# Hidden Alliances: RNA-Dependent Protein Interactions in Cancer Cells

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### Identifying RNA-Dependent Proteins from Proteomic Data

#### Why should we observe RNA-dependent proteins?

- Key Regulators: RBPs control RNA metabolism & gene expression.
- Disease Links: Missregulation is tied to cancer & neurodegeneration.
- Functional Clues: New RBPs hint at RNA's role in specific pathways.
- Molecular Insights: Deepens the understanding of cell cycle and cellular behavior

#### **Key Characteristics of Our Dataset**

- Dataset was created using the R-Deep approach and non-synchronized HeLa cells
- Contains 4765 proteins and their intensity values under normal (control) and RNase treated conditions in 25 sucrose density fractions, each fraction measured in triplicates

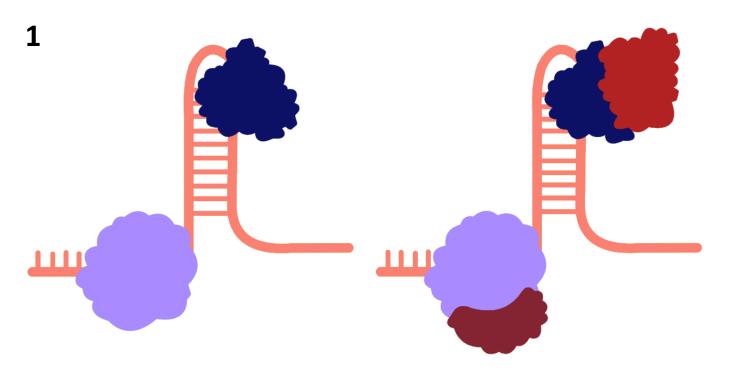


Fig. 1 Schematic illustration of RNA-dependency.

A protein is considered RNA-dependent if its interactome is dependent on RNA. It is either directly or indirectly attached to RNA & its functionality in biological context is associated with RNA

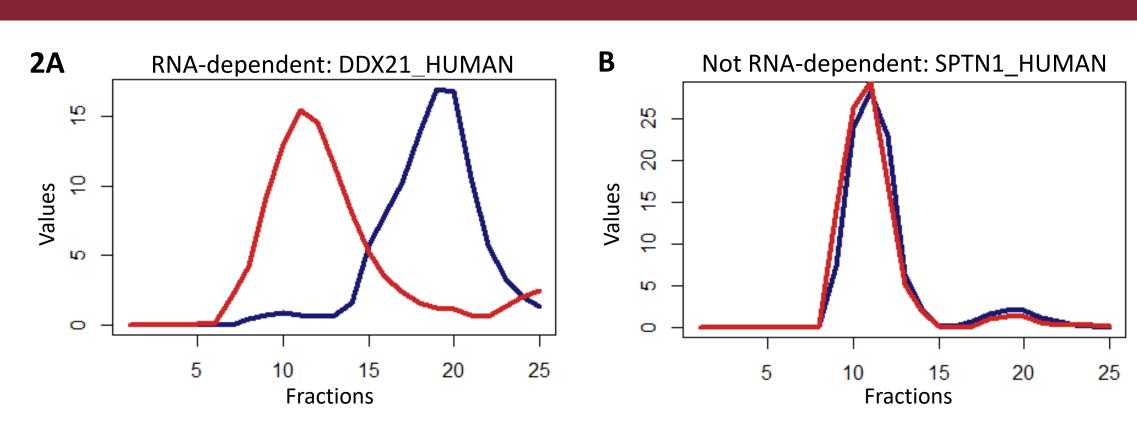


Fig. 2 Plot of proteins in data set. The data has been cleaned and normalized beforehand.

A) An RNA-dependent protein exhibits a shift for its intensity values between control (red) and RNasetreated (blue) group. B) A nor RNA-dependent protein exhibits no such shift.

