

Introduction to Bioequivalence

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1 Introduction

We recommend first reading the [documentation](#) which goes over how to download and install the module.

1.1 Quick Start

We first explore an example from the `pumas_be` function documentation.

Let us load the module through

```
using Pkg
Pkg.activate(joinpath(@__DIR__, "tutorials", "bioequivalence"))
using Bioequivalence
```

The first example comes from a two sequences, two periods, two formulations crossover design (RT|TR) from Schütz, Labes, and Fuglsang (2014).

```
SLF2014 = Bioequivalence.testdata("SLF2014_8")
show(SLF2014)
```

```
1434×4 DataFrames.DataFrame
Row    id    period  sequence  AUC
      Int64  Int64   Cat...   Float64

1      1      1      TR      168.407
2      1      2      TR      210.919
3      2      1      TR      131.031
4      2      2      TR      67.4314
5      3      1      TR      151.737
6      3      2      TR      85.1296
7      4      1      TR      375.575
8      4      2      TR      147.502
9      5      1      TR      50.023
10     5      2      TR      20.0107
⋮
1424   995      2      RT      47.0775
1425   996      1      RT      128.804
1426   996      2      RT      102.543
1427   997      1      RT      85.6709
1428   997      2      RT      45.7204
1429   998      1      RT      286.78
```

1430	998	2	RT	73.9542
1431	999	1	RT	63.1119
1432	999	2	RT	13.3009
1433	1000	1	RT	55.8031
1434	1000	2	RT	50.0577

The `id` variable specifies the identifier for each subject.

The `sequence` must be a `String` with each `Character` representing a formulation.

For example, "RTTR" represents R formulation in the first period, T formulation in the second and third periods, and R formulation in the fourth period.

The `period` variable must be integers specifying the period based on the sequence.

For example,

```
id = 1
period = 1
sequence = "TR"
endpoint = 168.407
```

means that the endpoint for subject with ID 1 in the first period when receiving the T formulation recorded the value 168.407.

Notice that the study design is fully characterized by the unique values of the `sequence` variable.

In order to perform the bioequivalence analysis we can pass the data to the `pumas_be` function.

```
Crossover = pumas_be(SLF2014)
show(Crossover)
```

Design: RT|TR

Sequences: RT|TR (2)

Periods: 1:2 (2)

Subjects per Sequence: (RT = 288, TR = 429)

Average Bioequivalence

	PE	SE	lnLB	lnUB	GMR	LB	UB
T - R	-0.0680309	0.044609	-0.141501	0.00543947	0.934232	0.868	1.0055

When passing only the data argument it will use the default values for all the other arguments.

For example, if the dataset had the variable indicating the subject ID as `subj` instead of `id`,

```
using DataFrames
rename!(SLF2014, "id" => "subj")
try
    pumas_be(SLF2014)
catch err
    err
end
```

Error: ArgumentError: data must contain id, sequence, period, and endpoint

In this case we can pass the variable position (i.e., 1) or the name of the variable as a `Symbol` to the function.

```
Crossover = pumas_be(SLF2014, id = :subj)
show(Crossover)
```

Design: RT|TR

Sequences: RT|TR (2)

Periods: 1:2 (2)

Subjects per Sequence: (RT = 288, TR = 429)

Average Bioequivalence

	PE	SE	lnLB	lnUB	GMR	LB	UB
T - R	-0.0680309	0.044609	-0.141501	0.00543947	0.934232	0.868	1.0055

1.2 Examining the Results

The `BioequivalenceStudy` will show the study design as well as how many subjects were in each sequence.

For convenience it also shows the formulation and periods as well as how many there are for that design.

A coefficients table will also be showing the parameter estimates and related values for each comparison among each non-reference formulation and the reference formulation. Those will include the point estimate (PE), standard error (SE), the 90% confidence interval in the natural log scale (i.e., `lnLB`, `lnUB`), the geometric means ratio (`GMR`), and the 90% confidence intervals in the natural scale (i.e., `LB`, `UB`).

