordinary Differential Equations*, Biophysical journal, 2008.
- A hybrid model;
  - Some model components described in great detail and biophysical rigour
  - Some components deliberately simplified and phenomenological, to keep complexity at a reasonable level
- Conceptually simple (but the devil is in the details)

# The model has eight main state variables



+ 3 mechanical (Sarcomere length and crossbridge distortions).  
Calcium is considered an input variable.

## Two ODEs describe calcium binding and detachment

The model treats Ca binding sites with nearby cross-bridges separately from those without nearby cross-bridges:

$$\begin{aligned}\frac{dTRPN_{CaL}}{dt} &= k_{onT} Ca_i (1 - TRPN_{CaL}) - k_{offLT} TRPN_{CaL} \\ \frac{dTRPN_{CaH}}{dt} &= k_{onT} Ca_i (1 - TRPN_{CaH}) - k_{offHT} TRPN_{CaH}\end{aligned}$$

The sites with bound cross-bridges have a higher Ca affinity, represented by a lower detachment rate:  $k_{offHT} < k_{offLT}$ .



# Six ODEs describe cross-bridge (XB) cycling

$$\frac{dN_{NoXB}}{dt} = k_{pnT}P_{NoXB} - k_{npT}N_{NoXB}$$

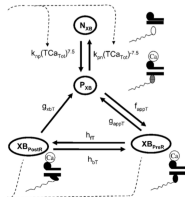
$$\frac{dP_{NoXB}}{dt} = k_{npT}N_{NoXB} - k_{pnT}P_{NoXB}$$

$$\frac{dN}{dt} = k_{pn}P - k_{npT}N$$

$$\frac{dP}{dt} = k_{npT}N - k_{pn}P - f_{appT}P + g_{appT}XB_{prer} + g_{xbT}) * XB_{postr}$$

$$\frac{dXB_{prer}}{dt} = f_{appT}P + h_{bT}XB_{postr} - (g_{appT} + h_{fT})XB_{prer}$$

$$\frac{dXB_{postr}}{dt} = h_{fT}XB_{prer} - (h_{bT}XB_{postr} + g_{xbT}) * XB_{postr}$$



## Only strongly-bound XBs develop force

Force is proportional to the number of strongly bound XBs

$$F \sim (XB_{prer} + XB_{postr})$$

The Rice model describes the XBs as linear springs. The force developed by a cross bridge is equal to a spring constant multiplied with its distortion (strain):

$$F \sim k_{XB}(XB_{prer} \times XB_{prer} + XB_{postr} \times XB_{postr})$$

Concept originally presented by Razumova et al (1999).

## Cross bridge distortion comes from two sources

- Lengthening or shortening a muscle will change the distortion of all strongly bound XBs. In the model the XB distortions depend on  $dSL/dt$ , with  $SL$  being sarcomere length.
- Cross-bridges going from pre- to post-rotation state (and back) will induce a distortion ( $x_0$ )

$$\frac{dxXB_{prer}}{dt} \sim \frac{dSL}{dt} + (-f_{appT}xXB_{prer} + hbT * (xXB_{postr} - (x_0 + xXB_{prer})))$$
$$\frac{dxXB_{postr}}{dt} \sim \frac{dSL}{dt} + h_{fT}(xXB_{prer} + x_0 - xXB_{postr})$$

Warning: The published model includes much more complex expressions. The important part is the overall concept.

# Demo of classical cell mechanics experiments

From Rice et al (2008).

## Coupling to electrophysiology

- Coupling of the Rice model to an electrophysiology (EP) model is straight-forward; just take Ca from EP model as input to Rice model.
- Often some overlap, since most EP models describe binding of Ca to Troponin; we need to adjust the models slightly to get the right Ca dynamics.
- Can be extended with more mechanisms of mechano-electric feedback (MEF), such as stretch-activated channels.