# Our Approach and Results

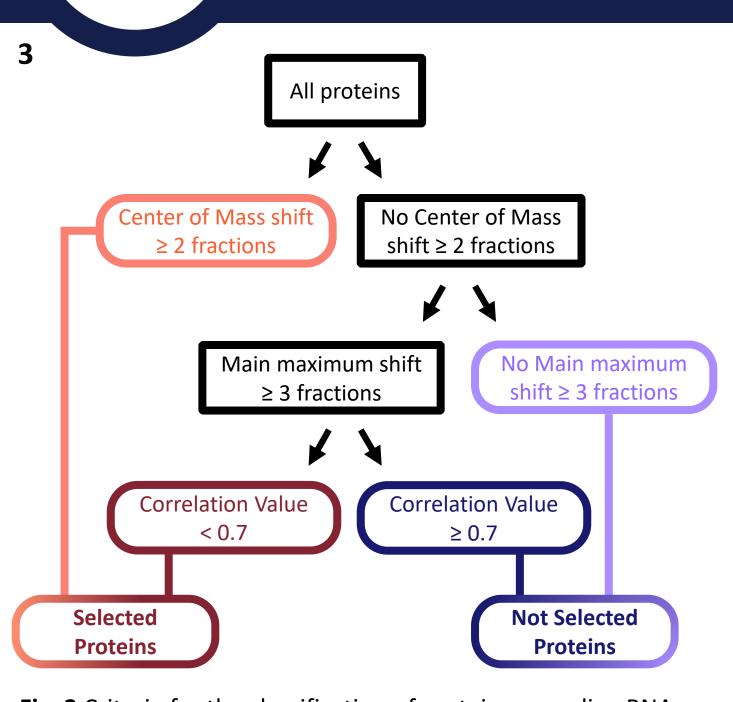


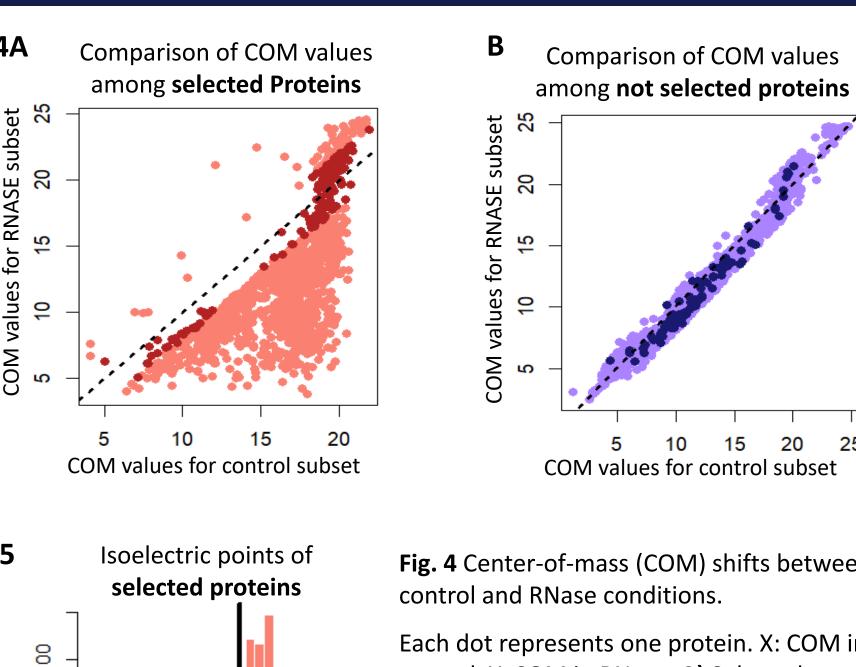
Fig. 3 Criteria for the classification of proteins regarding RNAdependency

### **Selected (RNA-dependent):**

- Center of mass (COM) shift ≥ 2 fractions (orange)
- or: no COM shift, but main peak shift ≥ 3 fractions and correlation < 0.7 (red)

#### **Not selected (not RNA-dependent):**

- No COM shift ≥ 2 and no main peak shift ≥ 3 (purple) • or: peak main shift without COM shift and correlation ≥ 0.7
- (dark blue)



0 2 4 6 8 10

Isoelectric point

Fig. 4 Center-of-mass (COM) shifts between

Each dot represents one protein. X: COM in control, Y: COM in RNase. A) Selected proteins. **B)** Not selected proteins

Fig 5. Isoelectric point (pI) distribution of selected proteins.

Mean pl = 8.16 (black line); one-sided t-test against pl = 7.0, p-value = 2.2e-16, indicates that selected proteins have significantly higher isoelectric points.

