

## MIKIAS H. WOLDETENSAE T.

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## **CAREER SUMMARY**

### Wet-lab & Dry-lab Research • Health/Medicine • Design/Optimization • Regulatory/Protocols

- A passionate biologist with an engineering perspective that was developed through 6+ years of wet lab and dry lab research experience and a lifetime of asking how things work on the inside.
- A proactive contributor that views every problem as a personal problem and finds creative solutions that reduce effort, cost and increase efficiency.
- An enthusiastic student with endless energy for new challenges; instinctively weaving new lessons into a preexisting tapestry of foundational concepts.

## **AREAS OF EXPERTISE**

### **METHODS**

- Oligos | Plasmids
- PCR | qPCR | RTqPCR
- Spectroscopy
- Chromatography
- Protein Assays
- Western Blot
- ELISA | ELISpot
- Cell Assays
- Confocal Microscopy
- Flow Cytometry

# MODELS

- Transgenic Mice
- Mammalian Cell Cultures
- 3D Cell Cultures

#### **DATA ANALYSIS**

- R | Python
- Power BI
- Statistical Modeling
- Cytometry
- RNA Sequencing
- Spatial Sequencing

#### **LAB MANAGEMENT**

- Safety Regulations
- IACUC Protocol
- Experimental Protocol Design
- Optimization
- Inventory Management
- Cost Reduction
- Trainer | Instructor

#### **DESIGN**

- 3D Printing
- Arduino

# **PROFESSIONAL EXPERIENCE**

### Benarova Research Institute, Ziegler Lab, Seattle, WA

Aug 2019 - Present

A nonprofit biomedical research institute focused on advancing science to predict, prevent, reverse and cure diseases of the immune system.

#### **Research Technician**

Served standard and investigative laboratory support roles in aid of scientific research. Acted as safety representative. Maintained 60+/- transgenic mouse models. Performed independent research projects.

- *Performed risk assessment of lab spaces* per CDC recommendations and updated Institutional Biosafety Committee protocols and Biosafety level ratings to maintain regulatory compliance.
- Validated and characterized three novel humanized mouse models to study Rhinoviruses, conducting In Vitro and In Vivo infections with 7 different species of Rhinovirus as proof of concept.
- Reverse engineered and optimized chromatographic extraction of DNA, RNA, and/or Protein to reduce costs by more than 80% per sample and increase flexibility.
- Prepared and maintained skin and lung, fibroblast, and epithelial primary cell cultures, to confirm infective capability of mouse models in Vitro.
- Streamlined inventory management methods resulting in ~20% reduction in wasted resources.
- *Designed and optimized various multiplexed oligonucleotides*, allowing for the interrogation of DNA and mRNA products at tissue or single cell resolution.

#### Bloodwork's Northwest, North Seattle, WA

June 2017 – Aug 2019

Nonprofit organization that provides health care services to over 90 hospitals in the Northwest by collecting, testing, and distributing lifesaving blood.

#### Phlebotomist | Medical Assistant

Perform health and history assessment for volunteer and medically prescribed blood donors, Following strict SOP and HIPPA regulations. Collect Whole Blood, Plasma, Platelets, and/or Red Blood Cells.

- Collected over 3500 units of blood components | 98% percentile success rate.
- Trained 18 Phlebotomists on proper venipuncture techniques and SOP/HIPPA regulations.
- Suggested and proved the benefit of syncope preventative measures that were not included in introductory training | Increased confidence in various donors with tendencies for adverse reactions.

#### **UW Human Photonics Lab**, Seattle, WA

**Sept 2011 – March 2013** 

UW Mechanical Engineering research laboratory focused on medical devices and optical technologies in the areas of enhancing human performance, detecting diseases, and guiding their treatments.

#### **Undergraduate Researcher**

Read relevant research articles,. Propose and lead novel experiments with Cancerous and Non-Cancerous cells, collect data, analyze results, present & publish findings.

- *Optimized power conditions of a proprietary Scanning Fiber Endoscope* for the fluorescence induced apoptosis and necrosis of cancer cells treated with a cancer biomarker 5-aminolevulinic acid and its photosensitive metabolite, Protoporphyrin IX.
- *Published Paper* M. H. Woldetensae et al., "Fluorescence image-guided photodynamic therapy of cancer cells using a scanning fiber endoscope," Proc. SPIE 8576, 85760L (March 20, 2013).

## **EDUCATION**

Bachelor of Science, Molecular, Cellular & Developmental Biology

2011-2016

University of Washington, Seattle, WA

Minor in Mathematics

**Bachelor of Art, Psychology** 

2011-2016

University of Washington, Seattle, WA

# **TEACHING EXPERIANCE**

Bioethics Instructor June-Aug 2016

University of Washington, ALVA & Clean Energy, GenOM Summer Program

First-Year Interest Group Instructor

University of Washington, First Year Programs

Sept-Dec 2015

# **SERVICE EXPERIENCE**

### **Safety Committee Member**

Nov 2021 - Nov 2022

Benaroya Research Institute, Seattle WA

# **COMMUNICATION**

**Tigrinya:** Native language **Dutch:** 1999 - 2004 **English:** 2004 - present