

# Three-year fitness of *Panicum virgatum* infected with switchgrass mosaic virus

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## 1 R

- The version of R used to make this document is 4.2.1.
- The version of R package `rmarkdown` used to make this document is 2.19.
- The version of R package `knitr` used to make this document is 1.41.
- The version of R package `aster` used to make this document is 1.1.2.
- The version of R package `trust` used to make this document is 0.1.8.
- The version of R package `numDeriv` used to make this document is 2016.8.1.1.
- The version of R package `freshr` used to make this document is 1.0.2.

Ensure a clean R global environment.

```
freshr::freshr()
```

Load R packages `aster` and `numDeriv`

```
library("aster")
library("numDeriv")
```

Set global option.

```
# don't need this with R-4.0.0, it's the default there and forevermore
# but it doesn't hurt and defends against users who haven't upgraded R
options(stringsAsFactors = FALSE)
```

## 2 Data

```
redata <- read.csv("redata_fin.csv")
# "symp.extent" is the proportion of diseased tillers. Taken initially,
# in early summer 2017, and, for 2017 and 2018,
# taken at the end of season around the time
# we collected fitness metrics (panicle counts, lengths)
```

```
sapply(redata, class)
```

```
##      PlantID      Year symp.extent      varb      resp      id
## "character" "integer"  "numeric" "character" "integer" "integer"
##           root
```

```
## "integer"
unique(redata$varb)

## [1] "late17.tillers"          "late17.panicles"
## [3] "late17.pans.sampled"     "late17.floret.count.total"
## [5] "late18.tillers"          "late18.panicles"
## [7] "late18.pans.sampled"     "late18.floret.count.total"
## [9] "late19.tillers"          "late19.panicles"
## [11] "late19.pans.sampled"     "late19.floret.count.total"

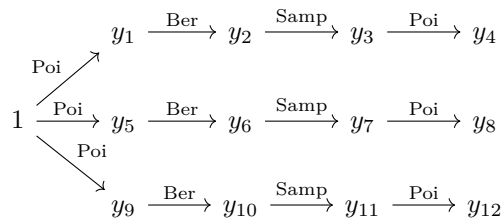
str(redata)

## 'data.frame': 120 obs. of 7 variables:
## $ PlantID : chr "A" "B" "C" "D" ...
## $ Year : int 2017 2017 2017 2017 2017 2017 2017 2017 2017 2017 ...
## $ symp.extent: num 0.42 0.42 0.39 0.68 0.19 0.05 0.14 0.04 0.22 0.22 ...
## $ varb : chr "late17.tillers" "late17.tillers" "late17.tillers" "late17.tillers" ...
## $ resp : int 56 45 77 79 148 114 172 157 90 219 ...
## $ id : int 1 2 3 4 5 6 7 8 9 10 ...
## $ root : int 1 1 1 1 1 1 1 1 1 1 ...

redata$PlantID <- as.factor(redata$PlantID)
redata$Year <- as.factor(redata$Year)
```

### 3 Graph

We use the following aster graph for one individual.



In this graph the “rows” are years (2017, 2018, and 2019) and the “columns” are data within years: first tillers ( $y_1$ ,  $y_5$ , and  $y_9$ ), then panicles ( $y_2$ ,  $y_6$ , and  $y_{10}$ ), then sub-sampled panicles ( $y_3$ ,  $y_7$ , and  $y_{11}$ ), and finally (the terminal nodes) floret count ( $y_4$ ,  $y_8$ , and  $y_{12}$ ).

After initial analysis we may change some Poisson to negative binomial (a kind of over-dispersed Poisson), but first we see whether that seems necessary.

It is somewhat problematic that tiller counts for different years are for the same plant and these should be dependent. We allow for such dependence (somewhat) by putting individual effects in the model.

```
pred <- c(0, 1, 2, 3, 0, 5, 6, 7, 0, 9, 10, 11)
fam <- rep(c(2, 1, 1, 2), times = 3)
fam
```

```
## [1] 2 1 1 2 2 1 1 2 2 1 1 2
```

```
vars <- unique(redata$varb)
vars
```

```
## [1] "late17.tillers"          "late17.panicles"
## [3] "late17.pans.sampled"     "late17.floret.count.total"
## [5] "late18.tillers"          "late18.panicles"
```

```
## [7] "late18.pans.sampled"      "late18.floret.count.total"
## [9] "late19.tillers"           "late19.panicles"
## [11] "late19.pans.sampled"      "late19.floret.count.total"

pred.names <- c("initial", vars)[pred + 1]
foo <- cbind(pred.names, vars, fam)
colnames(foo) <- c("predecessor", "successor", "family")
foo

##      predecessor      successor      family
## [1,] "initial"        "late17.tillers"    "2"
## [2,] "late17.tillers"  "late17.panicles"    "1"
## [3,] "late17.panicles" "late17.pans.sampled" "1"
## [4,] "late17.pans.sampled" "late17.floret.count.total" "2"
## [5,] "initial"        "late18.tillers"    "2"
## [6,] "late18.tillers"  "late18.panicles"    "1"
## [7,] "late18.panicles" "late18.pans.sampled" "1"
## [8,] "late18.pans.sampled" "late18.floret.count.total" "2"
## [9,] "initial"        "late19.tillers"    "2"
## [10,] "late19.tillers"  "late19.panicles"    "1"
## [11,] "late19.panicles" "late19.pans.sampled" "1"
## [12,] "late19.pans.sampled" "late19.floret.count.total" "2"

fit <- as.numeric(grepl("floret", as.character(redata$varb)))
redata <- data.frame(redata, fit = fit)
ind <- as.factor(redata$id)
redata <- data.frame(redata, ind = ind)

redata <- subset(redata, redata$varb %in% vars)

nnode <- length(vars)
nind <- length(unique(redata$id))
nnode * nind == nrow(redata)

## [1] TRUE
```