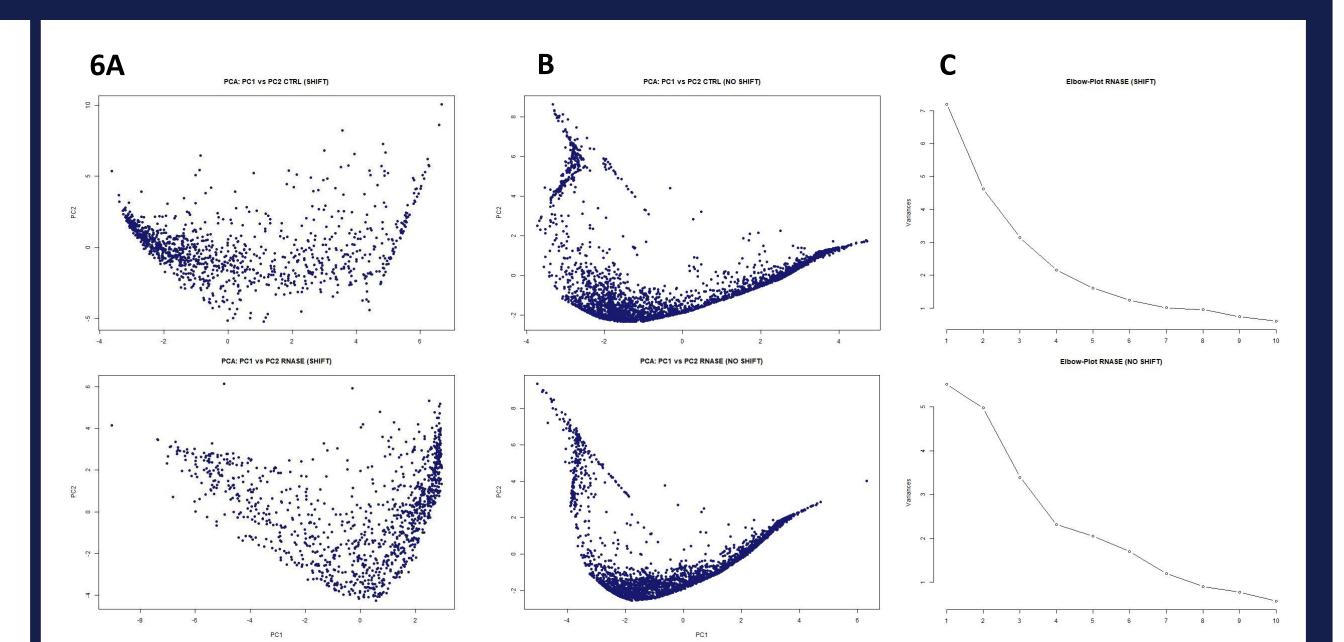


Fig. 6 Principal Component Analyses and Elbowplots of the selected and the non-selected proteins. RNASE and CTRL are plotted separately for comparison in the PCA. The Elbowplots shows only RNASE.

A) PCA of the selected proteins. The data points of the RNASE compared to the CTRL make up an overall similar shape, but a shift is visible in the density of the points. B) PCA of the non-selected proteins. The points of the RNASE and CTRL form mostly the same structure. C) Elbow-Plot of the RNASE. The knick of the elbow is between 3 and 4. To compare selected and non-selected proteins, we decided to use 3 clusters in the kmeans clustering.

RBP2GO-known non-RBPs

911

**Our Achievements** 

Not Selected RBP

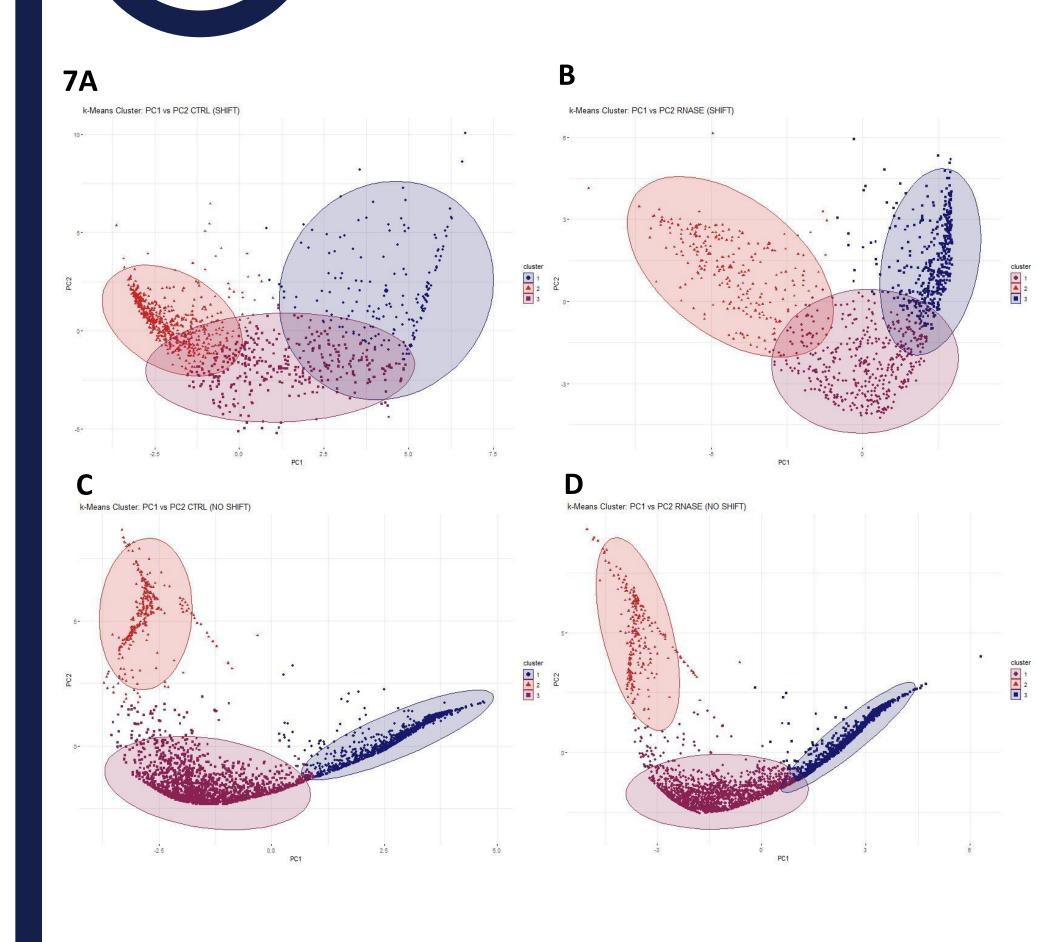
RBP2GO-annotated Non-RBPs in the dataset, showing wrongly selected (purple) versus not selected (blue).

