

# HYPERDIMENSIONAL COMPUTING FOR PROTEIN LANGUAGE MODELING

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# **SAMENVATTING**

nederlandse samenvatting





# **SUMMARY**

insert english summary here...



# 1. INTRODUCTION

## 1.1 outline

## 1.2 A historical perspective on bioinformatics

Many decades ago, around the 1950s, we did not know much about the molecules that carry our genetic information and how this would be translated into higher levels of biology. All that was known about deoxyribonucleic acid (DNA) was that it carries nucleotides in equimolar proportions so that there is as much guanine as cytosine and as much adenine as thymine. A major breakthrough came with Watson and Crick's discovery of the structure of DNA in 1953 [1]. Despite that, it took some more decades before the genetic code was deciphered and how this information is further transferred. Researchers came to know that DNA is essentially built up of a linear sequence of the four aforementioned nucleic acids. This sequence encodes information that undergoes transcription into ribonucleic acid (RNA) that in turn gets translated into proteins. The encoding of proteins is done in groups of three nucleic acids at a time, known as codons. All of this was later stated as the *central dogma of molecular biology*, also by Crick in 1958 [2]. This was hugely important for later research since it gives us more insight into how the genetic code is translated and transferred. In the same decade, major leaps were made in the research of protein structure and sequences. The first three-dimensional protein structures were determined via X-ray crystallography [3], which is still mostly the preferred method to this day. On top of that, the arrangement of the primary structure of a protein has been resolved after the first sequencing of a polypeptide. Sanger determined by sequencing insulin in 1953 [4] that a protein is built up of a sequence of amino acids, all connected by a peptide bond into a polypeptide. This established the idea that proteins are biological macromolecules that carry lots of information [5], which started a boom of research on more efficient methods for obtaining protein sequences. The most popular method of that time was the Edman degradation method [6]. A major issue with this method was

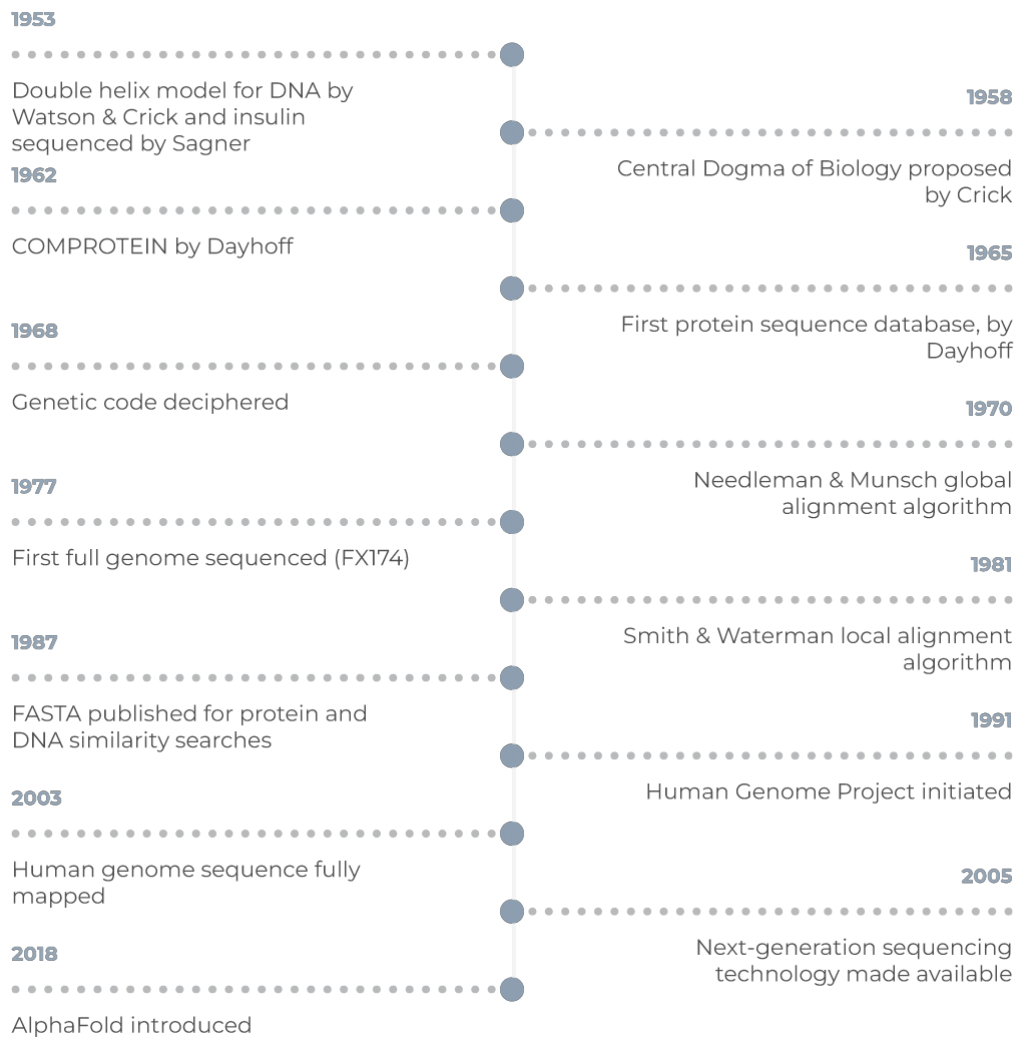


Figure 1.1: A brief timeline of important developments in bioinformatics.

that only a theoretical maximum of 50 to 60 sequential amino acids could be sequenced. Larger proteins had to be cleaved into fragments that were small enough to be sequenced. Tracing back the input sequence from this data was a cumbersome process and thus published Dayhoff the first computational program applied to biological data, COMPROTEIN [7] in 1962. This program was essentially a *de novo* sequence assembler for Edman degradation data. Furthermore, sequencing amino acids was also increasingly made automated later in the 1960s. These innovations assisted the creation of the first published protein sequence database [8] in 1965.

Further in the 1960s, researchers discovered the evolutionary value of having protein sequence data of different species. The problem to be solved back then was the quantification of the similarity between sequences. Pairwise alignment algorithms such as the algorithm of Needleman & Wun-

## 1. Introduction

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sch [9] for global alignments and that of Smith & Waterman [10] for local alignments from the 1970s solved this issue and alignment is still considered to be a key bioinformatics task to this day. Together with this, mathematical frameworks for amino acid substitutions in the context of evolution such as PAMs and BLOSUMs also contributed to bioinformatics. These pairwise alignment algorithms are sufficient for comparing two sequences but unfeasible for searching databases for homologous sequences, hence faster algorithms like FASTA [11], BLAST [12] were developed in the 1980s and 1990s. These methods were and are still important for discovering functional, structural and evolutionary information in biological sequences since sequences that are in some way similar, have a high chance to have the same biological function. This also means that such sequences might be derived from a common ancestor and it became commonplace that sequence patterns may lead to structural and functional relevance. A natural extension of pairwise alignments is multiple sequence alignment (MSA) [13], which is to align multiple related sequences. The most popular and still widely used tools today include Clustal [14] and MUSCLE [15]. This reveals much more information than pairwise alignment can. It allows for the identification of conserved sequence patterns and critical amino acid residues with much more statistical significance which is of great value for constructing phylogenetic profiles of gene families [16]. All of this showed the importance of computational biology and established bioinformatics as a beneficial field of science.

Development of DNA-based applications took some more time since the genetic code and how it translates to amino acids was not deciphered yet until 1968 [17]. Early DNA and RNA sequencing methods were first demonstrated in the 1970s [18, 19] and like with protein sequencing methods, these methods only became faster, more efficient and more scalable. The 1990s saw the appearance of whole genome sequencing and internet-accessible databases that we still use to this day, such as Genbank, Genomes and PubMed and so advances in computational biology followed the ever-increasing amount of data and need for processing power. With the advent of second-generation sequencing in the 2000s came even more Big Data issues, further challenging bioinformaticians by allowing us to sequence millions of DNA and RNA molecules in a single run. The sheer size and complexity of biological data can make it difficult to store and manage, as for instance implementing error identification, security, quality standards and easy data retrievability. On top of that, the analysis of such large-scale data is not straightforward and traditional software tools won't be sufficient

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anymore. Lastly, the need for high-performance computing resources to handle all of these computational demands will only rise.

Today we see that research in the field of biology is becoming more and more computationally driven and that this trend will not slow down any time soon. As for this thesis, we will focus on this matter, more specifically on the level of protein research which is discussed in the next section.

## 1.3 Protein biology

### 1.3.1 Protein structure levels

Proteins are an essential part of molecular biology and are responsible for almost all cellular functions. They are part of the important biological macromolecules that make up life, hence a lot of effort has gone towards trying to understand the functions of protein and disruptions in its mechanisms that lead to many kinds of diseases. Proteins are composed of a linear chain of amino acids (AA) with a length ranging from 50 to tens of thousands of AAs, all connected by peptide bonds into a polypeptide. This is also referred to as the *primary structure* of a protein as mentioned earlier [5]. A sequence of amino acids is mostly determined by the genetic code without considering post-translational and post-transcriptional modifications etc. In the genetic code of all living organisms, there are 20 different kinds of amino acids coded in that make up the 'language' of proteins. Each amino acid has the same backbone but differs by the chemical properties of its side chain, also known as the R-group. This sequence of amino acids does not occur as a mere linear chain of peptides and consists of much more intricacies, however. The polypeptide can form locally folded structures due to chemical interactions within the backbone (the polypeptide chain without the R-group), referred to as the *secondary structure* of a protein.  $\alpha$ -helices and  $\beta$ -sheets are the most well-known and common examples of these structures. The overall three-dimensional structure that forms out of these structures is referred to as the *tertiary structure*. These are formed due to interactions between the R-groups and are much harder to classify due to the quasi-infinite amount of different combinations that can occur in an amino acid sequence. And lastly, a protein can also be made up of multiple polypeptide chains, referred to as its subunits. These subunits together form the *quaternary structure* of a protein.