## 4 Initial Aster Models

### 4.1 Fit models (Poisson)

To start, we'll fit three models: a null model, a model with a fixed effect term for symptom extent (`symp.extent`) (a plant-level measure we assessed each growing season), and a third model that includes a term to describe variation at the individual level (`ind`).

```
anull <- aster(resp ~ varb,
  pred, fam, varb, id, root, data = redata)

aout.noind <- aster(resp ~ varb + fit : (symp.extent),
  pred, fam, varb, id, root, data = redata)

aout <- aster(resp ~ varb + fit : (symp.extent + ind),
  pred, fam, varb, id, root, data = redata)

anova(anull, aout.noind, aout)
```

```
## Analysis of Deviance Table
##
## Model 1: resp ~ varb
## Model 2: resp ~ varb + fit:(symp.extent)
## Model 3: resp ~ varb + fit:(symp.extent + ind)
##   Model Df Model Dev Df Deviance P(>|Chi|)
## 1      12  1283304
## 2      13  1283318  1   14.419 0.0001463 ***
## 3      22  1283360  9   41.634 3.833e-06 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

The hypothesis test says both `symp.extent` and `ind` are statistically significant. The `ind` term helps us model the dependency of the tiller counts at the individual plant level, which were different among plants at the start of the experiment. Additionally, we also know (from other experiments) that there is a lot of variation in panicle lengths and floret production at the individual plant level. Therefore, going forward, we'll use the `ind` model as the base model as we evaluate residuals and over-dispersion.

```
summary(aout, info.tol = 1e-9)
```

```
##
## Call:
## aster.formula(formula = resp ~ varb + fit:(symp.extent + ind),
##   pred = pred, fam = fam, varvar = varb, idvar = id, root = root,
##   data = redata)
##
##               Estimate Std. Error  z value Pr(>|z|)
## (Intercept)      6.006e+00  5.120e-03 1172.906 < 2e-16 ***
## varblate17.panicles -3.888e+00  7.796e-02 -49.870 < 2e-16 ***
## varblate17.pans.sampled -4.124e+02  2.021e+00 -204.073 < 2e-16 ***
## varblate17.tillers -2.292e+00  7.003e-02 -32.730 < 2e-16 ***
## varblate18.floret.count.total -3.922e-02  6.868e-03  -5.711 1.13e-08 ***
## varblate18.panicles -4.008e+00  6.770e-02 -59.208 < 2e-16 ***
## varblate18.pans.sampled -3.970e+02  1.852e+00 -214.407 < 2e-16 ***
## varblate18.tillers -1.984e+00  6.009e-02 -33.017 < 2e-16 ***
## varblate19.floret.count.total -2.994e-01  6.916e-03 -43.283 < 2e-16 ***
## varblate19.panicles -4.341e+00  5.463e-02 -79.463 < 2e-16 ***
## varblate19.pans.sampled -3.078e+02  1.448e+00 -212.613 < 2e-16 ***
## varblate19.tillers -1.471e+00  4.661e-02 -31.561 < 2e-16 ***
## fit:symp.extent -8.189e-03  4.020e-03  -2.037 0.041653 *
## fit:ind2      1.562e-03  8.528e-04   1.832 0.066964 .
## fit:ind3      1.750e-03  7.745e-04   2.260 0.023826 *
## fit:ind4      1.578e-03  1.771e-03   0.891 0.372840
## fit:ind5      8.084e-04  7.792e-04   1.037 0.299554
## fit:ind6     -1.225e-03  1.186e-03  -1.033 0.301691
## fit:ind7      1.379e-03  8.850e-04   1.558 0.119230
## fit:ind8     -8.420e-04  1.125e-03  -0.748 0.454169
## fit:ind9      4.081e-04  8.167e-04   0.500 0.617302
## fit:ind10     2.356e-03  6.985e-04   3.373 0.000744 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## 4.2 Get Conditional and Unconditional Mean Value Parameters

```
pout.cond <- predict(aout, model.type = "conditional",
  is.always.parameter = TRUE, gradient = TRUE)
xi <- pout.cond$fit
class(xi)

## [1] "numeric"

length(xi) == nind * nnode

## [1] TRUE

xi <- matrix(xi, nrow = nind)
colnames(xi) <- vars
xi
```

	late17.tillers	late17.panicles	late17.pans.sampled
## [1,]	109.6623	0.8130624	0.04443901
## [2,]	113.1557	0.8188337	0.08046659
## [3,]	114.5872	0.8210970	0.09445717
## [4,]	108.8709	0.8117036	0.03588178
## [5,]	117.4830	0.8255067	0.12149555
## [6,]	113.9266	0.8200595	0.08805372
## [7,]	123.2334	0.8336490	0.17066907
## [8,]	115.6305	0.8227111	0.10438804
## [9,]	114.7692	0.8213806	0.09620495
## [10,]	125.6813	0.8368890	0.18997002

	late17.floret.count.total	late18.tillers	late18.panicles
## [1,]	404.3292	138.8843	0.7991133
## [2,]	404.9614	140.3235	0.8011737
## [3,]	405.1370	142.4651	0.8041626
## [4,]	404.1064	134.6663	0.7928213
## [5,]	405.4191	142.2782	0.8039053
## [6,]	405.0598	140.0113	0.8007304
## [7,]	405.8165	151.9228	0.8163542
## [8,]	405.2480	140.2137	0.8010180
## [9,]	405.1573	140.2947	0.8011329
## [10,]	405.9472	149.9400	0.8139256