A) Pie chart of RBP2GO-annotated RBPs in the dataset, showing selected (orange) versus not selected (red). B) Pie chart of

Selected RBP

Fig. 9 Selection Status of RBPs and Non-RBPs According to the RBP2GO Database

## Discussion



lm(formula = target ~ ctrl\_1 + rnase\_1, data = df\_regression\_selected\_11)

1Q Median 3Q Max

Estimate Std. Error t value Pr(>|t|)

-1.50827 0.02997 -50.321 < 2e-16 \*\*\*

-15.254 -0.896 -0.091 1.100 12.043

Residuals:

Fig. 7 kmeans clustering of the selected and not-selected proteins

**A)** Shows the 3 clusters of the CTRL of the selected proteins. **B)** Shows the 3 clusters of the RNASE of the selected proteins. A significant shift in form and location of the clusters is noticeable. **C)** Shows the 3 clusters of the CTRL of the not-selected proteins. **D)** Shows the 3 clusters of the RNASE of the not-selected proteins. No shift in form and location is noticeable.

Fig. 8 Linear regression analyses between the selected proteins and the not-selected proteins each, with global maxima of the selected CTRL proteins as target variable

A) The regression analysis for the selected proteins describes the target variable well. **B)** The analysis of the notselected proteins does not describe the target variable well. This proves, that there is a difference between the selected and not-selected proteins, and therefore, that the selection criteria worked.

References Sternburg et al., Global Approaches in Studying RNA-Binding Protein Interaction Networks, 2020, Trends in Biochemical Sciences Caudron-Herger et al., R-DeeP Proteome-wide and Quantitative Identification of RNA-Dependent Proteins by Density Gradient

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analysis of RNA-dependent proteins by RNase treatment and density

Corley et al., How RNA-Binding Proteins Interact with RNA Molecules

database on RNA-binding proteins, their interactions and functions,

gradient ultracentrifugation using R-DeeP, 2020, Nature Protocols

Caudron-Herger et al., RBP2GO: a comprehensive pan-species

Ultracentrifugation, 2019, Molecular Cell

and Mechanisms, 2020, Molecular Cell

2021, Nucleic Acids Research

RBP2GO-known RBPs

2438

1129



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Not Selected RBP

Selected RBP

lm(formula = target ~ ctrl\_ns\_1 + rnase\_ns\_2, data = df\_regression\_not\_selected\_12) Residuals:

1Q Median 3Q Max -15.404 -3.423 1.761 2.808 8.949 Coefficients:

Estimate Std. Error t value Pr(>|t|)(Intercept) 16.2759487 0.0707927 229.910 <2e-16 \*\*\* rnase\_ns\_2 -0.0607672 0.0320809 -1.894 0.0583 .

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Multiple R-squared: 0.001019, Adjusted R-squared: 0.000461

0.14991 0.02935 5.108 3.79e-07 \*\*\* Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1 Residual standard error: 2.061 on 1178 degrees of freedom Residual standard error: 4.238 on 3581 degrees of freedom Multiple R-squared: 0.7648, Adjusted R-squared: 0.7644 F-statistic: 1916 on 2 and 1178 DF, p-value: < 2.2e-16 F-statistic: 1.826 on 2 and 3581 DF, p-value: 0.1612