The sought-after biochemical and cellular functions of a protein emerge from a combination of all of these structures. While a protein's 3D structure and function are dynamic and dependent on its surroundings such as the cellular state and other proteins and molecules, it is still defined by its underlying sequence. This means that a lot of the 3D-structural and functional information of a protein should be retrievable from its amino acid sequence [20]. Understanding how a protein's sequence translates to its structure and function, otherwise known as the 'protein folding problem', is the central problem of protein biology and is crucial for understanding disease mechanisms and designing proteins and drugs for therapeutic and bioengineering applications. Therefore, a lot of effort has gone into computational methods for structure and function predictions from protein sequences but this sequence-structure-function relationship continues to challenge bioinformaticians (insert visual aid). Further in this chapter, we will discuss traditional and state-of-the-art bioinformatics methods to obtain more knowledge in this field.

### 1.3.2 Computational methods for protein research

As mentioned earlier, the Edman degradation method was mostly used before to determine the primary structure of a protein. The current state-of-the-art methods for the identification of protein sequences are *de novo sequencing* algorithms applied to tandem mass spectrometry data [21] and allow simultaneous sequencing of thousands of proteins per given sample. Plenty of valuable biological and evolutionary information is already retrievable from just the primary structure *via* traditional tools such as BLAST and MSA as discussed earlier and could provide more information for the prediction of secondary and tertiary structures. To cope with the number of recorded protein sequences rising exponentially, far more compute-efficient methods based on multiple sequence alignments had to be developed like PSI-BLAST [22], HHblits [23] and MMseqs [24]. However, these methods might not be able to keep up with the ever-increasing number of protein sequences stored in databases.

At the other end of the spectrum, the most common way to determine the 3D structure of a protein has remained to be X-ray crystallography for more than half a century [3], with cryo-electron microscopy now catching up rapidly [25]. However, these kinds of laboratory approaches for structure determination of proteins are complex, expensive and in some cases not possible for the protein in question whilst sequence determination is

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relatively much easier to perform. Because of that, the structures and functions for a large fraction of the approximately 20000 known human proteins remain unknown. The number of verified three-dimensional structures in protein databases consequently has not kept up with the explosive growth in sequence information, further increasing the demand for computational structure/function prediction models.

A lot of methods have been developed to tackle this problem. Until recently, these methods were mainly based on statistical sequence models or physics-based structural simulations. *Ab initio* physics-based approaches such as ROSETTA [26] solve this problem by searching the protein's conformational space using atom energy functions and minimizing the total free energy of the system. ROSETTA has shown to be effective at predicting unknown structures and has been widely used for varying applications, but also assumes simplified energy models, is extremely computationally intensive and has limited accuracies [27]. Statistical sequence modeling of a set of related proteins, on the other hand, has proven to be very useful for discovering evolutionary constraints, homology searches and predicting residue-residue contacts. Improvement of these models has mainly been data-driven; exploiting databases to build large deep learning systems which culminated in the recent success of DeepMind's AlphaFold2 [28] at the Critical Assessment of protein Structure Prediction (CASP) 14. This success of AlphaFold continued into more recently CASP 15 [29]. Even though, DeepMind did not participate, the most successful participants integrated AlphaFold into their methods. However, all of these models are supervised methods that require labels. Labeled protein structure data essentially means retrieving the 3D coordinates of every atom in a protein which is very labor-intensive and time-consuming. Also, such kind of models would likely perform poorly when working with completely unrelated proteins since the model won't be trained on this kind of data.

One underlying theme of these computational methods for protein structure prediction is being able to translate the protein 'language' into numerical representations which computers can learn from. This is now known as the field of protein language modeling which is more thoroughly discussed in section 1.4.



### 1.4 State-of-the-art protein language modeling

It is intuitive to represent a protein as a sequence of letters with each letter corresponding to an amino acid. As with natural languages, we can find common elements between naturally evolved proteins. Noticeable patterns reoccurring in multiple (related) protein sequences are highly likely to be biologically relevant. These motifs and domains are essential to many biological processes and can easily be represented as words, phrases or sentences of amino acids in a language model perspective. This is why researchers are taking inspiration from the recent successes of natural language processing (NLP) and applying this to a biological context. NLP is a branch of artificial intelligence (AI) concerning itself with creating the ability for computers to learn and understand human languages by using statistical, machine learning and in recent years deep learning models [30]. Common tasks in NLP include part-of-speech tagging (grouping words based on their function in a sentence), named entity recognition (recognizing specific entities in a sentence such as locations, persons, dates etc.) and natural language generation (letter/word prediction).

As with protein modeling, applying labels to millions of natural language containing web pages, articles, journals etc. is a labor-intensive procedure and thus state-of-the-art NLP models use a form of *self-supervised learning*, a form of unsupervised learning in which the context of the text is learned to fill in missing words, predict the next word in a sentence etc. during the training as shown in figure 1.2. Well-known NLP methods of this kind include bi-directional long-short term recurrent neural networks (biLSTMs) such as ELMo [31] and more recently transformers such as Google's BERT [32] and OpenAI's GPT-3 [33]. Despite the simplicity of these tasks, it is found to develop interesting capabilities as the scale of the model increases together with very little training on a specific task, now mostly referred to as *few-shot learning*.

These deep learning methods also show promise in the field of protein biology with the most notable latest projects being TAPE [34], ProtTrans [35] and Meta AI's ESM-2 [36], one of the most recent and largest protein language models to ever have been developed at the time of writing. ESM-2 has shown to accurately capture evolutionary information and to perform well on structure prediction tasks. It consists of transformer protein lan-

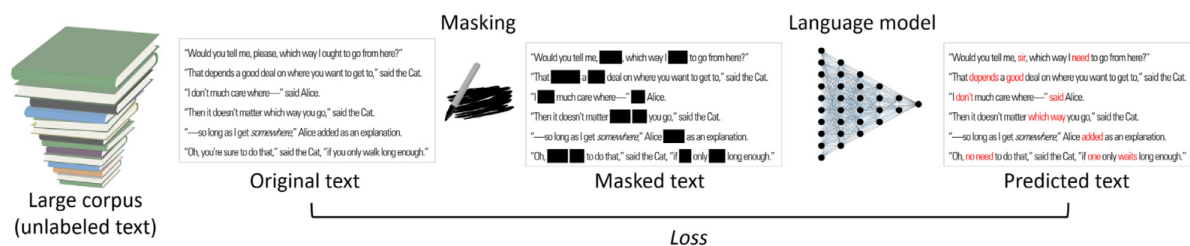


Figure 1.2: Demonstration of a self-supervised masked language learning model. A fraction of the unlabeled text is masked randomly and the language model will attempt to predict these masked tokens. The loss function is a method to assess the performance of the model on its predictions

language models with up to 15 billion parameters trained on 65 million unique sequences. Like with natural language models, it has been observed that larger models yield consistent improvement and that the performance does not seem to saturate with even the largest models [30]. The amount of computing power needed to train such models and to perform predictions with them rises exponentially with the number of parameters, close to the point that modern computers cannot sustainably handle anymore. For example, it is estimated that it would take at least 34 days with 1024 NVIDIA A100 GPUs to train the 175 billion parameter GPT-3 model with 300 billion tokens [37]. And although, these developments in computational modeling are very impressive, they are very costly too and cause a substantial strain on our environment in terms of the energy and materials needed to sustain these systems.

## **2. HYPERDIMENSIONAL** **COMPUTING**

Hyperdimensional computing (HDC) is a relatively new paradigm of computing in which data is represented and manipulated by high-dimensional (or hyperdimensional) vectors in the range of tens of thousands bit. This framework, developed by Kanerva [38], is inspired by the workings of the human brain and its ability to adapt, learn fast and easily understand semantic relations. The human brain consists of about 100 billion neurons (nerve cells) and 1000 trillion synapses that connect these neurons. Each neuron is connected to up to 10000 other neurons, creating massive circuits. This is likely fundamental to the workings of the human brain and what separates our brains from modern von Neumann computer architectures which operate on 8 to 64-bit vectors. This becomes clear when we compare the relative simplicity for a human to learn a language compared to computers. Computers use a large and complicated set of arithmetic operations in the form of deep learning networks which require terabytes of data and thousands of Watts of computing power to come close to mastering a language whilst a human can recognize other languages relatively easily when they don't even speak it. Likewise languages, we can very easily memorize and compare other intrinsically complex and contextual concepts such as images. A computer would have a hard time finding similarities between a set of images and faces because this requires very complex machine learning models. The human brain can do this all with a very large efficiency by consuming only roughly 20 W of energy.