	late18.pans.sampled	late18.floret.count.total	late19.tillers
## [1,]	0.07374258	389.5105	180.9998
## [2,]	0.08560038	389.6725	184.5408
## [3,]	0.10269359	389.8735	187.0294
## [4,]	0.03714979	388.7862	176.8308
## [5,]	0.10122746	389.8574	183.7563
## [6,]	0.08305444	389.6395	181.2938
## [7,]	0.17112048	390.4634	188.1139
## [8,]	0.08470643	389.6610	181.7812
## [9,]	0.08536599	389.6695	185.6416
## [10,]	0.15765326	390.3653	191.0123

	late19.panicles	late19.pans.sampled	late19.floret.count.total
## [1,]	0.7425412	0.08407609	300.3656
## [2,]	0.7474814	0.10758842	300.6382
## [3,]	0.7508413	0.12340281	300.7933
## [4,]	0.7364712	0.05475488	299.9053
## [5,]	0.7464032	0.10248368	300.5839

```
## [6,]      0.7429587      0.08607541      300.3913
## [7,]      0.7522778      0.13012098      300.8540
## [8,]      0.7436478      0.08937021      300.4325
## [9,]      0.7489787      0.11465373      300.7098
## [10,]     0.7560367      0.14757983      301.0001
```

```
pout.unco <- predict(aout, gradient = TRUE)
mu <- pout.unco$fit
mu <- matrix(mu, nrow = nind)
colnames(mu) <- vars
mu
```

```
##      late17.tillers late17.panicles late17.pans.sampled
## [1,]      109.6623      89.16228      3.962284
## [2,]      113.1557      92.65569      7.455688
## [3,]      114.5872      94.08721      8.887212
## [4,]      108.8709      88.37091      3.170905
## [5,]      117.4830      96.98300     11.783003
## [6,]      113.9266      93.42656      8.226556
## [7,]      123.2334     102.73342     17.533416
## [8,]      115.6305      95.13048      9.930485
## [9,]      114.7692      94.26916      9.069160
## [10,]     125.6813     105.18129     19.981292
##      late17.floret.count.total late18.tillers late18.panicles
## [1,]           1602.067      138.8843      110.9843
## [2,]           3019.266      140.3235      112.4235
## [3,]           3600.538      142.4651      114.5651
## [4,]           1281.383      134.6663      106.7663
## [5,]           4777.054      142.2782      114.3782
## [6,]           3332.247      140.0113      112.1113
## [7,]           7115.350      151.9228      124.0228
## [8,]           4024.309      140.2137      112.3137
## [9,]           3674.436      140.2947      112.3947
## [10,]          8111.349      149.9400      122.0400
##      late18.pans.sampled late18.floret.count.total late19.tillers
## [1,]           8.184266      3187.858      180.9998
## [2,]           9.623494      3750.011      184.5408
## [3,]          11.765102      4586.901      187.0294
## [4,]           3.966348      1542.061      176.8308
## [5,]          11.578217      4513.854      183.7563
## [6,]           9.311345      3628.068      181.2938
## [7,]          21.222849      8286.746      188.1139
## [8,]           9.513691      3707.115      181.7812
## [9,]           9.594683      3738.756      185.6416
## [10,]          19.240005      7510.630      191.0123
##      late19.panicles late19.pans.sampled late19.floret.count.total
## [1,]           134.3998      11.299811      3394.075
## [2,]           137.9408      14.840837      4461.723
## [3,]           140.4294      17.329381      5212.561
## [4,]           130.2308       7.130771      2138.556
## [5,]           137.1563      14.056280      4225.092
## [6,]           134.6938      11.593826      3482.685
## [7,]           141.5139      18.413932      5539.904
## [8,]           135.1812      12.081169      3629.576
## [9,]           139.0416      15.941643      4793.808
```

```
## [10,]          144.4123          21.312350          6415.020
```

### 4.3 Correct for Sub-sampling

#### 4.3.1 Point Estimates

The fundamental relationship between conditional and unconditional means is

$$\mu_j = \mu_{p(j)} \xi_j$$

So we get unconditional means by multiplying together the corresponding conditional mean and the unconditional mean for the predecessor.

To correct for sub-sampling, we want to do the same thing except we want to leave out the sub-sampling arrows. That is

$$\mu_{\text{florets}} = \mu_{\text{panicles}} \xi_{\text{florets}}$$

So first we obtain these quantities.

```
is.florets <- grep("florete", vars)
is.panicles <- grep("panicles", vars)
is.florets
```

```
## [1] 4 8 12
```

```
is.panicles
```

```
## [1] 2 6 10
```

```
mu.panicles <- mu[, is.panicles]
xi.florets <- xi[, is.florets]
mu.florets <- mu.panicles * xi.florets
mu.florets
```

```
##      late17.panicles late18.panicles late19.panicles
## [1,]      36050.92      43229.54      40369.09
## [2,]      37521.98      43808.35      41470.29
## [3,]      38118.21      44665.89      42240.21
## [4,]      35711.25      41509.28      39056.89
## [5,]      39318.76      44591.20      41226.97
## [6,]      37843.34      43683.01      40460.86
## [7,]      41690.92      48426.38      42575.03
## [8,]      38551.44      43764.27      40612.82
## [9,]      38193.84      43796.78      41811.18
## [10,]     42698.05      47640.18      43468.14
```