The average bioequivalence is assessed based on the lower and upper bounds of the 90% percent confidence interval. Based on the regulatory guidelines for the FDA, the confidence intervals are reported in percentage form with two decimal places. In other words, if the lower bound value is 0.85793 the percentage form would be 85.793% and be rounded down to 85.79%. If the upper bound value is 1.20522 the percentage form would be 120.522% and be rounded up to 120.53%. The Bioequivalence results will have correct percentage format in decimal form (e.g. 1.2053 for 120.53%).

Besides the bioequivalence results one can query a number of properties.

We can access the information about the design

```
show(Crossover.design)
```

(RT = 288, TR = 429)

The data used for the model

```
show(Crossover.data)
```

1434×5 DataFrames.DataFrame

Row	id	sequence	period	AUC	formulation
	Cat...	Cat...	Cat...	Float64	Cat...

```

1      1      TR      1      5.12638 'T'
2      1      TR      2      5.35148 'R'
3      2      TR      1      4.87543 'T'
4      2      TR      2      4.21111 'R'
5      3      TR      1      5.02215 'T'
6      3      TR      2      4.44418 'R'
7      4      TR      1      5.92846 'T'
8      4      TR      2      4.99384 'R'
9      5      TR      1      3.91248 'T'
10     5      TR      2      2.99627 'R'
:
1424   995    RT      2      3.8518  'T'
1425   996    RT      1      4.85829 'R'
1426   996    RT      2      4.63028 'T'
1427   997    RT      1      4.45051 'R'
1428   997    RT      2      3.82254 'T'
1429   998    RT      1      5.65872 'R'
1430   998    RT      2      4.30345 'T'
1431   999    RT      1      4.14491 'R'
1432   999    RT      2      2.58783 'T'
1433  1000    RT      1      4.02183 'R'
1434  1000    RT      2      3.91318 'T'

```

Statistics about the data such as:

how many observations did the initial data had.

```
show(Crossover.data_stats.total)
```

```
1434
```

how many observations did the analysis actually used (e.g., total - count of missing values).

```
show(Crossover.data_stats.used_for_analysis)
```

```
1434
```

as well as summary statistics of the endpoint distribution by sequence, period or formulation.

```
show(Crossover.data_stats.sequence)
```

```
2×9 DataFrames.DataFrame
```

Row	sequence	mean	std	min	q25	median	q75	max	n
	Cat...	Float64	Float64	Float64	Float64	Float64	Float64	Float64	
Int64									
1	RT	4.25295	0.858753	2.22401	3.77021	4.25043	4.71928	17.7519	576
2	TR	5.44929	3.12075	2.42739	4.32815	4.8114	5.24877	20.1913	858

```
show(Crossover.data_stats.period)
```

```
2×9 DataFrames.DataFrame
```

Row	period	mean	std	min	q25	median	q75	max	n
	Cat...	Float64	Float64	Float64	Float64	Float64	Float64	Float64	Int64
1	1	5.32608	2.57664	3.27928	4.50087	4.89085	5.27117	20.1913	717
2	2	4.61142	2.45832	2.22401	3.77503	4.22118	4.67236	19.9725	717

```
show(Crossover.data_stats.formulation)
```

```
2×9 DataFrames.DataFrame
```

Row	formulation	mean	std	min	q25	median	q75	max	n
	Cat...	Float64	Float64	Float64	Float64	Float64	Float64	Float64	
Int64									
1	'R'	4.93118	2.43666	2.42739	4.14032	4.52236	4.96409	19.9725	
2	'T'	5.00632	2.64537	2.22401	4.00392	4.67886	5.19103	20.1913	

One can also query the model used for inference. The exact API for extracting values from the models will depend on the design. The most common API is documented [here](#). For example, we can obtain the coefficient of determination (R²) through the API.

```
using StatsBase
r2(Crossover.model)
```

```
0.947066878314061
```

likewise we can obtain the deviance of the model through

```
deviance(Crossover.model)
```

```
490.35529438202303
```

Derived statistics for the model such as the Wald test for join-significance for the sequence, period, and formulation covariates

```
show(Crossover.model_stats.Wald)
```

	Wald	Distribution	p-value
Formulation	2.32578	FDist($\nu_1=1.0$, $\nu_2=715.0$)	0.1277
Sequence	2.3571	FDist($\nu_1=1.0$, $\nu_2=715.0$)	0.1252
Period	266.357	FDist($\nu_1=1.0$, $\nu_2=715.0$)	<1e-50
Subject	16.5092	FDist($\nu_1=715.0$, $\nu_2=715.0$)	<1e-99

and the linear regression mean estimates per formulation

```
show(Crossover.model_stats.lsmmeans)
```

	Mean	Standard Deviation	t-statistic	Distribution	p-value
R	4.63723	0.414669	11.183	TDist($\nu=715.0$)	<1e-26
T	4.5692	0.414669	11.0189	TDist($\nu=715.0$)	<1e-25

are available as shown above.