Achieving these kinds of flexible brain-like models based on high dimensionality is not entirely new and is being explored since the 1990s. Some of these earlier models include Holographic Reduced Representations [39], Spatter Code [40] and others. A hyperdimensional vector (HDV) can represent anything from a scalar number to any kind of concept. This vector is initially made up of totally random elements, but with a simple set of operations which will be explained later, we can use other vectors to combine some concepts into new similar or dissimilar concepts. For example, to show

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the essence of HDC and how it tries to simulate the brain, we can compare the concept of a *table* to the concept of a *broccoli*. We would not immediately conclude that they are in any way similar but as humans, we can trace back *table* to *plate* which has some similarities with *food* from which we can easily extract the concept of *broccoli* as in equation 2.1. These kinds of operations are not very obvious for a classical computer but creating these semantic pathways and recognizing links between distant objects are rather easy for humans.

$$\begin{aligned} \text{table} &\neq \text{broccoli} \\ \text{table} &\approx \text{plate} \approx \text{food} \approx \text{broccoli} \end{aligned} \tag{2.1}$$

The elements in an HDV can be made up of binary bits (values from the set 0, 1) like in classical computing but also of bipolar (values from the set -1, 1) or real numbers. The choice of the nature of the elements has also implications on the nature of the different operations and possibly the results.

An initial HDV is made up fully randomly. This *holistic* or *holographic* representation of a concept smeared out over a vector consisting of thousands of bits gives rise to interesting properties such as its robustness. These kinds of systems are very tolerant to noise and failure of bits since we introduce a lot of redundancy in the vector just by stochastics. This is very unlike classical computing where every bit counts and one failure in a bit can lead to immediate data corruption. Besides its robustness, it also has the potential to show much faster and more efficient calculations than traditional computer systems since it allows for more efficient data storage by encoding multiple objects into a vector.

## 2.1 Operations on hyperdimensional vectors

The interesting properties of HDC are based on only four basic operations we can perform on HDVs. We will discuss these for bipolar and binary vectors.

### Addition

Also referred to as *bundling* or *aggregation*, the element-wise addition as in equation 2.2 of  $n$  input vectors  $\{X_1 + X_2 + \dots + X_n\}$  creates a vector  $X$  that

## 2. Hyperdimensional computing

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is similar to the input vectors.

$$X = X_1 + X_2 + \dots + X_n \quad (2.2)$$

For bipolar vectors, this entails a straightforward element-wise addition. The resulting vector is restricted to a bipolar nature too depending on the sign of each element, thus containing only  $-1$ ,  $1$  but allowing  $0$  for elements that are in disagreement as shown in the following 6-dimensional example.

$X_1 =$	$+1$	$-1$	$+1$	$+1$	$-1$	$-1$
$X_2 =$	$+1$	$+1$	$+1$	$-1$	$-1$	$-1$
$X_3 =$	$-1$	$-1$	$+1$	$+1$	$-1$	$+1$
$X_4 =$	$-1$	$-1$	$-1$	$+1$	$-1$	$+1$
<hr/>						
$X_1 + X_2 + X_3 + X_4 =$	$0$	$-1$	$+1$	$+1$	$-1$	$0$

For binary vectors, the vectors are element-wise bundled based on the majority element. This is no problem if an odd number of input vectors are considered but ambiguity rises when bundling an even set of vectors. This can be solved by setting the element in question randomly. [41] Another possibility is to add another random vector however this may seem to add more unnecessary noise, especially when bundling a low number of vectors. We can also reverse this by an *inverse addition*. For bipolar vectors, this means just multiplying the vector of interest by  $-1$ . A binary vector can be flipped bit-wise.

Similar to an ordinary arithmetic summation, the bundling addition of hyperdimensional vectors is commutative so the result is not dependent on the order of addition.

$$X_1 + X_2 = X = X_2 + X_1 \quad (2.3)$$

### Multiplication

Also referred to as *binding*, two vectors can be multiplied element-wise resulting in a vector maximally dissimilar to the input vectors. Vectors  $X$  and  $Y$  are bound together forming  $Z$  being orthogonal to  $X$  and  $Y$  as shown in equation 2.4.

$$Z = X * Y \quad (2.4)$$

This *binding* operation translates to a simple arithmetic element-wise multiplication for bipolar vectors. For binary vectors, this is represented by a *XOR* bit-operation shown as follows.

$$\begin{array}{rcccccc}
 X = & 1 & 0 & 1 & 1 & 0 & 0 \\
 Y = & 1 & 1 & 0 & 1 & 0 & 1 \\
 \hline
 X * Y = & 0 & 1 & 1 & 0 & 0 & 1
 \end{array}$$

This operation can also be undone by multiplying with the same vector again. It is its own inverse so that

$$A * A = O \text{ where } O \text{ is a vector containing only 0s} \quad (2.5)$$

Likewise an ordinary multiplication, this operation is commutative and distributive over additions, meaning that transforming a bundle of concepts with binding is equivalent to binding every element before bundling.

$$A = Z * (X + Y) = XZ + YZ \quad (2.6)$$

## Permutation

The permutation operation of an HDV, also known as *shifting*, is a simple reordering of the HDV. This can be random but a circular shift is widely employed [42] and makes the operation easily reversible. This results in a vector technically dissimilar from the input vector but still encoding its information. This will become important later when it will be used to encode sequential information such as tokens in a text. This operation will be denoted by  $\Pi$ .

$$\begin{array}{rcccccc}
 X = & 1 & 0 & 1 & 1 & 0 & 0 \\
 \hline
 \Pi(X) = & 0 & 1 & 0 & 1 & 1 & 0
 \end{array}$$

## Similarity measurement

For many kinds of problems, it will be necessary to quantify the similarity between two HDVs. The method depends on the nature of the vectors. For binary vectors, the *Hamming distance* defined as in equation 2.7 is widely used.

## 2. Hyperdimensional computing

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$$Ham(A, B) = \frac{1}{d} \sum_{i=1}^d 1_{A(i) \neq B(i)} \quad (2.7)$$

The *cosine distance* as defined in equation 2.8 is most commonly used for bipolar vectors.

$$cos(A, B) = \frac{A \cdot B}{||A|| * ||B||} \quad (2.8)$$

The results of both of these measurements are summarized in table 2.1.

Table 2.1: Overview of similarity measurements in HDC depending on the nature of the HDVs

Measurement	Dissimilar	Orthogonal	Similar
Hamming distance	1	0.5	0
Cosine similarity	-1	0	1

It is important to note that two random HDVs will be quasi-orthogonal to each other just by stochastics. Also notice that the first quantifies a distance and the latter a similarity.

## 2.2 Examples

There are many interesting possibilities given the relative simplicity of all these operations. We shall illustrate some applications and examples. From here, all implementations are written in the programming language Julia unless noted otherwise. Julia makes it possible to write incredibly efficient programs whilst still being a high-level and interpreter-based programming language with many packages suited for mathematical operations included. Also note that all examples have been implemented with binary hyperdimensional vectors unless stated otherwise. However with minimal changes, bipolar vectors could also be applied. All source code is provided in the appendices.

### Simple example with simulated data

To get a feel for the operations, assume  $A, B, C, X, Y$  and  $Z$  to be random 10000-dimensional bipolar hypervectors and that  $D = X * A + Y * B + Z * C$ , let us then try to retrieve  $A$  from  $D$  by using the defined operations. We

generate for  $A, B, C, X, Y$  and  $Z$  each a random 10000-D vector. To retrieve an approximation of  $A$ ,  $D$  can be multiplied by  $X$  and the rest of the included vectors are then regarded as noise as done in equations 2.9. Because of the robustness of hyperdimensional vectors, a lot of information of  $A$  should still be contained within  $D$ .

$$\begin{aligned}
 A' &= X * D \\
 &= X * (X * A + Y * B + Z * C) \\
 &= \underbrace{X * X * A}_A + \underbrace{X * Y * B + X * Z * C}_{\text{noise}} \\
 &\approx A
 \end{aligned} \tag{2.9}$$

The procedure of equation 2.9 is repeated 10000 times because of the stochastic nature of these vectors. The results are illustrated in figure 2.1. We see that we can retrieve a lot of information with most of the Hamming

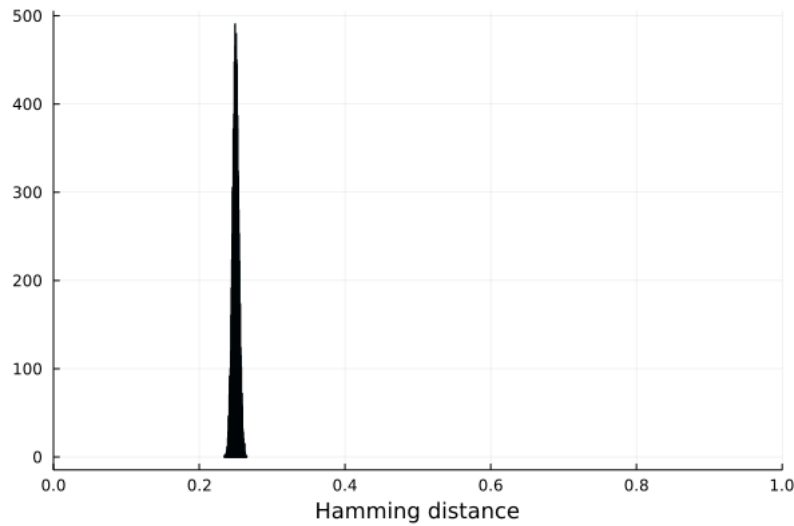


Figure 2.1: 10000 cases of random 10000-dimensional binary vectors are made and each time implemented following equation 2.9. The resulting Hamming distances between  $A$  and  $A'$  are then plotted in a histogram.

distances centering around 0.25. Notice that two completely random HDVs would have a distance close to 1 just by stochastics. A Hamming distance of 0 would mean that we retrieved all bits of  $A$  correctly, which is impossible in this model due to the consideration of noise. Although we work with a very constrained model, vector  $A$  is roughly 75 % retrievable and the calculations are very efficient as all of these can be completed in less than 2 seconds on a laptop. This same experiment was done with random bipolar 10000-dimensional vectors and it performs slightly slower as expected but retained the accuracy.