Then (the best surrogate of) fitness (in these data) is the sum of these for each individual.

```
mu.fit <- rowSums(mu.florets)
mu.fit
```

```
## [1] 119649.5 122800.6 125024.3 116277.4 125136.9 121987.2 132692.3 122928.5
## [9] 123801.8 133806.4
```

### 4.4 Standard Errors

For reasons that will soon become apparent, we make an R function to do the preceding calculation.

```
foo <- function(x) {
  # x is xi and mu strung out as one vector
```

```

xi <- x[1:length(xi)]
mu <- x[- (1:length(xi))]
xi <- matrix(xi, nrow = nind)
mu <- matrix(mu, nrow = nind)
mu.panicles <- mu[ , is.panicles]
xi.florets <- xi[ , is.florets]
mu.florets <- mu.panicles * xi.florets
mu.fit <- rowSums(mu.florets)
}

```

And we check that it does indeed give the same calculation as above.

```

ximu <- c(xi, mu)
all.equal(foo(ximu), mu.fit)

```

```
## [1] TRUE
```

In order to derive standard errors using the delta method, we need Jacobian matrices (matrices of partial derivatives). Rather than do any calculus, we let R package `numDeriv` figure out the Jacobian matrix for this transformation. We also need the Jacobian matrix for the transformation from the “coefficients” vector to the vector `ximu`.

```

jac.foo <- jacobian(foo, ximu)
jac.ximu <- rbind(pout.cond$gradient, pout.unco$gradient)

```

Now the chain rule from multivariate calculus says the Jacobian for the overall transformation is the product of the Jacobians for the parts.

```
jac.total <- jac.foo %*% jac.ximu
```

Now the delta method says the variance-covariance matrix of all the fitnesses (the vector estimate `mu.fit`) is  $JJ^{-1}J^T$ , where  $J$  is the overall Jacobian matrix `jac.total` and  $I$  is Fisher information for the “coefficients” vector

```
V <- jac.total %*% solve(aout$fisher) %*% t(jac.total)
```

and the standard errors are square roots of the variances (the diagonal elements of `V`)

```

se <- sqrt(diag(V))
bar.pois <- cbind(mu.fit, se)
colnames(bar.pois) <- c("Estimate", "SE")

```

Table 1: Estimated Fitness with Standard Error for Different Individuals (Poisson Distributions for Tillers and Florets)

Estimate	SE
119649.5	2702.672
122800.6	2903.538
125024.3	3037.413
116277.4	2478.442
125136.9	3062.308
121987.2	2865.781
132692.3	3495.365
122928.5	2927.697
123801.8	2965.634
133806.4	3547.607



## 5 Checking for Over-dispersion

Following the theory for the negative binomial distribution, if the conditional mean value parameter is  $\xi$  and the shape parameter is  $\alpha$  and the data are  $y$ , then the conditional variance is

$$\xi \left( 1 + \frac{\xi}{\alpha} \right)$$

We use this to estimate the shape parameter. Let  $A$  be a set of nodes all of which we think might be negative binomial with the same shape parameter, and let  $\hat{\xi}$  be the estimated conditional mean value parameter vector assuming the Poisson distribution. Then we equate empirical conditional variance with the formula above

$$\sum_{j \in A} (y_j - y_{p(j)} \hat{\xi}_j)^2 = \sum_{j \in A} y_{p(j)} \hat{\xi}_j \left( 1 + \frac{\hat{\xi}_j}{\alpha} \right) \quad (*)$$

where  $p(j)$  is the predecessor of  $j$ . The right-hand side is a decreasing function of  $\alpha$  and has infimum

$$\sum_{j \in A} y_{p(j)} \hat{\xi}_j \quad (**)$$

So long as the left-hand side of (\*) is greater than (\*\*) there will be a unique solution for  $\alpha$ . Otherwise there is no solution, in which case the  $y_j$  values are under-dispersed rather than over-dispersed, and negative binomial is not appropriate.

### 5.1 Tillers

#### 5.1.1 Observed and Conditional Mean Values

We assume the three arrows to the tillers nodes have the same over-dispersion. For these arrows, the predecessor is the constant 1. We have  $y_{p(j)} = 1$  in (\*) and (\*\*).

```
is.tiller <- grepl("tiller", redata$varb)
y.tiller <- redata$resp[is.tiller]
xi.tiller <- xi[is.tiller]
```

#### 5.1.2 Pearson Residuals

So-called Pearson residuals are deviations from the (estimated) mean divided by the (estimated) standard error. For Poisson (which we used for the fitted model we are diagnosing now) the standard deviation is the square root of the mean, hence

```
resid.pois.t <- (y.tiller - xi.tiller) / sqrt(xi.tiller)
stem(resid.pois.t, scale = 2)
```

```
##
## The decimal point is at the |
##
## -7 | 0
## -6 | 554
## -5 | 1
## -4 | 700
## -3 | 953
## -2 | 9430
## -1 |
## -0 | 44
## 0 | 0
## 1 |
```

```
##      2 | 28
##      3 | 588
##      4 | 47
##      5 |
##      6 |
##      7 | 124
##      8 | 34

resid.pois.tills <- stem(resid.pois.t, scale = 2)
```

```
##
## The decimal point is at the |
##
## -7 | 0
## -6 | 554
## -5 | 1
## -4 | 700
## -3 | 953
## -2 | 9430
## -1 |
## -0 | 44
##  0 | 0
##  1 |
##  2 | 28
##  3 | 588
##  4 | 47
##  5 |
##  6 |
##  7 | 124
##  8 | 34
```

We do not expect such large residuals in such a small sample. Thus we think we need negative binomial.

