

0) Files description

Target file

The group you want to find candidate regions

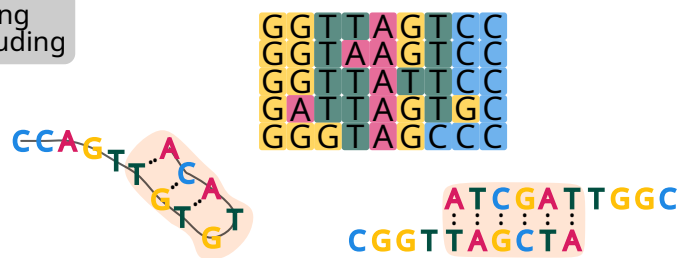
Excluding file

The sequences you want to exclude (or the rest of the diversity)

2) Test oligos

Test oligonucleotides for self-dimers, hairpin, mismatches, ...

testOligo -f primers -e excluding
testThorough -f oligos -e excluding

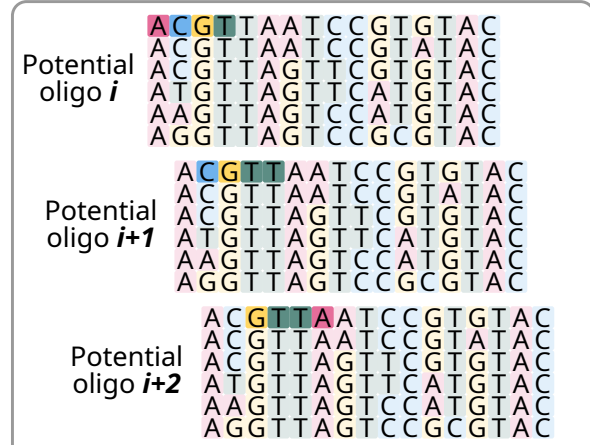
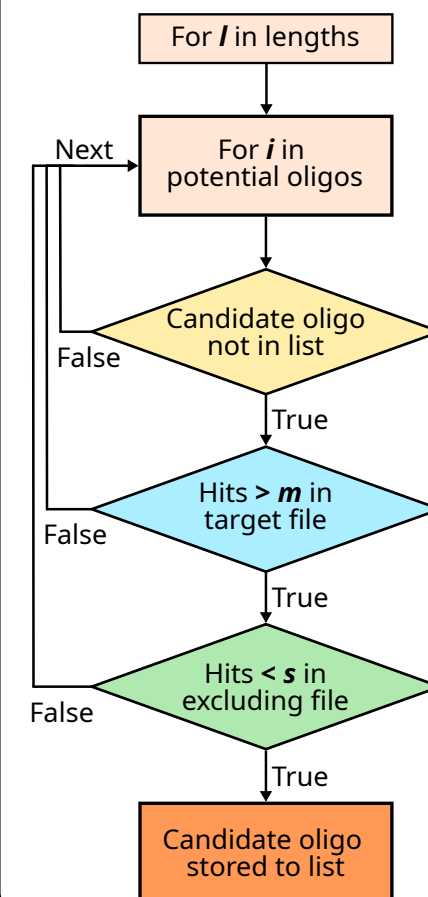


- Self-dimer count
- Hairpin count
- Max consecutive bases
- Hits allowing n mismatches
 - Position of the mismatch
 - Identity of the mismatch
- ...

1) Find oligos

Searches candidate oligonucleotides by sliding window

findOligo -t target -e excluding -l length -s specificity -m minMatch



- Sequence
- Reverse complement sequence
- GC content
- Basic melting temperature
- Hits in target file
- Hits in excluding file

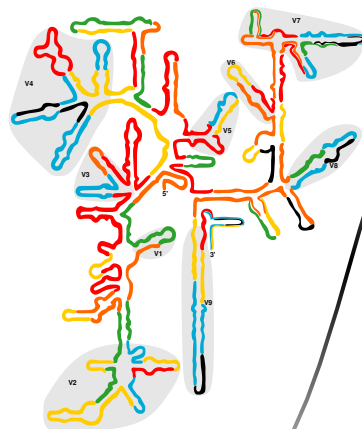
3) Rate accessibility

Compare candidate oligonucleotides to the accessibility map of *Saccharomyces cerevisiae* 18S rDNA or *Escherichia coli* 16S (Behrens et al., 2003)

```
alignPrimers -t target.fasta -p oligos.fasta -o oligos_align.fasta
rateAccess -f oligos_align.fasta -o oligos_access.tsv
```



- Accessibility score
- Region in the 18S rDNA gene



18S rDNA
accessibility:

4) Get homologs

Get all aligned positions from the excluding
file matching a HMM profile of a
given region of the target file

```
getHomolog -f region.fasta -e excluding.fasta
```



5) Selecting oligonucleotides

Merge all log files:

```
bindLogs -f oligos.tsv oligos_tested.tsv oligos_access.tsv -o oligos_log.tsv -r
```

The diagram illustrates the hierarchical structure of the data tables. It shows a large table on the left that branches into two smaller tables. The top table has columns: identifier, sequence, sequence_revCom, GC, Tm, hits_target_abs, hits_target_excluding, hits_ref_abs. The bottom table has columns: identifier, sequence, mismatch1, mismatch1_abs, mismatch2, mismatch2_abs. The bottom table further branches into two more tables. The top of these has columns: identifier, sequence, start_position, region, Scerevisae_spos, average_max_bright, average_min_bright, average_bright, class. The bottom of these has columns: identifier, length, sequence, sequence_revCom, GC, Tm, hits_target_abs, hits_target_excluding, hits_ref_abs, mismatch1, mismatch1_abs, mismatch2, mismatch2_abs, Scerevisae_spos, average_max_bright, average_min_bright, average_bright, class.

Selecting the best oligonucleotide:

- covering most of the targeted diversity
- with a high GC content
- with similar theoretical melting temperature
- with low hits to the excluding file allowing mismatches
- highly accessible

For example:

```
filterLog -l oligos log.tsv -s "0.4" -m "0.001" -M "0.0001" -c "III"
```

```
selectLog -l oligos_log_filtered.tsv -n "4"
```

6) Concluding remarks

Complementary softwares:

- **ARB** (Ludwig et al., 2004)
- **Decipher** (Wright et al., 2014)
- **primer3** (Untergasser et al., 2012)
- **oli2qo** (Hendling et al., 2018)

Post-hoc test:

- **OligoCalc**
- **BLAST** (blast.ncbi.nlm.nih.gov/Blast.cgi)
- **PR2-primers** (app.pr2-primers.org/)

Remember that oligo design is a tedious work that requires a final empirical test for its completion.

Therefore, bioinformatic pipelines will only provide theoretical candidate oligos, that have to be tested in the laboratory