### 2.3 Examples of hyperdimensional computing with real datasets

Now, the power of these simple operations will be demonstrated by applying them to a couple of relatively small real datasets.

#### Zoo animal classification

As the first example, we will consider a simple dataset[43] containing 101 animals with 17 descriptors such as their number of legs, their skin covering and other physical properties. In the end, we want to create a simple model that can classify these animals and other animals that are not present in the dataset based on their descriptors. To tackle this problem, we first assign to each descriptor a random hyperdimensional vector. For each animal, all of its features can be bundled to obtain a final vector representing the animal. For example, it is known that a chicken lays eggs, is covered with feathers and has two legs so then these features can be bundled as in the following equation.  $C$  is a vector representing a chicken,  $E$  the ability to lay eggs,  $F$  the possession of feathers and  $T$  the possession two legs:

$$C = E + F + T \quad (2.10)$$

This is simple for all the binary features but the feature for the number of legs is variable. Although it is possible to assign completely random vectors to each number of legs, it would make a slightly more biologically realistic model if an animal with 2 legs would be more similar to one with 4 legs than to one with 8 legs. To address this, a range of numbers would have to be representable by hyperdimensional vectors, the range from 0 to 8 in this case. First, a random hyperdimensional vector representing the lower bound of the interval is generated. Next, a vector representing the next step in the interval is constructed by replacing a fraction of the vector with random bits. This last step is then repeated to obtain a vector of each number in a range.

In biology, it is possible to find higher order of concepts which are combinations of directly observable characteristics. For example, an animal could lay eggs or be dependent on mother's milk, but (almost) never both. So, the growth and development of an animal depends on these characteristics. This property can be easily implemented into this HDC model by binding a

HDV of a higher order concept to the descriptor HDV. So as said previously, the 'milk' ( $M$ ), and 'egg' ( $E$ ) features give us information about the growth of the animal, so we will bind these features to another vector representing the growth feature ( $G$ ) to obtain a more expanded model. This is also done for the skin protection features ( $S$ ) and all the features considering the limbs ( $L$ ). This also gives us the possibility to retrieve some features of the animals as in the procedure shown in the previous example. Thus, equation 2.10 can be expanded into:

$$C = G * E + S * F + L * T \quad (2.11)$$

After all these procedures, there are 101 animals with each a 10000-dimensional vector as a feature. To effectively analyze this data, a principal component analysis (PCA) has been applied to essentially reduce the 10000 features into 2. This projection can then be easily shown in a 2D plot, resulting in figure 2.2. This shows 3 distinguishable clusters of animal classes that make sense from an evolutionary standpoint. The model could be easily improved on this part by including more features that should show more separation of these classes.

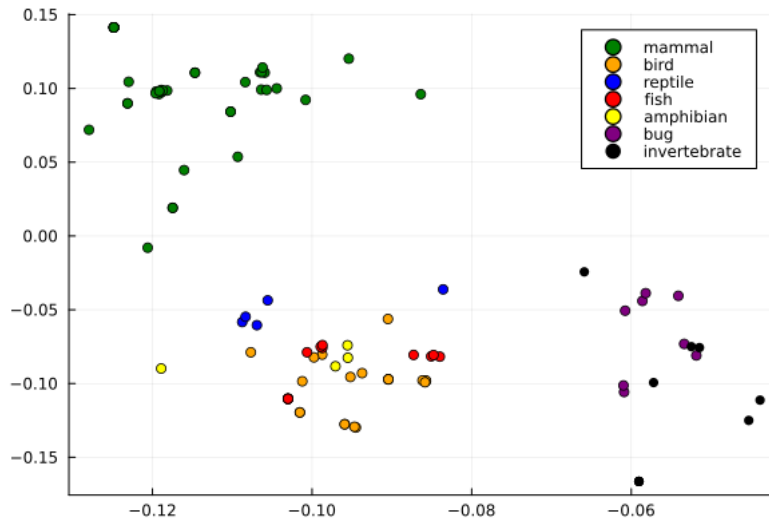


Figure 2.2: Scatter-plot of the first two principal components (PCs) of a  $101 \times 10000$  matrix containing hyperdimensional vectors for every animal in the zoo dataset after a PCA procedure. These PCs account for roughly 48 % of the variance.

After an HDV has been made for every animal, all animals of the same class can be bundled to obtain an HDV representing the said class. So for example, if we have a hyperdimensional vector for a pigeon ( $P$ ), chicken ( $C$ ) and

## 2. Hyperdimensional computing

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a kiwi ( $K$ ), an HDV representing birds ( $B$ ) can be made by doing:

$$B = P + C + K \quad (2.12)$$

To classify an animal, its HDV can be compared to the HDVs of every class and the most similar vector is then assumed to be its class. The dataset was stratified and split into a training set (comprising 80 % of the sequences) and a test set. With 100 runs, it could predict the class on average 94 % of the test animals. For further improvement, it would be possible to generate a set of animals not present in the dataset and test those in order to further understand how this model can be improved. On top of this, it would also be possible to generate a confusion matrix to understand where we could use more distinguishing descriptors. From the PCA, we could already predict that reptiles and amphibians would be easily confused, as for invertebrate and bugs.

### Protein classification

To illustrate an example more akin to this research topic, a model based on the principles of hyperdimensional computing will be built to classify a protein sequence dataset[44]. It contains 949 manually curated peptide sequences with their membranolytic anti-breast cancer activity level (very active, moderately active, experimentally inactive and virtually inactive). The virtually inactive peptides are predicted to be inactive. The model will be built with mostly the same procedure as for the animal classifier, but instead of animals, sequences have to be encoded into HDVs now. First, a random HDV is generated for every amino acid. Physicochemical properties, evolutionary constraints etc. could be introduced to make this model more realistic but that is not necessary for this demonstration. Next, a peptide sequence is to be considered as a bag of trimers as seen in figure 2.4. A vector representing a trimer is generated by binding the three amino acids whilst retaining sequential information by shifting as in equation 2.13. All retrievable trimers from a given sequence are then bundled together, forming a vector representative of the sequence.

$$ABC = A * \Pi(B) * \Pi(\Pi(C)) \quad (2.13)$$

From here on, the same procedures as in the last example can be applied here too, so all HDVs of a class are bundled for further analysis. The PCA

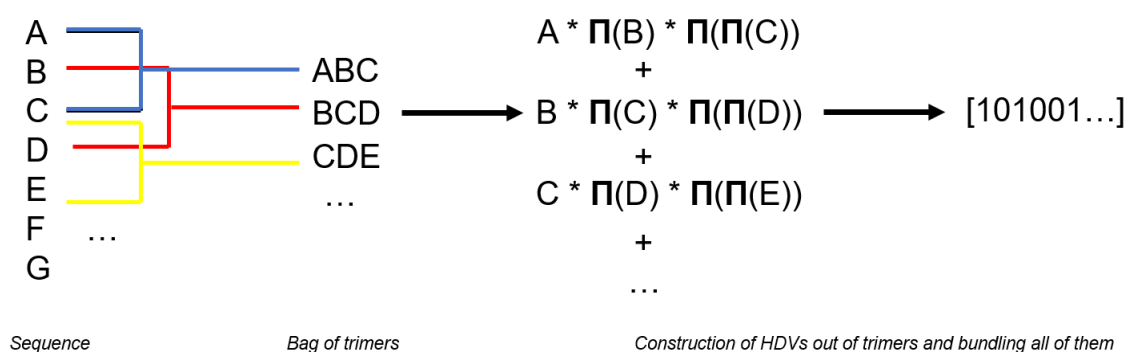


Figure 2.3: Overview of operations done to obtain an HDV of a protein sequence for the example concerning the real peptide dataset. First, a random HDV is generated for every amino acid. Next, a peptide sequence is to be considered as a bag of trimers. A vector representing a trimer is generated by binding the three amino acids whilst retaining sequential information by shifting as in equation 2.13. All retrievable trimers from a given sequence are then bundled together, forming a hyperdimensional vector representative of the sequence.

procedure did not generate interesting results because the two first principal components explained only 5 % of the variance, aside from the clear clustering of the predicted inactive peptides. This means that it is not feasible to reduce the 10000 dimensions of the vectors to 2, likely because the information is too smeared out over the vectors. This occurrence is highly dependent on the training data. Nevertheless, this follows the philosophy of hyperdimensional computing in keeping holistic representations of concepts.