### 5.1.3 Estimating Shape Parameter for Tillers

```
lhs <- sum((y.tiller - xi.tiller)^2)
rhs.min <- sum(xi.tiller)
lhs > rhs.min
```

```
## [1] TRUE
```

Thus we can fit negative binomial. Write a function the zero of which is our estimate of the shape parameter.

```
baz <- function(alpha) lhs - sum(xi.tiller * (1 + xi.tiller / alpha))
```

Then we find two points where this function has opposite signs and feed it to R function uniroot.

```
baz(1)
```

```
## [1] -573518.5
```

```
baz(10)
```

```
## [1] 34305.9
```

```
uout <- uniroot(baz, c(1, 10), tol = sqrt(.Machine$double.eps))
uout
```

```
## $root
```

```
## [1] 6.631457
##
## $f.root
## [1] 0
##
## $iter
## [1] 8
##
## $init.it
## [1] NA
##
## $estim.prec
## [1] 0.002350784
```

Looks like we want negative binomial with shape parameter 6.6314567 for this first arrow.

```
famlist <- list(fam.bernoulli(), fam.poisson(),
               fam.negative.binomial(uout$root))
# assign the nb to all tiller nodes
fam[grep("tillers", vars)] <- 3
famlist
```

```
## [[1]]
## [1] "bernoulli"
##
## [[2]]
## [1] "poisson"
##
## [[3]]
## [1] "negative.binomial(size = 6.63145669255945)"
```

#### 5.1.4 Model fitting

Now do everything all over again, and check for over-dispersion for florets.

```
aout <- aster(resp ~ varb + fit : (symp.extent + ind),
             pred, fam, varb, id, root, data = redata, famlist = famlist)
```

```
## Warning in aster.default(x, root, pred, fam, modmat, parm, type, famlist, :
## Algorithm did not converge
```

To avoid convergence trouble, let's bump up the max iterations and then see if convergence comes more easily after we finalize the shape parameter estimates.

```
aout <- aster(resp ~ varb + fit : (symp.extent + ind),
             pred, fam, varb, id, root, data = redata, famlist = famlist, maxiter = 20000)
```

## 5.2 Florets

Now we look at terminal arrows, using the same shape parameter for all years.

### 5.2.1 Observed, Predecessors, and Conditional Mean Values

```
is.floret <- grep("floret.count.total", redata$varb)
is.floret.pred <- grep("pans.sampled", redata$varb)
y.floret <- redata$resp[is.floret]
```

```
y.floret.pred <- redata$resp[is.floret.pred]
xi.floret <- xi[is.floret]
```

### 5.2.2 Pearson Residuals

```
resid.pois.f <- (y.floret - y.floret.pred * xi.floret) / sqrt(y.floret.pred * xi.floret)
summary(resid.pois.f)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
## -36.646  -9.606   1.307   1.047  10.897   72.895
```

```
stem(resid.pois.f, scale = 2)
```

```
##
## The decimal point is 1 digit(s) to the right of the |
##
## -3 | 70
## -2 | 5
## -1 | 86550
## -0 | 988754
## 0 | 1145799
## 1 | 013347
## 2 | 08
## 3 |
## 4 |
## 5 |
## 6 |
## 7 | 3
```

```
resid.pois.florets <- stem(resid.pois.f, scale = 2)
```

```
##
## The decimal point is 1 digit(s) to the right of the |
##
## -3 | 70
## -2 | 5
## -1 | 86550
## -0 | 988754
## 0 | 1145799
## 1 | 013347
## 2 | 08
## 3 |
## 4 |
## 5 |
## 6 |
## 7 | 3
```

We do not expect such large residuals (more than 4 standard deviations from the mean) in such a small sample. Thus we think we need negative binomial.

### 5.2.3 Estimating Shape Parameter for Florets

```
lhs.f <- sum((y.floret - y.floret.pred * xi.floret)^2)
rhs.min.f <- sum(y.floret.pred * xi.floret)
lhs.f > rhs.min.f
```

```
## [1] TRUE
```

Thus we can fit negative binomial. Write a function the zero of which is our estimate of the shape parameter.

```
baz.f <- function(alpha) lhs.f -  
  sum(y.floret.pred * xi.floret * (1 + xi.floret / alpha))
```

Then we find two points where this function has opposite signs and feed it to R function uniroot.

```
baz.f(1)
```

```
## [1] -4097496
```

```
baz.f(10)
```

```
## [1] 37998248
```

```
uout.f <- uniroot(baz.f, c(1, 10), tol = sqrt(.Machine$double.eps))  
uout.f
```

```
## $root  
## [1] 1.096015  
##  
## $f.root  
## [1] 0.0002074242  
##  
## $iter  
## [1] 7  
##  
## $init.it  
## [1] NA  
##  
## $estim.prec  
## [1] 7.450581e-09
```

Looks like we want negative binomial with shape parameter 1.0960151 for these terminal arrows.

```
famlist <- c(famlist, list(fam.negative.binomial(uout.f$root)))  
famlist
```

```
## [[1]]  
## [1] "bernoulli"  
##  
## [[2]]  
## [1] "poisson"  
##  
## [[3]]  
## [1] "negative.binomial(size = 6.63145669255945)"  
##  
## [[4]]  
## [1] "negative.binomial(size = 1.09601506363191)"  
fam[grepl("floret", vars)] <- 4  
fam
```

```
## [1] 3 1 1 4 3 1 1 4 3 1 1 4
```