You can also run models for various endpoints easily as following

```
data = Bioequivalence.testdata("PJ2017_4_3")
show(data)
```

68x5 DataFrames.DataFrame

Row	id	sequence	period	AUC	Cmax
	Int64	Cat...	Int64	Int64?	Int64?
1	1	RTTR	1	10671	817
2	1	RTTR	2	12772	1439
3	1	RTTR	3	13151	1310
4	1	RTTR	4	11206	1502
5	2	TRRT	1	6518	1393
6	2	TRRT	2	6068	1372
7	2	TRRT	3	5996	1056
8	2	TRRT	4	5844	1310
9	3	TRRT	1	4939	1481
10	3	TRRT	2	5728	1377
:					
58	15	RTTR	2	7905	1065
59	15	RTTR	3	6550	830
60	15	RTTR	4	7515	1247
61	16	RTTR	1	11473	1368
62	16	RTTR	2	9698	1281
63	16	RTTR	3	10355	1083
64	16	RTTR	4	10365	1418
65	18	TRRT	1	5210	668
66	18	TRRT	2	5120	842
67	18	TRRT	3	5420	1176
68	18	TRRT	4	missing	missing

```
auc = pumas_be(data)
cmax = pumas_be(data, endpoint = :Cmax)
output = Dict{endpoint => pumas_be(data, endpoint = endpoint) for endpoint in
setdiff(propertynames(data), (:id, :sequence, :period))}
```

```
Dict{Symbol,Bioequivalence.BioequivalenceStudy{NamedTuple{(:total, :used_for_analysis, :
formulation, :sequence, :period),Tuple{Int
64,Int64,DataFrames.DataFrame,DataFrames.DataFrame,DataFrames.DataFrame}},NamedTuple{(:
RTTR, :TRRT),Tuple{Int64,Int64}},MixedModel
s.LinearMixedModel{Float64},NamedTuple{(:Wald, :lsmeans),Tuple{StatsBase.CoeffTable,
StatsBase.CoeffTable}},Tuple{Float64,Float64}}}
with 2 entries:
  :AUC => Design: RTTR|TRRT...
  :Cmax => Design: RTTR|TRRT...
```

```
show(output[:AUC])
```

```
Design: RTTR|TRRT
```

```
Sequences: RTTR|TRRT (2)
```

```
Periods: 1:4 (4)
```

```
Subjects per Sequence: (RTTR = 8, TRRT = 9)
```

```
Regulatory constant for reference-scaled estimates: 0.10
```

```
Auxiliary parameter for reference-scaled estimates: 1.11
```

Average Bioequivalence

	PE	SE	lnLB	lnUB	GMR	LB	UB	scLB
scUB	Varratio	VarLB	VarUB					

```
T - R  0.0356935  0.0226361  -0.00230475  0.0736918  1.03634  0.9976  1.0765  0.8682
1.1518   1.34546  0.8641  2.1116
```

```
show(output[:Cmax])
```

```
Design: RTTR|TRRT
```

```
Sequences: RTTR|TRRT (2)
```

```
Periods: 1:4 (4)
```

```
Subjects per Sequence: (RTTR = 8, TRRT = 9)
```

```
Regulatory constant for reference-scaled estimates: 0.10
```

```
Auxiliary parameter for reference-scaled estimates: 1.11
```

```
Average Bioequivalence
```

		PE	SE	lnLB	lnUB	GMR	LB	UB	scLB
scUB	Varratio	VarLB	VarUB						
T - R	-0.0917895	0.0524819	-0.179889	-0.00369023	0.912297	0.8353	0.9964	0.7005	
	1.4274	1.25641	0.8069	1.9719					