## 2. Hyperdimensional computing

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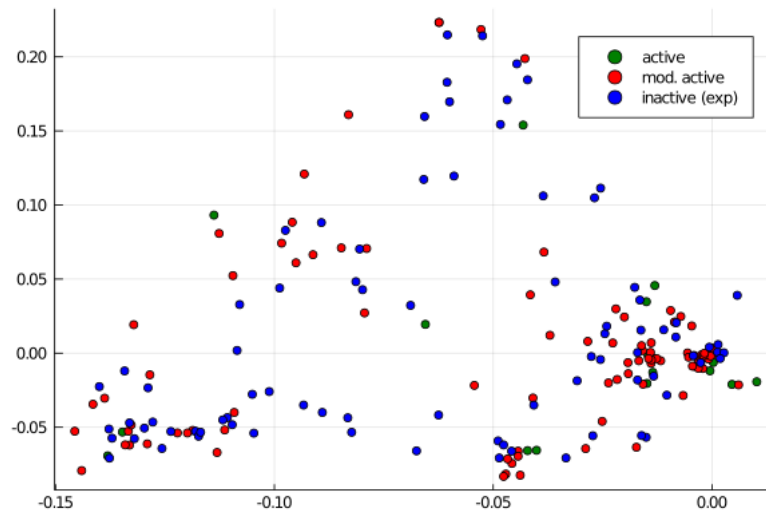


Figure 2.4: Scatter-plot of the first two principal components, decomposing the 10000-dimensional vectors for every peptide into 2. These PCs account for roughly 5 % of the total variance

Next, a classifier was made correspondingly. The dataset was stratified and split into a training set (comprising 80 % of the sequences) and a test set. With 100 runs, it could predict the class on average 85 % of the test sequences. It has to be taken into account that the predicted inactive peptides account for 80 % of the sequences of the dataset, thus this model performs slightly better than if we would predict at random. There are many possible improvements to be made however, such as using more suitable performance metrics, introducing similarities between amino acids instead of setting them randomly and using more suitable frameworks for our protein language models, which will all be research topics further on in this project.

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## 3. RESULTS

### 3.1 Hyperdimensional embeddings on the level of amino acids

#### Encoding biological information into a vector

Currently, in most of the research in hyperdimensional computing, there is an emphasis on creating and assigning hyperdimensional vectors fully randomly to certain concepts. This is useful for optimizing speed and efficiency and is not a problem for many cases such as natural language processing, where it is usually assumed that a letter does not have varying degrees of similarities to other letters in the alphabet. For protein language modeling, however, this assumption is not valid since some amino acids are chemically more similar to each other than to others, but also in an evolutionary sense. It is not uncommon to spot substitutions on the level of amino acids within the same proteins between different species and to barely notice changes in structure and function.

To account for this, we tried to encode the biological information of amino acids into HDVs by extending existing embeddings into hyperdimensionality and comparing these to baseline random hypervectors. To assess this, the vectors for each amino acids are reduced in dimensionality *via* PCA into 2 dimensions and then plotted as seen in figures 3.1 and 3.2. At first glance, there is not much to spot. Yet, there is significantly more variance encoded into the first two principal components of the ESM embeddings compared to the random vectors, meaning that there should be a significant amount of similarity encoded into the hyperdimensional vectors. This may be shown more clearly when used and compared in real-world problems.

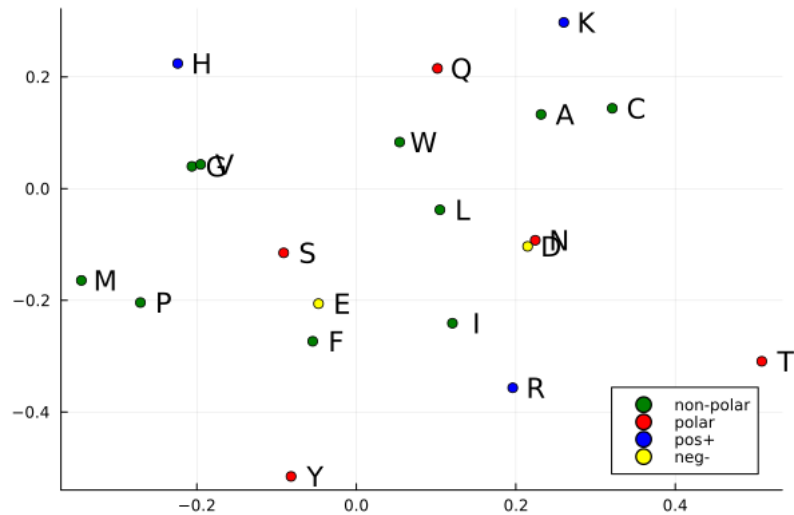


Figure 3.1: Scatter-plot of the first two principal components of random hyperdimensional vectors assigned to amino acids to set a baseline level. These PCs account for roughly 10.5 % of the total variance. The amino acids are annotated and colored based on their chemical property of polarity. This plot can look completely different depending on the run.

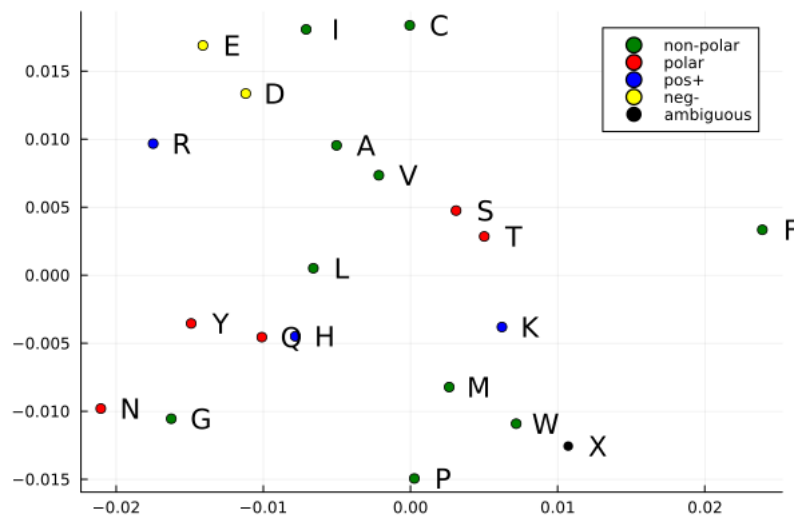


Figure 3.2: Scatter-plot of the first two principal components of ESM embeddings extended into hyperdimensionality. These PCs account for roughly 22 % of the total variance. The amino acids are annotated and colored based on their chemical property of polarity.

## Encoding interactions of a residue's neighborhood into a vector

State-of-the-art protein language models have the ability to gather information on long-range dependencies around a single amino acid and encode this



### 3. Results

information into neural networks and in embeddings. To investigate the possibilities of developing embeddings on the level of amino acids, we propose a novel encoding technique within the hyperdimensional computing framework. It encodes interactions of a given amino acid in a sequence to other amino acids in its neighborhood for a predetermined range. This method thus tries to learn information about an amino acid within a sequence in an unsupervised manner.

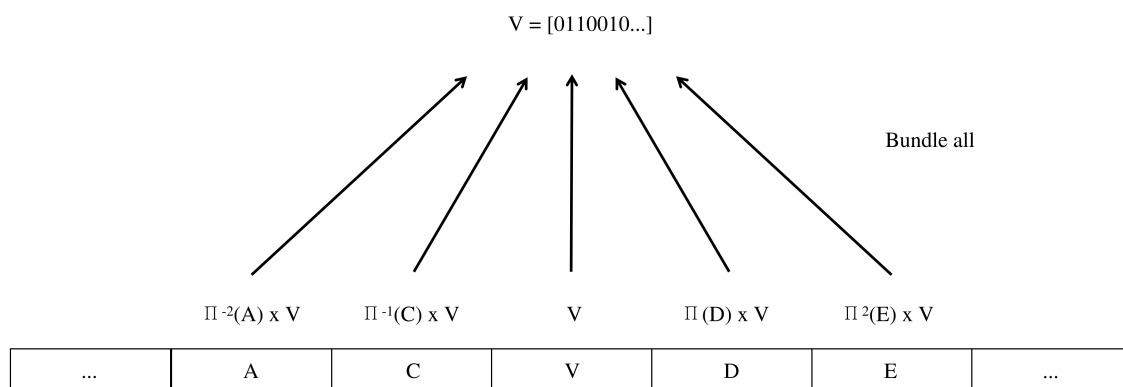


Figure 3.3: A simple demonstration of our amino acid encoder. It considers an amino acid and all amino acids in a predetermined distance (here  $k = 2$ ). It produces all possible interactions of the central amino acid in the window by binding and then bundles all the pairwise interactions into one hyperdimensional vector that represents the central amino acid.

Windows of  $k = 4$  and  $k = 50$  were considered for illustration purposes in figures 3.4 and 3.5. Every single residue in the human reference proteome was encoded with information within the  $k$ -range window and an average vector was made for every amino acid. For all 20591 peptides in the reference proteome, this procedure took only 3 hours for  $k = 4$ , but upwards to 20 hours for  $k = 50$  on a high-performance computing cluster (HPC). Both random HDVs and extended ESM embeddings were tested as base vectors. At first glance, the charged amino acids seem to be grouped vertically (PC 2), roughly dividing the polar and non-polar amino acids, albeit slightly more pronounced for  $k = 50$ . MORE TO ADD LATER WHEN TRIED WITH FULLY RANDOM VECTORS: Also interesting, the starting ESM embeddings are extended into hyperdimensionality in a random manner, so to see similar groupings in the PCA plots indicates that the first four amino acids before and after a residue are the most crucial.