### 5.2.4 Model Fitting

Fit model with NB distributions for tillers and florets

```
aout <- aster(resp ~ varb + fit : (symp.extent + ind),
  pred, fam, varb, id, root, data = redata, famlist = famlist)
```

### 5.3 Redo Conditional and Unconditional Mean Value Parameters

```
pout.cond <- predict(aout, model.type = "conditional",
  is.always.parameter = TRUE, gradient = TRUE)
xi <- pout.cond$fit
pout.unco <- predict(aout, gradient = TRUE)
mu <- pout.unco$fit
ximu <- c(xi, mu)
```

### 5.4 Redo Jacobian matrices

```
jac.foo <- jacobian(foo, ximu)
jac.ximu <- rbind(pout.cond$gradient, pout.unco$gradient)
jac.total <- jac.foo %*% jac.ximu
```

### 5.5 Estimate fitness

Re-do the delta method to estimate fitness.

```
V <- jac.total %*% solve(aout$fisher) %*% t(jac.total)

se <- sqrt(diag(V))
bar.nb1 <- cbind(foo(ximu), se)
colnames(bar.nb1) <- c("Estimate", "SE")
```

Table 2: Estimated Fitness with Standard Error for Different Individuals (Initial Negative Binomial Distributions for Tillers and Florets)

Estimate	SE
89147.71	23208.04
112386.97	28338.03
128489.18	32061.15
62850.23	17022.06
128860.78	31942.63
106362.79	26877.86
181763.34	44533.71
113017.06	28360.42
119496.72	29896.84
189029.06	45572.24

## 6 Re-estimating over-dispersion for final model

Now that we have fit the `ind` model with two negative binomial distributions, we

- re-estimate  $\xi$ ,
- re-estimate the negative binomial shape parameters based on this new  $\hat{\xi}$ .

And we do this repeatedly until the estimates of shape parameters converge. We'll use the initial estimates for the NB shape parameters as the starting point for the model.

```

shapes.save <- lapply(famlist, function(x) x$size)
shapes.save <- unlist(shapes.save)

for (i in 1:7) {
xi <- predict(aout, model.type = "conditional", is.always.parameter = TRUE)

# tillers
xi.tiller <- xi[is.tiller]

lhs <- sum((y.tiller - xi.tiller)^2)
rhs.min <- sum(xi.tiller)
stopifnot(lhs > rhs.min)

baz <- function(alpha) lhs - sum(xi.tiller * (1 + xi.tiller / alpha))
uout <- uniroot(baz, c(2, 20), tol = sqrt(.Machine$double.eps),
  extendInt = "yes")
famlist[[3]] <- fam.negative.binomial(uout$root)

# florets
xi.floret <- xi[is.floret]

lhs <- sum((y.floret - y.floret.pred * xi.floret)^2)
rhs.min <- sum(y.floret.pred * xi.floret)
stopifnot(lhs > rhs.min)

baz <- function(alpha) lhs -
  sum(y.floret.pred * xi.floret * (1 + xi.floret / alpha))
uout <- uniroot(baz, c(1/2, 2), tol = sqrt(.Machine$double.eps),
  extendInt = "yes")
famlist[[4]] <- fam.negative.binomial(uout$root)

aout <- aster(resp ~ varb + fit : (symp.extent + ind),
  pred, fam, varb, id, root, data = redata, famlist = famlist,
  maxiter = 20000)

shapes.tmp <- lapply(famlist, function(x) x$size)
shapes.save <- rbind(shapes.save, unlist(shapes.tmp))
}
rownames(shapes.save) <- NULL
colnames(shapes.save) <- c("tillers", "florets")

```

Let's take a look at where the shape parameters converged for the negative binomial model.

Table 3: Estimated shape parameters of negative binomial distributions, each row one iteration

tillers	florets
6.6315	1.0960
11.9381	1.2094
10.1647	1.1872
10.6408	1.1950

tillers	florets
10.5046	1.1930
10.5430	1.1936
10.5322	1.1935
10.5352	1.1935

The initial NB shape parameter for tillers was 6.631, and it converged at 10.535. The change in the shape parameter for florets was less drastic. Initially, it was 1.096, and it converged at 1.193.

## 7 Evaluating final model

The famlist was automatically updated during the convergence process, so we just need to rerun the null and final model in order to compare them.

```
anull <- aster(resp ~ varb, pred, fam, varb, id, root, data = redata, famlist = famlist)

aout <- aster(resp ~ varb + fit : (symp.extent + ind),
  pred, fam, varb, id, root, data = redata, famlist = famlist)

anova(anull, aout)
```

```
## Analysis of Deviance Table
##
## Model 1: resp ~ varb
## Model 2: resp ~ varb + fit:(symp.extent + ind)
##   Model Df Model Dev Df Deviance P(>|Chi|)
## 1      12    -14909
## 2      22   -14889 10    19.719   0.03203 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

With the final shape parameters for tillers and florets for the ind model, we didn't have any convergence trouble. We also see that our final model is still explaining a significant amount of variation relative the null.

## 8 Final fitness estimates

### 8.1 Get Conditional and Unconditional Mean Value Parameters

```
pout.cond.f <- predict(aout, model.type = "conditional",
  is.always.parameter = TRUE, gradient = TRUE)
xi <- pout.cond.f$fit
class(xi)