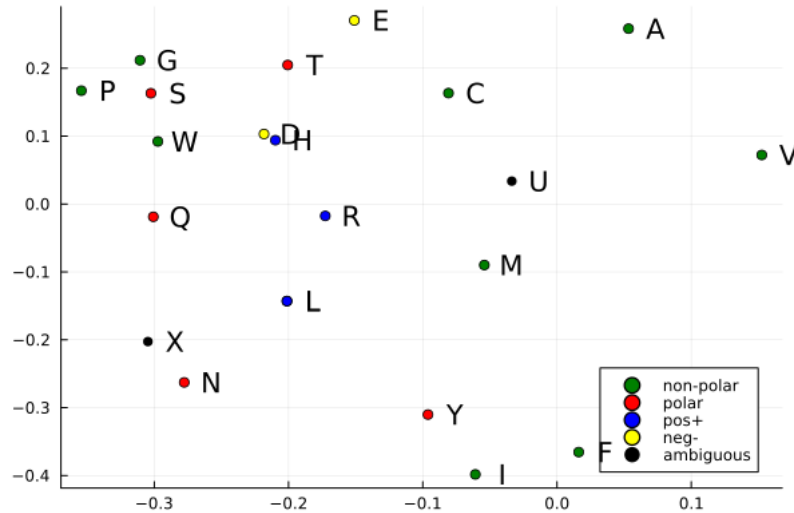


Figure 3.4: Scatter-plot of the first two principal components of the average amino acid HDVs with neighborhood-information of  $k = 4$  encoded, trained on the human reference proteome. These PCs account for roughly 21 % of the total variance. The amino acids are annotated and colored based on their chemical property of polarity.

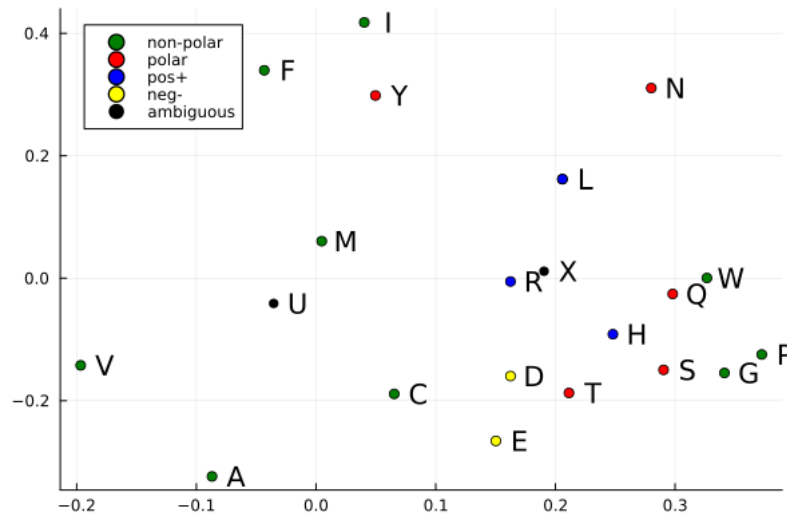


Figure 3.5: Scatter-plot of the first two principal components of the average amino acid HDVs with neighborhood-information of  $k = 50$  encoded, trained on the human reference proteome. These PCs account for roughly 21 % of the total variance. The amino acids are annotated and colored based on their chemical property of polarity.

## 3.2 Case study: PhaLP dataset

To implement and evaluate hyperdimensional computing in real-life problems, the potential of hyperdimensional computing will be evaluated on the

PhaLP dataset [45] for this chapter. PhaLP is a comprehensive database currently comprising more than 17000 entries of phage lytic proteins including much of their information such as their type, domains and tertiary structures. Phage lytic proteins are used by bacteriophages to infect bacterial cells. To cross the bacterial cell walls, phages use two different types of phage lytic proteins: virion-associated lysins (VALs) and endolysins. Phage lytic proteins also comprise one or more functional domains categorized into two classes: enzymatically active domains (EADs) and cell wall binding domains (CBDs).

All 17356 unique protein sequences were embedded into hyperdimensional vectors. This took only a few minutes for every method on a consumer laptop.

## Type classification

Only a fraction of the database is manually annotated to include the protein's type because the amount of phage lytic proteins whose type is described in the literature is relatively small. The developers of PhaLP resorted to a machine learning approach for the classification of unannotated sequences. They embedded each protein sequence *via* SeqVec [46] and trained a random forest classifier with 100 estimators and balanced weights to classify the proteins whose types were unknown. For this case study, we attempted to simulate their experiments of classifying the proteins into types based on their sequence using several methods.

First, we used several sequence encoding techniques tested on several kinds of base vectors. There was no visual difference between the PCA plots for the bag-of-words method and the convolutional method, but there is a clear difference between the different starting embeddings. The sequence embeddings from both the bag-of-words method and the convolutional method made with ESM AA embeddings seem to capture more of the variance between the sequences. UMAP projections were also tested and they showed all data points on one single line, thus failing to accurately reduce the high-dimensional data in our case compared to PCA.

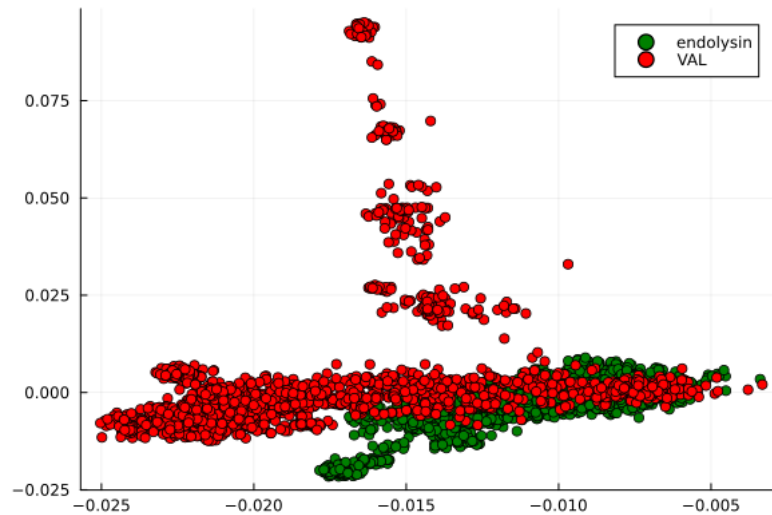


Figure 3.6: Scatter-plot of the first two principal components the encoded phage lytic protein. The sequences were encoded via the bag-of-words method starting from random hyperdimensional vectors. Only manually annotated phage lytic proteins were considered and are color-coded based on their type. These PCs account for roughly 7 % of the total variance in the system.

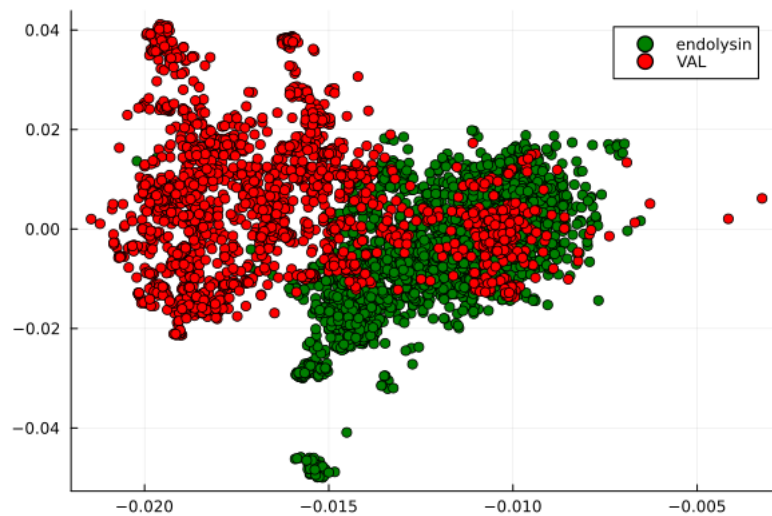


Figure 3.7: Scatter-plot of the first two principal components the encoded phage lytic protein. The sequences were encoded via the bag-of-words method starting from hyperdimensionally-extended ESM embeddings. Only manually annotated phage lytic proteins were considered and are color-coded based on their type. These PCs account for roughly 15.5 % of the total variance in the system.

### 3. Results

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Table 3.1: Results of type classifications using the principal classification technique of hyperdimensional computing and an XGBoost classifier with several kinds of embeddings

F1-scores	BoW random	BoW ESM	Convolutional random	Convolutional ESM
<b>Purely HDC</b>	0.1458	0.1468	0.1461	0.1461
<b>XGBoost classifier</b>	0.9667	0.9754	0.9661	0.986

#### Purely hyperdimensional

It is feasible to learn the classes of every sequence using only operations within the hyperdimensional computing framework. This is done by using the same techniques as in chapter 2.3. Evaluating our model using stratified 10-fold cross-validation results in F1-scores of around 0.14 for every kind of hyperdimensional embedding. This low result is likely due to the possibility of oversaturation of the class vectors.