## [1] "numeric"

length(xi) == nind * nnode

## [1] TRUE

xi <- matrix(xi, nrow = nind)
colnames(xi) <- vars
xi

##      late17.tillers late17.panicles late17.pans.sampled
```



```
## [1,] 90.67461 0.8187365 0.07986425
## [2,] 105.28175 0.8213544 0.09604337
## [3,] 110.67019 0.8221456 0.10091307
## [4,] 85.22798 0.8175307 0.07237712
## [5,] 124.38953 0.8238506 0.11137532
## [6,] 108.97500 0.8219051 0.09943399
## [7,] 145.55836 0.8258509 0.12359413
## [8,] 116.43644 0.8229112 0.10561608
## [9,] 111.89728 0.8223151 0.10195526
## [10,] 157.88888 0.8267688 0.12918153
## late17.floret.count.total late18.tillers late18.panicles
## [1,] 307.3460 125.2207 0.8018327
## [2,] 364.3105 133.3189 0.8027668
## [3,] 381.5037 143.5432 0.8037956
## [4,] 280.9967 102.9123 0.7984990
## [5,] 418.5753 143.7408 0.8038141
## [6,] 376.2781 132.4406 0.8026710
## [7,] 462.1763 194.4716 0.8073089
## [8,] 398.1430 133.5357 0.8027903
## [9,] 385.1878 133.0589 0.8027386
## [10,] 482.2485 178.7573 0.8064384
## late18.pans.sampled late18.floret.count.total late19.tillers
## [1,] 0.08937983 346.1512 163.5410
## [2,] 0.09472708 365.2839 188.0090
## [3,] 0.10060202 386.3426 205.2266
## [4,] 0.07019538 277.5825 135.8233
## [5,] 0.10070720 386.7200 182.9424
## [6,] 0.09417931 363.3227 167.5384
## [7,] 0.12055089 458.3223 205.0083
## [8,] 0.09486116 365.7640 170.6834
## [9,] 0.09456566 364.7059 195.7690
## [10,] 0.11562430 440.4627 226.4587
## late19.panicles late19.pans.sampled late19.floret.count.total
## [1,] 0.7451543 0.09655191 276.6657
## [2,] 0.7471616 0.10607578 302.1346
## [3,] 0.7482871 0.11139389 316.4095
## [4,] 0.7420068 0.08151413 236.6059
## [5,] 0.7467900 0.10431668 297.4219
## [6,] 0.7455223 0.09830178 281.3375
## [7,] 0.7482740 0.11133216 316.2436
## [8,] 0.7457997 0.09961978 284.8583
## [9,] 0.7476934 0.10859044 308.8791
## [10,] 0.7494395 0.11682196 331.0277
```

```
pout.unco.f <- predict(aout, gradient = TRUE)
mu <- pout.unco.f$fit
mu <- matrix(mu, nrow = nind)
colnames(mu) <- vars
mu
```

```
## late17.tillers late17.panicles late17.pans.sampled
## [1,] 90.67461 74.23861 5.929011
## [2,] 105.28175 86.47362 8.305218
## [3,] 110.67019 90.98700 9.181778
## [4,] 85.22798 69.67649 5.042984
```

```
## [5,]      124.38953      102.47839      11.413563
## [6,]      108.97500       89.56711       8.906015
## [7,]      145.55836      120.20950      14.857189
## [8,]      116.43644       95.81684      10.119799
## [9,]      111.89728       92.01482       9.381395
## [10,]     157.88888      130.53761      16.863048
##      late17.floret.count.total late18.tillers late18.panicles
## [1,]              1822.258       125.2207      100.40607
## [2,]              3025.678       133.3189      107.02402
## [3,]              3502.882       143.5432      115.37937
## [4,]              1417.062       102.9123       82.17538
## [5,]              4777.436       143.7408      115.54086
## [6,]              3351.138       132.4406      106.30625
## [7,]              6866.640       194.4716      156.99862
## [8,]              4029.127       133.5357      107.20119
## [9,]              3613.599       133.0589      106.81150
## [10,]             8132.179       178.7573      144.15673
##      late18.pans.sampled late18.floret.count.total late19.tillers
## [1,]             8.974277          3106.457      163.5410
## [2,]            10.138073          3703.275      188.0090
## [3,]            11.607398          4484.432      205.2266
## [4,]             5.768332          1601.188      135.8233
## [5,]            11.635796          4499.795      182.9424
## [6,]            10.011850          3637.532      167.5384
## [7,]            18.926324          8674.356      205.0083
## [8,]            10.169230          3719.538      170.6834
## [9,]            10.100700          3683.785      195.7690
## [10,]           16.668020          7341.641      226.4587
##      late19.panicles late19.pans.sampled late19.floret.count.total
## [1,]             121.8633          11.766132      3255.285
## [2,]             140.4731          14.900797      4502.047
## [3,]             153.5684          17.106582      5412.685
## [4,]             100.7818           8.215139      1943.750
## [5,]             136.6196          14.251701      4238.768
## [6,]             124.9036          12.278243      3454.330
## [7,]             153.4024          17.078621      5401.004
## [8,]             127.2956          12.681160      3612.334
## [9,]             146.3752          15.894944      4909.616
## [10,]            169.7171          19.826680      6563.180
```

## 8.2 Correcting for Sub-sampling

### 8.2.1 Point Estimates

```
is.florets <- grep("floret", vars)
is.panicles <- grep("panicles", vars)
is.florets
```

```
## [1] 4 8 12
```

```
is.panicles
```

```
## [1] 2 6 10
```

```
mu.panicles <- mu[ , is.panicles]
xi.florets <- xi[ , is.florets]
```

```
mu.florets <- mu.panicles * xi.florets
mu.florets
```

```
##      late17.panicles late18.panicles late19.panicles
## [1,]      22816.94      34755.68      33715.39
## [2,]      31503.25      39094.15      42441.80
## [3,]      34711.88      44575.96      48590.51
## [4,]      19578.86      22810.45      23845.56
## [5,]      42894.93      44681.97      40633.66
## [6,]      33702.14      38623.47      35140.06
## [7,]      55557.98      71955.96      48512.53
## [8,]      38148.81      39210.34      36261.22
## [9,]      35442.99      38954.79      45212.23
## [10,]     62951.56      63495.66      56181.05
```

Now calculate the final fitness estimates.