We can predict the angle between a class vector and a randomly selected vector from said class by  $\Theta = \arccos((\binom{2k}{k})/2^{2k})$  with  $2k + 1$  equal to the number of sequences in the class [47]. This approximation is valid for binary or bipolar vectors in hyperdimensions ( $\geq 10000$ ). This equation also suggests that an increase in dimensions will not influence the angle. Evaluating this equation by considering random 1001 vectors in a class, so  $k = 500$ , results in an angle of  $88.6^\circ$ . This indicates that a vector has a limited capacity: the more vectors we bundle together, the closer the angle will be to  $90^\circ$  and thus the more dissimilar the class vector becomes to its components. This equation assumes that the class vector is a bundle of purely random vectors which is not the case for our embeddings; however, it provides us a rough idea about the bundling capacity of a hyperdimensional vector. Thus, using the rudimentary model works only for very small datasets, like seen in the examples in chapter 2.3. To encode larger datasets, the algorithm has to be more refined.

#### Machine learning models with binary hyperdimensional embeddings

The baseline hyperdimensional classification model has been compared to a more established model, the XGBoost classifier. The results for every embedding with this model are much more comparable to the results of the experiment in the PhaLP paper. This is an indication that hyperdimensional

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computing can provide a very fast and reliable method of embedding protein sequences, even without prior biological information.

## 4. MATERIALS AND METHODS

Every random hyperdimensional vector produced is made to be binary and 10000-dimensional unless noted otherwise.

### 4.1 Amino acid embeddings

To encode biological information into the vectors, we used the last layer of the 3 billion-parameter ESM-2 model [36] of every amino acid, resulting in 1024-dimensional real-valued embeddings for every amino acid. To extend these into hyperdimensionality, a simple matrix multiplication has been employed:  $A_{1 \times 1024} \times B_{1024 \times 10000} = C_{1 \times 10000}$  where  $A$  is an 1024-dimensional ESM-2 embedding and  $B$  a matrix of 1024 random 10000-D vectors. The resulting vectors are then min-max scaled and rounded depending on the desired nature of the vectors.

To encode the surroundings of an amino acid in a sequence, all possible pairwise interactions with the central amino acid in question in a given window were made *via* binding and then all encoded into one vector *via* bundling as shown figure 3.3, starting from ESM embeddings extended into hyperdimensionality as explained earlier. This encoder was tested on the human reference proteome in UniProt, entry *UP000005640*, containing 20591 proteins. After all amino acids were encoded, an element-wise average was made for every amino acid. The resulting hyperdimensional vectors were kept to a real-numbered nature to not lose information for illustration purposes.

### 4.2 Case study: PhaLP dataset

As of March 2023, the latest version of the PhaLP database, *v2021\_04*, has been used to test our models.

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## Sequence embeddings

In chapter 2.3, a method of embedding sequences of amino acids has already been discussed. Here, a sequence of amino acids is considered to be a bag of k-mers. Within a k-mer, the amino acids (presented as randomly generated hyperdimensional vectors) are bonded together with sequential information included. All possible k-mers are all then bundled together, the result is then a hyperdimensional vector representing the whole sequence. We also introduce a novel sequence embedding method within the framework of hyperdimensional computing. It is similar to the bag-of-words method in the sense that it bundles vectors of k-mers, but here, the k-mer's positional information will be encoded into the k-mer before bundling.

## Type classification

Out of the 11549 unambiguous UniParc accessions in the newest version of the database, 4829 are manually annotated on their type. Out of these manually annotated proteins, 2803 are endolysins and 2026 are VALs. As a baseline level, we use the rudimentary HDV classification technique as seen in chapter 2.3: the HDVs of sequences of the same class are bundled to construct single HDVs representative of every class. Then, a sequence's class is inferred by comparing the sequence's HDV to both class HDV *via* a similarity measure based on the assumption that the class vector is maximally similar to its components. This model was evaluated *via* a stratified 10-fold cross validation. The classification with an XGBoost classifier is done via the default XGBoost classifier from *XGBoost.jl* v2.2.5 and is evaluated *via* *MLJ.jl* v0.19.5 with also a stratified 10-fold cross validation.



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## APPENDIX A

# DEFINE BASIC OPERATIONS OF HYPERDIMENSIONAL COMPUTING

All provided code is written in the programming language Julia unless noted otherwise.

## A.1 Binary hypervectors

```
using Random
"""
Construct a binary vector. By default 10000 elements long.
"""
bitHDV(N::Int=10000) = bitrand(N)
"""
Bundles binary hyperdimensional vectors based on the
↳ element-wise majority rule.
"""
function bitadd(vectors::BitVector ...)
    v = reduce(+, vectors)
    n = length(vectors) / 2
    x = [i > n ? 1 : i < n ? 0 : rand(0:1) for i in v]
    return convert(BitVector, x)
end
"""
Binds binary hyperdimensional vectors based on an element-wise
↳ XOR gate.
"""
bitbind(vectors::BitVector ...) = reduce(.⊕, vectors)
"""
```

---

```

Permutates a binary hyperdimensional vector by an adjustable
↳ circular shift.
"""
bitperm(vector::BitVector, k::Int=1) = circshift(vector, k)
"""

Calculates the Hamming distance between two binary vectors.
"""
hamming(x::BitVector, y::BitVector) = sum(x .!= y)/length(x)

```

## A.2 Bipolar hypervectors

```

using Random
using LinearAlgebra
"""

Construct a bipolar vector. By default 10000 elements long.
"""
hdv(N::Int=10000) = rand((-1,1), 1, N)
"""

Bundles bipolar hyperdimensional vectors.
"""
add(vectors::Vector{Int}...) = reduce(+, vectors) .|> sign
"""

Binds bipolar hyperdimensional vectors.
"""
multiply(vectors::Vector{Int}...) = reduce(*, vectors)
"""

Permutates a bipolar hyperdimensional vector by an adjustable
↳ circular shift.
"""
perm(vector::Vector, k::Int=1) = circshift(vector, (0, k))
"""

Calculates the cosine similarity between two bipolar vectors.
"""
cosine(x::Vector{Int}, y::Vector{Int}) = dot(x, y) / (norm(x) *
↳ norm(y))

```



## APPENDIX B

# EXAMPLES OF HYPERDIMENSIONAL COMPUTING ON DATASETS

### B.1 Simple example

```
v = Vector()
for i in 1:10000
    a, b, c, x, y, z = bitHDV(), bitHDV(), bitHDV(), bitHDV(),
        ↪ bitHDV(), bitHDV()
    d = bitadd(bitbind(a, x), bitbind(b, y), bitbind(c, z))
    ap = bitbind(x, d)
    score = hamming(ap, a)
    append!(v, score)
end

using Plots
let
    p = histogram(v, xlims=(0,1), label="", xlabel="Hamming
        ↪ distance")
end
```

### B.2 Zoo example

#### B.2.1 Data preprocessing

```
using DataFrames, CSV
data = CSV.read("zoodata/zoo.csv", DataFrame)
```

---

```

classls = data.class_type

mammal_idx = [i for i in 1:101 if data.class_type[i] == 1]
bird_idx = [i for i in 1:101 if data.class_type[i] == 2]
reptile_idx = [i for i in 1:101 if data.class_type[i] == 3]
fish_idx = [i for i in 1:101 if data.class_type[i] == 4]
amphibian_idx = [i for i in 1:101 if data.class_type[i] == 5]
bug_idx = [i for i in 1:101 if data.class_type[i] == 6]
invertebrate_idx = [i for i in 1:101 if data.class_type[i] ==
    ↪ 7]

indices = [mammal_idx, bird_idx, reptile_idx, fish_idx,
    ↪ amphibian_idx, bug_idx, invertebrate_idx]

legs0 = [i == 0 ? 1 : 0 for i in data.legs]
legs2 = [i == 2 ? 1 : 0 for i in data.legs]
legs4 = [i == 4 ? 1 : 0 for i in data.legs]
legs5 = [i == 5 ? 1 : 0 for i in data.legs]
legs6 = [i == 6 ? 1 : 0 for i in data.legs]
legs8 = [i == 8 ? 1 : 0 for i in data.legs]

select!(data, Not(:legs))

data[:, :legs0] = legs0
data[:, :legs2] = legs2
data[:, :legs4] = legs4
data[:, :legs5] = legs5
data[:, :legs6] = legs6
data[:, :legs8] = legs8

```

## B.2.2 Assigning HDVs to features

```

"""
Creates a list of binary hyperdimensional vectors representing
    ↪ an interval of numbers by constructing a random HDV
    ↪ representing the lower bound of the interval and replacing
    ↪ a fraction of the vector with random bits.
"""

```

## B. Examples of hyperdimensional computing on datasets

---

```
function range_hdvs(steps)
    k = length(steps) - 1
    V = [bitHDV() for i in 1:k+1]
    for i in 2:k+1
        for j in 1:10000
            V[i][j] = rand(0:1)
        end
    end
    return V
end

legs_steps = 0:1:8
legs_hdvs = range_hdvs(legs_steps)

feature_hdv=[bitHDV() for i in 1:15]
append!(feature_hdv, [legs_hdvs[1], legs_hdvs[3], legs_hdvs[5],
    ↪ legs_hdvs[6], legs_hdvs[7], legs_hdvs[9]])

growth_hdv = bitHDV()
skin_protection = bitHDV()
limbs = bitHDV()

for i in 1:2
    feature_hdv = replace!(feature_hdv, feature_hdv[i] =>
    ↪ bitbind(feature_hdv[i], skin_protection))
end

for i in 3:4
    feature_hdv = replace!(feature_hdv, feature_hdv[i] =>
    ↪ bitbind(feature_hdv[i], growth_hdv))
end

for i in 12:13;16:21
    feature_hdv = replace!(feature_hdv, feature_hdv[i] =>
    ↪ bitbind(feature_hdv[i], limbs))
end

hdv = BitVector[]
for i in 1:101
```

---

```

v = BitVector[]
for j in 2:16;18:23
    if data[:, j][i] == 1
        if j < 18
            push!(v, feature_hdv[j-1])
        end
        if j > 17
            push!(v, feature_hdv[j-2])
        end
    end
end
x = bitadd(v...)
push!(hdv, x)
end

data[:, :species_hdv] = hdv

function grouper(group::Int)
    v = BitVector[]
    for i in 1:length(indices[group])
        push!(v, data.species_hdv[indices[group][i]])
    end
    x = bitadd(v...)
    return x
end

mammal_hdv = grouper(1)
bird_hdv = grouper(2)
reptile_hdv = grouper(3)
fish_hdv = grouper(4)
amphibian_hdv = grouper(5)
bug_hdv = grouper(6)
invertebrate_hdv = grouper(7)

```

### B.2.3 Data analysis

```
list_group_hdvs = [mammal_hdv, bird_hdv, reptile_hdv, fish_hdv,  
↳ amphibian_hdv, bug_hdv, invertebrate_hdv]  
matrix_groups = permutedims(hcat(list_group_hdvs...))
```

**data.species\_hdv[94] refers to the 94th animal in the dataset, in this case that is vampire.**

```
println("Similarity to mammal =  
↳ ",string(hamming(data.species_hdv[94], mammal_hdv)))  
println("Similarity to bird  
↳ =",string(hamming(data.species_hdv[94], bird_hdv)))  
println("Similarity to reptile  
↳ =",string(hamming(data.species_hdv[94], reptile_hdv)))  
println("Similarity to amphibian  
↳ =",string(hamming(data.species_hdv[94], amphibian_hdv)))  
println("Similarity to bug  
↳ =",string(hamming(data.species_hdv[94], bug_hdv)))  
println("Similarity to fish  
↳ =",string(hamming(data.species_hdv[94], fish_hdv)))  
println("Similarity to invertebrate  
↳ =",string(hamming(data.species_hdv[94], invertebrate_hdv)))  
  
axolotl_features = [0, 0, 1, 0, 0, 1, 0, 1, 1, 1, 0, 0, 1, 0,  
↳ 0, 0, 0, 1, 0, 0, 0]  
v = BitVector[]  
    for j in 1:16  
        if axolotl_features[j] == 1  
            push!(v, feature_hdv[j])  
        end  
    end  
axolotl_hdv = bitadd(v...)
```

```
using MultivariateStats  
M = fit(PCA, matrix_groups ; maxoutdim=2)  
proj = projection(M)
```

---

```

using Plots
function plotter()
    colors = [:green, :orange, :blue, :red, :yellow, :purple,
        ↪ :black]
    labels = ["mammal", "bird", "reptile", "fish", "amphibian",
        ↪ "bug", "invertebrate"]
    fig = Plots.plot()

    for i in 1:7
        scatter!(fig, (proj[i, 1], proj[i, 2]),
            ↪ label=labels[i], mc = colors[i])
    end
    return fig
end
plotter()

```

```

matrix_species = permutedims(hcat(hdv...))
S = fit(PCA, matrix_species ; maxoutdim=2)
projS = projection(S)

```

```

using Plots
function plotter()
    colors = [:green, :orange, :blue, :red, :yellow, :purple,
        ↪ :black]
    labels = ["mammal", "bird", "reptile", "fish", "amphibian",
        ↪ "bug", "invertebrate"]
    fig = Plots.plot()

    for i in 1:7
        scatter!(fig, projS[indices[i], 1], projS[indices[i],
            ↪ 2], mc = colors[i], label=labels[i])
    end
    return fig
end
plotter()

```

### B.2.4 Animal classifier

```
function predict(animal)
    y = [hamming(mammal_hdv, animal), hamming(bird_hdv,
        ↪ animal), hamming(reptile_hdv, animal),
        ↪ hamming(fish_hdv, animal), hamming(amphibian_hdv,
        ↪ animal), hamming(bug_hdv, animal),
        ↪ hamming(invertebrate_hdv, animal)]
    return findmin(y)[2]
end
using StatsBase
pred = [predict(i) for i in data.species_hdv]
mean(classls .== pred)
```

## B.3 Anticancer protein example

### B.3.1 Data preprocessing

```
using DataFrames, CSV
data = CSV.read("ProtExdata/ACPs_Breast_cancer.csv", DataFrame)

unique(data.class)
class_num = [i == "very active" ? 1 : i == "mod. active" ? 2 :
    ↪ i == "inactive - exp" ? 3 : 4 for i in data.class]
data[:, :class_num] = class_num
```

### B.3.2 Assigning HDVs to features

```
AA_list = ['A', 'R', 'N', 'D', 'C', 'Q', 'E', 'G', 'H', 'I',
    ↪ 'L', 'K', 'M', 'F', 'P', 'S', 'T', 'W', 'Y', 'V', 'O', 'U',
    ↪ 'B', 'J', 'Z', 'X']
AA_hdv = [bitHDV() for i in AA_list]
AA_dict = Dict(zip(AA_list, AA_hdv))

trimer_hdvs = Dict(aa1 * aa2 * aa3 =>
```

---

```

bitbind(AA_dict[aa1], bitperm(AA_dict[aa2]),
  ↳ bitperm(AA_dict[aa3], 2))
for aa1 in AA_list for aa2 in AA_list for aa3 in AA_list)

function embedder(sequence)
  l = [trimer_hdvs[sequence[i:i+2]] for i in
  ↳ 1:length(sequence)-2]
  v = bitadd(hcat(l)...)
  return v
end
l = BitVector[]
for i in data.sequence
  push!(l, embedder(i))
end
data[!, :hdv] = l

active_hdv = bitadd(hcat([i for i in data[data.class_num .== 1,
  ↳ :hdv]]))...)
modactive_hdv = bitadd(hcat([i for i in data[data.class_num .==
  ↳ 2, :hdv]]))...)
notactive_exp_hdv = bitadd(hcat([i for i in data[data.class_num
  ↳ .== 3, :hdv]]))...)
notactive_virt_hdv = bitadd(hcat([i for i in
  ↳ data[data.class_num .== 4, :hdv]]))...)

```

### B.3.3 Data analysis

**Print the similarity of the HDV of sequence "KWKLFFKKILKFLH-LAKKF" to every class HDV**

```

println(hamming(data[data.sequence .== , :hdv]..., active_hdv))
println(hamming(data[data.sequence .== "KWKLFFKKILKFLH-LAKKF",
  ↳ :hdv]..., modactive_hdv))
println(hamming(data[data.sequence .== "KWKLFFKKILKFLH-LAKKF",
  ↳ :hdv]..., notactive_exp_hdv))
println(hamming(data[data.sequence .== "KWKLFFKKILKFLH-LAKKF",
  ↳ :hdv]..., notactive_virt_hdv))

```



**using** MultivariateStats

```
matrix_all = permutedims(hcat(data.hdv...))
```

```
S = fit(PCA, matrix_all; maxoutdim=2)
```

```
projS = projection(S)
```

```
indices = [[i for i in 1:nrow(data) if data.class_num[i] == j]
  ↪ for j in 1:4]
```

**using** Plots

**function** plotter()

```
    colors = [:green, :black, :blue, :red]
```

```
    labels = ["active", "mod. active", "inactive (exp)",
```

```
    ↪ "inactive (virt)"]
```

```
    fig = Plots.plot()
```

```
    for i in 1:4
```

```
        scatter!(fig, (projS[indices[i], 1], projS[indices[i],
```

```
        ↪ 2]), label=labels[i], mc = colors[i])
```

```
    end
```

```
    return fig
```

**end**

```
plotter()
```

### B.3.4 Classifier

```
n = nrow(data)
```

```
train = rand(n) .< 0.8
```

```
test = train = .! train
```

```
train_df = data[[i for i in 1:n if train[i] == 1], :]
```

```
test_df = data[[i for i in 1:n if test[i] == 1], :]
```

```
active_hdv_t = bitadd(hcat([i for i in
```

```
    ↪ train_df[train_df.class_num .== 1, :hdv]))...
```

```
modactive_hdv_t = bitadd(hcat([i for i in
```

```
    ↪ train_df[train_df.class_num .== 2, :hdv]))...
```

---

```
notactive_exp_hdv_t = bitadd(hcat([i for i in
    ↳ train_df[train_df.class_num .== 3, :hdv]])...))
notactive_virt_hdv_t = bitadd(hcat([i for i in
    ↳ train_df[train_df.class_num .== 4, :hdv]])...))

function predict(seq)
    y = [hamming(active_hdv_t, seq), hamming(modactive_hdv_t,
        ↳ seq), hamming(notactive_exp_hdv_t, seq),
        ↳ hamming(notactive_virt_hdv_t, seq)]
    return findmin(y)[2]
end

using StatsBase
pred = [predict(i) for i in test_df.hdv]
mean(test_df.class_num .== pred)
```