```
mu.fit <- rowSums(mu.florets)
mu.fit
```

```
## [1] 91288.01 113039.20 127878.35 66234.87 128210.55 107465.67 176026.47
## [8] 113620.36 119610.00 182628.26
```

Check

```
ximu <- c(xi, mu)
all.equal(foo(ximu), mu.fit)
```

```
## [1] TRUE
```

Final Jacobian matrices

```
jac.foo <- jacobian(foo, ximu)
jac.ximu <- rbind(pout.cond.f$gradient, pout.unco.f$gradient)
jac.total <- jac.foo %*% jac.ximu
```

Delta method for final fitness estimates.

```
V <- jac.total %*% solve(aout$fisher) %*% t(jac.total)
```

The standard errors are square roots of the variances (the diagonal elements of V)

```
se.final <- sqrt(diag(V))
bar.final <- cbind(mu.fit, se.final)
colnames(bar.final) <- c("Estimate", "SE")
```

Table 4: Estimated Fitness with Standard Error for Different Individuals (Final Negative Binomial Distributions for Tillers and Florets)

Estimate	SE
91288.01	19816.16
113039.20	23636.53
127878.35	26353.40
66234.88	15136.14
128210.55	26297.39
107465.66	22584.03
176026.47	35303.11
113620.36	23671.55

Estimate	SE
119610.00	24776.94
182628.26	36099.52

**Caution:** Standard errors involving negative binomial arrows do not account for estimating the shape parameters of these negative binomial distributions. Whenever such are presented, some academic weasel wording must be emitted to refer to this fact. More precisely, the standard errors in Table 3 assume the size parameters of the negative binomial distributions are known rather than estimated. They do correctly account for sampling variability under that assumption, asymptotically (for sufficiently large sample size).

## 9 Comparing Pearson residuals for models

```
is.tiller <- grepl("tiller", redata$varb)
y.tiller <- redata$resp[is.tiller]
xi.tiller <- xi[is.tiller]
resid.t.final <- (y.tiller - xi.tiller) /
  sqrt(xi.tiller * (1 + xi.tiller / famlist[[3]]$size))
summary(resid.t.final)
```

```
##      Min.   1st Qu.   Median     Mean  3rd Qu.     Max.
## -1.77192 -0.87328 -0.24797 -0.07649  0.72515  2.14036
```

```
stem(resid.t.final, scale = 1)
```

```
##
## The decimal point is at the |
##
## -1 | 876
## -1 | 321
## -0 | 997765
## -0 | 4432
## 0 | 12
## 0 | 666778
## 1 | 11122
## 1 |
## 2 | 1
```

Compare these to the Pearson residuals for tillers from the initial Poisson model.

```
stem(resid.pois.t, scale = 2)
```

```
##
## The decimal point is at the |
##
## -7 | 0
## -6 | 554
## -5 | 1
## -4 | 700
## -3 | 953
## -2 | 9430
## -1 |
## -0 | 44
## 0 | 0
## 1 |
```

```
## 2 | 28
## 3 | 588
## 4 | 47
## 5 |
## 6 |
## 7 | 124
## 8 | 34
```

The residual analysis for the original model (with Poisson arrows for tillers and florets) clearly showed the model did not fit the data because the residuals were far larger than standard normal (which they would be only for very large sample sizes, which we do not have here, but still the residuals are far larger than they should be). The residual analysis for the final model (with negative binomial arrows for tillers and florets) shows no lack of fit of the model data because the residuals are the same size as standard normal residuals would be, although not quite standard normal in distribution, perhaps. But there isn't anywhere else in aster models to go. So we declare this model fits and move on (at least as far as tillers are concerned).

On to florets.

```
resid.f.final <-
  (y.floret - y.floret.pred * xi.floret) /
  sqrt(y.floret.pred * xi.floret * (1 + xi.floret / famlist[[4]]$size))
summary(resid.f.final)
```

```
##      Min.   1st Qu.     Median       Mean    3rd Qu.      Max.
## -2.164408 -0.654005 -0.006392  0.134414  0.738515  4.545859
```

```
stem(resid.f.final, scale = 2)
```

```
##
## The decimal point is at the |
##
## -2 | 2
## -1 | 5
## -1 | 2
## -0 | 9887755
## -0 | 444300
## 0 | 333
## 0 | 577889
## 1 | 234
## 1 | 5
## 2 |
## 2 |
## 3 |
## 3 |
## 4 |
## 4 | 5
```

Compare these to initial residuals from Poisson model

```
stem(resid.pois.f, scale = 2)
```

```
##
## The decimal point is 1 digit(s) to the right of the |
##
## -3 | 70
## -2 | 5
## -1 | 86550
## -0 | 988754
```

```
##    0 | 1145799
##    1 | 013347
##    2 | 08
##    3 |
##    4 |
##    5 |
##    6 |
##    7 | 3
```

We still have the one outlier. Clearly that one observation does not fit either Poisson or negative binomial model. But the final model shows no other issues. The residuals are (except for the outlier) about the same size as standard normal residuals.